



Draft Genome Sequence of the Ectomycorrhizal Fungus *Astraeus odoratus* from Northern Thailand

 Teeratas Kijpornyongpan,^{a*}  Weerapong Juntachai^b

^aDepartment of Botany and Plant Pathology, Purdue University, West Lafayette, Indiana, USA

^bDepartment of Biology, Faculty of Science and Technology, Chiang Mai Rajabhat University, Chiang Mai, Thailand

ABSTRACT We report the draft genome sequence of *Astraeus odoratus*, an edible ectomycorrhizal fungus from northern Thailand. The assembled genome has a size of 45.1 Mb and 13,403 annotated protein-coding genes. This reference genome will provide a better understanding of the biology of mushroom-forming ectomycorrhizal fungi in the family Diplocystidiaceae.

A *straeus odoratus* is an endemic ectomycorrhizal fungus (Diplocystidiaceae, Boletales, Basidiomycota) that forms a mutualistic relationship with the roots of dipterocarp trees in the northern regions of Thailand. The fungus forms fruiting bodies under the host trees from May to June (1). Despite being a popular delicacy in northern Thailand, artificial cultivation of *A. odoratus* remains unsuccessful.

To sequence the genome of *A. odoratus*, we utilized the vouchered specimen MTA3-1 collected from a community forest in Mae Tha District, Lamphun Province, Thailand. An inner tissue of the fruiting body was ground in liquid nitrogen, and DNA was extracted immediately using the GF-1 plant DNA extraction kit (Vivantis Technologies, Malaysia) using the manufacturer's protocol. The DNA quality and quantity were determined using NanoDrop spectrophotometry and agarose gel electrophoresis. The DNA was sent to Novogene, Singapore, for high-throughput (HT) sequencing. Around 1.5 µg of DNA was used for the library preparation using the TruSeq Nano DNA high-throughput sample preparation kit (Illumina, San Diego, CA), following the manufacturer's protocol. Sequencing was performed in paired-end (PE) mode (2 × 150 bp) on the Illumina NovaSeq 6000 platform. The 89,745,059 reads generated were assessed using FastQC 0.11.7 (2). Neither adapter sequences nor low-quality bases were detected in the reads. Ten thousand high-quality paired-end reads were randomly selected and used for BLAST searches in the NCBI nucleotide

TABLE 1 Summary statistics for the genome assembly of *A. odoratus*

Property	Value
Assembly length (bp)	45,099,382 ^a
No. of scaffolds	10,076
Largest scaffold (bp)	258,675
N_{50} (bp)	9,406
GC content (%)	49.12
Coverage (×)	107 ^a
No. of predicted protein-coding genes	13,403
Completeness of genome assembly based on:	
BUSCO conserved gene set for the phylum Basidiomycota	1,617/1,764 (91.7%)
BUSCO conserved gene set for the kingdom Fungi	659/758 (87.0%)

^aThere are 89,745,059 reads from the sequencing facility. Only 67,554,725 reads were used for assembly after removing the potential contaminants.

Citation Kijpornyongpan T, Juntachai W. 2021. Draft genome sequence of the ectomycorrhizal fungus *Astraeus odoratus* from northern Thailand. *Microbiol Resour Announc* 10:e00044-21. <https://doi.org/10.1128/MRA.00044-21>.

Editor Antonis Rokas, Vanderbilt University

Copyright © 2021 Kijpornyongpan and Juntachai. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

Address correspondence to Weerapong Juntachai, weerapong_jun@cmru.ac.th.

* Present address: Teeratas Kijpornyongpan, National Renewable Energy Laboratory, Golden, Colorado, USA.

Received 13 January 2021

Accepted 25 May 2021

Published 1 July 2021

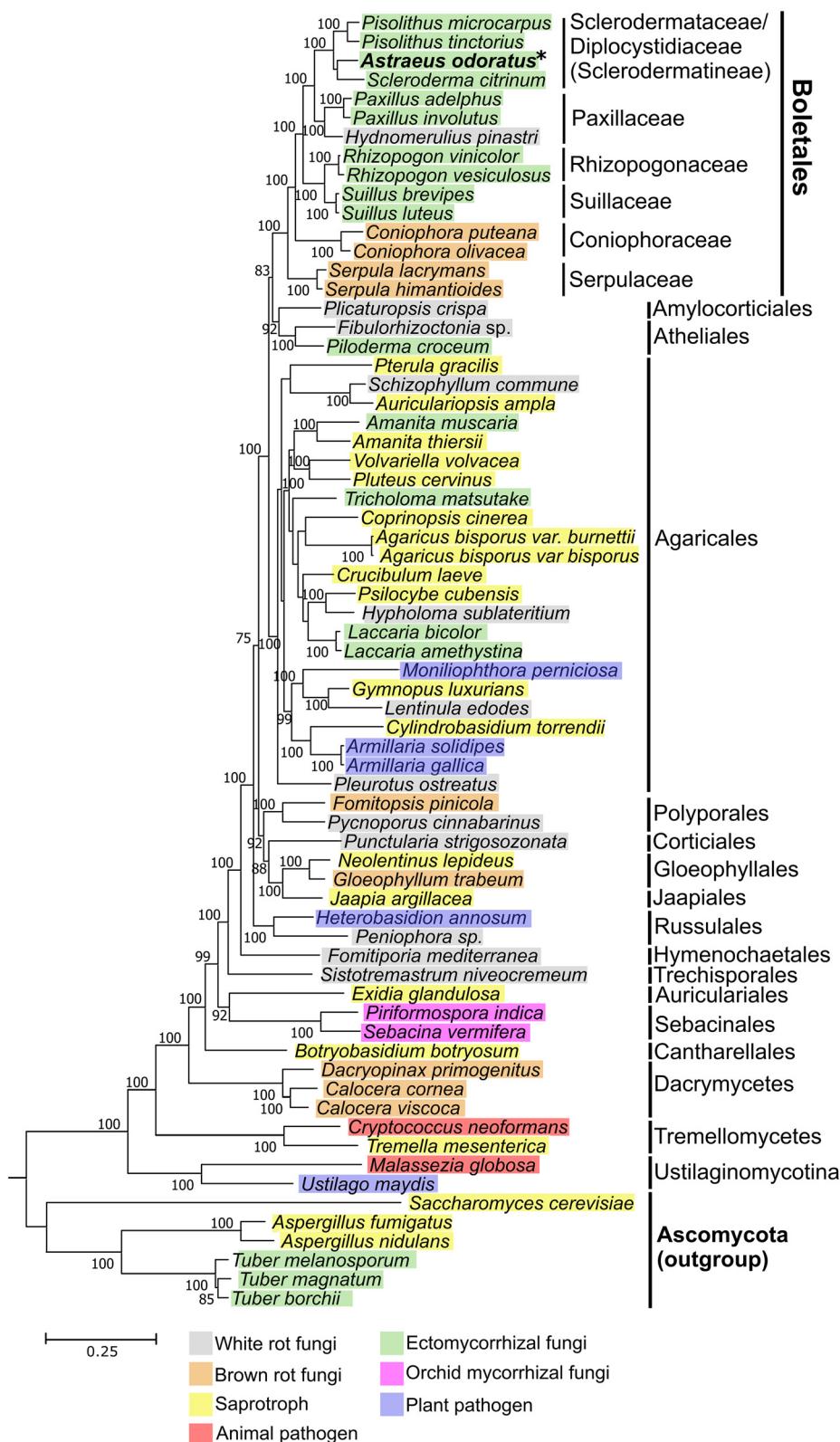


FIG 1 Phylogenetic placement of *Astraeus odoratus*. Sixty-eight fungal genomes were included in the phylogenomic analyses. The 124 single-copy BUSCO genes were used for phylogenomic reconstruction using the concatenation-RAXML maximum likelihood method. Only bootstrap values greater than 70 are shown as numbers next to the nodes. The taxonomic group and nutritional mode of each fungal species are indicated in the tree. The genome of *A. odoratus*, sequenced in this study, is shown in bold text with an asterisk.

database. Of this number, 16.28%, 12.84%, and 3.10% of reads had hits to *Enterobacter cloacae*, *Enterobacter asburiae*, and *Pseudomonas* phage OBP, respectively. This indicates potential contaminants in our data.

The genomes of *Enterobacter cloacae* ATCC 13047 (GenBank accession no. [KN150799](#)), *Enterobacter asburiae* ATCC 35953 (BioProject accession no. [PRJNA285282](#)), and *Pseudomonas* phage OBP (GenBank accession no. [NC_016571.1](#)) were utilized as references for contaminant removal (3–5). The Illumina reads were mapped to these references through Bowtie 2 2.3.5.1 using --fast-local mode and -N 1 (6). Reads not mapped to the references were *de novo* assembled using ABySS 2.2.4 with the following parameters: k = 102, q = 10, and j = 8 (7). Scaffolds having a length less than 1 kb were discarded from the final assembly. The quality assessment was determined by genome statistics using QUAST 3.2 (8) and by the completeness of conserved genes using BUSCO 2.0 with the kingdom Fungi and the phylum Basidiomycota as the data sets (9). We utilized MAKER 2.31.10 for gene prediction (10). The annotation pipeline incorporated *ab initio* gene predictions from GeneMark 4.46 using fungal gene finding mode (11) and Augustus 3.2.1 using the previously trained model of *Coprinopsis cinerea* as an external hint (12), with homology BLAST-based prediction using the protein model of *Pisolithus tinctorius* (13) (GenBank accession no. [PRJNA207840](#)) as a reference. Default parameters were used for the genome assembly and annotation unless otherwise noted. Summary statistics of the assembly are provided in Table 1.

We conducted phylogenomic analyses using 124 customized benchmarking universal single-copy ortholog (BUSCO) genes found in representative genomes from Basidiomycota, with some Ascomycota fungi as the outgroup (13–46). We downloaded these reference genomes from the JGI MycoCosm fungal genome portal (47). Protein sequences from each gene were aligned using MAFFT 6.903 (48); the alignments were trimmed using GBLOCKs 0.91 (49) with default parameters. Trimmed alignments of the 124 genes were concatenated and used for tree reconstruction using RAxML 8.2.9 (50) with the LG matrix with a gamma distribution for a heterogeneous rate across the sites (LGG) as a substitution model and 1,000-replicate bootstrapping. The tree (Fig. 1) confirms the phylogenetic placement of *A. odoratus* in Boletales as proposed by a previous study (51). Protein sequences, alignments, and phylogenetic trees are available in the FigShare repository ([10.6084/m9.figshare.14069180](#)).

This report represents the reference genome for the genus *Astraeus* and the family Diplocystidiaceae. This genome will be utilized to understand the biology of this ectomycorrhizal fungus, as well as how it forms fruiting bodies, which would be beneficial for industries in northern Thailand.

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession no. [JADGIP000000000.1](#). The version described in this paper is [JADGIP010000000](#). The sequencing reads are available under the BioProject accession no. [PRJNA663954](#) (BioSample accession no. [SAMN16189912](#)).

ACKNOWLEDGMENTS

The work was supported in part by a research grant (36/63) from the Chiang Mai Rajabhat University Research Fund. Computing resources for this study were provided by the Purdue Research Computing Facility (RCAC).

REFERENCES

1. Phosri C, Watling R, Martín MP, Whalley AJS. 2004. The genus *Astraeus* in Thailand. *Mycotaxon* 89:453–463.
2. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data.
3. Ren Y, Ren Y, Zhou Z, Guo X, Li Y, Feng L, Wang L. 2010. Complete genome sequence of *Enterobacter cloacae* subsp. *cloacae* type strain ATCC 13047. *J Bacteriol* 192:2463–2464. <https://doi.org/10.1128/JB.00067-10>.
4. Cornelissen A, Hardies SC, Shaburova OV, Krylov VN, Mattheus W, Kropinski AM, Lavigne R. 2012. Complete genome sequence of the giant virus OBP and comparative genome analysis of the diverse φKZ-related phages. *J Virol* 86:1844–1852. <https://doi.org/10.1128/JVI.06330-11>.
5. Zhu B, Wang S, Li O, Hussain A, Hussain A, Shen J, Ibrahim M. 2017. High-quality genome sequence of human pathogen *Enterobacter asburiae* type strain 1497-78^T. *J Glob Antimicrob Resist* 8:104–105. <https://doi.org/10.1016/j.jgar.2016.12.003>.
6. Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. *Nat Methods* 9:357–359. <https://doi.org/10.1038/nmeth.1923>.
7. Jackman SD, Vandervalk BP, Mohamadi H, Chu J, Yeo S, Hammond SA, Jahesh G, Khan H, Coombe L, Warren RL, Birrell I. 2017. ABySS 2.0: resource-efficient assembly of large genomes using a Bloom filter. *Genome Res* 27:768–777. <https://doi.org/10.1101/gr.214346.116>.
8. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
9. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness

- with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>.
10. Cantarel BL, Korf I, Robb SMC, Parra G, Ross E, Moore B, Holt C, Alvarado AS, Yandell M. 2008. MAKER: an easy-to-use annotation pipeline designed for emerging model organism genomes. *Genome Res* 18:188–196. <https://doi.org/10.1101/gr.674390>.
 11. Tang JD, Perkins AD, Sonstegard TS, Schroeder SG, Burgess SC, Diehl SV. 2012. Short-read sequencing for genomic analysis of the brown rot fungus *Fibroporia radiculosa*. *Appl Environ Microbiol* 78:2272–2281. <https://doi.org/10.1128/AEM.06745-11>.
 12. Stanek M, Schöffmann O, Morgenstern B, Waack S. 2006. Gene prediction in eukaryotes with a generalized hidden Markov model that uses hints from external sources. *BMC Bioinformatics* 7:62. <https://doi.org/10.1186/1471-2105-7-62>.
 13. Kohler A, Kuo A, Nagy LG, Morin E, Barry KW, Buscot F, Canbäck B, Choi C, Cichocki N, Clum A, Colpaert J, Copeland A, Costa MD, Doré J, Floudas D, Gay G, Girlanda M, Henrissat B, Herrmann S, Hess J, Höglberg N, Johansson T, Khouja H-R, LaButti K, Lahrmann U, Levasseur A, Lindquist EA, Lipzen A, Marimeisse R, Martino E, Murat C, Ngan CY, Nehls U, Plett JM, Pringle A, Ohm RA, Perotto S, Peter M, Riley R, Rineau F, Ruytinx J, Salamov A, Shah F, Sun H, Tarkka M, Tritt A, Veneault-Fourrey C, Zuccaro A, Mycorrhizal Genomics Initiative Consortium, Tunlid A, Grigoriev IV, Hibbett DS, Martin F. 2015. Convergent losses of decay mechanisms and rapid turnover of symbiosis genes in mycorrhizal mutualists. *Nat Genet* 47:410–415. <https://doi.org/10.1038/ng.3223>.
 14. Castanera R, Pérez G, López-Varas L, Amselem J, LaButti K, Singan V, Lipzen A, Haridas S, Barry K, Grigoriev IV, Pisabarro AG, Ramírez L. 2017. Comparative genomics of Coniophora olivacea reveals different patterns of genome expansion in Boletales. *BMC Genomics* 18:883. <https://doi.org/10.1186/s12864-017-4243-z>.
 15. Floudas D, Binder M, Riley R, Barry K, Blanchette RA, Henrissat B, Martinez AT, Otillar R, Spatafora JW, Yadav JS, Aerts A, Benoit I, Boyd A, Carlson A, Copeland A, Coutinho PM, de Vries RP, Ferreira P, Findley K, Foster B, Gaskell J, Glotzer D, Gorecki P, Heitman J, Hesse C, Hori C, Igashiki K, Jurgens JA, Kallen N, Kersten P, Kohler A, Kues U, Arun Kumar TK, Kuo A, LaButti K, Larrondo LF, Lindquist E, Ling A, Lombard V, Lucas S, Lundell T, Martin R, McLaughlin DJ, Morgenstern I, Morin E, Murat C, Nagy LG, Nolan M, Ohm RA, Patyshakulyeva A, Rokas A, Ruiz-Dueñas FJ, Sabat G, Salamov A, Samejima M, Schmutz J, Slot JC, St John F, Stenlid J, Sun H, Sun S, Syed K, Tsang A, Wiebenga A, Young D, Pisabarro A, Eastwood DC, Martin F, Cullen D, Grigoriev IV, Hibbett DS. 2012. The Paleozoic origin of enzymatic lignin decomposition reconstructed from 31 fungal genomes. *Science* 336:1715–1719. <https://doi.org/10.1126/science.1221748>.
 16. Mujic AB, Kuo A, Tritt A, Lipzen A, Chen C, Johnson J, Sharma A, Barry K, Grigoriev IV, Spatafora JW. 2017. Comparative genomics of the ectomycorrhizal sister species *Rhizophagus vinicolor* and *Rhizophagus vesiculosus* (Basidiomycota: Boletales) reveals a divergence of the mating type B locus. *G3 Bethesda* 7:1775–1789. <https://doi.org/10.1534/g3.117.039396>.
 17. Balasundaram SV, Hess J, Durling MB, Moody SC, Thorbek L, Progida C, LaButti K, Aerts A, Barry K, Grigoriev IV, Boddy L, Höglberg N, Kauserud H, Eastwood DC, Skrede I. 2018. The fungus that came from the cold: dry rot's pre-adapted ability to invade buildings. *ISME J* 12:791–801. <https://doi.org/10.1038/s41396-017-0006-8>.
 18. Eastwood DC, Floudas D, Binder M, Majcherczyk A, Schneider P, Aerts A, Asiegbu FO, Baker SE, Barry K, Bendiksby M, Blumentritt M, Coutinho PM, Cullen D, de Vries RP, Gathman A, Goodell B, Henrissat B, Ihrmark K, Kauserud H, Kohler A, LaButti K, Lapidus A, Lavin JL, Lee Y-H, Lindquist E, Lilly W, Lucas S, Morin E, Murat C, Oguiza JA, Park J, Pisabarro AG, Riley R, Rosling A, Salamov A, Schmidt O, Schmutz J, Skrede I, Stenlid J, Wiebenga A, Xie X, Kües U, Hibbett DS, Hoffmeister D, Höglberg N, Martin F, Grigoriev IV, Watkinson SC. 2011. The plant cell wall-decomposing machinery underlies the functional diversity of forest fungi. *Science* 333:762–765. <https://doi.org/10.1126/science.1205411>.
 19. Branco S, Gladieux P, Ellison CE, Kuo A, LaButti K, Lipzen A, Grigoriev IV, Liao H-L, Vilgalys R, Peay KG, Taylor JW, Bruns TD. 2015. Genetic isolation between two recently diverged populations of a symbiotic fungus. *Mol Ecol* 24:2747–2758. <https://doi.org/10.1111/mec.13132>.
 20. Morin E, Kohler A, Baker AR, Foulongne-Oriol M, Lombard V, Nagy LG, Ohm RA, Patyshakulyeva A, Brun A, Aerts AL, Bailey AM, Billette C, Coutinho PM, Deakin G, Doddapaneni H, Floudas D, Grimwood J, Hildén K, Kües U, Labutti KM, Lapidus A, Lindquist EA, Lucas SM, Murat C, Riley RW, Salamov AA, Schmutz J, Subramanian V, Wösten HAB, Xu J, Eastwood DC, Foster GD, Sonnenberg ASM, Cullen D, de Vries RP, Lundell T, Hibbett DS, Henrissat B, Burton KS, Kerrigan RW, Challen MP, Grigoriev IV, Martin F. 2012. Genome sequence of the button mushroom *Agaricus bisporus* reveals mechanisms governing adaptation to a humic-rich ecological niche. *Proc Natl Acad Sci U S A* 109:17501–17506. <https://doi.org/10.1073/pnas.1206847109>.
 21. Hess J, Skrede I, Wolfe BE, LaButti K, Ohm RA, Grigoriev IV, Pringle A. 2014. Transposable element dynamics among asymbiotic and ectomycorrhizal *Amanita* fungi. *Genome Biol Evol* 6:1564–1578. <https://doi.org/10.1093/gbe/evu121>.
 22. Sipos G, Prasanna AN, Walter MC, O'Connor E, Bálint B, Krizsán K, Kiss B, Hess J, Varga T, Slot J, Riley R, Bóka B, Rigling D, Barry K, Lee J, Mihaltcheva S, LaButti K, Lipzen A, Waldron R, Moloney NM, Sperisen C, Kredics L, Vágvölgyi C, Patrignani A, Fitzpatrick D, Nagy I, Doyle S, Anderson JB, Grigoriev IV, Güldener U, Münsterkötter M, Nagy LG. 2017. Genome expansion and lineage-specific genetic innovations in the forest pathogenic fungi *Armillaria*. *Nat Ecol Evol* 1:1931–1941. <https://doi.org/10.1038/s41559-017-0347-8>.
 23. Collins C, Keane TM, Turner DJ, O'Keeffe G, Fitzpatrick DA, Doyle S. 2013. Genomic and proteomic dissection of the ubiquitous plant pathogen, *Armillaria mellea*: toward a new infection model system. *J Proteome Res* 12:2552–2570. <https://doi.org/10.1021/pr301131t>.
 24. Almási É, Sahu N, Krizsán K, Bálint B, Kovács GM, Kiss B, Cseklye J, Drula E, Henrissat B, Nagy I, Chovatia M, Adam C, LaButti K, Lipzen A, Riley R, Grigoriev IV, Nagy LG. 2019. Comparative genomics reveals unique wood-decay strategies and fruiting body development in the Schizophyllaceae. *New Phytol* 224:902–915. <https://doi.org/10.1111/nph.16032>.
 25. Stajich JE, Wilke SK, Ahren D, Au CH, Birren BW, Borodovsky M, Burns C, Canback B, Casselton LA, Cheng CK, Deng J, Dietrich FS, Fargo DC, Farman ML, Gathman AC, Goldberg J, Guigo R, Hoegger PJ, Hooker JB, Huggins A, James TY, Kamada T, Kilaru S, Kodira C, Kues U, Kupfer D, Kwan HS, Lomsadze A, Li W, Lilly WW, Ma L-J, Mackey AJ, Manning G, Martin F, Muraguchi H, Natvig DO, Palmerini H, Ramesh MA, Rehmeyer CJ, Roe BA, Shenoy N, Stanke M, Ter-Hovhannisyan V, Tunlid A, Velagapudi R, Vision TJ, Zeng Q, Zolan ME, Pukkila PJ. 2010. Insights into evolution of multicellular fungi from the assembled chromosomes of the mushroom *Coprinopsis cinerea* (*Coprinus cinereus*). *Proc Natl Acad Sci U S A* 107:11889–11894. <https://doi.org/10.1073/pnas.1003391107>.
 26. Varga T, Krizsán K, Földi C, Dima B, Sánchez-García M, Sánchez-Ramírez S, Szöllősi GJ, Szarkándi JG, Papp V, Albert L, Andreopoulos W, Angelini C, Antonín V, Barry KW, Bouger NL, Buchanan P, Buyck B, Bense V, Catcheside P, Chovatia M, Cooper J, Dämön W, Desjardin D, Finy P, Geml J, Haridas S, Hughes K, Justo A, Karasiński D, Kautmanova I, Kiss B, Kocsimbé S, Kotiranta H, LaButti KM, Lechner BE, Liimatainen K, Lipzen A, Lukács Z, Mihaltcheva S, Morgado LN, Niskanen T, Noordeloos ME, Ohm RA, Ortiz-Santana B, Ovrebo C, Rácz N, Riley R, Savchenko A, Shiryav A, Soop K, et al. 2019. Megaphylogeny resolves global patterns of mushroom evolution. *Nat Ecol Evol* 3:668–678. <https://doi.org/10.1038/s41559-019-0834-1>.
 27. Floudas D, Held BW, Riley R, Nagy LG, Koehler G, Ransdell AS, Younus H, Chow J, Chiniquy J, Lipzen A, Tritt A, Sun H, Haridas S, LaButti K, Ohm RA, Kües U, Blanchette RA, Grigoriev IV, Minto RE, Hibbett DS. 2015. Evolution of novel wood decay mechanisms in Agaricales revealed by the genome sequences of *Fistulina hepatica* and *Cylindrobasidium torrendii*. *Fungal Genet Biol* 76:78–92. <https://doi.org/10.1016/j.fgb.2015.02.002>.
 28. Martin F, Aerts A, Ahrén D, Brun A, Danchin EGJ, Duchaussouy F, Gibon J, Kohler A, Lindquist E, Pereda V, Salamov A, Shapiro HJ, Wyuys J, Blaudez D, Buée M, Brokstein P, Canbäck B, Cohen D, Courte PE, Coutinho PM, Delaruelle C, Detter JC, Deveau A, DiFazio S, Duplessis S, Fraissinet-Tachet L, Lucic E, Frey-Klett P, Fourrey C, Feussner I, Gay G, Grimwood J, Hoegger PJ, Jain P, Kilaru S, Labbé J, Lin YC, Legué V, Le Tacon F, Marimeisse R, Melayah D, Montanini B, Muratet M, Nehls U, Niculita-Hirzel H, Oudot-Le Secq MP, Peter M, Quesneville H, Rajashekhar B, Reich M, Rouhier N, Schmutz J, Yin T, Chalot M, Henrissat B, Kües U, Lucas S, Van de Peer Y, Podila GK, Polle A, Pukkila PJ, Richardson PM, Rouzé P, Sanders IR, Stajich JE, Tunlid A, Tuskan G, Grigoriev IV. 2008. The genome of *Laccaria bicolor* provides insights into mycorrhizal symbiosis. *Nature* 452:88–92. <https://doi.org/10.1038/nature06556>.
 29. Park S-G, Il Yoo S, Ryu DS, Lee H, Ahn YJ, Ryu H, Ko J, Hong CP. 2017. Long-read transcriptome data for improved gene prediction in *Lentinula edodes*. *Data Brief* 15:454–458. <https://doi.org/10.1016/j.dib.2017.09.052>.
 30. Mondego JMC, Carazzolle MF, Costa GGL, Formighieri EF, Parizzi LP, Rincones J, Cotomacci C, Carrasco DM, Cunha AF, Carrer H, Vidal RO, Estrela RC, Garcia O, Thomazella DPT, de Oliveira BV, Pires ABL, Rio MCS, Araujo MRR, de Moraes MH, Castro LAB, Gramacho KP, Goncalves MS, Moura Neto JP, Goes Neto A, Barbosa LV, Guiltinan MJ, Bailey BA, mra.asm.org 4

- Meinhardt LW, Cascardo JCM, Pereira GAG. 2008. A genome survey of *Moniliophthora perniciosa* gives new insights into witches' broom disease of cacao. *BMC Genomics* 9:548. <https://doi.org/10.1186/1471-2164-9-548>.
31. Riley R, Salamov AA, Brown DW, Nagy LG, Floudas D, Held BW, Levasseur A, Lombard V, Morin E, Otillar R, Lindquist EA, Sun H, LaButti KM, Schmutz J, Jabbour D, Luo H, Baker SE, Pisabarro AG, Walton JD, Blanchette RA, Henrissat B, Martin F, Cullen D, Hibbett DS, Grigoriev IV. 2014. Extensive sampling of basidiomycete genomes demonstrates inadequacy of the white-rot/brown-rot paradigm for wood decay fungi. *Proc Natl Acad Sci U S A* 111:9923–9928. <https://doi.org/10.1073/pnas.1400592111>.
 32. Fricke J, Blei F, Hoffmeister D. 2017. Enzymatic synthesis of psilocybin. *Angew Chem Int Ed Engl* 56:12352–12355. <https://doi.org/10.1002/anie.201705489>.
 33. Ohm RA, de Jong JF, Lugones LG, Aerts A, Kothe E, Stajich JE, de Vries RP, Record E, Levasseur A, Baker SE, Bartholomew KA, Coutinho PM, Erdmann S, Fowler TJ, Gathman AC, Lombard V, Henrissat B, Knabe N, Kües U, Lilly WW, Lindquist E, Lucas S, Magnuson JK, Piumi F, Raudaskoski M, Salamov A, Schmutz J, Schwarze FWMR, vanKuyk PA, Horton JS, Grigoriev IV, Wösten HAB. 2010. Genome sequence of the model mushroom *Schizophyllum commune*. *Nat Biotechnol* 28:957–963. <https://doi.org/10.1038/nbt.1643>.
 34. Miyauchi S, Kiss E, Kuo A, Drula E, Kohler A, Sánchez-García M, Morin E, Andreopoulos B, Barry KW, Bonito G, Buée M, Carver A, Chen C, Cichocki N, Clum A, Culley D, Crous PW, Fauchery L, Girlanda M, Hayes RD, Kéri Z, LaButti K, Lipzen A, Lombard V, Magnuson J, Maillard F, Murat C, Nolan M, Ohm RA, Pangilinan J, Pereira MDF, Perotto S, Peter M, Pfister S, Riley R, Sitrit Y, Stielow JB, Szöllősi G, Žífcáková L, Štursová M, Spatafora JW, Tedersoo L, Vaario L-M, Yamada A, Yan M, Wang P, Xu J, Bruns T, Baldrian P, Vilgalys R, et al. 2020. Large-scale genome sequencing of mycorrhizal fungi provides insights into the early evolution of symbiotic traits. *Nat Commun* 11:5125. <https://doi.org/10.1038/s41467-020-18795-w>.
 35. Bao D, Gong M, Zheng H, Chen M, Zhang L, Wang H, Jiang J, Wu L, Zhu Y, Zhu G, Zhou Y, Li C, Wang S, Zhao Y, Zhao G, Tan Q. 2013. Sequencing and comparative analysis of the straw mushroom (*Volvariella volvacea*) genome. *PLoS One* 8:e58294. <https://doi.org/10.1371/journal.pone.0058294>.
 36. Nagy LG, Riley R, Tritt A, Adam C, Daum C, Floudas D, Sun H, Yadav JS, Pangilinan J, Larsson K-H, Matsuura K, Barry K, Labutti K, Kuo R, Ohm RA, Bhattacharya SS, Shirouzu T, Yoshinaga Y, Martin FM, Grigoriev IV, Hibbett DS. 2016. Comparative genomics of early-diverging mushroom-forming fungi provides insights into the origins of lignocellulose decay capabilities. *Mol Biol Evol* 33:959–970. <https://doi.org/10.1093/molbev/msv337>.
 37. Levasseur A, Drula E, Lombard V, Coutinho PM, Henrissat B. 2013. Expansion of the enzymatic repertoire of the CAZy database to integrate auxiliary redox enzymes. *Biotechnol Biofuels* 6:41. <https://doi.org/10.1186/1754-6834-6-41>.
 38. Olson Å, Aerts A, Asiegbu F, Belbahri L, Bouzid O, Broberg A, Canbäck B, Coutinho PM, Cullen D, Dalman K, Deflorio G, van Diepen LTA, Dunand C, Duplessis S, Durling M, Gonthier P, Grimwood J, Fosssdal CG, Hansson D, Henrissat B, Hietala A, Himmelstrand K, Hoffmeister D, Höglberg N, James TY, Karlsson M, Kohler A, Kües U, Lee Y-H, Lin Y-C, Lind M, Lindquist E, Lombard V, Lucas S, Lundén K, Morin E, Murat C, Park J, Raffaello T, Rouzé P, Salamov A, Schmutz J, Solheim H, Ståhlberg J, Vélez H, de Vries RP, Wiebenga A, Woodward S, Yakovlev I, Garbelotto M, et al. 2012. Insight into trade-off between wood decay and parasitism from the genome of a fungal forest pathogen. *New Phytol* 194:1001–1013. <https://doi.org/10.1111/j.1469-8137.2012.04128.x>.
 39. Zuccaro A, Lahrmann U, Guldener U, Langen G, Pfiffi S, Biedenkopf D, Wong P, Samans B, Grimm C, Basiewicz M, Murat C, Martin F, Kogel K-H. 2011. Endophytic life strategies decoded by genome and transcriptome analyses of the mutualistic root symbiont *Piriformospora indica*. *PLoS Pathog* 7:e1002290. <https://doi.org/10.1371/journal.ppat.1002290>.
 40. Loftus BJ, Fung E, Roncaglia P, Rowley D, Amedeo P, Bruno D, Vamathevan J, Miranda M, Anderson JJ, Fraser JA, Allen JE, Bosdet IE, Brent MR, Chiu R, Doering TL, Donlin MJ, D'Souza CA, Fox DS, Grinberg V, Fu J, Fukushima M, Haas BJ, Huang JC, Janbon G, Jones SJM, Koo HL, Krzywinski MI, Kwon-Chung JK, Lengeler KB, Maiti R, Marra MA, Marra RE, Mathewson CA, Mitchell TG, Pertea M, Riggs FR, Salzberg SL, Schein JE, Shvartsbeyn A, Shin H, Shumway M, Specht CA, Suh BB, Tenney A, Utterback TR, Wickes BL, Wortman JR, Wye NH, Kronstad JW, Lodge JK, et al. 2005. The genome of the basidiomycetous yeast and human pathogen *Cryptococcus neoformans*. *Science* 307:1321–1324. <https://doi.org/10.1126/science.1103773>.
 41. Kämper J, Kahmann R, Böker M, Ma L-J, Brefort T, Saville BJ, Banuett F, Kronstad JW, Gold SE, Müller O, Perlín MH, Wösten HAB, de Vries R, Ruiz-Herrera J, Reynaga-Peña CG, Snetselaar K, McCann M, Pérez-Martín J, Feldbrügge M, Basse CW, Steinberg G, Ibeas JL, Holloman W, Guzman P, Farman M, Stajich JE, Sentandreu R, González-Prieto JM, Kennell JC, Molina L, Schirawski J, Mendoza-Mendoza A, Greilinger D, Münch K, Rössel N, Scherer M, Vranes M, Ladendorf O, Vincon V, Fuchs U, Sandrock B, Meng S, Ho ECH, Cahill MJ, Boyce KJ, Klose J, Klosterman SJ, Deelstra HJ, Ortiz-Castellanos L, Li W, Sanchez-Alonso P, Schreier PH, Häuser-Hahn I, Vaupel M, Koopmann E, Friedrich G, Voss H, Schlüter T, Margolis J, Platt D, Swimmer C, Girke A, Chen F, Vysotskaia V, Mannhaupt G, Guldener U, Münterktötter M, Haase D, Oesterheld M, Mewes HW, Mauceli EW, DeCaprio D, Wade CM, Butler J, Young S, Jaffe DB, Calvo S, Nusbaum C, Galagan J, Birren BW. 2006. Insights from the genome of the biotrophic fungal plant pathogen *Ustilago maydis*. *Nature* 444:97–101. <https://doi.org/10.1038/nature05248>.
 42. Xu J, Saunders CW, Hu P, Grant RA, Boekhout T, Kuramae EE, Kronstad JW, DeAngelis YM, Reeder NL, Johnstone KR, Leland M, Fieno AM, Begley WM, Sun Y, Lacey MP, Chaudhary T, Keough T, Chu L, Sears R, Yuan B, Dawson TL, Jr. 2007. Dandruff-associated *Malassezia* genomes reveal convergent and divergent virulence traits shared with plant and human fungal pathogens. *Proc Natl Acad Sci U S A* 104:18730–18735. <https://doi.org/10.1073/pnas.0706756104>.
 43. Fedorova ND, Khalidi N, Joardar VS, Maiti R, Amedeo P, Anderson MJ, Crabtree J, Silva JC, Badger JH, Albarraq A, Angioli S, Bussey H, Bowyer P, Cotty PJ, Dyer PS, Egan A, Galens K, Fraser-Liggett CM, Haas BJ, Inman JM, Kent R, Lemieux S, Malavazi I, Orvis J, Roemer T, Ronning CM, Sundaram JP, Sutton G, Turner G, Venter JC, White OR, Whitty BR, Youngman P, Wolfe KH, Goldman GH, Wortman JR, Jiang B, Denning DW, Nierman WC. 2008. Genomic islands in the pathogenic filamentous fungus *Aspergillus fumigatus*. *PLoS Genet* 4:e1000046. <https://doi.org/10.1371/journal.pgen.1000046>.
 44. Arnaud MB, Cerqueira GC, Inglis DO, Skrzypek MS, Binkley J, Chibucus MC, Crabtree J, Howarth C, Orvis J, Shah P, Wymore F, Binkley G, Miyasato SR, Simison M, Sherlock G, Wortman JR. 2012. The *Aspergillus* Genome Database (AspGD): recent developments in comprehensive multispecies curation, comparative genomics and community resources. *Nucleic Acids Res* 40:D653–D659. <https://doi.org/10.1093/nar/gkr875>.
 45. Goffeau A, Barrell BG, Bussey H, Davis RW, Dujon B, Feldmann H, Galibert F, Hoheisel JD, Jacq C, Johnston M, Louis EJ, Mewes HW, Murakami Y, Philippsen P, Tettelin H, Oliver SG. 1996. Life with 6000 genes. *Science* 274:546–567. <https://doi.org/10.1126/science.274.5287.546>.
 46. Murat C, Payen T, Noel B, Kuo A, Morin E, Chen J, Kohler A, Krizsán K, Balestrini R, Da Silva C, Montanini B, Hainaut M, Levati E, Barry KW, Belfiori B, Cichocki N, Clum A, Dockter RB, Fauchery L, Guy J, Iotti M, Le Tacon F, Lindquist EA, Lipzen A, Malagnac F, Mello A, Molinier V, Miyauchi S, Poulaïn J, Riccioni C, Rubini A, Sitrit Y, Splivallo R, Traeger S, Wang M, Žífcáková L, Wipf D, Zambonelli A, Paolocci F, Nowrouzian M, Ottonello S, Baldrian P, Spatafora JW, Henrissat B, Nagy LG, Aury J-M, Wincker P, Grigoriev IV, Bonfante P, Martin FM. 2018. Pezizomycetes genomes reveal the molecular basis of ectomycorrhizal truffle lifestyle. *Nat Ecol Evol* 2:1956–1965. <https://doi.org/10.1038/s41559-018-0710-4>.
 47. Grigoriev IV, Nikitin R, Haridas S, Kuo A, Ohm R, Otillar R, Riley R, Salamov A, Zhao X, Korzeniewski F, Smirnova T, Nordberg H, Dubchak I, Shabalov I. 2014. MycoCosm portal: gearing up for 1000 fungal genomes. *Nucleic Acids Res* 42:D699–D704. <https://doi.org/10.1093/nar/gkt1183>.
 48. Katoh K, Toh H. 2008. Improved accuracy of multiple ncRNA alignment by incorporating structural information into a MAFFT-based framework. *BMC Bioinformatics* 9:212. <https://doi.org/10.1186/1471-2105-9-212>.
 49. Talavera G, Castresana J. 2007. Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. *Syst Biol* 56:564–577. <https://doi.org/10.1080/10635150701472164>.
 50. Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30:1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>.
 51. Phosri C, Watling R, Suwannasai N, Wilson A, Martín MP. 2014. A new representative of star-shaped fungi: *Astraeus sirindhorniae* sp. nov. from Thailand. *PLoS One* 9:e71160. <https://doi.org/10.1371/journal.pone.0071160>.