Research Paper

Metformin administration during pregnancy attenuated the longterm maternal metabolic and cognitive impairments in a mouse model of gestational diabetes

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ABSTRACT

Background: Gestational diabetes mellitus (GDM) is a metabolic disease that can have long-term adverse effects on the cognitive function of mothers. In our study, we explored the changes in metabolic health and cognitive function in mice of middle- and old- age after exposure to GDM, and whether metformin therapy during pregnancy provided long-term benefits.

Results: Mice with GDM demonstrated significant cognitive impairment in old age, which was associated with insulin resistance. Gestational metformin therapy was shown to increase insulin sensitivity and improve cognition. The ovarian aging rate was also accelerated in mice exposed to GDM during pregnancy, which may be related to fatty acid metabolism in the ovaries.

Conclusion: Treatment with metformin during pregnancy was shown to improve fatty acid metabolism in ovarian tissues.

Method: During pregnancy, mice were fed with a high-fat diet (GDM group) or a low-fat diet (Control group), and a third group received metformin while receiving a high-fat diet (Treatment group). At 12 months old, the mice completed an oral glucose tolerance test, insulin tolerance test, Morris water maze test, female sex hormones were measured, and metabolite profiles of tissue from the ovaries, hypothalamus, and pituitary glands were analysed using gas chromatography-mass spectrometry.

INTRODUCTION

Gestational diabetes mellitus (GDM) is a metabolic disease diagnosed when glucose intolerance is observed with onset or first occurrence during pregnancy. The global incidence of GDM is around 7 % [1]. However,

in recent years, the incidence of GDM has been rising in China, with prevalence as high as 17.5 % in some areas [2]. Women who have GDM are also at an increased risk of developing other pregnancy complications including gestational hypertension, premature delivery, macrosomia, and neonatal respiratory distress syndrome [3–5]. Despite most GDM patients postpartum blood glucose levels returning to normal, these women are at a significantly higher risk of developing type 2 diabetes mellitus (T2DM) post-delivery. In women who have experienced GDM during their pregnancy, the incidence of developing diabetes was 3.7% at 9 months postpartum, 4.9 % at 15 months postpartum, 13.1 % at 5.2 years postpartum, and 18.9 % at 9 years postpartum, while the incidence of diabetes remained at 2.0 % in women without a history of GDM [6].

Women who develop GDM experience metabolic disturbances similar to that of T2DM and there seem to be equivalent pathophysiological mechanisms contributing to the complications of both medical conditions. One such complication is later life cognitive dysfunction [7–9]. It is generally accepted that persistent hyperglycemia and insulin resistance lead to brain degeneration and dysfunction, and subsequently impaired cognition. Mouse models of diabetes established by genetic modifications or fed with a highfat diet (HFD), perform poorly on various learning and memory behavioural tests [10, 11]. Other factors contributing to cognitive dysfunction are sex-steroid hormones and aging. The main regulatory loop of the female sex hormones includes the hypothalamus, and ovary (HPO axis). pituitary gland, The hypothalamus receives information from the central nervous system and secretes a gonadotropin-releasing hormone (GnRH), which in turn stimulates the secretion of gonadotropic hormones in the pituitary gland and ultimately affects ovarian function. Hypofunction of the HPO axis is one of the important signs of female aging. Studies have found that levels of estrogen and progesterone begin to reduce as ovarian function declines [12]. Growing evidence suggests that large declines in female sex hormones that occur with menopause and aging render females more prone to diabetes, neurodegeneration, cognitive impairment, and memory disorders [13-15]. Moreover, low levels of sex steroid hormones are considered a risk factor for neurodegenerative diseases (e.g. Alzheimer's Disease), whilst therapeutic interventions using estrogen and progesterone have been shown to be neuroprotective [15–17]. However, there is no research investigating how these risk factors will influence the long-term cognitive health of women with a history of GDM.

In this study, we aimed to investigate the associations of diabetic risk factors including hyperglycemia, insulin resistance, hormonal changes, and age, together with GDM, on the cognitive function of postpartum women during middle-old age. A GDM mouse model was established using a high-fat diet. Blood glucose levels, insulin resistance, sex hormones, and metabolic profiles of the HPO axis were measured in mice at 12 months of age. In addition, we investigated whether metformin administration during pregnancy could provide longterm protection against GDM-associated cognitive dysfunction.

RESULTS

Body weight, oral glucose tolerance test (OGTT), and insulin tolerance test (ITT) results of mice during pregnancy and immediately after delivery

As depicted in Figure 1, the body weight of maternal mice was significantly lower in the low-fat diet group (LFD+vehicle) compared to the high-fat diet group (HFD+vehicle) and metformin treatment group (HFD+metformin) during the peripartum period. At day 16.5 of pregnancy, the OGTT and ITT results exhibited a similar trend; levels at 30 min and 60 min were significantly higher in the HFD group compared to the LFD and metformin treatment groups, as were the AUCs (Figure 2A and 2D). After delivery, the HFD group had significantly higher 90 min OGTT results and both 15 min and 30 min ITT results, compared to the LFD and metformin treatment groups (Figure 2E).

OGTT and IIT results of mice at 12 months of age

As shown in Figure 2C, the blood glucose concentrations and area under the curve (AUC) values for the OGTT were not significantly different between the three groups (F=1.624, p-value=0.230). In contrast, the blood glucose concentrations at 30 and 60 min for the ITT were significantly lower in the LFD group compared to the HFD and metformin treatment groups (Figure 2F). There was also a decreased trend in blood glucose AUC values for the ITT when comparing the LFD group to the HFD group (Figure 2F).

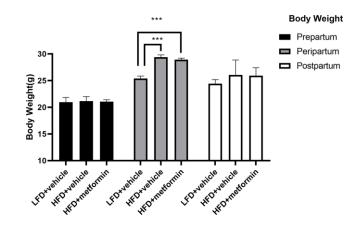
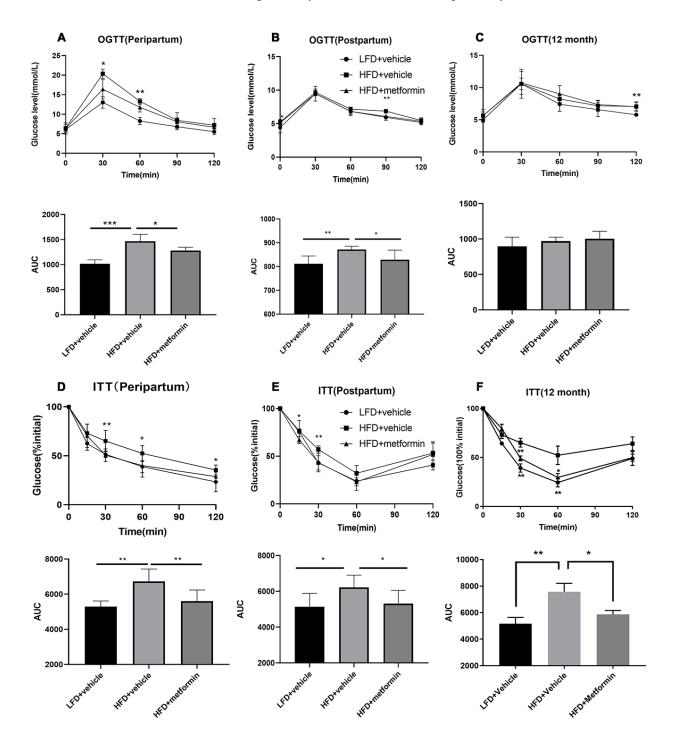


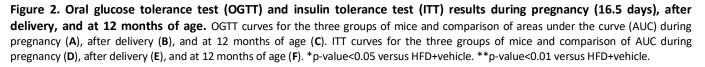
Figure 1. The body weight of maternal mice before, during, and after pregnancy. ***p-value<0.001.

Follicle-stimulating hormone (FSH), estrogen (E2), and progesterone (P) levels of mice at 12 months

There were no significant differences in serum levels of FSH or E2 across the three experimental groups (Figure 3A and 3B). However, the serum P level was significantly

higher in the LFD group compared to both the HFD and metformin treatment groups (Figure 3C). These results showed that serum P levels were significantly lower in 12-month-old mice exposed to a HFD (GDM mouse model) during pregnancy and that this lower hormone level was not improved by metformin.





Morris water maze performance

To evaluate cognitive function of the mice at 12 months of age, we conducted the Morris water maze experiment. The results of the training day during the first five days are displayed in Figure 4. There was a significantly longer latency to escape onto the hidden platform in the HFD group when compared to both the LFD and metformin treatment groups at day 3 (p-value <0.05) and day 4 (p-value <0.05) of the training period. However, 2-way ANOVA showed that there was no statistical significance of the interaction between training days and escape latency (p-value>0.05). On experimental day one (sixth day) when the swim times and distance covered to reach the removed platform were measured (Supplementary Figure 4), the mice fed with a HFD during pregnancy exhibited a longer swim time and swimming distance (Supplementary Table 1). The HFD group also spent more time on the quadrant where the platform was previously located, as well as a greater number of times crossing the platform (Supplementary Figure 4). These findings indicate that GDM mice expressed poor spatial learning and memory capability in later life and metformin administration during pregnancy could reverse this neurocognitive deficiency.

Metabolite profiles of the hypothalamus, pituitary gland, and ovarian tissues

A total of 125 compounds were identified in the hypothalamus, pituitary gland and ovary. The PCA demonstrated no obvious separation of the hypothalamus and pituitary gland tissue between the LFD, HFD, and metformin treatment groups (Figure 5A and 5B). The metformin treatment group and LFD group were clustered together and separated away from the HFD group in the ovary metabolite profile (PC1 and PC2 explained 62.5% and 13.3% of variance, respectively; Figure 5C). We then utilised OPLS-DA and S-plot to screen for statistically important metabolites leading the discrimination of each of the LFD and metformin treatment groups from the HFD group. As demonstrated in Figure 6, both OPLS-DA models (First model, LFD+vehicle vs HFD+vehicle; Second model, HFD+metformin vs HFD+ vehicle) vielded good class separation and statistical validation (R2 = 0.89, Q2 = 0.87; R2 = 0.82, Q2 = 0.78,

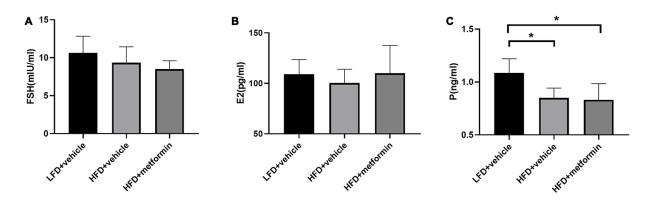
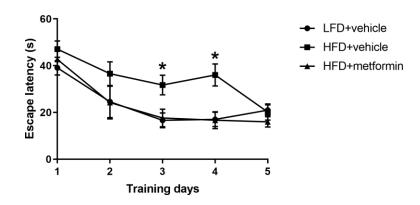
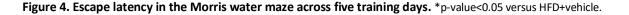


Figure 3. The serum level of follicle-stimulating hormone (A), estrogen (B), and progesterone (C) at 12 months of age. *p-value <0.05.





respectively). By combining VIP scores of the OPLS-DA (VIP > 1), Student's-T test (p-value >0.05), and covariance of the S-plot (p (corr)| > 0.5), a shortlist of significant metabolites was generated. A list of 25 and 15 metabolites passed the selection criteria for the LFD+vehicle vs HFD+vehicle and HFD+metformin vs HFD+vehicle, respectively (Figure 6A and 6B). Finally, a SUS-plot was constructed to identify the significant metabolites that were shared and unique in the two OPLS-DA models (Figure 7A). Four metabolites: 4-oxo pentanoic acid, 9-cis-hexadecenoic acid, isobutyl methyl phthalate, and 5,8,11,14,1,7-cis-eicosapentaenoic acid were significant in both models. The heatmap in Figure 7B shows that all of the

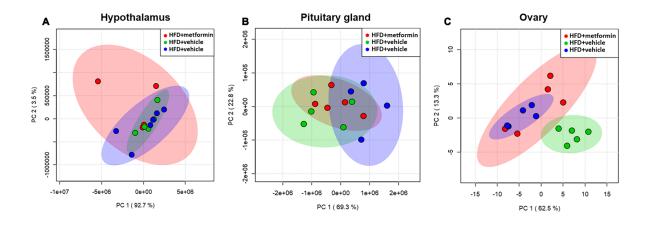


Figure 5. Principal component analysis of the hypothalamus (**A**), pituitary gland (**B**), and ovary (**C**) collected from maternal mice at 12 months of age. The color legends of experimental mice groups are listed as follows; Red color indicates maternal mice fed with a high-fat diet and metformin during pregnancy (HFD+metformin); Green color indicates maternal mice fed with a low-fat diet during pregnancy (LFD); Blue color indicates maternal mice fed with a high-fat diet during pregnancy (HFD).

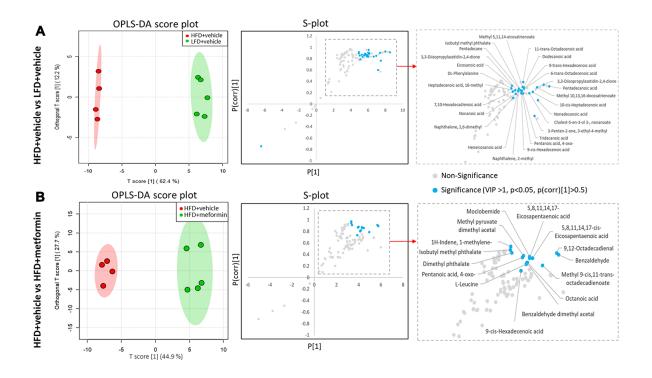


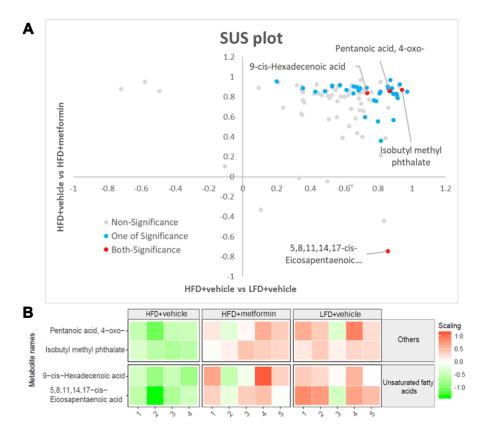
Figure 6. Identification of significant metabolites in the ovary. (A) OPLS-DA and S-plot were used to detect metabolites that discriminated between HFD+vehicle (red) and LFD+vehicle (green). (B) OPLS-DA and S-plot were used to detect metabolites that discriminated between HFD+vehicle (red) and HFD+metformin (green). In S-plot, the blue circles indicate significant metabolites with a VIP of > 1, p-value <0.05, and P(corr) >0.5.

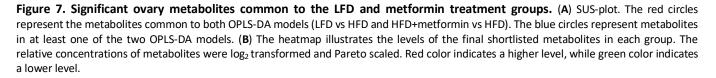
shortlisted metabolites were in lower concentrations in the HFD group compared to the LFD and metformin treatment groups. The changes in these metabolites between the HFD group and the metformin treatment group may represent the effect of metformin treatment on the ovaries of 12 month old mice exposed to GDM during pregnancy.

DISCUSSION

In this study, we investigated how risk factors such as hyperglycemia, insulin resistance, aging, and abnormal levels of female sex hormones contribute to the acceleration of cognitive decline in mice of middle-old age who had previously had GDM. Results of the Morris water maze test showed that compared to the control group, the cognitive function of mice in the GDM group was significantly reduced after entering middle-old age. We also observed impaired insulin tolerance and greater metabolic changes downstream of the HPO axis at 12 months of age. Treatment with metformin during pregnancy significantly improved the long-term cognitive function of mice exposed to GDM during pregnancy.

We have successfully established a mouse model that resembles the pathophysiological state of GDM; including having a higher body weight and confirmed hyperglycemia (Figures 1 and 2). No difference in glucose tolerance was observed between GDM and control mice at 12 months (Figure 2C). Hyperglycemia in GDM only occurred temporarily during gestation and blood glucose levels gradually returned to normal postdelivery. Thus, hyperglycemia in later life did not seem to be responsible for the GDM-related cognitive impairment. Moreover, of the hormones analysed, only progesterone levels were significantly reduced in the GDM and metformin treatment groups at 12 months (Figure 3). Progesterone typically declines earlier than estrogen in the first stages of perimenopause. Studies have suggested that cognitive impairment related to the perimenopausal period is related to the reduction of estrogen [18, 19]. Estrogen is thought to be involved in maintaining higher cognitive functions by inducing spinogenesis and synaptogenesis in the prefrontal cortex and hippocampus, via estrogen receptors [19]. A plethora of evidence has been presented which shows that estrogen loss resulting from menopause or the





surgical removal of an ovary/ovaries, can accelerate age-related cognitive decline. Estrogen replacement therapy has been shown to improve the cognitive function of these women [20]. Reductions in progesterone without reductions in estrogen merely represent early-stage functional decline of the ovaries, as part of the natural process of early perimenopause. Therefore, our results suggest that hyperglycemia, female sex hormones, and aging may not be the key factors leading to the cognitive impairments observed in the GDM group in later life.

Although hyperglycemia observed during GDM resolves postpartum, insulin resistance persists [21, 22]. Insulin resistance appears to be the most likely mediator of the cognitive impairment observed in later life. after exposure to GDM during pregnancy. Our results indicate that the GDM group had significantly reduced insulin sensitivity at 12 months of age (Figure 2F). Therefore, a mother who develops GDM manifests similar metabolic disturbances to T2DM, which is characterized by insulin resistance and insulin signalling deficiency [23, 24]. Many animal studies have emphasized the direct association between insulin resistance and cognitive dysfunction [25, 26]. Studies using mouse models of T2DM have revealed that hippocampal insulin resistance leads to neuroplasticity impairment [27-29]. HFD-induced models of diabetes have observed a reduction in cell proliferation and neurogenesis in the dentate gyrus of the hippocampus [30]. Furthermore, peripheral insulin resistance can directly contribute to brain insulin resistance by compromising transport of insulin into the central nervous system [31]. The restoration of insulin signalling activities in the hippocampus has been shown to alleviate the cognitive decrements observed in a T2DM mouse model [32]. These studies show that insulin resistance causes structural and functional abnormalities in the brain. Therefore, persistent insulin resistance appears to be an important factor in the longterm reduction in cognitive function observed following GDM. In addition, recent studies have shown aberrant gut microbiota can result in the development of cognitive dysfunction and T2DM in diabetic mouse models [33, 34]. The findings from these studies suggest that the gut microbiota could also influence host insulin sensitivity and cognitive function.

Despite the sex hormones remaining at a normal physiological level in the GDM group at 12 months of age, a sex hormone regulation and feedback system "HPO axis" was investigated to see whether it was adversely affected by exposure to GDM. A cascade amplification effect on the downstream regulation of HPO was observed in the early perimenopausal period. In particular, substantial metabolic changes were

detected in ovarian tissues of both normal and treatment groups when compared to GDM groups (Figure 5). Pentanoic acid, 9-cis-hexadecenoic acid, and isobutyl methyl phthalate were significantly lower in ovarian tissue from the GDM group (Figure 7). 9-cishexadecenoic acid (palmitoleic acid) is a common unsaturated fatty acid in the ovary that regulates gonadotropin-stimulated progesterone biosynthesis in granulosa cells [35]. 9-cis-hexadecenoic acid can also synergistically stimulate cell proliferation via IGF-1 [36]. Liu et al (2001) [37] has previously reported that pentanoic acid has a potent effect on stimulating protein synthesis without accelerating cell apoptosis in a rodent ovarian cell culture. Since these fatty acids are involved in protein biosynthesis, anti-apoptosis, hormone production, and cell proliferation, lower levels of these metabolites may accelerate the aging process in the ovaries of women with a history of GDM. The process of female reproductive aging is believed to be predominated by the age-associated decline in ovarian function. The number of follicles in the ovary are diminished and oocyte quality declines as chronological age increases. This leads to a reduction in ovarian hormones and subsequently compromises the negative feedback from the ovarian sex-steroids to the hypothalamic-pituitary axis [38]. Because maternal mice in our study were sampled at the premenopausal stage, the ovarian feedback status had not diminished enough to dysregulate the upstream components of the HPO axis. Based on these findings, we postulate that GDM manifestation results in a negative metabolic influence on the downstream aspects of the HPO axis and thereby accelerates postpartum ovarian aging.

To investigate the therapeutic effect of metformin, a drug known to increase peripheral target cell insulin sensitivity, mice with GDM were administrated metformin during pregnancy. Our results demonstrated that insulin sensitivity was returned to a normal level in the metformin treatment group, as shown in Figure 4C. Metformin has been reported to reduce insulin resistance by activating AMPK and SIRT1 to inhibit hepatic gluconeogenesis, suppress lipid biosynthesis, and enhance glucose metabolism, thus decreasing circulating glucose and lipid levels [39-41]. Metformin can also reduce insulin resistance by downregulating chemerin and suppressing endoplasmic reticulum stress in adipose tissue and liver cells [39, 42]. A recent human gut metagenomic study indicated that metformin treatment can improve insulin sensitivity by favouring gut Escherichia species. In doing so, the authors observed an increase in microbial short-chain fatty acid production, attenuation of intestinal lipid absorption, lipopolysaccharide (LPS)-induced and reduced inflammation [43]. Importantly, we also found that GDM mice administrated metformin during pregnancy

had spatial learning and memory capability scores equivalent to normal mice at 12 months of age (Figure 4). Although clinical studies have shown that the administration of metformin during pregnancy can effectively control blood glucose and improve insulin sensitivity in GDM [44-46], our findings highlight the protective effects metformin long-term of administration on cognitive health. Evidence strongly supports the role of insulin resistance in cognitive decline and it has been suggested that an insulin sensitizer may prevent against cognitive decline in prediabetic and diabetic patients [10, 47-48]. Metformin's proposed mechanisms of action on neurons and microglia include AMPK-associated neural proliferation, differentiation, self-renewal, autophagy, and energy homeostasis [10, 49–57]. Metformin inhibits inflammation by suppressing NF-KB [58] as well as promoting glucose consumption, lactate production, and reducing oxidative phosphorylation, thereby favoring glycolytic metabolism in astrocytes and microglia [59-61]. Based on these findings, we speculate that metformin therapy during GDM pregnancy may reverse the long-term maternal cognitive impairments in middle-old age by attenuating persistent postpartum insulin resistance. Despite metformin treatment during pregnancy not raising progesterone levels in the GDM group, treatment with metformin seemed to protect the downstream aspects of the HPO axis, including ovarian tissue metabolism. These findings could be explained by metformin's ability to reduce blood glucose levels by minimizing the absorption of glucose in the intestine and inhibiting gluconeogenesis in the liver, without stimulating the secretion of insulin [62–64].

The results of our study demonstrated that despite postpartum weight, diet, and blood glucose returning to a normal status after GDM, insulin resistance persisted. We propose that prolonged insulin resistance is likely responsible for the adverse effects on long-term endocrine and cognitive functions (Figure 8). The potential clinical translation of this finding is that medical professionals should monitor women with a history of GDM, even if their weight and blood glucose return to normal after delivery. In particular, their insulin tolerance should be regularly monitored. If insulin resistance remains persistent, it should be corrected in order to protect the endocrine and neurocognitive functions of women. More importantly, persistent postpartum insulin resistance may be minimized or avoided in GDM mothers by administrating metformin during pregnancy. Future research should include mechanistic studies such as gene knockout and cell models to test whether persistent postpartum insulin resistance is the primary cause of cognitive impairment and ovarian aging. Additional studies are also needed to test whether a higher dose of metformin supplementation during pregnancy or extending its treatment time to the postpartum period could improve maternal hormonal levels.

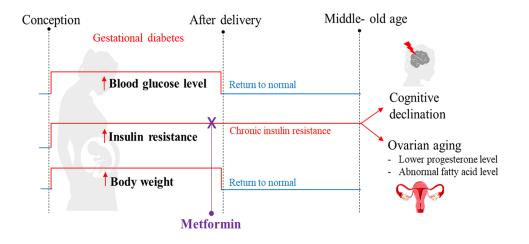


Figure 8. Summary of the proposed mechanism explaining how gestational diabetes mellitus (GDM) may lead to cognitive impairment and ovarian aging later in maternal life. Our high-fat diet (HFD) mouse model exhibited the pathophysiological phenotype that resembles GDM, including hyperglycemia, being overweight, and experiencing insulin resistance during pregnancy. After delivery, all mice were reverted back to a standard diet. In the postpartum period, the blood glucose level and body weight returned to normal, but insulin resistance persisted and reduced cognitive function was observed at 12 months of age. A reduced progesterone level was also observed; an early indication of perimenopause. Through metabolome profiling of the hypothalamus, pituitary gland, and ovarian (HPO) axis, a downstream dysregulation of the HPO axis was revealed. In particular, ovarian fatty acid levels were reduced. All these adverse outcomes were prevented when insulin resistance during pregnancy was treated with metformin. These phenotypes were not directly associated with hyperglycemia or high-fat diet during gestation, nor differences in bodyweight after pregnancy. Based on these observations, we hypothesize that persistent insulin resistance postpartum is the primary cause of GDM-related cognitive impairment and accelerated ovarian decline.

In conclusion, the cognitive function of mice exposed to GDM was significantly impaired compared to controls after entering middle- old age. Our findings suggest that the cognitive impairments were mainly related to persistent insulin resistance after delivery. Treatment of GDM with metformin during pregnancy significantly improved postpartum insulin resistance and was protective of long-term cognitive health. In addition, GDM altered the long-term metabolism of the ovary and thus accelerated the chronological aging process. Although metformin treatment of GDM during pregnancy improved the metabolite pattern of the HPO axis, whether it can further affect the function of the HPO axis remains to be investigated.

MATERIALS AND METHODS

Overall experimental design

Eighteen C57BL/6 mice were randomly divided into three groups: control (low-fat diet (LFD) + vehicle, n=6), GDM (high-fat diet (HFD) + vehicle, n=6) and metformin treatment (HFD+metformin, n=6). There was no metformin treatment for the LFD group because our pilot study showed that metformin treatment did not change the blood glucose level or insulin resistance in the mice fed with a LFD during pregnancy (Supplementary Figure 1). The control mice were fed a low-fat diet (Research Diets AIN-93G, consisting of 20.3 % protein, 63.9 % carbohydrate, and 15.8 % fat) for one week prior to mating and throughout pregnancy (18.5 days), while both GDM and treatment groups were fed a high-fat diet (Research Diets D12451, consisting of 20% protein, 35 % carbohydrate, and 45 % fat). All mice were given free access to 100 grams of

fresh diet and 250 ml of fresh water daily per cage (Five mice per cage). From 11.5 to 17.5 days of pregnancy, the treatment group received a 300 mg dose of metformin solution each day via oral gavage, whilst the control and GDM group received the same dose of a vehicle (phosphate buffer solution). After delivery, all maternal mice were reversed back to normal chow (Research Diet 1022, consisting of 18% protein, 78% carbohydrate, and 4 % fat) until the age of 12 months. At 12 months of age, the mice performed the Morris water maze experiment, and underwent an oral glucose tolerance test (OGTT) and insulin tolerance test (ITT). In addition, the body weight of the mice was measured prepartum, peripartum and postpartum. After the mice were sacrificed, 200 µl blood was collected by cardiac puncture. The blood was centrifuged at 1300 g for 10 min at 4 °C in order to obtain serum for hormone measurements such as follicle-stimulating hormone (FSH), estrogen, and progesterone. Furthermore, the ovaries, hypothalamus and pituitary gland were collected and stored in a -80 °C freezer until gas chromatography-mass spectrometry (GC-MS) based metabolomics was performed (Figure 9). The handling of animals in this research was in accordance with the guidelines approved by the ethics committee of the First Affiliated Hospital of Chongqing Medical University (Ethics number 2016-41). All efforts were taken to minimize the number of mice sacrificed and any potential suffering.

Oral glucose tolerance test (OGTT) and insulin tolerance test (ITT)

An OGTT and ITT were performed during pregnancy at 16.5 days, immediately after delivery, and at 12 months

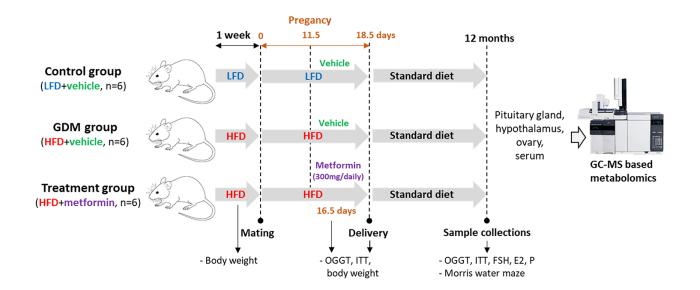


Figure 9. Overall experimental design.

of age. OGTT was performed by first fasting mice for six hours then administering glucose (2g/kg body weight) via gavage. Blood samples were collected at 0, 30, 60, 90, and 120 minutes from the tail vein, and blood glucose concentration was measured using a glucometer (Nova StatStrip Xpress; Nova Biomedical, Waltham, UK). To perform ITT, the mice were fasted for 6 hours and then insulin (2.5 U/kg body weight) was administered via an intraperitoneal injection. Blood samples were collected at 0, 15, 30, 60, and 120 minutes from the tail vein, and blood glucose concentration was measured using a glucometer.

Morris water maze experiment

The Morris water maze experiment was conducted in a constant room temperature swimming pool with a white inner wall which had a diameter of 120 cm, a height of 60 cm, and a data collection and analysis system. The entire pool was divided into four quadrants by four equally spaced points (indicated by different patterns). The hidden platform (10 cm in diameter and 24 cm in height) was placed 0.5 cm below the water surface. The animals had five continuous days of maze training where they were placed in the middle of each quadrant of the maze. During the training days, the mice were guided to the hidden platform if they failed to locate the platform within one minute and stayed on the hidden platform for 10 s. During the experiment day (sixth day), the hidden platform was eliminated, and mice were swimming in the maze for 60 s. This experiment was repeated four times a day.

Measurement of serum follicle-stimulating hormone (FSH), estrogen (E2), and progesterone (P)

Serum levels of FSH, E2, and P were analysed using ELISA kits (cat. no. CSB_E06871m; CSB_E05109m; CSB_E05104m). All analyses were performed in strict accordance with the manufacturer's protocols.

Metabolite extraction and derivatization

An approximately 10 mg sample of the ovary, hypothalamus, and pituitary gland from each animal were weighed at 4 °C. Metabolites were extracted from the samples using 2 mL of methanol/toluene 4:1 v/v containing solution two internal standards: nonadecanoic acid (20 µg/mL) and tridecanoic acid (20 µg/mL). 200 µL of acetyl chloride (Adamas Reagent Co.) was added to each sample, followed by a 1 min vortex. The tubes were then incubated at 100 °C for 1 h. After cooling in tap water, 5 mL of an aqueous solution of 6% potassium carbonate (Adamas Reagent Co.) was added into each tube. After vortexing for 10 s and centrifuging at 2000 rpm for 10 min at room temperature, the upper toluene phase was extracted for GC-MS analysis. Negative controls and quality control (QC) samples were prepared by replicating the sample preparation using empty tubes and a pooled biological sample respectively.

Gas chromatography-mass spectrometry (GCMS) analysis

The derivatized biospecimens were analysed using an Agilent 7890B Gas Chromatograph linked to an Agilent 5977A Mass Spectrometer. A RESTEK Rtx®-2330 column (90%) biscyanopropyl/10% phenylcyano propylpolysiloxane, 100 m, 0.25 mm ID, 0.2 um df) was installed to separate derivatised metabolites. The sample injection, inlet mode, oven temperature, and mass spectrometry parameters were operated according to Han et al. (2012) and Smart et al. (2010). The GC-MS chromatographic peaks were extracted, deconvoluted and identified using AMDIS and Agilent ChemStation. The peaks were identified based on two criteria; >85% match to the fatty acid library spectra and within a 30-second window of the library chromatographic retention time using the in-house lipid library and NIST library (https://www.nist.gov/nist-research-library). The relative concentrations of metabolites were quantified via our inhouse R based script (MassOmics) that uses the most abundant ion fragments within an appropriate retention time.

Data normalization and statistical analysis

Metabolite levels were first normalized either by the nonadecanoic acid or tridecanoic acid internal standard. determined based on their correlation with metabolites in the QC samples. Subsequently, median centering was performed using nine OC samples to correct for batch variation. Then, correction was applied using blank samples to remove contaminants and any carryover from identified metabolites. The tissue weights were accounted for, to eliminate the volume difference among samples. Prior to multivariate statistical analysis. the metabolite profiles of the HPO axis were log transformed and Pareto scaled since this combined scaling strategy resulted in a normal distribution of our dataset. Principal component analysis (PCA), orthogonal partial least squares discriminant analysis (OPLS-DA), variable importance projection (VIP) scores, one-way ANOVA, Turkey HSD, S-plots, and SUS-plots were performed using Metaboanalyst (https://www.metaboanalyst.ca/) and our in-house R scripts. A power analysis for the two-sample t-test was performed using the "pwr.t.test" command available in the R package "pwr2" [65]. The power analysis was performed based on the result of Zhu et al (2018), which utilized a HFD to establish a GDM mouse model,

to evaluate the effect of maternal obesity on cognitive development of the offspring. Our power calculation showed that a minimum sample size of six mice per group provided 80% power with an alpha value less than 0.05 in order to have 95% confidence of a true difference between HFD and LFD groups for both ITT and OGTT (Supplementary Figures 2 and 3).

Abbreviations

GDM: gestational diabetes mellitus; T2DM: type 2 diabetes mellitus; HFD: high-fat diet ; HPO axis: hypothalamus, pituitary gland, and ovary; GnRH: gonadotropin-releasing hormone; LFD: low-fat diet; OGTT: oral glucose tolerance test; ITT: insulin tolerance test; FSH: follicle-stimulating hormone; GC-MS: gas chromatography-mass spectrometry; PCA: principal component analysis; OPLS-DA: orthogonal partial least squares discriminant analysis; VIP: variable importance projection; AUC: area under the curve.

AUTHOR CONTRIBUTIONS

Y.Z., X.Z., and H.Z. designed and performed the experiments. Y.Z. and X.Y. built an animal model of GDM. Y.Z. Xue and X.C. completed the mass spectrometry experiment. Y.Z. and T-L.H. analyzed the data. Y.Z., T-L.H., H.Z., H.Q. and P.N.B. wrote the manuscript. All authors reviewed the manuscript. H.Z. is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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CONFLICTS OF INTEREST

No potential conflicts of interest relevant to this article were reported.

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REFERENCES

- Caissutti C, Berghella V. Scientific evidence for different options for GDM screening and management: controversies and review of the literature. Biomed Res Int. 2017; 2017:2746471. <u>https://doi.org/10.1155/2017/2746471</u> PMID:<u>28497042</u>
- Zhu W, Yang H, Wei Y, Wang Z, Li X, Wu H, Li N, Zhang M, Liu X, Zhang H, Wang Y, Niu J, Gan Y, et al. Comparing the diagnostic criteria for gestational diabetes mellitus of world health organization 2013 with 1999 in chinese population. Chin Med J (Engl). 2015; 128:125–27. <u>https://doi.org/10.4103/0366-6999.147858</u> PMID:25563325
- Kc K, Shakya S, Zhang H. Gestational diabetes mellitus and macrosomia: a literature review. Ann Nutr Metab. 2015 (Suppl 2); 66:14–20. <u>https://doi.org/10.1159/000371628</u> PMID:<u>26045324</u>
- Wendland EM, Duncan BB, Belizán JM, Vigo A, Schmidt MI. Gestational diabetes and pre-eclampsia: common antecedents? Arq Bras Endocrinol Metabol. 2008; 52:975–84. <u>https://doi.org/10.1590/s0004-27302008000600008</u> PMID:18820808
- Jahan M, Mahbuba S, Shaha S, Akhter H, Ahsan GU, Islam T, Kuehl TJ, Uddin MN. ID: 57: Pre-gestational diabetes and preeclampsia in Bangladeshi patients: A Retrospective Study. J Investig Med. 2016; 64:952–960. https://doi.org/10.1136/jim-2016-000120.99
- Feig DS, Zinman B, Wang X, Hux JE. Risk of development of diabetes mellitus after diagnosis of gestational diabetes. CMAJ. 2008; 179:229–34. <u>https://doi.org/10.1503/cmaj.080012</u> PMID:18663202
- Kodl CT, Seaquist ER. Cognitive dysfunction and diabetes mellitus. Endocr Rev. 2008; 29:494–511. <u>https://doi.org/10.1210/er.2007-0034</u> PMID:<u>18436709</u>
- Hiroyuki U, Takahiko K, Naoko K, Toshitaka U, Akio K, Takahisa S. Factors associated with cognitive decline in elderly diabetics. Dement Geriatr Cogn Dis Extra. 2011; 1:1–9. <u>https://doi.org/10.1159/000323188</u> PMID:22163228

9. Shalimova A, Graff B, Gasecki D, Wolf J, Sabisz A,

Szurowska E, Jodzio K, Narkiewicz K. Cognitive dysfunction in type 1 diabetes mellitus. J Clin Endocrinol Metab. 2019; 104:2239–49. https://doi.org/10.1210/jc.2018-01315 PMID:<u>30657922</u>

- Denver P, Gault VA, McClean PL. Sustained high-fat diet modulates inflammation, insulin signalling and cognition in mice and a modified xenin peptide ameliorates neuropathology in a chronic high-fat model. Diabetes Obes Metab. 2018; 20:1166–1175. <u>https://doi.org/10.1111/dom.13210</u> PMID:<u>29316242</u>
- 11. Zheng Y, Yang Y, Dong B, Zheng H, Lin X, Du Y, Li X, Zhao L, Gao H. Metabonomic profiles delineate potential role of glutamate-glutamine cycle in db/db mice with diabetes-associated cognitive decline. Mol Brain. 2016; 9:40. <u>https://doi.org/10.1186/s13041-016-0223-5</u> PMID:27090642
- Shideler SE, Gee NA, Chen J, Lasley BL. Estrogen and progesterone metabolites and follicle-stimulating hormone in the aged macaque female. Biol Reprod. 2001; 65:1718–25. <u>https://doi.org/10.1095/biolreprod65.6.1718</u>
 PMID:11717133
- Ding J, Strachan MW, Reynolds RM, Frier BM, Deary IJ, Fowkes FG, Lee AJ, McKnight J, Halpin P, Swa K, Price JF, and Edinburgh Type 2 Diabetes Study Investigators. Diabetic retinopathy and cognitive decline in older people with type 2 diabetes: the edinburgh type 2 diabetes study. Diabetes. 2010; 59:2883–89. https://doi.org/10.2337/db10-0752 PMID:20798334
- López-Grueso R, Borrás C, Gambini J, Viña J. [Aging and ovariectomy cause a decrease in brain glucose consumption in vivo in wistar rats]. Rev Esp Geriatr Gerontol. 2010; 45:136–40. <u>https://doi.org/10.1016/j.regg.2009.12.005</u> PMID:20206415
- Sakata A, Mogi M, Iwanami J, Tsukuda K, Min LJ, Jing F, Iwai M, Ito M, Horiuchi M. Female exhibited severe cognitive impairment in type 2 diabetes mellitus mice. Life Sci. 2010; 86:638–45. <u>https://doi.org/10.1016/j.lfs.2010.03.003</u> PMID:20226793
- 16. Ghazvini H, Khaksari M, Esmaeilpour K, Shabani M, Asadi-Shekaari M, Khodamoradi M, Sheibani V. Effects of treatment with estrogen and progesterone on the methamphetamine-induced cognitive impairment in ovariectomized rats. Neurosci Lett. 2016; 619:60–67. <u>https://doi.org/10.1016/j.neulet.2016.02.057</u> PMID:<u>26944454</u>
- 17. Hiroi R, Weyrich G, Koebele SV, Mennenga SE,

Talboom JS, Hewitt LT, Lavery CN, Mendoza P, Jordan A, Bimonte-Nelson HA. Benefits of hormone therapy estrogens depend on estrogen type: 17β -estradiol and conjugated equine estrogens have differential effects on cognitive, anxiety-like, and depressive-like behaviors and increase tryptophan hydroxylase-2 mRNA levels in dorsal raphe nucleus subregions. Front Neurosci. 2016; 10:517.

https://doi.org/10.3389/fnins.2016.00517 PMID:28008302

 Dumas J, Hancur-Bucci C, Naylor M, Sites C, Newhouse P. Estrogen treatment effects on anticholinergicinduced cognitive dysfunction in normal postmenopausal women. Neuropsychopharmacology. 2006; 31:2065–78. https://doi.org/10.1038/sj.npp.1301042

PMID:16482084

- Hara Y, Waters EM, McEwen BS, Morrison JH. Estrogen effects on cognitive and synaptic health over the lifecourse. Physiol Rev. 2015; 95:785–807. <u>https://doi.org/10.1152/physrev.00036.2014</u> PMID:<u>26109339</u>
- 20. Phillips SM, Sherwin BB. Effects of estrogen on memory function in surgically menopausal women. Psychoneuroendocrinology. 1992; 17:485–95. <u>https://doi.org/10.1016/0306-4530(92)90007-t</u> PMID:<u>1484915</u>
- Pallardo LF, Herranz L, Martin-Vaquero P, Garcia-Ingelmo T, Grande C, Jañez M. Impaired fasting glucose and impaired glucose tolerance in women with prior gestational diabetes are associated with a different cardiovascular profile. Diabetes Care. 2003; 26:2318–22.

https://doi.org/10.2337/diacare.26.8.2318 PMID:<u>12882855</u>

- 22. Buchanan TA, Xiang AH. Gestational diabetes mellitus. J Clin Invest. 2005; 115:485–91. <u>https://doi.org/10.1172/JCI24531</u> PMID:<u>15765129</u>
- 23. Catalano PM, Kirwan JP, Haugel-de Mouzon S, King J. Gestational diabetes and insulin resistance: role in short- and long-term implications for mother and fetus. J Nutr. 2003; 133:16745–83S. <u>https://doi.org/10.1093/jn/133.5.1674S</u> PMID:12730484
- Tang XW, Qin QX. miR-335-5p induces insulin resistance and pancreatic islet β-cell secretion in gestational diabetes mellitus mice through VASH1mediated TGF-β signaling pathway. J Cell Physiol. 2019; 234:6654–66. https://doi.org/10.1002/jcp.27406 PMID:30341900
- 25. Herath H, Herath R, Wickremasinghe R. Gestational

diabetes mellitus and risk of type 2 diabetes 10 years after the index pregnancy in Sri Lankan women-a community based retrospective cohort study. PLoS One. 2017; 12:e0179647.

https://doi.org/10.1371/journal.pone.0179647 PMID:<u>28644881</u>

- Pintana H, Pratchayasakul W, Sa-nguanmoo P, Pongkan W, Tawinvisan R, Chattipakorn N, Chattipakorn SC. Testosterone deprivation has neither additive nor synergistic effects with obesity on the cognitive impairment in orchiectomized and/or obese male rats. Metabolism. 2016; 65:54–67. https://doi.org/10.1016/j.metabol.2015.10.015 PMID:26773929
- Winocur G, Greenwood CE, Piroli GG, Grillo CA, Reznikov LR, Reagan LP, McEwen BS. Memory impairment in obese zucker rats: an investigation of cognitive function in an animal model of insulin resistance and obesity. Behav Neurosci. 2005; 119:1389–95. <u>https://doi.org/10.1037/0735-7044.119.5.1389</u> PMID:16300445
 - PIVIID:<u>16300445</u>
- Stranahan AM, Norman ED, Lee K, Cutler RG, Telljohann RS, Egan JM, Mattson MP. Diet-induced insulin resistance impairs hippocampal synaptic plasticity and cognition in middle-aged rats. Hippocampus. 2008; 18:1085–88. <u>https://doi.org/10.1002/hipo.20470</u> PMID:<u>18651634</u>
- 29. Kamal A, Ramakers GM, Gispen WH, Biessels GJ. Hyperinsulinemia in rats causes impairment of spatial memory and learning with defects in hippocampal synaptic plasticity by involvement of postsynaptic mechanisms. Exp Brain Res. 2013; 226:45–51. https://doi.org/10.1007/s00221-013-3409-4 PMID:23371746
- Hwang IK, Kim IY, Kim DW, Yoo KY, Kim YN, Yi SS, Won MH, Lee IS, Yoon YS, Seong JK. Strain-specific differences in cell proliferation and differentiation in the dentate gyrus of C57BL/6N and C3H/HeN mice fed a high fat diet. Brain Res. 2008; 1241:1–6. <u>https://doi.org/10.1016/j.brainres.2008.08.024</u> PMID:<u>18761331</u>
- Talbot K, Wang HY, Kazi H, Han LY, Bakshi KP, Stucky A, Fuino RL, Kawaguchi KR, Samoyedny AJ, Wilson RS, Arvanitakis Z, Schneider JA, Wolf BA, et al. Demonstrated brain insulin resistance in Alzheimer's disease patients is associated with IGF-1 resistance, IRS-1 dysregulation, and cognitive decline. J Clin Invest. 2012; 122:1316–38. <u>https://doi.org/10.1172/JCI59903</u> PMID:22476197
- 32. Cholerton B, Baker LD, Craft S. Insulin, cognition, and

dementia. Eur J Pharmacol. 2013; 719:170–79. https://doi.org/10.1016/j.ejphar.2013.08.008 PMID:24070815

- Yu F, Han W, Zhan G, Li S, Xiang S, Zhu B, Jiang X, Yang L, Luo A, Hua F, Yang C. Abnormal gut microbiota composition contributes to cognitive dysfunction in streptozotocin-induced diabetic mice. Aging (Albany NY). 2019; 11:3262–79. https://doi.org/10.18632/aging.101978 PMID:31123221
- 34. Yu F, Han W, Zhan G, Li S, Jiang X, Wang L, Xiang S, Zhu B, Yang L, Luo A, Hua F, Yang C. Abnormal gut microbiota composition contributes to the development of type 2 diabetes mellitus in db/db mice. Aging (Albany NY). 2019; 11:10454–67. https://doi.org/10.18632/aging.102469 PMID:31760385
- Johnson AL, Tilly JL. Arachidonic acid inhibits luteinizing hormone-stimulated progesterone production in hen granulosa cells. Biol Reprod. 1990; 42:458–64. <u>https://doi.org/10.1095/biolreprod42.3.458</u> PMID:<u>2111185</u>
- 36. Meln I, Wolff G, Gajek T, Koddebusch J, Lerch S, Harbrecht L, Hong W, Bayindir-Buchhalter I, Krunic D, Augustin HG, Vegiopoulos A. Dietary calories and lipids synergistically shape adipose tissue cellularity during postnatal growth. Mol Metab. 2019; 24:139–48. <u>https://doi.org/10.1016/j.molmet.2019.03.012</u> PMID:<u>31003943</u>
- Liu C, Chu I, Hwang S. Pentanoic acid, a novel protein synthesis stimulant for chinese hamster ovary (CHO) cells. J Biosci Bioeng. 2001; 91:71–75. <u>https://doi.org/10.1263/jbb.91.71</u> PMID:<u>16232949</u>
- Broekmans FJ, Soules MR, Fauser BC. Ovarian aging: mechanisms and clinical consequences. Endocr Rev. 2009; 30:465–93. <u>https://doi.org/10.1210/er.2009-0006</u> PMID:19589949
- 39. Zang M, Zuccollo A, Hou X, Nagata D, Walsh K, Herscovitz H, Brecher P, Ruderman NB, Cohen RA. AMP-activated protein kinase is required for the lipidlowering effect of metformin in insulin-resistant human HepG2 cells. J Biol Chem. 2004; 279:47898–905. <u>https://doi.org/10.1074/jbc.M408149200</u> PMID:15371448
- 40. Gallagher EJ, LeRoith D. Diabetes, cancer, and metformin: connections of metabolism and cell proliferation. Ann N Y Acad Sci. 2011; 1243:54–68. <u>https://doi.org/10.1111/j.1749-6632.2011.06285.x</u> PMID:<u>22211893</u>
- 41. Rogacka D, Audzeyenka I, Rychłowski M, Rachubik P, Szrejder M, Angielski S, Piwkowska A. Metformin

overcomes high glucose-induced insulin resistance of podocytes by pleiotropic effects on SIRT1 and AMPK. Biochim Biophys Acta Mol Basis Dis. 2018; 1864:115–25.

https://doi.org/10.1016/j.bbadis.2017.10.014 PMID:29032153

- Pei L, Yang J, Du J, Liu H, Ao N, Zhang Y. Downregulation of chemerin and alleviation of endoplasmic reticulum stress by metformin in adipose tissue of rats. Diabetes Res Clin Pract. 2012; 97:267–75. <u>https://doi.org/10.1016/j.diabres.2012.02.023</u> PMID:<u>22445233</u>
- Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, Prifti E, Vieira-Silva S, Gudmundsdottir V, Pedersen HK, Arumugam M, Kristiansen K, Voigt AY, et al, and MetaHIT consortium. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. Nature. 2015; 528:262–66.

https://doi.org/10.1038/nature15766 PMID:26633628

- 44. Feng Y, Yang H. Metformin a potentially effective drug for gestational diabetes mellitus: a systematic review and meta-analysis. J Matern Fetal Neonatal Med. 2017; 30:1874–1881. https://doi.org/10.1080/14767058.2016.1228061 PMID:27549367
- 45. Naderpoor N, Shorakae S, de Courten B, Misso ML, Moran LJ, Teede HJ. Metformin and lifestyle modification in polycystic ovary syndrome: systematic review and meta-analysis. Hum Reprod Update. 2015; 21:560–74. https://doi.org/10.1093/humupd/dmv025

PMID:26060208

- Wu MS, Johnston P, Sheu WH, Hollenbeck CB, Jeng CY, Goldfine ID, Chen YD, Reaven GM. Effect of metformin on carbohydrate and lipoprotein metabolism in NIDDM patients. Diabetes Care. 1990; 13:1–8. <u>https://doi.org/10.2337/diacare.13.1.1</u> PMID:<u>2404714</u>
- 47. Lin Y, Wang K, Ma C, Wang X, Gong Z, Zhang R, Zang D, Cheng Y. Evaluation of Metformin on Cognitive Improvement in Patients With Non-dementia Vascular Cognitive Impairment and Abnormal Glucose Metabolism. Front Aging Neurosci. 2018; 10:227. https://doi.org/10.3389/fnagi.2018.00227 PMID:30100873
- Ekblad LL, Rinne JO, Puukka P, Laine H, Ahtiluoto S, Sulkava R, Viitanen M, Jula A. Insulin Resistance Predicts Cognitive Decline: An 11-Year Follow-up of a Nationally Representative Adult Population Sample. Diabetes care. 2017; 40:751–758. <u>https://doi.org/10.2337/dc16-2001</u>

PMID:28381479

- 49. Hardie DG. AMP-activated protein kinase: maintaining energy homeostasis at the cellular and whole-body levels. Annu Rev Nutr. 2014; 34:31–55. <u>https://doi.org/10.1146/annurev-nutr-071812-161148</u> PMID:<u>24850385</u>
- Cantó C, Gerhart-Hines Z, Feige JN, Lagouge M, Noriega L, Milne JC, Elliott PJ, Puigserver P, Auwerx J. AMPK regulates energy expenditure by modulating NAD+ metabolism and SIRT1 activity. Nature. 2009; 458:1056–60. https://doi.org/10.1038/nature07813

PMID:19262508

- 51. Ou Z, Kong X, Sun X, He X, Zhang L, Gong Z, Huang J, Xu B, Long D, Li J, Li Q, Xu L, Xuan A. Metformin treatment prevents amyloid plaque deposition and memory impairment in APP/PS1 mice. Brain Behav Immun. 2018; 69:351–63. <u>https://doi.org/10.1016/j.bbi.2017.12.009</u> PMID:29253574
- 52. Chaudhari K, Reynolds CD, Yang SH. Metformin and cognition from the perspectives of sex, age, and disease. Geroscience. 2020; 42:97–116. <u>https://doi.org/10.1007/s11357-019-00146-3</u> PMID:31897861
- 53. Fatt M, Hsu K, He L, Wondisford F, Miller FD, Kaplan DR, Wang J. Metformin acts on two different molecular pathways to enhance adult neural precursor proliferation/self-renewal and differentiation. Stem Cell Reports. 2015; 5:988–95. https://doi.org/10.1016/j.stemcr.2015.10.014 PMID:26677765
- 54. Wang J, Gallagher D, DeVito LM, Cancino GI, Tsui D, He L, Keller GM, Frankland PW, Kaplan DR, Miller FD. Metformin activates an atypical PKC-CBP pathway to promote neurogenesis and enhance spatial memory formation. Cell Stem Cell. 2012; 11:23–35. <u>https://doi.org/10.1016/j.stem.2012.03.016</u> PMID:22770240
- 55. Potts MB, Lim DA. An old drug for new ideas: metformin promotes adult neurogenesis and spatial memory formation. Cell Stem Cell. 2012; 11:5–6. <u>https://doi.org/10.1016/j.stem.2012.06.003</u> PMID:<u>22770236</u>
- 56. Yan Q, Han C, Wang G, Waddington JL, Zheng L, Zhen X. Activation of AMPK/mTORC1-mediated autophagy by metformin reverses Clk1 deficiency-sensitized dopaminergic neuronal death. Mol Pharmacol. 2017; 92:640–52. https://doi.org/10.1124/mol.117.109512

PMID:<u>29025968</u>

57. Song YM, Lee YH, Kim JW, Ham DS, Kang ES, Cha BS,

Lee HC, Lee BW. Metformin alleviates hepatosteatosis by restoring SIRT1-mediated autophagy induction via an AMP-activated protein kinase-independent pathway. Autophagy. 2015; 11:46–59. https://doi.org/10.4161/15548627.2014.984271 PMID:25484077

- 58. Zhu XC, Jiang T, Zhang QQ, Cao L, Tan MS, Wang HF, Ding ZZ, Tan L, Yu JT. Chronic metformin preconditioning provides neuroprotection via suppression of NF-κB-mediated inflammatory pathway in rats with permanent cerebral ischemia. Mol Neurobiol. 2015; 52:375–85. <u>https://doi.org/10.1007/s12035-014-8866-7</u> PMID:25172433
- Li W, Chaudhari K, Shetty R, Winters A, Gao X, Hu Z, Ge WP, Sumien N, Forster M, Liu R, Yang SH. Metformin alters locomotor and cognitive function and brain metabolism in normoglycemic mice. Aging Dis. 2019; 10:949–63. https://doi.org/10.14336/AD.2019.0120

PMID:31595194

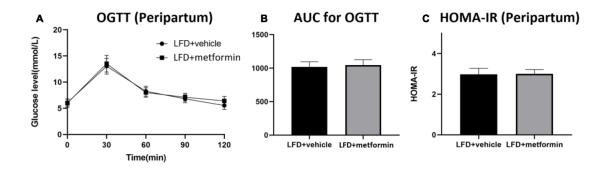
 Hohnholt MC, Blumrich EM, Waagepetersen HS, Dringen R. The antidiabetic drug metformin decreases mitochondrial respiration and tricarboxylic acid cycle activity in cultured primary rat astrocytes. J Neurosci Res. 2017; 95:2307–20. https://doi.org/10.1002/jnr.24050

PMID:28316081

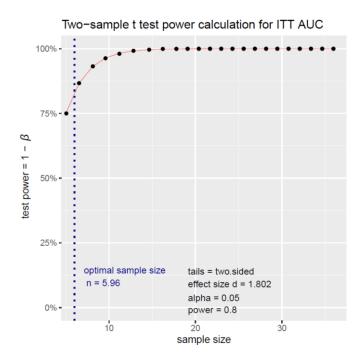
- 61. Westhaus A, Blumrich EM, Dringen R. The antidiabetic drug metformin stimulates glycolytic lactate production in cultured primary rat astrocytes. Neurochem Res. 2017; 42:294–305. <u>https://doi.org/10.1007/s11064-015-1733-8</u> PMID:<u>26433380</u>
- Viollet B, Guigas B, Sanz Garcia N, Leclerc J, Foretz M, Andreelli F. Cellular and molecular mechanisms of metformin: an overview. Clin Sci (Lond). 2012; 122:253–70. <u>https://doi.org/10.1042/CS20110386</u> PMID:<u>22117616</u>
- 63. Gong L, Goswami S, Giacomini KM, Altman RB, Klein TE. Metformin pathways: pharmacokinetics and pharmacodynamics. Pharmacogenet Genomics. 2012; 22:820–27. https://doi.org/10.1097/FPC.0b013e3283559b22 PMID:22722338
- 64. Kintiraki E, Goulis DG. Gestational diabetes mellitus: multi-disciplinary treatment approaches. Metabolism. 2018; 86:91–101. https://doi.org/10.1016/j.metabol.2018.03.025 PMID:29627447
- 65. Team RC. (2018). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

SUPPLEMENTARYMATERIALS

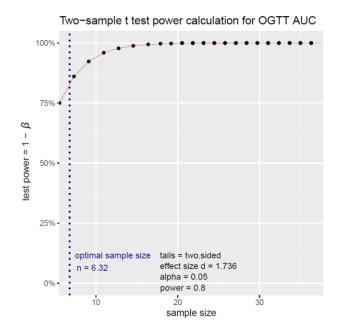
Supplementary Figures



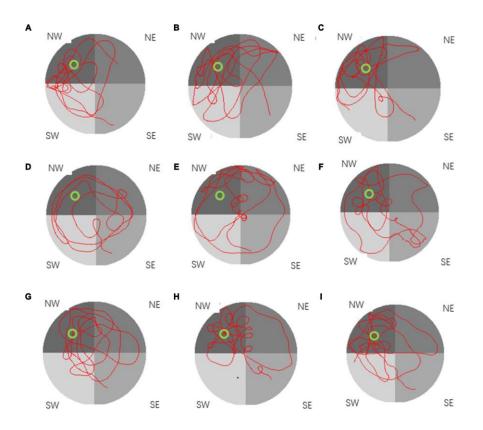
Supplementary Figure 1. Oral glucose tolerance test (OGTT) and homeostasis model assessment of insulin resistance (HOMA-IR) during pregnancy. OGTT curves between the LFD+vehicle and LFD+metformin treatment groups (A) and comparison of areas under the curve (AUC) at 16.5 gestational days (B). HOMA-IR results between the LFD+vehicle and LFD+metformin treatment groups at 18.5 gestational days (C).



Supplementary Figure 2. Power analysis of ITT between HFD and LFD mice at peripartum, based on data from Zhu et al (2018). The curve displays the relationship between changes of power (y-axis) along with changes in sample size (x-axis). The blue vertical line indicates sample size required to have 80% power with an alpha value less than 0.05 for a given mean difference between two groups using a two-sided Student's T test. n= number. AUC = area under the curve.



Supplementary Figure 3. Power analysis of OGTT between HFD and LFD mice at peripartum, based on data from Zhu et al (2018). The curve displays the relationship between changes of power (y-axis) along with changes in sample size (x-axis). The blue vertical line indicates sample size required to have 80% power with an alpha value less than 0.05 for a given mean difference between two groups using a two-sided Student's T test. n= number. AUC = area under the curve.



Supplementary Figure 4. Representative swimming tracks in the Morris water maze. Swimming tracks of mice in the control group (A–C). Swimming tracks of mice in the GDM group (D–F). Swimming tracks of mice in the treatment group (G–I). The circle represents the area of the water maze, divided into four quadrants, the green circle represents the location of the original platform, and the red line represents the swimming route of the mouse.

Supplementary Table

Supplementary Table 1. Morris water maze results.

	Control (n=7)	GDM (n=6)	Treatment (n=6)	F	p-value
Time (s)	$4.0{\pm}1.1^{*}$	1.3±1.5	2.3±0.8	7.903	0.005
Time (s in %)	$54.0{\pm}7.3^{*}$	35.3±14.7	$54.1{\pm}8.7^{*}$	6.100	0.012
Distance (cm in %)	$51.2\pm8.3^{**}$	34.8±13.3	$51.3 \pm 8.2^{**}$	5.209	0.019

Time (s in %) is calculated as swimming time in the quadrant of the original platform/total swimming time*100. Distance (cm in %) is calculated as swimming distance in the quadrant of the original platform/total swimming distance*100. F represents the statistic of one-way Anova, *p-value<0.05 versus HFD+Vehicle, **p-value<0.01 versus HFD+Vehicle