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Database on the taxonomical characterisation and potential toxigenic capacities of microorganisms used for the industrial production of food enzymes and feed additives, which do not have a recommendation for Qualified Presumption of Safety

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Abstract

The present work constitutes the external scientific report of the EFSA open call OC/EFSA/FEED/2015/01. The aim of the call was to provide EFSA with a database from a review on the taxonomical description and potential toxigenic capacities of microorganisms used for the industrial production of feed additives and food enzymes. The review includes microorganisms used as source of feed additives and food enzymes for which EFSA has received or can potentially receive applications for safety assessment, and which have not been recommended for Qualified Presumption of Safety status. The database also comprises the molecular taxonomical identifiers and biosynthetic pathways involved in the production of toxic compounds and the responsible genes. The main result of the project is shown as a database developed according to the EFSA data structure. The methodological aspects and the queries used in the systematic search, as well as the procedure applied for the screening of scientific documents retrieved are described in this report. Details are available in supplementary appendices.

In total, 22970 scientific documents were screened in the literature search, from which 411 were initially selected for providing pertinent data for the scope of the project. From the review of the selected articles, 474 bioactive secondary metabolites were recorded and 59 compounds were further studied in order to obtain data on their toxicology and the conditions in which they are produced by the microorganisms used in industrial fermentations. The database generated in this project comprises details that characterise, when available, the production conditions, genes involved and toxicity of these 59 compounds. This provides information that can be used to establish safety measures when using potentially toxigenic microorganisms in industrial fermentations

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Key words: Systematic review, literature search, safety, QPS, risk assessment, data collection

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Summary

The use of microorganisms in industrial fermentations for the production of food and feed additives (enzymes, vitamins, antimicrobial substances, etc.) with beneficial effects for agriculture, farming and primary food production is in continuous development. However, the production of secondary metabolites by these microorganisms should be studied and the safety of the products obtained during fermentation should be assessed properly before its commercialization.

In order to define appropriate endpoints for the safety assessment of microbial fermentation products, it is necessary to identify the production microorganisms and the range of toxic compounds which can be produced by them. In addition, further information on the molecular and genetic basis of the production of such metabolites helps to evaluate and compare the efficiency of any genetic manipulation that could be designed and performed to avoid their synthesis. Thus, obtaining a database from a review on the appropriate taxonomical identifiers and potential toxigenic capacities of these microorganisms provides information that can be used to establish safety measures when using potentially toxigenic microorganisms in industrial fermentations.

Regulation (EC) No 1831/2003 establishes the rules for the authorisation of feed additives in the European Union (EU) according to the principles in Regulation (EC) No 429/2008. In addition, Regulation (EC) No 1332/2008 on food enzymes constitutes the legal basis for the authorisation and use of food enzymes in the European Union (EU). The procedural aspects concerning the authorisation and the evaluation procedure of food enzymes are laid down in Regulation (EC) No 1331/2008, which establishes a common authorisation procedure for food additives, food enzymes and food flavourings.

This report is the external scientific report of the European Food Safety Authority (EFSA) contract OC/EFSA/FEED/2015/01, and describes the methodology and results of a project aiming to provide EFSA with a database from a review on the correct taxonomical description and potential toxigenic capacities of microorganisms used for the industrial production of feed additives and food enzymes. Data collection is an important task of EFSA and a fundamental component of risk assessment (Articles 22 and 23 of Regulation (EC) No 178/2002). The extraction of the scientific information was retrieved in a database, which structure of the data model and its terminology was established according to EFSA standard coding systems.

The methodology applied in this project consisted in the following steps: systematic search by the application of a well-defined searching strategy, the study of the information obtained and extraction methods and the integration procedure to store the relevant information.

The database contains detailed information on (1) taxonomical descriptions and potential toxigenic capacities of microorganisms used for the industrial production of feed additives and food enzymes; (2) the type of toxic compounds that can be produced; (3) the physiological conditions in which each identified toxic compound is produced; (4) the biosynthetic pathways and corresponding genes leading to the production of the toxic compound; and (5) the mechanism of toxic action of the identified toxic compound and the dose under which harmful effects are observed in humans and animals.

The systematic search has been done with the aid of an *ad hoc* searching platform (iWatch) to perform automatic searching that supports the data searching, extraction and management of the workflow. This automatic searching tool was set up to access eight searching engines as well as 45 web sites and provides a way for organising the data in a temporary local database that could be then exported to be integrated in the final database.

In total, 22970 scientific documents have been retrieved with the searching platform and manual search, classified by microorganism and reviewed, to obtain pertinent data. A total of 411 scientific documents were selected that contain data on 474 secondary metabolites with bioactivity. A selection

of 59 toxic or potentially toxic compounds identified in these documents was further examined for retrieving information on toxicity doses, biosynthesis and conditions under which they are produced. All suitable data extracted from the relevant articles was introduced in the database developed in this project.

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1. Introduction

1.1. Background and Terms of Reference as provided by the requestor

The aim of the contract was the preparation of a database from the results of a systematic search set to establish potential toxicological profile of microorganisms for which EFSA has received or can potentially receive applications for safety assessment, and which have not been recommended for Qualified Presumption of Safety (QPS) status, excluding the microorganisms for which EFSA has already established a guidance for the assessment of their potential toxicity. The prokaryotic and eukaryotic microorganisms that should be considered as minimum are alphabetically listed as follows:

Actinomadura yumaensis, Actinomadura roseorufa, Arthrobacter ramosus, Aspergillus aculeatus, Aspergillus japonicus, Aspergillus melleus, Aspergillus niger, Aspergillus oryzae, Aspergillus sojae, Candida cylindracea, Candida lipolytica, Candida rugosa, Cellulosimicrobium cellulans, Chaetomium erraticum, Chaetomium gracile, Chryseobacterium proteolyticum, Corynebacterium glutamicum, Cryphonectria parasitica, Disporotrichum dimorphosporum, Escherichia coli, Fusarium venenatum, Geobacillus stearothermophilus, Geobacillus caldoproteolyticus, Geobacillus pallidus, Hansenula polymorpha, Humicola insolens, Klebsiella planticola, Klebsiella pneumoniae, Leptographium procerum, Leuconostoc citreum, Microbacterium imperiale, Micrococcus luteus, Mucor javanicus, Paenibacillus alginolyticus, Paenibacillus lentus, Paenibacillus macerans, Penicillium camemberti, Penicillium chrysogenum, Penicillium citrinum, Penicillium decumbens, Penicillium funiculosum, Penicillium multicolor, Penicillium lilacinum, Penicillium roqueforti, Pseudomonas aeruginosa, Pseudomonas amyloclavata, Pullulanibacillus naganoensis, Rhizomucor miehei, Rhizopus niveus, Rhizopus oryzae, Sphingobacterium multivorum, Sporobolomyces singularis, Streptomyces albus, Streptomyces aureofaciens, Streptomyces chrestomyceticus, Streptomyces chromofuscus, Streptomyces cinnamomensis, Streptomyces cinnamoneus, Streptomyces lasaliensis, Streptomyces lividans, Streptomyces mobaraensis, Streptomyces murinus, Streptomyces netropsis, Streptomyces olivochromogenes, Streptomyces rubiginosus, Streptomyces violaceoruber, Streptoverticillium mobaraense, Trametes hirsuta, Talaromyces emersonii, Talaromyces cellulolyticus, Talaromyces versatilis, Trametes versicolor, Trichoderma citrinoviride, Trichoderma harzianum, Trichoderma koningii, Trichoderma longibranchiatum, Trichoderma reesei, Trichoderma viride.

The particular objectives of this project were:

- To perform a systematic search and to collect all relevant scientific information with respect to the characterization of microorganisms used to produce food enzymes and feed additives, with the following requisites:
 - a) To include as a minimum the microorganisms listed above.
 - b) To cover a time span of the last 20 years, only reduced in cases that the amount of literature references retrieved for a given species was higher than 500.
- To extract pertinent scientific information retrieved in an Excel database, with the following requisites:
 - a) A structure of the data model and its terminology in agreement with EFSA and including the use of EFSA standard coding systems for the items when possible.
 - b) The contents of the database with the following information for each considered microbial species:
 - Valid scientific name, scientific synonyms (when appropriate) and the existing methods for its taxonomical identification.
 - Toxic compounds or secondary metabolites potentially toxic for humans or animals and harmful doses.

- Physiological conditions of the microorganism for the production (or potential production) of toxic compounds or relevant substances.
- Biosynthetic pathways and genes implicated in the production of toxic compound and relevant substances.
- To prepare a collection of module maps containing graphical representations of the biosynthetic pathways leading to the integrated synthesis of the different toxic compounds included in the database, including the list of genes and/or clusters involved in such synthesis, highlighting matching genes between pathways.
- To prepare an external scientific report to support the preparation of Guidance Documents on the subject, with the following requisites:
 - a) Protocol for the search and data integration.
 - b) Inclusion and exclusion criteria for selecting references.
 - c) Summary of the literature review and data collection and main findings.
 - d) Complete list with the relevant sequences and literature references, per species.
 - e) A proposal to keep the databases updated in an efficient manner

This contract/grant was awarded by EFSA to:

AINIA, centro tecnológico. Parque Tecnológico de Valencia, Paterna, Valencia, Spain.

Beneficiaries: AINIA, Universidad de Navarra

Contract/Grant title: Database on the taxonomical identification and potential toxigenic capacities of microorganisms used for the industrial production of food and feed additives, which do not have a qualified presumption of safety

Contract/Grant number: OC/EFSA/FEED/2015/01.

2. Methodology

The principles of Systematic Review methodology (EFSA, 2010) have been applied to the literature search protocol (Figure 1). This involves the following steps:

- defining the review objectives and developing the eligibility criteria for studies;
- searching for research studies;
- selecting studies for inclusion or exclusion in the review;
- assessing validity and quality of included studies;
- collecting data from the included studies and creating evidence tables;
- synthesising data from included studies;
- presenting data and results.

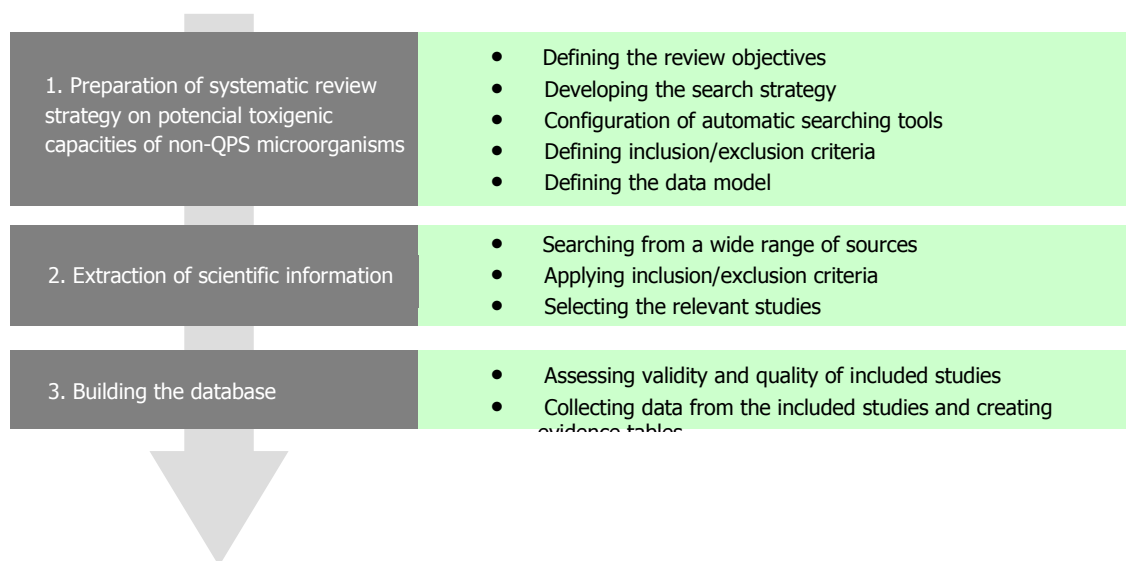


Figure 1: Methodology workflow for the literature search and data collection.

2.1. Defining the review objectives

The first step of the literature review process consisted of the analysis of the five review objectives in order to identify the key elements and to clarify their scopes.

REVIEW OBJECTIVE 1: Description of the current valid scientific name, taxon assignment, synonyms and the existing methods for the taxonomical description of microorganisms used to produce food enzymes and feed additives, which have not been recommended for Qualified Presumption of Safety (QPS) status.

Key aspects:

- To collect updated and relevant scientific information about the identification, classification and nomenclature of the microorganisms used to produce food enzymes and feed additives.

REVIEW OBJECTIVE 2: Identification of toxins or potentially toxic secondary metabolites/substances produced by the microorganisms used to produce food enzymes and feed additives.

Key aspects:

- To identify and characterise substances with toxic or other undesirable properties that are synthesised by the microorganisms used in biotechnological processes (some of them may be produced during the fermentative processes for obtaining food enzymes & feed additives). This review objective focuses on the safety assessment of unintentionally produced substances, therefore the review does not consider the safety of the food enzyme or feed additive itself.

- Some of the species of microorganisms used for the production of substances of interest have many different strains, within them could be present pathogenic strains. This review does not include the production of well-defined toxins or undesirable substances by recognised pathogenic strains.

REVIEW OBJECTIVE 3: Identification of the conditions under which the microorganism can produce the toxic compound.

Key aspects:

- Identification of culture conditions (life cycle and physiological conditions) as well as external factors (temperature, aeration, etc.) in which the identified toxic compounds are produced.

REVIEW OBJECTIVE 4: Toxicity characterisation of toxic compounds produced by the microorganism.

Key aspects:

- To identify the adverse effects of each identified toxic compound and the dose under which harmful effects are observed in humans and animals.

REVIEW OBJECTIVE 5: Biosynthetic pathway and genetic characterisation of the toxic compounds produced by the microorganism.

Key aspects:

- Acquisition of knowledge on the genetic basis of the production of toxic metabolites to assess the efficiency of any genetic manipulation performed in the microorganisms used in fermentative processes, in order to prevent the synthesis of these undesirable products.

2.2. Developing the searching strategy

The methodology applied is summarised in Figure 2.

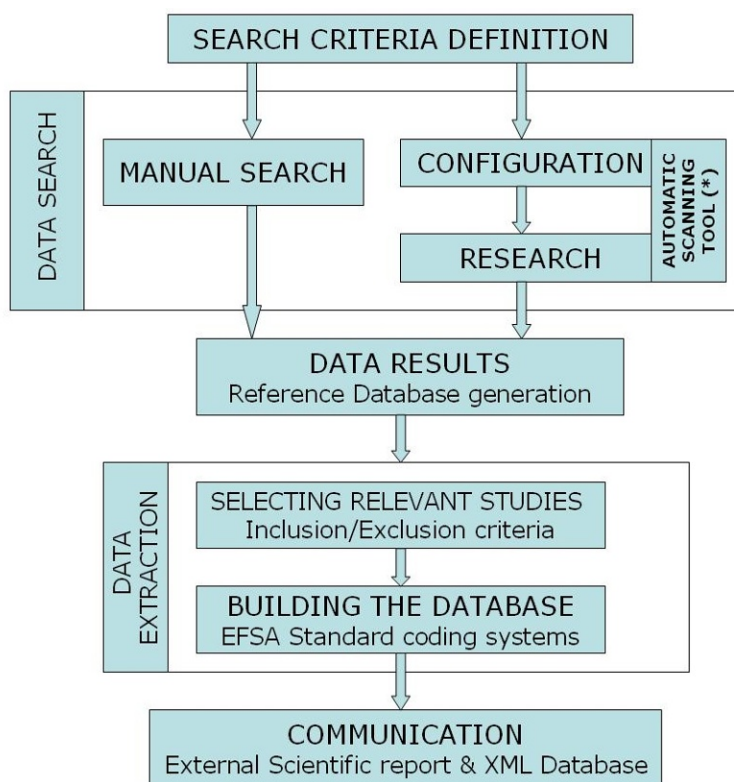


Figure 2: Workflow for the search strategy and data extraction.

As shown in Figure 2, after defining search criteria according to the required information and validated by EFSA, the systematic data search (A) was performed by two ways, either by manual searching or by the automatic scanning tool of the iWatch platform (Web Crawler, marked with an asterisk), which was used in all the cases where structured data sources allowed data scrapping. The scanning tool was configured with the information sources (scientific searching engines as well as scientific web pages) that were accessible to the crawling tool designed as well as with the keywords defined in the searching strategy. This system allowed for rapid data collection from multiple sources and easy

access to the information. Searches in engines or other information sources which did not allow automatic crawling were done manually.

To allow the management of all references (both automatic and manual searches) iWatch Platform provided a user interface as a collaborative environment for the users. On one side, the platform facilitated the integration of all results and their analysis after its configuration with the working methodology based on the inclusion/exclusion criteria. On the other side, the system permitted users to add information by filling formularies configured to include the required information for the data base.

2.2.1. Outline of the searching strategy and definition of the keywords

The searching strategy was defined after a preliminary study in which, general information about the fermentative process involving the microorganisms within the scope was obtained. This allowed to identify possible problems that can arise when retrieving data from this heterogeneous group of microorganisms, for instance, the large amount of literature associated to some of the microorganisms of the scope of this review (*Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Aspergillus niger*), and that is not relevant to the subject of the project, as well as the limited number of data available in the case of other microorganisms.

In this previous preliminary study performed to prepare the searching strategy, a review of the entries obtained by each species in the searching engine PubMed was done. This preliminary work helped to identify useful terms for the search as well as which microorganisms are more or less reported in the scientific documents.

According to these findings, several groups of species and groups of keywords were established to perform the searching strategy. The groups of keywords are the following:

- Keywords group 1: Terms related to toxin production and hazards
- Keywords group 2: Terms related to feed additives and food enzymes
- Keywords group 3: Terms related to fermentative processes
- Keywords group 4: Terms related to toxicology
- Keywords group 5: Terms related to biosynthetic pathways

The names of the species of **microorganisms** considered for the scientific search are listed in Section 1.1 of this document, as well as synonyms found for some of these species (see Appendix A), and the following considerations were taken into account:

The species *Corynebacterium glutamicum* and *Leuconostoc citreum* originally included in the list of microorganisms considered for this procurement, have been excluded as they are classified and confirmed as QPS in the latest revision 2016 (EFSA BIOHAZ Panel, 2013c). On the contrary, in the preliminary study the species *Penicillium salami* (Perrone *et al.* 2015) has been identified as a new species to be included and has been added to the list. Regarding the species *Escherichia coli*, the term *K12* was used to exclude documents related to the pathogenic nature of many of the serogroups and strains of *E. coli*.

The microbial species have been divided into 3 groups, Species I, Species II, and Species III, according to the preliminary outcome in PubMed search:

The microorganisms that belong to **Species I**: Microorganisms that produced ≤ 200 entries when searched by just their scientific name. This group is integrated by the following microorganisms: *Actinomadura yumaensis*, *Actinomadura roseorufa*, *Arthrobacter ramosus*, *Aspergillus japonicus*, *Aspergillus melleus*, *Aspergillus sojae*, *Cellulosimicrobium cellulans*, *Chaetomium erraticum*, *Chaetomium gracile*, *Chryseobacterium proteolyticum*, *Disporotrichum dimorphosporum*, *Fusarium*

venenatum, Geobacillus caldoproteolyticus, Geobacillus pallidus, Humicola insolens, Klebsiella planticola, Leptographium procerum, Microbacterium imperiale, Mucor javanicus, Paenibacillus alginolyticus, Paenibacillus lentus, Paenibacillus macerans, Penicillium camemberti, Penicillium decumbens, Penicillium funiculosum, Penicillium multicolour, Penicillium lilacinum, Penicillium salamii, Pseudomonas amyloclavata, Pullulanibacillus naganoensis, Rhizopus niveus, Sphingobacterium multivorum, Sporobolomyces singularis, Streptomyces chrestomyceticus, Streptomyces chromofuscus, Streptomyces cinnamonensis, Streptomyces cinnamoneus, Streptomyces lasaliensis, Streptomyces mobaraensis, Streptomyces murinus, Streptomyces netropsis, Streptomyces olivochromogenes, Streptomyces rubiginosus, Streptomyces violaceoruber, Trametes hirsuta, Talaromyces cellulolyticus, Talaromyces emersonii, Talaromyces cellulolyticus, Talaromyces versatilis, Trichoderma citrinoviride, Trichoderma koningii, Trichoderma longibranchiatum.

The microorganisms that belong to **Species II**: Microorganisms that produced > 200 but ≤ 500 entries when searched by scientific name and keywords from group 1. This group is integrated by the following microorganisms: *Aspergillus aculeatus, Aspergillus oryzae, Candida lipolytica, Candida rugosa, Cryphonectria parasitica, Geobacillus stearothermophilus, Hansenula polymorpha, Penicillium chrysogenum, Penicillium citrinum, Penicillium roqueforti, Rhizomucor miehei, Rhizopus oryzae, Streptomyces albus, Streptomyces aureofaciens, Streptomyces lividans, Trametes versicolor, Trichoderma harzianum, Trichoderma reesei, Trichoderma viride.*

The microorganisms that belong to **Species III**: Microorganisms that produced > 500 entries when searched by scientific name and keywords from group 1. This group is integrated by the following microorganisms: *Aspergillus niger, Escherichia coli K12, Klebsiella pneumoniae, Micrococcus luteus* and *Pseudomonas aeruginosa.*

With the considerations of the preliminary work and in order to retrieve information in the most effective way, a stepwise search was proposed. Figure 3 shows a flowchart of the search.

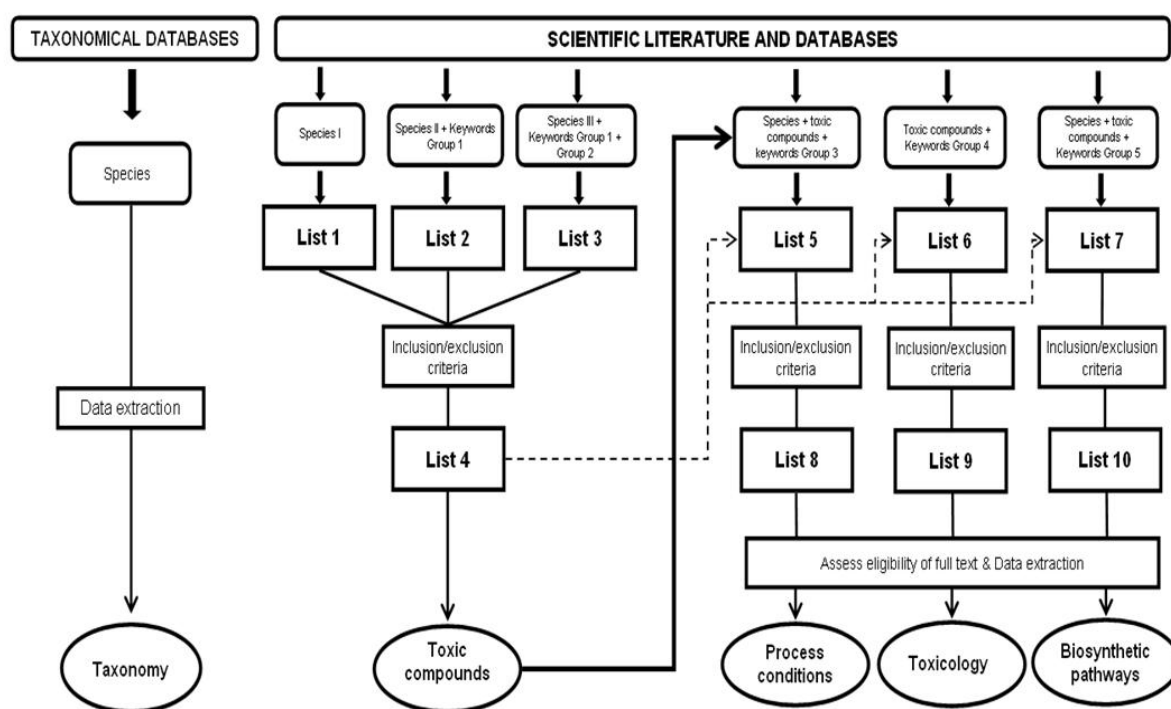


Figure 3: Searching strategy. Oval shape figures represent the main outcome descriptors containing endpoints.

For the purpose of the present review, a specific search to obtain taxonomical data was performed in databases specific for taxonomy (see Section 2.2.3) by searching each of the names of the species considered in the scope of the review. The information obtained by this search provided data for the review objective 1.

For the rest of the review objectives, the search was carried out using the different groups of keywords described above, that cover a range of searching terms in relation to toxins, additives, fermentative processes, biosynthetic pathways and toxicology.

A combination of the name of the species and the different groups of keywords allowed to obtain several lists of documents that were further screened by applying the inclusion and exclusion criteria (Section 2.3).

For the group **Species I**, the lists of documents were obtained by searching only the name of microorganism (**List 1**). For **Species II**, the **List 2** was obtained by searching the name of the microorganism AND keywords group 1. For **Species III**, the **List 3** was obtained by searching the microorganism name AND keywords group 1 AND keywords group 2. Once obtained these three lists, the inclusion/exclusion criteria defined for this first screening (see Table 2 in section 2.3.1) were applied to obtain the **List 4**. The documents from this **List 4** give information of the toxins or potentially toxic compounds that can be produced by the microorganism. They could be directly included in the **Lists 5, 6 or 7**, depending on the type of information provided. The toxic compounds retrieved from these documents were used as keywords for three new searches:

- **List 5:** toxic compounds combined with keywords group 3 (process conditions) and the name of the species. The documents resulting from this search were subjected to the corresponding inclusion/exclusion criteria ensuring that they cover conditions of production of the toxic substance when produced by the microorganisms within the scope of the project. This fed **List 8** that went for data extraction.
- **List 6:** toxic compounds combined with keywords group 4 (toxicology). The documents resulting from this search were subjected to the corresponding inclusion/exclusion criteria to feed **List 9** that went for data extraction.
- **List 7:** toxic compounds combined with keywords group 5 (biosynthetic pathways) and the name of the species. The documents resulting from this search were subjected to the corresponding inclusion/exclusion criteria ensuring that they covered genes and pathways for the production of the toxic substance by the microorganisms within the scope of the project. This fed **List 10** that went for data extraction.

The documents included in List 8, 9 and 10 were full-text screened for quality and adequacy before data extraction.

The search keywords used in relation to toxic compounds, food & feed additives, fermentation conditions and toxicology, are listed below.

- For toxic compounds relation (Keywords Group 1: TOXINS & BIOHAZARD):

adverse effect OR aflatoxin OR allerg* OR allergy OR bacteremi* OR biogenic amine* OR biohazard OR candidaemia OR carcinogen* OR coccidios* OR colitis OR co-ocurrence of toxin* OR cytotox* OR death OR developmental toxicity OR disease* OR emerging OR endocrine disruption OR endotoxin OR enterotoxin OR fungemia OR genotox* OR health OR hepatotox* OR hypersensitivity OR immune OR immunotox* OR infection* OR morbid* OR mortal* OR mutagen* OR mycosis OR mycotoxin OR necro* OR nephrotox* OR neurotox* OR ochratoxin OR opportunistic OR pathogen* OR poison* OR qualified presumption of safety OR reprotox* OR risk* OR safety OR safety assessment OR sepsis OR septicaemia OR toxic peptid* OR toxic* OR toxicokinetic* OR toxigenic potential OR toxigenic OR toxin OR undesirable co-product* OR undesirable substance* OR virulence.*

- For food and feed relation (Keywords Group 2: ADDITIVES):

additive OR amino acid* OR aminopeptidase OR amylase OR antibiotic* OR arginine OR cellulose OR enzyme* OR galactosidase OR glucanase OR glucosidase OR hydrolase OR inulinase OR laccase* OR lasalocid OR lipase* OR lipopolysaccharide OR lysine OR mannanase OR metabol* OR methionine OR monensin OR mutant* OR narasin OR paromomycin OR pectinase OR penicillin OR phytase OR polygalacturonase OR proline OR protease* OR salinomycin OR threonine OR transglutaminase OR tryptophan OR valine OR vitamin* OR xylanase OR carbapenemase OR semduramicin OR salinomycin OR glutaminase OR pullulanase OR phospholipase OR phosphodiesterase OR glucoamylase OR nuclease OR asparaginase OR trypsin OR urease OR biosynthesis OR bioprod* OR fermentation.*

- For fermentative process relation (Keywords Group 3: FERMENTATIVE PROCESS):

aerobic OR aerobiosis OR anaerobic* OR anaerobiosis OR aeration OR bioprocess OR bioprod* OR bioreact* OR biosynthesis OR by-product OR culture conditions OR culture media OR culture medium OR downstream OR end product OR engineer* OR feed OR fermentation OR food OR food industry OR immobil* OR life cycle OR life stage OR metabol* OR performance* OR pH OR physicochemical OR processing OR productivity OR purif* OR raw material* OR recover* OR semisolid OR solid state fermentation OR spore OR sporulat* OR submerged OR synthe* OR temperature OR worker exposure OR yield.*

- For toxicology relation (Keywords Group 4: TOXICOLOGY):

abiotic OR absorbed dose OR absorption OR action OR action level OR activation OR active ingredient OR active metabolite OR acute toxicity OR administration OR allergen OR allergy OR antago OR antidote OR carcinogen* OR chronic toxicity OR concentration OR critical effect OR cumulative effect OR adverse effect* OR harmful effects OR cytotoxicity OR dose OR dose-response OR epidemiology OR genotoxicity OR reprotoxic OR hepatotox* OR immunotox* OR nephrotox* OR neurotox* OR incidence OR ingestion OR inhalation OR intake OR intestinal immunity OR morbidity OR mortality OR mutagen* OR worker exposure OR prevalence OR sensitization OR source of contamination OR sub-chronic toxicity OR susceptibility OR reproductive toxicity OR tolerance OR toxicolog* OR toxicity OR toxicity assay OR toxicity testing OR toxicological profile OR toxicokinetic* OR toxigenic OR toxigenic potential OR risk assessment OR safety OR safety assessment OR developmental toxicity OR hypersensitivity.*

- For biosynthetic pathways (Keywords Group 5: BIOSYNTHETIC PATHWAYS):

precursor OR enzyme OR synthesis OR gene OR expression OR structure OR pathway OR molecular OR biosynthesis OR promoter OR sequence* OR DNA OR cloning OR encode* OR biosynthetic OR route OR regulation OR biochemical OR biogenesis OR coenzyme OR molecule OR cofactor.*

2.2.2. Automatic searching tool – Web Crawler

The web crawler is integrated in an “ad hoc” platform called iWatch that allows the automatic search of scientific data on structured web sources by providing a tool for web scrapping (Web Crawler), parameterized with the search strategy explained in the previous section. For this purpose, the tool was configured in order to allow searching from the proposed databases and web sources (see Appendix B) applying a matching algorithm that filters and classifies the articles within the different lists. To this end, as required by the strategy, the algorithm checks title and abstract looking for any of the keywords depending on the microorganism type.

For all the articles searched (which included at least one of the microorganisms or synonyms considered for this tender), the Web Crawler searched on title or abstract the required keywords depending on its group. The following figure displays the Web Crawler architecture where experts specified both the keywords and the search strategy:

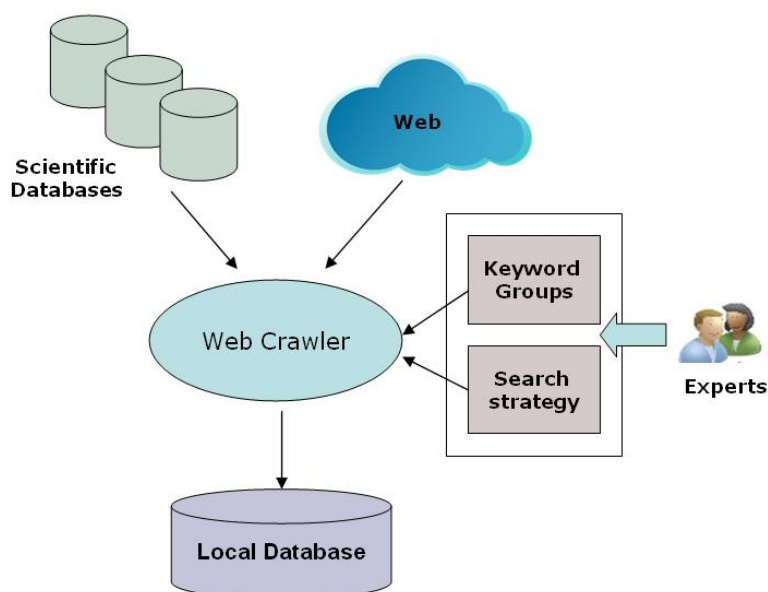


Figure 4: Automatic data collection architecture.

The result of the automatic search was stored in the iWatch database as a list of articles tagged by the microorganism and classified by the matched keywords, and was accessible using the iWatch user interface

In order to avoid double counting of studies, the scientific documents were compared for title, author names and publication area in which the study was conducted. The duplicates were discarded. This was done automatically by iWatch.

In addition to the review undertaken by the Web Crawler tool, a complementary manual literature search was carried out using the methodology explained above to search the sources that do not allow automatic searching

2.2.3. Searching sources

The search was conducted through a comprehensive list of searching engines, scientific web pages and other sources containing technical data.

For the taxonomic search (providing data for the review objective 1), an extensive search on the current taxonomy and nomenclature of microorganisms was done by using several websites and documents (Lapage *et al.*, 1992; Parker *et al.*, 2015). In the case of bacteria, a comprehensive and up-to-date presentation of the current taxonomy and nomenclature was obtained from LPSN (List of Prokaryotic names with Standing in Nomenclature, formerly List of Bacterial names with Standing in Nomenclature (LBSN)) (<http://www.bacterio.net>) and the list of validated prokaryotic names (IJSEM validated lists; <http://www.dsmz.de/bacterial-diversity/prokaryotic-nomenclature-up-to-date.html>) which contains the validly published names of bacteria compiled until September 2016. For fungi, the current taxonomic status and synonymy was obtained by using the online fungal nomenclature database Mycobank (<http://www.mycobank.org>) (Robert *et al.* 2013) and the International Code of Nomenclature for algae, fungi, and plants (ICN) (<http://www.iapt-taxon.org>) (McNeill *et al.* 2011). In both cases, to search for conserved genes, National Center of Biotechnology Information (NCBI) was used and complemented by a literature search using the terms "housekeeping" or "conserved genes" and the name of the microorganisms.

For the rest of the review objectives, the Table 1 shows the different categories of sources that have been used in the project. Additionally to this table, Appendix B includes the details of web sites that are configured in Web Crawler. The sources that do not allow data crawling have been searched manually.

In the case of the review objective 5, the databases KEGG (<http://www.genome.jp>) and MetaCyc (<https://metacyc.org>) containing metabolic pathways have been used to provide graphical representations.

Table 1: Searching engines and web sites that have been used as searching sources in the review. The web sites marked with an asterisk are automatically searched by iWatch.

| Searching Engines and sites for Scientific documents and reports |
|---|
| Web of Science*, Scopus*, PubMed*, ScienceDirect*, Embase, EspaceNet*, Medline, WorldWideScience, SpringerLink, Directory of Open Access Journals*, BASE: Bielefeld Academic Search Engine, SciELO, Science.gov High wire; Agencia EFE*; Agrodigital*; Agromeat-Agronews*; Albeitar*; Baden-Wuerttemberg*; CIDRAP*; ClubDarwin *; CODEX Alimentarius*; EDIPORC*; EFEAgro*; Food Business Review*; Harvard School Public Health*; Natural News*; New Scientist*; Nutraceutical Business Review* ;Nutraingredients*; RSSL Food & News*; Science Insider*; Sience Daily*; SINC; UNILEVER*;WIPO*. |
| Competent authorities, Food policy makers, Government databases, Food safety agencies, EU and Internationals Scientific Committees for Food Safety |
| European Commission*, FoodSafety (USA)*; FSA (UK)*; Health and Safety executive (UK)*; Health Protection Agency (UK)*; AHVLA (UK)*; ECDC, CDC (USA)* USDA(USA)*; USEPA(USA)*; SCIRI-AESAN (Spain); Health Canada*, AGES (Austria); AFSCA (Belgium); Ministry of Food, Agriculture & Fisheries (Denmark); National Food Agency Finland; ANSES (France)*; Bundesministerium für Ernährung, Landwirtschaft und Verbraucherschutz (Germany); Hellenic Food Authority (Greece); FSAI (Ireland)*; Livsmedelsverket (Sweden); VWA (The Netherlands); Norwegian Scientific Committee for Food Safety; CFIA Canadian Food Inspection*; FAO*; Risk Experts (AESAs; FDA*; SICURA Net), CSPINET (Center for Science in the Public interest), RASFF, FSANZ (Food Standard Australia New Zealand)*; BFR*. |
| Medical and Veterinary Databases |
| Environmental Health News *; Environmental Health Perspectives*; Infection Control Horizon Scanning*; OIE*; PigProgress*; WHO*; CABI (CAB Abstracts); EBSCOHost Databases with consultation of Food Science Source and Agricola database (National Agricultural Library); Clinical Evidence by BMJ; BMJ Learning; The New England Journal of Medicine; American Society of Microbiology (ASM); BioMedCentral; CAB Abstracts, FishBase (WorldFish Center). |
| Food industries information, food industry associations, trade organizations and Consultative Platform |
| Food Navigator (USA)*; Food quality news*; European Food Institutes (EFI), Spanish Agency for Consumer Affairs, Food Safety and Nutrition (AECOSAN); Spanish Agency of Medicines and Medical Devices (AEMPS), Bureau Européen des Unions de Consommateurs (BEUC); European Liaison Committee for Agricultural and Agri-Food Trade (CELCAA); Confederation of the Food and Drink Industries in the EU (CIAA); European Farmers - European Agri-Cooperatives (COPA-COGECA); European Federation of the Food,; European Public Health Alliance (EPHA); European Food Information Council (EUFIC); Eurogroup for animals; EuropaBIO; European Feed Manufacturers Federation (FEFAC); EU Association of Speciality Feed Ingredients and their Mixtures (FEFANA); Freshfel Europe. |
| Specific database on chemicals toxicity |
| Data bases on carcinogenic hazard characterization from the IARC*, International Agency for Research on Cancer (classification and monographs) or the NTP, U.S. National Toxicology Program (Studies and Report on Carcinogens); Developmental and Reproductive Toxicology Database (DART) and Hazardous Substances Data Base (HSDB) from TOXNET*, the U.S. National Library of Medicine Toxicology Data Network; JECFA* and EFSA scientific reports and monographs on chemical risks; Chemical watch*. |

Grey literature

Including theses and dissertations (University websites and research non-published reports), conference proceedings, newsletters, reports, informal communications, translations, census data, technical reports, standards, patents (e.g. European Patent Office, US Patent and Trademark Office).

Scientific opinions, scientific reports, statements and guidance documents from EFSA

As specified by the open call, the search has covered a time span of at least the last 20 years, only reduced to the last 10 years in case that the amount of literature references retrieved for a given species was ≥ 500 . In addition, extra information on toxicity of compounds produced by microorganisms has been retrieved manually from a cluster of Toxnet databases: HSDB (Hazardous Substances Data Bank) and ChemIDplus for general toxicity data information; CTD (Comparative Toxicogenomics Database) for data describing relationships between chemicals, genes and human diseases; Haz-Map for occupational information; CCRIS (Chemical Carcinogenesis Research Information System), CPDB (Carcinogenic Potency Database) and GENE-TOX (Genetic Toxicology Data Bank) for genotoxicity and carcinogenicity tests. No date filter was applied for this manual search. The information obtained with this search complemented the findings of the searching strategy and is summarised in Appendix C.

2.3. Selection of studies

The selection of the scientific documents recovered in the search was done following the criteria explained below.

The relevant studies selected were:

- referring to the microorganism of interest as a producer of a secondary metabolite/toxic compound
- providing data on its taxonomical identifiers and potential toxigenic capacities. Studies providing data on toxicity of compounds that have already been assessed by EFSA have been included in the database but extraction of data from the toxicological point of view has been considered not necessary.

The selection and extraction process has been conducted in two stages: Level 1 and Level 2.

Level 1: Screening of titles and abstracts: Inclusion/exclusion criteria were applied after reading either in the title or abstract of the documents retrieved at each stage of the searching (List 1-List 3 and List 5-List 7). Examination of title and abstracts of documents included in list 4 permitted the identification of the secondary metabolites produced by the microorganisms that might have a potential toxic effect. Some documents from List 4 were also included directly in Lists 5, 6 or 7, depending on the type of information provided. After having identified the potentially toxic metabolites from List 4, a second search was carried out with the corresponding keywords (see Figure 3), to obtain documents for Lists 5, 6 and 7. The title and abstract of these documents were read and the corresponding inclusion/exclusion criteria (see Table 2) applied to select the documents for Lists 8, 9 and 10, respectively.

Level 2: Full examination of documents: The full text of reports from Lists 8, 9 and 10 were examined at Level 2. Inclusion/exclusion criteria (see Table 3) were also applied at this stage, in order to select the documents that met all the conditions to extract the information for the database. The examination of the complete documents allowed in many cases to discard documents that passed the screening at Level 1, but that after a further examination were concluded as not containing relevant data. Documents containing any relevant information to provide data for the review objectives were used to extract information for the database. The quality of the information was assessed by the application of the inclusion/exclusion criteria.

For both levels, the lists of documents were evaluated by two independent reviewers (parallel review) from each area of specialization (microbiology, molecular biology and toxicology). In the case of disagreements between them, the scientific document was discussed to reach a consensus before proceeding to the next step of the screening process.

1.1.1. Screening of titles and abstracts for the relevance of the study questions: Level 1

The inclusion/exclusion criteria used to select relevant documents at title and abstract level are described in the following table (Table 2).

Table 2: Inclusion/Exclusion Criteria.

| | | |
|--------------------------------------|------------------|--|
| To be selected for List 4 | INCLUSION | Is the main aim of the document directly related to the microorganisms within the scope of the review? (Yes/No) |
| | | Is there a metabolite/substance produced by the microorganism regarded as toxic/causing adverse effect? (Yes/No/Unknown) |
| | EXCLUSION | Is the metabolite/substance produced by the microorganism included in the list of food enzymes & feed additives (Appendix D) ¹ ? (Yes/No) |
| | | Is the document related to the production of toxins or adverse effects outside the scope of the systematic search (antimicrobial, antifungal, responsible for the pathogenicity of the species)? |
| To be selected for List 8 | INCLUSION | Is the toxic metabolite/substance produced under specific conditions (process conditions detailed) described? (Yes/No/Unknown) |
| | EXCLUSION | Is the fermentation process not linked to the microorganism within the scope of the review? (Yes/No) |
| To be selected for List 9 | INCLUSION | Is the toxin/ secondary metabolite/substance evaluated from a toxicological point of view? (Yes/No) |
| | EXCLUSION | Is the toxicological study focused on ecotoxicology? (Yes/No) |
| | | Is the compound evaluated an undesirable substance in animal feed (Dir 2002/32/EC)? (Appendix E) (Yes/No) |
| | | Has the substance already been evaluated by IARC (Monographs on the Evaluation of Carcinogenic Risks to humans), NTP (Report on Carcinogens) or EFSA (Appendix E)? (Yes/No) |
| To be selected for List 10 | INCLUSION | Does the document deal with the biochemistry or biosynthetic pathway of the toxic compound? (Yes/No/Unknown) |
| | EXCLUSION | Is the biosynthesis of the toxic compound not linked to the microorganism within the scope of the review? (Yes/No) |

¹List of food enzymes & feed additives provided in Appendix D is used as exclusion criteria. If the substance regarded as toxic is included in the list, the document is considered ineligible, because the scope of the review does not include an assessment of the toxicological aspects of the food enzyme or additive itself but other not intentionally produced substances that could occur during the fermentation.

The screening consisted of several steps corresponding to four lists of questions (Table 2). It is defined that if any inclusion criteria was not fulfilled (answered NO) it was not necessary to proceed with the screening and the document was considered ineligible (all inclusion criteria must be answered YES or UNKNOWN to be included in study). On the other hand, if any exclusion criteria was answered YES the document was also discarded (exclusion criteria must be answered NO or UNKNOWN to be included in the study).

In the majority of the cases, the screening did not identify precise information concerning specific eligibility criteria regarding the toxicity of the compound; therefore the answer was inconclusive (UNKNOWN) that led to the inclusion of the document. The document was selected and the secondary compounds mentioned in these scientific documents recorded in a form "ITEM ENTRY" (see Section 2.4.). From all the compounds initially recorded, the ones selected for the second search had to comply with at least one of these conditions:

- 1) The compound is referred as a toxin in the scientific document or is a known toxin or mycotoxin or is involved in the biosynthetic route of one toxin.
- 2) The compound has been found to be produced by more than one species.
- 3) More than one reference has been found indicating the production of the compound by one species.

2.3.1. Eligibility criteria for full text documents: Level 2

Documents that potentially met eligibility criteria at the abstract and title screening stage were retrieved (Lists 8-10) and the full text documents were reviewed for eligibility. The criteria for inclusion or rejection of full text documents are recorded in the following table (Table 3).

Table 3: Inclusion/Exclusion Criteria.

| | |
|------------------|---|
| INCLUSION | Is the toxin compound produced well identified/defined? (Yes/No) |
| | Are the experimental designs/protocols/material and methods reported? (Yes/No) |
| EXCLUSION | Does the document not report the designated outcomes: process conditions or toxicology or metabolic pathway of the toxic compound? (Yes/No) |
| | Is the document a preliminary toxicological assessment /toxicity is not clear or related to potential medical properties of the substance? (Yes/No) |
| | Is the occurrence of the toxic compound doubtful or anecdotic? (Yes/No) |

The reasons for rejection of full text articles are justified and recorded in Appendix F of the report, together with their references.

Compounds of potential concern but with no standard toxicity information to be included in the final database (*e.g.* a very preliminary toxicity assessment, study of interaction with a molecular target, etc) are reported in Appendix G. They were found at the first level of the reviewing process, to obtain the name of the secondary metabolites potentially toxic that might be produced by the different species. They were identified whether in the abstract or when reviewing the complete document at level 2.

2.3.2. Collecting data from the included studies and creating evidence tables

As explained before, all retrieved articles in Lists 8, 9 and 10 were submitted to the Level 2 review, conducted by examining the full-text of the articles. At this point, the quality of the paper was assessed by applying the inclusion/exclusion criteria (Table 3).

The assessment of full-text documents was carried out also by 2 different reviewers. Data extracted from each document was verified by a second person (sequential method). In case of disagreement, the document was discussed between the different reviewers involved in the screening to reach a

consensus. In every case possible the extraction of the data was aided by the collaboration platform iWatch as explained below.

Collaboration platform - iWatch

The interface provided by the iWatch Web platform eases the work of filtering, reviewing and extracting the data required by the tender as specified in the methodology. The platform provides the functionality required to manage the outcomes of the search strategy, allowing for the storage of the manual search results and the management of the automatic search results.

Regarding the collaboration tasks, the tool has allowed the following functionalities:

- Filtering of documents by the keywords groups used in the searching strategy, allowing for the distribution of the work within the experts groups.
- Creation of classifying folders in order to facilitate the reviewing cycle (experts should be able to review documents two times).
- Executing batch actions with a group of entries to allow selecting a group of entries and executing the same action with all of them.
- Searching documents by words within a defined list to facilitate the application of inclusion/exclusion criteria (batch actions allow removing or adding tags to identify them).
- Addition of tags to identify documents within a topic.
- Discard non useful documents or addition of documents found by manual search.
- Inclusion of comments into each entry to save important information found.

Also, **iWatch** tool was configured to facilitate the work of data extraction by allowing entering data through different forms. These forms were designed after the information required for the data model which included text and numerical fields and drop-down lists when required to select options for EFSA collections. Finally, the submissions sent by the users were also stored at the platform database.

The following forms for data registration were created:

- **Item Entry:** internal entry form that allows introducing and collecting possible toxic compounds of interest extracted from List 4. It includes information about the source, the microorganism and toxic compound. This information was used to obtain Appendix G.
- **Fact Entry:** entry form to include information extracted from selected articles of interest from Lists 8, 9 and 10, which will be included into the database. It includes information about the source, the microorganism, biosynthetic pathways, process conditions and toxicology.

To extract the data submitted by users, iWatch export module was configured in order to allow the processing of all entries and its storage in a Data Schema created for this project as defined in section 2.6. For each entry, the module processes all fields converting to the format required for the data base specifications when required (see Appendix H) and stores the result in the schema. Once stored, the Data Base Management System allowed the data extraction in the XLM format required by EFSA.

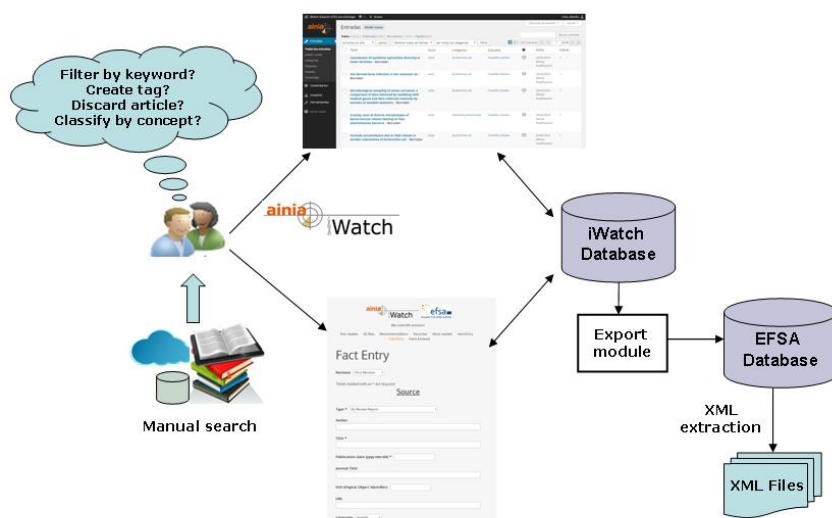


Figure 5: Collaboration Platform Architecture.

2.4. Assessing the validity and quality of the included studies

The overall development of this review was carried out under a framework of quality assurance system and under the supervision of a Quality Assurance Unit.

The main feature to control the quality of the review and the data extracted was a double check review done by two experts for each area of knowledge (microbiology, molecular biology and toxicology).

Moreover, and specifically for this project, a number of control mechanisms at the data entry level were implemented to guarantee high quality of deliverables and reduce unintended errors:

- Sharing the preliminary results with the requestor in order to obtain an external validation of the work that has already been done.
- Expert groups meetings to review the methodologies and conclusions.
- Furthermore, an external quality check was implemented by:
 - Automatic verification of data quality at the data entry level.
 - Manual revision of the collected data to be submitted.

Automatic verification entails the definition and implementation of rules aiming to minimise errors during data entry, thus maximizing data quality. In the data entry phase, errors that can be detected automatically are: type mismatches, missing values for mandatory fields, wrong format (e.g., dates), and length exceeding the maximum allowed size.

For these structural errors the nature itself of a Database Management System (DBMS) engine such as MySQL can prevent to store wrong data. In fact, at the database creation phase, it was specified for each fields its type, length, and format and if it is a mandatory field. In this way the DBMS will not allow storage of data that is not compliant with the corresponding field definition. In addition, more complex rules can be specified to the database engine using the SQL language, such as foreign keys. For catalogue fields, namely fields that can assume only a value chosen within a predefined list, it is possible to define catalogue tables and refer to them in the proper fields through the creation of foreign keys. A web-interface guides and supports the operator in the data entry phase.

2.5. Data model

A relational database (Appendix H) was designed to store the extracted information, following a relational methodology based in an iterative process, in which EFSA provided a framework and advice for the database structure. The database shares the same structure and some catalogues already built for other EFSA's databases, in particular the *Chemical hazards database* created to hold a summary hazard data from EFSA's chemical risk assessments in food and feed. This data collection covers the opinions and statements adopted by EFSA panels including FEEDAP Panel (Additives and Products or Substances used in Animal Feed), CEF Panel (Food Contact Materials, Enzymes, Flavourings and Processing Aids), ANS Panel (Food Additives and Nutrient Sources Added to Food) and NDA Panel (Dietetic Products, Nutrition and Allergies). The catalogues used to include some of the data in the database are available in EFSA's Data Collection Framework. All the suitable new terms encountered in this review that were not included in the catalogues have been introduced by sending the pertinent information to the EFSA DCF Data Manager.

The starting point for this database is the harmful substance or toxin produced by the microorganism as a secondary metabolite during the fermentative process. This substance is always going to be associated to the **PARAM substance definition** as provided in the standard sample description (EFSA, 2013). If a substance was not included in the EFSA PARAM catalogue, such substance was introduced as explained previously. The information to be provided to the EFSA DCF Data Manager for these substances is included in Appendix I and contains the following:

CAS Number: The Chemical Abstract Service registration number of the substance. The CAS number can be retrieved from publicly available databases such as: PubChem, ChemIDPlus, etc.

Molecular formula: The molecular formula is reported. The molecular formula can be retrieved from publicly available databases in particular PubChem (Bolton, 2008).

SMILES notation: The SMILES (Simplified Molecular Input Line Entry Specification) for the substance. The coded structure can be retrieved from publicly available databases such as: PubChem, ChemIDPlus. The structure code can be also obtained by manually drawing the structure in popular chemical drawing programs that allow for the SMILES generation such as ChemSpider. When this option is used, verification of the structures codified will be done. Together with the SMILES, the source of this notation will be also included.

Figure 6 shows the logical map that represents the intrinsic properties of the data to be structured in the database as extracted from this scientific review and describes the following tables of keywords:

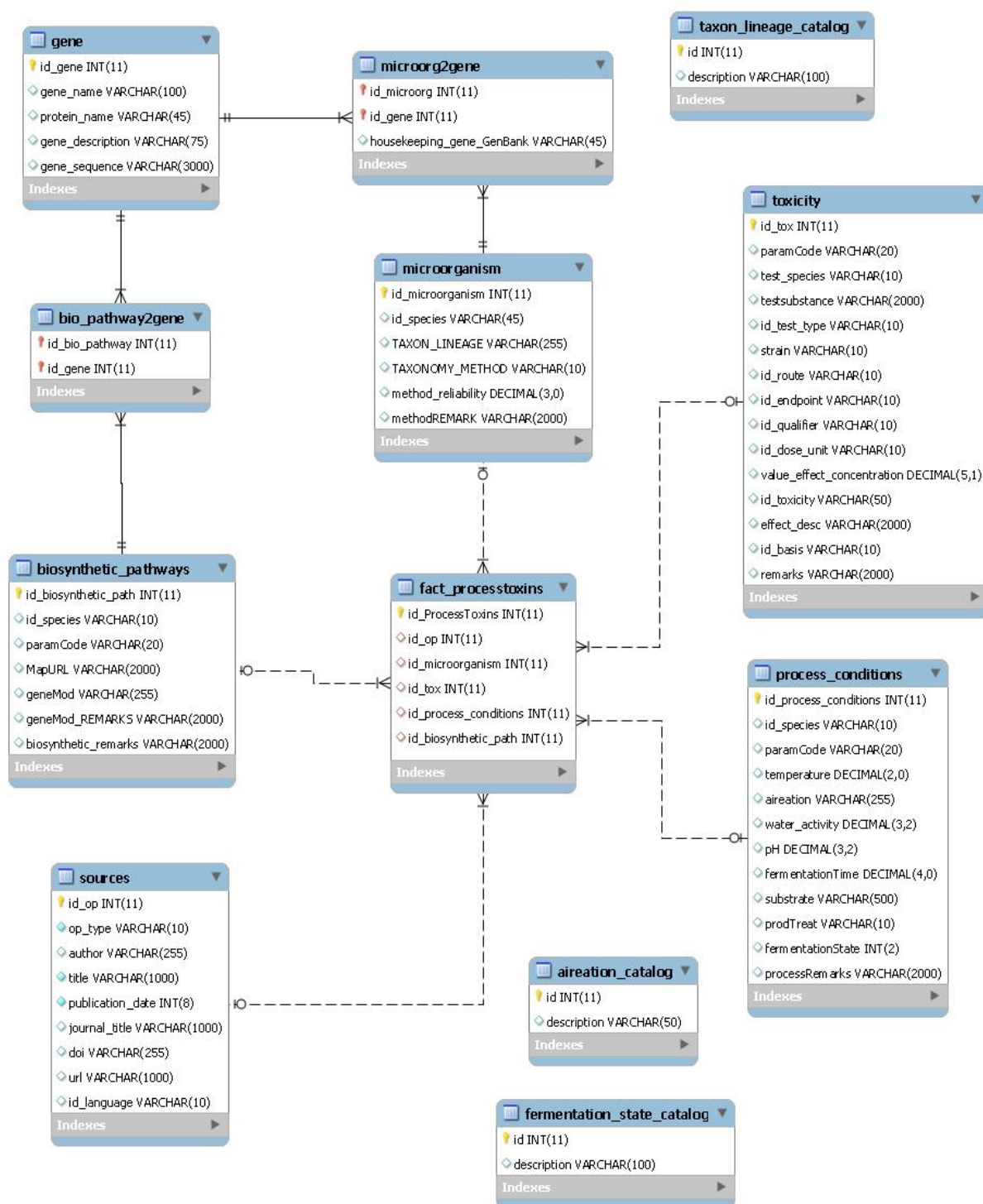


Figure 6: Data model for the review.

FACT_PROCESSTOXIN: Codification of each fact that is described in the database.

MICROORGANISM: Descriptors of the microorganism associated to the facts, normally one of the microbial species considered in the scope of this review.

BIOSYNTHETIC PATHWAYS: Descriptors of the metabolic route used by the microorganism for the synthesis of the toxic compounds associated to the facts.

PROCESSING CONDITIONS: Descriptors of the processing conditions in which the microorganism is able to synthesize the toxic compounds associated to the facts.

TOXICOLOGY: Descriptors of the toxicological parameters of the toxic compounds associated to the facts.

SOURCES: Descriptors of the document from which the information for the facts is obtained.

2.5.1. The sections of the data model – Description and general information

The descriptors chosen for each of the features (tables) forming the data model were the following (see also **Appendix H**; Excel file of the Data Model):

FACT_PROCESSTOXIN

- **id_ProcessToxins:** Unique ID for fact_processtoxin table.
- **id_op (FK):** Foreign key (FK) with a unique ID from the source table.
- **id_microorganism (FK):** Foreign key with a unique ID from the microorganism table.
- **id_tox (FK):** Foreign key with the unique ID from the toxicology data.
- **id_process conditions (FK):** Foreign key with a the unique ID from the process conditions table.
- **id_biosynthetic path (FK):** Foreign key with a the unique ID from the biosynthetic pathway table.

MICROORGANISM

- **id_microorganism (PK):** Primary key (PK) with a unique ID for the microorganism table.
- **id_species:** Microorganism used in the biotechnological process. The data type is a code from the **MTX** catalogue. MTX catalogue is filled with new entries of the microorganisms of this review that are not included yet. Data to be provided is:
 - a complete identification kingdom-order-family
 - the more recent recognized name
 - possible synonyms
- **TAXON_LINEAGE:** (picklist) General classification of the microorganism (filamentous fungi, yeast or bacteria) which provide information useful for the safety assessment.
- **TAXONOMY_METHOD:** Acceptable scientific methodologies and techniques for the identification of microorganisms. This data type is a code from the **ANLYMD** catalogue.
- **method_reliability:** (number) Measure of the consistency of the taxonomic method (% Homology/Closetest given in the molecular studies).
- **methodREMARKS:** (free text) Include the reference organism for the comparison and the homology threshold considered.

SOURCES

- **id_op (PK):** Unique ID for fact_processtoxin table.

- **op_TYPE:** Type of document (picklist): whether the document is an scientific article, report, book, thesis, EFSA opinion or statement etc. This field will be using one of the terms in the EFSA's REF-TYPE catalogue.
- **Author:** (free text) Author(s) of the journal article.
- **TITLE:** Title of the journal article or document (title, free text): the title of the document as reported in the published document (PDF file) is reported.
- **PUBLICATION_DATE:** (number) Date of the publication the publication. The format is yyyy-mm-dd.
- **JOURNAL_TITLE:** (free text) Title of the journal where has been published the scientific article.
- **DOI:** (free text) Digital Object Identifier of the journal article.
- **URL:** (free text) The journal link (internet address) or publisher.
- **id_LANGUAGE:** (picklist) Language used to fill in the free text fields (ISO-639-1) and **LANG** catalogue.

BIOSYNTHETIC PATHWAY

- **id_biosynthetic_path:** Identification of the synthesis pathway of the metabolite.
- **id_species:** Microorganism used in the biotechnological process. The data type is a code from the **MTX** catalogue.
- **paramCode:** Toxic compound described according to the Substance Code from **PARAM** catalogue.
- **MapURL:** Link to the diagram where the route of synthesis of the substance is represented.
- **geneMod:** (picklist) Genetic modification, where the gen has been genetically modified or not.
- **geneMod_REMARKS:** (free text) Genetic modification (information of genes inserted or deleted). It can be used to comment on the result of gene modification.
- **Biosynthetic_REMARKS:** (free text) Details of the biosynthetic pathway (promoters, etc).

PROCESS CONDITIONS

- **id_process conditions:** The data elements belonging to this section describe information related to the process conditions.
- **id_species:** Microorganism used in the biotechnological process. The data type is a code from the **MTX** catalogue.
- **temperature: (number)** Temperature in degrees centigrade.
- **aireation: (picklist)** Oxygen requirements for the microorganism during the fermentative process (*Aerobic*: $\geq 20\%$ oxygen; *Anaerobic*: $< 5\%$ oxygen; *Microaerophilic*: $20\% >$ oxygen $\geq 5\%$)
- **WaterActivity: (number)** Water activity, as it is expressed with numerical number from the range (0-1).
- **Substrate: (free text)** Type of substrate used in the biotechnological process. It makes reference to the substrate used in the fermentative process (Main protein source (yeast extract), polysaccharides (molasses), etc.)

- **pH: (number)** pH during the fermentative process, as it is expressed with numerical number from the range (0-14).
- **Fermentation time (h):** Time in hours of the fermentative process.
- **prodTreat:** Used to characterise a food product based on the treatment or processes applied to the product. This data will be obtained from the **PRODTR catalogue** catalogue.
- **fermentationState: (picklist)** Types of fermentative processes: *solid state* (microbial growth and product formation occur at the surface of solid substrates); *semi-solid* (this type of fermentation is a sort of SSF in which the free liquid content has been increased in order to facilitate nutrient availability and fermentation control), *submerged batch* (closed culture system which contains limited amount of nutrient medium); *submerged fed-batch* (batch culture to which is added new fresh nutrient medium without removing the growing microbial culture), *submerged continuous* (a culture in which nutrients are supplied and end products are continuously removed), *immobilized cell bioreactors* (bioreactors based on immobilized cells), *immobilized enzyme bioreactors* (continuous flow reactors based on immobilized enzymes).
- **paramCode: (free text)** Its data type is a code from the **PARAM** catalogue. It is used to describe the toxic substance aim of the study.
- **processRemarks: (free text)** Additional remarks on process conditions. Its data type is free text of up to 2000 characters. It can be used to comment on the process analysed.

TOXICITY

- **id_tox:** The data elements belonging to this section describe information related to toxicology.
- **paramCode: (free text)** Its data type is a code from the **PARAM** catalogue. It is used to describe the toxic substance aim of the study.
- **test_species:** Species of the organism/cell culture used as the test organism in the toxicological study
- **testsubstance:** Description of test material used in the toxicological study.
- **id_test_type:** Type of toxicological test. Its data type is a code from the **TEST_TYPE** catalogue.
- **Strain:** Identification of the strain used as the test organism.
- **id_route:** Indicator how the substance is administered to the organism (human/animals). Its data type is a code from the **ROUTE_EXP** catalogue.
- **id_endpoint:** Endpoint reported in the study to describe the reported values (e.g. dose level). The reference point is described in the EFSA documents and used to derive health-based guidance values, margin of exposure values for human health, margin of safety values for human or animal health. The opinion may also discuss a reference point or toxicity values for animal health. This data type is a code from the **ENDPOINT_HGV** catalogue.
- **id_qualifier:** Qualifier for the reported endpoint values (e.g. =, <=, >=). Its data type is a code from the **QUALIFIER** catalogue.
- **id_dose-unit:** Enumeration of group units for group assessment. Its data type is a code from the **UNIT** catalogue.
- **Value_effect_concentration:** Effect concentration.

- **id_toxicity:** This field refers to classification of critical effect according to toxicity target (Owens 2002) and the data type is a code from the **TOXICITY** catalogue.
- **Effect_desc:** (Free text) Description of the effects observed in the toxicological study.
- **id_basis:** Characterisation of the measures toxicological outcome measure. Its data type is a code from the **BASIC_EFFECT** catalogue.
- **Remarks:** Additional remarks on toxicological study

GENE

- **id_gene:** The data elements belonging to this section describe information related to the gene.
- **Gene_name:** Gene name involved in the synthesis of the substance.
- **Protein_name:** Name of protein synthesised from the gene.
- **Gene_description:** (Free text) Description of the gene function.
- **Gene_sequence:** Sequence of the involved in the synthesis route.

BIO_PATHWAY2GENE

- **id_bio_pathway:** The data elements belonging to this section describe information related to the identification of the biosynthetic pathway.
- **id_gene:** The data elements belonging to this section describe information related to the genes involved in the biosynthesis of the compound.

MICROORG2GENE

- **id_microorg:** The data elements belonging to this section describe information related to the taxonomy of the microorganism.
- **id_gene:** The data elements belonging to this section describe information related to the genes involved in the microorganism identification.
- **housekeeping_gene_GenBank: GenBank ID for the housekeeping gene**

3. Results

3.1. Taxonomic characterisation

A literature search was carried out for each microorganism included in the scope of the project with the purpose of assessing the available scientific information on the taxonomical characterisation (see section 2.2.3.). A list of microorganisms including information about its valid current name and the taxonomic ranks (kingdom, order, family), as well as other scientific names and synonyms, was prepared and sent to EFSA for the inclusion in the MTX catalogue.

The literature search on taxonomy data produced 123 references with the methods for its relevant and accurate taxonomical identifiers, genomes sequenced and housekeeping genes (see Appendix J, and Appendix K).

Bacterial taxonomy determination is mainly based on several molecular techniques, each one for retrieving the information at different levels (proteins, fatty acids, DNA) and the obtained results are combined and analysed to reach the correct taxonomic identification of the microorganism.

The taxonomy of fungi is currently very complex. There is still an ongoing need of unique identifiers harmonisation, in this regards, big data tools contribute to harmonise approaches (three official

registries are approved: MycoBank, Index Fungorum, Fungal Names). However, debate on species concepts in the mycological society results in a lack of a universally accepted fungal taxonomy.

Several studies have been dedicated to explore high-throughput expression profiles, in order to identify candidate housekeeping genes. The 16S ribosomal RNA (16S rRNA) gene sequence to study bacterial phylogeny and taxonomy has been universally accepted as the common housekeeping genetic marker. A commonly used reference gene for fungi is the 18S ribosomal RNA (18S rRNA). The accession numbers to the Genbank database for the sequences of the conserved genes found in the search is reported in Appendix J (Appendix J Bacteria, Appendix J Fungi). The appendix also includes the links to the available ID genome annotations for the type species of the microorganism when available. Those links allow downloading sequence and annotations for current versions of genome assemblies from the NCBI genomes FTP site.

The results of the taxonomic review were integrated into the database, and synonyms were also included as new keywords for the searching strategy.

3.2. Retrieval of relevant documents

According to the searching strategy explained in Section 2.2, IWatch tool was set up and configured for the retrieval of scientific documents that match with the different sets of keywords and non-searchable databases were manually searched in order to complement the search. The total number of scientific documents retrieved for the first level was 22970. The references were distributed into several groups using the microorganism name as a keyword.

The election of the articles for the revision by the two reviewers was also done by the IWatch tool that displayed the abstract and allowed direct access to the URL of the document. The reviewers labeled the articles as "first" or "second" revision and as "included" or "excluded" which facilitated the management of the review.

After having examined the titles and abstracts of Lists 1, 2 and 3 (22970 documents in total) and applying the inclusion/exclusion criteria, 411 documents (List 4) were selected. The revision of the articles allowed the identification of 474 bioactive secondary metabolites synthesised by the different microorganisms of the scope that were subjected to a further selection according to the criteria explained in section 2.3.1. Figure 7 shows the outcome of each searching and screening stage conducted in this project.

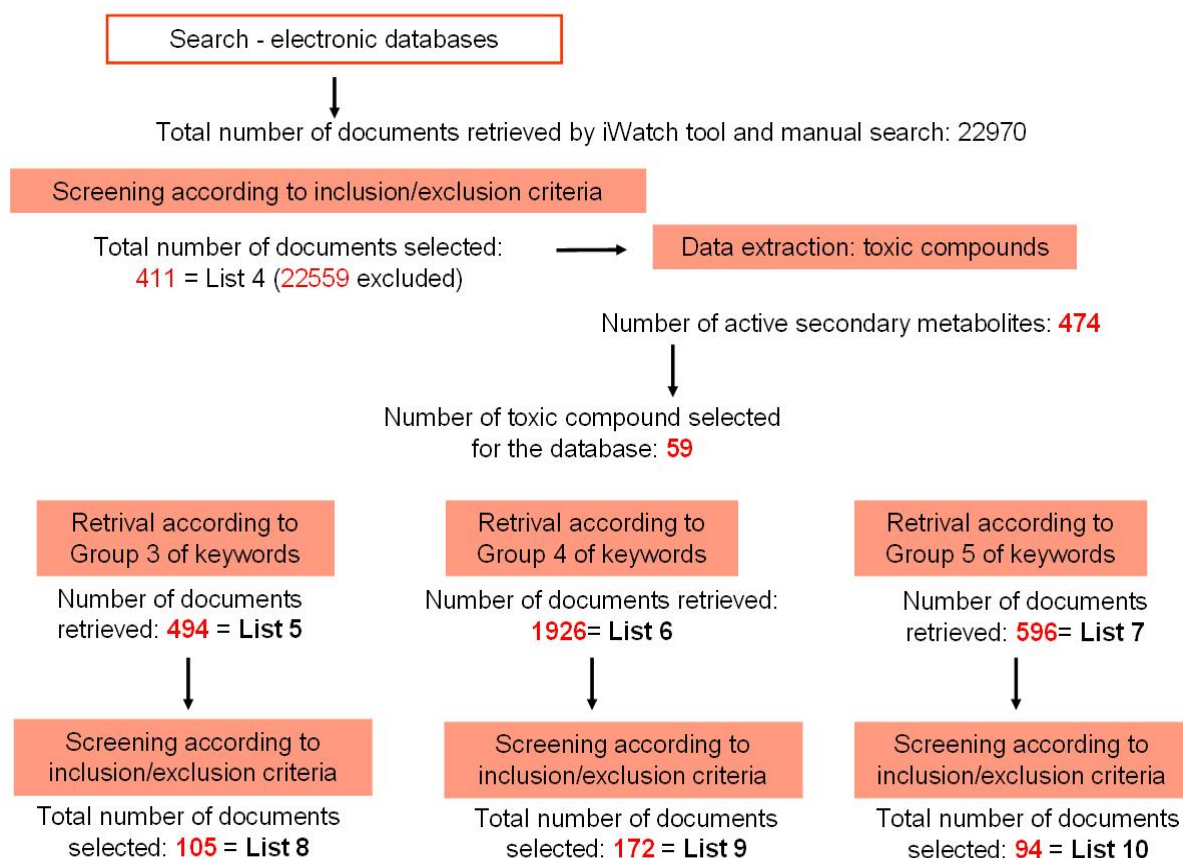


Figure 7: Flow-chart summarising results of the literature search and retrieval of relevant documents.

For the second search, aimed to obtain pertinent information about the toxicity characteristics of the compounds, biosynthetic pathways and fermentation conditions, 59 compounds (see below) were selected from the total number of bioactive metabolites identified, based on the data available about the toxicity or potential toxicity of these compounds or the role they may have on the biosynthetic routes, as explained in section 2.3.1. From these 59 compounds selected, 18 were mycotoxins or groups of mycotoxins that have already been characterised for their toxicological impact. Those toxins are (Appendix E):

Aflatoxin B1, Aflatoxin B2, Beauvericin, Citrinin, Deoxynivalenol, Enniatins, Ergot alkaloids, Fumonisin B1, Fumonisin B2, Fumonisin B4, Fusarin C, Ochratoxin A, Patulin, Penicillic acid, Sterigmatocystin, T-2 toxin, Trichothecenes, Zearalenone.

Another 41 compounds were toxins not yet fully characterised from the toxicological point of view:

1-Hydroxyphenazine, 3-Nitropropionic acid, Andrastin A, Aristolochene, Asperparaline A, Citreoviridin, Culmorin, Cyclophenol, Cyclopiazonic acid, Cytolysin A, Diacetoxyscirpenol, Endophenazine A, Festuclavine, Isofumigaclavine A, Isofumigaclavine B, Isotrichodermin, Isotricodermol, Kojic acid, LPS endotoxin, Malformin(s), Maltoryzine, Marcfortine(s), Mitorubrinic acid, Mycophenolic acid, Naphto-gamma-pyrones, Neoxaline, Nigerazines, Nigragillin, Orlandin, Oxalic acid, Perinadine A, Phenazine-1-carboxylic acid, Phenazines, PR-toxin, Pyocianin, Roquefortin, Sambucinol, Secalonic acid, Trichodermin, Trichorzin, Xanthomegnin.

In the second search to obtain List 5 and List 7, each of the 59 compounds were used as keywords, combined with the name of the microorganism(s) that produced the compound and the keywords

groups 3 and 5 to obtain the List 5 and List 7. List 6 was obtained for the compounds not included in Appendix E, as mentioned in Section 2.2.

Additional information regarding the molecular characterisation of these compounds (Molecular formula, CAS number, SMILES, INCHI and IUPAC) was also obtained for those compounds not included in the EFSA catalogues (Appendix I).

The URL with the toxicological information retrieved for each compound (as explained in section 2.2.3) was provided as supplementary data (Appendix C)¹.

3.3. Production of toxic secondary metabolites by microorganisms used in industrial fermentations

In this review, it has been found that many of the species of microorganisms studied produce a wide range of secondary metabolites, in addition to the substances that are intentionally produced for industrial purposes. Several of these secondary metabolites have been described as toxic to humans and animals.

In many cases, the metabolic role of these compounds is unknown and/or not well defined; however, biotechnological or biopharmaceutical interest in these compounds is considerable, as many of these substances have been studied for application in medical, industrial and/or agricultural areas. The benefits of these compounds as natural sources for pesticides, pigments, plant growth regulators, biocontrol agents, antitumor or antimicrobial products are in continuous research. There are still not enough concluding data about the possible adverse effects that these compounds could pose to mammals or humans, when unintentionally exposed to them through the diet or the environment. However, the unintentional presence of these substances in feed or food as contaminants from the fermentation product could raise safety concerns. The absence of toxicological information does not mean absence of toxicity, and thus more research will be needed for the toxicological characterization of these compounds.

On the other hand, some of the substances found to be produced by fungi such as *Aspergillus niger*, *A. oryzae* or *Fusarium venenatum* are well recognised mycotoxins, some of them already present in the current legislative frameworks. In these cases it is essential to obtain data about the genes involved in the biosynthetic pathways of these substances, as well as the culture conditions in which they are produced, in order to select the adequate strain and fermentation conditions that impair the production of these toxins. In this sense, it is common that industrial strains are genetically modified to avoid the expression of the genes responsible for the biosynthesis of mycotoxins.

A list of the 474 bioactive secondary metabolites produced by the microorganisms within the scope of this project that have been found during the literature search is presented in Appendix G. The 59 compounds selected for further searching are marked in bold.

3.3.1. Toxic secondary metabolites produced by fungi

In the present review, several mycotoxins and other toxic compounds have been found associated to the fungal species within the scope of this work. The most important and relevant facts in relation to the production of these substances are described below.

3.3.1.1. Substances produced by fungi not characterised in depth from a toxicological point of view

Several secondary substances produced by species of fungi used in industrial fermentations have been reported as toxic or potentially toxic substances, but still are not officially recognized as toxins and

¹ The databases CCRIS, CPDB and GENE-TOX are no longer updated

few information about their biosynthesis, production and toxicological data on humans and animals is available. Some characteristics of these compounds are summarised below:

3-Nitropropionic acid

3-nitropropionic acid is a mycotoxin that inactivates succinate dehydrogenase thus inhibiting mitochondrial respiration. It is a neurotoxin that induces rat striatal degeneration; systemic intoxication by 3-nitropropionic acid induces selective lesions of the striatum in most species. It can be used as an experimental pharmacological model of Huntington disease in rodents. Thus, many articles related to the mechanism of action underlying striatal lesions have been found.

In this project, 3-nitropropionic acid has been identified as a secondary metabolite of *A. oryzae*. Acute and subacute effects (5-days) were investigated in adult male Wistar rats after receiving 3-nitropropionic acid doses by i. p. route (Szabó *et al.*, 2005). Several neurological alterations and a significant decrease of the thymus weight were observed in the treated rats.

In another study, 3-nitropropionic acid was given by i.p. route to C57Bl/6 adult male mice in two different dose regimes: a low-dose (total dose of 340 mg/kg in 7 days) or a high-dose (total dose of 560 mg/kg in 7 days) (Fernagut *et al.*, 2002). The main objective of the study was to carefully characterise the motor disorder, its time course and the histopathological correlations. The low-dose regimen was not lethal but the high-dose regimen produced 37.5% of lethality to mice.

The effects of 3-nitropropionic acid were evaluated in males of four different mouse strains: BALB/c, C57BL/6, FVB/n and 129SVEMS mice (Gabrielson *et al.*, 2001). It was found that 3-nitropropionic acid induced brain and heart lesions and that cardiac pathology had a different degree depending on the strains; the follow up investigation on the biochemical mechanisms of cardiac toxicity suggested that the inhibition of the succinate dehydrogenase in heart mitochondria was also the cause.

Finally, acute intoxication of male and female adult Wistar rats was induced by the subcutaneous injection of 3-nitropropionic acid (20 mg/kg) once a day for 2 or 3 days (Nishino *et al.*, 1997). Intoxication produced motor symptoms and striatum-specific lesions; it also irreversibly increased astrocytic $[Ca^{2+}]_i$. The possible mechanisms conducting to astrocyte destruction are discussed in the article.

Andrastin A

Andrastin A is a farnesyltransferase inhibitor isolate from *Penicillium* species including *P. roqueforti* (Rasmussen *et al.*, 2011). Andrastin A inhibits the activity of the ras protein farnesyltransferase, and therefore these meroterpenoids are promising leads for anticancer drugs, since the farnesylation of the ras oncogene protein is essential for its function (Uchida *et al.*, 1996).

The first identification of the biosynthetic gene cluster for andrastins is based on genome database of *P. chrysogenum*. The cluster contains eleven genes, and nine of them are likely to be directly involved in the biosynthesis of andrastin A, the most complex andrastin molecule (Uchida *et al.*, 1996).

The industrial strains of *P. roqueforti* showed high levels of aspartylprotease AspA, whereas the culture collection strain showed barely detectable levels of this enzyme, as shown by proteolysis tests and by immunodetection with anti-AspA antibodies. The lipolytic activity was similar in the strains isolated from the three types of local blue cheeses (Fernández-Bodega *et al.*, 2009).

Andrastin A was produced by all strains analysed at different levels. *P. roqueforti* CECT 2905 showed high ability to synthesize this compound. Andrastin A was present in all industrial and local varieties of blue cheese.

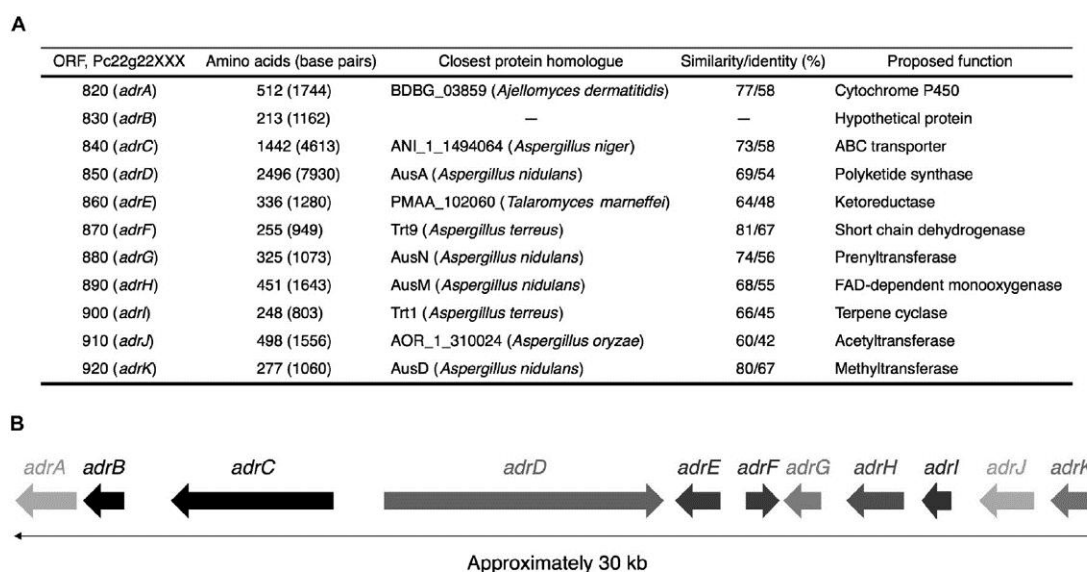


Figure 8: (A) Annotation of each protein in the *adr* gene cluster. Deduced function of each open reading frame (ORF) and amino acid sequence similarity/identity, compared with the closest homologues found by BLAST search at NCBI. (B) Schematic representation of the arrow indicates the direction from the start to the stop codon (Matsuda *et al.*, 2013).

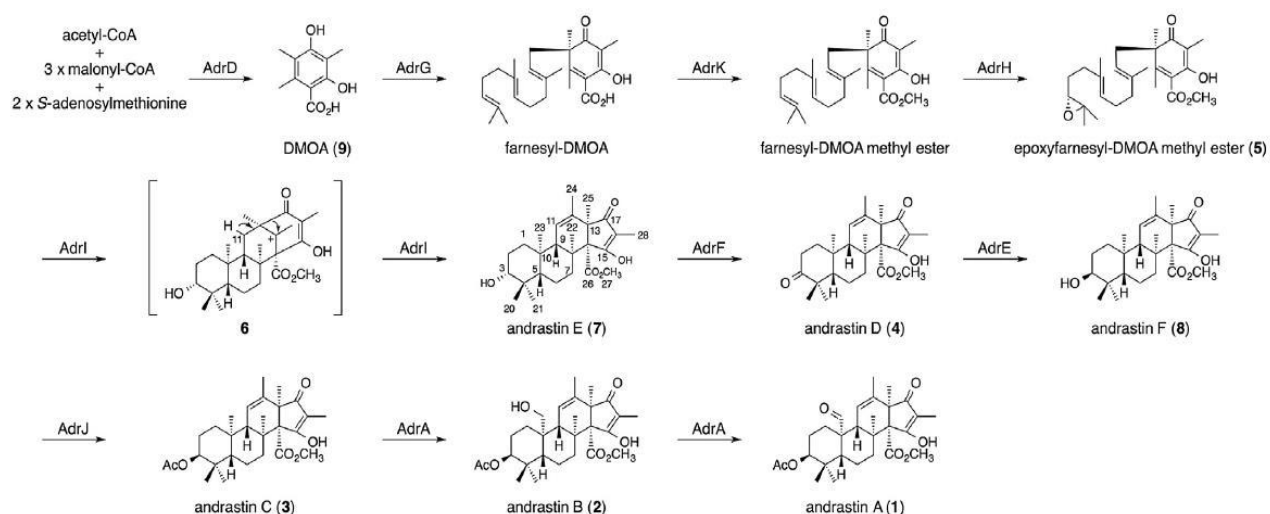


Figure 9: Proposed biosynthetic pathway of andrastin A (Matsuda *et al.*, 2013).

Aristolochene

Aristolochene is a sesquiterpene biosynthesized from farnesyl pyrophosphate by aristolochene synthase and is the precursor of a range of sesquiterpenoid toxins produced by filamentous fungi (Hohn *et al.*, 1991), including PR-toxin produced by *P. roqueforti*.

Asperparalines

Asperparalines A, B and C, were isolated from okara (the insoluble residue of whole soybean) that had been fermented with *A. japonicus* JV-23, and they were shown to have paralytic activities in silk worms (Hayashi *et al.*, 1997 and 2000). More recently, it has been demonstrated that asperparaline A selectively blocks insect nicotinic acetylcholine receptors (Hirata *et al.*, 2011).

No standard toxicity studies have been found in this review.

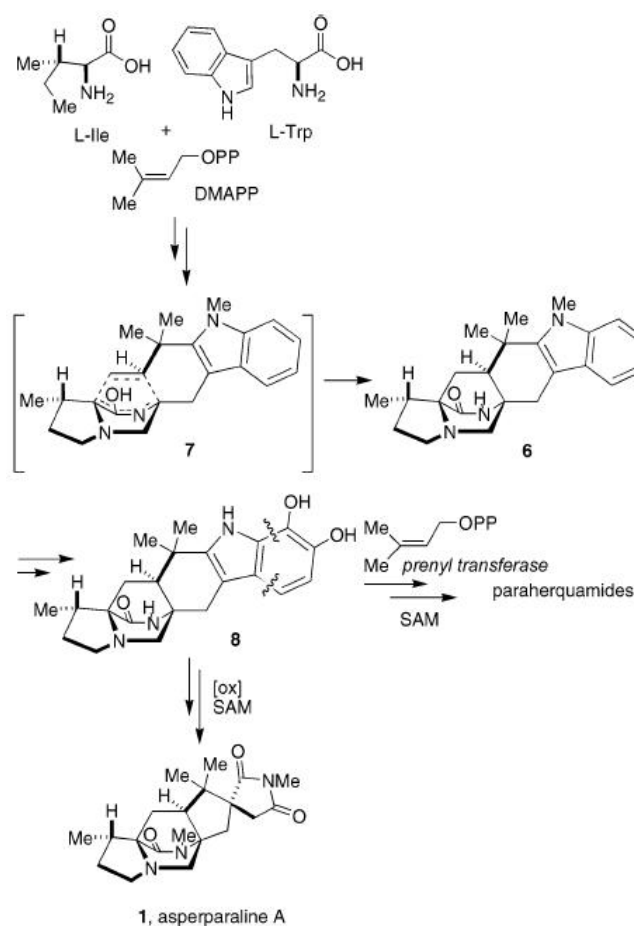


Figure 10: Studies on the biosynthesis of Asperparaline A: Origin of the Spirosuccinimide Ring System (from Gray *et al.*, 2003).

Citreoviridin

Citreoviridin A is a mycotoxin found in cereal and bread grains. It exhibits pro-oxidative, anticancer chemotherapeutic, antibacterial, antifungal, and antiviral activities. *In vitro*, citreoviridin A increases ROS levels and decreases glutathione, inducing oxidative stress, destabilizing the lysosome and mitochondria and causing DNA damage.

It is a neurotoxin, acutely toxic to mice, with intraperitoneal and oral LD₅₀ of 7.5 mg/kg and 20 mg/kg respectively (Ueno and Ueno, 1972).

Culmorin

The sesquiterpene culmorin was first isolated by Ashley *et al.* (1937) from *Fusarium culmorum*. It has been shown to be produced also by *F. venenatum* (Greenhalgh *et al.*, 1984; Lauren *et al.*, 1992; Miller and Mackenzie, 2000). This compound has been detected in cereals.

Strongman *et al.* (1987) showed the antifungal activity of culmorin against a variety of marine and medically relevant fungi. The only reported study on the toxicity of culmorin in mammals was published by Rotter *et al.* (1992). Culmorin has a low toxicity in several biological assays (Pedersen and Miller, 1999) but a synergistic effect with deoxynivalenol has been demonstrated (Dowd *et al.*, 1989)

Cyclophenol

Cyclophenol is a benzodiazepine metabolite produced in several species of *Penicillium*. It displays antimicrobial and phytotoxic properties.

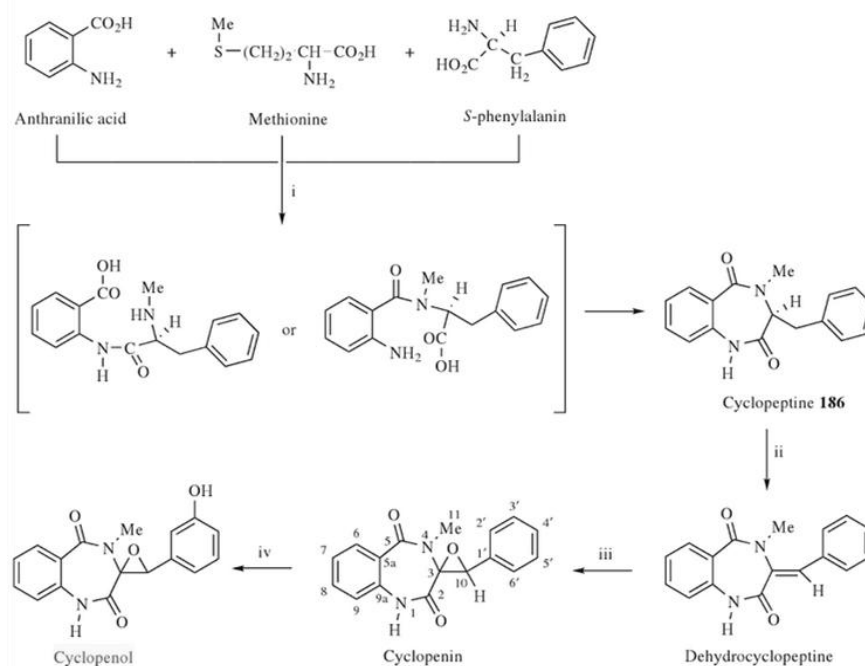


Figure 11: Biosynthetic pathway to cyclophenin and cyclophenol (modified from Diana P and Cirrincione G. Biosynthesis of Heterocycles: From Isolation to Gene Cluster. Wiley, 2015)

Cyclopiazonic Acid (α -cyclopiazonic acid)

Cyclopiazonic acid (CPA) is a mycotoxin produced by the genera *Aspergillus* and *Penicillium* which has been shown to be toxic in several animal species including swine, chickens, turkeys, guinea pigs, rats, and dogs.

CPA may be produced by the same species that produce aflatoxins. No standard toxicity studies have been found in this revision, apart from some in vitro cytotoxicity studies in porcine lymphocytes or human cells. A discussion on the article regarding safety assessment of the CPA that had been published by Burdock and Flamm in 2000 has been found (De Waal, 2002).

Cyclopiazonic acid is an indole-tetramic acid mycotoxin and it has been reported to be produced by *A. oryzae* and *P. camembertii*, both species used in food fermentation (Bockelmann *et al.*, 1999; Chang *et al.*, 2009; Shimada *et al.*, 2000)

There have been identified three clustered biosynthetic genes in *A. flavus* and closely related *A. oryzae* (Chang *et al.*, 2009; Tokuoka *et al.*, 2008)

The toxicity of CPA is attributed to its ability to alter normal intracellular calcium flux through the specific inhibition of sarcoplasmic or endoplasmic reticulum calcium-dependent ATPase (SERCA) essential for calcium uptake as in the muscle contraction-relaxation cycle, which results in increased muscle contraction.

Regarding the biosynthesis, it has been reported that the gene cluster involved in CPA biosynthesis is adjacent to the aflatoxin gene cluster (Chang *et al.*, 2009) and that the dimethylallyl-cycloacetoacetyl-L-tryptophan synthase (DCAT-S) gene from an *A. oryzae* is involved in the CPA production, as disruption of this gene resulted in the loss of production of this toxin.

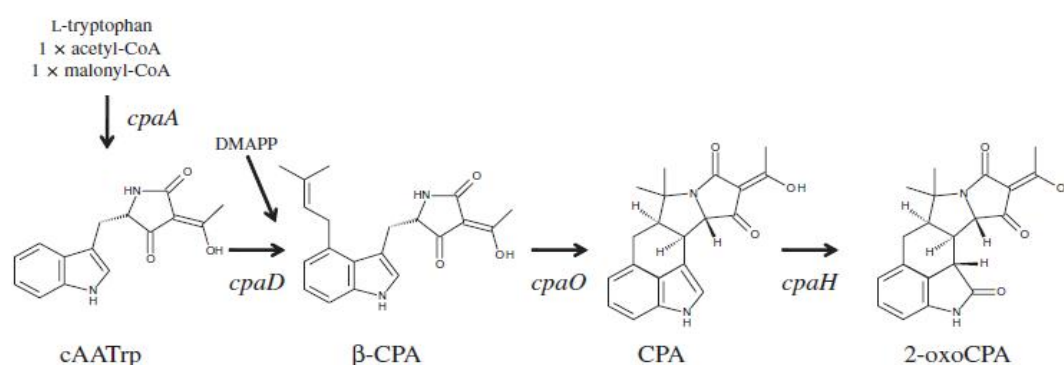


Figure 12: Cyclopiazonic Acid Biosynthesis Gene Cluster in *A. oryzae*. Structures of the intermediates in the biosynthetic pathway of 2-oxocyclopiazonic acid (2-oxoCPA) in *A. oryzae*. Genes in CPA cluster corresponding to each pathway are described beside arrows. Abbreviations: cAATrp, cyclo-acetoacetyl-L-tryptophan; B-CPA, B-cyclopiazonic acid; CPA, cyclopiazonic acid; 2-oxoCPA, 2-oxocyclopiazonic acid; DMAPP, dimethylallyl diphosphate) (from Shinohara *et al.*, 2011)

These authors reported that the co-localization of the two gene clusters explains why strains of *A. flavus* and *A. oryzae* have different abilities to produce aflatoxin and CPA. This understanding has significant health implications. The genetic diversity of *A. flavus* and *A. oryzae* in the region adjoining the CPA gene cluster suggests a divergence of *A. flavus* from *A. oryzae*. The conclusion is that *A. oryzae* most likely descended from an ancestor that was the precursor of the *Aspergillus* SBG variant, while *A. flavus* descended from a precursor of *A. parasiticus* (Chang *et al.*, 2009).

Regarding *A. niger*, it has been reported that it can be modified by mutagenesis, resulting in the deletion of gene clusters required for the synthesis of this mycotoxin (EFSA Journal 2014b).

Isotrichodermin/Isotricodermol

These compounds belong to the class of organic compounds known as trichothecenes. These are sesquiterpene mycotoxins structurally characterized by the presence of an epoxide ring and a benzopyran derivative with a variant number of hydroxyl, acetyl, or other substituents. The most important structural features causing the biological activities of trichothecenes are the 12,13-epoxy ring, the presence of hydroxyl or acetyl groups at appropriate positions on the trichothecene nucleus and the structure and position of the side-chain.

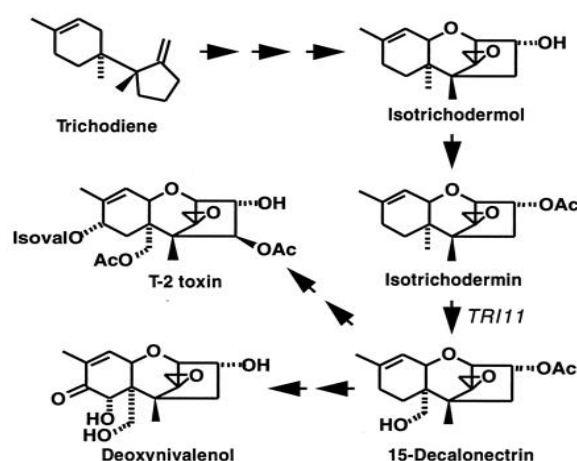


Figure 13: Trichothecene pathway (from Alexander NJ *et al.*, 1998)

Kojic acid

Kojic acid is produced by several *Aspergillus* and *Penicillium* species. It is a tyrosinase inhibitor thus inhibiting the production of pigments in animals and vegetables. Among other applications, it is used as a food additive to avoid browning of vegetables and crustaceans, and as a cosmetic ingredient for skin lightening. It was evaluated by the IARC and classified in the Group 3 *not classifiable as to its carcinogenicity to humans*, due to inadequate evidence in humans and limited evidence in experimental animals (IARC, 2001).

In this project, kojic acid has been identified as a secondary metabolite of *A. oryzae* (Chaves *et al.*, 2012) and three reports giving information on toxic effects of this compound have been retrieved.

In a 55-week chronic toxicity study performed on male F344/DuCrj rats, kojic acid was given at 0 (control group), 0.5% and 2% in the diet (Ota *et al.*, 2009). Dose-dependent toxic effects were observed in liver, kidney, thyroids and adrenals, with very significant differences at the highest dose. A no observed adverse effect level (NOAEL) below 0.5%, which is equivalent to 227 mg/kg body weight/day in male rats, was determined.

In a 26-week chronic toxicity study performed on male heterozygous p53-deficient CBA [p53(+/-)] mice and wild-type [p53(+/+)] mice, kojic acid was given at 0 (control group), 1.5% and 3% in the diet, and the tumorigenic potential in the thyroids and the liver was determined (Takizawa *et al.*, 2003). It was observed that kojic acid induced diffuse hypertrophy and hyperplasia of thyroid follicular epithelial cells with decreased serum thyroxine levels in both p53 (+/-) and p53 (+/+) mice, but caused no thyroid tumors. In the liver, the incidence of altered hepatocellular foci and hepatocellular adenomas was significantly increased, and the heterozygous p53 (+/-) mice appeared to be more susceptible in terms of the tumorigenic dose of kojic acid, with a greater prevalence of hepatic proliferative lesions. Thus, the contribution of genotoxicity on hepatocellular tumor development could not be ruled out.

Finally, a thorough investigation, under Good Laboratory Practice (GLP) conditions, of the genotoxicity and general toxicity of kojic acid, has been performed in order to estimate a margin of safety for topical exposure in humans to this compound (Nohynek *et al.*, 2004). The authors concluded that the genotoxic risk for humans using kojic acid as a skin lightening agent is negligible, and is in any case much less than from exposure to kojic acid from fermented foods. They also pointed out that the data obtained also suggested that consumer exposure from fermented foods does not pose a significant risk to human health.

Malformins

Malformins are a group of secondary metabolites originally discovered and isolated from culture filtrate of the fungus *A. niger* (Blumenthal, 2004).

No standard toxicity studies have been found in this review. The only *in vivo* study in which some toxicological endpoints were evaluated (Wang *et al.*, 2015) in BDF1 mice concluded that malformin C has potent cell growth inhibition activity, but the therapeutic index is too low to be an anti-cancer drug.

Some of the compiled articles evaluated malformins for their potential therapeutic properties (anti-cancer, fibrinolytic, anti-HIV and antimicrobial activities). Three different types of malformins (A, C and E) have been studied.

An LD₅₀ (3.1 mg/kg, i.p, mouse) (from 1975) has been found in ChemIDplus database.

Maltoryzine

This compound was first isolated from the culture broth of a strain of *A. oryzae* (Blumenthal, 2004). No standard toxicity studies have been found in this review. The only original document reporting toxic effects found is a study from 1962 (Iizuka *et al.*, 1962).

An LD₅₀ (> 3mg/kg, i.p, mouse) (from 1969) has been found in ChemIDplus database.

Mycophenolic acid

Mycophenolic acid (MPA) is a toxic metabolite produced by many *Penicillium* species such as *P. stoloniferum*, *P. viridicatum*, *P. brevicompactum*, *P. carneum*, *P. raciborskii* and some strains of *P. roqueforti*. Mycophenolic acid (MPA) is an approved therapeutic drug used as an immunosuppressant and numerous articles devoted to its therapeutic effects have been found. Its main mechanism of action is related to a reversible inhibition of inosine monophosphate dehydrogenase. More toxicological data can be obtained from its preclinical and clinical assessment, mainly from mycophenolate mofetil, which is its prodrug.

In this review, it has been found two reports on developmental defects in zebrafish embryos have been retrieved. Morphological defects including tail curvature and severe pericardial edema in zebrafish embryos were found in a dose-dependent manner (3.7-11.1 µmol/L). The teratogenic index (25% lethal concentration value (LC25)/no observed adverse effect level ratio) was 16, which indicated mycophenolic acid as a teratogen (Jiang *et al.*, 2016). In a second study (Schmidt *et al.*, 2013) in which MPA was evaluated at concentrations of 0.1; 0.25; 0.5; 0.75 mg/ L (0.31; 0.78; 1.56; 2.34 µM), all zebrafish embryos showed dysmorphic changes at 0.75 mg MPA/ L medium. Embryos exposed to 0.25 mg MPA/ L medium showed impaired development of nerves, and at 0.1 mg/ L, no effects were detectable.

Other report with preliminary toxicity assessments, such as cytotoxicity in zebrafish, mechanistic information related to damage of insulin-secreting cells of MPA or gastrointestinal tract toxic effects in rats have been found. A recent review providing an update of the literature on the pharmacology and toxicology of the prodrug of MPA, mycophenolate, in organ transplant recipients has been found (Staatz and Tett, 2014).

Some information (updated in 1995) related to its genotoxic effects can be found in CCRIS and GENE-TOX databases (see Appendix C). No conclusions were obtained from an Ames test from 1978, and negative results have been found in an Ames test (from 1983) performed with TA98 and TA100 *Salmonella typhimurium* strains in presence of metabolic activation. Negative results were obtained in Rec-assay (from 1981), while positive results were obtained in a chromosome aberrations test performed in vitro (from 1981).

Various LD₅₀ (all previous to 1983) in different animal species and with different routes of exposures are collected in ChemIDplus database (see Appendix C).

In this review, reports have been found on the production of MPA in blue-veined cheeses due to the fact that *P. roqueforti* (a producer of MPA) is used in the production of this type of cheeses (Malekinejad *et al.*, 2015).

Naphtho- γ -pyrones

These compounds have been associated with *A. niger* and are defined as a group of aromatic compounds able to cause central nervous system signs in albino mice and rats leading to death when dosed intraperitoneally (Blumenthal, 2004).

No standard toxicity studies have been found in this review. It is a generic name of naturally-occurring structurally-related compounds.

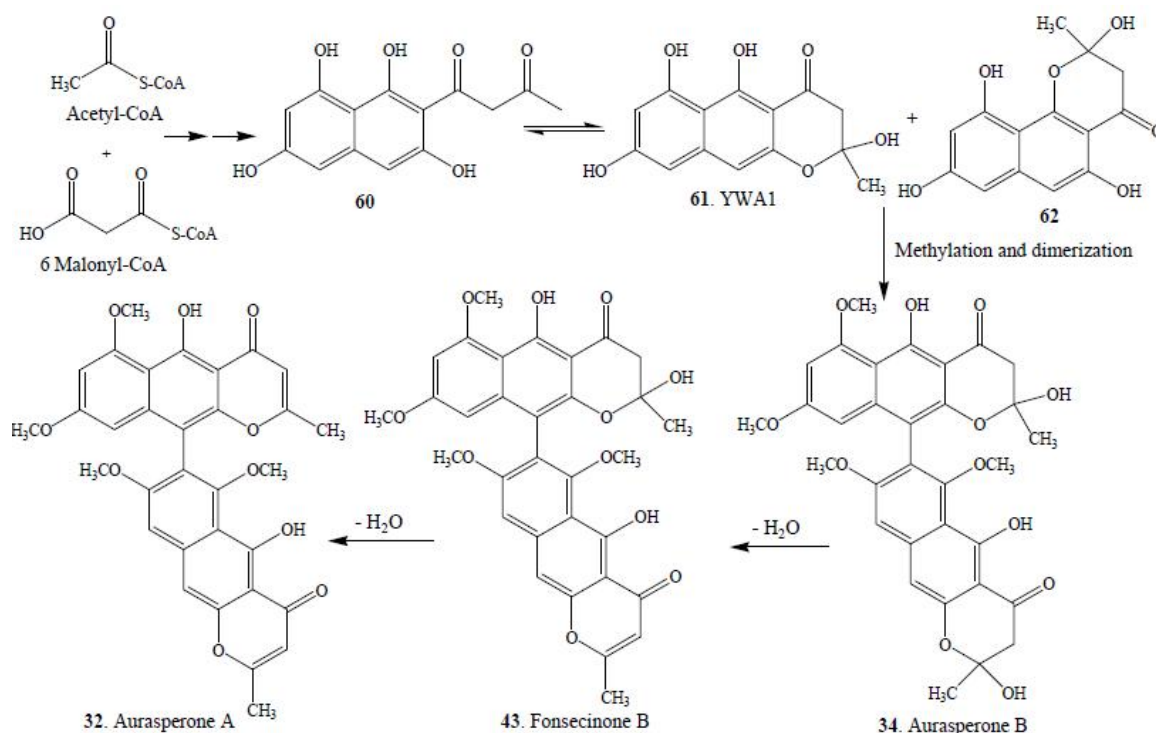


Figure 14: Proposed biosynthetic pathway of asperpyrone-type bis-naphtho- γ -pyrones in *A. niger*. (from Lu *et al.*, 2014)

Neoxaline

This compound has been described as an alkaloid produced by *A. japonicus* (Hirano *et al.*, 1969). No standard toxicity studies have been found in this review. An LD₅₀ (> 200 mg/kg, i.p, mouse) (from 1979) has been found in ChemIDplus database.

Neoxaline has been reported to have antiproliferative activity (in an MTT assay) and arrest the cell cycle at the G2/M phase in Jurkat cells (Koizumi *et al.*, 2004)

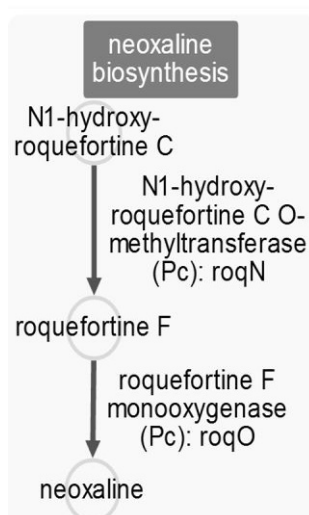


Figure 15: Biosynthetic pathway of neoxaline. From Caspi *et al.*, 2014, "The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of Pathway/Genome Databases, " *Nucleic Acids Research* 42:D459-D471.

Nigerazines

Nigerazines were first isolated by Iwamoto *et al.* (1983, 1985) from *A. niger* and found to inhibit root growth of lettuce seedlings. Nigerazines can be extracted from the mycelia of *A. niger* (Iwamoto *et al.*, 1983). Acute toxicity (LD₅₀) in mice was determined to be 75 ~ 150 mg/kg.

Nigragillin

This toxin was obtained in 1969 by Caesar *et al.* as a fungal alkaloid from the *A. niger* group. It can be extracted from culture filtrate and was demonstrated to be toxic to silkworm larvae (Isogai *et al.*, 1975). The compound was also found to be produced by *A. phoenicis* which is a species of Aspergilli Section Nigri (Cole and Cox, 1981).

Orlandin

This compound has been identified as a secondary metabolite produced by *A. niger* together with other toxins, such as fumonisin B 2 (Sørensen *et al.*, 2009). No standard toxicity studies have been found in this review, an LD₅₀ (> 125 mg/kg, oral, chicken) (from 1979) has been found in ChemIDplus database.

Oxalic acid

Oxalic acid is a toxic organic compound found in certain plants and natural sources. Although there have been reports of oxalic acid poisoning involving the consumption of food, toxicological effects in humans are relatively uncommon but include gastrointestinal effects, hypocalcemia secondary to calcium oxalate crystal formation and renal toxicity. *A. niger* is associated with pulmonary oxalosis consisting of calcium oxalate crystals, which generate local oxidants that cause cell injury (Blumenthal, 2004).

In this review, no standard toxicity studies have been found. Some information related to its genotoxic effects can be found in CCRIS (updated for this compound in 2006). Three different Ames tests (from 1987, 1988 and 1997) were negative with and without metabolic activation using different bacteria strains (*S. typhimurium*: TA 97, TA 98, TA 100, TA 102, TA 104, TA 1535, TA 1537, *E.coli*: WP2 UVRA,

WP2 UVRA/PKM101). A previous study from 1983 cited by the National Toxicology Program (NTP) also reported negative results for the Ames test.

Another study from the NTP (1985) in which the effects of oxalic acid on reproduction and fertility were assessed in CD-1 mice via drinking water concluded that oxalic acid is a reproductive toxicant in Swiss mice at concentrations that reduce parental water consumption, but that cause few other somatic effects.

Various LD₅₀ (previous to 1990) in different animal species and with different routes of exposures are collected in ChemIDplus database (see Appendix C). The LD₅₀ ranged from 112 mg/kg in cats (subcutaneous administration) to 7500 mg/kg in rats (oral administration). In humans, a lethal dose of 600 mg/kg was reported in 1980 (females).

Several occupational limit values have been established by different occupational safety organizations (information in HAZ-MAP database).

The production of oxalic acid in *A. niger* involves the enzyme oxaloacetate acetylhydrolase (Ruijter *et al.*, 1999), but a second pathway that has been elucidated demonstrates the production of oxalate from pentoses via glycolate and glyoxylate as intermediates. There are several reports that provide the optimised parameters for transformation of different substrates, including wastes containing free glucose or sugars to oxalic acid by *A. niger* strains (Mandal and Banerjee, 2005; Cameselle *et al.*, 1998)

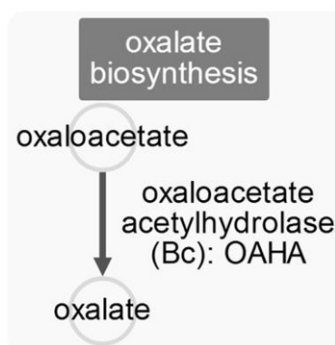


Figure 16: Biosynthetic pathway of oxalic acid (oxalate). From Caspi *et al.*, 2014, "The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of Pathway/Genome Databases, " *Nucleic Acids Research* 42:D459-D471.

Perinadine A

Perinadine A is an alkaloid isolated from *P. citrinum* in 2006 (Sasaki M *et al.*, 2005). No standard toxicity studies have been found. No data were available for this compound in any of the databases consulted.

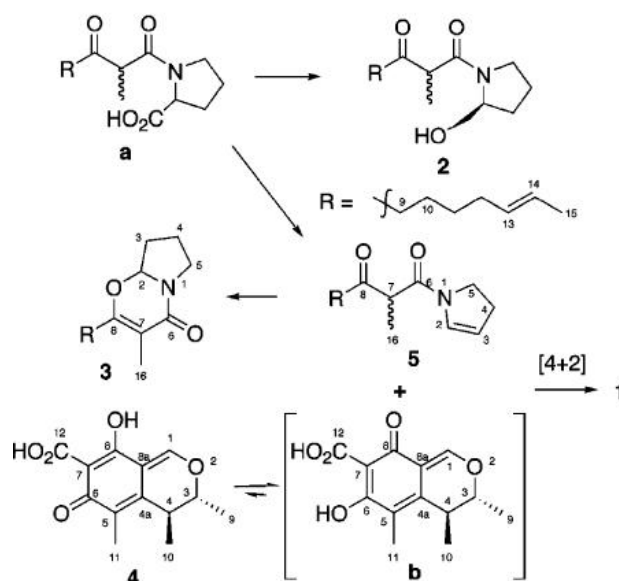


Figure 17: Synthesis of perinadine A. Perinadine A (1) may be derived from a known pyrrolidine alkaloid (5) isolated from *P. brevicompactum* and citrinin (4), a well-known mycotoxin. An intermediate a, which may be derived from glutamic acid or proline and a pentaketide, are considered to be converted into 5 by decarboxylation at C-16. Intermolecular cyclization between 5 and an intermediate b equal to citrinin (4) may give perinadine A (1). On the other hand, scalusamide A (2) may be generated from a through reduction of the carboxyl Group at C-16, while pyrrolo[2,1-*b*]oxazines (3) is likely to be converted from 5 through Michael-type cyclization between C-2 and a carbonyl oxygen at C-8 (from Sasaki *et al.*, 2005).

PR-toxin

This compound produced by *P. roqueforti* is a sesquiterpenoid mycotoxin. PR-toxin produced acute toxic effects in animals via an increase of capillary permeability and direct damage to lungs, heart, liver and kidneys.

In this review no standard toxicity studies have been found. Some studies evaluating the effects on health status and animal performance found that PR-toxin did not affect rumen fermentation pattern (Gallo *et al.*, 2015; Gallo *et al.*, 2015b)

PR-toxin selectively exhibited cytotoxic activity towards certain cancer cell lines (Darsih *et al.*, 2015) and Caco-2 cells (Rasmussen *et al.*, 2011). Some information related to its genotoxic effects can be found in CCRIS (updated in 1994) and GENE-TOX (updated in 1998) databases. PR-toxin gave positive Ames tests with TA 98 and TA100 strains. Positive results were also obtained in Rec-assay in *Bacillus subtilis* and gene conversions assays in *Saccharomyces cerevisiae* and *Neurospora crassa*.

Various LD₅₀ (previous to 1990) in rodents and with different routes of exposures are collected in ChemIDplus database. The LD₅₀ ranged in mice from 2 mg/kg (intravenous and intraperitoneal administration) to 72 mg/kg in after oral administration. In rats, LD₅₀ ranged from 8.2 mg/kg (intravenous) to 11.6 mg/kg (intraperitoneal). A LD_{LO} of 115mg/kg has been reported in rats.

There have been reported the conditions of PR-toxin production, which only happens in stationary cultures, within the pH range of 4.5-9.0 and temperature range of 10°-30°C with the optimum temperature at 24°C. In the database it is also included the PR-toxin biosynthetic pathway by *P. roqueforti*, through the action of aristolochene synthase.

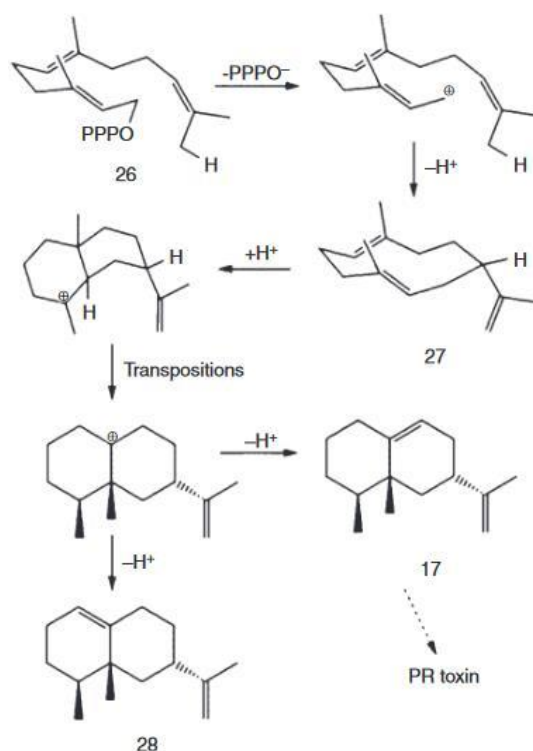


Figure 18: Biogenesis of volatile sesquiterpenes involved in the pathway of PR-toxin in toxic strains of *P. roqueforti*. [farne-syl pyrophosphate (26); aristolochene (17); AS enzyme with germacrene A (27), valencene (28)] (from Mioso *et al.*, 2015)

Roquefortine C

Roquefortine C is a neurotoxic mycotoxin derived from the diketopiperazine cycle (Trp-dehydro-His) that causes a paralytic syndrome. Its mechanism of action is unknown but it can interact with cytochrome P450 and other haemoproteins (Aninat *et al.*, 2001). It can also activate P-glicoprotein transport system (Aninat *et al.*, 2005).

Roquefortine-type alkaloids are produced by several *Penicillium* species including species of industrial applications such as *P. roqueforti* (Sumarah *et al.*, 2005) and *P. chrysogenum* (Bringmann *et al.*, 2005).

P. roqueforti roquefortine cluster contains only four genes (*rds*, *rdh*, *rpt*, and *gmt*) encoding the roquefortine dipeptide synthetase, roquefortine D dehydrogenase, roquefortine prenyltransferase, and a methyltransferase, respectively.

Silencing of the *rds* or *rpt* genes by the RNAi strategy reduced roquefortine C production by 50%, confirming the involvement of these two key genes in roquefortine biosynthesis.

P. roqueforti lacks the genes that encode the enzymes for the conversion of roquefortine C to roquefortine L (or glandicoline A) and meleagrín, a difference between other *Penicillium* species with the same gene cluster as *P. chrysogenum*.

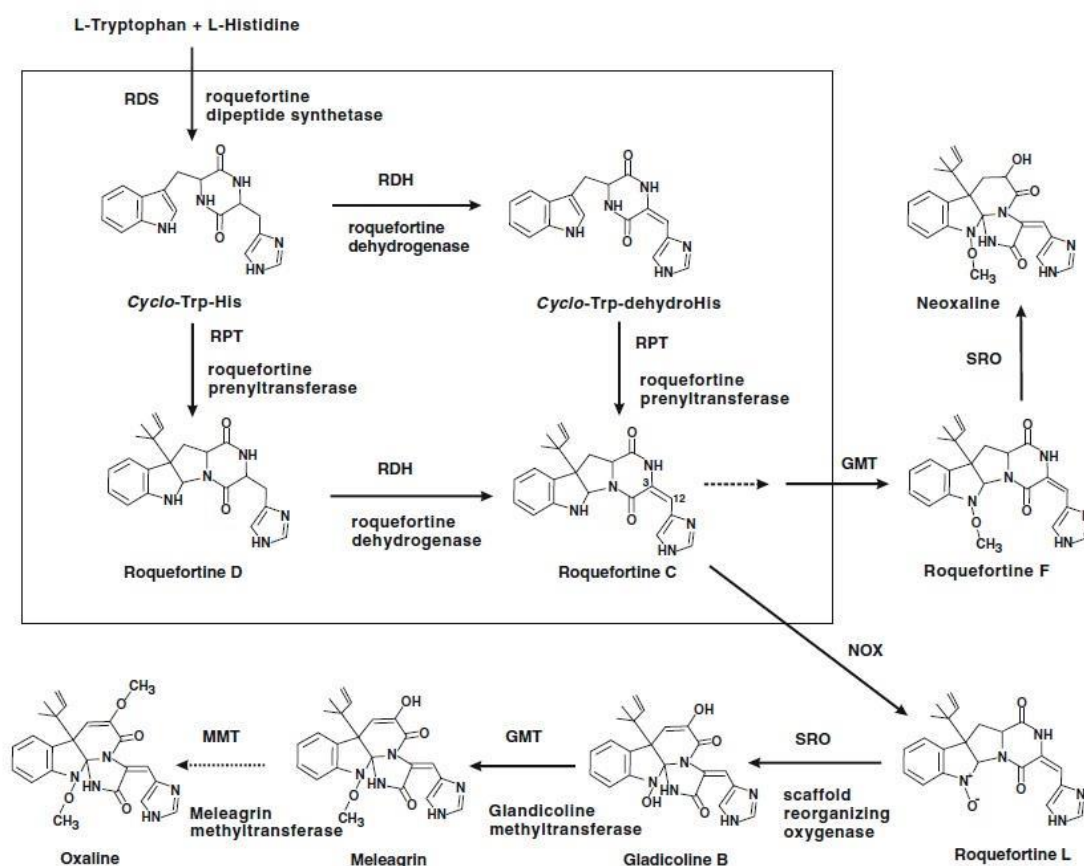


Figure 19: Biosynthetic pathways of roquefortine C. The meleagrins pathway (from roquefortine C) is shown below and the neoxaline branch on the right side. The putative conversion of meleagrins into oxalines is indicated by a dashed arrow at the left side below the box. This conversion is catalyzed by a second methyltransferase (MMT, meleagrins methyl transferase) (from Martín *et al.*, 2016)

Derivatives of roquefortine

- **Isfumigaclavine A and B:** Isfumigaclavine A is another alkaloid produced by *P. roqueforti*. This toxin and the product of its hydrolysis, isfumigaclavine B, are identical with roquefortines A and B, respectively.
- **Festoclavine and Marcfortine A:** Festoclavine is an intermediate metabolite for the synthesis of ergot alkaloids and Marcfortine A is clavine alkaloid toxins produced by *P. roqueforti*. No standard toxicity studies have been found in this review; however, Marcfortine has been reported to have a potent antihelmintic activity.
- The mitorubins are a unique subclass of azaphilones (Marsini *et al.*, 2006) isolated from a variety of fungal species. In this review, **mitorubrinic acid** has been identified in the fermentation broth of *P. funiculosum*, together with other bioactive metabolites (Lesová *et al.*, 2001). No standard toxicity studies have been found in this review. It is a polyketide and has been considered, together with mitorubrinol, as virulence factors of *P. marneffei* (Marsini *et al.*, 2006). A recent review was done by Tam EW *et al.*, 2015.

Sambucinol

This is a trichothecene metabolite produced by several species of *Fusarium* such as *F. venenatum*. The metabolic route of this compound has been studied and postulated that 2a-hydroxytrichodiene is the first oxygenated step in its biosynthesis (Zamir et al. 1999).

Secalonic acid D

Secalonic acid D (SAD), a metabolite of *P. oxalicum*, although it has been reported that can be produced by *A. aculeatus* (Gao et al., 2014).

In this review, no standard toxicity studies have been found. However, some mechanistically-relevant articles and reviews have been retrieved from the systematic search. Secalonic acids, mainly secalonic D, have been evaluated for potential anticancer activity in vitro (antiangiogenic activity, cytotoxicity, ABCG2 downregulation, cell signaling disruption, DNA topoisomerase I inhibitor, apoptosis, cell cycle arrest at G1). In vivo, a preliminar preclinical validation and mechanistic definition of the antiangiogenic activity of secalonic acid D with potential application as a cancer-selective therapeutic agent was carried out. Treatment of SAD at all experimental doses was nontoxic in solid tumor model animals (mice) as no mortality (0/7) occurred during the course of the experiment (Guru et al., 2015).

Secalonic A has shown to antagonize the cytotoxicity of colchicine in the rat cortical neurons through inhibition of phosphorylation of JNK and p38 MAPKs, calcium influx, and the activation of caspase-3.

Another set of articles found focused on unraveling the mechanism of teratogenesis (cleft palate induction) of SAD in mice.

Some information related to genotoxic effects of secalonic acid D can be found in CCRIS (updated in 1993) and GENE-TOX (updated in 1995) databases: negative results with and without metabolic activation were obtained in an Ames test (TA 100) (study from 1992). No conclusions were drawn from a previous study performed in 1978.

An LD_{LO} (15.6 mg/kg, mouse, intraperitoneal) (from 1952) has been found in ChemIDplus database.

Trichodermin

As mentioned in the previous section, Trichodermin is a trichothecene (4 β -acetoxy-12, 13-epoxy- Δ 9-trichothecene). A review of the understanding of the relationship between structure and known function of these compounds has been found (Alexander NJ et al., 1998). No standard toxicity studies have been found in this review. Some reports evaluated trichodermin for its potential therapeutic effects as antifungal or antitumoral.

An LD₅₀ (500 mg/kg, mouse, intraperitoneal and subcutaneous) (from 1986) have been found in ChemIDplus database.

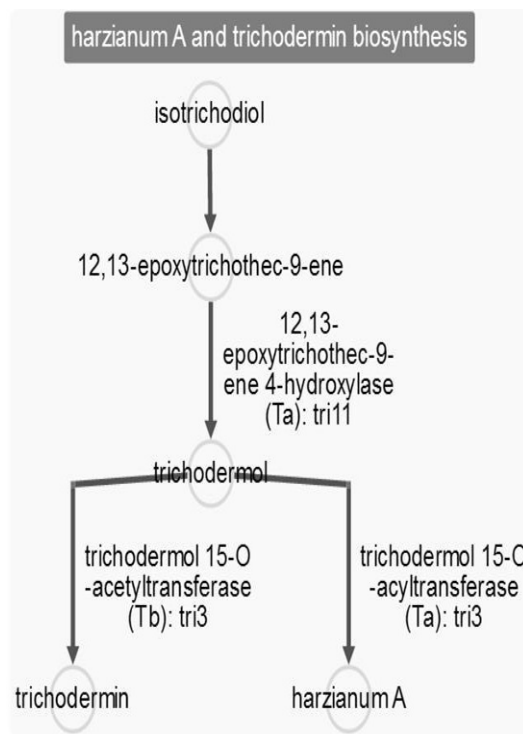


Figure 20: Biosynthetic pathway of trichodermin. From Caspi *et al.*, 2014, "The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of Pathway/Genome Databases, " *Nucleic Acids Research* 42:D459-D471.

Trichorzin

This substance reported as a mycotoxin by Bisht *et al.* 2011, belongs to a peptaibol family and has been studied for different functionalities as calcitonin (CT) agonist (Katayama *et al.* 2001) and antibiotic activity against phytopathogenic fungi as well as several bacteria (Bisht *et al.* 2011).

Xanthomegnin

This compound has been identified as a secondary metabolite of *A. melleus* (Neidig *et al.*, 2013).

Xanthomegnin has shown to have potent inhibitory activity in the inducible nitric oxide synthase (iNOS) assay (Alvi *et al.*, 2000). No standard toxicity studies have been found in this review.

This compound has been studied in the past. No teratogenic effects were observed in ICR mice (Bolin *et al.*, 1991) and it has been proposed has a potential genotoxic (Carlton *et al.*, 1976; Mori *et al.*, 1983). Some hepatic alterations produced in mice by xanthomegnin were evaluated in 1976 (Carlton *et al.*, 1976)

3.3.1.2. Toxic substances produced by fungi fully characterised from the toxicological point of view (Appendix E)

There are several recognised toxic compounds, some of them legislated under European regulations for food and feed and that have been reported to be produced by species of fungi used in industrial fermentations. Some of their characteristics and biosynthesis are described below.

Aflatoxins

Aflatoxins are one of the main mycotoxins, produced by some species of *Aspergillus* Section Flavi.

Aflatoxin B1 is the most common in food and widely studied mycotoxin. It is produced primarily by the two species: *A. flavus* and *A. parasiticus*, which are especially found in areas with hot and humid climates. *A. flavus* is ubiquitous, favouring the aerial parts of plants (leaves, flowers) and produces B aflatoxins. *A. parasiticus* produces both B and G aflatoxins. Aflatoxins are classified by the International Agency for Research on Cancer in the Group 1 “carcinogenic to human”, producing liver cancer (IARC, 2012). Aflatoxins are also considered as “known to be human carcinogens” by the NTP (2016).

Based on the existing evidence related to the toxicological effects of these mycotoxins, The European Regulation (EC) 1881/2006 and its modifications have established maximum permitted levels in certain foodstuffs, such peanuts, pistachios, tree nuts, cereals, maize, rice and milk.

The section Flavi of genus *Aspergillus* also includes *A. oryzae* and *A. sojae*, which are used for the production of various industrial enzymes and fermented foods in eastern Asia. Although is generally recognized by the scientific community that *Aspergillus oryzae* and *A. sojae* do not produce aflatoxins, some controversy has arisen in the past from reports by some authors (Atalla *et al.*, 2003). These fungi are taxonomically similar to aflatoxin-producing species; therefore, it is necessary to determine whether these strains have the potential to make toxins.

Amadi and Adeniyi (2009) reported the production of aflatoxin B1 from *A. oryzae* isolated from stored grains (rice, maize and millet), that was determined by thin layer chromatography (TLC).

In this review, it has been found several reports (Matsushima *et al.*, 2001; Kim *et al.*, 2014) that described the natural mutations in the *afR* gene (main transcriptional regulator of aflatoxin biosynthesis) that many strains of *A. oryzae* and *A. sojae* have and that prevents these microorganisms from producing aflatoxins.

The main conclusion from this data is that *afR* gene could be examined to assess possible aflatoxin production by industrial strains within the group *Aspergillus* section Flavi.

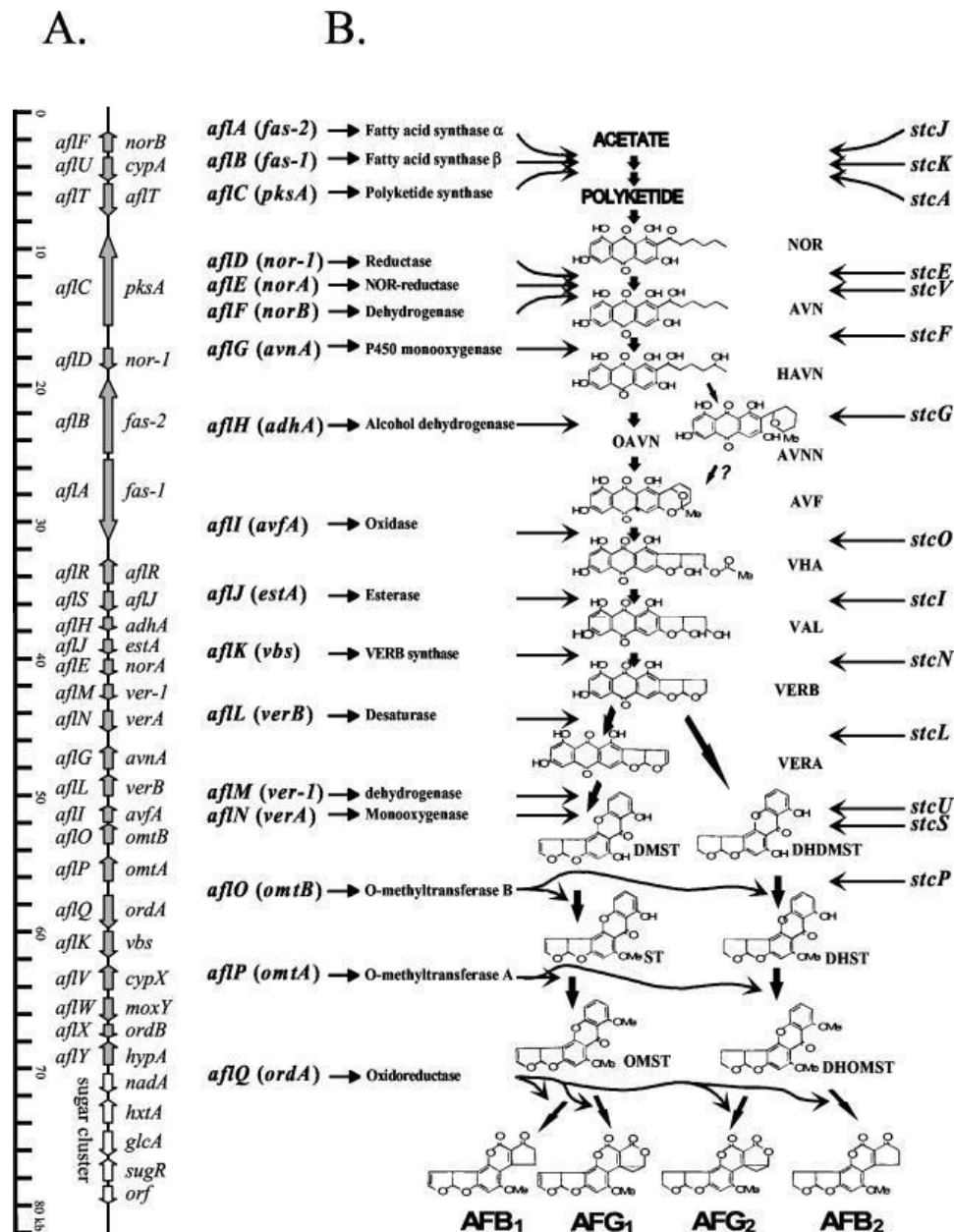


Figure 21: Clustered genes (A) and the aflatoxin biosynthetic pathway (B). The generally accepted pathway for aflatoxin and ST biosynthesis is presented in panel B. The corresponding genes and their enzymes involved in each bioconversion step are shown in panel A. The vertical line represents the 82-kb aflatoxin biosynthetic pathway gene cluster and sugar utilization gene cluster in *A. parasiticus* and *A. flavus*. The new gene names are given on the left of the vertical line and the old gene names are given on the right. Arrows along the vertical line indicate the direction of gene transcription. The ruler at far left indicates the relative sizes of these genes in kilobases. The ST biosynthetic pathway genes in *A. nidulans* are indicated at the right of panel B. Arrows in panel B indicate the connections from the genes to the enzymes they encode, from the enzymes to the bioconversion steps they are involved in, and from the intermediates to the products in the aflatoxin bioconversion steps. Abbreviations: NOR, norsolorinic acid; AVN, averantin; HAVN, 5'-hydroxyaverantin; OAVN, oxoaverantin; AVNN, averufanin; AVF, averufin; VHA, versiconal hemiacetal acetate; VAL, versiconal; VERB, versicolorin B; VERA, versicolorin A; DMST, demethylsterigmatocystin; DHDMST, dihydrodemethylsterigmatocystin; ST, sterigmatocystin; DHST, dihydrosterigmatocystin; OMST, O-methylsterigmatocystin; DHOMST, dihydro-O-methylsterigmatocystin; AFB₁, aflatoxin B₁; AFB₂, aflatoxin B₂; AFG₁, aflatoxin G₁; AFG₂, aflatoxin G₂ (from Yu *et al.*, 2004)

Beauvericin and Enniatins

These secondary metabolites are produced by *Fusarium* species, typical mycotoxin producing fungi. They are predominantly found in cereal grains and their products.

Beauvericin and enniatins possess a wide range of biological activities. These substances are known as ionophores, antimicrobial and antibiotic compounds, enzyme inhibitors, and as compounds inducing oxidative stress. They also have cytotoxic activity towards different cell types inducing apoptosis.

There are no reports on mycotoxicoses associated with these toxins, although different toxic actions are known *in vitro* for enniatins and beauvericin, probably related to their ionophoric properties. For beauvericin, the LD₅₀ for acute toxicity was 100 mg/kg b.w. upon oral administration to mice. An LD₅₀ of 350 mg/kg b.w. was reported for fusafungine (a mixture of enniatins) in mice upon oral administration. A sub-acute toxicity study on enniatin A with some limitations showed no adverse effects. Beauvericin is a strong insecticidal product (EFSA Journal 2014c).

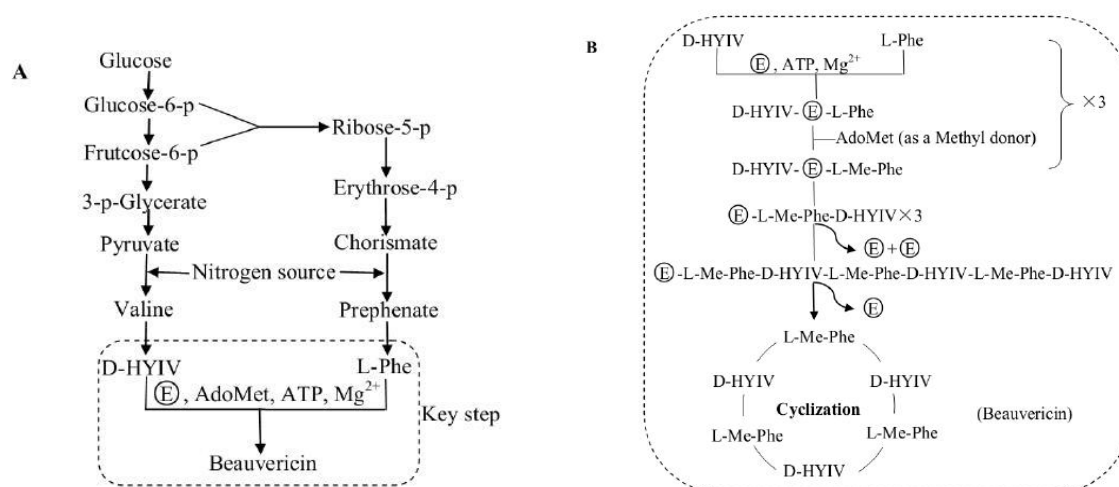


Figure 22: Beauvericin biosynthesis pathway; B, The specific substeps of the key step (E represents the beauvericin synthetase; the dashed frame indicates the key step of beauvericin synthesis); C, The possible structure of the beauvericin synthetase (E1 is the D-HYIV module; E2 is the L-Phe module; SH1,2,3 are the 4'-phosphopantetheine residues corresponding to D-HYIV, L-Phe, and the linear hexadepsipeptide acceptor; M is the N-methyltransferase domain; Cy is the cyclization cavity). Adapted from Wang Q and Xu L, 2012.

Citrinin

Citrinin is a nephrotoxic mycotoxin produced by several species of the genera *Aspergillus*, *Penicillium* and *Monascus*. It has been implicated in porcine nephropathy and has been found as a natural contaminant of corn, rice, wheat, rye, barley, and oats. Citrinin was classified as group 3 (not classifiable as to its carcinogenicity to humans) by IARC (IARC, 1987) and was more recently evaluated by EFSA (EFSA Journal 2012b).

Maximum levels of citrinin in food supplements based on rice fermented with red yeast *Monascus purpureus* have been laid down in the EU Commission regulation 212/2014 (amending Regulation (EC) No 1881/2006).

Citrinin is a polyketide that has been reported to be produced by the two fungal species *P. citrinum*, *P. chrysogenum*, but besides citrinin, a large number of citrinin derivatives have been isolated from different fungal species, citrinin decomposes during heat treatment to form other complex compounds, such as citrinin H1 and citrinin H2, respectively with higher and weaker cytotoxicity than the original citrinin.

There are some reports on the biosynthesis of citrinin by *P. citrinum* and the effect of several process conditions on the production of this mycotoxin by microorganisms used in industrial fermentations (Reddy *et al.*, 2010; Prabha *et al.*, 2009; Pimentel *et al.*, 1996; Panda *et al.*, 2015).

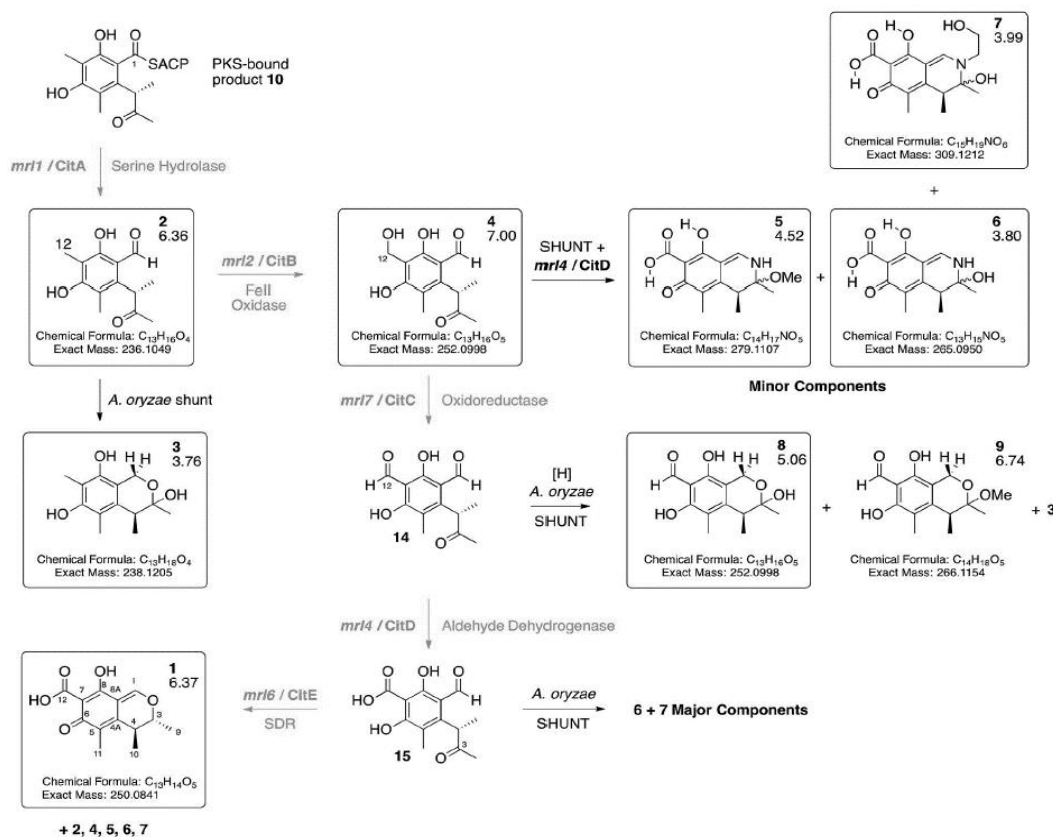


Figure 23: Deduced biosynthetic pathway to citrinin 1 (red steps). Boxed compounds purified and characterised. Black steps represent shunts branching from the main pathway. Figures below compound numbers re LCMS-Rt values. Unboxed compounds structure inferred (from He and Cox, 2016).

Ergot alkaloids

Ergot alkaloids (EA) belong to a diverse group of mycotoxins with neurological effect on humans, but also with a range of biological activities that have important applications in medicine and agriculture. EAs are produced by a variety of plant associated fungi, mainly of the genera *Claviceps* and *Aspergillus*. In this review it has been found to be produced by *A. japonicus* (Nielsen *et al.*, 2014) and *P. roqueforti* (Mioso *et al.*, 2015)

Ergot alkaloids in food and feed have been assessed by EFSA (EFSA Journal 2012a). EAs are not listed in Directive 2002/32/EC, however, the Directive sets a maximum content for rye ergot (*Claviceps purpurea*) of 1000 mg/kg in all feed containing unground cereals. The maximum content relates to a feed with a moisture content of 12%.

The production of ergot alkaloids has been linked to biosynthetic gene clusters found in several species, containing eight genes, seven of which (*dmaW*, *easF*, *easE*, *easC*, *easD*, *easA*, *easG*) are homologous to genes previously implicated in the biosynthesis of festuclavine or agroclavine in other filamentous fungi (Jakubczyk *et al.*, 2014).

All ergot alkaloids are derived from the common biosynthetic intermediate chanoclavine-I, and the structural diversity within the ergot alkaloids results from the elaborate chemical derivatization of this intermediate. Cycloclavine has been observed in only one species of filamentous fungus, *A. japonicus*. Inspection of the *A. japonicus* genome revealed a 16.8-kb biosynthetic cluster containing eight genes.

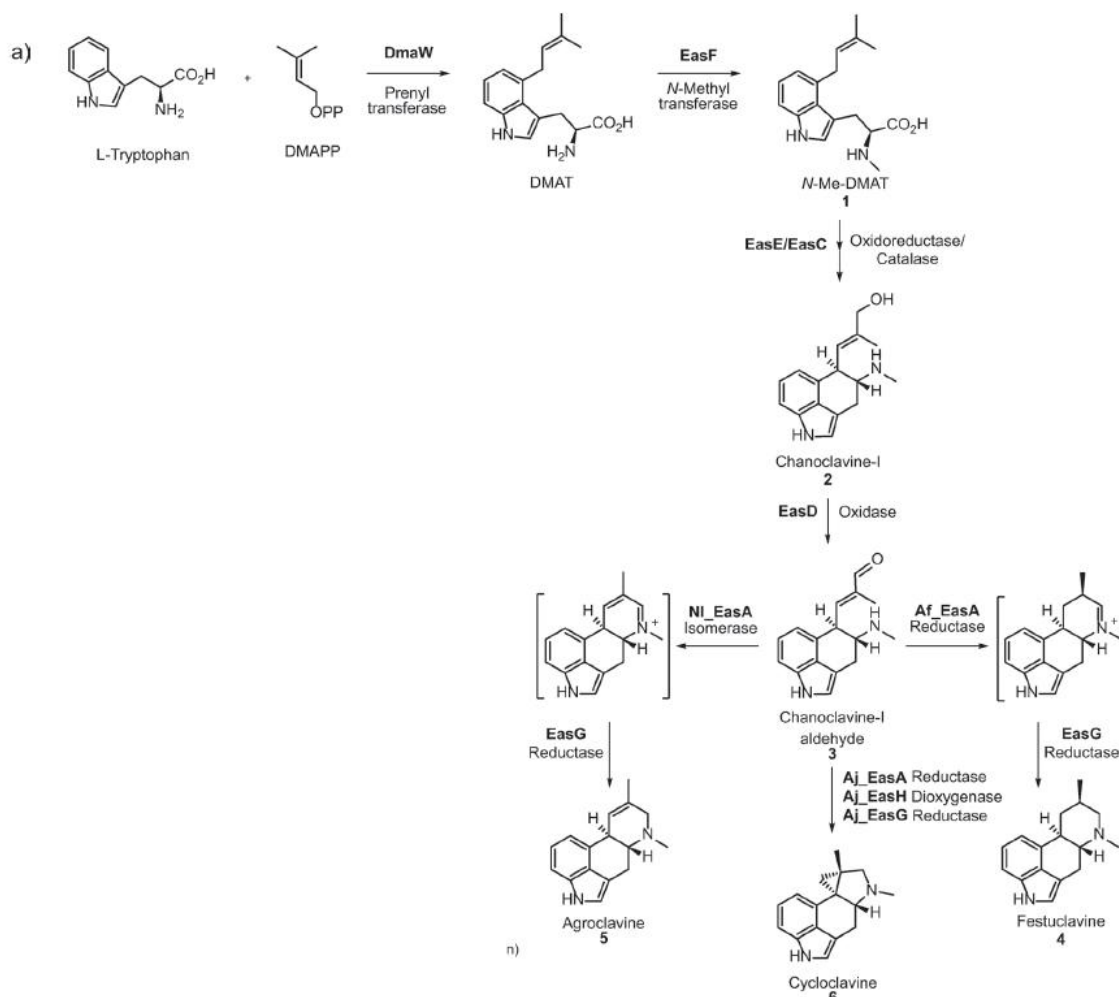


Figure 24: Ergot alkaloid biosynthetic pathway. A) Biosynthesis of festuclavine (4), agroclavine(5), and cycloclavine (6) from L-tryptophan and dimethylallyl pyrophosphate (DMAPP) (from Jakubczyk *et al.*, 2015).

Fumonisin

Fumonisin are formed mainly by *F. verticillioides* (*syn. F. moniliforme*) and *F. proliferatum*. At least 12 fumonisin analogues are known, the most important being the B series (fumonisins B1, B2 and B3). The most significant crop, in which fumonisins occur, is maize, particularly when grown in warmer regions. Fumonisin B1, B2 and fusarin C were classified by IARC as group 2B (possibly carcinogenic to humans) (IARC, 1993). This classification was maintained for fumonisin B1 in 2002 (IARC, 2002).

Fumonisin have been evaluated by EFSA in an Opinion of the Scientific Panel on contaminants in the food chain (CONTAM) related to fumonisins as undesirable substances in animal feed (EFSA Journal 2014a). EU legislation has set maximum levels for fumonisins in foodstuffs and more in particular for the sum of fumonisin B1 and B2, in maize and maize-based foods (EC 1881/2006 and its

modifications). Recently a scientific statement on the increase of risk for public health related to a possible temporary derogation from the maximum level fumonisins in maize and maize products has been published (EFSA Journal 2014a).

Some guidance values for the sum of fumonisin B1 and 2 in feed materials have been proposed in an EU Commission Recommendation (2006/576/EC).

It has been reported that *A. niger* strains produce fumonisins B2, B4, and B6 but not B1 (Frisvad *et al.*, 2011) (Pel *et al.*, 2007; Fanelli *et al.*, 2012; Sørensen *et al.*, 2009). In this revision papers relating *A. niger* strains and fumonisin B1 production have not been found. Mogensen *et al.* (2009) found that *A. niger* had the highest production of FB2 was at 25-30°C in PDA agar and the addition of 2.5-5% NaCl, or 10-20% sucrose increased the production of the toxin but glycerol has an opposite effect.

It has been reported that some of the most frequently used strains in industry, *A. niger* NRRL 337, 3112 and 3122 produced fumonisins and several strains used for citric acid production were among the highest producers of fumonisins in pure agar culture. Most strains used for other biotechnological processes also produced fumonisins. These mycotoxins can be produced under conditions of citric acid fermentation so it is recommended to use strains of *A. niger* with inactive or inactivated gene clusters for fumonisins and ochratoxins (Frisvad *et al.*, 2011).

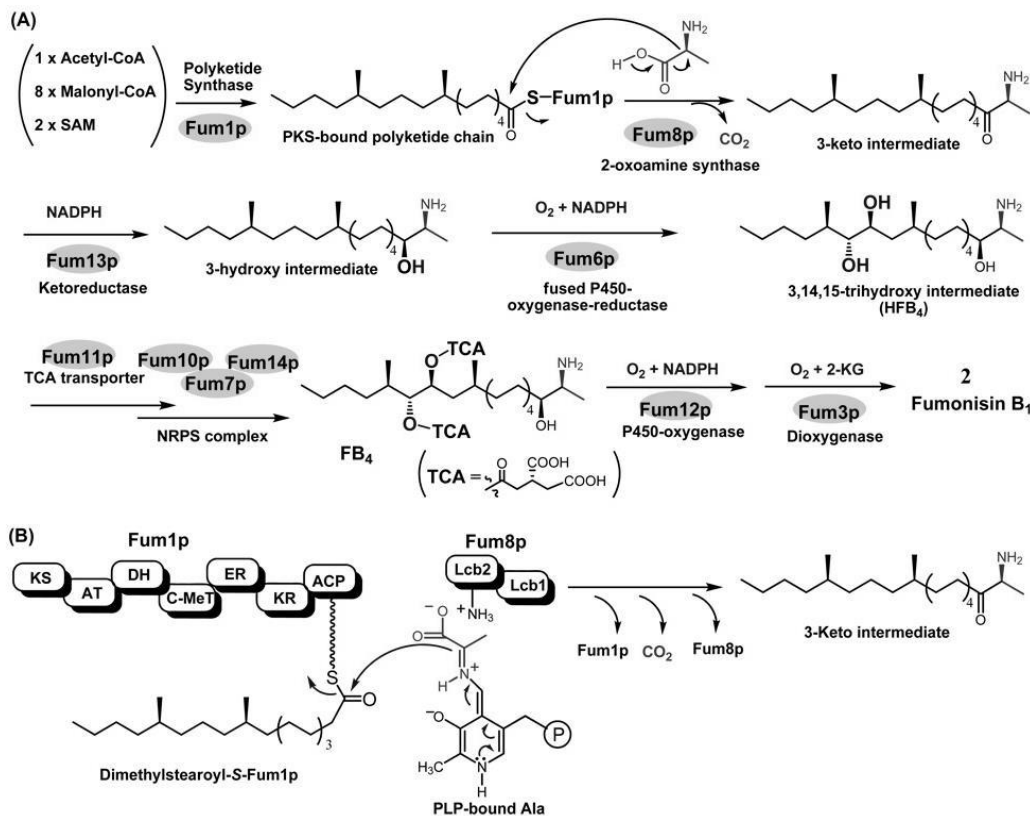


Figure 25: Biosynthetic mechanism for fumonisins. A, the biosynthetic pathway for FB1; B, the PLP-dependent polyketide chain-releasing mechanism in fumonisin biosynthesis (from Huffman J *et al.*, 2010)

Fusarin C

Fusarin C is a mycotoxin produced by several *Fusarium* species. Fusarin C, like other mycotoxins produced by *F. moniliforme* was classified as possible carcinogenic to humans (Group 2B) by the IARC

(1996). It gave positive results in several genotoxicity assays and was able to induce induced papilloma and carcinoma of the oesophagus and forestomach in mice and rats (IARC, 1996). The biosynthesis of fusarin C has been partially resolved and is performed by a polyketide synthase/nonribosomal peptide synthetase system.

In this review, it has been found a report elucidating the biosynthesis of Fusarin C by the species *F. venenatum*. A homologous gene of *fusA* in *F. moniliforme* is responsible for the early stages of fusarin biosynthesis in *F. venenatum* (Song *et al.*, 2004).

Ochratoxin A

Ochratoxin A (OTA) is a nephrotoxic mycotoxin, potentially carcinogenic, hepatotoxic, and teratogenic toxin with immunosuppressive activities. It is produced by several fungal species in the *Penicillium* and *Aspergillus* genera, primarily *P. verrucosum*, *A. ochraceus* and *Aspergilli* of the section *Nigri*, including *A. niger*.

OTA is classified by the International Agency for Research on Cancer (IARC) as possibly carcinogenic to humans (group 2B), based on sufficient evidence for carcinogenicity in animal studies and has been considered as reasonably anticipated to be a human carcinogen by the NTP (National Toxicology Program, 2016). It has been evaluated by EFSA in 2004 and 2006 (EFSA Journal 2004; EFSA Journal 2006).

The European Regulation (EC) 1881/2006 and its modifications, sets the maximum levels for OTA in cereals, vine fruit, coffee, and certain spices. Some guidance values have been proposed for products intended for animal feeding (EU Commission Regulation 2006/576/EC and its modifications).

Expression of the *pks* gene appears to be correlated with OTA production in *A. niger*; therefore this finding provides data to find a route for avoiding the production of OTA in industrial strains. The genome sequencing of *A. niger* strain CBS 513.88 revealed the presence of a *pks* gene An15g07920 that has a strong similarity to the *pks* gene of *A. ochraceus* involved in OTA biosynthesis. It has been reported that a fragment of this gene is specific for ochratoxigenic strains of *A. niger* and therefore deletion of this fragment should be sought for avoiding OTA in industrial processes (Shinohara *et al.*, 2011).

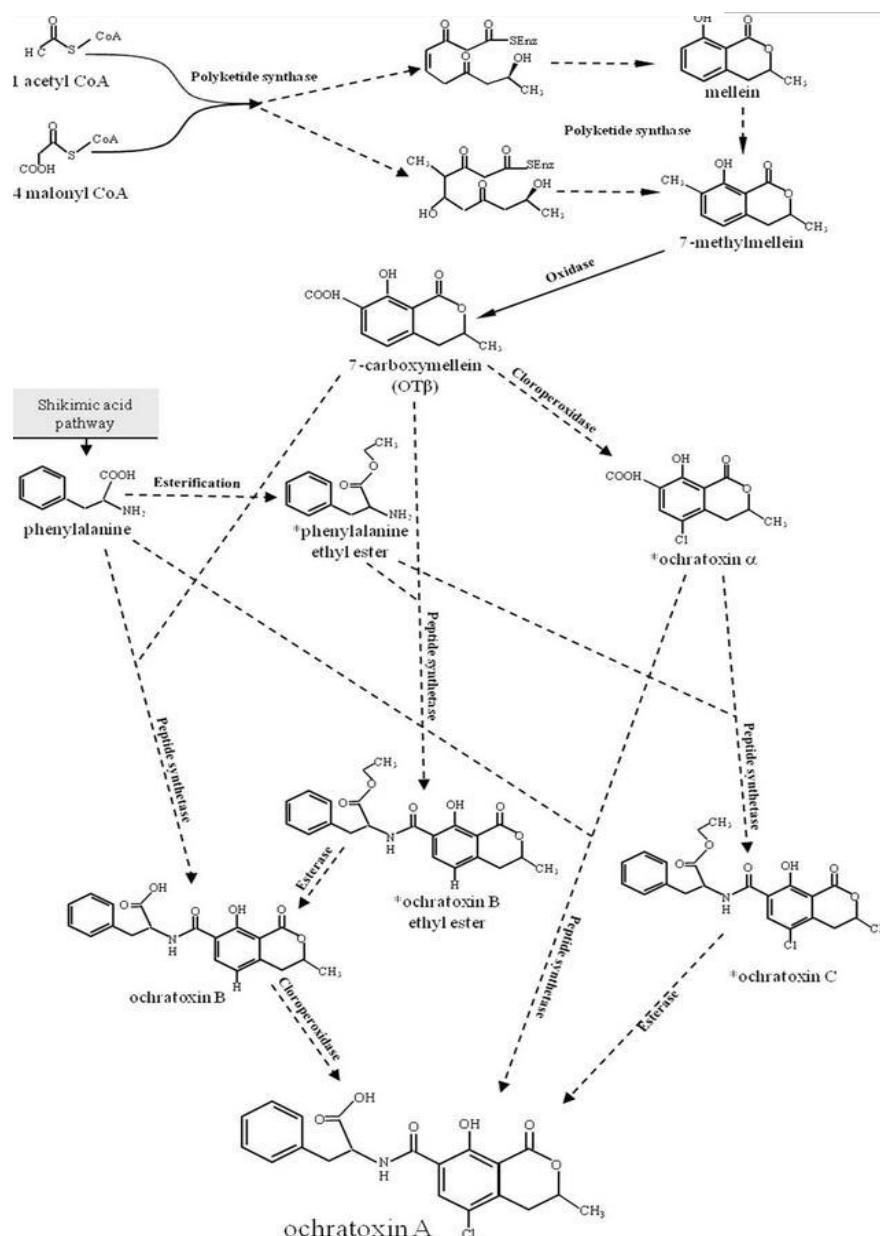


Figure 26: Scheme showing all the different hypotheses of the OTA biosynthesis pathway according to the literature data available, hypothetical intermediary compounds (Gallo *et al.*, 2012)

Regarding the production of OTA by *P. nordicum*, there is a study that shows the effect of sodium chloride on the regulation of ochratoxin A biosynthesis in *Penicillium* species. High amounts of ochratoxin A are produced by *P. nordicum* over a wide concentration range of NaCl (5 to 100 g/L), with an optimum at about 20 g/L on YES medium (Schmidt-Heydt and Geisen, 2007).

Patulin

Patulin is a toxic chemical contaminant produced by several species of mold, especially within *Aspergillus* and *Penicillium*. It is the most common mycotoxin found in apples and apple-derived products such as juice, cider, compotes and other food intended for young children. Exposure to this

mycotoxin is associated with immunological, neurological and gastrointestinal outcomes (Puel *et al.*, 2010).

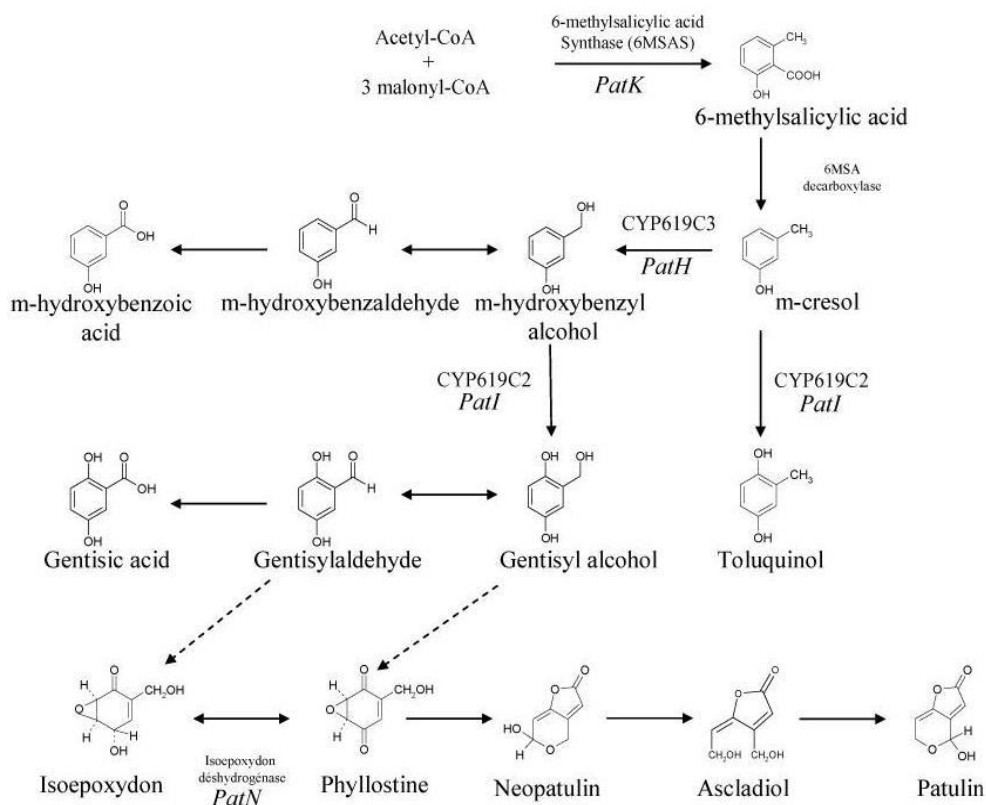


Figure 27: Scheme of patulin biosynthetic pathways. From Puel O *et al.*, 2010.

Penicillic acid

Penicillic acid (PA) is a polyketide mycotoxin produced by several species of *Aspergillus* and *Penicillium*. This mycotoxin is toxic in experimental animals and has also been reported to be carcinogenic. It can cause generalized hepatic necrosis in the mouse and cytotoxicity in cultured cells, including hepatocytes.

The cytotoxicity of penicillic acid was studied in rat alveolar macrophages (AM) in vitro (Sorenson *et al.*, 1986). The data demonstrate that penicillic acid is toxic to rat alveolar macrophages in vitro and suggest the possibility of a respiratory hazard to agricultural workers exposed to contaminated grain. This mycotoxin was also evaluated for its toxicity in broiler chickens (Huff *et al.*, 1980). The data suggest that penicillic acid by itself has little toxicity in chickens (less than 1% compared with aflatoxin).

Sterigmatocystin

Sterigmatocystin is a mycotoxin produced mainly by *Aspergillus* fungi, and is an intermediate in the biosynthesis of aflatoxin B1. Major producing microbes include *A. versicolor* and *A. nidulans*.

The structure and the bioactivity of sterigmatocystin are similar to those of aflatoxin, but its toxicity is weaker than aflatoxin and is observed in the liver and the lung. Its toxicity is said to be 1/125 and the carcinogenicity to be 1/250 of aflatoxin B1, and its acute toxicity is low (The Pharmaceutical Society of Japan: Methods of Analysis in Health Science, 2010). Carcinogenicity is reported in animal studies.

Sterigmatocystin has been detected in feed materials including maize, wheat, barley, milo, cassava, corn gluten feed, corn gluten meal, bran, and soybean meal.

Sterigmatocystin is classified as Group 2B with possible carcinogenicity in humans by the International Agency for Research on Cancer (IARC: Summaries & Evaluations, 10 (1987)). Effects of the toxin on human health are not well known. It has been reported that it causes apoptosis in human peripheral lymphocytes (Sun *et al.*, 2002).

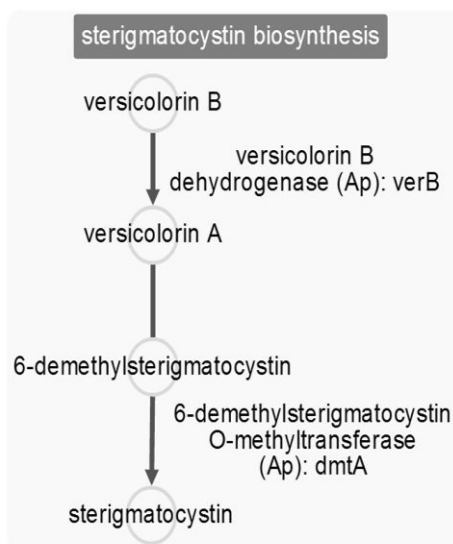


Figure 28: Biosynthetic pathway of sterigmatocystin. From Caspi *et al.*, 2014, "The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of Pathway/Genome Databases," *Nucleic Acids Research* 42:D459-D471.

Trichothecenes

Trichothecenes are one of the major classes of mycotoxins, produced by several fungi genera including *Fusarium*, *Myrothecium*, *Spicellum*, *Stachybotrys*, *Cephalosporium*, *Trichoderma*, and *Trichothecium* and are potent inhibitors of eukaryotic protein synthesis.

Trichothecenes are a family of over 200 toxins with a common tricyclic 12,13-epoxytrichothec-9-ene (EPT) core structure that have been classified into four groups (Types A, B, C, and D).

Type A trichothecenes include **T-2 toxin**, neosolaniol, **trichodermin**, **diacetoxyscirpenol (DAS)**, and harzianum A. In this review, several reports on the production of these toxins by the microorganisms within the scope of the review, such as *F. venenatum* (DAS) (Miller and Mackenzie, 2000); *T. viride* (DAS and T-2 toxin) and *T. harzium* and *T. reesei* (Trichodermin) (Mohamed *et al.*, 2006; Blumenthal, 2004)

Type B trichothecenes include nivalenol, **deoxynivalenol (DON)**, that can be produced by *A. oryzae* (Atalla *et al.*, 2003), and trichothecin.

Type C trichothecenes include crotocin.

Type D trichothecenes include roridin A, verrucarin A, satratoxin H.

Some of these compounds have been evaluated by IARC as "toxins derived from *Fusarium*" and have been classified as group 3 (not classifiable as to its carcinogenicity to humans) (IARC, 1993).

The European Regulation (EC) 1881/2006 and its modifications, sets the maximum levels for certain types of trichothecenes, such DON, ZEA and T-2 and HT-2.

Zearalenone

Zearalenone (ZEA) is a mycotoxin produced mainly by fungi belonging to the genus *Fusarium* in foods and feeds. It is frequently implicated in reproductive disorders of farm animals and occasionally in hyperoestrogenic syndromes in humans. There is evidence that ZEA and its metabolites possess oestrogenic activity in pigs, cattle and sheep. However, ZEA is of a relatively low acute toxicity after oral or interperitoneal administration in mice, rat and pig. Although ZEA is toxic, it globally presents a potential danger for animal and human health only when it is absorbed in high amounts or over a long time of exposure (Zinedine *et al.*, 2007)

The Joint FAO/WHO Expert Committee on Food Additives (JECFA) 2016 established a provisional maximum tolerable daily intake (PMTDI) for ZEA of 0.5 µg/kg of body weight.

3.3.2. Toxic Secondary metabolites produced by bacteria

Potentially toxic substances produced by bacteria that have not been fully evaluated from the toxicological point of view have been identified in this review. The majority are associated with virulence factors, such as signaling molecules (that are only elicited in the presence of competitors), or adhesion proteins that allow bacteria the attachment, favoring the formation of biofilms and therefore, providing more resistance to external aggression. Moreover, information on possible effects of the LPS (lipopolysaccharide endotoxins), released by the microorganism species of the scope of the project, would need to be taken into account as relevant information.

A short description of the most relevant compounds synthesized by bacteria within the scope of this project is given below.

Cytolysin A

Cytolysin A (ClyA) is a hemolytic protein of *Escherichia coli* K-12, they damage host cells by forming holes in the cytoplasmic membrane, which may result in cell death and osmotic cell lysis. It has been reported that this pore-forming cytotoxic protein encoded by the *clyA* gene protein, is not expressed under normal laboratory conditions. Genetic analysis suggested that *clyA* is silenced by the nucleoid protein H-NS. Purified H-NS protein showed preferential binding to *clyA* sequences in the promoter region.

In this review, it has been found a study that test the purified cytotoxic of ClyA and a ClyA-expressing *E. coli* strain to both human and murine macrophages, showing the induction of a massive amount of apoptosis and host cell DNA fragmentation (Enow *et al.*, 2014; Ludwig *et al.*, 2010; Oscarsson *et al.*, 1999)

Lipopolysaccharide endotoxins

Endotoxin refers to the lipopolysaccharide (LPS) that constitutes the outer leaflet of the outer membrane of most Gram-negative bacteria. The biological activity of endotoxin is associated with the LPS. Toxicity is associated with the lipid component (Lipid A) and immunogenicity is associated with the polysaccharide components. Both Lipid A (the toxic component of LPS) and the polysaccharide side chains (the nontoxic but immunogenic portion of LPS) act as determinants of virulence in Gram-negative bacteria (Table 4).

Table 4: Characteristics of bacterial endotoxins.

| PROPERTY | ENDOTOXIN |
|----------------------|-------------------------|
| CHEMICAL NATURE | Lipopolysaccharide |
| RELATIONSHIP TO CELL | Part of outer membrane |
| DENATURED BY BOILING | No |
| ANTIGENIC | Yes |
| FORM TOXOID | No |
| POTENCY | Relatively low (>100ug) |
| SPECIFICITY | Low degree |
| ENZYMATIC ACTIVITY | No |
| PYROGENICITY | Yes |

Most enzymes and genes related to the biosynthesis and export of lipopolysaccharide have been identified in *E. coli*, and they are shared by most Gram-negative bacteria based on available genetic information. However, the detailed structure of lipopolysaccharide differs from one bacterium to another, suggesting that additional enzymes that can modify the basic structure of lipopolysaccharide exist in bacteria, especially some pathogens. These structural modifications of lipopolysaccharide are sometimes tightly regulated. They are not required for survival but closely related to the virulence of bacteria.

Table 5: Information on nine enzymes required for the biosynthesis of Lipid A in *E. coli* (from Wang and Quinn, 2010).

| ENZYME | GENE | FUNCTION |
|--------|-------------|-----------------------|
| LpxA | <i>lpxA</i> | Acyltransferase |
| LpxC | <i>lpxC</i> | Deacetylase |
| LpxD | <i>lpxD</i> | Acyltransferase |
| LpxH | <i>lpxH</i> | Pyrophosphatase |
| LpxB | <i>lpxB</i> | Disaccharide synthase |
| LpxK | <i>lpxK</i> | 4'-Kinase |
| KdtA | <i>kdtA</i> | Kdo transferase |
| LpxL | <i>lpxL</i> | Acyltransferase |
| LpxM | <i>lpxM</i> | Acyltransferase |

The first three enzymes LpxA, LpxC and LpxD of the LPS biosynthetic pathway have been purified, and their structures have been characterised by X-ray diffraction and NMR methods. Based on the structural information from these proteins, research into developing new antibiotics is being carried out.

LPS from well-known species can cause diseases such as septic shock, multiple organ dysfunction and failure. Understanding the biochemistry of LPS modifications and their impact on pathogenesis could lead to novel treatment options for these diseases.

Phenazines

Phenazines are pigmented, redox-active, heterocyclic, nitrogen containing molecules secreted by a considerable number of *Streptomyces*, (Sarmin *et al.*, 2013) and multiple fluorescent *Pseudomonas* (Cezairliyan *et al.*, 2013). Several derivatives have been found in this review, such as **1-Hydroxyphenazine**, **Phenazine-1-carboxylic acid** and **Endophenazine A**.

Phenazines display a broad spectrum of (toxic) activity toward prokaryotic and eukaryotic organisms, varying according to the nature and position of the substituent on the heterocyclic ring. They are considered as virulent factor of *P. aeruginosa* (Ballok and O'Toole, 2013), but no standard toxicity data have been found in this report for these substances. Phenazines are considered redox-active compounds as well as a cytotoxic pigment. Pyocyanin, (1-hydroxy-5-methylphenazine) is the best-studied natural phenazine.

In this review it has been found that several of the articles related to virulence of *P. aeruginosa*, do not include toxicological evaluation. Others are related with cystic fibrosis lung (patients are specially affected by *P. aeruginosa* infection).

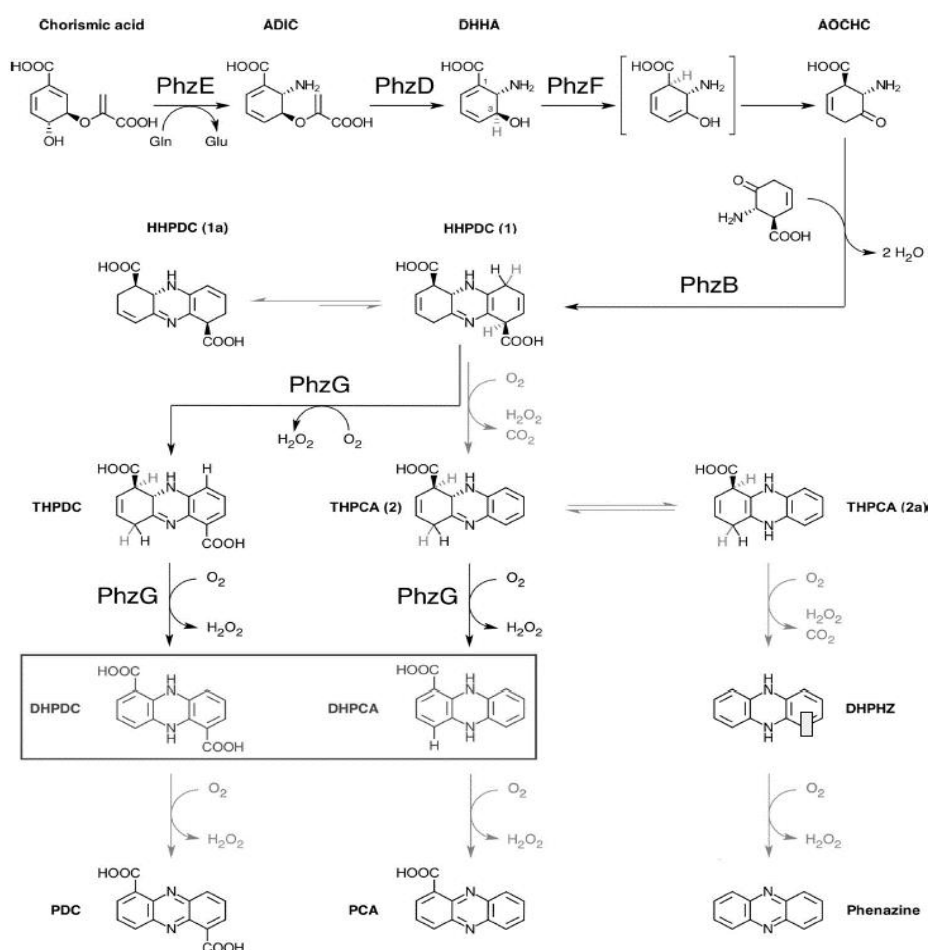


Figure 29: Enzime biosynthesis. Abstracted or shifted hydrogens are shown in red, grey arrows indicate uncatalyzed steps. 5,10-Dihydro-PCA and 5,10-dihydro-PDC are the final products of the pathway (green) (from Blankenfeldt and Parsons, 2014)

Pyocyanin

Pyocyanin (5-*N*-methyl-1-hydroxyphenazine), the first and most-studied member of the phenazine family,. Phenazines are low-molecular-weight secondary metabolites including nitrogen-containing heterocyclic pigments. Almost all phenazines exhibit broad spectrum activity against plant pathogenic bacteria and fungi.

Pyocyanin is produced only by *P. aeruginosa* and 90 to 95% of *P. aeruginosa* strains produce pyocyanin. This specificity that has been useful in the rapid diagnosis of this opportunistic pathogen.

Pyocyanin produced in the rhizosphere of plants plays an important role in the biological control activity of *P. aeruginosa* against *Fusarium* wilt of chickpea and *Pythium* damping off of bean.

This blue phenazine is one of the major virulence factors in this pathogen, contributing to both acute and chronic infections. It suppresses lymphocyte proliferation, damages epithelial cells as a consequence of hydroxyl radical formation, inactivates protease inhibitors (consequently causing tissue damage by endogenous proteases), and targets multiple cellular functions.

The redox-active pyocyanin (PCN) secreted by the respiratory pathogen *P. aeruginosa* generates reactive oxygen species (ROS) and causes oxidative stress to pulmonary epithelial cells.

Pyocyanin is able to produce oxidative stress damage in mammalian tissues. Several toxicological endpoints were detected in mechanistic studies (related mainly with sepsis and bacteria pathogenicity/virulence): neurotoxicity, haemotoxicity, inhibition of ATPase and immunotoxicity (neutrophil extracellular traps promotion).

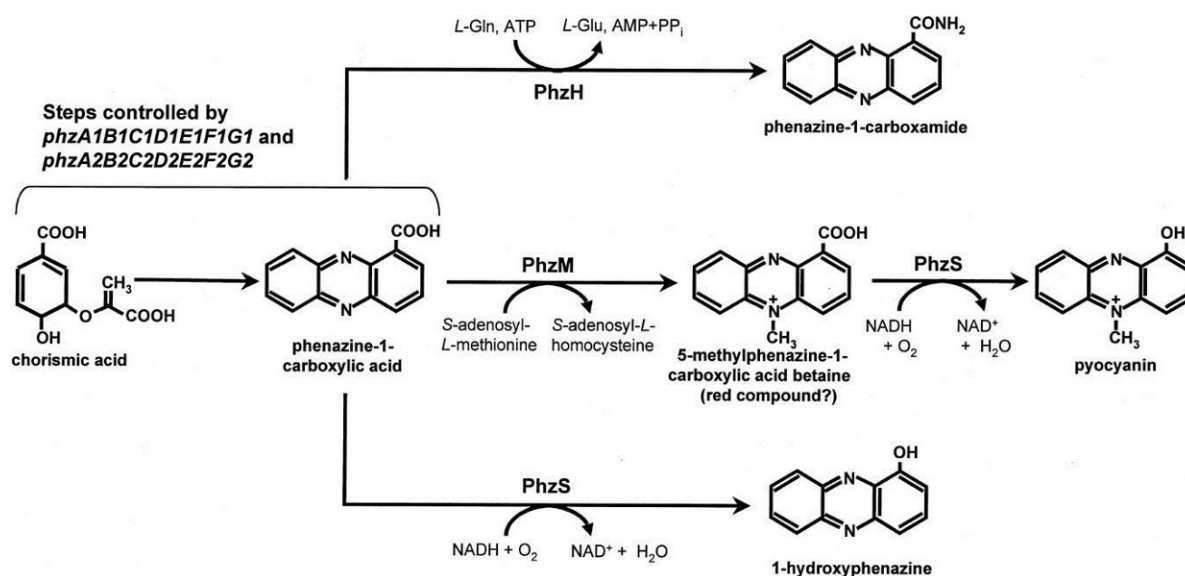


Figure 30: Proposed mechanism for the synthesis of pyocyanin, 1-OH-PHZ, and PCN in *P. aeruginosa* PAO1 (from Mavrodi *et al.*, 2001).

4. Proposal for the sustainability of the database

As a result of this project, different methodologies, studies and technologies have been developed and integrated, as described below:

- a. A well-defined searching strategy proved to be suitable for the purpose of this project that included sets of suitable keywords identified in the areas of toxicology, industrial

- fermentation as well as biosynthetic pathways, which can help to retrieve useful information in future searches.
- b. A customized technology based on a collaboration platform (iWatch) capable of extracting information from different web databases. This platform provides two tools: a crawler tool implemented and configured to collect and classify all documents as required by the search strategy and a user interface parameterized to implement the working methodology. Both allow continuing with extraction of information efficiently.
 - c. A working methodology based on inclusion/exclusion criteria to allow experts the review, the analysis and classification of all extracted documents. This can be useful to continue efficiently with the same workflow in future searches.
 - d. A validated Database to store all data extracted by experts regarding taxonomy, process conditions, biosynthetic pathways and toxicology on non-QPS microorganisms. Also a tool to extract, convert and store the information retrieved in collaboration with the platform (iWatch) integrated into the database. All can be useful to continue efficiently updating the database with new information included by the experts.
 - e. A report on more than 400 bioactive secondary metabolites produced by industrial microorganisms have been identified and recorded. Most of these compounds have been reported in relation to some metabolic aspects and studies related to their toxicity were not found, and therefore only a small part of the identified compounds are included in the database, however, new information could be gathered in the future and the name of this compounds could be used as new keywords for future searches.

Considering the above aspects, the sustainability of the database can be ensured by updating it periodically (every year) maintaining the same structure.

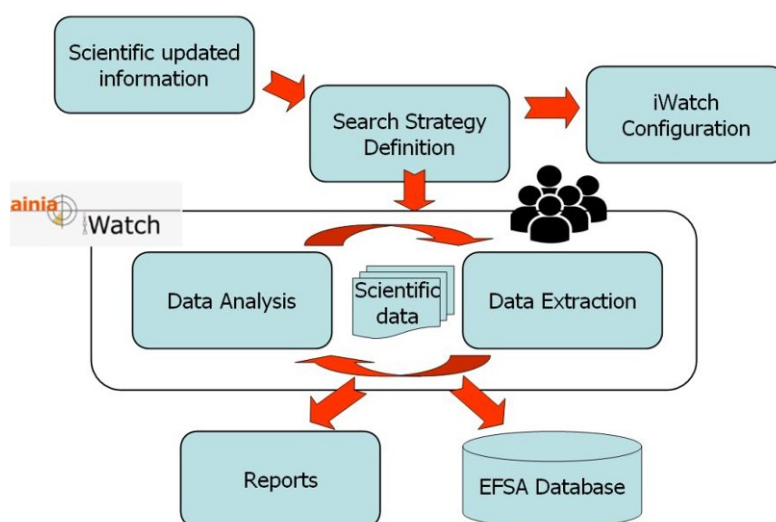


Figure 31: Sustainability of the EFSA Database.

5. Conclusions

In the present project, it has been developed an integrative and useful database for supporting safety assessment that includes relevant and accurate information about taxonomical characterisation and potential toxigenic capacities of microorganisms used for the industrial production of feed additives and food enzymes. The structure of the data model and its terminology includes the use of EFSA standard coding system.

The data model has been aimed to include the following information:

- Description of microbial species used to produce food enzymes and feed additives which have not been recommended for Qualified Presumption of Safety (QPS) status, with its main identifiers: the current scientific name, taxon assignation, synonyms, existing methods for the taxonomical characterisation and accession numbers for housekeeping genes.
- Identification of toxins or potentially toxic secondary metabolites/substances produced by the microorganisms used to produce food enzymes and feed additives
- Identification of the conditions under which the microorganisms can produced the toxic compound/s.
- Biosynthetic pathway and genetic characterization of the toxic compounds produced by the microorganism.

The contents of the database have been retrieved by the performance of a systematic search that has been aided by the use of an "ad hoc" automatic tool.

For the microbial taxonomic review, a total of 123 scientific documents have been obtained to extract the relevant information for the database. For the identification of secondary metabolites, 22970 scientific documents have been recovered combining the searching platform and manual searching, classified by microorganism species names and systematically revised, to obtain pertinent data. A total of 411 scientific documents were selected that contain relevant data on potentially toxic secondary metabolites.

A total of 474 bioactive secondary metabolites were found to be produced by the microbial species within the scope of this project, from which 59 were selected for further searches for data on their toxicity and biosynthesis. Fungal species are the most predominant group to produce toxic metabolites, including well known mycotoxins. However, in the majority of the cases, industrial strains contain a natural or artificially induced safeguard mutation or genetic modification which prevents the production of the toxin under defined fermentation conditions.

The majority of secondary metabolites produced by fungi found in this review are studied for their biotechnological and biopharmaceutical potential due to their properties such as antimicrobial, antinematode, antitumor, etc.

Reports on secondary metabolites produced by bacterial species within the scope of this project were less common and in most of the cases those metabolites are identified as virulent factors, expressed only under certain conditions

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Abbreviations

| | |
|-----------------|--|
| AEMPS | Spanish Agency of Medicines and Medical Devices |
| AESAN | Agencia española de Consumo, Seguridad alimentaria y Nutrición. (Spanish Agency for Consumer Affairs, Food Safety and Nutrition) |
| AFSCA | Federal Agency for the safety of the food chain |
| AGES | Austrian Agency for Health and Food Safety |
| AHVLA | Animal Health and Veterinary Laboratories Agency |
| AINIA | Asociación Investigación Industria Agroalimentaria (Food Industry Research Association) |
| ANSES | French Agency for Food, Environmental and Occupational Health & Safety |
| BEUC | Bureau Européen des Unions de Consommateurs (The European Consumer Organisation) |
| BFR | The Federal Institute for Risk Assessment |
| CAS | (Number) Chemical Abstract Service |
| CCRIS | Chemical Carcinogenesis Research Information System |
| CDC | Centers for Disease Control and Prevention |
| CELCAA | European Liaison Committee for Agricultural and Agri-Food Trade |
| CFIA | Canadian Food Inspection |
| CIAA | Confederation of the Food and Drink Industries in the EU |
| ClyA | Cytolysin A |
| COPA- COGECA | European Farmers - European Agri-Cooperatives |
| CSPINET | Center for Science in the Public interest |
| DART | Developmental and Reproductive Toxicology Database |
| DAS | Diacetoxyscirpenol |
| DBMS | Database Management System |
| DCF | Data Collection Framework (EFSA) |
| DNA | Deoxyribonucleic acid |
| DON | Deoxynivalenol |
| EC | European Commission |
| ECDC | European Centre for Disease Prevention and Control |
| EFI | European Food Institutes |
| EFSA | European Food Safety Authority |
| EPHA | European Public Health Alliance |

| | |
|--------|---|
| EU | European Union |
| EUFIC | European Food Information Council |
| FAO | Food and Agriculture Organization of the United Nations |
| FDA | U S Food and Drug Administration |
| FEEDAP | Panel on Additives and Products or Substances used in Animal Feed |
| FEFAC | European Feed Manufacturers Federation |
| FEFANA | EU Association of Speciality Feed Ingredients and their Mixtures |
| FK | Foreign key |
| FSA | Food Standards Agency (UK) |
| FSAI | Food Safety Authority of Ireland |
| FSANZ | Food Standard Australia New Zealand |
| HPA | Hydroxypropionaldehyde |
| HSDB | Hazardous Substances Data Base |
| ICAR | International Agency for research on Cancer |
| ID | Identification |
| iNOS | Nitric oxide synthase |
| ISO | International Organization for Standardization |
| JECFA | Joint FAO/WHO Expert Committee on Food Additives |
| LBSN | List of Bacterial names with Standing in Nomenclature |
| LPS | Lipopolysaccharide |
| NTP | National Toxicology Program |
| OTA | Ochratoxin A |
| PSK | polysaccharide-K |
| QMS | Quality Management System |
| QPS | Qualified Presumption of safety |
| RASFF | Food and Feed Safety Alerts - European Commission |
| SICURA | Spanish Network for Food Security |
| SQL | Structured Query Language |
| TOXNET | TOXicology Data NETwork |
| URL | Uniform Resource Locator |
| USDA | U.S. Department of Agriculture |
| USEPA | US Environmental Protection Agency |
| VWA | The Netherlands Food and Consumer Product Safety Authority |

Appendices

Appendix A – Search in Iwatch: number of entries per microorganism (annexed)

Appendix B – Websites and Databases list for automatic search by IWatch (annexed)

Appendix C – Additional toxicological information databases (annexed)

Appendix D – List of food and feed additives (annexed)

Appendix E – List of toxins evaluated by EFSA, IARC or NTP or included in Directive 2002/32/EC

Appendix F – List of excluded articles. Level 2 (annexed)

Appendix G – Summary of secondary metabolites with bioactivity (annexed)

Appendix H – Relational database (annexed)

Appendix I – Attributes for Param Catalogue (annexed)

Appendix J – Microorganism taxonomical identifiers (annexed)

Appendix K – Taxonomy references (annexed)

Appendix A – Search in IWatch: number of entries per microorganism

| MICROORGANISMS CLASSIFICATION | | | | | | | |
|---|-----------|------------|-------------|--|-----------|------------|-------------|
| SEARCH KEYWORDS | | | | | | | |
| Species I: scientific name of the microorganism | | | | | | | |
| Species II: scientific name of the microorganism AND (keywords group 1) | | | | | | | |
| Species III: scientific name of the microorganism AND (keywords group 1) AND (keywords group 2) | | | | | | | |
| MICROORGANISMS | SEARCH | | | MICROORGANISMS | SEARCH | | |
| | Species I | Species II | Species III | | Species I | Species II | Species III |
| <i>Actinomadura roseorufa</i> | 0 | - | - | <i>Penicillium chrysogenum</i> | - | 446 | - |
| <i>Actinomadura yumaensis</i> | 5 | - | - | <i>Penicillium citrinum</i> | - | 236 | - |
| <i>Aeribacillus pallidus</i> (Syn. <i>Geobacillus pallidus</i>) | 17 | - | - | <i>Penicillium decumbens</i> | 145 | - | - |
| <i>Arthrobacter ramosus</i> | 12 | - | - | <i>Penicillium funiculosum</i> | 313 | - | - |
| <i>Aspergillus aculeatus</i> | - | 31 | - | <i>Penicillium lilacinum</i> | 16 | - | - |
| <i>Aspergillus japonicus</i> | 173 | - | - | <i>Penicillium multicolor</i> | 32 | - | - |
| <i>Aspergillus melleus</i> | 57 | - | - | <i>Penicillium notatum</i> | - | 118 | - |
| <i>Aspergillus niger</i> | - | - | 1186 | <i>Penicillium roqueforti</i> | - | 112 | - |
| <i>Aspergillus oryzae</i> | - | 523 | - | <i>Penicillium salamii</i> | 3 | - | - |
| <i>Aspergillus sojae</i> | 156 | - | - | <i>Pseudomonas aeruginosa</i> | - | - | 9426 |
| <i>Bacillus macerans</i> | 237 | - | - | <i>Pseudomonas amyloclavata</i> | 14 | - | - |
| <i>Bacillus naganensis</i> | 12 | - | - | <i>Pullulanibacillus naganensis</i> | 6 | - | - |
| <i>Bacillus soli</i> (syn. <i>Paenibacillus macerans</i>) | 8 | - | - | <i>Rhizomucor miehei</i> | - | 38 | - |
| <i>Bullera singularis</i> (syn. <i>Sporobolomyces singularis</i>) | 6 | - | - | <i>Rhizopus niveus</i> | 92 | - | - |
| <i>Candida cylindracea</i> (syn. <i>Candida rugosa</i>) | - | 274 | - | <i>Rhizopus oryzae</i> | - | 330 | - |
| <i>Candida lipolytica</i> | - | 39 | - | <i>Sphingobacterium multivorum</i> | 58 | - | - |
| <i>Candida paralipolytica</i> (syn. <i>Candida lipolytica</i>) | - | 1 | - | <i>Sporobolomyces singularis</i> | 11 | - | - |
| <i>Candida rugosa</i> | - | 135 | - | <i>Sporotrichum dimorphosporum</i> | 1 | - | - |
| <i>Cellulosimicrobium cellulans</i> | 77 | - | - | <i>Streptomyces albus</i> | - | 41 | - |
| <i>Chaetomium erraticum</i> | 8 | - | - | <i>Streptomyces aureofaciens</i> | - | 24 | - |
| <i>Chaetomium gracile</i> | 9 | - | - | <i>Streptomyces chrestomyceticus</i> | 3 | - | - |
| <i>Chryseobacterium proteolyticum</i> | 8 | - | - | <i>Streptomyces chromofuscus</i> | 85 | - | - |
| <i>Cryphonectria parasitica</i> | - | 370 | - | <i>Streptomyces cinnamomensis</i> | 43 | - | - |
| <i>Disporotrichum dimorphosporum</i> | 1 | - | - | <i>Streptomyces cinnamomeus</i> | 14 | - | - |
| <i>Endothia parasitica</i> (syn. <i>Cryphonectria parasitica</i>) | - | 21 | - | <i>Streptomyces lasaliensis</i> | 8 | - | - |
| <i>Escherichia coli</i> | - | - | 778 | <i>Streptomyces lividans</i> | - | 89 | - |
| <i>Flavobacterium multivorum</i> | 9 | - | - | <i>Streptomyces mobaraensis</i> | 70 | - | - |
| <i>Fusarium venenatum</i> | 56 | - | - | <i>Streptomyces murinus</i> | 13 | - | - |
| <i>Geobacillus caldoproteolyticus</i> | 2 | - | - | <i>Streptomyces netropsis</i> | 10 | - | - |
| <i>Geobacillus pallidus</i> | 26 | - | - | <i>Streptomyces olivochromogenes</i> | 17 | - | - |
| <i>Geobacillus stearothermophilus</i> | - | 173 | - | <i>Streptomyces rubiginosus</i> | 35 | - | - |
| <i>Hansenula polymorpha</i> | - | 132 | - | <i>Streptomyces violaceoruber</i> | 40 | - | - |
| <i>Humicola insolens</i> | 254 | - | - | <i>Streptoverticillium cinnamomeum</i> (syn. <i>Streptomyces cinnamomeus</i>) | 77 | - | - |
| <i>Klebsiella planticola</i> | 76 | - | - | <i>Streptoverticillium mobaraense</i> (syn. <i>Streptomyces mobaraensis</i>) | 57 | - | - |
| <i>Klebsiella pneumoniae</i> | - | - | 3436 | <i>Talaromyces cellulolyticus</i> | 16 | - | - |
| <i>Leptographium procerum</i> | 44 | - | - | <i>Talaromyces emersonii</i> | 89 | - | - |
| <i>Microbacterium imperiale</i> | 20 | - | - | <i>Talaromyces versatilis</i> | 6 | - | - |
| <i>Micrococcus luteus</i> | - | - | 290 | <i>Trametes hirsuta</i> | 291 | - | - |
| <i>Mucor circinelloides</i> | 107 | - | - | <i>Trametes versicolor</i> | - | 232 | - |
| <i>Mucor javanicus</i> | 97 | - | - | <i>Trichoderma citrinoviride</i> | 43 | - | - |
| <i>Ogataea polymorpha</i> (syn. <i>Hansenula polymorpha</i>) | - | 9 | - | <i>Trichoderma harzianum</i> | - | 950 | - |
| <i>Paecilomyces lilacinus</i> (syn. <i>Penicillium lilacinum</i>) | 275 | - | - | <i>Trichoderma koningii</i> | 213 | - | - |
| <i>Paenibacillus alginolyticus</i> | 8 | - | - | <i>Trichoderma longibrachiatum</i> | 285 | - | - |
| <i>Paenibacillus lentus</i> | 1 | - | - | <i>Trichoderma reesei</i> | - | 196 | - |
| <i>Paenibacillus macerans</i> | 107 | - | - | <i>Trichoderma viride</i> | - | 435 | - |
| <i>Penicillium camemberti</i> | 73 | - | - | <i>Yarrowia lipolytica</i> | - | 205 | - |

Appendix B – Websites and Databases list for automatic search by Iwatch

| NAME | URL | DESCRIPTION |
|------------------------------------|---|---|
| EFE | http://www.efe.com/efe/noticias/english/4 | EFE Agency |
| Agrodigital | www.agrodigital.com | Agrodigital |
| Agromeat - Agronews | http://www.agromeat.com | Agromeat - Agronews website |
| AHVLA | http://www.defra.gov.uk/ahvla-en | Animal Health and Veterinary Laboratories Agency |
| Albeitar | http://albeitar.portalveterinaria.com/ | Veterinary website |
| ANSES FRANCE | https://www.anses.fr/en | National Food Agency France |
| Baden-Wuerttemberg | http://www.cvuas.de/pub/default.asp?subid=1&lang=EN | Investigative office for food control and animal health |
| BFR | http://www.bfr.bund.de/en/home.html | The Federal Institute for Risk Assessment |
| CDC (USA) | http://www.cdc.gov/niosh/nioshtic-2/20023037.html | Center of Disease Control (USA) |
| CFIA | http://www.inspection.gc.ca/about-the-cfia/newsroom | Canadian Food Inspection Agency |
| Chemical Watch | http://chemicalwatch.com | Chemical Watch |
| CIDRAP | http://www.cidrap.umn.edu/cidrap/index.html | Center for Infectious Disease Research and Policy |
| ClubDarwin | http://www.clubdarwin.net/ | ClubDarwin |
| CODEX Alimentarius | http://www.codexalimentarius.org/ | Codex Alimentarius Reports |
| EDIPORC | http://www.ediporca.com/ | Technical Farming Editions |
| EFEAgro | http://www.efeaagro.com/ | Agrofood news |
| EFSA | http://www.efsa.europa.eu/en/newsletters/highlights.htm | European Food Safety Agency reports |
| EHN | http://www.environmentalhealthnews.org | Environmental Health News |
| EHP | http://ehp.niehs.nih.gov/ | Environmental Health Perspectives |
| Espacenet | https://worldwide.espacenet.com/ | European Patent Office |
| Eurekalert | http://www.eurekalert.org/rss.php | Online, global news service operated by AAAS |
| European Comission | http://ec.europa.eu/ | European comission news alert |
| EPHA | http://www.ephac.org/ | European Public Health Alliance |
| FAO | http://www.fao.org | Food and Agriculture Organization of the United Nations |
| FDA | http://www.fda.gov/Safety/Recalls/default.htm | Recalls, Market Withdrawals, & Safety Alerts |
| Food Business Review | http://www.food-business-review.com/ | Food Business Review |
| Food Navigator | http://www.foodnavigator.com/ | Food Navigator |
| FoodNavigator | http://www.foodnavigator.com/ | Daily online news service |
| FoodQuality | http://www.foodqualitynews.com/ | Food Quality |
| FoodSafety | http://www.foodsafety.gov/recalls/recent/index.html | FoodSafety.gov |
| FSA | http://www.food.gov.uk/enforcement/alerts/ | Food Standards Agency UK |
| FSAI | http://www.fsai.ie/news_centre/food_alerts.html | Food Safety Authority of Ireland |
| FSANZ | http://www.foodstandards.gov.au | Food Standards Australia New Zealand |
| GlobalMeatNews | http://www.globalmeatnews.com | Global Meat News |
| Harvard School Public Health | www.hsph.harvard.edu | Harvard School Public Health |
| HSE UK | http://www.hse.gov.uk/index.htm | Health and Safety Executive UK |
| Health Canada | http://www.hc-sc.gc.ca/ | Health Canada |
| HPA | http://www.hpa.org.uk/Topics/InfectiousDiseases/InfectionsAZ | Health Protection Agency |
| IARC | http://www.iarc.fr/en/feeds/index.php | International Agency For Research on Cancer. |
| Infection Control Horizon Scanning | http://infectioncontrolhoriznwpctl.wordpress.com/ | Infection Control |
| JECFA | http://www.fao.org/fao-who-codexalimentarius/codex-home/en/ | International Food Standards |
| Natural News | http://www.naturalnews.com | Natural News |
| New Scientist | http://www.newscientist.com/ | New Scientist |
| Nutraceutical Business Review | http://www.nutraceuticalbusinessreview.com | Nutraceutical Business Review |
| Nutraingredients | http://www.nutraingredients.com | Breaking News on Supplements & Nutrition - Europe |
| OIE | http://www.oie.int/wahis_2/public | Animal Health Information |
| PigProgress | http://www.pigprogress.net | PigProgress |
| Pubmed | http://www.ncbi.nlm.nih.gov/pubmed | Searching Engine |
| RSSL Food e News | http://www.rssl.com/services/food/foode-news | RSSL Food e News |
| ScienceDirect | http://www.sciencedirect.com/ | Searching Engine |
| Science Insider | http://news.sciencemag.org/ | Science Insider |
| Scopus | www.scopus.com | Searching Engine |
| Science Daily | http://www.sciencedaily.com | Science Daily |
| SINC | http://www.agenciasinc.es | Sinc Science |
| Toxnet | http://toxnet.nlm.nih.gov/ | Toxicology data network |
| Unilever | http://www.unilever.com/ | Unilever Resources |
| USDA | http://www.fsis.usda.gov/Fsis_Recalls/index.asp | Food Safety and Inspection Service (FSIS) |
| USEPA | http://www.epa.gov/ | U.S Environmental Protection Agency |
| Web of science | https://apps.webofknowledge.com | Searching Engine |
| WHO | http://www.who.int/csr/don/2013_06_02_ncov/en/index.html | Global Alert and response GAR |
| WIPO | http://www.wipo.int/portal/en/index.html | World Intellectual Property Organization |

Appendix C – Additional toxicological information databases

| Compound | Title | Source. <i>Last update</i> | Information | URL |
|--|---------------------------------------|---|---|---|
| 1-hydroxyphenazine | 1-HYDROXYPHENAZINE CASRN: 528-71-2 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/528-71-2 |
| | | CTD (TOXNET) | genomics | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ctd:@term+@DOCNO+CTD/C050093 |
| 3-nitropropionic acid | 3-NITROPROPIONIC ACID CASRN: 504-88-1 | HSDB (TOXNET). <i>Updated 2003</i> | -- | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+hsdb:@term+@DOCNO+4147 |
| | | CCRIS (TOXNET). <i>Updated 2003</i> | carcinogenicity /genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+454 |
| | | GENE-TOX (TOXNET). <i>Updated 1992</i> | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+genetox:@term+@DOCNO+1519 |
| | | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/504-88-1 |
| | | CPDB (TOXNET) | carcinogenicity / genotoxicity | https://toxnet.nlm.nih.gov/cpdb/chempages/3-NITROPROPIONIC%20ACID.html |
| | | CTD (TOXNET) | genomics | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ctd:@term+@DOCNO+CTD/C015392 |
| | | HAZ-MAP (TOXNET) | occupational | https://hazmap.nlm.nih.gov/category-details?table=copytblagents&id=3470 |
| | | Bioassay of 3-nitropropionic acid for possible carcinogenicity (CAS No. 504-88-1) | NTP | carcinogenicity |
| NTP Assessments 3-Nitropropionic acid - 10620-G. | NTP | general toxicity / genotoxicity / carcinogenicity | http://ntp.niehs.nih.gov/testing/status/agents/ts-10620-g.html | |
| Target Organs and Levels of Evidence for TR-052 | NTP | carcinogenicity | http://ntp.niehs.nih.gov/results/summaries/chronicstudies/tr0099/bydate/tr052levels/index.html | |
| andrastin A | -- | -- | -- | -- |
| aristolochene | ARISTOLOCHENE CASRN: 26620-71-3 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/26620-71-3 |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Compound | Title | Source. Last update | Information | URL |
|---------------------------|--------------------------------------|--|----------------|---|
| asperparaline(s) | -- | -- | -- | -- |
| citreoviridin | CITREOVIRIDIN CASRN: 25425-12-1 | GENE-TOX (TOXNET). <i>Updated 1991</i> | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+genetox:@term+@DOCNO+3451 |
| | | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/25425-12-1 |
| culmorin | CULMORIN CASRN: 18374-83-9 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/18374-83-9 |
| cyclophenol | CYCLOPENOL CASRN: 20007-85-6 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/20007-85-6 |
| cyclopiazonic acid | CYCLOPIAZONIC ACID CASRN: 18172-33-3 | HSDB (TOXNET). <i>Updated 2005</i> | -- | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+hsdb:@term+@DOCNO+7248 |
| | | CCRIS (TOXNET). <i>Updated 1993</i> | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+4942 |
| | | GENE-TOX (TOXNET). <i>Updated 1995</i> | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+genetox:@term+@DOCNO+3284 |
| | | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/18172-33-3 |
| | | CTD (TOXNET) | genomics | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ctd:@term+@DOCNO+CTD/C000543 |
| | | HAZ-MAP (TOXNET) | occupational | https://hazmap.nlm.nih.gov/category-details?table=copytblagents&id=4609 |
| cytolysin A | -- | -- | -- | -- |
| endophenazine A | -- | -- | -- | -- |
| festuclavine | FESTUCLAVINE CASRN: 569-26-6 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/569-26-6 |
| | | CCRIS (TOXNET). <i>Updated 1995</i> | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+6640 |
| isofumigaclavine | ISOFUMIGACLAVINE CASRN: 72626-13-2 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/72626-13-2 |
| | ISOFUMIGACLAVINE B CASRN: 58800-20-7 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/58800-20-7 |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Compound | Title | Source. <i>Last update</i> | Information | URL |
|--------------------------|------------------------------------|--|--------------------------------|---|
| isotrichodermin | ISOTRICHODERMIN CASRN: 91423-90-4 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/91423-90-4 |
| isotrichodermol | ISOTRICHODERMOL CASRN: 104155-10-4 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/104155-10-4 |
| kojic acid | KOJIC ACID CASRN: 501-30-4 | HSDB (TOXNET). <i>Updated 2009</i> | -- | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+hsdb:@term+@DOCNO+7664 |
| | | CCRIS (TOXNET). <i>Updated 2009</i> | carcinogenicity / genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+4131 |
| | | GENE-TOX (TOXNET). <i>Updated 1995</i> | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+genetox:@term+@DOCNO+1514 |
| | | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/501-30-4 |
| | | CPDB (TOXNET) | carcinogenicity / genotoxicity | https://toxnet.nlm.nih.gov/cpdb/chempage/s/KOJIC%20ACID.html |
| | | CTD (TOXNET) | genomics | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ctd:@term+@DOCNO+CTD/C011890 |
| | Volume 79. Some Thyrotropic Agents | IARC monographs. <i>Updated 2001</i> | carcinogenicity | http://monographs.iarc.fr/ENG/Monographs/vol79/index.php |
| LPS endotoxin | ENDOTOXINS CASRN: 11034-88-1 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/11034-88-1 |
| | | HAZ-MAP (TOXNET) | occupational | https://hazmap.nlm.nih.gov/category-details?table=copytblagents&id=1358 |
| | | ENDOTOXINS NO CASRN | CTD (TOXNET) | genomics |
| malformin(s) | MALFORMINS CASRN: 3022-92-2 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/3022-92-2 |
| maltoryzine | MALTORYZINE CASRN: 6826-42-2 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/6826-42-2 |
| marcfortine | MARCFORTINE A CASRN: 75731-43-0 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/75731-43-0 |
| mitorubrinic acid | -- | -- | -- | -- |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Compound | Title | Source. Last update | Information | URL | |
|--------------------------|---|---|---|---|---|
| mycophenolic acid | MYCOPHENOLIC ACID CASRN: 24280-93-1 | CCRIS (TOXNET). Updated 1994 | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+5565 | |
| | | GENE-TOX (TOXNET). Updated 1995 | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+genetox:@term+@DOCNO+3414 | |
| | | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/24280-93-1 | |
| | | CTD (TOXNET) | genomics | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ctd:@term+@DOCNO+CTD/D009173 | |
| naphtho-pyrones | -- | -- | -- | -- | |
| neoxaline | NEOXALINE CASRN: 71812-10-7 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/71812-10-7 | |
| nigerazine | B: N-Methyl-trans-2,5-dimethyl-N'-cinnamoylpiperazine CASRN | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/85982-75-8 | |
| nigragillin | NIGRAGILLIN(E) CASRN: 24779-38-2 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/24779-38-2 | |
| orlandin | ORLANDIN CASRN: 69975-77-5 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/69975-77-5 | |
| oxalic acid | OXALIC ACID CASRN: 144-62-7 | HSDB (TOXNET). Updated 2009 | -- | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+hsdb:@term+@DOCNO+1100 | |
| | | CCRIS (TOXNET). Updated 2006 | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+1454 | |
| | | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/144-62-7 | |
| | | CTD (TOXNET) | genomics | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ctd:@term+@DOCNO+CTD/D019815 | |
| | | HAZ-MAP (TOXNET) | occupational | https://hazmap.nlm.nih.gov/category-details?table=copytblagents&id=582 | |
| | | NTP Assessments Oxalic acid - 10049-C | NTP | genotoxicity / reproductive toxicity | http://ntp.niehs.nih.gov/testing/status/agents/ts-10049-c.html |
| | | Oxalic Acid (CAS #144-62-7): Reproduction and Fertility Assessment in CD-1 Mice When Ad | NTP | reproductive toxicology | http://ntp.niehs.nih.gov/testing/types/repro/abstracts/racb84064/index-32.html |
| Oxalic acid ICSC: 0529 | ICSC (Inchem). Updated 2009 | safety card | http://www.inchem.org/documents/icsc/icsc/eics0529.htm | | |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Compound | Title | Source. Last update | Information | URL |
|---------------------------|---|---------------------------------|----------------|---|
| perinadine A | -- | -- | -- | -- |
| phenazine-1-carboxylic ac | -- | -- | -- | -- |
| phenazines | PHENAZINE CASRN: 92-82-00 | HAZ-MAP (TOXNET) | occupational | https://hazmap.nlm.nih.gov/category-details?table=copytblagents&id=16513 |
| PR toxin | PR-TOXIN CASRN: 56299-00-4 | HSDB (TOXNET). Updated 2014 | | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+hsdb:@term+@DOCNO+7247 |
| | | CCRIS (TOXNET). Updated 1994 | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+4938 |
| | | GENE-TOX (TOXNET). Updated 1998 | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+genetox:@term+@DOCNO+3920 |
| | | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/56299-00-4 |
| | | HAZ-MAP (TOXNET) | occupational | https://hazmap.nlm.nih.gov/category-details?table=copytblagents&id=6825 |
| pyocianin | -- | -- | -- | -- |
| roquefortine | ROQUEFORTINE CASRN: 58735-64-1 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/58735-64-1 |
| | | HAZ-MAP (TOXNET) | occupational | https://hazmap.nlm.nih.gov/category-details?table=copytblagents&id=6976 |
| sambucinol | SAMBUCINOL CASRN: 90044-33-0 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/90044-33-0 |
| secalonic acid | SECALONIC ACID D CASRN: 35287-69-5 | CCRIS (TOXNET). Updated 1993 | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+ccris:@term+@DOCNO+4937 |
| | | GENE-TOX (TOXNET). Updated 1995 | genotoxicity | http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?dbs+genetox:@term+@DOCNO+3636 |
| | SECALONIC ACID CASRN: 56283-72-8 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/56283-72-8 |
| trichodermin | TRICHODERMIN CASRN: 4682-50-2 | ChemIDplus (TOXNET) | acute toxicity | https://chem.nlm.nih.gov/chemidplus/rn/4682-50-2 |
| | International Programme on Chemical safety. Environmental Health Criteria 105. Selected mycotoxins: ochratoxins, tricothecenes, ergot | IPCS (Inchem) | -- | http://www.inchem.org/documents/ehc/ehc/ehc105.htm |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Compound | Title | Source. <i>Last update</i> | Information | URL |
|--------------|------------------------------|----------------------------|-------------|---|
| xanthomegnin | XANTOMEGNIN CASRN: 1685-91-2 | ChemIDplus (TOXNET) | -- | https://chem.nlm.nih.gov/chemidplus/rn/1685-91-2 |

Appendix D. List of food enzymes & feed additives used as exclusion criteria.

| Product |
|--|
| (1→4)-α-D-glucan 1-α-D-glucosylmutase |
| 1,4-alpha-Glucan branching enzyme |
| 3-Phytase |
| 4-alpha-glucanotransferase |
| 4-Phytase |
| 6-phytase |
| Acetolactate decarboxylase |
| Acetylhexosaminidase (beta-L-N) |
| Acid prolyl endopeptidase |
| Acylglycerol lipase |
| Alginate lyase |
| Alpha-trehalase |
| Alpha-amylase |
| Alpha-galactosidase |
| Alpha-glucosidase |
| Alpha-L-arabinofuranosidase |
| Alpha-L-rhamnosidase |
| Alternansucrase |
| Aminoacylase |
| Aminopeptidase |
| AMP deaminase |
| Aqualysin 1 |
| Arabinanase |
| Arabinofuranosidase |
| Arginine |
| Asparaginase |
| Asparaginase |
| Aspergillopepsin I |
| Aspergillus nuclease S1 |
| Bacillolysin |
| Beta-amylase |
| Beta-galactosidase (lactase) |
| Beta-glucanase |
| Beta-Glucosidase |
| Beta-mannanase |
| Bromelain |
| Carboxypeptidase (serine-type) |
| Carboxypeptidase C |
| Catalase |
| Cellobiose dehydrogenase |
| Cellulase |
| Chitinase |
| Chymosin |
| Chymotrypsin |
| Cyclomaltoextrin glucanotransferase |
| Dextranase |
| D-Fructose 4-Epimerase |
| D-psicose 3-epimerase |
| Endo-1,3(4)-β-glucanase |
| Endo-1,4-beta-xylanase |
| Endothiapepsin |
| Ferulic acid esterase |
| Ficin |
| Food enzyme consisting of Aspergillopepsin I and Aspergillopepsin II |

| Product |
|--|
| Food enzyme consisting of cellulase as a main activity and cellulose 1,4- β -cellobiosidase and β -glucosidase as subsidiary activities) |
| Food enzyme consisting of Cellulase, Endo-1,3(4)- β -glucanase, and Xylanase |
| Food enzyme consisting of Cellulase, Glucanase and Hemicellulase covering Xylanase and Mannanase |
| Food enzyme consisting of Pectinase, Polygalacturonase, Pectinesterase, Pectin Lyase and Arabanase |
| Food enzyme consisting of Protease, Leucyl aminopeptidase, Oryzin and Aspergillopepsin I |
| Food enzyme consisting of Trypsin and Chymotrypsin |
| Food enzyme consisting of Trypsin, Chymotrypsin, Elastase and Carboxypeptidase |
| Food enzyme consisting of Xylanase, Endo-1,3(4)- β -glucanase and Glucan 1,3- β -glucosidase |
| Glucan 1,4- α -glucosidase |
| Glucan 1,4- α -maltotetrahydrolase |
| Glucan 1,4- α -maltohydrolase |
| Glucanase |
| Glucoamylase |
| Glucose isomerase |
| Glucose oxidase |
| Glucosidase (alpha) |
| Glucosidase (beta) |
| Glucosyltransferase or Transglucosidase |
| Glutaminase |
| Hemicellulase |
| Hexose oxidase |
| Inulase |
| Inulinase |
| Invertase |
| Isoamylase |
| Isomaltulose synthase |
| Laccase |
| Lactase or Galactosidase (beta) |
| Lactoperoxidase |
| Isalocid sodium |
| L-ascorbate oxidase |
| Leucyl aminopeptidase |
| Lipase |
| Lipase monoacylglycerol |
| Lipase triacylglycerol |
| Lipoxygenase |
| Lysine |
| Lysophospholipase |
| Lysozyme |
| Maduramicin |
| Maltogenic amylase |
| Mannanase (endo-1.4-beta) |
| Membrane alanyl aminopeptidase |
| Methionine |
| Microbial collagenase |
| Monensin |
| Mucorpepsin |
| Narasin |
| Organophosphate esterase |
| Oryzin |
| Pancreatin |
| Papain |
| Paromomycin |
| Pectate lyase |

| Product |
|---|
| Pectin lyase |
| Pectin methylesterase or Pectinesterase |
| Pectinase |
| Penicillin amidase |
| Pentosanase |
| Peroxidase |
| Phosphodiesterase |
| Phospholipase A |
| Phospholipase B |
| Phospholipase C |
| Phospholipase D |
| Plant coagulant (Phytopsin) |
| Polygalacturonase |
| Protease |
| Protein glutaminase |
| Pullulanase |
| Rennet |
| Rhizopuspepsin |
| Ribonuclease P |
| Salinomycin |
| Semduramicin |
| Subtilisin |
| Tannase |
| Thermolysin |
| Threonine |
| Thrombin |
| Transglutaminase |
| Trypsin |
| Tryptophan |
| Urease |
| Valine |
| Xaa-Pro dipeptidase |
| Xylanase |
| Xylose isomerase |

Appendix E – List of toxins evaluated by EFSA, IARC or NTP or included in Directive 2002/32/EC

Table 1E. List of toxins evaluated by IARC (Monographs on the Evaluation of Carcinogenic Risks to humans), NTP (Report on Carcinogens) or EFSA , or included in the Directive 2002/32/EC

| TOXINS | CAS NUMBER | Agency (year) | Evaluation title/classification |
|------------------------------|---|-------------------------------------|---|
| Aflatoxin B1 ¹ | 1162-65-8 | | |
| Aflatoxins | 1402-68-2 | IARC (2012) | Group 1 ⁴ |
| | | NTP (2016) | Known to be human carcinogens |
| | | EFSA (2006) | Opinion of the scientific panel on contaminants in the food chain [CONTAM] related to the potential increase of consumer health risk by a possible increase of the existing maximum levels for aflatoxins in almonds, hazelnuts and pistachios and derived products |
| | | EFSA (2004) | Opinion of the Scientific Panel on contaminants in the food chain [CONTAM] related to Aflatoxin B1 as undesirable substance in animal feed |
| Beauvericin | 26048-05-5 | EFSA (2014) | Scientific Opinion on the risks to human and animal health related to the presence of beauvericin and enniatins in food and feed |
| Citrinin | 518-75-2 | IARC (1987) | Group 3 ⁵ |
| | | EFSA (2012) | Scientific Opinion on the risks for public and animal health related to the presence of citrinin in food and feed |
| Deoxynivalenol | 51481-10-8 | IARC (1993) ² | Group 3 |
| | | EFSA (2007) | Opinion of the Scientific Panel on Contaminants in the Food Chain on a request from the Commission related to Deoxynivalenol (DON) as undesirable substance in animal feed |
| Enniatin A, A1, B, B1 | 2503-13-1; 4530-21-6; 917-13-5; 19914-20-6 | EFSA (2014) | Scientific Opinion on the risks to human and animal health related to the presence of beauvericin and enniatins in food and feed |
| Ergot alkaloids ¹ | | EFSA (2011) | Scientific Opinion on Ergot alkaloids in food and feed |
| Fumonisin B1 | 116355-83-0 | IARC (1993) ³ , 2002) | Group 2B ⁶ |
| Fumonisin B2 | 116355-84-1 | IARC (1993) ³ | Group 2B |
| Fumonisin | | IARC (1993) ³ | Group 2B |
| | | EFSA (2005) | Opinion of the Scientific Panel on contaminants in the food chain [CONTAM] related to fumonisins as undesirable substances in animal feed |
| Fusarenone X | | IARC (1993) ² | Group 3 |
| Fusarin C | 116355-84-1 | IARC (1993) ³ | Group 2B |
| Nivalenol | 23282-20-4 | IARC (1993) ² | Group 3 |
| | | EFSA (2013) | Scientific Opinion on risks for animal and public health related to the presence of nivalenol in food and feed |
| Ochratoxin A | 303-47-9 | IARC (1993) | Group 2B |
| | | NTP | Reasonably anticipated to be HC |

| TOXINS | CAS NUMBER | Agency (year) | Evaluation title/classification |
|--------------------|---------------------------|--------------------------|---|
| | | EFSA (2006) | Opinion of the Scientific Panel on contaminants in the food chain [CONTAM] related to ochratoxin A in food |
| | | EFSA (2004) | Opinion of the Scientific Panel on contaminants in the food chain [CONTAM] related to ochratoxin A (OTA) as undesirable substance in animal feed. |
| Patulin | 149-29-1 | IARC (1987) | Group 3 |
| Penicillic acid | 90-65-3 | IARC (1987) | Group 3 |
| Phomopsis A | 64925-80-0 | EFSA (2012) | Scientific Opinion on the risks for animal and public health related to the presence of phomopsins in feed and food |
| Sterigmatocystin | 10048-13-2 | IARC (1987) | Group 2B |
| T2 and HT-2 toxins | 21259-20-1; 26934-87-2 | EFSA (2011) | Scientific Opinion on the risks for animal and public health related to the presence of T-2 and HT-2 toxin in food and feed |
| T2-toxin | 21259-20-1 | IARC (1993) | Group 3 |
| Zearalenone | 17924-92-4 | IARC (1993) ² | Group 3 |
| | | EFSA (2011) | Scientific Opinion on the risks for public health related to the presence of zearalenone in food |
| | | EFSA (2004) | Opinion of the Scientific Panel on contaminants in the food chain [CONTAM] related to Zearalenone as undesirable substance in animal feed. |

¹Included in Directive 2002/32/EC

²IARC agent "Fusarium graminearum, F. culmorum, and F. crookwellense, toxins derived from (zearalenone, deoxynivalenol, nivalenol, and fusarenone X)"

³IARC agent "Fusarium moniliforme, toxins derived from (fumonisin B1, fumonisin B2, and fusarin C)"

⁴Carcinogenic to humans

⁵Not classifiable as to its carcinogenicity to humans

⁶Possibly carcinogenic to humans

Appendix F – List of excluded articles. Level 2

Table F1: List of articles excluded according to the criteria established for phase 2 of the revision, and reasons of exclusion

| CITATION | REASON OF EXCLUSION |
|-----------------------------------|--|
| Abbas, 2015. | Document does not report designated outcomes |
| Albillos, <i>et al.</i> , 2011. | Data not well defined/reported for inclusion in the DB |
| Alvi <i>et al.</i> , 2000 | Document does not report designated outcomes |
| Aninat <i>et al.</i> , 2005 | Document does not report designated outcomes |
| Bai <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| Ballok <i>et al.</i> , 2013 | Document does not report designated outcomes |
| Basu <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Bauer <i>et al.</i> , 2001 | Document does not report designated outcomes |
| Bernhoft <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Bianchi <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| Blankenfeldt <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Bohlmann <i>et al.</i> , 1998 | Data not well defined/reported for inclusion in the DB |
| Britigan <i>et al.</i> , 1999 | Data not well defined/reported for inclusion in the DB |
| Britigan <i>et al.</i> , 1997 | Document does not report designated outcomes |
| Burdock <i>et al.</i> , 2001 | Data not well defined/reported for inclusion in the DB |
| Burnett <i>et al.</i> , 2010 | Document does not report designated outcomes |
| Burrowes <i>et al.</i> , 2005 | Data not well defined/reported for inclusion in the DB |
| Cakmakci <i>et al.</i> , 2012 | Document does not report designated outcomes |
| Castellá <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| Cezairliyan <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Chan <i>et al.</i> , 2005 | Document does not report designated outcomes |
| Chang <i>et al.</i> , 2009 | Data not well defined/reported for inclusion in the DB |
| Chang, 2004 | Document does not report designated outcomes |
| Chang <i>et al.</i> , 1996 | Data not well defined/reported for inclusion in the DB |
| Chang <i>et al.</i> , 1998 | Data not well defined/reported for inclusion in the DB |
| Cheluvappa <i>et al.</i> , 2010 | Data not well defined/reported for inclusion in the DB |
| Cheluvappa, 2014 | Data not well defined/reported for inclusion in the DB |
| Chen <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Cherigo <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| Chieda <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| Choi <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Cugini <i>et al.</i> , 2010 | Document does not report designated outcomes |
| Cui <i>et al.</i> , 2009 | Document does not report designated outcomes |
| Cui Q <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| Danguilan <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Dantasa <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |

| CITATION | REASON OF EXCLUSION |
|---------------------------------------|--|
| Darsih <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| Dassanayake <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| De Waal, 2002 | Data not well defined/reported for inclusion in the DB |
| Demetrius <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Deshpande <i>et al.</i> , 2006 | Data not well defined/reported for inclusion in the DB |
| Dhulipala, 2005 | Data not well defined/reported for inclusion in the DB |
| Dhulipala <i>et al.</i> , 2004a | Data not well defined/reported for inclusion in the DB |
| Dhulipala <i>et al.</i> , 2004b | Data not well defined/reported for inclusion in the DB |
| Dhulipala <i>et al.</i> , 2006 | Data not well defined/reported for inclusion in the DB |
| Dirheimer, 1998 | Data not well defined/reported for inclusion in the DB |
| Dobretsov <i>et al.</i> , 2016 | Document does not report designated outcomes |
| Du <i>et al.</i> , 2010 | Document does not report designated outcomes |
| Du <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| Eadon <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| Ehrlich <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| El-Elimat <i>et al.</i> , 2015 | Document does not report designated outcomes |
| Erdogan <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Filler <i>et al.</i> , 2007 | Document does not report designated outcomes |
| Filler <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Folch <i>et al.</i> , 2013 | Document does not report designated outcomes |
| Fontaine <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| Forbes <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Fuse <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Gałczyńska <i>et al.</i> , 2015 | Document does not report designated outcomes |
| Gallo <i>et al.</i> , 2015a | Document does not report designated outcomes |
| Gallo <i>et al.</i> , 2015b | Document does not report designated outcomes |
| Gallo <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Gallo <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| Gao <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Gao <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| García-Estrada <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Geisen, 1996 | Document does not report designated outcomes |
| Geiser <i>et al.</i> , 2000 | Document does not report designated outcomes |
| Gloyne <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Gohain <i>et al.</i> , 2006a | Document does not report designated outcomes |
| Gohain <i>et al.</i> , 2006b | Document does not report designated outcomes |
| Gomes <i>et al.</i> , 2015 | Document does not report designated outcomes |
| Gomez <i>et al.</i> , 1996 | Data not well defined/reported for inclusion in the DB |
| Gruber <i>et al.</i> , 2016 | Document does not report designated outcomes |
| Gruber-Dorninger <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |

| CITATION | REASON OF EXCLUSION |
|---------------------------------------|--|
| Guo <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Guo <i>et al.</i> , 2007 | Data not well defined/reported for inclusion in the DB |
| Gupta <i>et al.</i> , 2000 | Document does not report designated outcomes |
| Guragain <i>et al.</i> , 2016 | Document does not report designated outcomes |
| Guru <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| Hagimori <i>et al.</i> , 2007 | Data not well defined/reported for inclusion in the DB |
| Hanumegowda <i>et al.</i> , 2002a | Document does not report designated outcomes |
| Hanumegowda <i>et al.</i> , 2002b | Document does not report designated outcomes |
| Hendrickson <i>et al.</i> , 2001 | Document does not report designated outcomes |
| Herman-Edelstein <i>et al.</i> , 2012 | Document does not report designated outcomes |
| Hong, 2011 | Document does not report designated outcomes |
| Hoshi <i>et al.</i> , 2005 | Data not well defined/reported for inclusion in the DB |
| Hou <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Hu <i>et al.</i> , 2013 | Document does not report designated outcomes |
| Hu <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Huang J, et al 2012 | Data not well defined/reported for inclusion in the DB |
| Huh <i>et al.</i> , 2013 | Document does not report designated outcomes |
| Hymery <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Ismail <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Jayaseelan <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Jelen, 2002 | Document does not report designated outcomes |
| Jelen <i>et al.</i> , 2002 | Data not well defined/reported for inclusion in the DB |
| Jiang <i>et al.</i> , 1999 | Document does not report designated outcomes |
| Jimenez <i>et al.</i> , 2012 | Document does not report designated outcomes |
| Joshi <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Kamal <i>et al.</i> , 2012 | Document does not report designated outcomes |
| Karolewicz <i>et al.</i> , 2005 | Data not well defined/reported for inclusion in the DB |
| Kasper <i>et al.</i> , 2016 | Document does not report designated outcomes |
| Katri <i>et al.</i> , 2007 | Data not well defined/reported for inclusion in the DB |
| Keblys <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Kharwar <i>et al.</i> , 2011 | Document does not report designated outcomes |
| Kim <i>et al.</i> , 2015 | Document does not report designated outcomes |
| Kim <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Kobayashi <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Koizumi <i>et al.</i> , 2002 | Data not well defined/reported for inclusion in the DB |
| Koizumi <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Koizumi <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Kojima <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| Kong <i>et al.</i> , 2006 | Data not well defined/reported for inclusion in the DB |
| Kuilman-Wahls <i>et al.</i> , 2002 | Data not well defined/reported for inclusion in the DB |

| CITATION | REASON OF EXCLUSION |
|---|--|
| Kutty <i>et al.</i> , 2015 | Document does not report designated outcomes |
| Lahouar <i>et al.</i> , 2015 | Document does not report designated outcomes |
| Leangon <i>et al.</i> , 1999 | Data not well defined/reported for inclusion in the DB |
| Lee <i>et al.</i> , 2006 | Data not well defined/reported for inclusion in the DB |
| Lee <i>et al.</i> , 2011 | Document does not report designated outcomes |
| Lee <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Levif <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Li <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| Liang <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| Liao <i>et al.</i> , 2010. | Data not well defined/reported for inclusion in the DB |
| Lin <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Liu <i>et al.</i> , 2010. | Document does not report designated outcomes |
| Liu <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Lundgren <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Luo <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| Ma <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Mabrouk <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Mai <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Massi <i>et al.</i> , 2016a | Data not well defined/reported for inclusion in the DB |
| Massi <i>et al.</i> , 2016b | Data not well defined/reported for inclusion in the DB |
| McFarland <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| McFarland <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| McKenzie <i>et al.</i> , 1997 | Document does not report designated outcomes |
| Meisel-Mikolajczyk <i>et al.</i> , 1999 | Data not well defined/reported for inclusion in the DB |
| Miller <i>et al.</i> , 2000 | Data not well defined/reported for inclusion in the DB |
| Millot <i>et al.</i> , 2009 | Document does not report designated outcomes |
| Mirandola <i>et al.</i> , 2010 | Data not well defined/reported for inclusion in the DB |
| Mokhtar <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| Montis <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Moura-Alves <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Mudge <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Müller <i>et al.</i> , 1997 | Document does not report designated outcomes |
| Müller, 2011 | Data not well defined/reported for inclusion in the DB |
| Munday <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| Neerman <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Niatsetskaia <i>et al.</i> , 2010 | Data not well defined/reported for inclusion in the DB |
| Nielsen <i>et al.</i> , 2005 | Data not well defined/reported for inclusion in the DB |
| Nielsen <i>et al.</i> , 2009 | Data not well defined/reported for inclusion in the DB |
| Nielsen <i>et al.</i> , 2006 | Document does not report designated outcomes |
| O'Brien <i>et al.</i> , 2006 | Data not well defined/reported for inclusion in the DB |

| CITATION | REASON OF EXCLUSION |
|-------------------------------------|--|
| Okegbe <i>et al.</i> , 2012 | Document does not report designated outcomes |
| Olmeda <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| O'Malley <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Palumbo, 2013 | Data not well defined/reported for inclusion in the DB |
| Parsons <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Parsons <i>et al.</i> , 2007 | Data not well defined/reported for inclusion in the DB |
| Patrauchan <i>et al.</i> , 2007 | Document does not report designated outcomes |
| Pedersen <i>et al.</i> , 1999 | Data not well defined/reported for inclusion in the DB |
| Pel <i>et al.</i> , 2007 | Document does not report designated outcomes |
| Pérez-Martínez <i>et al.</i> , 2011 | Document does not report designated outcomes |
| Pertz, 1996 | Document does not report designated outcomes |
| Podgorski <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Prakash <i>et al.</i> , 2005 | Document does not report designated outcomes |
| Prince <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| Priyaja <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Pulido <i>et al.</i> , 2012 | Document does not report designated outcomes |
| Pustelny <i>et al.</i> , 2013 | Document does not report designated outcomes |
| Qadri <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Rada <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Rahman <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Rahman <i>et al.</i> , 2009 | Document does not report designated outcomes |
| Ran <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Rasmussen <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Reddy <i>et al.</i> , 2010 | Data not well defined/reported for inclusion in the DB |
| Reddy, 2005 | Document does not report designated outcomes |
| Reimann <i>et al.</i> , 1997 | Data not well defined/reported for inclusion in the DB |
| Ren <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Ren <i>et al.</i> , 2006 | Data not well defined/reported for inclusion in the DB |
| Reszka <i>et al.</i> , 2010 | Data not well defined/reported for inclusion in the DB |
| Reszka <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Reszka <i>et al.</i> , 2006 | Document does not report designated outcomes |
| Richard <i>et al.</i> , 2004 | Data not well defined/reported for inclusion in the DB |
| Rudrappa <i>et al.</i> , 2008 | Document does not report designated outcomes |
| Ryan <i>et al.</i> , 2009 | Document does not report designated outcomes |
| Rymowicz <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Sabater <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Santoro <i>et al.</i> , 1999 | Data not well defined/reported for inclusion in the DB |
| Santos <i>et al.</i> , 2002 | Document does not report designated outcomes |
| Sarkisova <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Schmidt-Heydt <i>et al.</i> , 2009 | Data not well defined/reported for inclusion in the DB |

| CITATION | REASON OF EXCLUSION |
|----------------------------------|--|
| Schulz <i>et al.</i> , 1996 | Data not well defined/reported for inclusion in the DB |
| Schweizer <i>et al.</i> , 2011 | Document does not report designated outcomes |
| Seshime <i>et al.</i> , 2009 | Data not well defined/reported for inclusion in the DB |
| Shank <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Shen <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Shipkova <i>et al.</i> , 2005 | Document does not report designated outcomes |
| Simmons <i>et al.</i> , 1997 | Document does not report designated outcomes |
| Singh <i>et al.</i> , 2009 | Data not well defined/reported for inclusion in the DB |
| Sio <i>et al.</i> , 2006 | Document does not report designated outcomes |
| Sismaet <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Sollinger, 1996 | Data not well defined/reported for inclusion in the DB |
| Sommerer <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Sonnleitner <i>et al.</i> , 2003 | Document does not report designated outcomes |
| Staatz <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Stern <i>et al.</i> , 2007 | Data not well defined/reported for inclusion in the DB |
| Su <i>et al.</i> , 2013. | Data not well defined/reported for inclusion in the DB |
| Susca <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Takahashi <i>et al.</i> , 2002 | Document does not report designated outcomes |
| Takaya <i>et al.</i> , 2008 | Document does not report designated outcomes |
| Takemoto <i>et al.</i> , 1998 | Document does not report designated outcomes |
| Tanaka <i>et al.</i> , 2006 | Document does not report designated outcomes |
| Tao <i>et al.</i> , 2014 | Document does not report designated outcomes |
| Tayabali <i>et al.</i> , 2015 | Data not well defined/reported for inclusion in the DB |
| To <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Tokuoka <i>et al.</i> , 2008 | Data not well defined/reported for inclusion in the DB |
| Uchida <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Ueda <i>et al.</i> , 2008 | Document does not report designated outcomes |
| Vaideeswar <i>et al.</i> , 2009 | Document does not report designated outcomes |
| Vinckx <i>et al.</i> , 2010 | Document does not report designated outcomes |
| Wadia <i>et al.</i> , 2009 | Data not well defined/reported for inclusion in the DB |
| Wan <i>et al.</i> 2011 | Data not well defined/reported for inclusion in the DB |
| Wang <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Wang <i>et al.</i> , 2012 | Document does not report designated outcomes |
| Wang <i>et al.</i> , 2013 | Document does not report designated outcomes |
| Xu, 2013 | Data not well defined/reported for inclusion in the DB |
| Xu <i>et al.</i> , 2013 | Document does not report designated outcomes |
| Yamamoto <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Yang <i>et al.</i> , 2016 | Data not well defined/reported for inclusion in the DB |
| Young <i>et al.</i> , 2003 | Data not well defined/reported for inclusion in the DB |
| Yoza <i>et al.</i> , 2006 | Data not well defined/reported for inclusion in the DB |

| CITATION | REASON OF EXCLUSION |
|--------------------------------|--|
| Yu <i>et al.</i> , 2012 | Data not well defined/reported for inclusion in the DB |
| Zacharias <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Zhai <i>et al.</i> , 2011 | Data not well defined/reported for inclusion in the DB |
| Zhai <i>et al.</i> , 2013 | Data not well defined/reported for inclusion in the DB |
| Zhang <i>et al.</i> , 2009 | Data not well defined/reported for inclusion in the DB |
| Zhang <i>et al.</i> , 2014 | Data not well defined/reported for inclusion in the DB |
| Zheng <i>et al.</i> , 2007 | Document does not report designated outcomes |
| Zizzo <i>et al.</i> , 2010 | Data not well defined/reported for inclusion in the DB |

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Appendix G – Summary of secondary metabolites with bioactivity

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--|---|-----------------------|------|
| <i>Actinomadura roseorufa</i> | reports not found | | |
| <i>Actinomadura yumaensis</i> | reports not found | | |
| <i>Aeribacillus pallidus</i> Syn. <i>Geobacillus pallidus</i> | reports not found | | |
| <i>Arthrobacter ramosus</i> | reports not found | | |
| <i>Aspergillus aculeatus</i> | (5aS,6S,7S)-3,7-dihydroxy-6-methoxy-1,4,6,9-tetramethyl-6,7-dihydro-5aHdibenzo[b,e][1,4]dioxepine-8,11-dione | bioactive compound | [1] |
| | 15a hydroxy 22E, 24R ergosta 3, 5, 8 14, 22 tetraen 7 one 1 | bioactive compound | [2] |
| | 2,5-dimethyl-1,3-benzenediol | bioactive compound | [1] |
| | 3,8-dihydroxy-1,4,6,9-tetramethyldibenzo[b,e][1,4]dioxepin-11-one | bioactive compound | [1] |
| | 4-O-demethylbarbatic acid | bioactive compound | [1] |
| | aculeatusquinones A-D | bioactive compound | [1] |
| | aculenes A-D (sesquiterpenoid) | bioactive compound | [3] |
| | asperaculane A (sesquiterpenoid) | bioactive compound | [3] |
| | asperaculane B (sesquiterpenoid) | bioactive compound | [3] |
| | atraric acid | bioactive compound | [1] |
| | nordaucane one (sesquiterpenoid) | bioactive compound | [3] |
| | okaramines H and I | insecticidal activity | [4] |
| secalonic acid D | mycotoxin | [3] | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|------------------------------|-------------------------|------------------------------|-----------|
| <i>Aspergillus japonicus</i> | asperparalines | bioactive compound | [5, 6, 7] |
| | chanoclavine-I | ergot alkaloids intermediate | [8] |
| | cycloclavin | ergot alkaloid | [9] |
| | ergot alkaloids | mycotoxin | [8] |
| | festuclavine | ergot alkaloid | [9] |
| | neoxaline | alkaloid | [10, 11] |
| | ochratoxin A | mycotoxin | [12] |
| | preparaheerquamide | alkaloid | [13] |
| | secalonic acid F | mycotoxin | [14, 15] |
| | sterigmatocystin | mycotoxin | [16] |
| <i>Aspergillus melleus</i> | aspyrone | nematicidal activity | [17] |
| | mellamide | insecticidal activity | [18] |
| | ochratoxin A | mycotoxin | [18] |
| | viomellin | bioactive compound | [18] |
| | xanthomegnin | bioactive compound | [18] |
| <i>Aspergillus niger</i> | anthraquinones | bioactive compound | [19] |
| | asperenone | bioactive compound | [20] |
| | aspernigerin | bioactive compound | [21] |
| | aspernigrins | bioactive compound | [22] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---------------------------------|------------------------------|-------------------------------|---------------------------------|
| <i>Aspergillus niger</i> | aurasperone B ;F; G | naphtho-gamma-pyrone | [23, 24] |
| | campyrone A;B;C | bioactive compound | [25] |
| | coprogen B | iron chelator | [26] |
| | desmethylkotanin | bioactive compound | [23] |
| | ferrichrome | iron chelator | [26] |
| | fonsecin | bioactive compound | [22] |
| | fumonisin; B2; B4 | mycotoxin | [23, 27-37] |
| | funalenone | collagenase inhibitor | [31] |
| | hexylitaconic acid | plant growth regulator | [38] |
| | kotanin | bioactive compound | [23, 31, 34] |
| | malformin A; C | cause malformations in plants | [22, 23, 31, 34] |
| | maltoryzine | mycotoxin | [39] |
| | naphtho-gamma-pyrones | toxic aromatic compounds | [31, 34, 40] |
| | nigerasperone A;B;C | naphtho-gamma-pyrones | [41] |
| | nigerazine B | plant growth inhibitor | [42] |
| | nigragillin | alkaloid | [40, 42] |
| | ochratoxin A;alpha | mycotoxin | [23, 24, 29, 31, 33, 34, 43-53] |
| orlandin | bioactive compound | [23, 40] | |
| oxalates and oxalic acid | toxic organic compound | [54-57] | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---------------------------|-----------------------------------|---------------------------------------|------------------|
| <i>Aspergillus niger</i> | pyranonigrin A | antioxidative activity | [23, 31] |
| | rubrofusarins | bioactive compound | [22] |
| | saponins | foaming glucoside | [19] |
| | tensidol B | antimicrobial compound | [23, 31] |
| | tensyuc acids | antibiotic | [38] |
| | yanuthone K;L;M;X2 | antifungal compound | [58] |
| <i>Aspergillus oryzae</i> | 13-desoxypaxilline | intermediate metabolite | [59] |
| | 2-oxocyclopiazonic acid | less toxic analogue of CPA | [60] |
| | 3-nitropropionic acid | mycotoxin | [42] |
| | aflatoxins B1 and B2 | mycotoxin | [61, 62] |
| | aspergillomarasmine | phytotoxins | [42] |
| | aspirochlorine | inhibitor of fungal protein synthesis | [63] |
| | astellolides A; B; C-E; F-I | sesquiterpenoids | [64] |
| | bisdethio(bismethylthio)gliotoxin | bioactive compound | [65] |
| | cyclo-(Pro,Val) | bioactive compound | [65] |
| | cyclo-(Tryp,Tyr) | bioactive compound | [65] |
| | cyclopiazonic acid | mycotoxin | [42, 60, 65- 69] |
| | deoxynivalenol (DON) | mycotoxin | [62] |
| | ditryptophenaline | alkaloid | [65] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--|-----------------------|-------------------------------|------------------|
| <i>Aspergillus oryzae</i> | kojic acid | tyrosinase inhibitor | [42, 65, 66, 70] |
| | maltoryzine | cause food poisoning in cows | [42] |
| | oryzaeins | bioactive compound | [71] |
| | parasiticolide A | secondary metabolite | [59] |
| | pseurotin A | bioactive compound | [65] |
| | speradines B -H | bioactive compounds | [72, 73] |
| | T-2 toxin | mycotoxin | [62] |
| | uridine | bioactive compound | [65] |
| | violacetin | bioactive compound | [42] |
| | zearalenone | mycotoxin | [62] |
| <i>Aspergillus sojae</i> | aflatoxin | mycotoxin | [74] |
| | glyceollins | isoflavone bioactive compound | [75, 76] |
| <i>Bacillus macerans</i> Syn. <i>Paenibacillus macerans</i> | reports not found | | |
| <i>Bacillus naganoensis</i> Syn. <i>Pullulanibacillus naganoensis</i> | reports not found | | |
| <i>Bacillus soli</i> Syn. <i>Paenibacillus macerans</i> | reports not found | | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--|--|---------------------------------|----------|
| <i>Bullera singularis</i> Syn. <i>Sporobolomyces singularis</i> | reports not found | | |
| <i>Candida cylindracea</i> Syn. <i>Candida rugosa</i> | reports not found | | |
| <i>Candida lipolytica</i> Syn. <i>Mycotorula lipolytica</i> | tryptanthrin | agonists of the dioxin receptor | [77, 78] |
| <i>Candida paralipolytica</i> Syn. <i>Candida lipolytica</i> | reports not found | | |
| <i>Candida rugosa</i> | reports not found | | |
| <i>Cellulosimicrobium cellulans</i> | anthranilic acid | bioactive compound | [79] |
| | cyclo-(dehydroala-I-Leu) | bioactive compound | [79] |
| | cyclo-(I-Pro-I-Leu) | bioactive compound | [79] |
| | cyclo-(I-Pro-I-Tyr) | bioactive compound | [79] |
| <i>Cellulosimicrobium cellulans</i> | cyclo-(I-Pro-I-Val) | bioactive compound | [79] |
| | L-phenylalanine | bioactive compound | [79] |
| <i>Chaetomium erraticum</i> | reports not found | | |
| <i>Chaetomium gracile</i> | 22E,24R)-ergosta-7,22-diene-3 β ,5alfa,6beta-triol | antimicrobial substance | [80] |
| | adenosine | antimicrobial substance | [80] |
| | chaetochromin A | antimicrobial substance | [80] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---------------------------------------|---------------------------------------|------------------------------------|----------|
| | chaetoquadrin | antimicrobial substance | [80] |
| | ergosterol | antimicrobial substance | [80] |
| | eugenitol | antimicrobial substance | [80] |
| | glycerol monopalmitate | antimicrobial substance | [80] |
| | indole-3-carboxylic acid | antimicrobial substance | [80] |
| | p-hydroxylbenzaldehyde | antimicrobial substance | [80] |
| <i>Cryphonectria parasitica</i> | A (1 --> 3)-beta-D-glucan | immunostimulant | [81] |
| Syn. <i>Endothia parasitica</i> | cryparin | hydrophobin protein | [82, 83] |
| | cryphonectric acid | phytotoxin | [84] |
| | diaporthin | produce canker in leaves | [84] |
| <i>Cryphonectria parasitica</i> | L- p-hydroxyphenyllactic acid (HOPLA) | phytotoxin | [84] |
| | orthosporin | produce canker in leaves | [84] |
| | phleichrome | pigment (quinone) | [85] |
| <i>Chryseobacterium proteolyticum</i> | endotoxin | (virulence factor), low production | [86] |
| <i>Disporotrichum dimorphosporum</i> | reports not found | | |
| <i>Endothia parasitica</i> | reports not found | | |
| Syn. <i>Cryphonectria parasitica</i> | | | |
| Escherichia coli K-12 | curli proteins | adhesion protein | [87] |
| | cytolysin A (ClyA) | cytotoxin | [88, 89] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|----------------------------------|--|----------------------------------|--------------|
| | lipopolysaccharide (LPS) or endotoxin | endotoxin | [90] |
| | SheA hemolysin | cytotoxin | [91] |
| | Tsh protein | hemagglutinin (virulence factor) | [92] |
| <i>Flavobacterium multivorum</i> | reports not found | | |
| <i>Fusarium venenatum</i> | apotriconothecene | mycotoxin (trichothecenes) | [93] |
| | beauvericin | mycotoxin | [94] |
| | butenedioic acid | bioactive compound | [95] |
| | culmorin | sesquiterpene | [93] |
| | culmorone | sesquiterpene | [93] |
| | diacetoxyscirpenol | mycotoxin (trichothecenes) | [93, 95, 96] |
| | enniatin B | sesquiterpene | [93] |
| | fusarin C | mycotoxin | [97] |
| | hexadecane | bioactive compound | [95] |
| | hexadecanoic acid | bioactive compound | [95] |
| <i>Fusarium venenatum</i> | imidazole | bioactive compound | [95] |
| | isotrichodermin | mycotoxin (trichothecenes) | [93] |
| | isotrichodermol | mycotoxin (trichothecenes) | [93] |
| | phenol, 2,5-bis (1,1- dimethylethyl) | bioactive compound | [95] |
| | phthalic acid | bioactive compound | [95] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---------------------------------------|---------------------------------|----------------------------|-------|
| | sambucinol | mycotoxin (trichothecenes) | [93] |
| | tetradecanoic acid | bioactive compound | [95] |
| <i>Geobacillus caldoproteolyticus</i> | reports not found | | |
| <i>Geobacillus palidus</i> | antimicrobial polipeptide | antimicrobial | [98] |
| <i>Geobacillus stearothermophilus</i> | reports not found | | |
| <i>Hansenula polymorpha</i> | reports not found | | |
| <i>Humicola insolens</i> | reports not found | | |
| <i>Klebsiella planticola</i> | indole-3-acetic acid (IAA) | auxin (plant hormone) | [99] |
| | 3-hydroxypropionaldehyde (HPA) | antimicrobial | [100] |
| <i>Klebsiella pneumoniae</i> | capsule polysaccharide | virulence factor | [101] |
| | lipopolysaccharide (LPS) | virulence factor | [101] |
| <i>Leptographium procerum</i> | reports not found | | |
| <i>Microbacterium imperiale</i> | reports not found | | |
| <i>Micrococcus luteus</i> | reports not found | | |
| <i>Mucor circinelloides</i> | 3-nitropropionic acid | mycotoxin | [102] |
| Syn. <i>Mucor javanicus</i> | | | |
| <i>Mucor javanicus</i> | reports not found | | |
| Syn. <i>Mucor circinelloides</i> | | | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--|--|------------------------|------------|
| <i>Mycotorula lipolytica</i> Syn. <i>Candida lipolytica</i> | reports not found | | |
| <i>Ogataea polymorpha</i> Syn. <i>Hansenula polymorpha</i> | reports not found | | |
| <i>Paecilomyces lilacinus</i> Syn. <i>Penicillium lilacinum</i> | leucinostatins | bioactive compounds | [103, 104] |
| <i>Paenibacillus alginolyticus</i> | reports not found | | |
| <i>Paenibacillus lentus</i> | reports not found | | |
| <i>Paenibacillus macerans</i> | anti-phytopathogenic compounds | | [105] |
| | nematicidal compounds | | [106] |
| <i>Penicillium camemberti</i> | cyclopiazonic acid | mycotoxin | [107] |
| <i>Penicillium chrysogenum</i> | (3R, 4R)-3,4,8-trihydroxy-3,4-dihydronaphthalen-1 (2H)-one | bioactive compound | [108] |
| | (9Z,12Z)-2,3-dihydroxypropyl octadeca-9,12-dienoate | neuroprotective effect | [109] |
| | (E)-N-(4-hydroxystyryl) formamide | bioactive compound | [108] |
| | (Z)-N-(4-hydroxy styryl) formamide | bioactive compound | [108] |
| | 1,2,4 dihydroxy 6 methylbenzoyl glycerol | antifungal compound | [110] |
| | 11 bromoroquefortine C | bioactive compound | [111] |
| | 2,2,4 dihydroxy 6 methylbenzoyl glycerol | antifungal compound | [110] |
| | 2-(4-hydroxyphenyl)acetamides | bioactive compound | [112] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|------------------------------------|--|------------------------|------------|
| <i>Penicillium chrysogenum</i> | 2-(4-hydroxyphenyl) acetonitrile | bioactive compound | [108] |
| | 2,3-dihydrosorbicillin | neuroprotective effect | [109] |
| | 2-[(2-hydroxypropionyl) amino] benzamide | neuroprotective effect | [109] |
| | 24 epicyclotrinol 9 | steroid | [113] |
| | 2S,3R oxaline | bioactive compound | [108] |
| | 4-(2-hydroxyethyl) benzene-1,2-diol | bioactive compound | [108] |
| | bisvertinolone | bioactive compound | [114] |
| | BMS 182123 | TNF-alpha inhibitor | [115] |
| | chrysogenamide A | neuroprotective effect | [109] |
| | chrysotriazoles A and B | bioactive compound | [112] |
| | circumdatin G | neuroprotective effect | [109] |
| | citrinin | mycotoxin | [116-118] |
| | conidiogenones | diterpenes | [119] |
| | conidogenol | diterpenes | [119] |
| | cyclocitrinol | steroid | [113] |
| | emodin | bioactive compound | [108, 120] |
| | ergosterol endoperoxide | bioactive compound | [121] |
| erythro 11 hydroxyneocyclocitrinol | steroid | [113] | |
| linolenic acid | bioactive compound | [121] | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--------------------------------|------------------------------------|-----------------------------------|------------|
| <i>Penicillium chrysogenum</i> | lovastatin | bioactive compound | [122] |
| | meleagrins | bioactive compound | [120, 123] |
| | methyl 2-(4-hydroxyphenyl) acetate | bioactive compound | [108] |
| | N 4 hydroxystyryl formamides | bioactive compound | [112] |
| | neocyclocitrinols A-D | steroid | [113] |
| | norcyclocitrinol A | steroid | [113] |
| | ochratoxin A | mycotoxin | [124, 125] |
| | oxosorbicillinol | bioactive compound | [114] |
| | penicillin G | antibiotic | [123] |
| | penicimonoterpene | antifungal compound | [110] |
| | penicisteroid A, B | cytotoxic polyoxygenated steroids | [126] |
| | penicitide A, B | cytotoxic and antimicrobial | [110] |
| | penicitols A and C | citrinin analogues | [127] |
| | penimethavone A | flavone | [128] |
| | penixanacid A | xanthone derivative | [127] |
| | pesudocyclocitrinol A | cytotoxic polyoxygenated steroids | [113] |
| | phenoxyacetic acid | antifungal compound | [129] |
| | phenylacetic acid | citrinin analogues | [129] |
| | PR toxin | sesquiterpenoid mycotoxin | [130] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--------------------------------|--|--------------------|--------------------------|
| <i>Penicillium chrysogenum</i> | quinazolinones | bioactive compound | [112] |
| | roquefortine C | mycotoxin | [114, 120, 123, 131-133] |
| | rugulosin | bioactive compound | [134] |
| | secalonic acid | mycotoxin | [120, 135] |
| | skyrin | bioactive compound | [134] |
| | sorbicillactones A and B | bioactive compound | [114] |
| | sorbicillin | bioactive compound | [114] |
| | sorbicillinol | bioactive compound | [114] |
| | sorbivinetone | bioactive compound | [114] |
| | xanthocillin X | bioactive compound | [123] |
| <i>Penicillium citrinum</i> | (3S)-4,6-dihydro-8-methoxy-3,5-dimethyl-6-oxo-3H-2-benzopyran | bioactive compound | [136] |
| | (3S)-6-hydroxy-8-methoxy-3,5-dimethylisochroman | bioactive compound | [136] |
| | 1,2,3,11b-tetrahydroquinolactacide | bioactive compound | [136] |
| | 2-(hept-5-enyl)-3-methyl-4-oxo-6,7,8,8a-tetrahydro-4H-pyrrolo[2,1-b]-1,3-oxazine | bioactive compound | [137] |
| | 2,4,5-trimethylbenzene-1,3-diol | bioactive compound | [137] |
| | 2,4-dihydroxy-3, 5,6-trimethylbenzoic acid | bioactive compound | [137] |
| | 3-methoxy-2-methyl-4H-pyran-4-one | bioactive compound | [137] |
| | 4-hydroxyquinolin-2(1H)-one | bioactive compound | [136] |
| 5-methyl alternariol ether | bioactive compound | [137] | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--|---|---------------------|------------|
| <i>Penicillium citrinum</i> | 6-methylcurvulinic acid | bioactive compound | [136] |
| | 8-methoxy-3,5-dimethylisoquinolin-6-ol | bioactive compound | [136] |
| | ω -hydroxyemodin | bioactive compound | [137] |
| | alternariol | bioactive compound | [137] |
| | arohynapene D | bioactive compound | [136] |
| | citrinamide | bioactive compound | [136] |
| | citrinin, H1, H2 | mycotoxin | [137-147] |
| | citriquinochroman | citrinin derivative | [136] |
| | conioxanthone A | bioactive compound | [137] |
| | decarboxycitrinin | citrinin derivative | [137, 142] |
| | decarboxydihydrocitrinone | citrinin derivative | [145] |
| | dicitrinin | citrinin derivative | [142] |
| | dicitrinol A and B | citrinin derivative | [142] |
| | ergot alkaloids | mycotoxin | [148] |
| | indole acetic acid methyl ester | bioactive compound | [136] |
| | methyl 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylate | bioactive compound | [137] |
| | methylpenicillin | bioactive compound | [136] |
| mevastatine | statins | [149] | |
| Pen c 1, 2, 3, 13, n18, 22, 24, 30, 32 | allergen | [150-157] | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---------------------------------|---------------------------------------|----------------------------|-----------------|
| <i>Penicillium citrinum</i> | penicnoline | bioactive compound | [136] |
| | penicitrinol B, C, D, E13, G, L | cytotoxic compound | [142, 145, 158] |
| | penicitrinone A | cytotoxic compound | [158] |
| | penidicitrinin B | cytotoxic compound | [158] |
| | perinadine A | alkaloid | [137, 159] |
| | phenol A, phenol A acid | bioactive compound | [142] |
| | quinolactacide | TNF inhibitor | [136, 160] |
| | quinolactacin A | TNF inhibitor | [137] |
| | redoxcitrinin | citrinin derivative | [137] |
| | sclerotinin A, B | bioactive compound | [137] |
| | sydowinin B | bioactive compound | [137] |
| | tanzawaic acid A, B, F, G, H | anti-inflammatory activity | [136, 147] |
| | trichodermamide | bioactive compound | [136] |
| | tricitrinol A, B | bioactive compound | [142] |
| vanillic acid | bioactive compound | [136] | |
| <i>Penicillium decumbens</i> | (16R)-16-hydroxy-24-epi-cyclocitrinol | bioactive compound | [161] |
| Syn. <i>Penicillium indicum</i> | 12R-hydroxycyclocitrinol | bioactive compound | [161] |
| | 20-O-methyl-24-cyclocitrinol | bioactive compound | [161] |
| | 20-O-methyl-24-epi-cyclocitrinol | bioactive compound | [161] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---|--|--------------------------------------|-------------------|
| <i>Penicillium decumbens</i> | 22-acetyliscyclocitrinol A | bioactive compound | [161] |
| | 24-epi-cyclocitrinol | bioactive compound | [161] |
| | cyclocitrinol | bioactive compound | [161] |
| | cyclopenicillone | bioactive compound | [161, 162] |
| | decumbenones A and B | fungal inhibitor | [163] |
| | isocyclocitrinol A, B | bioactive steroid | [161] |
| | neocyclocitrinol A, B, C, D | bioactive steroid | [161] |
| | OH-15 derivative of 24-epi-cyclocitrinol | bioactive compound | [161] |
| <i>Penicillium funiculosum</i> Syn. <i>Talaromyces funiculosus</i> | versiol | fungal inhibitor | [163] |
| | 2,3,4 trihydroxybutanamide | antimicrobial compound | [164] |
| | chrodriamans I and J | bioactive compound | [165] |
| | citreoviridin | mycotoxin | [166] |
| | gibberellins (diterpenoid acids) | plant growth promoter | [167] |
| | mitorubrinic acid | derivative of roquefortine | [168, 169] |
| | OR 1 | a mixture of esters of glyceric acid | [169, 170] |
| | patulin | mycotoxin | [171] |
| | prenyleudesmane diterpenoids 1 and 2 | terpenoid | [172] |
| | siderophores | iron chelator | [173] |
| | <i>Penicillium funiculosum</i> | wortmannin | cytotoxic steroid |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---------------------------------------|--|----------------------|-----------------|
| <i>Penicillium lilacinum</i> | leucinostatins, A | statin | [103, 104, 175] |
| Syn. <i>Purpureocillium lilacinum</i> | phytotoxins | agonist to nematodes | [176] |
| Syn. <i>Paecylomices lilalycinus</i> | | | |
| <i>Penicillium multicolor</i> | (S)-6-((1S,2S)-1,2-dihydroxypentyl)-4-methoxy-5,6-dihydro-2H-pyran-2-one | antitumor | [177] |
| | 5 bromochrephilone | bioactive compound | [178] |
| | 5-hydroxy-4-methoxy-5,6-dihydro-2H-pyran-2-one | bioactive compound | [177] |
| | 8-O-methylsclerotiorinamine | bioactive compound | [179] |
| | dechloroisochromophilone II- IV (azaphilone) | bioactive compound | [177, 178] |
| | epi-isochromophilone III (azaphilone) | bioactive compound | [177] |
| | ergosterol | bioactive compound | [177] |
| | ergosterol peroxide | bioactive compound | [177] |
| | isochromophilone I-IV, VI, VIII (azaphilone) | bioactive compound | [177, 178, 180] |
| | Isorotiorin (azaphilone) | bioactive compound | [178] |
| | Ochrephilone (azaphilone) | bioactive compound | [177] |
| | sclerotioramine (azaphilone) | bioactive compound | [177] |
| <i>Penicillium multicolor</i> | sclerotiorin (azaphilone) | bioactive compound | [177, 180] |
| <i>Penicillium nordicum</i> | ochratoxin A | mycotoxin | [181-191] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|-------------------------------------|---|---------------------------------------|---|
| <i>Penicillium notatum</i> | reports not found | | |
| <i>Syn. Penicillium chrysogenum</i> | | | |
| <i>Penicillium roqueforti</i> | 1-hydroxyeremophil-7(11),9(10)-dien-8-one | | [192] |
| | andrastins A-D | bioactive compounds | [192-195] |
| | aristolochene | in the biosynthetic route of PR toxin | [130, 196] |
| | eremofortines A-E | alkaloid | [197] |
| | festuclavine | ergot alkaloid | [197-199] |
| | isofumigaclavines A and B | ergot alkaloids | [197, 198] |
| | marcfortines | derivatives of roquefortine | [195, 200] |
| | mycophenolic acid | mycotoxin | [192, 200-208] |
| | patulin | mycotoxin | [204, 205, 208] |
| | penicillic acid | mycotoxin | [204, 205, 208] |
| | PR toxin | mycotoxin | [130, 192, 193, 196, 198, 200, 205, 209-212] |
| | roquefortine A, C, D | mycotoxin | [192, 193, 195, 197, 198, 200, 204, 206-209, 212-219] |
| <i>Penicillium salamii</i> | reports not found | | |
| <i>Penicillium sclerotiorum</i> | cyclophenol | benzodiazepine alkaloid | [220] |
| <i>Syn. Penicillium multicolor</i> | ergosterol | bioactive alkaloid | [220] |
| | ergosterol peroxide | bioactive alkaloid | [220] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|----------------------------------|-------------------------------------|----------------------------|------------|
| | isochromophilone VI | antimicrobial substance | [221] |
| | pencolide | antimicrobial substance | [221] |
| | sclerotiorin | aldose reductase inhibitor | [221, 222] |
| | verrucosidin | bioactive alkaloid | [220] |
| | viridicatin | bioactive alkaloid | [220] |
| | viridicatol | bioactive alkaloid | [220] |
| <i>Pichia angusta</i> | reports not found | | |
| <i>Syn. Hansenula polymorpha</i> | | | |
| <i>Pseudomonas aeruginosa</i> | 2-aminoacetophenone | virulence factor | [223] |
| | alkylquinoline | signaling molecule | [224] |
| | CFTR inhibitory factor | virulence factor | [225, 226] |
| | elastase | | [227] |
| | ETA | | [228] |
| | ExoS | virulence factor | [228-231] |
| | ExoT | virulence factor | [228, 229] |
| | Exotoxin A | virulence factor | [227, 232] |
| | ExoU | virulence factor | [233-237] |
| | L-2-amino-4-methoxy-3-butenoic acid | | [238] |
| | lipopolysaccharide (LPS) | endotoxin | [239, 240] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---|----------------------------------|---------------------------------|----------------|
| <i>Pseudomonas aeruginosa</i> | lipotoxin | | [241] |
| | metallo- β -Lactamase | antimicrobial resistance factor | [242] |
| | PcrV (type III secretion system) | virulence factor | [243] |
| | phenazine and derivates | virulence factor | [244-246] |
| | pseudoverdine | virulence factor | [247] |
| | pyochelin (siderophore) | virulence factor | [248] |
| | pyocyanin (phenazide) | virulence factor | [223, 249-262] |
| | pyoluteorin | | [263] |
| | pyoverdine | virulence factor | [227, 264-266] |
| | rhamnolipids | bacterial surfactants | [227, 267] |
| | Tse2 | virulence factor | [268] |
| | TypA | virulence factor | [269] |
| Type II secretion system | virulence factor | [270] | |
| Type III secretion system | virulence factor | [271-274] | |
| <i>Pseudomonas amyloclavata</i> | reports not found | | |
| <i>Pullulanibacillus naganoensis</i> | reports not found | | |
| <i>Purpureocillium lilacinum</i> <i>Syn. Penicillium lilacinum</i> <i>Syn. Paecylomices</i> | reports not found | | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|---|--|--------------------------------|-------|
| <i>Raoultella planticola</i> <i>Syn. Klebsiella planticola</i> | reports not found | | |
| <i>Rhizomucor miehei</i> | reports not found | | |
| <i>Rhizopus niveus</i> | reports not found | | |
| <i>Rhizopus oryzae</i> | gamma-linolenic acid methyl ester (methyl GLA) | antiapoptotic effect | [275] |
| | linoleic acid | cytotoxic and antiinflammatory | [276] |
| <i>Rhizopus oryzae</i> | linolenic acid methyl ester | cytotoxic and antiinflammatory | [276] |
| | palmitic acid | cytotoxic and antiinflammatory | [276] |
| | palmitic acid methyl ester | cytotoxic and antiinflammatory | [276] |
| | Rhi o 1 | allergen | [277] |
| <i>Sphingobacterium multivorum</i> | reports not found | | |
| <i>Sporobolomyces singularis</i> | reports not found | | |
| <i>Streptomyces albus</i> | bacteriocin | antibacterial compound | [278] |
| | pyranones | antitumor | [279] |
| | rebeccamycin (indolocarbazole alkaloids) | antitumor | [280] |
| | siderophore | iron chelant | [278] |
| | terpene | bioactive compound | [278] |
| <i>Streptomyces aureofaciens</i> | reports not found | | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--------------------------------------|--|------------------------------|------------|
| <i>Streptomyces chrestomyceticus</i> | albofungin | antibiotic | [281] |
| | chloroalbofungin | antibiotic | [281] |
| | chrestoxanthones A-C | antifungal | [281] |
| <i>Streptomyces chromofuscus</i> | carazostatin | carbazole alkaloids | [282] |
| <i>Streptomyces chromofuscus</i> | herboxidiene | bioactive compound | [283] |
| <i>Streptomyces cinnamonensis</i> | 2-demethylmonensins A, B | minor congeners of monensins | [284] |
| | arcyriaflavin E | indolocarbazole alkaloid | [285] |
| | endophenazine A | antimicrobial compound | [286, 287] |
| | furanonaphthoquinone I (FNQ I) | bioactive compound | [286, 287] |
| <i>Streptomyces cinnamoneus</i> | reports not found | | |
| <i>Streptomyces lasaliensis</i> | manumycin A and lycorine | antimicrobial substance | [288] |
| <i>Streptomyces lividans</i> | colabomycin E | anti inflammatory agent | [289] |
| <i>Streptomyces mobaraensis</i> | bleomycin | anticancer substance | [290] |
| <i>Streptomyces murinus</i> | reports not found | | |
| <i>Streptomyces netropsis</i> | congoicidone analogs (pyrrolamide compounds) | antibiotics | [291] |
| | congoicidone (pyrrolamide compound) | antibiotic | [292] |
| | disgocidone (pyrrolamide compound) | antibiotic | [292] |
| | distamycin (pyrrolamide compound) | antibiotic | [291, 292] |
| <i>Streptomyces olivochromogenes</i> | reports not found | | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--|------------------------------|-----------------------------|------------|
| <i>Streptomyces rubiginosus</i> | reports not found | | |
| <i>Streptomyces violaceoruber</i> | granaticin A, C | bioactive polyketide | [293, 294] |
| <i>Streptomyces violaceoruber</i> | granatomycin E | bioactive polyketide | [293] |
| | kendomycin | bioactive polyketide | [295] |
| | metenaticin A, B and C | bioactive polyketide | [293] |
| <i>Streptoverticillium cinnamoneum</i> | antibiotic HA 94 | antimicrobial substance | [296] |
| <i>Streptoverticillium mobaraense</i> | reports not found | | |
| <i>Talaromyces cellulolyticus</i> | reports not found | | |
| <i>Talaromyces emersonii</i> | reports not found | | |
| Syn. <i>Rasamsonia emersonii</i> | | | |
| <i>Talaromyces versatilis</i> | reports not found | | |
| <i>Torula lypolitica</i> | reports not found | | |
| <i>Trametes hirsuta</i> | 1-phenyl-3-methylpyrazolones | oxidoreductase mediators | [297] |
| | aryl tetral226in lignans | bioactive compounds | [298] |
| | podophyllotoxin | bioactive compounds | [298] |
| <i>Trametes versicolor</i> | funatrol D | bioactive compound | [299] |
| | isodrimenediol | sesquiterpenes (anticancer) | [299] |
| | tramspiroins | sesquiterpenes (anticancer) | [299] |
| | triterpenes | bioactive compounds | [300] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|----------------------------------|---|-----------------------------|------------|
| | rosenonolactone 15,16-acetonide | sesquiterpenes (anticancer) | [299] |
| <i>Trichoderma citrinoviride</i> | 24-methylenecycloartanol | | [301] |
| | 5-hydroxy-2,3-dimethyl-7-methoxychromone | | [301] |
| | bislongiquinolide | insecticide activity | [302] |
| | citrantifidiene | insecticide activity | [303] |
| | citrantifidiol | insecticide activity | [303] |
| | citrinoviric acid | cytotoxic metabolite | [304] |
| | citrostadienol | antimicrobial activity | [301] |
| | cyclo Leu Pro, cyclo Ile Pro, cyclo Phe Pro | | [304] |
| | dihydrotrichodimerol | insecticide activity | [302] |
| | lignoren | | [304] |
| <i>Trichoderma citrinoviride</i> | nafuredin | antimicrobial activity | [301] |
| | penicillenol B1; B2, D | cytotoxic metabolite | [304] |
| | peptaibols | bioactive compounds | [305, 306] |
| | trichocitrin | antimicrobial activity | [301] |
| | trichoderiol A, C | | [304] |
| | trichodimerol | insecticide activity | [302] |
| <i>Trichoderma harzianum</i> | 1,8-dihydroxy-3-methyl-anthraquinone | antifungal | [307-309] |
| | 1-hydroxy-3-methylanthraquinone | antifungal | [307-309] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|------------------------------|--|--------------------|------------|
| | 2(5H)-furanone | antifungal | [308] |
| | 2,4-di-tert-butylphenol | bioactive compound | [310] |
| | 2,6,10-trimethylundeca-5,9-dienal | bioactive compound | [310] |
| | 2-phenylethanol | bioactive compound | [311] |
| | 6-pentyl-alpha-pyrone | bioactive compound | [312, 313] |
| | 6-methyl-1,3,8-trihydroxyanthraquinone | antifungal | [308] |
| | 6-n-pentyl-a-pyrone | bioactive compound | [311] |
| | 6-pentyl-2H-pyran-2-one | antifungal | [308] |
| | alpha-cubebene | bioactive compound | [310] |
| | alpha-curcumene | bioactive compound | [310] |
| | anthraquinones | antifungal | [314] |
| | azaphilone | antifungal | [307, 309] |
| | b-bisabolene | bioactive compound | [310] |
| | biformen (6CI) | bioactive compound | [310] |
| | butenolide | antifungal | [307, 309] |
| | chrysophanol | antifungal | [314, 315] |
| | cytochalasin analogs | cytotoxic compound | [316] |
| <i>Trichoderma harzianum</i> | decanolactone | antifungal | [308] |
| | emodin | antifungal | [314] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|------------------------------|-------------------------------|---------------------|-----------------|
| <i>Trichoderma harzianum</i> | ergosterol | antifungal | [308, 311, 317] |
| | fleophilone | bioactive compound | [318] |
| | gliotoxin | bioactive compound | [319] |
| | gliovirin | bioactive compound | [319, 320] |
| | harzianic acid | siderophore | [321] |
| | harzianolide | anti-phytopathogen | [307, 309, 322] |
| | harzianopyridone | anti-phytopathogen | [307-309] |
| | harziphilone | bioactive compound | [318] |
| | hexahydrofarnesol | bioactive compound | [310] |
| | indol acetic acid | anti-phytopathogen | [323-325] |
| | lignocerane | bioactive compound | [310] |
| | nerolidol | bioactive compound | [310] |
| | pachybasin, hydroxypachybasin | anti-phytopathogen | [314, 315] |
| | pentadecane | bioactive compound | [310] |
| | peptaibols | bioactive compounds | [326, 327] |
| | pristane | bioactive compound | [310] |
| | sitosterol | antifungal | [308] |
| sorbicillin | bioactive compound | [311] | |
| stigmasterol | antifungal | [308] | |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|-------------------------------------|--|------------------------------|-----------------|
| | tandyukisins B-D (decalin derivative) | cytotoxic | [328] |
| | trichodenone A, B, C | cytotoxic | [329] |
| | trichodermin | trichotecene toxin | [319, 330] |
| | trichodermol (detected in hydrolysed crude extracts) | trichotecene toxin | [331] |
| | trichorzin | peptaibol | [332] |
| | tyrosol | bioactive compound | [311] |
| | verticillol | bioactive compound | [310] |
| | viridiofungin A | anti-phytopathogen | [333] |
| <i>Trichoderma koningii</i> | 6, 4-oxopentyl-2H-pyran-2-one | antifungal | [334] |
| | 6-pentyl-pyranone | antifungal | [334] |
| | 7-O-methylkoninginin D (polyketide derivative) | antimicrobial | [335] |
| | alpha-aminoisobutyric acid | | [336] |
| | amino alcohol (peptaibols) | | [336] |
| | decanolactone | antifungal | [334] |
| | koninginins A, D, E, F, L, M | phospholipase A 2 inhibitors | [335, 337, 338] |
| | stigmasterol | antifungal | [334] |
| | trichodermaketones A-D (polyketide derivatives) | antimicrobial | [335, 337] |
| | trichokonins (peptaibols) | antimicrobial | [339-341] |
| <i>Trichoderma longibranchiatum</i> | longibranchins (LGA I-V and LGB II-III) (peptaibols) | bioactive compounds | [342-345] |

| Microorganism | Secondary Metabolites | Comments | Ref. |
|--------------------------------|-------------------------------------|------------------------|-------|
| | trichobrachin A, C (peptaibols) | biactive compounds | [346] |
| | trichogin GA IV (peptaiboils) | bioactive compound | [347] |
| | trilongins BI-BIV (peptaibols) | mitochondriotoxic | [348] |
| <i>Trichoderma reesei</i> | fusarinine | siderophore | [349] |
| | kojic acid | antibacterial activity | [350] |
| | trichoderide A (cyclotetrapeptide) | bioactive compound | [351] |
| | trichodermatide A-D | bioactive compound | [352] |
| | trichodermin (trichothecene) | mycotoxin | [42] |
| <i>Trichoderma viride</i> | diacetoxyscirpenol | mycotoxin | [353] |
| | kojic acid | antibacterial activity | [350] |
| | lovastatin | bioactive compound | [122] |
| | trichovirins II (peptaibols) | antimicrobial | [354] |
| | T2-toxin | mycotoxin | [124] |
| <i>Yarrowia lipolytica</i> | Y-decolactone | | [355] |
| Syn. <i>Candida lipolytica</i> | | | |

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Appendix H – Relational database

| Entity | Name | description | dataType | catalogueCode | isNullable | precision | scale | maxLength |
|---------------|-----------------------|---|-----------|---------------|------------|-----------|-------|-----------|
| SOURCES | id_op (PK) | Primary key for the source | VARCHAR2 | | FALSE | | | 20 |
| SOURCES | op_type | Type of document | VARCHAR2 | REF_TYPE | FALSE | | | 400 |
| SOURCES | author | Author of the journal article | VARCHAR2 | | TRUE | | | 255 |
| SOURCES | TITLE | Title of the journal article | VARCHAR2 | | FALSE | | | 1000 |
| SOURCES | PUBLICATION_DATE | Date of the publication (yyyymmdd) | NUMBER | | FALSE | 8 | 0 | |
| SOURCES | JOURNAL_TITLE | Title of the journal where has been published the article | VARCHAR2 | | TRUE | | | 1000 |
| SOURCES | DOI | Digital Object Identifier of the journal article | VARCHAR2 | | TRUE | | | 255 |
| SOURCES | URL | the jornal link (internet address) or publisher | VARCHAR2 | | TRUE | | | 1000 |
| SOURCES | id_language | Language used to fill in the free text fields (ISO-639-1) from LANG catalogue | CATALOGUE | LANG | TRUE | | | 400 |
| MICROORGANISM | id_microorganism (PK) | Primary key for the identification of the microorganism | NUMBER | | FALSE | | | 20 |
| MICROORGANISM | id_species | microorganism used in the biotechnological process | CATALOGUE | MTX | FALSE | | | |
| MICROORGANISM | TAXON_LINEAGE | Contains a general classification of the microorganism (molds/yeast/bacteria) | VARCHAR2 | | FALSE | | | 255 |
| MICROORGANISM | TAXONOMY_METHOD | Acceptable scientific methodologies and techniques for the identification of microorganisms | CATALOGUE | ANLYMD | TRUE | | | |
| MICROORGANISM | method_reliability | Measure of the consistency of the taxonomic method (% Homology/Clostenest) | NUMBER | | TRUE | 3 | 10 | |
| MICROORGANISM | methodREMARK | Additional remarks on taxonomy study | VARCHAR2 | | TRUE | | | 2000 |
| TOXICITY | id_tox (PK) | Primary key for the identification of the toxicology data | NUMBER | | FALSE | | | 20 |
| TOXICITY | paramCode | Toxic compound described according to the Substance Code from PARAM catalogue | CATALOGUE | PARAM | FALSE | | | 400 |
| TOXICITY | test_species | Organism/cell culture used in the toxicological study | CATALOGUE | MTX | TRUE | | | |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Entity | Name | description | dataType | catalogueCode | isNullable | precision | scale | maxLength |
|--------------------|----------------------------|--|-----------|---------------|------------|-----------|-------|-----------|
| TOXICITY | testsubstance | Description of test material used in the toxicological study | VARCHAR2 | | TRUE | | | 2000 |
| TOXICITY | id_test_type | Type of toxicological test | CATALOGUE | TEST_TYPE | FALSE | | | 400 |
| TOXICITY | strain | The strain of the organism tested | CATALOGUE | STRAIN | TRUE | | | |
| TOXICITY | id_route | Indicator how the substance is administered to the organism (human/animals) | CATALOGUE | ROUTE_EXP | TRUE | | | |
| TOXICITY | id_endpoint | Endpoint reported in the study to describe the reported values (e.g. NOAEL, dose level) | CATALOGUE | ENDPOINT_HGV | FALSE | | | 400 |
| TOXICITY | id_qualifier | Qualifier for the reported endpoint values (e.g. =, <=, >=) | CATALOGUE | QUALIFIER | TRUE | | | 400 |
| TOXICITY | id_dose_unit | Enumeration of group units for group assessment | CATALOGUE | UNIT | TRUE | | | |
| TOXICITY | value_effect_concentration | Effect concentration | NUMBER | | TRUE | 5 | 1 | |
| TOXICITY | id_toxicity | Classification of critical effect according to toxicity target (Owens 2002) | CATALOGUE | TOXICITY | TRUE | | | |
| TOXICITY | effect_desc | Free text to describe effects observed in the toxicological study | VARCHAR2 | | TRUE | | | 2000 |
| TOXICITY | id_basis | Characterisation of the measures toxicological outcome measure | CATALOGUE | BASIC_EFFECT | FALSE | | | 400 |
| TOXICITY | remarks | Additional remarks on toxicological study. Free text on hazard assessment including (if necessary): 1) short explanation on how the study has been carried on; 2) any conclusions on the hazard identification | VARCHAR2 | | TRUE | | | 2000 |
| PROCESS CONDITIONS | id_process conditions (PK) | Primary key for the identification of the process conditions | NUMBER | | FALSE | | | 20 |
| PROCESS CONDITIONS | id_species | microorganism used in the biotechnological process | CATALOGUE | MTX | FALSE | | | |
| PROCESS CONDITIONS | temperature | Temperature in degrees centigrade | NUMBER | | TRUE | 2 | 10 | |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Entity | Name | description | dataType | catalogueCode | isNullable | precision | scale | maxLength |
|-----------------------|---------------------------|--|-----------|------------------|------------|-----------|-------|-----------|
| PROCESS CONDITIONS | aeration | Aerobic/Anaerobic/Microaerophilic conditions | VARCHAR2 | | TRUE | | | 255 |
| PROCESS CONDITIONS | waterActivity | Water activity | NUMBER | | TRUE | 1 | 10 | |
| PROCESS CONDITIONS | substrate | Type of substrate used in the biotechnological process | VARCHAR2 | | TRUE | | | 300 |
| PROCESS CONDITIONS | pH | pH during the fermentation process | NUMBER | | TRUE | 2 | 10 | |
| PROCESS CONDITIONS | FermentationTime | Time in hours of the fermentation process | NUMBER | | TRUE | 20 | 10 | |
| PROCESS CONDITIONS | prodTreat | Used to characterise a food product based on the treatment or processes applied to the product or any indexed ingredient from PRODTR catalogue | CATALOGUE | PRODTR catalogue | TRUE | | | |
| PROCESS CONDITIONS | fermentationState | Types of fermentation: solid State, semi-solid, submerged batch, submerged fed-batch, submerged continuous, immobilized cell bioreactors, immobilized enzyme bioreactors | NUMBER | | TRUE | 1 | 0 | |
| PROCESS CONDITIONS | paramCode | Toxic compound described according to the Substance Code from PARAM catalogue | CATALOGUE | PARAM | FALSE | | | |
| PROCESS CONDITIONS | processRemarks | Additional remarks on process conditions. Free text | VARCHAR2 | | TRUE | | | 2000 |
| BIOSYNTHETIC PATHWAYS | id_biosynthetic path (PK) | Primary key for the identification of the synthesis pathway of the metabolite | NUMBER | | FALSE | | | 20 |
| BIOSYNTHETIC PATHWAYS | id_species | microorganism used in the biotechnological process | CATALOGUE | MTX | FALSE | | | |
| BIOSYNTHETIC PATHWAYS | paramCode | Toxic compound described according to the Substance Code from PARAM catalogue | CATALOGUE | PARAM | FALSE | | | |
| BIOSYNTHETIC PATHWAYS | MapURL | link to the diagram where the route of synthesis of the substance is represented | VARCHAR2 | | TRUE | | | 2000 |
| BIOSYNTHETIC PATHWAYS | geneMod | Genetic modification (YES/NO) | VARCHAR2 | | TRUE | | | 255 |

Database on potential toxigenic capacities of microorganisms used for industrial production

| Entity | Name | description | dataType | catalogueCode | isNullable | precision | scale | maxLength |
|-----------------------|----------------------------|--|----------|---------------|------------|-----------|-------|-----------|
| BIOSYNTHETIC PATHWAYS | geneMod_REMARKS | Genetic modification (information of genes inserted or deleted) | VARCHAR2 | | TRUE | | | 2000 |
| BIOSYNTHETIC PATHWAYS | biosynthetic_remarks | Additional remarks on biosynthetic pathways. Free text | VARCHAR3 | | TRUE | | | 2000 |
| FACT_PROCESSTOXINS | id_ProcessToxins (PK) | Unique ID for fact table | NUMBER | | FALSE | | | 20 |
| FACT_PROCESSTOXINS | id_op (FK) | Foreing key for the source | NUMBER | | FALSE | | | 20 |
| FACT_PROCESSTOXINS | id_microorganism (FK) | Foreing key for the identification of the microorganism | NUMBER | | TRUE | | | 20 |
| FACT_PROCESSTOXINS | id_tox (FK) | Foreing key for the identification of the toxicology data | NUMBER | | TRUE | | | 20 |
| FACT_PROCESSTOXINS | id_process conditions (FK) | Foreing key for the identification of the process conditions | NUMBER | | TRUE | | | 20 |
| FACT_PROCESSTOXINS | id_biosysthetic path (FK) | Foreing key for the identification of the synthesis pathway of the metabolite | NUMBER | | TRUE | | | 20 |
| GENE | id_gene (PK) | Primary key for the identification of the gene | NUMBER | | FALSE | | | 20 |
| GENE | gene_name | Gene name involved in the synthesis of the substance | VARCHAR2 | | FALSE | | | 100 |
| GENE | protein_name | Name of protein synthesised from the gene | VARCHAR2 | | TRUE | | | 45 |
| GENE | gene_description | Free text to describe the function of the gene | VARCHAR2 | | TRUE | | | 75 |
| GENE | gene_sequence | sequence of the involved in the synthesis route | VARCHAR2 | | TRUE | | | 3000 |
| BIO_PATHWAY2GENE | id_bio_pathway (PK & FK) | Foreign key for the identification of the biosynthetic pathway and Primary key in the relationship | NUMBER | | FALSE | | | 20 |
| BIO_PATHWAY2GENE | id_gene (PK & FK) | Foreing key for the identification of the gene and Primary key in the relationship | NUMBER | | FALSE | | | 20 |
| MICROORG2GENE | id_microorg (PK & FK) | Foreing key for the identification of the microorganism taxonomy and Primary key in the relationship | NUMBER | | FALSE | | | 20 |
| MICROORG2GENE | id_gene (PK & FK) | Foreing key for the identification of the gene and Primary key in the relationship | NUMBER | | FALSE | | | 20 |
| MICROORG2GENE | housekeeping_gene_GenBank | GenBank ID for the housekeeping gene | VARCHAR2 | | TRUE | | | 45 |

Appendix I – Attributes for Param Catalogue

| code | extendedName | shortName | reportable | IUPAC | CAS | MOLECULAR_FORMULA | SMILES_NOTATION | INCHI |
|-----------------|--------------------|-----------|------------|---|-----------------|-------------------|---|---|
| RF-00004558-PAR | 1-Hydroxyphenazine | (1-HP) | Yes | 5H-phenazin-1-one | CAS 528-71-2 | C12H8N2O | C1=CC=C2C(=C1)NC3=CC=CC(=O)C3=N2 | NAD |
| RF-00004547-PAR | Andrastin A | - | Yes | (3S,5R,8S,9R,10S,13R,14R)-methyl 3-acetoxy-10-formyl-4,4,8,12,13,16-hexamethyl-15,17-dioxo-2,3,4,5,6,7,8,9,10,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthrene-14-carboxylate | CAS 174232-42-9 | C28H38O7 | CC1C([C@]2(C)C(C)=C[C@]3([H])[C@@]([C@@]([C@@]4[C@@]([C@@]3C(=O)CC[C@@H](OC(C)=O)C4(C)C)C)C)2(C(OC)=O)C1=O)=O | InChI=1S/C28H38O7/c1-15-13-19-25(6,28(23(33)34-8)22(32)16(2)21(31)26(15,28)7)11-9-18-24(4,5)20(35-17(3)30)10-12-27(18,19)14-29/h13-14,16,18-20H,9-12H2,1-8H3/t16?,18-,19-,20+,25+,26+,27+,28-/m1/s1 |
| RF-00004573-PAR | Aristolochene | - | Yes | (4S,4aR,6S)-4,4a-dimethyl-6-prop-1-en-2-yl-2,3,4,5,6,7-hexahydro-1H-naphthalene | NAD | C15H24 | C[C@H]1CCCC2=CC[C@@H](C[C@]12C)C(=C)C | InChI=1S/C15H24/c1-11(2)13-8-9-14-7-5-6-12(3)15(14,4)10-13/h9,12-13H,1,5-8,10H2,2-4H3/t12-,13-,15+/m0/s1 |
| RF-00004548-PAR | Asperparaline A | - | Yes | (1R,5aR,7R,8aR,9aS)-1,1',8,8,11-pentamethyltetrahydro-1H,2'H,5'H,8H,10H-spiro[5a,9a-(epiminomethano)cyclopenta[f]indolizine-7,3'-pyrrolidine]-2',5',10-trione | CAS 195966-93-9 | C20H29N3O3 | C[C@@H]1CCN2C[C@@]34C[C@@]5(CC(=O)N(C)C5=O)C(C)C[C@H]3C[C@@]12C(=O)N4C | InChI=1S/C20H29N3O3/c1-12-6-7-23-11-19-10-18(9-14(24)21(4)15(18)25)17(2,3)13(19)8-20(12,23)16(26)22(17)5/h12-13H,6-11H2,1-5H3/t12-,13-,18-,19+,20+/m0/s1 |
| RF-00004572-PAR | Citreoviridin | - | Yes | 6-[(1E,3E,5E,7E)-8-[(2S,3R,4R,5R)-3,4-dihydroxy-2,4,5-trimethylxolan-2-yl]-7-methylocta-1,3,5,7-tetraenyl]-4-methoxy-5-methylpyran-2-one | NAD | C23H30O6 | C[C@@H]1[C@@]([C@H]([C@]([C@]([C@]([C@]1O)C)C=C(C)C)C)C)C(=O)C(C)C(=O)C2=CC(=O)O2)OC(C)O(C)O | InChI=1S/C23H30O6/c1-15(14-22(4)21(25)23(5,26)17(3)29-22)11-9-7-8-10-12-18(16)2(19(27-6)13-20(24)28-18/h7-14,17,21,25-26H,1-6H3/b8-7+,11-9+,12-10+,15-14+/t17-,21+,22+,23+/m1/s1 |
| RF-00004574-PAR | Culmorin | - | Yes | NAD | NAD | C15H26O2 | CC1(CCCC2(C3C1C(C2(CC3O)C)O)C)C | InChI=1S/C15H26O2/c1-13(2)6-5-7-14(3)10-9(16)8-15(14,4)12(17)11(10)13/h9-12,16-17H,5-8H2,1-4H3 |
| RF-00004579-PAR | Cyclopenol | - | Yes | 3'-(3-hydroxyphenyl)-4-methylspiro[1H-1,4-benzodiazepine-3,2'-oxirane]-2,5-dione | NAD | C17H14N2O4 | CN1C(=O)C2=CC=CC=C2NC(=O)C13C(O3)C4=CC(=CC=C4)O | InChI=1S/C17H14N2O4/c1-19-15(21)12-7-2-3-8-13(12)18-16(22)17(19)14(23-17)10-5-4-6-11(20)9-10/h2-9,14,20H,1H3,(H,18,22) |
| RF-00004562-PAR | Cyclopiazonic acid | CPA | Yes | (6aR,11aS,11bR)-10-Acetyl-11-hydroxy-7,7-dimethyl-2,6,6a,7,11a,11b-hexahydro-9H-pyrrolo[1',2':2,3]isoindolo[4,5,6-cd]indol-9-one | CAS 18172-33-3 | C20H20N2O3 | CC(=O)/C1=C(O)[C@H]5N(C1=O)[C@@](C[C@@@H]4Cc2cccc3ncc(c23)[C@@H]45 | NAD |
| RF-00004559-PAR | Cytolysin A | ClyA | Yes | NAD | NAD | NAD | NAD | NAD |
| RF-00004560-PAR | Endophenazine A | - | Yes | 9-(3-Methyl-2-buten-1-yl)-1-phenazinecarboxylic acid | CAS 86125-71-5 | C18H16N2O2 | CC(=CCC1=C2C(=CC=C1)N=C3C=CC=C(C3=N2)C(=O)O)C | NAD |
| RF-00004549-PAR | Fusarin C | - | Yes | methyl (2E,3E,5E,7E,9E)-2-ethylidene-11-[(1R,4S,5R)-4-hydroxy-4-(2-hydroxyethyl)-2-oxo-6-oxa-3-azabicyclo[3.1.0]hexan-1-yl]-4,6,10-trimethyl-11-oxoundeca-3,5,7,9-tetraenoate | NAD | C23H29NO7 | C/C=C(C=C(C)/C)C=C(C)/C=C(C/C)C(=O)[C@@]12[C@@H]([O1][C@]([NC2=O](CCO)O)C(=O)OC | InChI=1S/C23H29NO7/c1-6-17(19(27)30)5)13-15(3)12-14(2)8-7-9-16(4)18(26)23-20(31-23)22(29,10-11-25)24-21(23)28/h6-9,12-13,20,25,29H,10-11H2,1-5H3,(H,24,28)/b8-7+,14-12+,15-13+,16-9+,17-6+/t20-,22-,23-/m0/s1 |
| RF-00004575-PAR | Isofumigaclavine A | - | Yes | [(6aR,9S,10R)-7,9-dimethyl-6,6a,8,9,10,10a-hexahydro-4H-indolo[4,3-fg]quinoline-10-yl] acetate | NAD | C18H22N2O2 | C[C@H]1CN([C@@H]2CC3=CNC4=CC=CC(=C34)C2[C@@H]1O(C=O)C)C | InChI=1S/C18H22N2O2/c1-10-9-20(3)15-7-12-8-19-14-6-4-5-13(16(12)14)17(15)18(10)22-11(2)21/h4-6,8,10,15,17-19H,7,9H2,1-3H3/t10-,15+,17?,18+/m0/s1 |

Database on potential toxigenic capacities of microorganisms used for industrial production

| code | extendedName | shortName | reportable | IUPAC | CAS | MOLECULAR_FORMULA | SMILES_NOTATION | INCHI |
|-----------------|--------------------|-----------|------------|---|----------------|-------------------|---|---|
| RF-00004576-PAR | Isofumigaclavine B | - | Yes | (6aR,9S,10R,10aR)-7,9-dimethyl-6,6a,8,9,10,10a-hexahydro-4H-indolo[4,3-fg]quinoline-10-ol | NAD | C16H20N2O | C[C@H]1CN([C@@H]2CC3=CNC4=CC=CC(=C34)C[C@@H]2[C@H]1O)C | InChI=1S/C16H20N2O/c1-9-8-18(2)13-6-10-7-17-12-5-3-4-11(14(10)12)15(13)16(9)19/h3-5,7,9,13,15-17,19H,6,8H2,1-2H3/t9-,13+,15+,16+/m0/s1 |
| RF-00004577-PAR | Isotrichodermin | - | Yes | NAD | NAD | C17H24O4 | CC1=C[C@@H]2[C@@](CC1)(C3[C@H]([C@H](C34CO4)O2)OC(=O)C)C | InChI=1S/C17H24O4/c1-10-5-6-15(3)13(7-10)21-14-12(20-11(2)18)8-16(15,4)17(14)9-19-17/h7,12-14H,5-6,8-9H2,1-4H3/t12-,13-,14-,15+,16?,17?/m1/s1 |
| RF-00004578-PAR | Isotricodermol | - | Yes | NAD | NAD | C15H22O3 | CC1=C[C@@H]2[C@@](CC1)([C@]3[C@H]([C@H](C34CO4)O2)O)C | InChI=1S/C15H22O3/c1-9-4-5-13(2)11(6-9)18-12-10(16)7-14(13,3)15(12)8-17-15/h6,10-12,16H,4-5,7-8H2,1-3H3/t10-,11-,12-,13+,14-,15?/m1/s1 |
| RF-00004561-PAR | Kojic acid | - | Yes | 5-Hydroxy-2-(hydroxymethyl)-4H-pyran-4-one | CAS 501-30-4 | C6H6O4 | O=C1C=C(O)C=C1O)CO | NAD |
| RF-00004563-PAR | Malformins | - | Yes | 10-butan-2-yl-7-(2-methylpropyl)-4-propan-2-yl-15,16-dithia-3,6,9,12,18-pentazabicyclo[11.4.2]nonadecane-2,5,8,11,19-pentone | CAS 3022-92-2 | C23H39N5O5S2 | CCC(C)C1C(=O)NC2CSSCC(C(=O)NC(C(=O)NC(C(=O)N)1CC(C)C)C)NC2=O | NAD |
| RF-00004564-PAR | Maltoryzine | - | Yes | 1-(2,3,6-trihydroxyphenyl)pent-3-en-1-one | CAS 6826-42-2 | C11H12O4 | OC1=C(O)C=CC(O)=C1C(/C=C/CC)=O | NAD |
| RF-00004580-PAR | Marcfortines | - | Yes | | CAS 75731-43-0 | C28H35N3O4 | NAD | NAD |
| RF-00004550-PAR | Mitorubrinic acid | - | Yes | (2E)-3-(7-[(2,4-Dihydroxy-6-methylbenzoyloxy]-7-methyl-6,8-dioxo-7,8-dihydro-6H-isochromen-3-yl)acrylic acid | CAS 58958-07-9 | C21H16O9 | Cc1cc(cc(c1C(=O)OC2(C(=O)C=C3C=C(OC=C3C2=O)/C=C(C(=O)O)C)O)O | InChI=1S/C21H16O9/c1-10-5-12(22)8-15(23)18(10)20(28)30-21(2)16(24)7-11-6-13(3-4-17(25)26)29-9-14(11)19(21)27/h3-9,22-23H,1-2H3,(H,25,26)/b4-3+ |
| RF-00004565-PAR | Naphtho-γ-pyrones | - | Yes | 1H-Benzo[f]chromen-1-one | NAD | C13H8O | O=C3/C=C\Oc2ccc1ccccc1c23 | NAD |
| RF-00004551-PAR | Neoxaline | - | Yes | (14E)-11-HYDROXY-14-[(1H-IMIDAZOL-5-YL)METHYLIDENE]-2-METHOXY-9-(2-METHYLBUT-3-EN-2-YL)-2,13,16-TRIAZATETRACYCLO[7.7.0.0.1 ¹³ .0 ³ .8]HEXADECAN-3,5,7-TRIFENE-12,15-DIONE | CAS 71812-10-7 | C23H25N5O4 | CC(C)(C=C)C12CC(C(=O)N)3C1(NC(=O)/C3=C(C4=CN=CN4)N(C5=CC=CC=C5)OC)O | InChI=1S/C23H25N5O4/c1-5-21(2,3)22-11-18(29)20(31)27-17(10-14-12-24-13-25-14)19(30)26-23(22,27)28(32-4)16-9-7-6-8-15(16)22/h5-10,12-13,18,29H,1,11H2,2-4H3,(H,24,25)(H,26,30)/b17-10+ |
| RF-00004566-PAR | Nigerazines | - | Yes | (E)-3-phenyl-1-[(2S,5R)-2,4,5-trimethylpiperazin-1-yl]prop-2-en-1-one | NAD | C16H22N2O | C[C@H]1CN([C@@H]([CN1C(=O)]/C=C/C2=CC=CC=C2)C | InChI=1S/C16H22N2O/c1-13-12-18(14(2)11-17(13)3)16(19)10-9-15-7-5-4-6-8-15/h4-10,13-14H,11-12H2,1-3H3/b10-9+/t13-,14+/m1/s1 |
| RF-00004567-PAR | Nigragillin | - | Yes | (2E,4E)-1-[(2R,5S)-2,4,5-trimethylpiperazin-1-yl]hexa-2,4-dien-1-one | CAS 24779-38-2 | C13H22N2O | C/C=C/C=C/C(=O)N1C[C@@H](N(C[C@H]1C)C)C | InChI=1S/C13H22N2O/c1-5-6-7-8-13(16)15-10-11(2)14(4)9-12(15)3/h5-8,11-12H,9-10H2,1-4H3/b6-5+,8-7+/t11-,12+/m0/s1 |
| RF-00004583-PAR | Orlandin | - | Yes | 7-hydroxy-8-(7-hydroxy-4-methoxy-5-methyl-2-oxochromen-8-yl)-4-methoxy-5-methylchromen-2-one | NAD | C22H18O8 | CC1=CC(=C(C2=C1C(=CC(=O)O2)OC)C3=C(C=C(C4=C3OC(=O)C=C4OC)C)O | InChI=1S/C22H18O8/c1-9-5-11(23)19(21-17(9)13(27-3)15(25)29-21)20-12(24)6-10(2)18-14(28-4)8-16(26)30-22(18)20/h5-8,23-24H,1-4H3 |
| RF-00004552-PAR | Penicillic acid | - | Yes | (2Z)-3-methoxy-5-methyl-4-oxohexa-2,5-dienoic acid | CAS 90-65-3 | C8H10O4 | CC(=C)C(=O)/C=C/C(=O)O)OC | InChI=1S/C8H10O4/c1-5(2)8(11)6(12-3)4-7(9)10/h4H,1H2,2-3H3,(H,9,10)/b6-4 |

Database on potential toxigenic capacities of microorganisms used for industrial production

| code | extendedName | shortName | reportable | IUPAC | CAS | MOLECULAR_FORMULA | SMILES_NOTATION | INCHI |
|-----------------|-----------------------------|-----------|------------|---|----------------|-------------------|---|--|
| RF-00004571-PAR | Perinadine A | - | Yes | NAD | NAD | C28H37NO7 | [C@H]12[C@H]3[C@H](N(CC3)C(=O)C(C(=O)CCCC/C=C/C)C)O3c1c([C@H]([C@H](O2)C)C)c(c(c3C(=O)O)O)C | InChI=1S/C28H37NO7/c1-6-7-8-9-10-11-19(30)15(3)26(32)29-13-12-18-24-21-20(14(2)17(5)35-24)16(4)23(31)22(28(33)34)25(21)36-27(18)29/h6-7,14-15,17-18,24,27,31H,8-13H2,1-5H3,(H,33,34)/b7-6+/t14-,15?,17-,18+,24+,27-/m1/s1 |
| RF-00004557-PAR | Phenazine-1-carboxylic acid | PCA | Yes | phenazine-1-carboxylic acid | CAS 2538-68-3 | C13H8N2O2 | C1=CC=C2C(=C1)N=C3C=CC=C(C3=N2)C(=O)O | NAD |
| RF-00004556-PAR | Phenazines | - | Yes | Phenazine | CAS 92-82-0 | C12H8N2 | n1c3c(nc2c1cccc2)cccc3 | NAD |
| RF-00004553-PAR | PR toxin | - | Yes | [(1aR,3R,3'S,3aR,5R,7bS)-3'-formyl-3,3',3a-trimethyl-6-oxospiro[2,3,4,7b-tetrahydro-1aH-naphtho[1,2-b]oxirene-5,2'-oxirane]-2-yl] acetate | CAS 56299-00-4 | C17H20O6 | C[C@H]1C[C@@H]2[C@@H](O2)C3=CC(=O)[C@@]4[C[C@]13C][C@@](O4)(C)C(=O)OC(=O)C | InChI=1S/C17H20O6/c1-8-12(21-9(2)19)14-13(22-14)10-5-11(20)17(6-15(8,10)3)16(4,7-18)23-17/h5,7-8,12-14H,6H2,1-4H3/t8-,12+,13-,14+,15+,16+,17-/m0/s1 |
| RF-00004555-PAR | Pyocyanin | PYO | Yes | 5-Methylphenazin-1-one | CAS 85-66-5 | C13H10N2O | CN1C2=CC=CC=C2N=C3C1=CC=CC3=O | NAD |
| RF-00004569-PAR | Roquefortine C | - | Yes | NAD | CAS 58735-64-1 | C22H23N5O3 | CC(C)(C=C)C12CC3C(=O)N/C(=C/C4=CN=CN4)/C(=O)N3C1N(C5=CC=CC=C5)O | InChI=1S/C22H23N5O3/c1-4-21(2,3)22-10-17-18(28)25-15(9-13-11-23-12-24-13)19(29)26(17)20(22)27(30)16-8-6-5-7-14(16)22/h4-9,11-12,17,20,30H,1,10H2,2-3H3,(H,23,24)(H,25,28)/b15-9+ |
| RF-00004582-PAR | Sambucinol | - | Yes | NAD | NAD | C15H22O4 | CC1=C[C@]23[C@](CC1)([C@]4[C[C@@H]([C@H]([C@]4(O2)CO)O3)O)C | InChI=1S/C15H22O4/c1-9-4-5-12(2)13(3)7-10(17)11-14(13,8-16)19-15(12,6-9)18-11/h6,10-11,16-17H,4-5,7-8H2,1-3H3/t10-,11+,12+,13+,14+,15+/m0/s1 |
| RF-00004554-PAR | Secalonic acid | - | Yes | methyl (3R,4S,4aS)-7-[(5S,6R,10aS)-1,5,9-trihydroxy-10a-methoxycarbonyl-6-methyl-8-oxo-6,7-dihydro-5H-xanthen-2-yl]-4,8,9-trihydroxy-3-methyl-1-oxo-3,4-dihydro-2H-xanthen-4a-carboxylate | CAS 35287-69-5 | C32H30O14 | C[C@@H]1CC(=O)C2=C(C3=C(C=CC(=C3O)C4=C(C5=C(C=C4)O[C@@]6([C@H]([C@@H](CC(=O)C6=C5O)C)O)C(=O)O)O)O[C@@]2([C@H]1O)C(=O)OC)O | InChI=1S/C32H30O14/c1-11-9-15(33)21-25(37)19-17(45-31(21,27(11)39)29(41)43-3)7-5-13(23(19)35)14-6-8-18-20(24(14)36)26(38)22-16(34)10-12(2)28(40)32(22,46-18)30(42)44-4/h5-8,11-12,27-28,35-40H,9-10H2,1-4H3/t11-,12-,27+,28+,31+,32+/m1/s1 |
| RF-00004568-PAR | Trichodermin | - | Yes | (4β,12R)-12,13-epoxytrichothec-9-en-4-yl acetate | CAS 4682-50-2 | C17H24O4 | CC1=C[C@@H]2[C@](CC1)([C@]3([C@@H](C[C@H]([C@@]34CO4)O2)OC(=O)C)C | NAD |
| RF-00004570-PAR | Xanthomegnin | - | Yes | (3R,3'R)-10,10'-Dihydroxy-7,7'-dimethoxy-3,3'-dimethyl-3,3',4,4'-tetrahydro-1H,1'H-8,8'-bibenzo[g]isochromene-1,1',6,6',9,9'-hexone | CAS 1685-91-2 | C30H22O12 | C[C@@H]1Cc2cc3c(c(c2C(=O)O1)O)C(=O)C(=C(C3=O)OC)C4=C(C(=O)c5cc6c(c(c5C4=O)O)C(=O)O[C@@H]([C@]6)C)OC | InChI=1S/C30H22O12/c1-9-5-11-7-13-17(23(33)15(11)29(37)41-9)25(35)19(27(39-3)21(13)31)20-26(36)18-14(22(32)28(20)40-4)8-12-6-10(2)42-30(38)16(12)24(18)34/h7-10,33-34H,5-6H2,1-4H3/t9-,10-/m1/s1 |

NAD: Non-available data

Database on potential toxigenic capacities of microorganisms used for industrial production

Appendix J - Microorganism taxonomical identifiers

Table J1. BACTERIA

| MICROORGANISMS NAME BY PROCUREMENT/ CURRENT NAME | SYNONYM NAMES | TAXONOMY KINGDOM PHYLUM/ORDER/FAMILY | TAXONOMY REFERENCES | METHODS SUMMARY | HOUSEKEEPING GENES SEQUENCE ID GENBANK | HOUSEKEEPING GENES | REFERENCES HOUSEKEEPING GENES** | GENOME ID | CONCLUSION |
|--|---|---|---------------------|--|--|---|---------------------------------|---|---|
| <i>Actinomadura yumaensis</i> | <i>Actinomadura yumaense</i> | BACTERIA/Actinobacteria/Actinomycetales/Thermomonosporaceae | [1] | Scanning electron microscopy, cell chemistry, physiological tests | AF163122.1 | 16S rRNA gene | [85] | NAD | Current taxonomy valid update <i>Actinomadura yumaensis</i> 35:335* |
| <i>Actinomadura roseorufa</i> | NAD | - | [2] | Information from a Patent | NAD | NAD | NAD | NAD | No Taxonomy data available/ Not validated name |
| <i>Arthrobacter ramosus</i> | NAD | BACTERIA/Actinobacteria/Actinomycetales/Micrococcaceae | [3] | 16S rRNA gene for the type strain: AM039435. | AM039435 | 16S rRNA gene | [86] | NAD | Current taxonomy valid update <i>Arthrobacter ramosus</i> 30:254 (AL) |
| <i>Cellulosimicrobium cellulans</i> | <i>Brevibacterium fermentans</i> <i>Brevibacterium lyticum</i> | BACTERIA/Actinobacteria/Actinomycetales/Promicromonosporaceae | [4] | 16S rRNA sequence analyses | X83809 | 16S rRNA gene | [80] | https://www.ncbi.nlm.nih.gov/genome/genomes/15565 | Current taxonomy valid update <i>Brevibacterium lyticum</i> 30:269 (AL) = <i>Cellulosimicrobium cellulans</i> |
| <i>Chryseobacterium proteolyticum</i> | NAD | BACTERIA/Bacteroidetes/Flavobacteriales/Flavobacteriaceae | [5] | Physiological and biochemical characteristics, 16S rRNA gene sequence analysis | AB039830 | 16S rRNA gene | [5] | NAD | Not validated name |
| <i>Escherichia coli</i> | NAD | BACTERIA/Proteobacteria/Enterobacteriales/Enterobacteriaceae | [6] | Comparative analysis of 5S and 16S rRNA sequences | X80725 | 16S rRNA gene | [79] | https://www.ncbi.nlm.nih.gov/genome/genomes/167 | Current taxonomy valid update <i>Escherichia coli</i> 30:296 (AL) |
| | | | | | KJ858767.1 | <i>adk</i> (adenylate kinase) | [57; 83] | | |
| | | | | | KJ868320.1 | <i>fumC</i> (fumarate hydratase) | [57] | | |
| | | | | | DQ447149.1 | <i>gyrB</i> (DNA gyrase) | [57] | | |
| | | | | | KJ868480.1 | <i>icd</i> (isocitrate/isopropylmalate dehydrogenase) | [57] | | |
| | | | | | KJ868560.1 | <i>mdh</i> (malate dehydrogenase) | [57] | | |
| | | | | | KJ868640.1 | <i>purA</i> (adenylosuccinate dehydrogenase) | [57] | | |
| | | | | | JQ907532.1 | <i>recA</i> (ATP/GTP binding motif) | [57] | | |
| <i>Geobacillus stearothermophilus</i> | <i>Bacillus stearothermophilus</i> | BACTERIA/Firmicutes/Bacillales/Bacillaceae | [7] | Phenotypic characterization, Chemotaxonomic characterization (fatty acid methyl ester analysis), Genotypic characterization (16S rRNA gene sequencing) | FN428694 | 16S rRNA gene | [78] | https://www.ncbi.nlm.nih.gov/genome/genomes/673 | Current taxonomy valid update <i>Geobacillus stearothermophilus</i> 51:443* |
| | | | | | - | genes of carbohydrate metabolism | [58; 59] | | |
| <i>Geobacillus caldoproteolyticus</i> | <i>Anoxybacillus caldoproteolyticus</i> | BACTERIA/Firmicutes/Bacillales/Bacillaceae | [119; 7] | 16S rRNA gene sequence analysis; and rpoB sequencing | FN428698 | 16S rRNA gene | [7] | NAD | Current taxonomy valid update <i>Anoxybacillus caldoproteolyticus</i> 62:1483* |
| | | | | | DQ642082.1 | rpoB gene | [58; 59; 120] | | |
| <i>Geobacillus pallidus</i> | <i>Aeribacillus pallidus</i> | BACTERIA/Firmicutes/Bacillales/Bacillaceae | [7] | 16S rRNA sequence analyses | Z26930 | 16S rRNA gene | [69] | https://www.ncbi.nlm.nih.gov/genome/genomes/14084 | Current taxonomy valid update <i>Geobacillus pallidus</i> 54:2200 ≡ <i>Aeribacillus pallidus</i> |
| | | | | | DQ642059.1 | rpoB gene | [58; 59; 120] | | |

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|--|--|--|---------------------|---|--|---|--|---|---|
| <i>Klebsiella planticola</i> | <i>Raoultella planticola</i> | BACTERIA/Proteobacteria/Enterobacteriales/Enterobacteriaceae | [8] | Biochemical tests, 16S rRNA and rpoB sequencing. | AF129443.1 | 16S rRNA gene | [8] | https://www.ncbi.nlm.nih.gov/genome/genomes/11487 | Current taxonomy valid update <i>Klebsiella planticola</i> 32:266 ≡ <i>Raoultella planticola</i> |
| <i>Klebsiella pneumoniae</i> | NAD | BACTERIA/Proteobacteria/Enterobacteriales/Enterobacteriaceae | [8] | Biochemical tests, 16S rRNA and rpoB sequencing; MLSA. | X87276 | 16S rRNA gene | [77] | https://www.ncbi.nlm.nih.gov/genome/genomes/815 | Current taxonomy valid update <i>Klebsiella pneumoniae</i> 34:355 |
| | | | | | - | <i>folA</i> (dihydrofolate reductase) | [60; 61; 62] | | |
| | | | | | AF146532.1 | <i>waa</i> gene cluster | [60; 61; 62] | | |
| | | | | | FJ769969.1 | <i>gnd</i> (6-phosphogluconate dehydrogenase) | [60; 61; 62] | | |
| - | <i>ugd</i> (UDP-glucose 6-dehydrogenase) | [60; 61; 62] | | | | | | | |
| <i>Microbacterium imperiale</i> | <i>Brevibacterium imperiale</i> | BACTERIA/Actinobacteria/Actinomycetales/MicroBACTERIA/ceae | [9] | sequence comparison of <i>gyrB</i> , <i>rpoB</i> , <i>recA</i> and <i>ppk</i> and 16S rRNA genes | X77442 | 16S rRNA gene | [76] | NAD | Current taxonomy valid update <i>Brevibacterium imperiale</i> 30:269 (AL) ≡ <i>Microbacterium imperiale</i> |
| <i>Micrococcus luteus</i> | <i>Micrococcus lysodeikticus</i> | BACTERIA/Actinobacteria/Actinomycetales/Micrococcaceae | [10] | morphological, cultural, and biochemical studies | AJ536198 | 16S rRNA gene *Type strain | Swiderski J. Unpublished (2003) | https://www.ncbi.nlm.nih.gov/genome/genomes/888 | Current taxonomy valid update <i>Micrococcus luteus</i> 30:320 (AL) |
| <i>Paenibacillus alginolyticus</i> | <i>Bacillus alginolyticus</i> | BACTERIA/Firmicutes/Bacillales Paenibacillaceae | [11] | 16S rRNA gene sequence and cellular fatty acid composition analyses | AB073362 | 16S rRNA gene | [75] | https://www.ncbi.nlm.nih.gov/genome/genomes/14779 | Current taxonomy valid update/ <i>Paenibacillus alginolyticus</i> 47:295* |
| <i>Paenibacillus lentus</i> | NAD | BACTERIA/Firmicutes/Bacillales/Paenibacillaceae | [12] | 16S rRNA gene sequence analysis | KC800716 | 16S rRNA gene | [12] | NAD | Current taxonomy valid update / <i>Paenibacillus lentus</i> 64:1171* |
| <i>Paenibacillus macerans</i> | <i>Bacillus macerans</i> , <i>Aerobacillus macerans</i> , <i>Zymobacillus macerans</i> , <i>Bactrillum macerans</i> , <i>Bacillus acetoethylicum</i> , <i>Bacillus betanigrificans</i> , <i>Bacillus soli</i> , <i>Aerobacillus schuykilliensis</i> , <i>Bacillus vagans</i> | BACTERIA/Firmicutes/Bacillales/Paenibacillaceae | [23] | sequence analysis of the <i>rplK</i> , sequence analysis of the 26 N-terminal amino acid residues of ribosomal L30 proteins | AB073196.1 | 16S rRNA gene | [75] | https://www.ncbi.nlm.nih.gov/genome/genomes/33271 | Current taxonomy valid update / <i>Paenibacillus macerans</i> 45:197 |
| <i>Pseudomonas aeruginosa</i> | NAD | BACTERIA/Proteobacteria/Pseudomonadales/Pseudomonadaceae | [13; 122] | 16S rRNA sequence analyses; Housekeeping genes: <i>rpoD</i> ; <i>gyrB</i> MLSA. | HE978271 /NR_026078.1 | 16S rRNA <i>gene</i> | Swiderski J. Unpublished (2012)/ [121] | https://www.ncbi.nlm.nih.gov/genome/genomes/187 | Current taxonomy valid update / <i>Pseudomonas aeruginosa</i> 30:349 (AL) |
| | | | | | - | phzA1-G1 | [63] | | |
| | | | | | - | phzA2-G2 | [63] | | |
| | | | | | KR824168.1 | phzM | [63] | | |
| | | | | | AF116283.1 | mexG | [63] | | |
| | | | | | U89892.1 | vfr | [63] | | |
| | | | | | FJ649224.1 | algU | [63] | | |
| | | | | | U27988.1 | gacA | [63] | | |
| | | | | | JN868722.1 | phoP | [63] | | |
| | | | | | AY280952.1 | mvaT | [63] | | |
| - | algR | [63] | | | | | | | |
| - | phoB | [63] | | | | | | | |

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|--|---------------------------------------|---|---------------------|--|--|--------------------|---|---|--|
| <i>Pseudomonas amyloclavata</i> | NAD | BACTERIA/ Proteobacteria/ Pseudomonadales/ Pseudomonadaceae | [13] | 16S rRNA sequence analyses: Housekeeping genes and MLSA. | NAD | NAD | NAD | NAD | No Taxonomy data available/ Not validated name |
| <i>Pullulanibacillus naganoensis</i> | <i>Bacillus naganoensis</i> | BACTERIA/Firmicutes/ Bacillales/Bacillaceae | [14] | 16S rRNA sequence analyses | AB021193/ NR_024694.1 | 16S rRNA gene | [74] | NAD | Current taxonomy valid update / <i>Pullulanibacillus naganoensis</i> 56:2550* |
| <i>Sphingobacterium multivorum</i> | <i>Flavobacterium multivorum</i> | BACTERIA/Bacteroidetes/ Sphingobacteriales/ Sphingobacteriaceae | [15; 16] | phenotypic characters, DNA-DNA hybridization | AB100738 | 16S rRNA gene | [87] | https://img.igi.doe.gov/cgi-bin/m/main.cgi?section=ANI&page=cliqDetails&cliq_id=8537 | Current taxonomy valid update / <i>Flavobacterium multivorum</i> 31:33* ≡ <i>Sphingobacterium multivorum</i> |
| <i>Streptomyces albus</i> | NAD | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [17; 18] | Chemotaxonomic methods, DNA-DNA hybridization, LFRFA, RAPD-PCR assays, 16S rRNA, 23S rRNA, 5S rRNA genes, RFLPs of rRNA | AJ621602 | 16S rRNA gene | Swiderski J. Direct Submission (2004) | https://www.ncbi.nlm.nih.gov/genome/genomes/1821 | Current taxonomy valid update / <i>Streptomyces albus</i> 30:371 (AL) |
| <i>Streptomyces aureofaciens</i> | NAD | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [17] | Chemotaxonomic methods, DNA-DNA hybridizations, LFRFA, RAPD-PCR assays, 16S rRNA, 23S rRNA, 5S rRNA genes, RFLPs of rRNA | AY207608 | 16S rRNA gene | Song J, Lee S-C, Kang J-W and Suh J-W. Direct Submission (2002) | https://www.ncbi.nlm.nih.gov/genome/genomes/32330 | Current taxonomy valid update / <i>Streptomyces aureofaciens</i> 30:373 (AL) |
| <i>Streptomyces chrestomyceticus</i> | NAD | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [17] | Chemotaxonomic methods, DNA-DNA hybridizations, LFRFA, RAPD-PCR assays, 16S rRNA, 23S rRNA, 5S rRNA genes, RFLPs of rRNA | AJ621609 | 16S rRNA gene | Swiderski, J. Direct Submission (2004) | https://www.ncbi.nlm.nih.gov/genome/genomes/40695 | Current taxonomy valid update / <i>Streptomyces chrestomyceticus</i> 30:376 (AL) |
| <i>Streptomyces chromofuscus</i> | NAD | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [17] | Chemotaxonomic methods, DNA-DNA hybridizations, LFRFA, RAPD-PCR assays, 16S rRNA, 23S rRNA, 5S rRNA genes, RFLPs of rRNA | AB184194 | 16S rRNA gene | [88] | NAD | Current taxonomy valid update / <i>Streptomyces chromofuscus</i> 30:376 (AL) |
| <i>Streptomyces cinnamonensis</i> | NAD | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [17] | Chemotaxonomic methods, DNA-DNA hybridizations, LFRFA, RAPD-PCR assays, 16S rRNA, 23S rRNA, 5S rRNA genes, RFLPs of rRNA | AB184707 | 16S rRNA gene | [88] | NAD | Current taxonomy valid update / <i>Streptomyces cinnamonensis</i> 30:377 (AL) |
| <i>Streptomyces cinnamoneus</i> | <i>Streptomyces cinnamoneum</i> | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [19] | phenotypes, DNA-DNA hybridization data and partial sequences of gyrB | AB184850 | 16S rRNA gene | [88] | https://www.ncbi.nlm.nih.gov/genome/genomes/50490 | Current taxonomy valid update / <i>Streptomyces cinnamoneus</i> 41:456 |
| <i>Streptomyces lasaliensis</i> | NAD | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [65] | 16S rRNA sequence analyses | HQ537060 | 16S rRNA gene | [68] | NAD | Not validated name |
| <i>Streptomyces lividans</i> | <i>Actinomyces lividans</i> | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [17] | Chemotaxonomic methods, DNA-DNA hybridizations, LFRFA, RAPD-PCR assays, 16S rRNA, 23S rRNA, 5S rRNA genes, RFLPs of rRNA | EU790486.1 | 16S rRNA gene | [89] | https://www.ncbi.nlm.nih.gov/genome/genomes/1888 | Not validated name |
| <i>Streptomyces mobaraensis</i> | <i>Streptoverticillium mobaraense</i> | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [19] | phenotypes, DNA-DNA hybridization data and partial sequences of gyrB | DQ442528 | 16S rRNA gene | [90] | https://www.ncbi.nlm.nih.gov/genome/genomes/14887 | Current taxonomy valid update / <i>Streptomyces mobaraensis</i> 41:456 |
| <i>Streptomyces murinus</i> | NAD | BACTERIA/Actinobacteria/ Actinomycetales/ Streptomycetaceae | [21] | 16S rRNA sequence analyses | AB184155 | 16S rRNA gene | [88] | NAD | Current taxonomy valid update / <i>Streptomyces murinus</i> 30:393 (AL) |

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|--|-------------------------------------|---|---------------------|--|--|--------------------|---------------------------------|---|--|
| <i>Streptomyces netropsis</i> | NAD | BACTERIA/Actinobacteria/Actinomycetales/Streptomycetaceae | [19] | phenotypes, DNA-DNA hybridization data and partial sequences of gyrB | EF178671 | 16S rRNA gene | [90] | NAD | Current taxonomy valid update / <i>Streptomyces netropsis</i> 41:456 |
| <i>Streptomyces olivochromogenes</i> | <i>Actinomyces olivochromogenus</i> | BACTERIA/Actinobacteria/Actinomycetales/Streptomycetaceae | [20] | DNA-DNA hybridization, partial sequences of the gyrB gene, 16S rRNA fragment | AY094370 | 16S rRNA gene | [91] | https://www.ncbi.nlm.nih.gov/genome/genomes/42751 | Current taxonomy valid update / <i>Streptomyces olivochromogenes</i> 30:395 (AL) |
| <i>Streptomyces rubiginosus</i> | <i>Actinomyces rubiginosus</i> | BACTERIA/Actinobacteria/Actinomycetales/Streptomycetaceae | [21] | 16S rRNA sequence analyses | AB122724.1 | 16S rRNA gene | [115] | NAD | Current taxonomy valid update / <i>Streptomyces rubiginosus</i> 30:400 (AL) |
| <i>Streptomyces violaceoruber</i> | <i>Actinomyces violaceus-ruber</i> | BACTERIA/Actinobacteria/Actinomycetales/Streptomycetaceae | [22] | 16S rRNA sequence analyses | AF503492 | 16S rRNA gene | [92] | https://www.ncbi.nlm.nih.gov/genome/genomes/16431 | Current taxonomy valid update / <i>Streptomyces violaceoruber</i> 30:404 (AL) |

Methods Summary highlights molecular methodology that complements basic phenotypic and classical characterisation

NAD: Non-available data/ No relevant data

NAD*: Non-available data/ No Teleomorph synonyms needed

Not validated name: According to International Code of Nomenclature, IJSEM validated list

AL: Included in the Approved Lists (Skerman et. al. (1980), (1989); Euzéby, J.P. (1997))

***: Original publication in IJSB or IJSEM Volume:page**

(≡) homotypic (formerly: objective) synonym; the original name is indicated as a basonym. International Code of Nomenclature of Bacteria (1990 revision); Tindall, B.J. (1999).

(=) heterotypic (formerly: subjective) synonym; the name published first has priority over. International Code of Nomenclature of Bacteria (1990 revision); Tindall, B.J. (1999).

****:** Direct submissions to the GenBank Database. Not cited in articles.

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Appendix J - Microorganism taxonomical identifiers

Table J2. FUNGI

| MICROORGANISMS NAME BY PROCUREMENT/ CURRENT NAME | SYNONYM NAMES | TAXONOMY KINGDOM PHYLUM/ORDER/FAMILY | TAXONOMY REFERENCES | METHODS SUMMARY | HOUSEKEEPING GENES SEQUENCE ID GENBANK | HOUSEKEEPING GENES | REFERENCES HOUSEKEEPING GENES** | GENOME ID | CONCLUSION |
|--|---|--|---------------------|---|--|---|---|---|---|
| <i>Aspergillus aculeatus</i> | <i>Aspergillus japonicus</i> var. <i>aculeatus</i> | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [24; 26] | ITS and 5.8S rRNA gene | EU714394.1 | 18S rRNA; ITS1; 5.8S rRNA gene; ITS2; 28S rRNA gene | [55; 56; 114] | https://www.ncbi.nlm.nih.gov/genome/genomes/12515 | Current/Legitimate name: <i>Aspergillus aculeatus</i> |
| <i>Aspergillus japonicus</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [25; 26] | ITS1-5.8S rRNA gene-18S rRNA gene-ITS2 region, RFLPs of nuclear DNA, and secondary-metabolite profiles | AJ279993 | 18S rRNA gene; 5.8S rRNA gene; ITS1; ITS2 | [55; 56; 25] | NAD | Current/Legitimate name: <i>Aspergillus japonicus</i> |
| <i>Aspergillus melleus</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [26] | nuc rRNA internal transcribed spacer rRNA region (ITS1-5.8S-ITS2), calmodulin (CaM), β -tubulin (BenA), RNA polymerase II second largest subunit (RPB2) | AF203796 | 18S rRNA; ITS1; 5.8S rRNA gene; ITS2; 28S rRNA gene | [73] | NAD | Current/Legitimate name: <i>Aspergillus melleus</i> |
| <i>Aspergillus niger</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [26; 27] | RFLP analysis; 18S rRNA | JN587346 - | 18S rRNA gene - | [72] [54; 55; 56] | https://www.ncbi.nlm.nih.gov/genome/genomes/429 | Current/Legitimate name: <i>Aspergillus niger</i> |
| <i>Aspergillus oryzae</i> | <i>Aspergillus flavus</i> var. <i>oryzae</i> | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [28] | nuc rRNA internal transcribed spacer rRNA region (ITS1-5.8S-ITS2), calmodulin (CaM), β -tubulin (BenA), RNA polymerase II second largest subunit (RPB2) | DQ155287.1 | 18S rRNA; ITS1; 5.8S rRNA gene | [52; 113] | https://www.ncbi.nlm.nih.gov/genome/genomes/526 | Current/Legitimate name: <i>Aspergillus flavus</i> var. <i>oryzae</i> |
| <i>Aspergillus sojae</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [29] | scanning electron micrograph (morphological characteristics) and physiological characteristics | AB008419.1 | 18S rRNA gene | [52] | https://www.ncbi.nlm.nih.gov/genome/genomes/10858 | Current/Legitimate name: <i>Aspergillus sojae</i> |
| <i>Candida cylindracea</i> | NAD* | FUNGI/ Ascomycota/ Saccharomycetales/ Saccharomycetaceae | [30] | SSU rRNA sequence, 18S rRNA sequence, 5S rRNA sequences, CUG codon | X64703.1 X64704.1 X66006.1 X66007.1 X66008.1 | <i>lip1</i> <i>lip2</i> <i>lip3</i> <i>lip4</i> <i>lip5</i> | [71; 116] [71; 116] [71; 116] [71; 116] [71; 116] | NAD | Current/Legitimate name: <i>Candida cylindracea</i> |
| <i>Candida lipolytica</i> | <i>Yarrowia lipolytica</i> , <i>Torula lipolytica</i> , <i>Mycotorula lipolytica</i> , <i>Candida paralipolytica</i> , <i>Candida oleae</i> | FUNGI/ Ascomycota/ Saccharomycetales/ Saccharomycetaceae | [31] | ITS-PCR | AJ250347.1 | <i>act1</i> | [93] | https://www.ncbi.nlm.nih.gov/genome/genomes/194 | Current/Legitimate name: <i>Candida deformans</i> |
| <i>Chaetomium erraticum</i> | <i>Chaetomium gracile</i> , <i>Chaetomium arcuatum</i> <i>Chaetomium virescens</i> var. <i>thielavioideum</i> | FUNGI/Ascomycota/ Sordariales/Chaetomiaceae | [32] | - | - | - | - | NAD | Current/Legitimate name: <i>Chaetomium virescens</i> var. <i>thielavioideum</i> |
| <i>Chaetomium gracile</i> | <i>Chaetomium erraticum</i> , <i>Chaetomium virescens</i> var. <i>Thielavioideum</i> | FUNGI/Ascomycota/ Sordariales/ Chaetomiaceae | [33] | - | JX536280.1 | 18S rRNA; ITS1; 5.8S rRNA gene; ITS2; 28S rRNA gene | [94] | NAD | Current/Legitimate name: <i>Chaetomium virescens</i> var. <i>thielavioideum</i> |

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|---|--|---|---------------------|---|--|---|---------------------------------|---|---|
| <i>Cryphonectria parasitica</i> | <i>Endothia parasitica</i> , <i>Diaporthe parasitica</i> , <i>Endothia gyrosa</i> var. <i>Parasitica</i> , <i>Valsonectria parasitica</i> | FUNGI/Ascomycota/ Diaporthales/Valsaceae | [34] | Ribosomal RNA (ITS1, 5.8S, ITS2) and b-tubulin gene sequencing. Morphological studies | L42441.1 | 18S rRNA gene | [95] | Mitochondrial Plasmid only available: AF218210 | Current/Legitimate name: <i>Cryphonectria parasitica</i> |
| <i>Disporotrichum dimorphosporum</i> | <i>Sporotrichum dimorphosporum</i> | FUNGI/Basidiomycota/ Agaricales/ Incertae sedis | [35] | - | NAD | NAD | NAD | NAD | No Taxonomy data available |
| <i>Fusarium venenatum</i> | NAD* | FUNGI/Ascomycota/ Hypocreales/ Nectriaceae | [36] | RAPDs/PCR using species-specific primers | GQ915554.1 | <i>tri5</i> | [52; 53; 67; 117] | NAD | Current Legitimate name: <i>Fusarium venenatum</i> |
| <i>Hansenula polymorpha</i> | <i>Pichia angusta</i> ; <i>Ogataea polymorpha</i> | FUNGI/Ascomycota/ Saccharomycetales/ Saccharomycetaceae | [37] | 18S rRNA gene/26S rRNA gene | FJ914915 | 18S rRNA gene | [37] | https://www.ncbi.nlm.nih.gov/genome/genomes/14402 | Current/Legitimate name: <i>Ogataea polymorpha</i> |
| <i>Humicola insolens</i> | NAD* | FUNGI/Ascomycota/ Sordariales/Chaetomiaceae | [38] | - | HQ850154.1 | 18S rRNA gene | [81] | NAD | Current/Legitimate name: <i>Humicola insolens</i> |
| <i>Leptographium procerum</i> | NAD* | FUNGI/Ascomycota/ Ophiostomatales/ Ophiostomataceae | [39] | Phylogenetic analyses of seven gene regions (ITS2-LSU, actin, β -tubulin, calmodulin, translation elongation factor 1- α , and the mating type genes <i>MAT1-1-3</i> and <i>MAT1-2-1</i>) | JF440600.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [96] | https://www.ncbi.nlm.nih.gov/genome/genomes/35500 | Current/Legitimate name: <i>Leptographium procerum</i> |
| <i>Mucor javanicus</i> | <i>Mucor circinelloides</i> *heterotypic synonyms | FUNGI/Zygomycota/ Mucorales/Mucoraceae | [40] | LSU, SSU, ITS1, ITS2, 5.8S rRNA, actin (act) and translation elongation factor 1-alpha (tef) | HM234130.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [123] | https://www.ncbi.nlm.nih.gov/genome/genomes/2804 | Current/Legitimate name: <i>Mucor circinelloides</i> f. <i>circinelloides</i> |
| <i>Penicillium camemberti</i> | NAD* | FUNGI/Ascomycota/Eurotiales/ Trichocomaceae | [41] | Macromorphology, Micromorphology, Internal transcribed spacer (ITS), β -tubulin (BenA), calmodulin (CaM) or the RNA polymerase II second largest subunit (RPB2) genes | AB479314 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [64; 70] | https://www.ncbi.nlm.nih.gov/genome/genomes/30945 | Current/Legitimate name: <i>Penicillium camemberti</i> |
| <i>Penicillium chrysogenum</i> | NAD* | FUNGI/Ascomycota/Eurotiales/ Trichocomaceae | [41] | Macromorphology, Micromorphology, Internal transcribed spacer (ITS), β -tubulin (BenA), calmodulin (CaM) or the RNA polymerase II second largest subunit (RPB2) genes | L76153.1 | 18S rRNA gene | [64; 118] | https://www.ncbi.nlm.nih.gov/genome/genomes/10820 | Current/Legitimate name: <i>Penicillium chrysogenum</i> |
| <i>Penicillium citrinum</i> | NAD* | FUNGI/Ascomycota/Eurotiales/ Trichocomaceae | [41; 82] | ITS regions, β -tubulin and calmodulin sequence analysis | LN835269 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [64; 112] | https://www.ncbi.nlm.nih.gov/genome/genomes/40785 | Current/Legitimate name: <i>Penicillium citrinum</i> |

Database on potential toxigenic capacities of microorganisms used for industrial production

| MICROORGANISMS NAME BY PROCUREMENT/ CURRENT NAME | SYNONYM NAMES | TAXONOMY KINGDOM PHYLUM/ORDER/FAMILY | TAXONOMY REFERENCES | METHODS SUMMARY | HOUSEKEEPING GENES SEQUENCE ID GENBANK | HOUSEKEEPING GENES | REFERENCES HOUSEKEEPING GENES** | GENOME ID | CONCLUSION |
|--|--|--|---------------------|---|--|---|---------------------------------|---|---|
| <i>Penicillium decumbens</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichomaceae | [41] | Macromorphology, Micromorphology, Internal transcribed spacer (ITS), β -tubulin (BenA), calmodulin (CaM) or the RNA polymerase II second largest subunit (RPB2) genes | KF284158.1 | 18S rRNA gene | [64; 111] | NAD | Legitimate name: <i>Penicillium decumbens</i> |
| <i>Penicillium funiculosum</i> | <i>Talaromyces funiculosus</i> | FUNGI/Ascomycota/Eurotiales/Trichomaceae | [42] | sequencing, RAPD, AFLP | KC013273.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [64; 110] | NAD | Current/Legitimate name: <i>Penicillium funiculosum</i> |
| <i>Penicillium multicolor</i> | <i>Penicillium implicatum</i> var. <i>aureomarginatum</i> , <i>Penicillium sclerotiorum</i> | FUNGI/Ascomycota/Eurotiales/Trichomaceae | [41] | Macromorphology, Micromorphology, Internal transcribed spacer rRNA area (ITS), β -tubulin (BenA), calmodulin (CaM) or the RNA polymerase II second largest subunit (RPB2) genes | KC790402.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [64; 109] | NAD | No Current/Legitimate name |
| <i>Penicillium lilacinum</i> | <i>Purpureocillium lilacinum</i> ; <i>Paecilomyces lilacinus</i> | FUNGI/Ascomycota/Eurotiales/Trichomaceae | [43] | 18S rRNA gene, internal transcribed spacer (ITS) and partial translation elongation factor 1-a (TEF) sequences | HQ842812 | 18S rRNA gene | [43; 64] | https://www.ncbi.nlm.nih.gov/genome/genomes/42115 | Current/Legitimate name: <i>Purpureocillium lilacinum</i> ; <i>Paecilomyces lilacinus</i> |
| <i>Penicillium roqueforti</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichomaceae | [41] | Macromorphology, Micromorphology, Internal transcribed spacer rDNA area (ITS), β -tubulin (BenA), calmodulin (CaM) or the RNA polymerase II second largest subunit (RPB2) genes | DQ119124.1 | 18S rRNA gene | [64; 108] | https://www.ncbi.nlm.nih.gov/genome/genomes/24568 | Current/Legitimate name: <i>Penicillium roqueforti</i> |
| <i>Penicillium salamii</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichomaceae | [44] | calmodulin, β tubulin and ITS sequences, phenotypic characters and extrolite patterns | | | [64] | NAD | No Current Legitimate name |
| <i>Penicillium nordicum</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichomaceae | [41] | Macromorphology, Micromorphology, Internal transcribed spacer rDNA area (ITS), β -tubulin (BenA), calmodulin (CaM) or the RNA polymerase II second largest subunit (RPB2) genes | KJ834513.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [41; 64] | https://www.ncbi.nlm.nih.gov/genome/genomes/32546 | Current/Legitimate name: <i>Penicillium nordicum</i> |
| <i>Rhizomucor miehei</i> | <i>Mucor miehei</i> | FUNGI/Zygomycota/Mucorales/Mucoraceae | [45] | 18S and 28S rRNA sequences | AJ278360 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [97] | https://www.ncbi.nlm.nih.gov/genome/genomes/11518 | Current/Legitimate name: <i>Rhizomucor miehei</i> |

Database on potential toxigenic capacities of microorganisms used for industrial production

| MICROORGANISMS NAME BY PROCUREMENT / CURRENT NAME | SYNONYM NAMES | TAXONOMY KINGDOM PHYLUM/ORDER/FAMILY | TAXONOMY REFERENCES | METHODS SUMMARY | HOUSEKEEPING GENES SEQUENCE ID GENBANK | HOUSEKEEPING GENES | REFERENCES HOUSEKEEPING GENES** | GENOME ID | CONCLUSION |
|---|--|--|---------------------|---|--|---|--|---|--|
| <i>Rhizopus niveus</i> | NAD* | FUNGI/Zygomycota/Mucorales/Mucoraceae | [46] | rRNA gene (ITS), actin, translation elongation factor 1 (EF-1) | DQ641284.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2; 28S rRNA | [107] | NAD | Current/Legitimate name: <i>Rhizopus niveus</i> |
| <i>Rhizopus oryzae</i> | NAD* | FUNGI/Zygomycota/Mucorales/Mucoraceae | [46] | 18S rRNA nucleotide sequences | AF113440.1 | 18S rRNA gene | [106] | https://www.ncbi.nlm.nih.gov/genome/genomes/322 | Current/Legitimate name: <i>Rhizopus oryzae</i> |
| <i>Sporobolomyces singularis</i> | <i>Bullera singularis</i> , <i>Hamamotoa singularis</i> | FUNGI/Basidiomycota/Sporidiobolales/Sporidiobolaceae | [47] | 18S rRNA nucleotide sequences | AB021690 | 18S rRNA gene | [47] | NAD | Current/Legitimate name: <i>Hamamotoa singularis</i> ' (MB 813188) |
| <i>Talaromyces emersonii</i> | <i>Rasamsonia emersonii</i> | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [49; 99] | Morphological analysis, TS, BenA and RPB2 multigene phylogeny; phenotypic characters, extrolite patterns, ITS and partial calmodulin and β -tubulin sequences | D88321.2 | 18S rRNA gene | [104] | https://www.ncbi.nlm.nih.gov/genome/genomes/36802 | Current/Legitimate name: <i>Rasamsonia emersonii</i> |
| <i>Talaromyces cellulolyticus</i> | <i>Acremonium cellulolyticus</i> | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [49; 84] | Morphological analysis, TS, BenA and RPB2 multigene phylogeny | KF811039.1 | 18S rRNA; ITS1; 5.8S rRNA gene; ITS2; 28S rRNA gene | [105] | https://www.ncbi.nlm.nih.gov/genome/genomes/35980 | Current/Legitimate name: <i>Talaromyces cellulolyticus</i> |
| <i>Talaromyces versatilis</i> | NAD* | FUNGI/Ascomycota/Eurotiales/Trichocomaceae | [49] | Morphological analysis, TS, BenA and RPB2 multigene phylogeny | KC962113.1 | 18S rRNA; ITS1; 5.8S rRNA gene; ITS2; 28S rRNA gene | [103] | NAD | Current/Legitimate name: <i>Talaromyces versatilis</i> |
| <i>Trametes hirsuta</i> | NAD* | FUNGI/Basidiomycota/Polyporales/Polyporaceae | [48] | 18S rRNA nucleotide sequences | EU771080.1 | 18S rRNA; ITS1; 5.8S rRNA gene; ITS2; 28S rRNA gene | Deng X, Song R and Sun H. Direct Submission (2008) | https://www.ncbi.nlm.nih.gov/genome/genomes/40253 | Current/Legitimate name: <i>Trametes hirsuta</i> |
| <i>Trametes versicolor</i> | NAD* | FUNGI/Basidiomycota/Polyporales/ Polyporaceae | [48] | ITS and mt SSU rDNA | KF638522.1 | rRNA genes, ITS1 and ITS2 DNA | [102] | https://www.ncbi.nlm.nih.gov/genome/genomes/3260 | Current/Legitimate name: <i>Trametes versicolor</i> |
| <i>Trichoderma citrinoviride</i> | NAD* | FUNGI/Ascomycota/Hypocreales/Hypocreaceae | [50] | rRNA ITS, actin, EF-1 α | JF745094.1 | 18S rRNA; ITS1; 5.8S rRNA gene; ITS2; 28S rRNA gene | [101] | NAD | Current/Legitimate name: <i>Trichoderma citrinoviride</i> |
| <i>Trichoderma harzianum</i> | NAD* | FUNGI/Ascomycota/Hypocreales/Hypocreaceae | [51] | rRNA ITS, actin, EF-1 α | AF508862.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2 | Kim S-J. Direct Submission (2002) | https://www.ncbi.nlm.nih.gov/genome/genomes/2441 | Current/Legitimate name: <i>Trichoderma harzianum</i> |
| <i>Trichoderma koningii</i> | NAD* | FUNGI/Ascomycota/Hypocreales/Hypocreaceae | [51] | rRNA ITS, actin, EF-1 α | KU314996.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2 | Garzoli L and Varese GC. Direct Submission (2015) | https://www.ncbi.nlm.nih.gov/genome/genomes/51821 | Current/Legitimate name: <i>Trichoderma koningii</i> |
| <i>Trichoderma longibrachiatum</i> | NAD* | FUNGI/Ascomycota/Hypocreales/Hypocreaceae | [51] | rRNA ITS, actin, EF-1 α | Z31019.1 | rRNA genes, ITS1 and ITS2 DNA | [98] | https://www.ncbi.nlm.nih.gov/genome/genomes/18220 | Current/Legitimate name: <i>Trichoderma longibrachiatum</i> |
| <i>Trichoderma reesei</i> | NAD* | FUNGI/Ascomycota/Hypocreales/Hypocreaceae | [50] | rRNA ITS, actin, EF-1 α | JX841312.1 | <i>lae1</i> | [66] | https://www.ncbi.nlm.nih.gov/genome/genomes/323 | Current/Legitimate name: <i>Trichoderma reesei</i> |
| <i>Trichoderma viride</i> | NAD* | FUNGI/Ascomycota/Hypocreales/Hypocreaceae | [51] | rRNA ITS, actin, EF-1 α | KF765423.1 | 18S rRNA; ITS1; 5.8S rRNA; ITS2 | [100] | NAD | Current/Legitimate name: <i>Trichoderma viride</i> |

Methods Summary highlights molecular methodology that complements basic phenotypic and classical characterisation

NAD: Non-available data/ No relevant data

NAD*: Non-available data/ No Teleomorph synonyms needed

Not validated name: According to International Code of Nomenclature, IJSEM validated list

AL: Included in the Approved Lists (Skerman et. al. (1980), (1989); Euzéby, J.P. (1997))

***: Original publication in IJSB or IJSEM Volume:page**

(=) homotypic (formerly: objective) synonym; the original name is indicated as a basonym. International Code of Nomenclature of Bacteria (1990 revision); Tindall, B.J. (1999).

(=) heterotypic (formerly:subjective) synonym; the name published first has priority over. International Code of Nomenclature of Bacteria (1990 revision); Tindall, B.J. (1999).

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