



Draft Genome Sequence of *Pseudomonas* sp. Strains MWU12-2319 and MWU12-2311, Isolated from a Wild Cranberry Bog in the Cape Cod National Seashore

Arin Pittman,^a Scott Soby^{b,c}

^aArizona College of Osteopathic Medicine, Midwestern University, Glendale, Arizona, USA ^bBiomedical Sciences, College of Graduate Studies, Midwestern University, Glendale, Arizona, USA ^cCollege of Veterinary Medicine, Midwestern University, Glendale, Arizona, USA

ABSTRACT *Pseudomonas* sp. strains MWU12-2319 and MWU12-2311 were isolated from the soil of a wild cranberry bog in the Cape Cod National Seashore as part of a culture-dependent bacterial population survey. The genomes exceed 7 Mbp and contain putative gene clusters for the biosurfactant orfamides A and C.

he genus Pseudomonas contains highly diverse species that are adapted to many different environments, but little is known about their evolution, rate of speciation, or how these bacteria influence the microbiomes with which they interact (1-3). As part of a larger microbiome analysis project, we sampled bacteria from wild cranberry bogs to analyze their genomes for taxonomic placement and to provide further insights into their potential ecological functions. Pseudomonas sp. strains MWU12-2319 and MWU12-2311 were isolated from sandy bog soil in the Cape Cod National Seashore (42.064742N, 70.117562W). Bacteria were isolated on King's medium B (KMB) agar containing 50 μ g mL⁻¹ each of cycloheximide and ampicillin, incubated at 26°C for 24 to 48 h, colony purified three times on KMB agar, and stored at -80° C in 34% glycerol. All kits described below were used according to the manufacturer's instructions. Isolates from frozen storage were recovered on KMB agar, and populations were inoculated into overnight KMB broth cultures for genomic DNA isolation with a DNeasy blood and tissue kit (Qiagen, USA). Illumina-compatible genomic DNA libraries were generated using a HyperPlus library preparation kit (Kapa Biosystems product number KK8514; Roche, USA). DNA was enzymatically sheared to \sim 500 bp, end repaired, and A-tailed. Illumina-compatible adapters with unique indexes (product number 00989130v2; Integrated DNA Technologies, Coralville, IA) were ligated to each sample. Adapter-ligated molecules were cleaned using KAPA pure beads (Kapa Biosystems product number KK8002) and amplified with KAPA HiFi enzyme (Kapa Biosystems product number KK2502). Library fragment sizes were determined on an Agilent TapeStation system, and fragments were quantified by quantitative PCR (KAPA library quantification kit [Kapa Biosystems product number (KK4835]) on a QuantStudio 5 system (Thermo Fisher Scientific, USA) before multiplex pooling and sequencing in a 2 \times 250-bp flow cell on the Illumina MiSeq platform. All software was used with default settings except as indicated. Raw reads were assembled with Unicycler v0.4.8 (4) and polished with Pilon v1.23 (5) within the PATRIC Comprehensive Genome Analysis pipeline v3.6.12 using default settings except for the trim setting, which was set to true (6). The Comprehensive Genome Analysis pipeline includes quality control and trimming by QUAST v5.0.2 (7) and Trim Galore v0.4.0 (8) and annotation by RASTtk v1.073 (9), supplemented with antiSMASH (https://antismash.secondarymetabolites.org/#!/start) for recognition of secondary metabolite gene clusters (10). Using the Type (Strain) Genome Server (TYGS) (11), isolates were placed with high confidence within the genus Pseudomonas. Digital DNA-DNA hybridization (dDDH) (formula d4) with Pseudomonas batumici UCM B-321^T (GenBank accession number JXDG0000000), the closest relative within the genus, yielded

Editor David A. Baltrus, University of Arizona Copyright © 2022 Pittman and Soby. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Scott Soby, ssobyx@midwestern.edu. The authors declare no conflict of interest. Received 19 August 2022 Accepted 27 September 2022 Published 12 October 2022

TABLE 1 Data summary

	BioSample accession	GenBank accession	SRA accession	Genome	No. of	N ₅₀	G+C	Total read	No. of	Coverage
Isolate	no.	no.	no.	size (bp)	contigs	(bp)	content (%)	length (bp)	reads	(X)
MWU12-2319	SAMN26896722	JALJES00000000	SRR18508975	7,276,095	83	299,807	61.24	716,456,692	3,085,670	98
MWU12-2311	SAMN27783457	JAMQRU00000000	SRR19044811	7,276,098	85	284,877	61.24	975,020,131	4,169,776	134

values of 42.1% for both isolates, indicating that MWU12-2319 and MWU12-2311 are not closely related to any named *Pseudomonas* species. The isolates contain putative gene clusters for synthesis of orfamide A and C, biosurfactant cyclic lipopeptides (CLPs) that have activity against oomycete zoospores and filamentous fungi and insecticidal activity against aphids, as found previously in the *Pseudomonas protegens* group (12, 13).

Data availability. The whole-genome shotgun project has been deposited in DDBJ/ EMBL/GenBank under BioProject accession number PRJNA691338, with the genome and SRA accession numbers presented in Table 1. The versions described in this paper are JALJES000000000.1 (MWU12-2319) and JAMQRU000000000.1 (MWU12-2311). RASTtk annotations are available under open license at Zenodo (https://doi.org/10.5281/zenodo.6413223 [MWU12-2319] and https://doi.org/10.5281/zenodo.6629972 [MWU12-2311]).

ACKNOWLEDGMENTS

This research was supported by the Office of Research and Sponsored Programs, College of Graduate Studies, and the Biomedical Sciences Program, Midwestern University.

We appreciatively acknowledge Alisha Harrison for isolation of genomic DNA. Library construction and Illumina sequencing were performed at the Arizona State University Genomics Core Facility.

This work meets a course requirement for Special Topics in Bacterial Genomics for A.P.

REFERENCES

- Passarelli-Araujo H, Franco GR, Venancio TM. 2022. Network analysis of ten thousand genomes shed light on *Pseudomonas* diversity and classification. Microbiol Res 254:126919. https://doi.org/10.1016/j.micres.2021.126919.
- Lopes LD, Davis EW, Pereira e Silva MC, Weisberg AJ, Bresciani L, Chang JH, Loper JE, Andreote FD. 2018. Tropical soils are a reservoir for fluorescent *Pseudomonas* spp. biodiversity. Environ Microbiol 20:62–74. https://doi.org/10.1111/ 1462-2920.13957.
- Hesse C, Schulz F, Bull CT, Shaffer BT, Yan Q, Shapiro N, Hassan KA, Varghese N, Elbourne LDH, Paulsen IT, Kyrpides N, Woyke T, Loper JE. 2018. Genome-based evolutionary history of *Pseudomonas* spp. Environ Microbiol 20:2142–2159. https://doi.org/10.1111/1462-2920.14130.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal.pone.0112963.
- Wattam AR, Davis JJ, Assaf R, Boisvert S, Brettin T, Bun C, Conrad N, Dietrich EM, Disz T, Gabbard JL, Gerdes S, Henry CS, Kenyon RW, Machi D, Mao C, Nordberg EK, Olsen GJ, Murphy-Olson DE, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Vonstein V, Warren A, Xia F, Yoo H, Stevens RL. 2017. Improvements to PATRIC, the all-bacterial Bioinformatics Database and Analysis Resource Center. Nucleic Acids Res 45:D535–D542. https://doi.org/10.1093/nar/qkw1017.
- 7. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment

tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10 .1093/bioinformatics/btt086.

- Krueger F. 2014. Trim Galore: a wrapper tool around Cutadapt and FastQC to consistently apply quality and adapter trimming to FastQ files, with some extra functionality for Mspl-digested RRBS-type (Reduced Representation Bisufite-Seq) libraries. https://www.bioinformatics.babraham.ac.uk/projects/trim_galore.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https://doi.org/10.1038/srep08365.
- Blin K, Shaw S, Kloosterman AM, Charlop-Powers Z, van Wezel GP, Medema MH, Weber T. 2021. antiSMASH 6.0: improving cluster detection and comparison capabilities. Nucleic Acids Res 49:W29–W35. https://doi.org/10.1093/nar/gkab335.
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. Nat Commun 10: 2182. https://doi.org/10.1038/s41467-019-10210-3.
- Ma Z, Geudens N, Kieu NP, Sinnaeve D, Ongena M, Martins JC, Höfte M. 2016. Biosynthesis, chemical structure, and structure-activity relationship of orfamide lipopeptides produced by *Pseudomonas protegens* and related species. Front Microbiol 7:382. https://doi.org/10.3389/fmicb.2016.00382.
- Gross H, Stockwell VO, Henkels MD, Nowak-Thompson B, Loper JE, Gerwick WH. 2007. The genomisotopic approach: a systematic method to isolate products of orphan biosynthetic gene clusters. Chem Biol 14:53–63. https://doi.org/10.1016/ j.chembiol.2006.11.007.