

Long-Term Depression in the Hippocampal CA1 Area of Aged Rats, Revisited: Contribution of Temporal Constraints Related to Slice Preparation

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Abstract

Background: The effects of low-frequency conditioning stimulation (LFS, 900 pulses at 1 Hz) of glutamatergic afferents in CA1 hippocampal area using slices from two different strains of adult (3–5 month-old) and aged (23–27 month-old) rats were reinvestigated regarding the discrepancies in the literature concerning the expression of long-term depression (LTD) in the aging brain.

Methodology/Principal Findings: N-methyl-D-aspartate receptor (NMDA-R) dependent LTD was examined in both adult ($n = 21$) and aged ($n = 22$) Sprague-Dawley rats. While equivalent amounts of LTD could be obtained in both ages, there was significant variability depending upon the time between the slices were made and when they were tested. LTD was not apparent if slices were tested within 3 hours of dissection. The amount of LTD increased over the next three hours but more in adult than in aged rats. This age-related impairment was abolished by exogenous D-serine, thus reflecting the reduced activation of the NMDA-R glycine-binding site by the endogenous agonist in aged rats. Then, the amount of LTD reached asymptote at 5–7 hours following dissection. Similar temporal profiles of LTD expression were seen in young and aged Wistar rats.

Conclusions/Significance: Taken together, these results sound a cautionary note regarding the existence of an experimental “window of opportunity” for studying the effects of aging on LTD expression in hippocampal slice preparation.

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Introduction

Activity-dependent changes in synaptic strength, such as long-term potentiation (LTP) and long-term depression (LTD) of synaptic transmission, are now considered leading candidates for the cellular mechanisms underlying learning and memory [1,2,3,4,5]. Competitive interactions between these specific forms of synaptic plasticity have been reported to underlie the storage of emotional memories and stress-induced amnesia [3,6]. In fact, LTD may significantly determine the efficacy of learning and memory by limiting acquisition and favoring the decline of memory [7,8,9]. Because aging is generally associated with memory impairment, changes in susceptibility to LTD have been postulated to occur in the aged brain (see [10]). Although this question has been repeatedly investigated in the rat hippocampus, how LTD expression is affected by age still remains the subject of debate. On the one hand, an increase in susceptibility to LTD induction has been reported in aged animals [11,12,13,14,15,16,17] whereas on the other hand, several studies point more towards a weaker ability of hippocampal neuronal networks to generate synaptic depression with age [18,19,20]. Recent evidence also shows that strong stimuli do not

induce any age-related changes in the magnitude of asymptotic LTD [21]. In their respective contributions, the authors have raised the possibility of inter-strain variability or methodological differences (such as different calcium/magnesium ($\text{Ca}^{2+}/\text{Mg}^{2+}$) ratio) to account for these contradictory results. However, it is not possible to confirm the relevance of these explanations since both increased and decreased susceptibility to LTD induction have been found in the same strain and/or for the same $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio (for a comparison of these studies see [15,17,18,20]).

In studying the effects of acute stress on metabotropic glutamate receptor (mGlu-R)-dependent LTD in hippocampal CA1 area, Chaouloff et al. have recently reported that application of the mGlu-R agonist (*RS*)-3,5-dihydrophenylglycine (DHPG) promotes LTD in slices from young animals subjected to a 4–7 h period of rest before recording whereas no effect was observed after a recovery period of 1–4 h period [22]. Although this point has not been discussed by the authors, it suggests that the ability of slices to express LTD is temporally related to rest conditions, possibly through basal intracellular free calcium $[\text{Ca}^{2+}]_i$ concentrations, since a moderate increase in $[\text{Ca}^{2+}]_i$ is a prerequisite for the induction of LTD [23,24,25,26]. Interestingly, the increase in

[Ca²⁺]_i levels is dependent on the period for which the slices are maintained *in vitro* and the magnitude of this effect is significantly larger in slices from aged than from young ones [27]. In the present study, I therefore investigated age-related changes in susceptibility to LTD as a function of the resting time of the slices, in order to establish whether the discrepancies in the literature were a reflection of different conditions of the slice preparation.

Methods

Experiments were carried out in accordance with the European Communities Council Directive (86/609/EEC) regarding the care and use of animals for experimental procedures and approved by the local ethical committee (Comité régional d'éthique en experimentation animale Paris Descartes, Université Paris 5). The experiments were conducted using adult (3–5 month-old, *n* = 21) and aged (23–27 month-old, *n* = 22) male Sprague-Dawley rats and adult (*n* = 8) and aged (*n* = 6) Wistar rats purchased from IFFA-CREDO.

One animal was studied per day. The rat was anesthetized with halothane and decapitated. The hippocampus was quickly removed and placed in ice-cold artificial cerebrospinal fluid (aCSF). The composition of the aCSF was as follows (in mM): NaCl 124, KCl 3.5, MgSO₄ 1.5, CaCl₂ 2.5, NaHCO₃ 26.2, NaH₂PO₄ 1.2, and glucose 11, pH 7.4 (Ca²⁺/Mg²⁺ ratio = 1.6). This solution had a pH of 7.35 by bubbling a gas mixture of 95% O₂/5% CO₂. Its osmolarity was adjusted to 300 mOsm with an osmometer. In one series of experiments, the concentration of MgSO₄ was raised to 2.5 mM so as to reach a Ca²⁺/Mg²⁺ ratio of 1. Slices (400 μm thick) were cut and placed into aCSF warmed at 28–30°C in a holding chamber to facilitate recovery for at least 1 hr. A single slice was then transferred to the recording chamber and continuously submerged with the pre-gassed aCSF.

Extracellular recordings were obtained at 25–28°C from the apical dendritic layer of CA1 area using glass micropipettes filled with 2 M NaCl (resistance 2–8 MΩ). Field excitatory postsynaptic potentials (fEPSPs) were evoked by electrical stimulation of afferent fibers (Schaffer collaterals and commissural fibers) located in the *stratum radiatum*. Stimuli (20 μs duration) were applied between the two poles of a bipolar tungsten electrode, with one pole inserted into the slice (350 μm diameter) and the other in the bath just above the slice.

Age-related effects on LTD were determined using test stimuli applied every 10 sec and adjusted to get a baseline slope of 0.2 mV/ms. After a period of at least 10 min for stabilization, the initial slope of three averaged fEPSPs was recorded and measured for 10 min using the Acquis 1 software (CNRS, Paris) before the delivery of a low-frequency conditioning stimulation (LFS, 900 pulses, 1 Hz at test intensity). Single pulses were then resumed for 40 min following LFS. The amplitude of the stimulus artifact was also continuously monitored during the recording to ensure that changes in the fEPSP slope did not reflect alterations in tissue resistance.

Pharmacological experiments were performed in the presence of the antagonist D-2-amino-5-phosphonovalerate (D-APV, 80 μM) to block NMDA-R or the NMDA-R co-agonist D-serine at 100 μM that saturates NMDA-R glycine binding sites [28]. Drugs were added to the aCSF 10 min prior to baseline acquisition and maintained until the end of the experiment.

Data are reported as means ± SEM. Susceptibility to LTD expression was statistically estimated using analysis of variance (ANOVA) for repeated measures, carried out between the 10 min of baseline recordings and the last 10 min of recordings (i.e. 30 to 40 min after LFS). Age-related differences were determined using

repeated measures ANOVA for the last 10 min of recordings. Post hoc analyses were performed using Scheffe's F-test with the threshold for significance set at *P* < 0.05.

Results

Age-related susceptibility to LTD in control aCSF

When applied to slices from adult Sprague-Dawley rats (29 slices/18 animals) in aCSF with a Ca²⁺/Mg²⁺ ratio of 1.6, LFS produced a significant depression of fEPSPs that persisted until the end of the recording [*F*_{1,57} = 39.6, *p* < 0.0001] (Figure 1A). In aged rats (27 slices, 17 animals), LFS also induced LTD [*F*_{1,44} = 15.7, *p* < 0.001], which was not significantly different from the depression exhibited by young animals [*F*_{1,49} = 2.1, *p* = 0.1] (Figure 1A). However, studying susceptibility to LTD as a function of the delay after slice isolation revealed clear differences between the two groups of animals (Figure 1B). LFS delivered to slices shortly after slicing (1–3 h) did not induce LTD either in slices from adult rats (*n* = 5) or from aged ones (*n* = 5). In slices that were allowed to rest for 3 to 5 hours, LTD was induced by LFS in adult rats (77.7 ± 3.1% of baseline value) [*F*_{1,8} = 0.2, *p* < 0.0001, *n* = 14] but not in old animals (92.6 ± 2.5% of baseline value) [*F*_{1,8} = 4.4, *p* = 0.9, *n* = 8]. As a consequence, the magnitude of the depression was significantly weaker in slices from aged than from adult ones [*F*_{1,17} = 7.9, *p* < 0.01] (Figure 1B). Longer rest periods after slicing (5 to 7 h) did not significantly increase LTD magnitude in adult rats (71.7 ± 3.8% of baseline value) [*F*_{1,20} = 1.4, *p* = 0.3, *n* = 8]. In contrast, the extent of LTD induced in slices from aged animals (76.1 ± 4.7% of baseline value) was significantly enhanced compared to the weak depression generated at shorter delays [*F*_{1,9} = 3.9, *p* < 0.05, *n* = 8] (Figure 1B). Consequently, the level of LTD expression was not significantly different in adult and aged rats at these delays between isolation and LFS induction [*F*_{1,12} = 1.1, *p* = 0.3] (Figure 1B). At longer recovery times (>7 h), the magnitude of LTD was stabilized both in adult (71.9 ± 5.9% of baseline value, *n* = 4) and aged (75.9 ± 7.6% of baseline value *n* = 6) animals (Figure 1B).

Effects of the NMDA-R antagonist D-APV

I then asked whether LTD induced by LFS depended solely on NMDA-R activation or whether additional sources could contribute to it as a function of recovery times of the slices. When all recordings were pooled (Figure 2A), LFS delivered in the presence of D-APV (80 μM) still initially depressed fEPSPs in adult rats (26 slices/11 animals) but responses rapidly returned to basal values, indicating that LTD expression was closely linked to NMDA-R activation. In aged rats (23 slices/8 animals), LTD was also completely antagonized by D-APV (Figure 2A). As illustrated in Figure 2B, no statistically relevant depression was induced by LFS in slices from either adult or aged rats treated with D-APV, regardless of the delay between slice isolation and LFS induction. These results indicate that under our experimental conditions, only NMDA-R was concerned by the LFS protocol. However, it is worth noting that unexpected and significant potentiations were induced in slices from adult and aged rats treated with D-APV and recorded 1–3 h after isolation (116.2 ± 9% of baseline value and 120.2 ± 6.3% respectively) suggesting the activation NMDA-R-independent mechanisms under these conditions (Figure 2B). On the other hand, the fact that LTD expression was abolished by D-APV after long periods of rest in both groups of animals indicates that mechanisms that lead to the restoration of robust LTD in slices from aged rats at these intervals do not involve NMDA-R independent activation but may involve related to changes in the resting conditions of the slices.

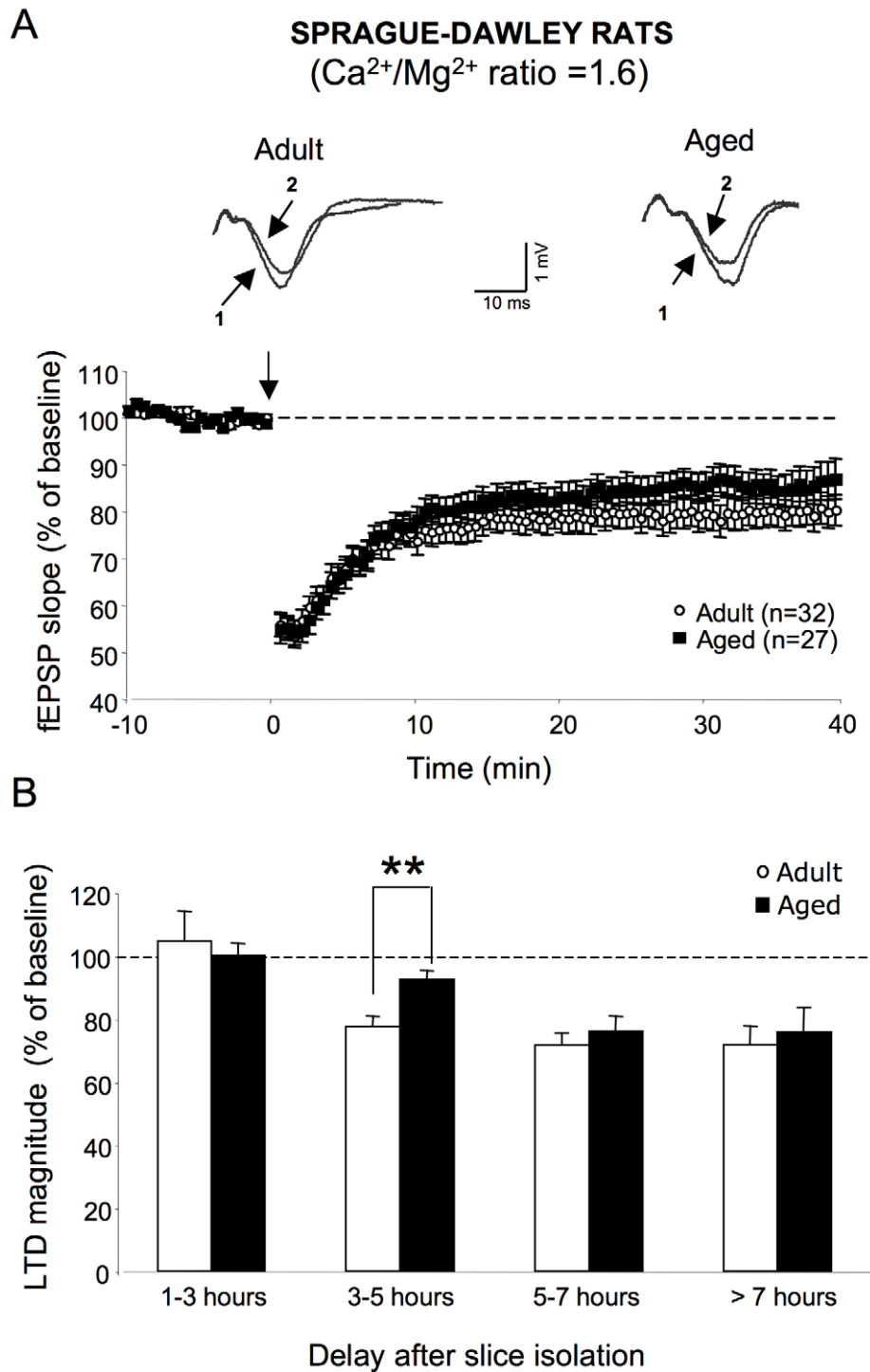


Figure 1. LFS-induced LTD is differentially expressed in slices from adult and aged Sprague-Dawley rats depending on the rest period. **A, top.** Superimposed sample traces of evoked fEPSPs induced by the electrical stimulation of glutamatergic afferents in the *stratum radiatum* of an adult (left) and aged Sprague-Dawley rat (right) before (1) and 40 min after LFS induction (2). **A, bottom.** Averaged LTD expressed as percent change in the slope of fEPSP vs. time, recorded in aCSF with a $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio of >1.5 in slices from adult (n = 32) and aged (n = 27) animals. Note that LTD is comparable in the two groups of animals. **B.** Comparison of LTD magnitude averaged from the last 10 min of recordings in adult and aged Sprague-Dawley rats as a function of the interval between slice isolation and LFS delivery (** $p < 0.01$). Note an age-related decrease in LTD expression in slices that have rested for 3 to 5 h, which is not observed at longer delays.
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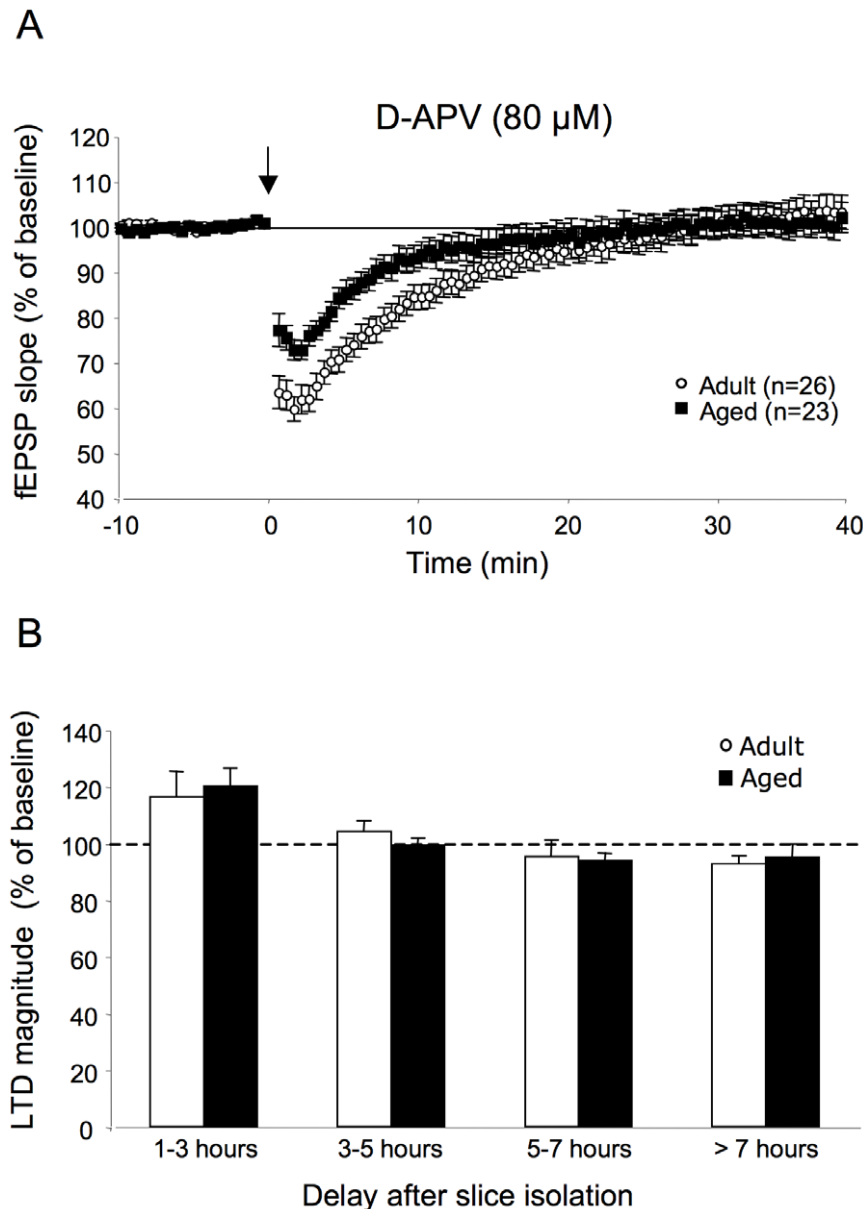


Figure 2. LFS-induced LTD is only mediated by NMDA-R activation in adult and aged rats. **A.** Averaged LTD expressed as percent change in the slope of fEPSP vs. time, recorded in slices from adult ($n = 26$) and aged ($n = 23$) Sprague-Dawley rats in the presence of the NMDA-R antagonist D-APV ($80 \mu\text{M}$). Note that D-APV prevents LTD expression in both groups of animals. **B.** Comparison of LTD expression as a function of the interval between slice isolation and LFS delivery in the two groups of animals in the presence of D-APV.
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Effects of the NMDA-R co-agonist D-serine

The possible mechanisms underlying the age-related decrease in LTD expression 3 to 5 h after the slice isolation were then addressed using the NMDA-R co-agonist D-serine at saturating doses ($100 \mu\text{M}$). This procedure has been reported to reverse the deficit in theta-burst stimulation-induced LTP in aged Sprague-Dawley [29] or Wistar rats [30].

When all recordings were pooled (except for the 1–3 h group, see below), the magnitude of the depression was not affected by D-serine in adult rats (24 slices/13 animals) [$F_{1,43} = 0.36$, $p = 0.6$] whereas it was significantly facilitated in aged animals (16 slices/11 animals) [$F_{1,41} = 14.5$, $p < 0.01$] (Figure 3A and 3B). Consequently, under these conditions of saturation of NMDA-R glycine binding

sites, the magnitude of LTD was significantly higher in slices from aged animals than in adult ones [$F_{1,33} = 5.62$, $p < 0.05$].

There were also striking differences between adult and aged Sprague-Dawley rats with respect to the effects of D-serine on LTD expression as a function of the delay after slice preparation (Figure 3C). After 1 to 3 h of rest, no effect of the NMDA-R co-agonist was detected in the former group [$F_{1,8} = 0.01$, $n = 5$, $p = 0.9$] whereas a significant potentiation ($141.4 \pm 9.1\%$ of baseline value) was induced in the latter [$F_{1,9} = 21.5$, $p < 0.01$, $n = 6$]. For periods from 3 to 5 h, LTD expression in adult animals was not changed by D-serine [$F_{1,19} = 2.24$, $p = 0.2$, $n = 7$] whereas the level of depression was significantly increased in old animals [$F_{1,13} = 9.83$, $p < 0.01$, $n = 7$]. As a consequence, the age-related

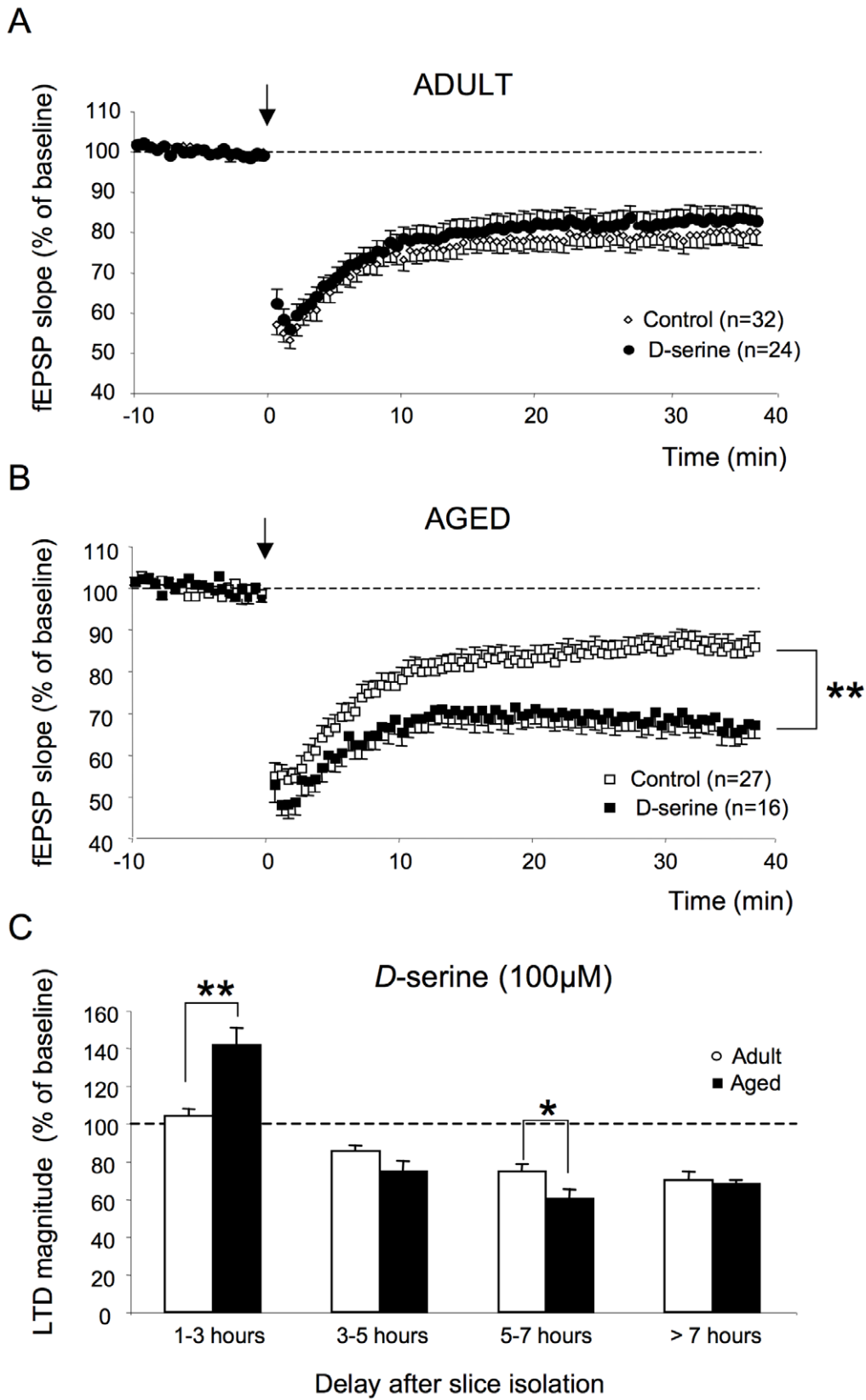


Figure 3. The NMDA-R co-agonist D-serine alters LTD expression in aged but not in adult rats. A. Averaged LTD expressed as percent change in the slope of fEPSP vs. time, recorded in adult Sprague-Dawley rats in control medium (n = 32) and in the presence of D-serine (n = 24). **B.** Same as in **A** but with recordings carried out in aged animals (control: n = 27; D-serine: n = 16). Note the significant increase in LTD magnitude in the presence of the NMDA-R co-agonist in aged rats but not in adults. (** $p < 0.01$). **C.** Comparison of LTD expression as a function of the interval between slice isolation and LFS delivery in the two groups of animals in the presence of D-serine (** $p < 0.01$, * $p < 0.05$). Note the higher LTD induced in aged rats at the 5–7 h interval.

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impairment that is normally observed under control conditions at these delays following slice recovery was alleviated in the presence of the NMDA-R co-agonist (Figure 3C). At longer rest periods (5–7 h), LTD expression with or without D-serine was again comparable in adult rats [$F_{1,14} = 0.28$, $p = 0.6$, $n = 8$] whereas LTD was still enhanced by the co-agonist in aged animals [$F_{1,12} = 5.25$, $p < 0.05$, $n = 6$]. Interestingly, LTD expression in the presence of D-serine at this point appeared to be significantly higher in aged than in adult animals [$F_{1,12} = 5.87$, $p < 0.05$] (Figure 3C). Finally, there was no significant effect of D-serine on LTD expression in either adult [$F_{1,17} = 0.29$, $p = 0.8$, $n = 4$] or aged [$F_{1,6} = 0.41$, $p = 0.5$, $n = 3$] rats at slice recovery delays of more than 7 h (Figure 3C).

Age-related susceptibility to LTD in Wistar rats

In order to determine whether the dependence of susceptibility to LTD on the delay between slicing and LFS induction is specific to Sprague-Dawley rats, a similar investigation was carried out in adult (n = 8) and aged (n = 6) Wistar rats.

When all recordings were pooled, LFS led to LTD in slices from both adult [$F_{1,62} = 79.8$, $p < 0.0001$, $n = 32$] and aged rats [$F_{1,44} = 18.2$, $p < 0.001$, $n = 19$]. Under these conditions, no age-related differences was observed [$F_{1,48} = 2.3$, $p = 0.1$] (Figure 4A). Regarding LTD susceptibility as a function of the delay after slice isolation, no noticeable depression was induced in either group of animals for slices analyzed 1 to 3 h after the isolation procedure (Figure 4B). In slices that were allowed to rest for 3 to 5 h, LTD was expressed in adult (74.2 ± 2.3% of baseline value) [$F_{1,14} = 126.7$, $p < 0.0001$, $n = 8$] as well as in old rats (86.26 ± 2.7% of baseline value) [$F_{1,108} = 29$, $p < 0.001$, $n = 6$]. However, the level of depression was significantly greater in slices from adults than in those of aged ones [$F_{1,12} = 11.5$, $p < 0.01$] (Figure 4B). At rest periods of 5 to 7 h, LTD magnitude was not changed in adult animals, as compared to earlier intervals (67.6 ± 3.8% of baseline value) [$F_{1,15} = 2.1$, $p = 0.2$, $n = 8$]. In contrast, the level of LTD was significantly enhanced in aged animals (74.5 ± 3.9% of baseline value) [$F_{1,10} = 5.9$, $p < 0.05$, $n = 6$] (Figure 4B). Consequently, LTD expression was comparable in adult and aged rats for these delays between slice isolation and LFS induction [$F_{1,12} = 1.2$, $p = 0.3$] (Figure 4B). At delays of more than 7 h, the level of LTD in adult (58.5 ± 2.6% of baseline value, $n = 7$) and aged (62.4 ± 4.6% of baseline value, $n = 4$) animals was stabilized, and no age-related difference was observed (Figure 4B). Because these results mostly reproduced those observed in Sprague-Dawley rats, they appear to reflect a general property of hippocampal tissues to the effects of aging on LTD susceptibility.

Effects of altering the $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio on susceptibility to LTD induction

Several lines of evidence in the literature indicate that LTD expression induced by a 1 Hz-conditioning stimulation, such as that used in the present study, is closely dependent on the $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio (see [31] for a review). It has been reported that LFS fails to induce LTD in young animals when this ratio is closed to 1, whereas significant depression may be induced under the same

conditions in aged rats [12,14,15]. In this section, I therefore investigated age-related changes in LTD induction as a function of the delay between slice isolation and LFS administration in the presence of varying $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio.

As illustrated in Figure 5A, no LTD was induced in adult (95.1 ± 2.3% of baseline value, $n = 26$) [$F_{1,48} = 3.53$, $p = 0.07$] or aged (96.6 ± 2.8% of baseline value, $n = 21$) [$F_{1,48} = 1.7$, $p = 0.2$] Sprague-Dawley rats in aCSF with a $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio of 1 when all recordings were pooled. This absence of depressive effect of LFS was also observed for periods of rest of up to 7 h (Figure 5B). At longer experimental times, LTD occurred in slices from adult animals (86.2 ± 2.5% of baseline value, $n = 6$) [$F_{1,10} = 30.8$, $p < 0.01$] as well as from aged ones (83.3 ± 3.7% of baseline value, $n = 5$) [$F_{1,10} = 22.1$, $p < 0.01$] with no statistically significant difference between the two groups [$F_{1,10} = 0.3$, $p = 0.6$] (Figure 5B).

Discussion

The present study provides evidence that the magnitude of LFS-induced LTD mediated by the NMDA subtype of glutamate receptors is related to the interval between slice isolation and LFS induction, and that this correlation exists regardless of the $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio of the perfusion medium. In addition, it shows that differences in susceptibility to LTD between adult and aged Sprague-Dawley or Wistar rats are present or absent depending on the time for which slices are allowed to rest before recording, which may partly explain the discrepancy seen in the literature concerning susceptibility to LTD and aging.

With respect to the expression of long-lasting depression in an CSF with a $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio of 1, no LTD occurs in adult animals, as repeatedly reported (see [32] for a review). However, weak but noticeable LTD can be induced by LFS, but only in slices that are allowed to rest for more than 7 h. A similar profile of LTD expression is found in slices from aged animals, a finding that is different from several reports showing an age-related increase in LTD expression under the same $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio conditions [11,12,13,14,15,16,17]. Nevertheless, the dependence of LTD expression on the period for which slices are allowed to rest, described in the present study, could account for this discrepancy. Indeed, it may be hypothesized that the age-related increase in LTD magnitude reported by others is simply the result of differences in the delay between the slice preparation and recording in the two groups of animals. In adult rats, in which it is possible to rapidly isolate slices and in which fEPSPs are easily recorded, LFS could be administered after a relatively short rest periods (<7 h), when no significant LTD is exhibited (see Figure 5B). In contrast, slices are usually less rapidly isolated in aged rats and electrophysiological recordings are often delayed in these animals due to difficulties in obtaining stable fEPSPs. Consequently, LFS could be induced in slices subjected to longer periods of rest (>7 h) than in those from than in adult animals, at a time when weak but significant LTD takes place.

In aCSF with a $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio of >1.5, LFS induces NMDA-R-dependent LTD in adult animals, as previously reported [18,20,33]. However, a minimal rest period of 3 h is required after slicing for the expression of a significant long-lasting depression. The magnitude of LTD then increases concomitantly

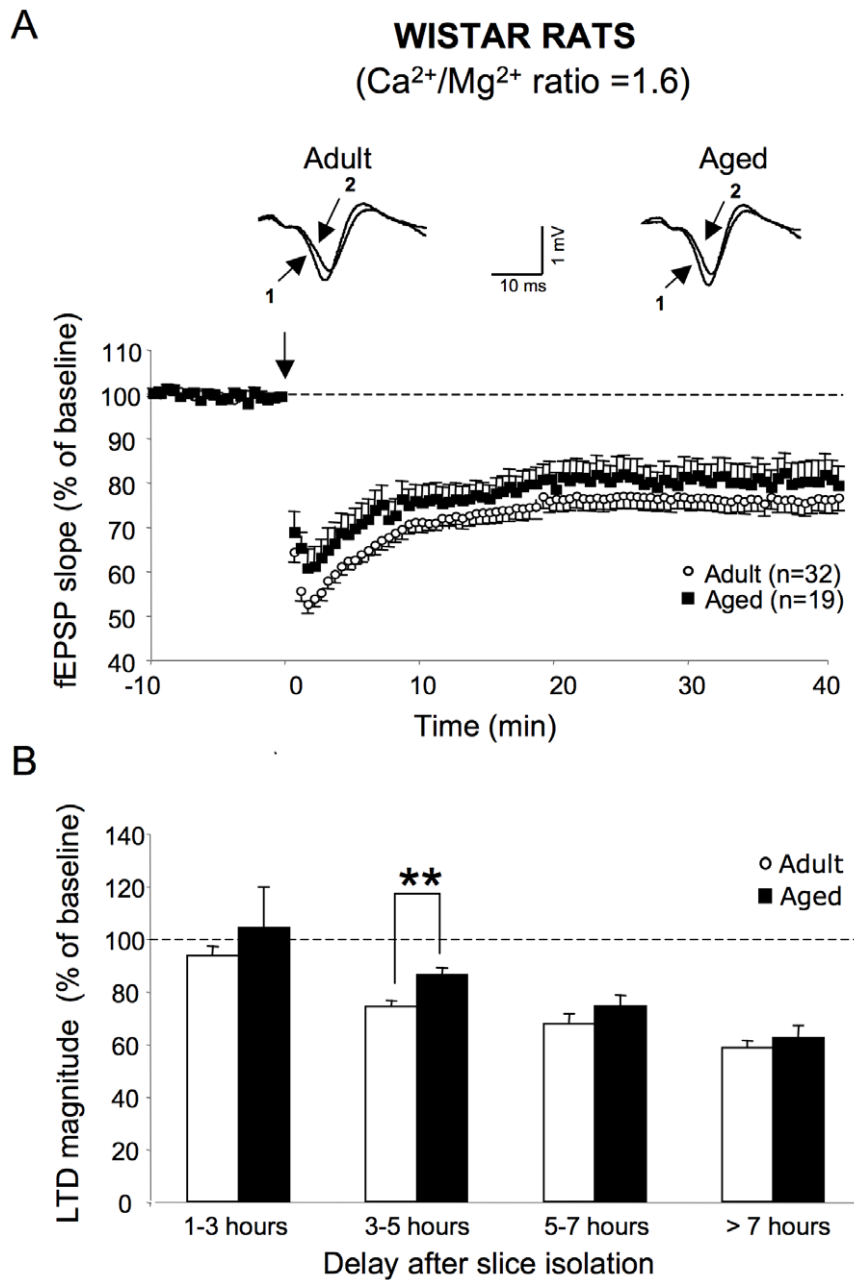


Figure 4. LFS-induced LTD is differentially expressed in slices from adult and aged Wistar rats depending on rest period. A, top. Superimposed sample traces of evoked fEPSPs induced by the electrical stimulation of glutamatergic afferents in the *stratum radiatum* of an adult (left) and aged Wistar rat (right) before (1) and 40 min after LFS induction (2). **A, bottom.** Averaged LTD expressed as percent change in the slope of fEPSP vs. time, recorded in aCSF with a Ca²⁺/Mg²⁺ ratio of >1.5 in slices from adult (n=32) and aged (n=19) animals. **B.** Comparison of LTD magnitude in adult and aged Wistar rats as a function of the interval between slice isolation and LFS delivery (** $p < 0.01$). Note the same time-course as in Figure 1B for Sprague-Dawley rats.
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with the rest period up to 5 h, and stabilizes over longer intervals. In the presence of this Ca²⁺/Mg²⁺ ratio, slices from aged rats behave differently. Even after a 5 h period of rest, no significant LTD is seen in these animals, due to the impaired activation of NMDA-R glycine binding sites. Indeed, age-related deficits in LTD are prevented when LFS is delivered in the presence of saturating concentrations of the endogenous NMDA-R co-agonist D-serine, similar to the effect of the related compound, D-cycloserine [18]. A reduction in the activation of NMDA-R glycine binding site by D-serine has recently been characterized in

aged Sprague-Dawley [28,29] and Wistar rats [30] accounting for the impaired theta-burst-induced LTP displayed by these animals.

Interestingly, when delivered to slices from aged animals allowed to rest more than 5 h, potent LTD occurs, comparable to the depression recorded in adult animals, indicating that compensatory mechanisms have taken place. At these delays separating slice isolation from LFS administration, it is interesting to note that LTD is significantly higher in aged than in adult rats if impaired NMDA-R activation is overcome by saturating doses of D-serine (see Figure 3C). NMDA-R-independent mechanisms,

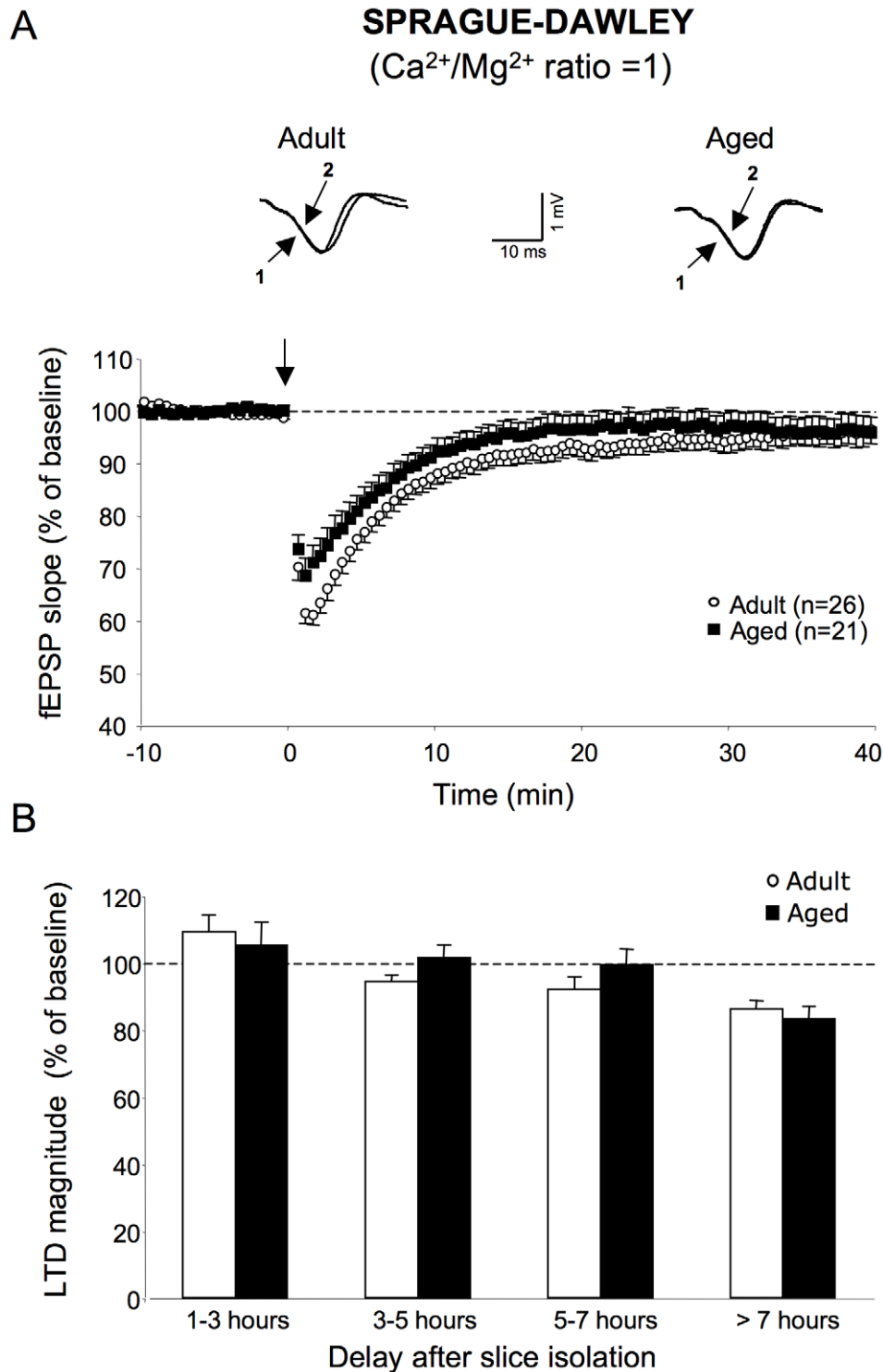


Figure 5. Effects of lowering the Ca²⁺/Mg²⁺ ratio to 1 on LTD expression in adult and aged Sprague-Dawley rats. **A, top.** Superimposed sample traces of evoked fEPSPs induced by the electrical stimulation of glutamatergic afferents in the *stratum radiatum* of an adult (left) and aged Sprague-Dawley rat (right) before (1) and 40 min after LFS induction (2). **A, bottom.** Averaged LTD expressed as percent change in the slope of fEPSP vs. time, recorded in slices from adult (n=26) and aged (n=21) animals. Note the absence of significant LTD in the two groups of animals. **B.** Comparison of LTD magnitude in slices as a function of the rest period (**p*<0.05). Note that significant LTD is induced only at the longest delays after slicing, and that it is comparable in magnitude in adult and aged rats.
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such as the activation of L-type Ca²⁺ channels [13] and intracellular Ca²⁺ stores [12], have been reported to contribute to LTD expression in aged rats. However, a significant

contribution of these pathways, or other such as the activation of metabotropic glutamate receptors [34,35] to the compensatory mechanisms is unlikely since D-APV completely antagonizes LTD

expression regardless of the recovery period of the slices. On the other hand, this result rather suggests that changes in resting conditions must instead be considered.

At CA3-CA1 synapses, a prolonged but moderate increase in postsynaptic intracellular free calcium ($[Ca^{2+}]_i$) is a prerequisite for the induction of LTD [23,24,25,26]. One can therefore intuitively postulate that changes in basal $[Ca^{2+}]_i$ will affect LTD expression by facilitating or preventing its induction as the basal $[Ca^{2+}]_i$ level increases or decreases. Interestingly, it has been reported that in cerebellar granule neurons in brain slices, basal $[Ca^{2+}]_i$ levels increase as a function of the time the slices are allowed to rest *in vitro*, and that after a delay of 3–4 h, the magnitude of this effect is significantly greater in slices from aged animals than in those from adult ones (see [27]). This weaker capacity of aged neurons to maintain a stable resting Ca^{2+} concentration has been demonstrated in slices as well as in cell culture preparations and has been primarily attributed to the inability of mitochondria to function as normal Ca^{2+} stores [36,37]. Although this possibility remains to be definitively demonstrated, it is thus conceivable that similar changes in basal $[Ca^{2+}]_i$ induced by slice isolation occur in hippocampal neurons, accounting for the time-dependence of LTD induction as well as for the weaker initial LTD in aged rats that is not observed at longer periods of slice recovery. At very short rest period (1–3 h), basal $[Ca^{2+}]_i$ levels in neurons could be too low to allow the NMDA-R-dependent Ca^{2+} entry induced by LFS to reach the threshold required for LTD expression. After 3 to 5 h of rest, as the resting $[Ca^{2+}]_i$ increases, this threshold may be reached in adult rats but not in aged ones, because of impaired NMDA-R activation. At longer times after slicing (5 to 7 h), the decrease in Ca^{2+} entry through the NMDA-R could be compensated by the greater increase in basal $[Ca^{2+}]_i$ in aged neurons, alleviating the age-related impairment of LTD. This hypothesis could also explain why LTD is higher in aged animals than in adult ones at long recovery times, when the impairment in NMDA-R activation is prevented by D-serine (see Figure 3C).

Although attractive, these hypotheses would be difficult to prove because the manipulation of the Ca^{2+} environment of neurons would affect not only basal $[Ca^{2+}]_i$ but also the Ca^{2+} dynamics induced by LFS. Alternatively, the age-related inability of mitochondria to function as normal Ca^{2+} stores could be reproduced in adult animals using the mitochondrial protonophore carbonyl cyanide p-(trifluoro-methoxy) phenylhydrazone (FCCP) [38,39] in order to determine its effects on LTD expression. However, FCCP not only modifies basal $[Ca^{2+}]_i$ in neurons but also significantly alters the amplitude of glutamate-induced Ca^{2+} transients [40], thus interfering with the process of LTD.

Although the use of slices rather than other preparations such as synaptosomes or dissociated neurons to investigate the physiology of aging has been the subject of debate [41,42], it is obvious that slices are an adequate preparation in which to simultaneously control $[Ca^{2+}]_i$ dynamics and electrophysiological responses. However, the present study points to an experimental “window of opportunity” for studying the effects of aging in hippocampal

slices, at least those dependent on changes in basal $[Ca^{2+}]_i$. The crucial point is to know whether basal Ca^{2+} is really enhanced in aged neurons *in vivo*? If the answer is yes, then age-related LTD susceptibility must be investigated in slices that have rested for long periods and that display higher values of $[Ca^{2+}]_i$ [27]. From the present study, it appears under these conditions that susceptibility to LTD is not modified in aged rats. Alternatively, if the answer is no, i.e. if basal $[Ca^{2+}]_i$ is not enhanced in aged neurons *in vivo*, LTD *in vitro* needs to be studied shortly after slice isolation, when basal $[Ca^{2+}]_i$ levels are similar in adult and aged neurons [27]. Under these conditions, we have shown that susceptibility to LTD is impaired in aging. Unfortunately, no data are yet available *in vivo* regarding the expression of LTD in the hippocampus of aged animals, that could favor one hypothesis from another. It is worth noting that *in vivo* recordings in young animals also lead to discrepancy since some studies have indicated failure to induce LTD [43,44] whereas others have shown successful induction in anesthetized or freely moving rats [45,46]. Variations in rat strains and also in the behavioral state of the animal have been proposed to underlie the discrepancy [47] but changes in basal $[Ca^{2+}]_i$ in neurons between studies could also be possibly be involved.

Although this study focuses on LTD expression, it is obvious that the length of time for which slices are allowed to rest may impact other Ca^{2+} -dependent functional events analyzed in the slice preparations such as post-burst afterhyperpolarization potentials (AHPs). Indeed, it is worth noting that an age-related increase in AHPs has been reported in some studies (reviewed in [31]) possibly reflecting the result of intracellular recordings carried out at different intervals after slicing. On the other hand, time-dependent changes in basal $[Ca^{2+}]_i$ may represent only one factor that interferes with the expression of synaptic plasticity in slice preparations. Indeed, a time-dependent loss of AMPA receptor subtypes GluR1 and GluR3 has been reported in slices from young rats suggesting that progressive changes may occur in the composition of major glutamate receptors [48]. This study also shows a time-dependent induction of mRNAs for the transcription factors *c-fos* and *zif268* and the neurotrophin brain-derived neurotrophic factor, which are all involved in the expression of synaptic plasticity. These modifications may therefore also contribute to the differences in LTD expression in slices from adult and aged animals as a function of time after slicing, although to what extent they occur in aged tissues has yet to be determined.

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Author Contributions

Conceived and designed the experiments: JmB. Performed the experiments: JmB. Analyzed the data: JmB. Contributed reagents/materials/analysis tools: JmB. Wrote the paper: JmB.

References

- Eichenbaum H (1996) Learning from LTP: a comment on recent attempts to identify cellular and molecular mechanisms of memory. *Learn Mem* 3: 61–73.
- Lynch MA (2004) Long-term potentiation and memory. *Physiol Rev* 84: 87–136.
- Braunewell KH, Manahan-Vaughan D (2001) Long-term depression: a cellular basis for learning? *Rev Neurosci* 12: 121–140.
- Neves G, Cooke SF, Bliss TV (2008) Synaptic plasticity, memory and the hippocampus: a neural network approach to causality. *Nat Rev Neurosci* 9: 65–75.
- Bliss TV, Collingridge GL (1993) A synaptic model of memory: long-term potentiation in the hippocampus. *Nature* 361: 31–39.
- Diamond DM, Park CR, Campbell AM, Woodson JC (2005) Competitive interactions between endogenous LTD and LTP in the hippocampus underlie the storage of emotional memories and stress-induced amnesia. *Hippocampus* 15: 1006–1025.
- Roman FS, Truchet B, Marchetti E, Chaillan FA, Soumireu-Mourat B (1999) Correlations between electrophysiological observations of synaptic plasticity modifications and behavioral performance in mammals. *Prog Neurobiol* 58: 61–87.
- Genoux D, Haditsch U, Knobloch M, Michalon A, Storm D, et al. (2002) Protein phosphatase 1 is a molecular constraint on learning and memory. *Nature* 418: 970–975.

9. Malleret G, Haditsch U, Genoux D, Jones MW, Bliss TV, et al. (2001) Inducible and reversible enhancement of learning, memory, and long-term potentiation by genetic inhibition of calcineurin. *Cell* 104: 675–686.
10. Foster TC (1999) Involvement of hippocampal synaptic plasticity in age-related memory decline. *Brain Res Brain Res Rev* 30: 236–249.
11. Foster TC, Kumar A (2007) Susceptibility to induction of long-term depression is associated with impaired memory in aged Fischer 344 rats. *Neurobiol Learn Mem* 87: 522–535.
12. Kumar A, Foster TC (2005) Intracellular calcium stores contribute to increased susceptibility to LTD induction during aging. *Brain Res* 1031: 125–128.
13. Norris CM, Halpain S, Foster TC (1998) Reversal of age-related alterations in synaptic plasticity by blockade of L-type Ca²⁺ channels. *J Neurosci* 18: 3171–3179.
14. Norris CM, Korol DL, Foster TC (1996) Increased susceptibility to induction of long-term depression and long-term potentiation reversal during aging. *J Neurosci* 16: 5382–5392.
15. Hsu KS, Huang CC, Liang YC, Wu HM, Chen YL, et al. (2002) Alterations in the balance of protein kinase and phosphatase activities and age-related impairments of synaptic transmission and long-term potentiation. *Hippocampus* 12: 787–802.
16. Vouimba RM, Foy MR, Foy JG, Thompson RF (2000) 17beta-estradiol suppresses expression of long-term depression in aged rats. *Brain Res Bull* 53: 783–787.
17. Foy MR, Baudry M, Foy JG, Thompson RF (2008) 17beta-estradiol modifies stress-induced and age-related changes in hippocampal synaptic plasticity. *Behav Neurosci* 122: 301–309.
18. Billard JM, Rouaud E (2007) Deficit of NMDA receptor activation in CA1 hippocampal area of aged rats is rescued by D-cycloserine. *Eur J Neurosci* 25: 2260–2268.
19. Kollen M, Stephan A, Faivre-Bauman A, Loudes C, Sinet PM, et al. (2010) Preserved memory capacities in aged Lou/C/Jall rats. *Neurobiol Aging* 31: 129–142.
20. Lee HK, Min SS, Gallagher M, Kirkwood A (2005) NMDA receptor-independent long-term depression correlates with successful aging in rats. *Nat Neurosci* 8: 1657–1659.
21. Kumar A, Thinschmidt JS, Foster TC, King MA (2007) Aging effects on the limits and stability of long-term synaptic potentiation and depression in rat hippocampal area CA1. *J Neurophysiol* 98: 594–601.
22. Chaouloff F, Hemar A, Manzoni O (2007) Acute stress facilitates hippocampal CA1 metabotropic glutamate receptor-dependent long-term depression. *J Neurosci* 27: 7130–7135.
23. Mizuno T, Kanazawa I, Sakurai M (2001) Differential induction of LTP and LTD is not determined solely by instantaneous calcium concentration: an essential involvement of a temporal factor. *Eur J Neurosci* 14: 701–708.
24. Hansel C, Artola A, Singer W (1996) Different threshold levels of postsynaptic [Ca²⁺]_i have to be reached to induce LTP and LTD in neocortical pyramidal cells. *J Physiol Paris* 90: 317–319.
25. Zucker RS (1999) Calcium- and activity-dependent synaptic plasticity. *Curr Opin Neurobiol* 9: 305–313.
26. Yang SN, Tang YG, Zucker RS (1999) Selective induction of LTP and LTD by postsynaptic [Ca²⁺]_i elevation. *J Neurophysiol* 81: 781–787.
27. Xiong J, Verkhratsky A, Toescu EC (2002) Changes in mitochondrial status associated with altered Ca²⁺ homeostasis in aged cerebellar granule neurons in brain slices. *J Neurosci* 22: 10761–10771.
28. Junjaud G, Rouaud E, Turpin F, Mothet JP, Billard JM (2006) Age-related effects of the neuromodulator D-serine on neurotransmission and synaptic potentiation in the CA1 hippocampal area of the rat. *J Neurochem* 98: 1159–1166.
29. Mothet JP, Rouaud E, Sinet PM, Potier B, Jouvenceau A, et al. (2006) A critical role for the glial-derived neuromodulator D-serine in the age-related deficits of cellular mechanisms of learning and memory. *Aging Cell* 5: 267–274.
30. Turpin FR, Potier B, Dulong JR, Sinet PM, Alliot J, et al. (2009) Reduced serine racemase expression contributes to age-related deficits in hippocampal cognitive function. *Neurobiol Aging* (2009): doi:10.1016/j.neurobiolaging.2009.09.001.
31. Billard JM (2006) Ageing, hippocampal synaptic activity and magnesium. *Magnes Res* 19: 199–215.
32. Massey PV, Bashir ZI (2007) Long-term depression: multiple forms and implications for brain function. *Trends Neurosci* 30: 176–184.
33. Kirkwood A, Dudek SM, Gold JT, Aizenman CD, Bear MF (1993) Common forms of synaptic plasticity in the hippocampus and neocortex in vitro. *Science* 260: 1518–1521.
34. Burke JP, Hablitz JJ (1994) Presynaptic depression of synaptic transmission mediated by activation of metabotropic glutamate receptors in rat neocortex. *J Neurosci* 14: 5120–5130.
35. van Dam EJ, Kamal A, Artola A, de Graan PN, Gispen WH, et al. (2004) Group I metabotropic glutamate receptors regulate the frequency-response function of hippocampal CA1 synapses for the induction of LTP and LTD. *Eur J Neurosci* 19: 112–118.
36. Xiong J, Camello PJ, Verkhratsky A, Toescu EC (2004) Mitochondrial polarisation status and [Ca²⁺]_i signalling in rat cerebellar granule neurones aged in vitro. *Neurobiol Aging* 25: 349–359.
37. Toescu EC, Verkhratsky A (2004) Ca²⁺ and mitochondria as substrates for deficits in synaptic plasticity in normal brain ageing. *J Cell Mol Med* 8: 181–190.
38. Budd SL, Nicholls DG (1996) A reevaluation of the role of mitochondria in neuronal Ca²⁺ homeostasis. *J Neurochem* 66: 403–411.
39. Schuchmann S, Luckermann M, Kulik A, Heinemann U, Ballanyi K (2000) Ca²⁺- and metabolism-related changes of mitochondrial potential in voltage-clamped CA1 pyramidal neurons in situ. *J Neurophysiol* 83: 1710–1721.
40. Wang GJ, Thayer SA (1996) Sequestration of glutamate-induced Ca²⁺ loads by mitochondria in cultured rat hippocampal neurons. *J Neurophysiol* 76: 1611–1621.
41. Verkhratsky A, Orkand RK, Kettenmann H (1998) Glial calcium: homeostasis and signaling function. *Physiol Rev* 78: 99–141.
42. Foster TC, Norris CM (1998) On slices, synaptosomes and dissociated neurones to study in vitro ageing physiology. *Trends Neurosci* 21: 286–287.
43. Errington ML, Bliss TV, Richter-Levin G, Yen K, Doyere V, et al. (1995) Stimulation at 1–5 Hz does not produce long-term depression or depotentiation in the hippocampus of the adult rat in vivo. *J Neurophysiol* 74: 1793–1799.
44. Doyle CA, Cullen WK, Rowan MJ, Anwyl R (1997) Low-frequency stimulation induces homosynaptic depotentiation but not long-term depression of synaptic transmission in the adult anaesthetized and awake rat hippocampus in vivo. *Neuroscience* 77: 75–85.
45. Heynen AJ, Abraham WC, Bear MF (1996) Bidirectional modification of CA1 synapses in the adult hippocampus in vivo. *Nature* 381: 163–166.
46. Thiels E, Xie X, Yeckel MF, Barrionuevo G, Berger TW (1996) NMDA receptor-dependent LTD in different subfields of hippocampus in vivo and in vitro. *Hippocampus* 6: 43–51.
47. Manahan-Vaughan D (2000) Long-term depression in freely moving rats is dependent upon strain variation, induction protocol and behavioral state. *Cereb Cortex* 10: 482–487.
48. Taubenfeld SM, Stevens KA, Pollonini G, Ruggiero J, Alberini CM (2002) Profound molecular changes following hippocampal slice preparation: loss of AMPA receptor subunits and uncoupled mRNA/protein expression. *J Neurochem* 81: 1348–1360.