

Exploration of the molecular linkage between endometriosis and Crohn disease by bioinformatics methods

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Abstract

Background: Endometriosis (EMT) is a common disease in reproductive-age woman and Crohn disease (CD) is a chronic inflammatory disorder in gastrointestinal tract. Previous studies reported that patients with EMT had an increased risk of CD. However, the linkage between EMT and CD remains unclear. In this study, we aimed to investigate the potential molecular mechanism of EMT and CD.

Methods: The microarray data of EMT and CD were downloaded from Gene Expression Omnibus. Common genes of EMT and CD were obtained to perform the Gene Ontology and Kyoto Encyclopedia of Gene Genomes enrichments. The protein-protein interaction network was constructed by Cytoscape software and the hub genes were identified by CytoHubba plug-in. Finally we predicted the transcription factors (TFs) of hub genes and constructed a TFs-hub genes regulation network.

Results: A total of 50 common genes were identified. Kyoto Encyclopedia of Gene Genomes enrichment showed that the common genes mainly enriched in MAPK pathway, VEGF pathway, Wnt pathway, TGF-beta pathway, and Ras pathway. Fifteen hub genes were collected from the protein-protein interaction network, including FMOD, FRZB, CPE, SST, ISG15, EFEMP1, KDR, ADRA2A, FZD7, AQP1, IGFBP5, NAMPT, PLUA, FGF9, and FHL2. Among them, FGF9, FZD7, IGFBP5, KDR, and NAMPT were both validated in the other 2 datasets. Finally TFs-hub genes regulation network were constructed.

Conclusion: Our findings firstly revealed the linkage between EMT and CD, including inflammation, angiogenesis, immune regulation, and cell behaviors, which may lead to the risk of CD in EMT. FGF9, FZD7, IGFBP5, KDR, and NAMPT may closely relate to the linkage.

Abbreviations: BP = biological process, CC = cellular component, CD = Crohn disease, DEGs = differential expression genes, EMT = endometriosis, GEO = Gene Expression Omnibus, GO = Gene Ontology, IBD = inflammatory bowel disease, KEGG = Kyoto Encyclopedia of Gene Genomes, MF = molecular function, PPI = protein-protein interaction, ROS = reactive oxygen species, TFs = transcription factors, TRRUST = Transcriptional Regulatory Relationships Unraveled by Sentence-based Text mining.

Keywords: bioinformatics analysis, Crohn disease, endometriosis, hub genes, shared pathogenesis, TFs-gene network

1. Introduction

Crohn disease (CD), a type of inflammatory bowel disease (IBD), is defined as a chronic inflammatory disease of gastrointestinal tract. The prevalence of CD has been increasing since the 20th century.^[1] A systematic review reported that the prevalence rate in Europe ranged from 137.17 to 322 per 100,000, while in Asia, it varied from 1.2 to 53.1 per 100,000.^[2] The cause and pathophysiology of CD are complex. Now it is believed to be the disturbed innate and adaptive immune response caused by multiple elements, including genetic susceptibility, environmental

factors, and intestinal microflora.^[3] Clinical presentations depend on the disease location and severity of inflammation. Most of CD patients experience intermittent abdominal pain, chronic diarrhea, and weight loss, profoundly impacting their quality of life.^[4]

Endometriosis (EMT) is a disease that endometrium-like tissue appears outside the uterus.^[5] EMT affects 10% of reproductive-age woman, with approximately 200 million women worldwide potentially suffering from this disease.^[6] EMT patients typically present chronic pain continually or

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All data generated or analyzed during this study are included in this published article [and its supplementary information files].

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intermittently until the menopause, dysmenorrhea, infertility, and abnormal menstruation. However, the etiology of EMT is largely unknown. Retrograde menstruation is the widely accepted theory, but increasing studies believe EMT is related with abnormal autoimmunity.^[7] Immune escape and abnormal expression of immune cells have been identified.^[8] Several studies have revealed that EMT is not solely linked to malignant tumors such as ovarian cancer, non-Hodgkin lymphoma, and breast cancer, but also some immune diseases like systemic lupus, rheumatoid arthritis, and IBD.^[9,10] A nationwide cohort in Danish reported that patients with EMT had an elevated risk of developing IBD. The standardized incidence ratio of ulcerative colitis was 1.5 and that of CD was 1.6.^[11] Although EMT patients have been observed to be more susceptible to CD, our understanding of the mechanism is very limited.

With the quick development of gene microarray technology, this method is widely applied in the genomics research in various diseases, which enables researchers to comprehend gene expression and make a further analysis. For instance, Antonio et al^[12] obtained intestinal tissue from IBD patients and utilized a high-throughput approach to identify a novel predictor classifier. Stefania et al^[13] discovered 2 important genes (BMP4 and GREM1) that were specifically deregulated in the ectopic endometrium by genome wide profiling analysis. As a consequence, we can investigate the relationship between diseases by integrating and analyzing the biological data.

Therefore, we used original microarray datasets from Gene Expression Omnibus (GEO) in order to reveal the molecular mechanism of CD and EMT. Common genes of CD and EMT were obtained, and analyzed by enrichment analysis and PPI network. Moreover, Transcription factors (TFs) of hub genes were predicted. The regulatory relationship between hub genes and TFs were displayed.

2. Materials and methods

2.1. Collection of CD and EMT related genes

Key words “inflammatory bowel disease,” “Crohn’s disease,” “CD,” “endometriosis,” and “EMT” were put into GEO database (<https://www.ncbi.nlm.nih.gov/geo/>) to search CD and EMT gene expression profiles. Selected profiles should meet the following criteria. Firstly, the gene expression profiling should include 2 groups. One is case group and the other is control group. Secondly, each group should contain at least 5 samples. Thirdly, the sample for sequencing should be intestinal tissue or endometriosis tissue. According to the criteria, 2 datasets of CD (GSE36807 and GSE59071) and 2 datasets of EMT (GSE25628 and GSE7305) were collected.

2.2. Differential expression genes

GSE36807 and GSE25628 were used for differential expression genes (DEGs) analysis by R soft. Raw data were read and pre-processed by R soft. Then “limma” package was performed for the data analysis. GSE36807 contained 13 CD and 7 control samples. P value < 0.05 and $|\log_2 \text{FC (fold change)}| > 0.58$ were set as the cutoff for DEGs.^[14] GSE25628 contained 16 EMT and 6 control samples. P value < 0.05 and $|\log_2 \text{FC (fold change)}| > 1$ were set as the cutoff for DEGs.^[15] Subsequently, we visualized the DEGs by a heatmap and a volcano map, respectively. Finally we overlapped the DEGs of CD and EMT in order to obtain common genes. A Venn diagram was constructed by R package “VennDiagram” to visualize the common genes.

2.3. GO and KEGG enrichment analysis of common genes

Gene Ontology (GO) function enrichment consisted of biological process (BP), molecular function (MF), and cellular component

(CC). Kyoto Encyclopedia of Gene Genomes (KEGG) was applied to explore metabolic pathways. GO and KEGG enrichment analysis were performed by R soft. R packages included “org.Hs.eg.db,” “clusterProfiler,” “enrichplot,” and “ggplot2.” P value < 0.05 was recognized as significant enrichment.^[16] The first 10 enrichment results were displayed by a bubble diagram or a bar chart.

2.4. Construction of the protein-protein interaction network

In order to explore the relationship between common genes, we searched these genes in the online website STRING (<https://cn.string-db.org>). All the common genes were put into STRING and we got the protein-protein interaction (PPI) network. The protein interaction data was analyzed by cytoscape_3.9.1 software. According to plug-in “Cytohubba,” we figured out hub genes.

2.5. GO and KEGG enrichment of hub genes

GO function and KEGG pathway enrichment were analyzed by R soft. P value < 0.05 was considered as significant enrichment.^[16] Subsequently we displayed the enrichment results by a bubble diagram or a bar chart.

2.6. Validation of hub genes expression

To enhance accuracy, the hub genes obtained from the PPI network were validated with other datasets. The expression of these hub genes was validated in GSE59071 for CD and GSE7305 for EMT. GSE59071 contained 8 CD and 11 control samples. GSE7305 contained 10 EMT and 10 control samples. The expression of hub genes was visualized by a violin chart.

2.7. Prediction of transcription factors

Transcriptional Regulatory Relationships Unraveled by Sentence-based Text mining (TRRUST) is a database that focuses on human and mouse transcriptional regulatory networks. We upload the hub genes to TRRUST and enrich the TFs. Subsequently, cytoscape_3.9.1 software was applied to construct the TFs-hub genes network. The flow chart is shown in Figure 1.

2.8. Ethical statement

This study does not involve in any patient’s personal data, cell, or animal experiments, and does not require any ethical approval.

3. Results

3.1. DEGs of CD and EMT

According to the inclusion criteria, GSE36807 dataset and GSE25628 dataset were selected for the DEGs analysis. Based on the cutoff criteria, there were 434 DEGs of CD, including 202 up-regulated genes and 232 down-regulated genes. There were 1782 DEGs of EMT, including 654 up-regulated genes and 1128 down-regulated genes. A volcano plot and a heatmap were constructed by R soft to visualize the distribution of DEGs (Fig. 2). Subsequently, we obtained 50 common genes by matching the DEGs of CD and EMT (Fig. 3, Supplement S1, <http://links.lww.com/MD/M497>).

3.2. GO and KEGG enrichment analysis of common genes

We utilized R soft to perform the GO function and KEGG enrichment to gain a deeper understanding of the common

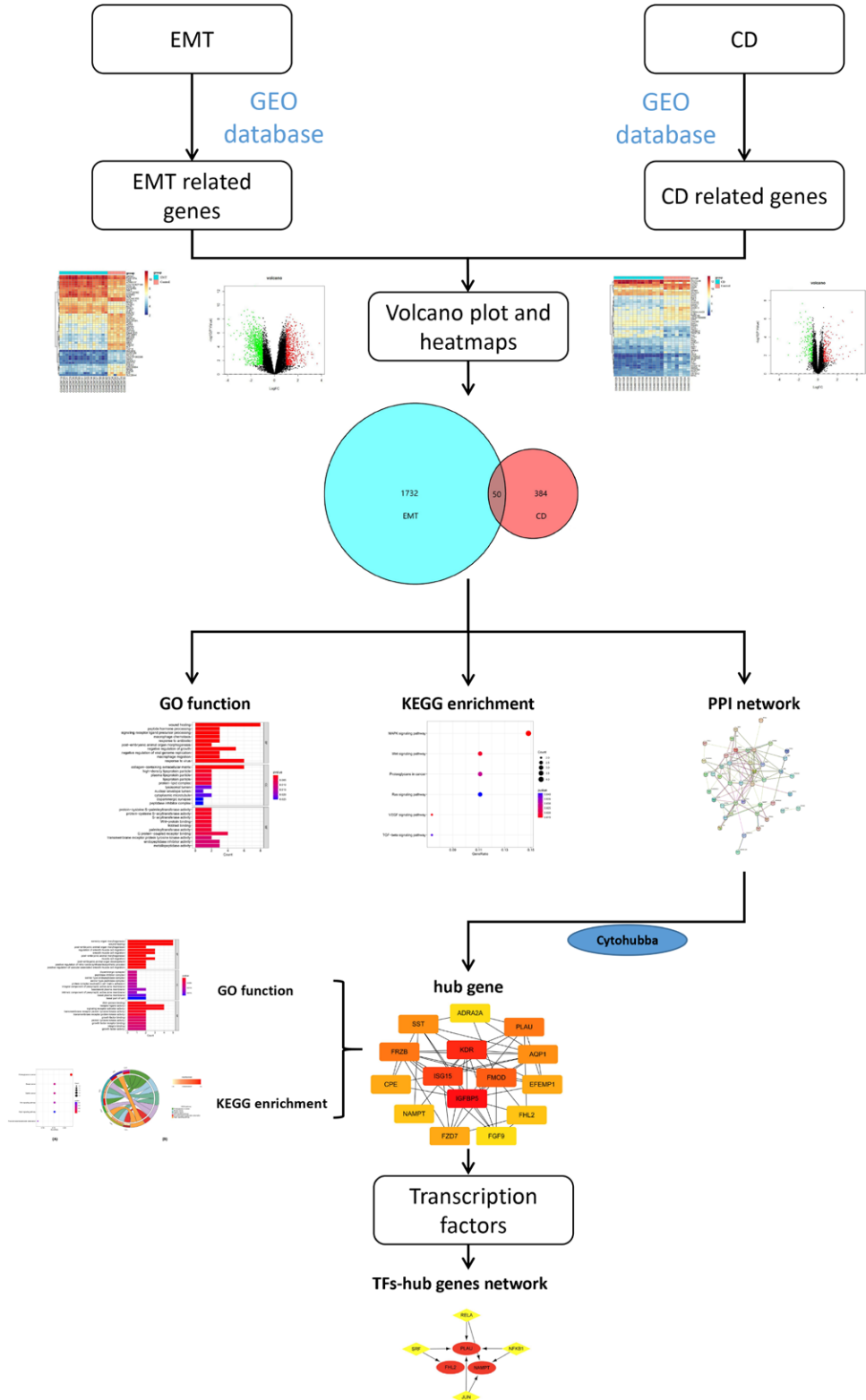


Figure 1. The flow chart of this study.

genes. The top 10 enriched GO terms are displayed in Figure 4. In terms of BP, the results chiefly enriched in wound healing, peptide hormone processing, signaling receptor ligand precursor processing, macrophage chemotaxis, and response to antibiotic. In terms of CC, these genes mainly associated with collagen-containing extracellular matrix, high-density lipoprotein particle, plasma lipoprotein particle, lipoprotein particle, and

protein-lipid complex. In terms of MF, these genes principally related with protein-cysteine S-palmitoyltransferase activity, protein-cysteine, S-acyltransferase activity, Wnt-protein binding, and frizzled binding. KEGG analysis enriched 5 pathways, including MAPK signaling pathway, VEGF signaling pathway, Wnt signaling pathway, TGF-beta signaling pathway, and Ras signaling pathway (Fig. 5).

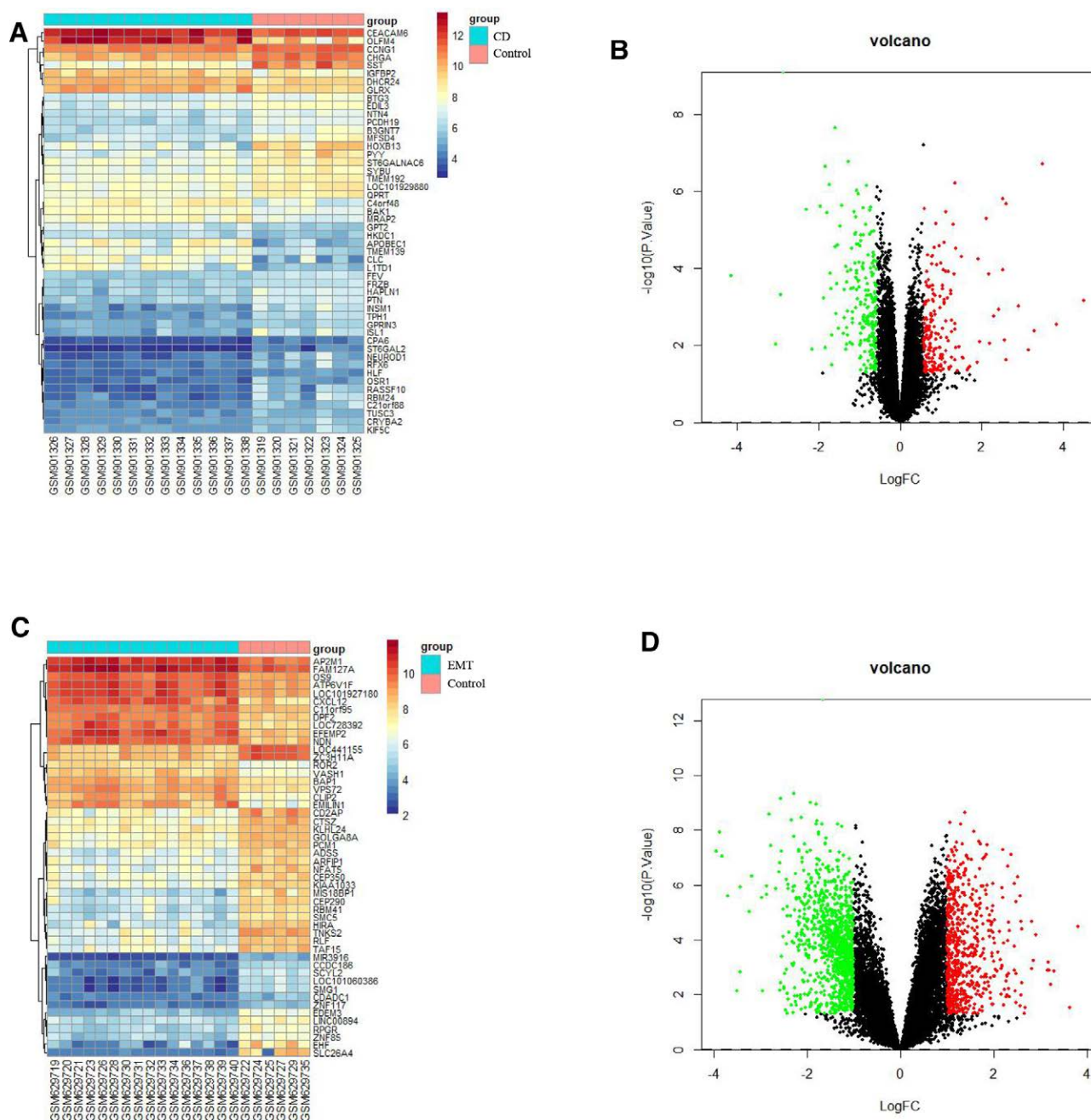


Figure 2. The heatmap and volcano plot of DEGs. Panel (A) heatmap of DEGs in GSE36807; Panel (B) volcano plot of DEGs in GSE36807; Panel (C) heatmap of DEGs in GSE25628; Panel (D) volcano plot of DEGs in GSE25628. Red dots: significant up-regulation; Green dots: significant down-regulation; Black dots: no significance. CD = Crohn disease; DEGs = differential expression genes; EMT = endometriosis.

3.3. Construction of PPI network and analysis

All the common genes were put into the STRING database and we got a PPI network, including 49 nodes and 109 edges (Fig. 6). Subsequently, the PPI network was analyzed by CytoHubba plug-in to figure out the hub genes. Fifteen genes were collected, including IGFBP5, ISG15, KDR, FMOD, FRZB, SST, ADRA2A, PLAU, AQP1, EFEMP1, FHL2, FGF9, FZD7, NAMPT, and CPE (Fig. 7).

GO analysis and KEGG enrichment were performed for the hub genes. In terms of the BP, the results mainly related with sensory organ morphogenesis, wound healing, postembryonic animal organ morphogenesis, regulation of smooth muscle cell migration and smooth muscle cell migration. In terms of CC, the results were associated with dopaminergic synapse, peptidase inhibitor complex, serine-type endopeptidase complex, serine-type

peptidase complex, and protein complex involved in cell-matrix adhesion. In terms of MF, these genes enriched in Wnt-protein binding, receptor ligand activity, signaling receptor activator activity, transmembrane receptor protein tyrosine kinase activity, and transmembrane receptor protein kinase activity (Fig. 8). From the results of KEGG analysis, hub genes chiefly related with Wnt signaling pathway and Rap1 signaling pathway (Fig. 9A). Furthermore, we displayed the relationship between hub genes and pathways (Fig. 9B). According to the results, KDR, FGF9, FZD7, and FRZB closely related with these pathways.

3.4. Validation of hub genes expression

Validation of hub genes was performed in GSE59071 for CD and GSE7305 for EMT. As for CD, the results showed

4 up-regulated (IGFBP5, KDR, NAMPT, and PLUA) and 3 down-regulated (FGF9, FZD7, and SST) genes (Fig. 10). As for EMT, The results showed 11 up-regulated (AQP1, CPE, EFEMP1, FHL2, FMOD, FRZB, FZD7, IGFBP5, ISG15, KDR, and NAMPT) and 1 down-regulated (FGF9) genes (Fig. 11). IGFBP5, FGF9, FZD7, KDR, and NAMPT exhibited differential expression in GSE59071 and GSE7305. Moreover, the expression of these 5 genes closely aligned with that of GSE36807 and GSE25628. We hypothesized that these genes may play an important role in the linkage between EMT and CD.

3.5. Prediction of TFs and analysis

The TFs of hub genes were predicted by online website TRRUST version 2. All the hub genes were uploaded to the website and 4 TFs were obtained, including serum response factor (SRF), jun

proto-oncogene (JUN), v-rel reticuloendotheliosis viral onco-gene homolog A (RELA), and nuclear factor of kappa light poly-peptide gene enhancer in B-cells 1 (NFKB1). According to the regulation of TFs and hub genes, TFs-hub genes network was constructed (Fig. 12). We found that every TF regulated 2 genes.

4. Discussion

EMT is a common disease in childbearing age women. CD has a peak onset between 20 and 40 years old, followed by a small peak from 50 to 60 years old.^[17] The epidemiology indicates that these 2 diseases share a similar onset age. There are more and more studies showing the correlation between EMT and CD. Previously a few cases reported the co-existence of EMT and CD.^[18,19] Nowadays a cohort study has found the increased incidence of CD in patients with EMT, and a meta-analysis has further corroborated this observation.^[11,20] However, the mechanism of the linkage between EMT and CD remains largely unknown. As a consequence, it is crucial to figure out the intricate relationship between EMT and CD in order to achieve early detection and diagnosis.

Our study is the first to explore the relationship between CD and EMT at genetic level. In this study, we downloaded 2 dependent gene expression data of CD and EMT from GEO. Fifty common genes were obtained from matching DEGs of CD and EMT. The KEGG pathways analysis showed that the common genes mainly enriched in the MAPK signaling pathway, Wnt signaling pathway, VEGF signaling pathway, TGF-beta signaling pathway, and Ras signaling pathway. These pathways were chiefly related to inflammation, cell behavior, angiogenesis, and immunoregulation.

The MAPK's family members consisted of extracellular signal-regulated kinase (ERK), c-Jun N-terminal Kinase (JNK), and p38 mitogen-activated protein kinase (p38MAPK). MAPK signaling pathway involved in a wide variety of cellular activities, including inflammation, cell proliferation, differentiation, migration, senescence, and apoptosis.^[21,22] EMT was regarded as a chronic inflammatory condition. A study showed that

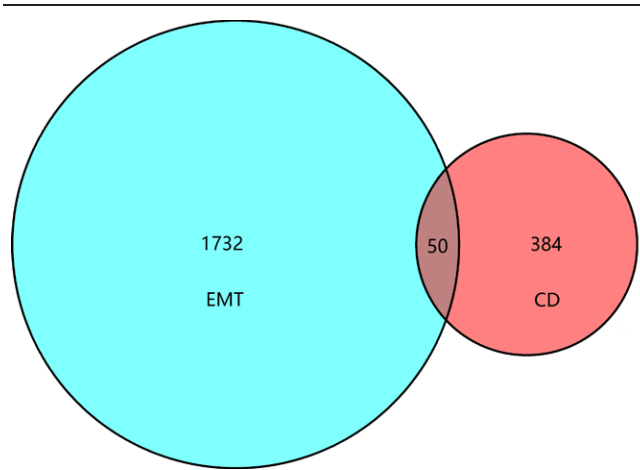


Figure 3. Venn diagram of DEGs in EMT and CD. CD = Crohn disease; DEGs = differentially expression genes; EMT = endometriosis.

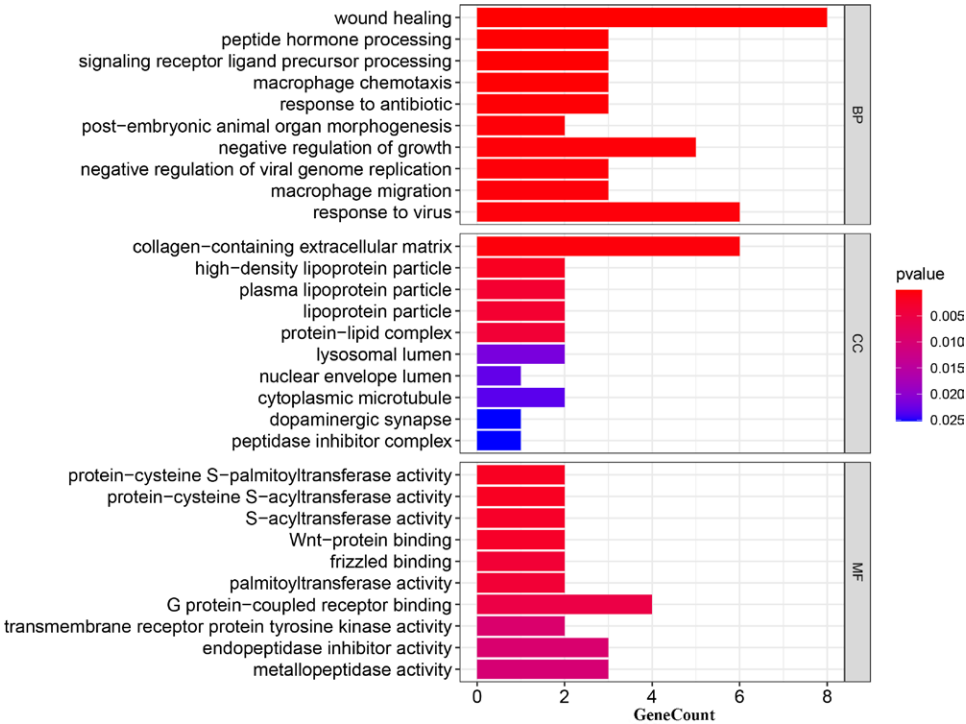


Figure 4. GO enrichment of DEGs. DEGs = differentially expression genes; GO = Gene Ontology.

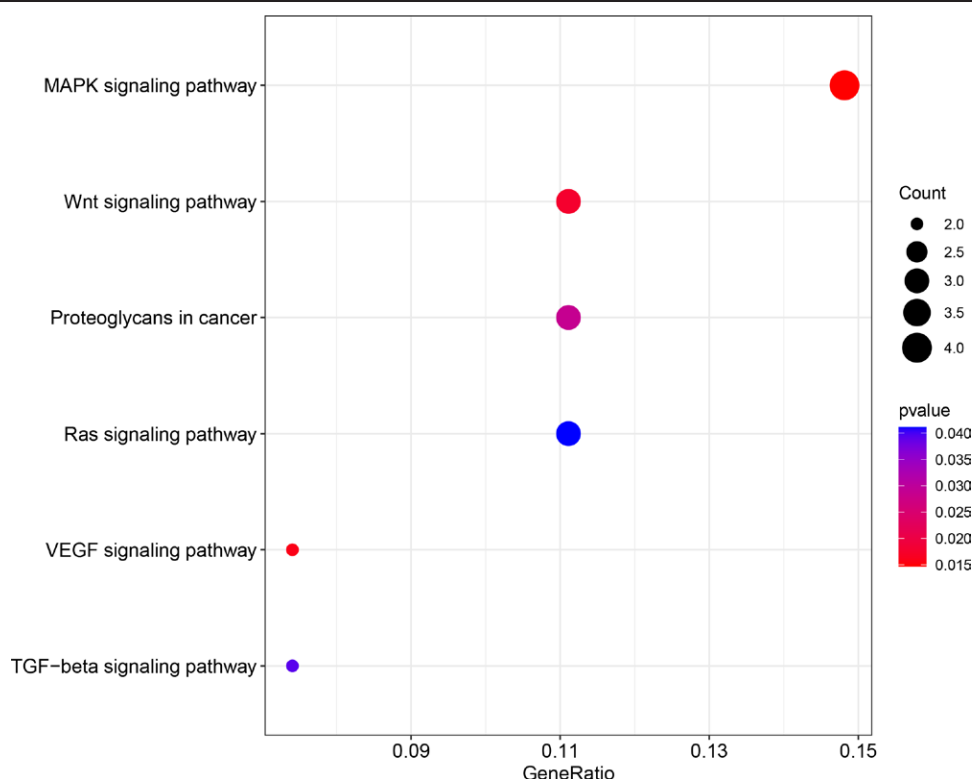


Figure 5. KEGG enrichment of DEGs. DEGs = differential expression genes; KEGG enrichment of hub genes.

estrogen-induced reactive oxygen species and then activated MAPK signaling pathways, which played a crucial role in the inflammation.^[23] Moreover, excessive production of estrogen in EMT profoundly disturbed the MAPK pathway, resulting in the stimulation of ectopic endometriotic cell proliferation.^[23] Activation of MAPK signaling pathway was also confirmed in CD patients, which triggered intestinal inflammation by up-regulating pro-inflammation cytokines.^[24]

Wnt signaling pathway consisted of ligand protein Wnt and membrane protein receptors, which involved in cell specification, stem cell self-renewal and tissue patterning.^[25] Studies showed that high expression of this pathway was observed in the tissue of CD or EMT. Abnormal cell behaviors like cell migration, invasion, and matrix metalloproteinase expression induced by Wnt signaling pathway may be the prerequisites to develop and maintained EMT.^[26] In the intestine of IBD, Wnt pathway was found to be the regulator of inflammation and was considered as a feature of chronic IBD.^[27] Moreover, continuous activation of Wnt pathway increased collagen-I expression in myofibroblasts and involved in intestinal fibrosis.^[28]

Ras was an important regulator of cell growth.^[29] However, over activation of Ras signaling pathway increased cell proliferation and migration in EMT.^[30] Similarly, a study found that Ras highly expressed in CD and contributed to intestinal fibrosis. Activated Ras pathway promoted extracellular matrix and myofibroblast formation in intestine.^[31]

VEGF signaling pathway played a leading role in angiogenesis, contributing to the progression of diseases and inflammation.^[32,33] Angiogenesis promoted ectopic endometrium cell proliferation, migration, and invasion.^[34] Among many pro-angiogenic factors, VEGF played an important role in the pathogenesis. Studies found that high expression of VEGF promoted the development of EMT.^[35,36] VEGF was also highly expressed in IBD.^[37] Angiogenesis significantly contributed to the recruitment of pro-inflammatory cells and led to the disruption of epithelial integrity, ultimately impacting the progression and severity of IBD.^[38]

TGF-beta was regarded as one of the most important cytokines in tissue. The TGF-beta signaling pathway played a critical role in cell development and homeostasis.^[39] However, higher expression of TGF-beta was found in endometriotic tissue. Abnormal expression of TGF-beta enhanced the migration, adhesion to mesothelium and invasive ability of ectopic endometrial cells.^[40,41] Moreover, TGF-beta regulated multiple immune activities to accelerate the progression of EMT.^[42-44] Overexpression of TGF-beta1 protected ectopic tissue survival from immune clearance via reducing natural killer cell and macrophage numbers.^[45] Similarly, dysregulated TGF-beta signaling in IBD led to systemic autoimmunity, abnormal intestinal microbial immune regulation, and barrier function impairment.^[44]

After conducting aforementioned analysis, we discovered several shared signaling pathways that involved in the progression of EMT and CD, thereby furnishing a pathophysiologic groundwork for the risk of CD in patients with EMT.

Subsequently, we obtained 15 hub genes in PPI network by cytoHubba plug-in. These hub genes mainly enriched in Wnt signaling pathway and Rap1 signaling pathway. Rap1 belonged to Ras family. These results were consistent with the KEGG enrichment of common genes. Furthermore, these hub genes were validated in other datasets and 5 genes exhibited differential expression in EMT and CD. These genes were FGF9, FZD7, IGFBP5, KDR, and NAMPT, which may closely relate to the common pathology of EMT and CD.

FGF9 belonged to the human fibroblast growth factor (FGF) and was a kind of secreted protein regulating embryonic development, cell proliferation, cell differentiation, and cell migration.^[46,47] FGF9 was required for many organs development, like lung, heart, and digestive system.^[46] However, FGF9 mutation was reported to be associated with colorectal, endometrial, and ovarian cancer, which indicated the importance of this gene in the intestinal and endometrial disease development.^[48] High expression of FGF9, especially in the early stage, promoted endometriotic stromal cell proliferation and maintain endometriosis.^[49,50] As for CD, there had been no research exploring the

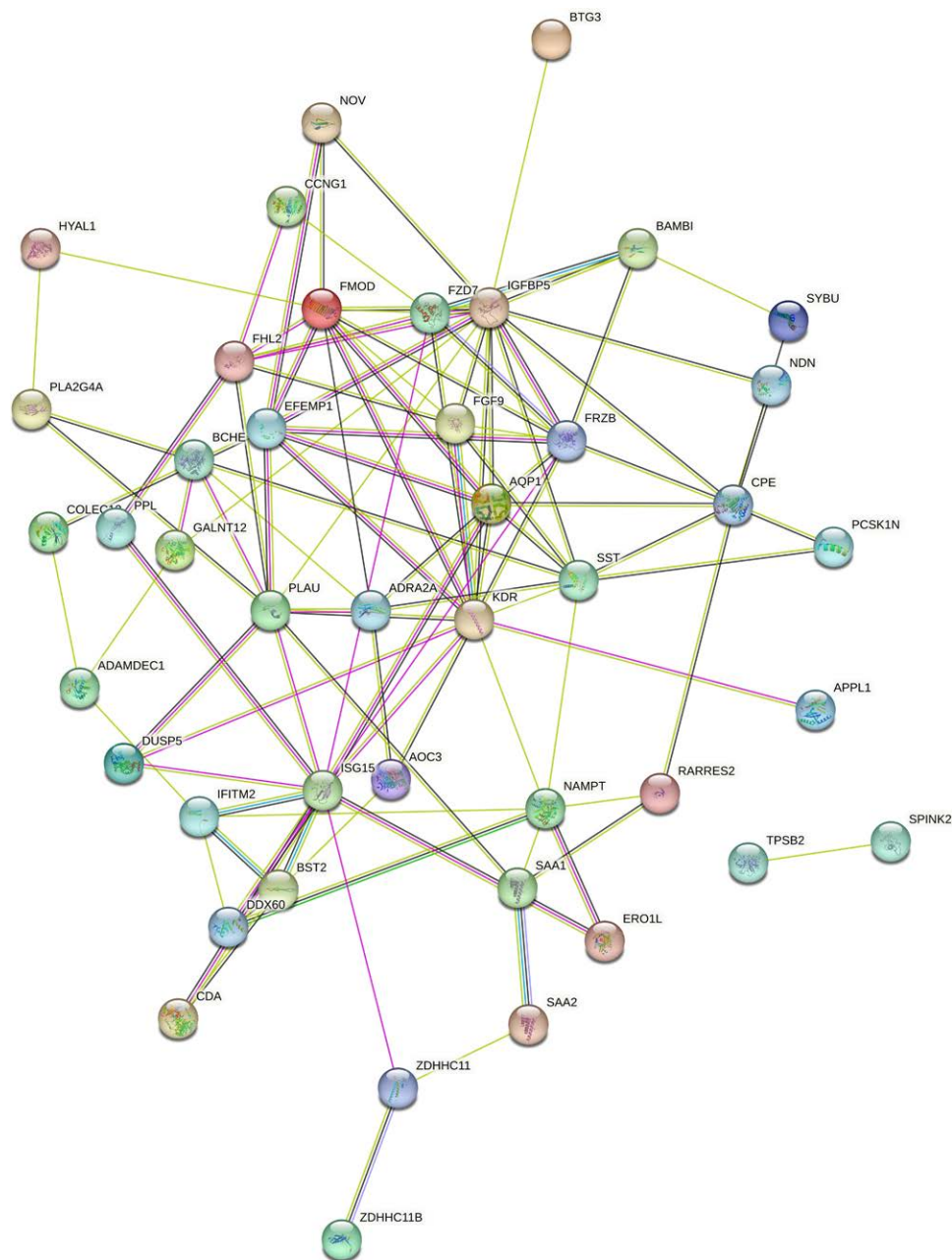


Figure 6. PPI network of DEGs. DEGs = differential expression genes; PPI = protein-protein interaction.

relationship between FGF9 and CD. However, a study found FGF9 regulated the small intestinal mesenchymal fibroblast proliferation and differentiation during the late stage embryogenesis.^[51] We suspected that FGF9 may promote intestinal fibrosis in CD by accelerating mesenchymal fibroblast proliferation. Therefore, we believed that abnormal FGF9 expression may promote tissue proliferation to maintain EMT or CD.

FZD7 was a receptor of Wnt protein and was one of the Frizzled family members.^[52] A study found that up-regulation of FZD7 could promote cell proliferation, migration, and invasion in ectopic human endometrium by activating Wnt signaling pathway.^[53] Similarly, our study showed that FZD7 was increased in EMT tissue. As for CD, FZD7 was decreased in our research. A study found that loss of FZD7 prevented intestinal stem cells from regenerating.^[54] In addition, down-regulation of FZD7 in intestine interrupted differentiation procession of intestinal stem cells and thus the homeostasis of the intestinal epithelium.^[55] Although no study has explored the role of FZD7

in CD, we hypothesized that it caused the disease by affecting the biological processes of intestinal stem cell.

IGFBP5 belonged to the insulin-like growth factor binding proteins (IGFBPs) and was a kind of secreted protein released by a variety of different cells.^[56] IGFBPs were associated with autoimmune diseases via its immunoregulation.^[57] Peripheral blood mononuclear cells, including lymphocytes and monocytes, formed the major innate and adaptive immune systems. In a vivo study, IGFBP5 could recruit mononuclear cell infiltration and accelerate peripheral blood mononuclear cells migration.^[58] Moreover, IGFBP5 induced the preferential migration of CD4 + T cells, which involved in the early phase of inflammation and fibrosis in a disease development.^[59] Studies found that IGFBP5 was highly expressed in inflamed and fibrotic intestine tissue in CD patients. IGFBP5 stimulated smooth muscle, fibroblasts, and myofibroblasts to increase collagen synthesis and cell proliferation.^[60] In addition, IGFBP5 also regulated smooth muscle proliferation via alpha-subunit Gi 3.^[61] Besides, IGFBP5

was found to dominate in the proliferation phase of endometrium.^[62] Nowadays, increasing evidence proved that EMT was

an autoimmune disease.^[63] We suspected IGFBP5 may affect the cell cycle of immune cells to trigger EMT.

KDR, a receptor tyrosine kinase gene, encoded a receptor vascular endothelial cell growth factor (VEGF).^[64] VEGF played an important role in angiogenesis, vascular development, vascular permeability, and embryonic hematopoiesis.^[65] As mentioned before, angiogenesis contributed to the development of EMT and CD.

NAMPT was recognized as a regulator of the intracellular nicotinamide adenine dinucleotide pool, in which NAMPT catalyzed rate-limiting step in the biosynthesis of nicotinamide adenine dinucleotide from nicotinamide (NAM).^[66] NAMPT was elevated in inflammatory disorders and represented damage-associated molecular patterns-like actions.^[67] A study showed that elevation of NAMPT reflected the activation and severity of CD.^[68] In contrast, NAMPT exerted different effect on endometrial cells. NAMPT regulated by estrogen and progesterone in uterus, elevated cellular proliferation by increasing antioxidant enzymes.^[69] This suggested that increased NAMPT may prompt ectopic endometrial cells survival.

TFs are the DNA-proteins that recognize specific DNA sequence and regulate the transcription of chromatin. They form a complex system that governs the genome expression.^[70] As a consequence, we predicted the TFs of hub genes, including JUN, NFKB1, RELA, and SRF, and established the TFs-genes

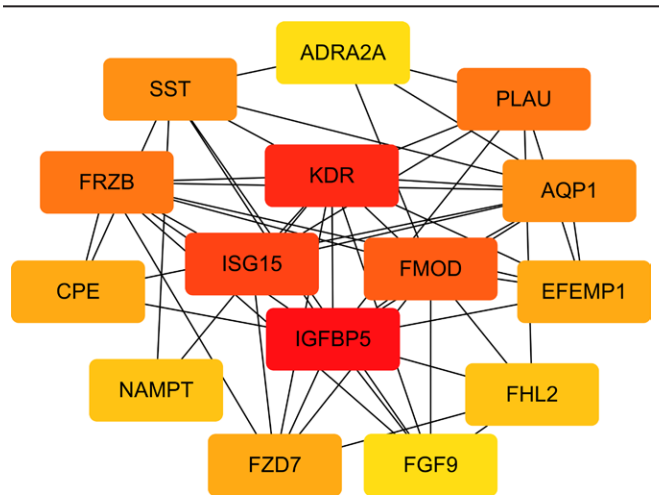


Figure 7. Top 15 hub genes of PPI network analyzed by cytoHubba. PPI = protein-protein interaction.

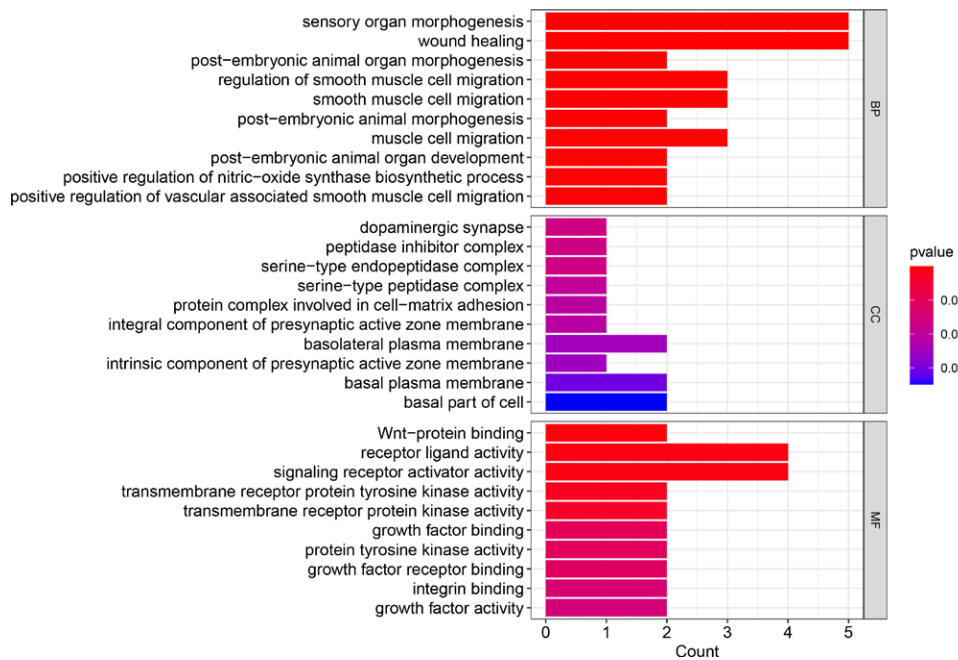


Figure 8. GO enrichment of 15 hub genes. GO = Gene Ontology.

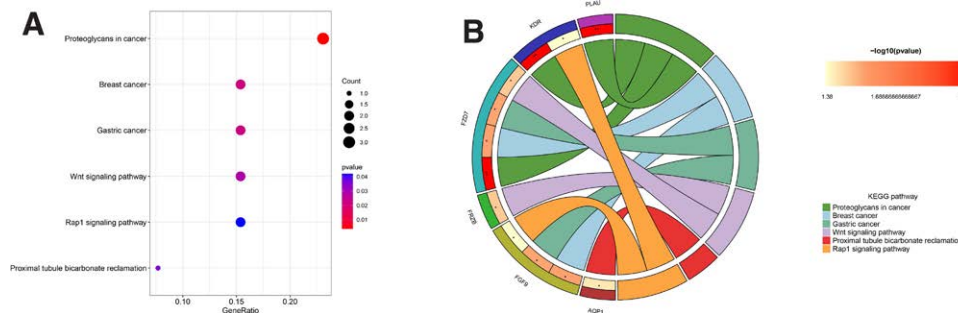


Figure 9. KEGG enrichment of 15 hub genes. Panel (A) KEGG enrichment of hub genes; Panel (B) the relationship of hub genes and signaling pathways. KEGG = Kyoto Encyclopedia of Gene Genomes.

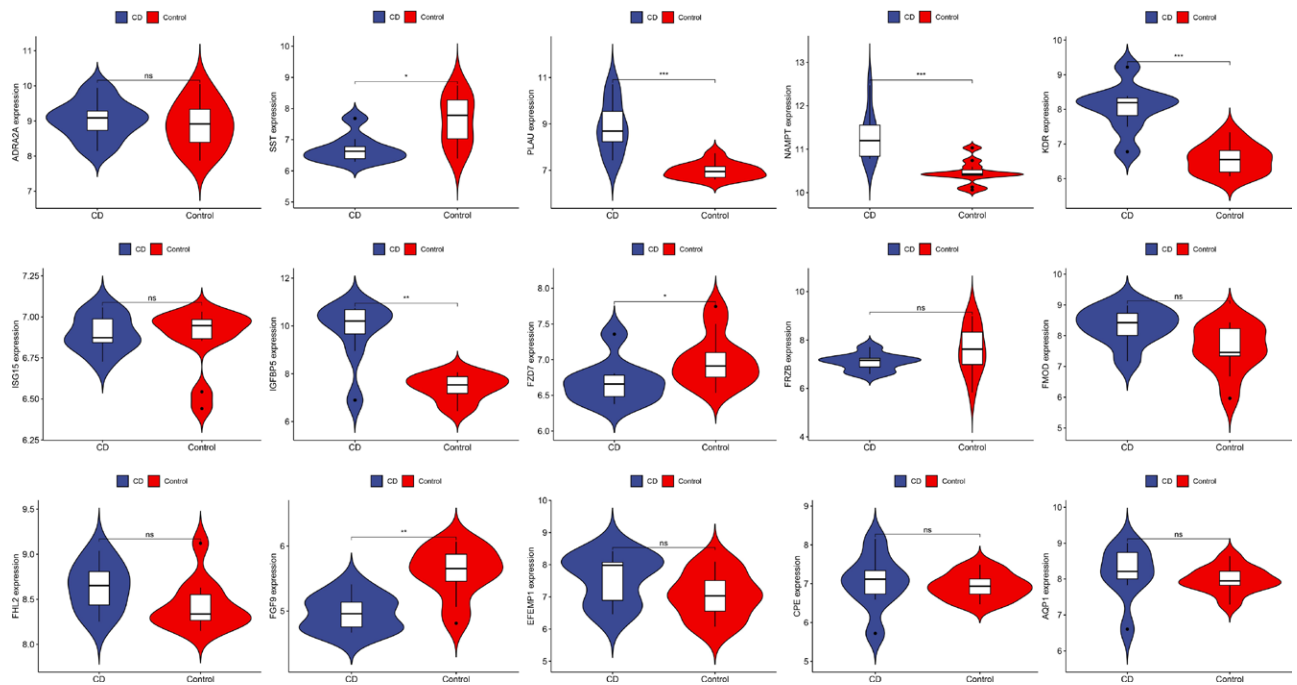


Figure 10. The expression of hub genes in GSE59071. ***: $P < .001$; **: $P < .01$; *: $P < .05$; CD = Crohn disease; Ns = no significance.

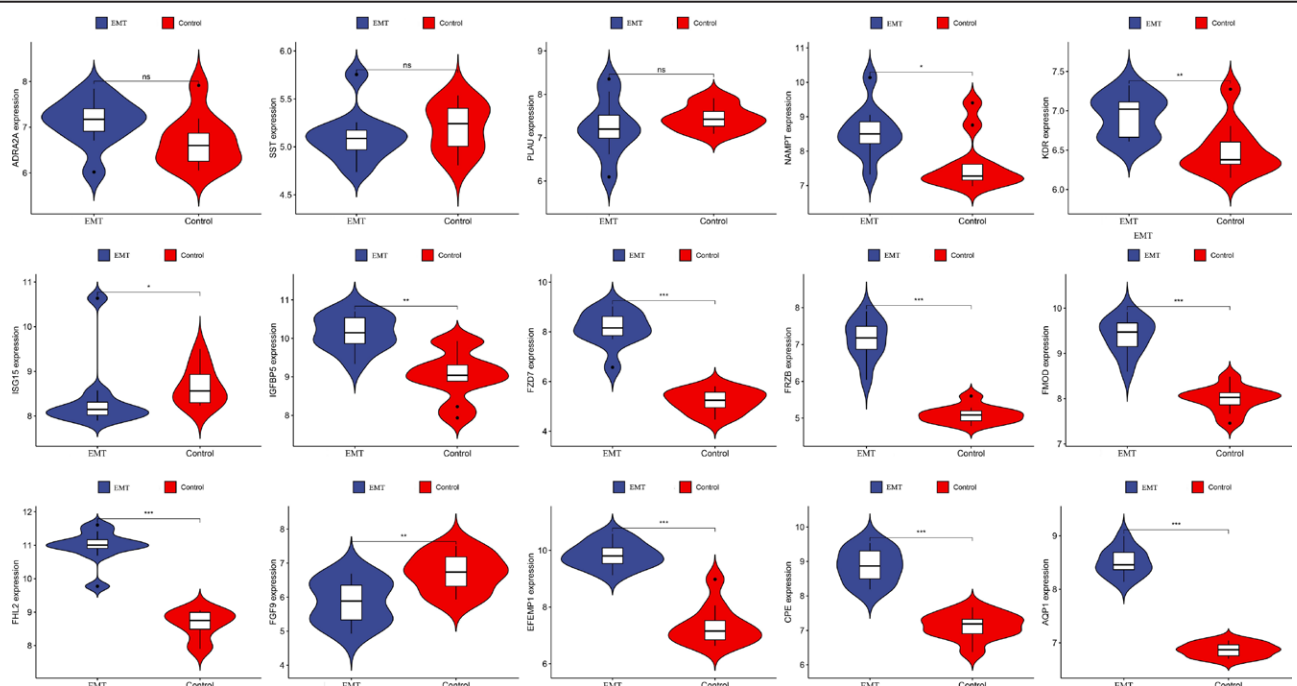


Figure 11. The expression of hub genes in GSE7305. ***: $P < .001$; **: $P < .01$; *: $P < .05$; EMT = endometriosis; Ns = no significance.

network. Pro-inflammatory TNF was a main contributor to CD and was regulated by JUN. Increased expression of JUN was found in CD patients and stimulated macrophages to product TNF.^[71] In addition, JUN interacted with the functional AP-1 transcription factor binding site to promote Metastasis-Associated in Colon Cancer 1 expression, which induced cell migration and metastasis in colorectal cancer.^[72] A study reported that transcription factor JUN contributed to aromatase P450 (P450arom) expression. P450arom was a key enzyme for biosynthesis of estrogen which was an important

element for the development of EMT.^[73] NFKB1 was a vital transcription regulator of immune response and its polymorphism was associated with the risk of CD.^[74] Additionally, NFKB1 polymorphism was also found to heighten susceptibility to EMT.^[75] RELA, also known as nuclear factor K κ -B p65 subunit, was strongly activated in IBD and played a central role in the persistent inflammation.^[76] Similarly, RELA was increased in EMT and mediated cell proliferation, invasion and inflammation of endometrial stromal cells.^[77] Transcription factor SRF was related with intestinal fibrosis. Increased SRF

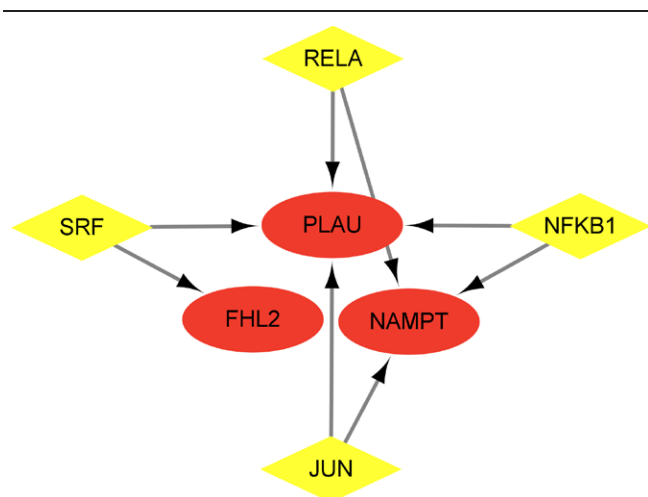


Figure 12. The TFs-hub genes regulation network. TFs = transcription factors.

stimulated myofibroblasts to synthesize collagen and fibronectin, promoting intestinal fibrosis in CD.^[78]

However, there are also some limitations in our study. On one hand, only 4 TFs were predicted via a public database. On the other hand, our study revealed the potential mechanism between EMT and CD only by nonexperiment methods. Therefore, our further study will utilize RT-qPCR to analyze the mRNA expression of hub genes in EMT and CD samples. Furthermore, we will investigate whether alterations in these hub genes would impact the enriched signaling pathways, ultimately influencing the development of these diseases.

5. Conclusion

In conclusion, our study uncovered a deep insight into the molecular mechanism between EMT and CD via bioinformatics analysis. Integrating multiple databases, we revealed the shared pathogenesis of EMT and CD, including inflammation, angiogenesis, immune regulation, cell behaviors, which may lead to the risk of CD in EMT. These genes (FGF9, FZD7, IGFBP5, KDR, and NAMPT) may closely relate to the common pathology of EMT and CD.

Author contributions

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Funding acquisition: Zhaotao Li.

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Validation: Weijie Zhou.

Visualization: Weijie Zhou.

Writing – original draft: Weijie Zhou.

Writing – review & editing: Weijie Zhou, Peizhu Su, Zhaotao Li, Liu Liu.

References

- [1] Kaplan G, Ng S. Understanding and preventing the global increase of inflammatory bowel disease. *Gastroenterology*. 2017;152:313–21.e2.
- [2] Ng S, Shi H, Hamidi N, et al. Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies. *Lancet (London, England)*. 2017;390:2769–78.
- [3] Baumgart D, Sandborn W. Crohn's disease. *Lancet (London, England)*. 2012;380:1590–605.

- [4] Torres J, Mehandru S, Colombel J, Peyrin-Biroulet L. Crohn's disease. *Lancet (London, England)*. 2017;389:1741–55.
- [5] Zondervan K, Becker C, Koga K, Missmer S, Taylor R, Viganò P. Endometriosis. *Nat Rev Dis Primers*. 2018;4:9.
- [6] Rogers P, D'Hooghe T, Fazleabas A, et al. Priorities for endometriosis research: recommendations from an international consensus workshop. *Reprod Sci (Thousand Oaks, Calif)*. 2009;16:335–46.
- [7] Horne A, Missmer S. Pathophysiology, diagnosis, and management of endometriosis. *BMJ (Clin Res Ed)*. 2022;379:e070750.
- [8] Izumi G, Koga K, Takamura M, et al. Involvement of immune cells in the pathogenesis of endometriosis. *J Obstet Gynaecol Res*. 2018;44:191–8.
- [9] Shigesu N, Kvaskoff M, Kirtley S, et al. The association between endometriosis and autoimmune diseases: a systematic review and meta-analysis. *Hum Reprod Update*. 2019;25:486–503.
- [10] Brinton L, Gridley G, Persson I, Baron J, Bergqvist A. Cancer risk after a hospital discharge diagnosis of endometriosis. *Am J Obstet Gynecol*. 1997;176:572–9.
- [11] Jess T, Frisch M, Jørgensen K, Pedersen B, Nielsen N. Increased risk of inflammatory bowel disease in women with endometriosis: a nationwide Danish cohort study. *Gut*. 2012;61:1279–83.
- [12] Montero-Meléndez T, Llor X, García-Planella E, Perretti M, Suárez A. Identification of novel predictor classifiers for inflammatory bowel disease by gene expression profiling. *PLoS One*. 2013;8:e76235.
- [13] Crispi S, Piccolo M, D'Avino A, et al. Transcriptional profiling of endometriosis tissues identifies genes related to organogenesis defects. *J Cell Physiol*. 2013;228:1927–34.
- [14] Kang YM, Lan A, Huang YH, Hsu KM, Chao Y, Lan KL. Identification of key genes and pathways associated with topotecan treatment using multiple bioinformatics tools. *J Chin Med Assoc*. 2020;83:446–53.
- [15] Wang Y, Zhou W, Chen Y, et al. Identification of susceptibility modules and hub genes of osteoarthritis by WGCNA analysis. *Front Genet*. 2022;13:1036156.
- [16] Liu Q, Zhang B, Liu C, Zhao D. Molecular mechanisms underlying the positive role of treadmill training in locomotor recovery after spinal cord injury. *Spinal Cord*. 2017;55:441–6.
- [17] Molodecky N, Soon I, Rabi D, et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology*. 2012;142:46–54.e42; quiz e30.
- [18] Oldani A, Monni M, Gentili S. Ileal endometriosis and Crohn's disease: an unusual association causing acute bowel obstruction. *Ann Ital Chir*. 2016;87:S2239253X1602510X.
- [19] Kaemmerer E, Westerkamp M, Kasperk R, Niepmann G, Scherer A, Gassler N. Coincidence of active Crohn's disease and florid endometriosis in the terminal ileum: a case report. *World J Gastroenterol*. 2013;19:4413–7.
- [20] Chiaffarino F, Cipriani S, Ricci E, et al. Endometriosis and inflammatory bowel disease: a systematic review of the literature. *Eur J Obstet Gynecol Reprod Biol*. 2020;252:246–51.
- [21] Sun Y, Liu W, Liu T, Feng X, Yang N, Zhou H. Signaling pathway of MAPK/ERK in cell proliferation, differentiation, migration, senescence and apoptosis. *J Recept Signal Transduct Res*. 2015;35:600–4.
- [22] Kim EK, Choi EJ. Pathological roles of MAPK signaling pathways in human diseases. *Biochim Biophys Acta*. 2010;1802:396–405.
- [23] Santulli P, Marcellin L, Tosti C, et al. MAP kinases and the inflammatory signaling cascade as targets for the treatment of endometriosis? *Expert Opin Ther Targets*. 2015;19:1465–83.
- [24] Feng Y, Li Y. The role of p38 mitogen-activated protein kinase in the pathogenesis of inflammatory bowel disease. *J Dig Dis*. 2011;12:327–32.
- [25] Xu X, Zhang M, Xu F, Jiang S. Wnt signaling in breast cancer: biological mechanisms, challenges and opportunities. *Mol Cancer*. 2020;19:165.
- [26] Klemmt P, Starzinski-Powitz A. Molecular and cellular pathogenesis of endometriosis. *Curr Women's Health Rev*. 2018;14:106–16.
- [27] Moparthi L, Koch S. Wnt signaling in intestinal inflammation. *Differentiation*. 2019;108:24–32.
- [28] Lewis A, Sánchez S, Berti G, et al. Small-molecule Wnt inhibitors are a potential novel therapy for intestinal fibrosis in Crohn's disease. *Clin Sci (London, England: 1979)*. 2022;136:1405–23.
- [29] Vojtek A, Der C. Increasing complexity of the Ras signaling pathway. *J Biol Chem*. 1998;273:19925–8.
- [30] Yotova I, Quan P, Leditznig N, Beer U, Wenzl R, Tschugguel W. Abnormal activation of Ras/Raf/MAPK and RhoA/ROCKII signalling pathways in eutopic endometrial stromal cells of patients with endometriosis. *Hum Reprod*. 2011;26:885–97.
- [31] Wang X, Lu Y, Wu L, et al. Moxibustion Inhibits the ERK signaling pathway and intestinal fibrosis in rats with crohn's disease. *Evid Based Complement Alternat Med*. 2013;2013:198282.

- [32] Liang X, Xu F, Li X, Ma C, Zhang Y, Xu W. VEGF signal system: the application of antiangiogenesis. *Curr Med Chem*. 2014;21:894–910.
- [33] Shaik-Dasthagirisahab Y, Varvara G, Murmura G, et al. Vascular endothelial growth factor (VEGF), mast cells and inflammation. *Int J Immunopathol Pharmacol*. 2013;26:327–35.
- [34] Hall M, Gourley C, McNeish I, et al. Targeted anti-vascular therapies for ovarian cancer: current evidence. *Br J Cancer*. 2013;108:250–8.
- [35] Hung S, Zhang R, Tan Z, Chung J, Zhang T, Wang C. Pharmaceuticals targeting signaling pathways of endometriosis as potential new medical treatment: a review. *Med Res Rev*. 2021;41:2489–564.
- [36] Vodolazkaia A, Yesilyurt B, Kyama C, et al. Vascular endothelial growth factor pathway in endometriosis: genetic variants and plasma biomarkers. *Fertil Steril*. 2016;105:988–96.
- [37] Cane G, Moal V, Pagès G, Servin A, Hofman P, Vouret-Craviari V. Up-regulation of intestinal vascular endothelial growth factor by Afa/Dr diffusely adhering *Escherichia coli*. *PLoS One*. 2007;2:e1359.
- [38] Sarnelli G, D'Alessandro A, Iuvone T, et al. Palmitoylethanolamide Modulates Inflammation-Associated Vascular Endothelial Growth Factor (VEGF) Signaling via the Akt/mTOR Pathway in a Selective Peroxisome Proliferator-Activated Receptor Alpha (PPAR- α)-Dependent Manner. *PLoS One*. 2016;11:e0156198.
- [39] Zi Z. Molecular Engineering of the TGF- β Signaling Pathway. *J Mol Biol*. 2019;431:2644–54.
- [40] Choi H, Park M, Kim B, et al. Transforming growth factor β 1 enhances adhesion of endometrial cells to mesothelium by regulating integrin expression. *BMB Rep*. 2017;50:429–34.
- [41] Liu Z, Yi L, Du M, Gong G, Zhu Y. Overexpression of TGF- β enhances the migration and invasive ability of ectopic endometrial cells via ERK/MAPK signaling pathway. *Exp Ther Med*. 2019;17:4457–64.
- [42] Sikora J, Smycz-Kubańska M, Mielczarek-Palacz A, Bednarek I, Kondera-Anasz Z. The involvement of multifunctional TGF- β and related cytokines in pathogenesis of endometriosis. *Immunol Lett*. 2018;201:31–7.
- [43] Mizumoto Y. Changes in NK activities and TGF- β concentrations in the peritoneal cavity in endometriosis and their interaction related with infertility. *Nihon Sanka Fujinka Gakkai Zasshi*. 1996;48:379–85.
- [44] Ihara S, Hirata Y, Koike K. TGF- β in inflammatory bowel disease: a key regulator of immune cells, epithelium, and the intestinal microbiota. *J Gastroenterol*. 2017;52:777–87.
- [45] Young V, Ahmad S, Duncan W, Horne A. The role of TGF- β in the pathophysiology of peritoneal endometriosis. *Hum Reprod Update*. 2017;23:548–59.
- [46] Zhang X, Ibrahim O, Olsen S, Umemori H, Mohammadi M, Ornitz D. Receptor specificity of the fibroblast growth factor family. The complete mammalian FGF family. *J Biol Chem*. 2006;281:15694–700.
- [47] Krejci P, Prochazkova J, Bryja V, Kozubik A, Wilcox W. Molecular pathology of the fibroblast growth factor family. *Hum Mutat*. 2009;30:1245–55.
- [48] Abdel-Rahman W, Kalinina J, Shoman S, et al. Somatic FGF9 mutations in colorectal and endometrial carcinomas associated with membranous beta-catenin. *Hum Mutat*. 2008;29:390–7.
- [49] Chuang P, Sun H, Chen T, Tsai S. Prostaglandin E2 induces fibroblast growth factor 9 via EP3-dependent protein kinase Cdelta and Elk-1 signaling. *Mol Cell Biol*. 2006;26:8281–92.
- [50] Wing L, Chuang P, Wu M, Chen H, Tsai S. Expression and mitogenic effect of fibroblast growth factor-9 in human endometriotic implant is regulated by aberrant production of estrogen. *J Clin Endocrinol Metab*. 2003;88:5547–54.
- [51] Geske M, Zhang X, Patel K, Ornitz D, Stappenbeck T. Fgf9 signaling regulates small intestinal elongation and mesenchymal development. *Development (Cambridge, England)*. 2008;135:2959–68.
- [52] Dong D, Na L, Zhou K, et al. FZD5 prevents epithelial-mesenchymal transition in gastric cancer. *Cell Commun Signal*. 2021;19:21.
- [53] Zhu H, Cao X, Liu J, Hua H. MicroRNA-488 inhibits endometrial glandular epithelial cell proliferation, migration, and invasion in endometriosis mice via Wnt by inhibiting FZD7. *J Cell Mol Med*. 2019;23:2419–30.
- [54] Flanagan D, Phesse T, Barker N, et al. Frizzled7 functions as a Wnt receptor in intestinal epithelial Lgr5(+) stem cells. *Stem Cell Rep*. 2015;4:759–67.
- [55] Gu N, Guo Y, Lin S, et al. Frizzled 7 modulates goblet and Paneth cell fate, and maintains homeostasis in mouse intestine. *Development (Cambridge, England)*. 2023;150:dev200932.
- [56] Duan C, Allard J. Insulin-like growth factor binding protein-5 in physiology and disease. *Front Endocrinol*. 2020;11:100.
- [57] Ding H, Wu T. Insulin-like growth factor binding proteins in autoimmune diseases. *Front Endocrinol*. 2018;9:499.
- [58] Yasuoka H, Zhou Z, Pilewski J, Oury T, Choi A, Feghali-Bostwick C. Insulin-like growth factor-binding protein-5 induces pulmonary fibrosis and triggers mononuclear cellular infiltration. *Am J Pathol*. 2006;169:1633–42.
- [59] Yasuoka H, Yamaguchi Y, Feghali-Bostwick C. The pro-fibrotic factor IGFBP-5 induces lung fibroblast and mononuclear cell migration. *Am J Respir Cell Mol Biol*. 2009;41:179–88.
- [60] Zimmermann E, Li L, Hou Y, Mohapatra N, Pucilowska J. Insulin-like growth factor I and insulin-like growth factor binding protein 5 in Crohn's disease. *Am J Physiol Gastrointest Liver Physiol*. 2001;280:G1022–9.
- [61] Flynn R, Mahavadi S, Murthy K, Kellum J, Kuemmerle J. Insulin-like growth factor-binding protein-5 stimulates growth of human intestinal muscle cells by activation of G[alpha]i3. *Am J Physiol Gastrointest Liver Physiol*. 2009;297:G1232–8.
- [62] Zhou J, Dsupin B, Giudice L, Bondy C. Insulin-like growth factor system gene expression in human endometrium during the menstrual cycle. *J Clin Endocrinol Metab*. 1994;79:1723–34.
- [63] Zhang T, De Carolis C, Man G, Wang C. The link between immunity, autoimmunity and endometriosis: a literature update. *Autoimmun Rev*. 2018;17:945–55.
- [64] Terman B, Dougher-Vermazen M, Carrion M, et al. Identification of the KDR tyrosine kinase as a receptor for vascular endothelial cell growth factor. *Biochem Biophys Res Commun*. 1992;187:1579–86.
- [65] Melincovici C, Boşca A, Şuşman S, et al. Vascular endothelial growth factor (VEGF) - key factor in normal and pathological angiogenesis. *Romanian J Morphol Embryol*. 2018;59:455–67.
- [66] Garten A, Schuster S, Penke M, Gorski T, de Giorgis T, Kiess W. Physiological and pathophysiological roles of NAMPT and NAD metabolism. *Nat Rev Endocrinol*. 2015;11:535–46.
- [67] Audrito V, Messana V, Deaglio S. NAMPT and NAPRT: two metabolic enzymes with key roles in inflammation. *Front Oncol*. 2020;10:358.
- [68] Neubauer K, Bednarek-Misa I, Walecka-Zacharska E, et al. Oversecretion and overexpression of Nicotinamide Phosphoribosyltransferase/Pre-B Colony-Enhancing Factor/Visfatin in inflammatory bowel disease reflects the disease activity, severity of inflammatory response and hypoxia. *Int J Mol Sci*. 2019;20:166.
- [69] Annie L, Gurusubramanian G, Roy V. Estrogen and progesterone dependent expression of visfatin/NAMPT regulates proliferation and apoptosis in mice uterus during estrous cycle. *J Steroid Biochem Mol Biol*. 2019;185:225–36.
- [70] Lambert S, Jolma A, Campitelli L, et al. The human transcription factors. *Cell*. 2018;172:650–65.
- [71] Lin Y, Thibodeaux C, Peña J, Ferry G, Versalovic J. Probiotic *Lactobacillus reuteri* suppress proinflammatory cytokines via c-Jun. *Inflamm Bowel Dis*. 2008;14:1068–83.
- [72] Kobelt D, Zhang C, Clayton-Lucey I, et al. Pro-inflammatory TNF- α and IFN- γ promote tumor growth and metastasis via Induction of MACC1. *Front Immunol*. 2020;11:980.
- [73] Yu C, Li Y, Chen H, Yang S, Xie G. Decreased expression of aromatase in the Ishikawa and RL95-2 cells by the isoflavone, puerarin, is associated with inhibition of c-jun expression and AP-1 activity. *Food Chem Toxicol*. 2008;46:3671–6.
- [74] Bank S, Skytt Andersen P, Burisch J, et al. Polymorphisms in the inflammatory pathway genes TLR2, TLR4, TLR9, LY96, NFKB1A, NFKB1, TNFA, TNFRSF1A, IL6R, IL10, IL23R, PTPN22, and PPARG are associated with susceptibility of inflammatory bowel disease in a Danish cohort. *PLoS One*. 2014;9:e98815.
- [75] Zhou B, Rao L, Peng Y, et al. A functional promoter polymorphism in NFKB1 increases susceptibility to endometriosis. *DNA Cell Biol*. 2010;29:235–9.
- [76] Borg S, Björkman J, Eriksson S, et al. Novel *Salmonella typhimurium* properties in host-parasite interactions. *Immunol Lett*. 1999;68:247–9.
- [77] Wu M, Zhang Y. MiR-182 inhibits proliferation, migration, invasion and inflammation of endometrial stromal cells through deactivation of NF- κ B signaling pathway in endometriosis. *Mol Cell Biochem*. 2021;476:1575–88.
- [78] Choi Y, Koo J, Kim H, et al. Umbilical cord/placenta-derived mesenchymal stem cells inhibit fibrogenic activation in human intestinal myofibroblasts via inhibition of myocardin-related transcription factor A. *Stem Cell Res Ther*. 2019;10:291.