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ORIGINAL ARTICLE

Sequence variations of *ABCB1*, *SLC6A2*, *SLC6A3*, *SLC6A4*, *CREB1*, *CRHR1* and *NTRK2*: association with major depression and antidepressant response in Mexican-Americans

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We studied seven genes that reflect events relevant to antidepressant action at four sequential levels: (1) entry into the brain, (2) binding to monoaminergic transporters, and (3) distal effects at the transcription level, resulting in (4) changes in neurotrophin and neuropeptide receptors. Those genes are ATP-binding cassette subfamily B member 1 (ABCB1), the noradrenaline, dopamine, and serotonin transporters (SLC6A2, SLC6A3 and SLC6A4), cyclic AMP-responsive element binding protein 1 (CREB1), corticotropin-releasing hormone receptor 1 (CRHR1) and neurotrophic tyrosine kinase type 2 receptor (NTRK2). Sequence variability for those genes was obtained in exonic and flanking regions. A total of 56 280 000 bp across were sequenced in 536 unrelated Mexican Americans from Los Angeles (264 controls and 272 major depressive disorder (MDD)). We detected in those individuals 419 single nucleotide polymorphisms (SNPs); the nucleotide diversity was 0.00054 ± 0.0001. Of those, a total of 204 novel SNPs were identified, corresponding to 49% of all previously reported SNPs in those genes: 72 were in untranslated regions, 19 were in coding sequences of which 7 were non-synonymous, 86 were intronic and 27 were in upstream/downstream regions. Several SNPs or haplotypes in ABCB1, SLC6A2, SLC6A3, SLC6A4, CREB1 and NTRK2 were associated with MDD, and in ABCB1, SLC6A2 and NTRK2 with antidepressant response. After controlling for age, gender and baseline 21-item Hamilton Depression Rating Scale (HAM-D21) score, as well as correcting for multiple testing, the relative reduction of HAM-D21 score remained significantly associated with two NTRK2-coding SNPs (rs2289657 and rs56142442) and the haplotype CAG at rs2289658 (splice site), rs2289657 and rs2289656. Further studies in larger independent samples will be needed to confirm these associations. Our data indicate that extensive assessment of sequence variability may contribute to increase understanding of disease susceptibility and drug response. Moreover, these results highlight the importance of direct re-sequencing of key candidate genes in ethnic minority groups in order to discover novel genetic variants that cannot be simply inferred from existing databases.

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Introduction

Major depressive disorder (MDD) is a common, complex and recurrent disorder of gene–environment interactions. The estimated heritability may range from 0.36 to 0.66.^{1,2} Following up on previous study on the pathophysiology of MDD and on the prevailing hypotheses for treatment response, we sought to

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identify genes that influence susceptibility for MDD or treatment response in the central nervous system pathways relevant to stress reactivity and to the pathways of action of antidepressant drugs. Current data point out to roles for genes involved in drug transport, serotonin neurotransmission, neurotrophin signaling and response to stress. Promising linkage results are located in several chromosomes,³ which highlight the multilocus nature of the genetic vulnerability to MDD.

Recently, rapid technological advances have started unraveling the contributions of common (frequency >1%) and rare genetic variants in complex disorders. In a topical review, Bodmer and Bonilla⁴ have synthesized current views, implications and

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integration of the competing hypotheses of common disease-common variant and common disease-rare variant. For most common variants, the diseaseassociated variant is unlikely to be functionally relevant; it may be closely linked to the functional variant, and it will cause a small increase in disease risk (odds ratio smaller than 2, generally between 1.1 and 1.4). In contrast, rare variants generally have functional and large phenotypic effects; in many cases they are missense variants that reflect aminoacid changes relevant to protein-protein interactions. Diverse scenarios may occur in the pathophysiology of common complex disorders: Common variants may be modifiers of genes with rare variant effects, such as recently described for the MC4R gene.⁵ Moreover, areas near common variants may contain candidate genes in which there are rare variants. The identification of rare variants may significantly affect our understanding of complex disease etiology.

We re-sequenced seven candidate genes of importance in the pathophysiology of MDD.⁶ Conceptually, we sought a group of genes that reflects a sequence of events relevant to drug action at four levels: (1) entry into the brain, (2) binding to monoaminergic transporters, and (3) distal effects at the transcription level, resulting in (4) changes in neurotrophin and neuropeptide receptors. Specifically, we studied a bloodbrain barrier drug transporter pump (ACCB1, also called MDR1), which regulates drug entry into the brain (level 1), the norepinephrine, dopamine, and serotonin transporters (SCL6A2, SLC6A3 and SL6A4) (level 2), an antidepressant-regulated transcription factor (cyclic AMP-responsive element binding protein 1 (*CREB1*)) (level 3) and two receptors (level 4): neurotrophic tyrosine kinase type 2 receptor (NTRK2), important in synaptic function and neural plasticity, and corticotropin-releasing hormone receptor 1 (*CRHR1*), which regulates the response to stress at the behavioral, neuroimmune and neuroendocrine—hypothalamic-pituitary-adrenalaxis levels.

Materials and methods

Patients and controls

The study consisted of 272 patients (66% female, 34% male; mean age: 38 ± 10) with MDD and 264 healthy control individuals (60% female, 40% male; average age: 36 ± 11). MDD was defined as a DSM-IV (Diagnostic and Statistical Manual of Mental Disorders, 4th Edition) diagnosis of current, unipolar major depressive episode and a 21-item Hamilton Depression Rating Scale (HAM-D21) score of ≥ 18 with item number 1 (depressed mood) rated ≥ 2 . All MDD patients were screened for the pharmacogenetic study of antidepressant treatment response as previously described.7 All MDD patients had comprehensive psychiatric and medical assessments in their primary language, on the basis of diagnostic and ratings instruments that had been fully validated in English and in Spanish. Exclusion criteria included active medical illnesses that could be etiologically related to the ongoing depressive episode, current or active suicidal ideation with a plan and strong intent, pregnancy, lactation, current use of medications with significant central nervous system activity, which interfere with electroencephalogram (EEG) activity (for example, benzodiazepines) or any other antidepressant treatment within the 2 weeks before enrollment, illicit drug use and/or alcohol abuse in the last 3 months or current enrollment in psychotherapy. All MDD patients were Mexican-Americans and had at least three grandparents born in Mexico.

All patients had an initial comprehensive psychiatric and medical assessment and, if enrolled in the pharmacogenetic study of antidepressant treatment response, had weekly structured follow-up assessments for 9 weeks. The study consisted of two phases: а 1-week single-blind placebo lead-in phase to minimize the impact of placebo responders followed. if subjects continued to meet the inclusion criteria after phase 1, by random assignment to one of the two treatment groups: fluoxetine 10–40 mg per day or desipramine 50-200 mg per day, administered in a double-blind manner for 8 weeks. Our primary clinical outcome measure was HAM-D21 score and clinical remission on antidepressants was defined as having a final (week 8) HAM-D21 score <8. In addition, the relative response change was also computed as the difference in HAM-D21 score between pre- and post-treatment divided by the pretreatment HAM-D21 score.

Age-, gender- and ethnicity-matched healthy control individuals were recruited from the same Mexican-American community in Los Angeles by the same bilingual clinical research team. Controls for our genomic studies were in general good health but were not screened for medical or psychiatric illness.

Genomic DNA collection, amplification and sequencing

At the initial visit, after informed consent was obtained from the participating individuals, blood samples were collected into EDTA (K2EDTA) BD Vacutainer EDTA tubes (Becton Dickinson, Franklin Lakes, NJ, USA), and genomic DNA was isolated by using Gentra Puregene DNA purification kits (Gentra Systems, Indianapolis, IN, USA). DNA sequencing for seven genes was carried out in collaboration with the Sanger Institute by following ExoSeq protocol (http:// www.sanger.ac.uk/humgen/exoseq/). Briefly, the protein-coding regions, novel known coding sequences and transcripts, exons and their flanking sequence were extracted from the Vega database (http://vega.sanger.ac.uk/index.html). Primers were designed automatically using Primer3 (http://frodo. wi.mit.edu/) to amplify DNA and primer pairs were checked for uniqueness before ordering and prescreened to determine the optimum conditions for amplification. After amplification, a sample of the products were visualized on an agarose gel to confirm the size of the PCR product. The remaining PCR product was then cleaned up using two enzymes,

Exonuclease 1 and Shrimp Alkaline Phosphatase. Bidirectional sequencing of amplicons was carried out using Big DyeTM chemistry (Big Dye Terminator, Version 3.1; Applied Biosystems, Foster City, CA, USA). Single nucleotide polymorphisms (SNPs) were called using ExoTrace http://www.sanger.ac.uk/ humgen/exoseq/analysis.shtml, a novel algorithm developed in-house for the detection of heterozygotes in sequence traces , which processes the sense and antisense sequence reads separately and subsequently, and combines the results to allow SNP scoring. All polymorphisms reported here had a genotyping rate of $\geq 80\%$ and an average nucleotide call rate of 93%.

Genomic control genotyping

To detect potential bias due to population stratification, two approaches were used to test for hidden stratification in our data. First, 54 independent SNPs across 22 autosomal chromosomes were selected to analyze a combined sample using the genotype data download from three HapMap ethnic samples using STRUCTURE program (http://pritch.bsd.uchicago. edu/software.html)8,9 and showed that three distinct clusters were well identified with an average proportion of at least 92% of individuals correctly assigned to the given ethnic populations (CEU, CHB+JPT, YRI). This panel of SNPs were then used as genomic control to test our sample and showed an almost equal proportion assigned to each clusters, given K=2, 3, 4 in both cases and controls. Second, genotype frequencies from each of the 54 unlinked SNPs were also compared between cases and controls using the method described by Pritchard and Rosenberg ¹⁰ and no significant difference was found based on an overall test statistic ($\chi^2 = 100.50$, d.f. = 108, P = 0.68). Therefore, no population stratification adjustment was necessary for our association analyses.

Nucleotide diversity, population differentiation and Hardy–Weinberg equilibrium

Nucleotide diversity (θ) and its standard deviation $(S(\theta))$ were calculated under the assumption of an infinite neutral allele model,^{11,12} and all calculations were based on n = 946 for all the sites given that the average sample size was 473 individuals across all the polymorphisms. Population differentiation estimation was based on the pairwise F_{ST} values for the dbSNPs (single nucleotide polymorphism database hosted at the National Center for Biotechnology Information), which were both detected in our Mexican-American sample and reported in HapMap sample. F_{ST} values were calculated as described byWeir,¹³ Weir and Cockerham,¹⁴ and Weir and Hill.¹⁵ In order to compare allele frequencies and to be able to treat chromosomes as independent observations, the genotype frequencies must be in Hardy–Weinberg equilibrium (HWE).¹⁶ Exact testing of HWE was performed separately for healthy controls and MDD patients using the PLINK program Version1.00 (http:// pngu.mgh.harvard.edu/~purcell/plink/).¹⁷ SNPs that

were not in HWE in the healthy control group were excluded from the allele-based association analyses of cases and controls.

Statistical analysis

Data preparation and descriptive statistics were carried out with SAS software (SAS Version 9.1.3, SAS Institute, Cary, NC, USA). For SNP-based association analyses of case vs control or remitter vs non-remitter, Fisher's exact test (two-tailed) was performed to compare allele and genotype distributions between depressed and healthy individuals using PLINK. In the allelic association analysis, each polymorphism was tested in controls to ensure the fitting with HWE; the odds ratio on the 2×2 contingency table of allele counts and its 95% confidence interval were estimated using Woolf's method or fitting exact logistic regression model with SAS software when the frequency in a table cell is zero.¹⁸ In genotypic association analysis, SNP effects were tested under a codominant model on the 2×3 contingency table of genotype counts.

For the quantitative outcome (relative reduction % in HAM-D21 scores between pre- and post-treatment), the analyses based on dominant model were performed, separately, for the joint sample of patients treated with desipramine or fluoxetine and for medication-specific sample. General linear regression models were used to examine the association between genotype and relative HAM-D21 score reduction by controlling for age, gender and baseline (pretreatment) HAM-D21 score using the PLINK program. The Benjamini and Hochberg method was used to control for false discovery rate and the significance threshold was set at FDR_BH $\leq 0.05^{19}$.

For haplotype-based association analysis, Haploview (Version 4.1, Broad Institute of MIT and Harvard, http://www.broad.mit.edu/mpg/haploview/), was first used to identify the haplotype blocks by applying the Four Gamete Rule²⁰ based on the SNPs with a minor allele frequency (MAF) ≥ 0.01 in the combined sample of cases and controls and HWE exact test P > 0.01 in controls. The PLINK program was then used to examine the association of specific haplotype with depression diagnosis, clinical remission, as well as quantitative outcome of antidepressant treatment.

Results

Identification of sequence variations

A total of 419 single nucleotide sequence variants (Table 1) were identified by re-sequencing of ~105 kb of exonic sequence and their flanking regions in the selected seven genes in an ethnically homogeneous sample of 264 healthy controls and 272 MDD patients. Among the 419 SNPs, 204 (49%) are novel polymorphisms, not previously described, including 86 in introns, 72 in untranslated regions (UTRs), 19 (12 synonymous) in coding regions, 18 in upstream and 9 in downstream regions. Overall, 95% of the novel polymorphisms had a MAF lower than 5%,

Gene (Location)											
	SNP ^a	Downstream	3' UTR	Intronic	SYN	SN	5' UTR	Upstream	AII	Sequence screened (kb)	Nucleotide diversity $(s.d.) imes 10^{-4}$
CREB1 (2q34) CREB1 (2q34)	New JbSNP Total	3 1 5	4 7 2	7 7 2	0 1 1	000	000	000	12 6 18	7.5	3.2 (0.9)
SLC6A3 (5p15.3) C c 1	New JbSNP Total	000	0 1 0	6 10 16	7 2 7	7 7 7	000	7 7 4	18 30 48	12.3	5.3 (1.2)
ABCB1 $(7q21.1)$ 1	New JbSNP Total	000	044	20 37 57	3 7 1	യവന	4 4 8	0 1 1	28 53 81	28.5	3.8 (0.8)
<i>NTRK2 (9q22.1)</i> 6 7	New JbSNP Total	000	43 24 67	10 6 16	0 2 4	0 0 7	000	000	57 32 89	23.6	5.1 (1.0)
SLC6A2 (16q12.2) C c 1	New JbSNP Total	000	448	10 9 19	2 2 4	თ 10 H	101	$\begin{array}{c} 11\\ 9\\ 20 \end{array}$	29 26 55	13.3	5.6 (1.2)
SLC6A4 (17q11.1-q12) 1 c 1	New JbSNP Total	ю 4 0	4 H U	23 23 46	044	4 L D	0 7 J	000	37 35 72	12.5	7.8 (1.6)
CRHR1 (17q12-q22) CRHR1 (17q12-q22)	New JbSNP Total	0 H O	8 12 20	12 16 28	0 ო ო	7 7 7	000	000	23 33 56	6.9	10.9(2.4)
All seven genes	New JbSNP Total	9 6 15	66 54 120	86 103 189	7 19 26	12 10 22	6 6 12	18 17 35	204 215 419	104.5	5.4 (1.0)

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Figure 1 Minor allele frequency (MAF), nucleotide diversity and F_{ST} measure in seven candidate genes in Mexican-American major depressive disorder (MDD) patients and controls. Histograms show the total number of single nucleotide polymorphisms (SNPs) detected in intronic (black bar) and exonic (gray bar) regions in the seven genes by MAF in 272 MDD patients and 264 healthy controls (a); the nucleotide diversity in noncoding (black bar) and coding (gray bar) (b) or intronic (black bar) and exonic (gray bar) regions (c) by gene in the combined sample of 272 MDD patients and 264 healthy controls; the total number of SNPs shared by Mexican-American (MA) sample and HapMap samples by pairwise F_{ST} value (d) or represent the average F_{ST} by gene (e) in MA vs CEU (black bar), MA vs HCB (dark gray bar), MA vs JPT (gray bar) and MA vs YRI (White bar).

whereas the corresponding proportion was 57% for dbSNPs (Supplementary Table 1). Similar distribution of MAFs was seen between cases and controls for both SNPs in intronic and in exonic regions (Figure 1a). Among the 419 SNPs, the proportion of SNPs with HWE exact test *P*-value ≥ 0.05 was 92% for controls and 91% for MDD cases (Supplementary Table 1).

Nucleotide diversity was estimated for each gene by correcting for both sample size and length of the screened site (Table 1). Nucleotide diversities were comparable in *SLC6A3* (0.00053 \pm 0.00012), *NTRK2* (0.00051 \pm 0.0001) and *SLC6A2* (0.00056 \pm 0.00012), but were lower in *CREB1* (0.00032 \pm 0.00009) and ATP-binding cassette subfamily B member 1 (*ABCB1*)

(0.00038 \pm 0.00008) and appeared higher in *SLC6A4* (0.00078 \pm 0.00016) and *CRHR1* (0.00109 \pm 0.00024). This led to an overall nucleotide diversity of 0.00054 for all the seven genes investigated. When the nucleotide diversity was estimated separately for coding and noncoding sequence, five out of seven genes (except for *SLC6A3* and *ABCB1*) showed higher nucleotide diversity in noncoding regions when compared with coding segments (Figure 1b). However, when the nucleotide diversity was estimated separately for exonic and intronic sequence, all the seven genes showed higher nucleotide diversity in exonic regions than in intronic segments (Figure 1c). This is because of the high nucleotide diversity in untranslated regions (0.00088 \pm 0.00017).

Among the 215 dbSNPs detected, 83 were reported in all four HapMap ethnic groups: CEU (Caucasian), YRI (African), CHB (Han Chinese) and JPT (Japanese) in the NCBI database as of 25 June 2008. Pairwise F_{ST} values between Mexican Americans (MA) and each HapMap ethnic sample were computed for the shared 83 dbSNPs. Overall, the greatest difference in allele frequencies was found between Mexican Americans and Africans with a highest mean F_{ST} of 0.126, compared with mean F_{ST} of 0.035 in MA vs CEU, 0.033 in MA vs CHB and 0.032 in MA vs JPT (Figure 1d). For the gene-specific mean F_{ST} in MA vs YRI, larger mean F_{ST} values were observed for *SLC6A3* (0.208) and *SLC6A4* (0.198), but much lower for *SLC6A2* (0.04) (Figure 1e).

SNP-based genetic association analyses of cases and controls

Single nucleotide polymorphism-based allelic and genotypic association analyses revealed that 16 polymorphisms were associated with MDD with a nominal P < 0.05 in five genes (Table 2), including two common 3' UTR polymorphisms in NTRK2 (rs7020204 and rs2013566) and one rare 5' UTR polymorphism in SLC6A4 (rs28914831). Among the nine SNPs with a nominal P < 0.05 in both allelic and genotypic tests, seven were uncommon polymorphisms with a MAF <0.03 in controls, including (rs3732076), two in one in CREB1 ABCB1 (rs4728697, rs58898486) and four in SLC6A4 (rs7212502, rs28914831, NT 010799.14 3288789 and rs56355214) (Table2 and Supplementary Table 1). Three SLC6A4 common polymorphisms (rs7224199 and rs3813034 in upstream and rs140701) showed genotypic association, but with a small allelic odds ratio <1.3 and allelic test nominal P>0.05. No associated SNPs remained significant after adjusting for multiple tests with an FDR_BH ≤ 0.05 .

SNP-based genetic association analysis of antidepressant response

In this study, there were 142 MDD patients who enrolled in the pharmacogenetic trial and completed 8-week antidepressant treatment (68 treated with desipramine and 74 treated with fluoxetine). For the discrete outcome (remission vs non-remission), SNP-based allelic or genotypic association analyses revealed that clinical remission status was associated with several polymorphisms in or near three genes, *ABCB1*, *NTRK2* and *SLC6A2* (Table 3). All of the nine associated *NTRK2* SNPs were in 3' UTR or coding regions except for rs2289658 at a splice site, whereas the two associated *SLC6A2* SNPs were in intron or upstream region. For the *ABCB1* gene, the associated SNPs included two in UTR, two in introns and one in coding sequence. No associated SNPs remained significant after adjusting for multiple tests with an FDR BH ≤ 0.05 in the discrete outcome analysis.

For the quantitative outcome (relative reduction in HAM-D21 score) after controlling for age, gender and baseline HAM-D21 score, general linear regression analyses revealed that relative reduction of HAM-D21 scores was associated with six *NTRK2* SNPs (three in 3' UTR, two synonymous and one intronic at splice site) and one SLC6A3 intronic SNP rs8179029 in desipramine-treated patients, two SLC6A2 upstream SNPs in fluoxetine-treated patients and one SLC6A3 intronic SNP rs8179029 for combined sample, with a nominal P < 0.01 (Table 4). Among the associated SNPs, only two NTRK2 synonymous SNPs, rs2289657 and rs56142442, remained statistically significant after correcting for multiple testing with an FDR_BH = 0.05 in the sample of patients treated with desipramine. Desipramine-treated patients who are homozygous for C allele at synonymous SNP rs2289657 or at rs56142442 had higher levels of improvement with 27% larger reduction in HAM-D21 scores, compared with those who are not homozygous for C allele at rs2289657 or rs56142442.

Haplotype-based analyses

Haplotype analysis identified a total of 17 haplotype blocks in the seven genes using the Four Gametes Rules with the Haploview program, including one block in CREB1, two blocks in each of SLC6A3, SLC6A4 and CRHR1, three blocks in each of ABCB1 and SLC6A2, and four blocks in NTRK2 (Figure 2). For the association analysis of case and control, the diagnosis of depression was found to be associated with five haplotypes with a nominal *P*-value between 0.01 and 0.05 in CREB1, SLC6A3, ABCB1, NTRK2 and SLC6A2. Among the five depression-associated haplotypes, four included at least one SNP showing an association with depression in the single SNP-based analysis (Table 5). For the association of remitter and non-remitter, eight haplotypes were found to be associated with remission status, including two in ABCB1 (ACA in block 1 for desipramine-treated patients and GCGCACACGAGAC in block 2 for fluoxetine-treated patients), two in NTRK2 (TCG and CAG in block 3 for desipramine-treated patients), one in SLC6A2 (GCCAGT in block 4 for desipraminetreated patients) and three in SLC6A4 (TAGC and TAGA in block 1 and ATTGTAACCC in block 2 for the combined sample of designamine- or fluoxetinetreated patients). Among the eight remission-associated haplotypes, three showed an association with a

Table 2	Polymorphisms associated	with depression	in Mexican-An	nericans						
						Risk allele	frequency			Р
Gene	SNP	Chromosome	Position	SNP type	Risk/non- risk allele	Case	Control	OR^{a} (95% CI)	Allelic	Genotypic
CREB1	rs3730276	2	208140591	INI	A/G	0.998	0.977	11.45(1.46-89.77)	0.004	0.003
SLC6A3	rs8179029 rs2550936	വ വ	1462985 1464256	INT INT	C/T A/C	$0.910 \\ 0.839$	0.856 0.799	$1.71 (1.12-2.58) \\1.31 (0.94-1.83)$	$0.02 \\ 0.13$	0.02 0.004
ABCB1	rs4728697 rs2032583 rs58898486		86986874 86998497 87063142	INI TNI TNI	A/G A/G G/T	0.060 0.936 0.998	0.028 0.899 0.984	$\begin{array}{c} 2.24 \ (1.18 - 4.28) \\ 1.64 \ (1.04 - 2.59) \\ 8.40 \ (1.05 - 67.39) \end{array}$	$\begin{array}{c} 0.01 \\ 0.04 \\ 0.02 \end{array}$	0.02 0.10 0.02
NTRK2	rs7020204 rs2013566	5 5	86616007 86616118	INT, 3' UTR INT, 3' UTR	C/T A/G	0.899 0.876	0.850 0.828	$1.57 (1.07 - 2.32) \\ 1.47 (1.02 - 2.10)$	$0.02 \\ 0.04$	0.05 0.13
SLC6A4	rs7224199 rs3813034 rs140701 rs7212502 rs28914831 rs2065713 rs2065713 rs2655214 rs56355214	17 177 117 117 117 117	25547852 25548930 25562658 25573388 25573388 2557791 2557791 25576325 25576325	Downstream Downstream INT 5/ UTR INT INT INT INT	G/T A/C A/G G/T G/A G/A	$\begin{array}{c} 0.411\\ 0.454\\ 0.454\\ 1.000\\ 1.000\\ 0.329\\ 1.000\\ 0.329\\ 1.000\\ 1.000\end{array}$	$\begin{array}{c} 0.369\\ 0.399\\ 0.400\\ 0.987\\ 0.988\\ 0.988\\ 0.263\\ 0.988\\ 0.988\\ 0.988\end{array}$	$\begin{array}{c} 1.19 \ (0.92-1.55) \\ 1.25 \ (0.98-1.60) \\ 1.25 \ (0.97-1.61) \\ 8.56 \ (1.23-\infty) \\ 8.37 \ (1.20-\infty) \\ 1.37 \ (1.04-1.80) \\ 8.63 \ (1.24-\infty) \\ 8.47 \ (1.22-\infty) \\ \end{array}$	$\begin{array}{c} 0.19\\ 0.08\\ 0.09\\ 0.01\\ 0.02\\ 0.03\\ 0.01\\ 0.01 \end{array}$	$\begin{array}{c} 0.004\\ 0.040\\ 0.01\\ 0.01\\ 0.01\\ 0.01\\ 0.004\\ 0.01\\ 0.01\end{array}$
Abbrević <i>NTRK2</i> , ^a OR: calo	ations: <i>ABCB1</i> , ATP-binding neurotrophic tyrosine kinas. culation based on allele cou	cassette subfam e type 2 receptor nt and 95% CI b	ily B member r; OR, odds rat ased on exact	1; CI, confidenc io; SNP, single logistic model v	e interval; <i>CR</i> nucleotide po when one cell	<i>EB1</i> , cyclic A lymorphism; count is 0.	MP-responsiv UTR, untran	ve element binding prc slated region.	otein 1; IN	T, Intronic;

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							Better all	ele freque	ncy	Fisher'	s exact P
Medication	Gene	SNP	Chromo- some	Position	SNP type	Better/poor response allei	Remitter le	Non- remitter	OR^{a} (95% CI)	Allelic	Genotypic
Desipramine or fluoxetine (N=142)	ABCB1	rs3842	~	86971302	3' UTR	C/T	0.203	0.094	2.44 (1.14–5.24)	0.02	0.11
	NTRK2		∿ o o o o o o	86971406 86618439 86618627 86618628 86618628 86619110 86677208 86677208	3' UTR INT, splice site	A/T A/C A/C A/C A/C C/C T/C	$\begin{array}{c} 0.100\\ 0.171\\ 0.177\\ 0.181\\ 0.321\\ 0.321\\ 0.961\\ 0.885\end{array}$	0.028 0.123 0.123 0.123 0.224 0.880 0.778	$\begin{array}{c} 3.81 \ (1.08-13.5) \\ 1.47 \ (0.74-2.94) \\ 1.53 \ (0.77-3.05) \\ 1.58 \ (0.77-3.15) \\ 1.58 \ (0.95-2.83) \\ 3.35 \ (1.14-9.86) \\ 3.35 \ (1.12-4.28) \end{array}$	$\begin{array}{c} 0.03\\ 0.31\\ 0.24\\ 0.24\\ 0.08\\ 0.02\\ 0.03\end{array}$	$\begin{array}{c} 0.11\\ 0.002\\ 0.002\\ 0.001\\ 0.021\\ 0.05\\ 0.05\end{array}$
Desipramine	ABCB1	rs17064	2	86971406	3' UTR	A/T	0.132	0.018	8.39 (1.03–68.4)	0.02	0.14
(xa = vi)	NTRK2	$\begin{array}{l} \mathrm{NT}_023935.17_16593339\\ \mathrm{NT}_023935.17_16593340\\ \mathrm{rs}11140793\\ \mathrm{rs}211440793\\ \mathrm{rs}2289658\\ \mathrm{rs}2289657\\ \mathrm{rs}56142442\\ \mathrm{rs}56142442\\ \mathrm{rs}56142442\end{array}$		86618627 86618628 8661300 86753190 86753280 86826085	INT, 3' UTR INT, 3' UTR INT, 3' UTR INT, 3' UTR INT, splice site SYN BYN	1/C 6/A 7/C C/A	0.157 0.167 0.882 0.909 0.955 0.957	0.113 0.113 0.726 0.742 0.823 0.902	$\begin{array}{c} 1.47 & (0.53-4.05) \\ 1.57 & (0.57-4.35) \\ 2.83 & (1.12-7.14) \\ 3.48 & (1.26-9.59) \\ 4.53 & (1.20-17.1) \\ 4.53 & (1.20-17.1) \\ 2.31 & (0.55-9.65) \\ 0.05 & (1.00) \\ 0.05$	0.61 0.45 0.03 0.02 0.31 0.31	0.03 0.05 0.04 0.04 0.05 0.05
Fluoxetine	ABCB1	rs1128503	07 1	87017537	SYN	G/A	0.522	0.304	2.49 (1.18–5.30)	0.02	0.05
(JN = /4)	NTRK2 SLC6A2	$\begin{array}{c} {\rm rs10276036} \\ {\rm rs2235020} \\ {\rm rs1624327} \\ {\rm NT_023935.17_16651920} \\ {\rm rs1362621} \end{array}$	16 9 16	87018134 87037201 86619110 86677208 54245985	INT INT INT, 3' UTR INT, 3' UTR Upstream	T/C T/A A/G G/C	0.550 0.539 0.348 0.972 0.872	0.320 0.318 0.212 0.864 0.731	$\begin{array}{c} 2.59 & (1.24{-}5.44) \\ 2.50 & (1.15{-}5.43) \\ 1.99 & (0.90{-}4.39) \\ 5.52 & (1.06{-}28.7) \\ 2.52 & (1.06{-}5.96) \\ \end{array}$	$\begin{array}{c} 0.01\\ 0.02\\ 0.09\\ 0.05\\ 0.04\end{array}$	0.04 0.09 0.04 0.03 0.03
Abbreviation: OR, odds rati ^a OR: calculati	s: ABCB1, ¹ o; SNP, sin _§ ion based o	ATP-binding cassette subfarr şle nucleotide polymorphisn n allele count and 95% CI b	iily B mem n; UTR, un ased on exe	ber 1; CI, co translated re act logistic n	onfidence interv gion; SYN, sync aodel when one	al; INT, intron mymous. cell count is (iic; <i>NTRK2</i> ,).	neurotrol	phic tyrosine kiné	ase type	2 I

Table 4 Poly	morphism	s associated with relative re	duction of	HAM-D21 s	core after 8-wee	ek antidepres	sant tr	eatment with d	esipramine or fluoxeti	ne	
Medication	Gene	SNP	Chromo- some	Position	SNP type	Genotype	N	Mean (s.d.)	$Delta^a$ (95% Cl)	P^b	FDR_BH^{c}
Desipramine or fluoxetine	ABCB1	rs2214103	7	87062884	INT	CC	132	64.26 (21.35)	21.45(5.29 - 37.61)	0.01	0.72
	NTRK2	IS9969765	6	86679605	INT, 3' UTR		46 146	$46.44 (32.92) \\ 69.48 (19.13) \\ 60.60 \\ 60 \\ 60 \\ 60 \\ 60 \\ 60 \\ 60 \\$	10.96 (3.11–18.81)	0.007	0.72
		rs2289657	6	86753280	SYN		8/ 106	58.79 (23.81) 65.38 (20.35)	12.74 $(3.32-22.15)$	0.009	0.72
	SLC6A2	${ m NT}_010498.15_9300464$	16	54243766	Upstream	TG TT	$\frac{20}{121}$	22.48 (29.11) 77.25 (21.72) 60.69 (22.06)	18.49 $(6.60 - 30.39)$	0.003	0.72
Desipramine	SLC6A3	rs8179029	Ŋ	1462985	INT	CC TT/TC	$\frac{54}{6}$	61.32 (20.05) 27.05 (39.26)	29.79 (9.04–50.54)	0.007	0.56
	NTRK2	rs7038236	6	86678222	INT, 3' UTR	CC A A / A C	37 1 0	63.52 (18.98)	16.97 (4.73 - 29.22)	0.009	0.56
		rs11140793	6	86681300	INT, 3' UTR	AA	43	40.02 (24.77) 62.71 (23.47) 46.20 (22.20)	18.30(5.74 - 30.86)	0.006	0.56
		rs2289658	6	86753190	INT,	TT	44	40.39 (23.30) 62.94 (20.19)	18.00(5.32 - 30.69)	0.007	0.56
		rs2289657	6	86753280	splice site	CC/TC CC	20 51	$\begin{array}{c} 43.57 \\ 62.11 \\ (19.93) \end{array}$	26.83 (13.39–40.27)	0.0002	0.05
		rs56142442	6	86826085	NYS	AA/AC CC TT/TC	13 59 8	$36.40 (29.89) \\ 60.70 (19.89) \\ 28.92 (34.90)$	33.17 (17.15–49.19)	0.0001	0.05
Fluoxetine	SLC6A2	$\mathrm{NT}_010498.15_9300464$	16	54243766	Upstream	ТG	10	85.09 (13.13) 65.60 (18.37)	20.55 (8.23–32.87)	0.002	0.46
		rs4783899	16	54244713	Upstream	TT GG/TG	24 48	76.99 (15.15) 64.22 (20.09)	15.02 (5.37–24.68)	0.003	0.47
Abbreviations <i>NTRK2</i> , neur ^a Delta: adjustu ^b <i>P</i> : based on { ^c Benjamini-H	:: <i>ABCB1</i> , otrophic ty ed mean d general lin ochberg fa	ATP-binding cassette subfi- rosine kinase type 2 recept ifference in relative reduct ear model after controlling lse discovery rate after corr	amily B me or; SNP, si ion of HAN for age, ge ecting for	ember 1; CI, ngle nucleot A-D21 score. nder and ba multiple test	confidence in ide polymorph seline HAM-D2 ing.	terval; HAM lism; UTR, u 21 score.	-D21, ntrans	21-item Hamil slated region; S	ton Depression Ratin, YN, synonymous.	g Scale;]	NT, intronic;

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Figure 2 Linkage disequilibrium (LD) pattern in seven genes: cyclic AMP-responsive element binding protein 1 (*CREB1*) (a), *SLC6A3* (b), ATP-binding cassette subfamily B member 1 (*ABCB1*) (c), neurotrophic tyrosine kinase type 2 receptor (*NTRK2*) (d), *SLC6A2* (e), *SLC6A4* (f) and corticotropin-releasing hormone receptor 1 (*CRHR1*) (g). Standard color scheme in Haploview program is used to display the level of logarithm of odds (LODs) and the D'. Shown in each box are estimated statistics of the D', which indicates the LD relationship between each pair of single nucleotide polymorphisms (SNPs) and are not labeled if D' = 1.00. Regions are shown in bright red, light blue, shades of pink/red and white for $D' = 1 + \text{LOD} \ge 2$, $D' = 1 + \text{LOD} \ge 2$, $D' < 1 + \text{LOD} \ge 2$ and D' < 1 + LOD < 2, respectively. Vertical lines on the long horizontal white indicate the relative positions of SNPs in the gene.

Sample	Gene	Block no.	<i>Haplotype</i> ^a	Fre	equency	χ^2	d.f.	Р
				Case	Control			
MDD patients and healthy controls	CREB1	Block 1	<u>G</u> CACGG	0.003	0.018	5.71	1	0.02
liourup controlo	SLC6A3	Block 1	CGTGCGGT	0.089	0.134	4.83	1	0.03
	ABCB1	Block 2	GCACACACGAGAC	0.060	0.028	6.12	1	0.01
	NTRK2	Block 1	GTTAGGGCA	0.094	0.133	3.96	1	0.05
	SLC6A2	Block 3	ACCAGA	0.004	0.017	3.98	1	0.05
				Remitter	Non-remitter			
MDD patients treated with desipramine or fluoxetine	ABCB1	Block 1	<u>A</u> CA	0.101	0.029	4.72	1	0.03
1	NTRK2	Block 3	TCG	0.839	0.717	5.63	1	0.02
	SLC6A4	Block 1	TAGC	0.006	0.047	5.11	1	0.02
		Block 1	TAGA	0.048	0.003	4.92	1	0.03
		Block 2	ATTGTAACC	0.028	0.081	4.00	1	0.05
MDD patients treated with desipramine	ABCB1	Block 1	<u>A</u> CA	0.132	0.018	5.31	1	0.02
I	NTRK2	Block 3	TCG	0.865	0.679	6.35	1	0.01
		Block 3	CAG	0.045	0.177	5.74	1	0.02
	SLC6A2	Block 3	GCCAGT	0.134	0.012	6.79	1	0.009
MDD patients treated with fluoxetine	ABCB1	Block 2	GCGCACACGAG <u>A</u> C	0.503	0.682	4.08	1	0.04
	SLC6A4	Block 1	TAGC	0.000	0.085	8.37	1	0.004

Table 5 Haplotypes associated with depression or clinical remission after 8-week antidepressant treatment

Abbreviations: *ABCB1*, ATP-binding cassette subfamily B member 1; *CREB1*, cyclic AMP-responsive element binding protein 1; MDD, major depressive disorder; *NTRK2*, neurotrophic tyrosine kinase type 2 receptor; SNP, single nucleotide polymorphism.

^aLetters with underline indicate the SNPs also showing an association with a nominal P < 0.05 in the corresponding SNP-based analysis.

nominal $P \leq 0.01$: TCG in block 3 of *NTRK2* and GCCAGT in block 4 of *SLC6A2* (P = 0.009) for desipramine-treated patients, and TAGC in block 1 of *SLC6A4* for fluoxetine-treated patients (P = 0.004) (Table 5).

For quantitative outcome analysis of antidepressant treatment, 15 haplotypes were found to be associated with the relative reduction in HAM-D21 score after controlling for age, gender and baseline HAM-D21 score (Table 6). Among the 15 associated haplotypes, 2 in *SLC6A3* and 3 in *NTRK2* showed a correlation with a nominal P < 0.004 in desipramine-treated patients, and 2 in *SLC6A2* showed an association with a nominal P < 0.008 in fluoxetine-treated patients. The most significant association was found between *NTRK2* haplotype CAG (rs2289658, rs2289657 and rs2289656) and relative reduction of HAM-D21 score with a nominal P = 0.0002 and an effect size of squared R = 0.20 (Table 6).

Discussion

In this study, we analyzed the fine structure of seven genes that are relevant to the pathophysiology of MDD or to antidepressant response at four sequential levels: (1) entry into the brain, (2) binding to monoaminergic transporters, and (3) distal effects at the transcription level, resulting in (4) changes in neurotrophin and neuropeptide receptors. We observed new alleles in all seven genes in Mexican-Americans. We described a total of 204 novel SNPs (Table 1), which almost doubled the number of reported SNPs in these genes that was detected in these individuals (total of dbSNPs was 215). The number of novel SNPs identified in these Mexican-American subjects ranged from 12 to 57, and in the case of *CREB1*, the total number of SNPs tripled from 6 to 18. Most of the novel SNPs reported here had MAF lower than 5% (Supplementary Table 1). Higher nucleotide diversity was found in the exonic regions of these genes, particularly in UTRs (Figure 2b). Only a small number of the novel SNPs were in coding regions¹⁹ and of those <40% (7) were non-synonymous. Analyses of HapMap data on four ethnic groups found different allele frequencies, with the greatest differences between Mexican Americans and Africans (Figure 1d).

Our analyses revealed nominal associations of eight SNPs and four haplotypes with susceptibility for MDD; those SNPs and haplotypes were located in four genes, *ABCB1*, *CREB1*, *NTRK2* and *SLC6A3*. In addition, eight SNPs in *SLC6A4* and one haplotype

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Medication	Gene	Block no.	<i>Haplotype</i> ^a	Ν	β^{b}	$R^{2 \ c}$	t	Р
Desipramine or fluoxetine	SLC6A3	Block 1	CGCGAAGT	133	40.95	0.07	3.14	0.002
		Block 1	CGTGCGGT	133	17.01	0.05	2.69	0.008
	ABCB1	Block 3	TCTTACCGATCG	141	27.17	0.05	2.58	0.01
	NTRK2	Block 2	GCACGCCATTGGCAC	140	-5.85	0.03	-2.20	0.03
		Block 3	CAG	132	14.58	0.07	3.16	0.002
		Block 3	TCG	132	-7.49	0.04	-2.35	0.02
		Block 4	CA	134	-11.22	0.04	-2.36	0.02
	SLC6A2	Block 1	GATGAT	140	-15.88	0.04	-2.46	0.01
		Block 1	TAGGGT	140	10.00	0.03	2.10	0.04
Desipramine	SLC6A3	Block 1	CGTGCGGT	65	36.51	0.15	3.29	0.002
-		Block 1	CGCGAGGT	65	-15.77	0.12	-2.99	0.004
	NTRK2	Block 2	G <u>AACCCGC</u> TCGGTAC	66	13.69	0.09	2.47	0.02
		Block 2	G <u>C</u> ACGCC <u>A</u> TTGGCAC	66	-9.68	0.08	-2.30	0.02
		Block 3	CAG	64	24.02	0.20	3.90	0.0002
		Block 3	TCG	64	-11.55	0.09	-2.49	0.02
		Block 4	CA	65	-20.49	0.15	-3.29	0.002
		Block 4	TA	65	22.05	0.13	3.02	0.004
	SLC6A4	Block 1	GĀGA	67	10.65	0.08	2.37	0.02
Fluoxetine	NTRK2	Block 2	CCACCCCACCATCTC	72	14.21	0.06	2.16	0.03
	SLC6A2	Block 1	GATGAT	73	-18.92	0.10	-2.88	0.005
		Block 1	TAGGGT	73	14.54	0.10	2.73	0.008
		Block 2	GT	67	-13.66	0.06	-2.03	0.05
	SLC6A4	Block 1	GAAA	74	28.05	0.06	2.08	0.04

 Table 6
 Haplotypes associated with relative reduction of HAM-D21 score after 8-week antidepressant treatment

Abbreviation: *ABCB1*, ATP-binding cassette subfamily B member 1; HAM-D21, 21-item Hamilton Depression Rating Scale; *NTRK2*, neurotrophic tyrosine kinase type 2 receptor; SNP, single nucleotide polymorphism.

^aLetters with underline indicate the SNPs also showing an association with a nominal P < 0.05 in the corresponding SNP-based analysis.

^bβ: regression coefficient after controlling for age, gender and baseline HAM-D21 score.

^c*R*²: proportion variance explained.

in *SLC6A2* were also associated with MDD (Tables 2 and 5). However, some of these SNPs were not very common (MAF < 0.03 in controls).

Nominal associations with several polymorphisms were also found for treatment response of 142 MDD patients who completed 8-week antidepressant treatment with desipramine or fluoxetine. Discrete outcome analyses (remitters vs non-remitters) showed that SNPs and haplotypes in *ABCB1* and *NTRK2* were associated with response. Variation in *SLC6A2* and one haplotype in *SLC6A4* were also associated with remission status. Quantitative outcome analyses showed that SNPs and haplotypes in *ABCB1*, *NTRK2* and *SLC6A2* were associated with relative HAM-D21 score reduction, but only two SNPs and one haplotype in *NTRK2* remained significant for desipramine treatment after correcting for multiple testing.

Our data show that variations in six out of seven genes were associated with MDD or antidepressant response. Briefly, (i) SNPs in *ABCB1* (located at 7q21.1), which is also called multidrug resistance 1, were associated with MDD and antidepressant response. *ABCB1* encodes a large transmembrane transporter protein that acts as an active efflux pump

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transporting a wide range of drugs from the brain to the blood. Polymorphisms in this gene have been reported to predict the response to antidepressant treatment to drugs that are substrates for this transporter.²¹ (ii) SNPs in the *CREB1* gene were associated with MDD. CREB (cyclic AMP response element-binding protein, located at 2q32.2-q34) encodes a transcription factor that modulates key growth factors important for synaptogenesis and neurogenesis. Sequence variations in the promoter and intronic regions of the CREB1 gene have previously been described to be cosegregated with mood disorders in women.²² 3) SNPs in NTRK2 (located at 9q22.1) were associated with susceptibility to MDD and antidepressant response. Furthermore, two SNPs and one haplotype in *NTRK2* continued to be significantly associated with relative reduction of HAM-D21 scores in the desipramine-treated group, after controlling for age, gender and baseline HAM-D21 scores. NTRK2, also known as tyrosine kinase receptor B, and its ligand, brain-derived neurotropic factor, regulate short- and long-term synaptic functions and neural plasticity. NTRK2 variants have been recently associated with obsessive-compulsive

disorder in female patients.²³ (iv) SNPs in SLC6A2 (noradrenaline transporter, located at 16q12.2) were associated with remission status and relative reduction of HAM-D21 scores, and one haplotype in this gene (ACCAGA) was associated with MDD. SLC6A2 gene encodes a transporter, which regulates norepinephrine (noradrenaline) homeostasis and the of norepinephrine into reuptake presynaptic nerve terminals.²⁴ SLC6A2 polymorphisms have been reported to be associated with depression^{25,26} and response to antidepressants.^{27,28} (v) SNPs or haplotypes in SLC6A3 (dopamine transporter or DAT1, located at 5p15.33), which encodes a transporter that is important in dopaminergic neurotransmission, were associated with risk for MDD or relative reduction of HAM-D21. This transporter mediates the active re-uptake of synaptic dopamine.²⁹ Variations in this gene have already been implicated in susceptibility for mood disorders^{30,31} and antidepressant action.³² Other neuropsychiatric conditions have also been associated with SLC6A3, such as parkinsonism,³³ attention-deficit hyperactivity disorder.^{34,35,36} Tourette's syndrome and addictive behavior.^{37,38} 6) SLC6A4 (serotonin transporter, located at 17q11.1-q12) encodes a transporter, which mediates antidepressant action, and behavioral effects of cocaine and amphetamines. Sequence variations in SLC6A4 have been extensively queried and they may be associated with several neuropsychiatric conditions, including MDD,^{39,40,41} anxietyrelated personality traits⁴² and antidepressant response.43,44 Our findings support that variations in SLC6A4 are associated with MDD risk. Haplotypes in SCL6A4 have also been associated with remission status and reduction of HAM-D21 scores.

The analyses presented here have not shown that variations in *CRHR1* (located at 17q12-q22) gene are associated with susceptibility to MDD or antidepressant response. It can be noted that the current analyses have not taken anxiety levels into consideration. *CRHR1* encodes the receptor of CRH, a key stress hormone that regulates the response to stress at the behavioral, immune, autonomic and neuroendocrine levels, through the activation of the hypothalamic-pituitary-adrenalaxis. Polymorphisms in *CRHR1* were reported to be associated with antidepressant response, but only when anxiety scores are taken in consideration,^{45,46} and with seasonal pattern and early onset of first depressive episode.⁴⁷

In summary, we show that substantial levels of sequence variation, especially those that are not very common (MAF >5%), are likely to be found in candidate genes in an ethnically defined and understudied group. In this population group, for example, half of the SNPs detected were novel. Therefore, deep sequencing data may be relevant to our understanding of common and complex disorders, such as major depression, particularly in minority populations. Our analyses showed that several sequence variations and haplotypes in six out of seven selected genes were nominally associated with MDD risk and/or

antidepressant treatment response and that after controlling for age, gender and baseline HAM-D21 score, as well as correcting for multiple testing, there was a significant association of antidepressant response with two *NTRK2*-coding SNPs and one haplotype. Our findings suggest that these variants may be implicated in the pathophysiology of MDD. The Mexican Americans are the most rapidly growing population group in the United States, but remain under represented in research studies. These results highlight the importance of direct re-sequencing of key candidate genes in ethnic minority groups in order to discover novel genetic variants that cannot be simply inferred from existing databases.

Conflict of interest

The authors declare no conflict of interest.

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