

MiR-17 and miR-93 Promote Tumor Progression by Targeting p21 in Patients with Chordoma

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Objective: MicroRNAs have been implicated in the progression of various cancers. However, the role of microRNAs in chordoma remains to be further elucidated. Here, we purposed to character the role of two microRNAs, miR-17 and miR-93, and their potential mechanisms in chordoma.

Methods: The expression and prognostic value of miR-17 and miR-93 were assessed by the quantitative real-time polymerase chain reaction, Kaplan–Meier survival curve, and Cox regression analysis. The effects of miR-17/93 mimics on chordoma cell proliferation, colony formation, and invasion were analyzed by CCK-8 assay, colony formation assay, and transwell assay. The downstream target of miR-17/93 was further explored via luciferase reporter assay.

Results: High expression of miR-17/93 was identified in chordoma tissues, and was associated with poor prognosis. Overexpression of miR-17/93 contributed to cell proliferation, colony formation, and invasion. Mechanistically, we demonstrated that miR-17/93 directly targeted p21 and decreased the expression of p21. Besides, the rescue assay further confirmed the essential role of the miR-17/93-p21 axis in chordoma.

Conclusion: Our results revealed the potential oncogenic effect of the miR-17/93 on chordoma progression, and suggested that the miR-17/93-p21 axis served as a promising therapeutic target in chordoma.

Keywords: microRNA, miR-17, miR-93, p21, chordoma

Introduction

Chordoma, identified as a rare and locally aggressive mesenchymal neoplasm, is developed from residual notochord tissues.^{1–3} Chordoma dominantly occurs at the midline skeleton axis, commonly skull base and sacrum with an incidence rate of 0.84 per million.³ Currently, the optimal treatment of chordoma is maximal surgical resection with optional proton/photon-beam radiotherapy for its nature of resistance to traditional radio/chemotherapy.^{2,4,5} However, due to the difficulty of radical resection and high locoregional recurrent rate, the chordoma patients harbor a dismal prognosis, especially for recurrent patients.³ Thus, a deeper exploration of chordoma's oncogenesis and more effective treatment options are needed to prolong patient survival in chordoma.

MicorRNAs (miRNAs), a cluster of small endogenous non-coding RNAs that diffusely existed in various species including animals and plants, are generally composed of 18–23 nucleotides and post-transcriptionally regulated the target gene expression via binding to the mRNA.^{6–8} Studies involving the role of miRNAs have attracted prominent attention in recent decades and miRNAs have been recognized to

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play an essential role as carcinogen or tumor suppressors in multiple tumors, such as acute myeloid leukemia, gastric cancer, colorectal carcinoma, and prostate cancer.^{9–12} Additionally, studies have demonstrated that miRNAs participate in various biological and tumor-associated processes, including cell growth, cell differentiation, invasion potential, angiogenesis, epithelial-mesenchymal transition and metastasis, and many aberrantly expressed miRNAs have shown the effective diagnostic value and prognostic performance in various cancers.¹³

Till now, the prognostic role and functions of miRNAs in chordoma are poorly understood. Recent studies have indicated that several miRNAs may contribute to oncogenesis and act as a valuable prognostic indicator in chordoma, such as miR-1, miR-1237-3p, miR-155.^{14–16} However, the role of miR-17 and miR-93 in chordoma remains unclear. MiR-17, encoded by miR-17-92 cluster, is highly expressed in tumors and involved in tumor progression, and high expression of which is associated with adverse outcome in T-cell lymphoblastic lymphoma and Burkitt lymphoma.^{17,18} For miR-93 encoded by miR-106b-25 cluster, a paralog of miR-17-92 cluster, increasing researches have identified the oncogenetic role of miR-93 and high expression of miR-93 acts as a risk variable for survival in hepatocellular carcinoma and gastric cancer.^{19,20} Additionally, miR-17 and miR-93 have been identified to directly target p21 in various cancers.^{21–24}

In the current study, we investigated the mRNA expression of miR-17, miR-93, and p21 in chordoma, and their clinical impact, survival performance were further analyzed. We also explored the functions of miR-17/93 and p21 in chordoma cells. Our findings identified that the miR-17/93-p21 axis act as a novel prognostic indicator and therapeutic target for chordoma.

Materials and Methods

Tissue Samples

After informed consent signature and ethics approval, fresh tumor tissues were collected from 30 patients who received surgical treatment at Tangshan People's Hospital between 2008 April and 2016 September. Patients with a prior history of radiotherapy/chemotherapy were excluded. All included patients were confirmed as chordoma by the pathologist, and 5 nucleus pulposus tissues were used as a control as the previous studies indicated.^{25–27} All tissues were immediately stored in the liquid nitrogen until RNA extraction.

Of the 30 chordoma patients, 21 patients were male and 9 female patients. Ten patients were skull base chordoma and the remaining 20 patients harbor spinal chordoma (sacrum, 16 patients; thoracic or cervical vertebra, 4 patients), and the ages ranged from 26 to 71 years old. All patients were histopathologically confirmed as conventional types of chordoma.² The detailed clinical characteristics of chordoma patients are shown in [Table 1](#) and [Supplementary Table 1](#). The follow-up information from each patient was collected for survival analysis and the median follow-up time was 50 months (4–96 months). During the follow-up, 13 (43.3%) patients died and the overall survival (OS) was defined as the interval from the first surgery to patient death.

Cell Culture and Transfection

Chordoma cell lines MUG-Chor1 (CRL-3219) and UM-Chor1 (CRL-3270) were acquired from American Type Culture Collection (ATCC), and the chordoma cells were cultured in IMDM (ATCC, USA) and RPMI-1640 (ATCC, USA) with a ratio of 4:1 containing 10% FBS (Gibco, USA) and 1% penicillin/streptomycin (Invitrogen, USA). Cells were routinely incubated under 5% CO₂, 37°C environment in a humidified incubator.

MiR-17 mimics, miR-93 mimics and miRNA mimics negative control (miR-NC) were purchased from Ribibio (Guangzhou, China). To overexpress p21, the pcDNA3.1-p21 overexpressed plasmid (p21) and control vector (Vector) were bought from Genepharma (Shanghai, China). 50 nM miRNA mimics or 2 µg of plasmid were transfected into MUG-Chor1 and UM-chor1 cells using Lipofectamine 3000 (Invitrogen, USA) according to the manufacturer's instruction. 48 hours after treatment, cells were gathered for RNA or protein extraction.

RNA Extraction and Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

Expression of miR-17/93 and p21 in chordoma tissues and cell lines were evaluated by qRT-PCR. Total RNA was extracted using TRIzol reagent (Invitrogen, USA) based on the protocol and the RNA quantify and quality was further examined. 1st strand cDNA was acquired via a HiScript II cDNA kit (Vazyme, Nanjing, China). PCR was carried out via the ABI 7500 PCR system (Applied Biosystems, USA) using SYBR-Green PCR mix (Vazyme, China). U6 small nuclear RNA and GAPDH were used to normalize the expression of miR-17/93 and p21, respectively. Primers of

Table 1 Correlation Between Expression of miR-17, miR-93 and P21 with Clinicopathological Features

Variables	Number	miR-17 Expression			miR-93 Expression			p21 Expression		
		High	Low	P value	High	Low	P value	High	Low	P value
Age (years)				0.060			0.263			0.710
>50	12	9	3		8	4		5	7	
≤50	18	6	12		7	11		10	8	
Sex				1.00			1.00			0.427
Male	21	10	11		11	10		9	12	
Female	9	5	4		4	5		6	3	
Tumor location				1.00			0.439			0.121
Skull base	10	5	5		4	6		7	3	
Spine	20	10	10		11	9		8	12	
Tumor size				0.456			0.456			0.264
≤3 cm	12	5	7		5	7		8	4	
>3 cm	18	10	8		10	8		7	11	
Primary tumor				0.035*			0.214			0.682
Yes	22	8	14		9	13		12	10	
No	8	7	1		6	2		3	5	
Total resection				1.00			0.651			0.651
Yes	6	3	3		2	4		2	4	
No	24	12	12		13	11		13	11	

Note: *Indicates statistical significance.

miRNA were purchased from Ribio company (Guangzhou, China) and the sequences for p21/GAPDH primers (BGI Genomics, Beijing, China) were as follows: p21, 5'-TCCAGCGACCTTCCTCATCCA-3' and 5'-TCCATAGCC TCTACTGCCACCA; GAPDH, 5'-AGGTCGGAGTCA ACGGATTTG-3' and 5'-GTGATGGCATGGACTGTGGT-3'. The expression of the target gene was calculated as $2^{-\Delta\Delta Ct}$.

CCK-8 Assay

Cell proliferation was evaluated by the CCK-8 assay. MUG-Chor1 (3×10^3 cells per well) and UM-Chor1 (2×10^3 cells per well) in medium without antibiotics were seeded into 96-well plate, and chordoma cells were then transfected with miR-17/93 mimics or miR-NC, p21/control vector triplicate. At each point (0, 24, 48, 72 h) after transfection, 10 μ L CCK-8 (Dojindo, Japan) was applied in wells, following the plates were incubated at 37°C for 2 h. Cells' absorbances at 450 nm were assessed using the microplate reader.

Colony Formation Assay

Six hundred MUG-Chor1 cells and 1×10^3 UM-Chor1 cells transfected with different groups (miR-NC, miR-17/93 mimics, with/without vector or p21) were plated in the

6-well plates. After a two-week incubation, cells were fixed via 4% methanol and subsequently stained using 0.1% crystal violet for 10 min.

Invasion Assay

Cell invasion activity of MUG-Chor1 and UM-chor1 was determined by transwell assay. 8 μ m 24-well transwell chamber (Corning, USA) with matrigel (BD, Biosciences, USA) was used according to the instruction. Briefly, transfected MUG-Chor1 (5×10^4 cells per well) and UM-Chor1 (2×10^4 cells per well) cells were plated onto the upper chamber with 100 μ L serum-free and antibiotic-free culture. For the bottom chamber, 500 μ L medium with 10% of FBS was applied. The transwell chamber was incubated for 48h at 37°C, and cells were fixed with methanol. After removing the cells on the upper chamber with the cotton swab, the cells on the inferior chamber were stained using crystal violet (0.1%) for a quarter, photographed, and countered with a microscope (Zeiss, Germany).

Western Blot Analysis

Total protein from chordoma cells was regularly lysed with RIPA buffer (Beyotime, China), and the concentrations were

quantified by BCA assay (Applygen, China). 30 μg protein was separated by 10% SDS-PAGE and then moved to the PVDF member (Sigma-Aldrich, USA) according to standard instruction. The anti-p21 antibody (1:1000, ab109520) and GAPDH antibody (1:8000, ab128915) were purchased from Abcam (UK). A chemiluminescence assay from Sigma-Aldrich (USA) was used for signal detection.

Luciferase Reporter Assay

The renilla/firefly luciferase reporter plasmid (psiCHECK2 vector, Promega) with 3'-UTR wild or mutant types of p21, and miR-17/93 mimics or miR-NC were routinely transfected in chordoma cells according to the protocol. 48h after transfection, cells were collected for assessment of firefly and renilla luciferase activities with the help of a dual-luciferase reporter assay system (Promega, Madison, USA). Then, ratios of firefly to renilla were calculated, and the average of three independent experiments was recognized as final luciferase activity.

Statistical Analysis

All statistical analyses were performed by SPSS 21.0 (IBM, NY, USA). The expression of miR-17/93 and p21 was separated into two subgroups (high level or low level) using the median value, respectively. The relationships between miR-17, miR-93, p21, and clinical features were analyzed via the chi-square test. To find potential correlations, Pearson's correlation analysis was used. Student's *t*-test was applied to compare data between two subgroups and for data with multi

groups, one-way ANOVA was applied for different analyses. Kaplan Meier analysis and Log rank test were used for OS analysis. Univariable and multivariable Cox regression analyses were performed for recognizing potential independent prognostic variables. A *p*-value of less than 0.05 was considered to be of statistical significance.

Results

Upregulation of MiR-17/93 and Downregulation of P21 Was Correlated with Poor Outcome in Chordoma Patients

We first evaluated the expression of miR-17/93 and p21 in 30 chordoma tissues and 5 normal tissues using qRT-PCR and our finding indicated that the expression of miR-17 was notably increased in chordoma tissues, compared with that in nucleus pulposus ($p < 0.01$, Figure 1A). Similarly, the level of miR-93 was also higher in chordoma tissues ($p < 0.05$, Figure 1B). In contrast, the expression of p21 was lower in chordoma tissues ($p < 0.01$, Figure 1C).

We then divided the 30 chordoma patients into high/low expression groups using the median values of miR-17/93 and p21 and further analyzed the relationships between their expressions with clinical parameters and survival in chordoma. We found that the expression of miR-17 was significantly correlated with primary/recurrent tumors ($p = 0.035$) (Table 1). Besides, there was a trend that miR-17 expression positively correlated with

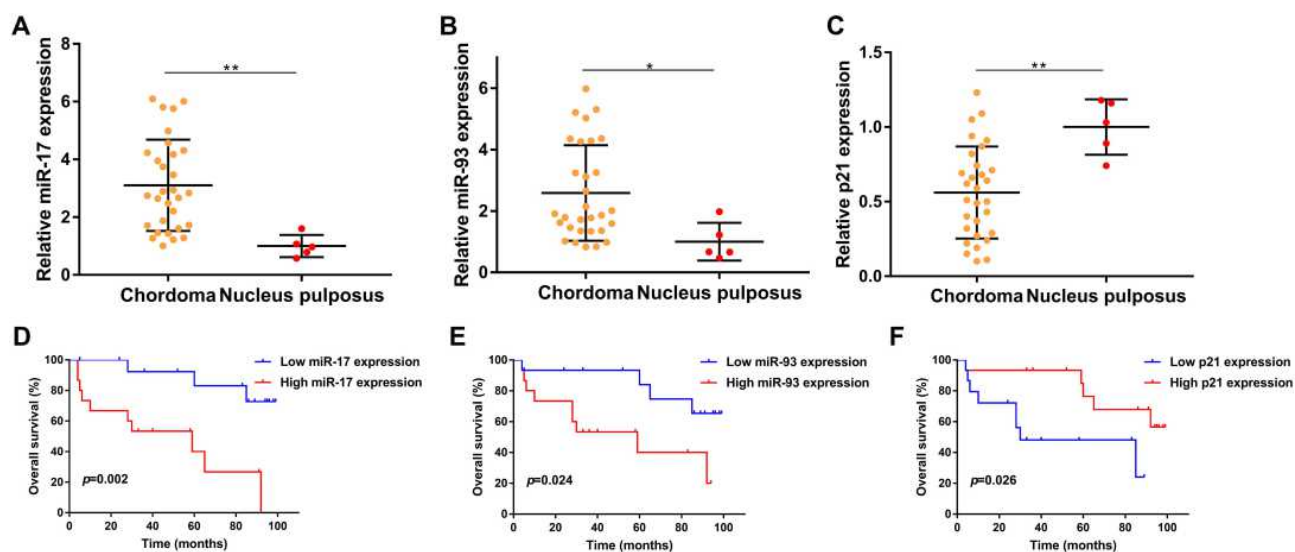


Figure 1 Prognostic value of miR-17, miR-93, and p21 in chordoma. (A–C) Expression of miR-17, miR-93, and p21 in chordoma tissues compared to nucleus pulposus tissues. (D–F) Kaplan-Meier survival analysis of miR-17, miR-93, and p21 in chordoma. * $p < 0.05$, ** $p < 0.01$.

patient age ($p=0.060$). However, no other correlations with clinical features including gender, location, tumor size, and types of resection were observed (Table 1). Kaplan Meier analysis indicated that patients with higher miR-17 expression had a shorter OS time than patients with lower miR-17 expression ($p=0.002$, Figure 1D). Moreover, higher miR-93 expression was also associated with poor overall survival ($p=0.024$, Figure 1E), while patients with lower p21 expression harbored a poor outcome than patients with a higher p21 level ($p=0.026$, Figure 1F).

The univariate Cox analysis revealed that expression of miR-17 ($p=0.006$), expression of miR-93 ($p=0.035$), p21 expression ($p=0.037$), types of resection ($p=0.048$), and tumor progression ($p<0.001$) were associated with survival of chordoma patients, while other clinical parameters showed no prognostic value. More importantly, multivariate Cox analysis found that the expression of miR-17 ($p=0.048$), tumor progression ($p<0.001$) were independent predicted parameters for OS of chordoma patients (Table 2).

MiR-17 and MiR-93 Promoted the Malignant Behavior of Chordoma Cells

Considering the prognostic roles of miR-17 and 93 in chordoma, we next investigated their potential function via overexpressing miR-17/93 in MUG-Chor1 and UM-Chor1 chordoma cells using miR-17/93 mimics. Using qRT-PCR, we confirmed the overexpression of miR-17/93 in miR-17/93 transfected cells (Figure 2A and B). CCK8 assay revealed that a remarked increase in cell viability of miR-17/93 group cells compared to the miR-NC cells (Figure 2C and D). Besides, colony formation assay measuring the cloning

ability of MUG-Chor1 and UM-Chor1 cells revealed a significant increase of cloning cells in miR-17/93 transfected cells compared to the miR-NC group cells (Figure 2E and F). Moreover, invasion assay showed that invasion abilities of MUG-Chor1 and UM-Chor1 cells were increased after miR-17/93 overexpression (Figure 2G and H). These data suggested that the oncogenic roles of miR-17 and miR-93 in chordoma cells in vitro.

MiR-17 and MiR-93 Directly Targeted P21 in Chordoma Cells

Bioinformatics analysis of starBase v2.0 and TargetScan showed p21 as a hypothetical target of miR-17 and miR-93 (Figure 3A),²⁸ and previous research demonstrated that miR-17/93 could function via directly targeting p21,^{22,24} thus, we explored whether p21 as a target of miR-17/93 in chordoma. As shown in Figure 3B, overexpression of miR-17 or miR-93 could decrease the mRNA expression of p21 in chordoma cells. Accordingly, the protein expression of p21 was also decreased (Figure 3C). Moreover, luciferase reporter plasmids containing wide type 3' UTR (p21-wt) and mutant 3' UTR (p21-mut) sequences of p21 were constructed, and chordoma cells were co-transfected with miR-17/93 mimics or miR-NC, and p21-wt or p21-mut plasmid. Co-transfection of miR-17/miR-93 and p21-wt showed decreased luciferase activity, while no significant decrease in luciferase activity was observed when co-transfection of miR-17/miR-93 mimics and p21-mut (Figure 3D and E). Besides, there were significant negative correlations between miR-17 ($r=-0.49$, $p=0.007$) or miR-93 ($r=-0.53$, $p=0.003$) expression and mRNA level of p21 in chordoma tissues (Figure 3F and G).

Table 2 Univariate and Multivariate Cox Analysis of OS in Chordoma

Variables	Univariate Analysis		Multivariate Analysis	
	HR (95% CI)	P value	HR (95% CI)	P value
Age (>50 vs ≤50 years)	2.131(0.563–8.063)	0.265		
Sex (female vs male)	0.850 (0.260–2.785)	0.789		
Tumor location (cranial vs spinal)	1.131(0.347–3.686)	0.838		
Tumor size (>3 vs ≤3 cm)	1.467(0.473–4.554)	0.507		
Primary tumor (yes vs no)	0.665(0.183–2.423)	0.537		
Total resection (no vs yes)	3.406(1.010–11.491)	0.048*	—	0.110
Progression (yes vs no)	9.549(3.482–26.183)	<0.001*	6.850(2.451–19.145)	<0.001*
miR-17 (high vs low)	6.456 (1.708–24.395)	0.006*	5.097 (1.014–25.611)	0.048*
miR-93 (high vs low)	3.690 (1.097–12.419)	0.035*	—	0.311
p21 expression (low vs high)	3.723(1.081–12.825)	0.037*	—	0.245

Note: *Indicates statistical significance.

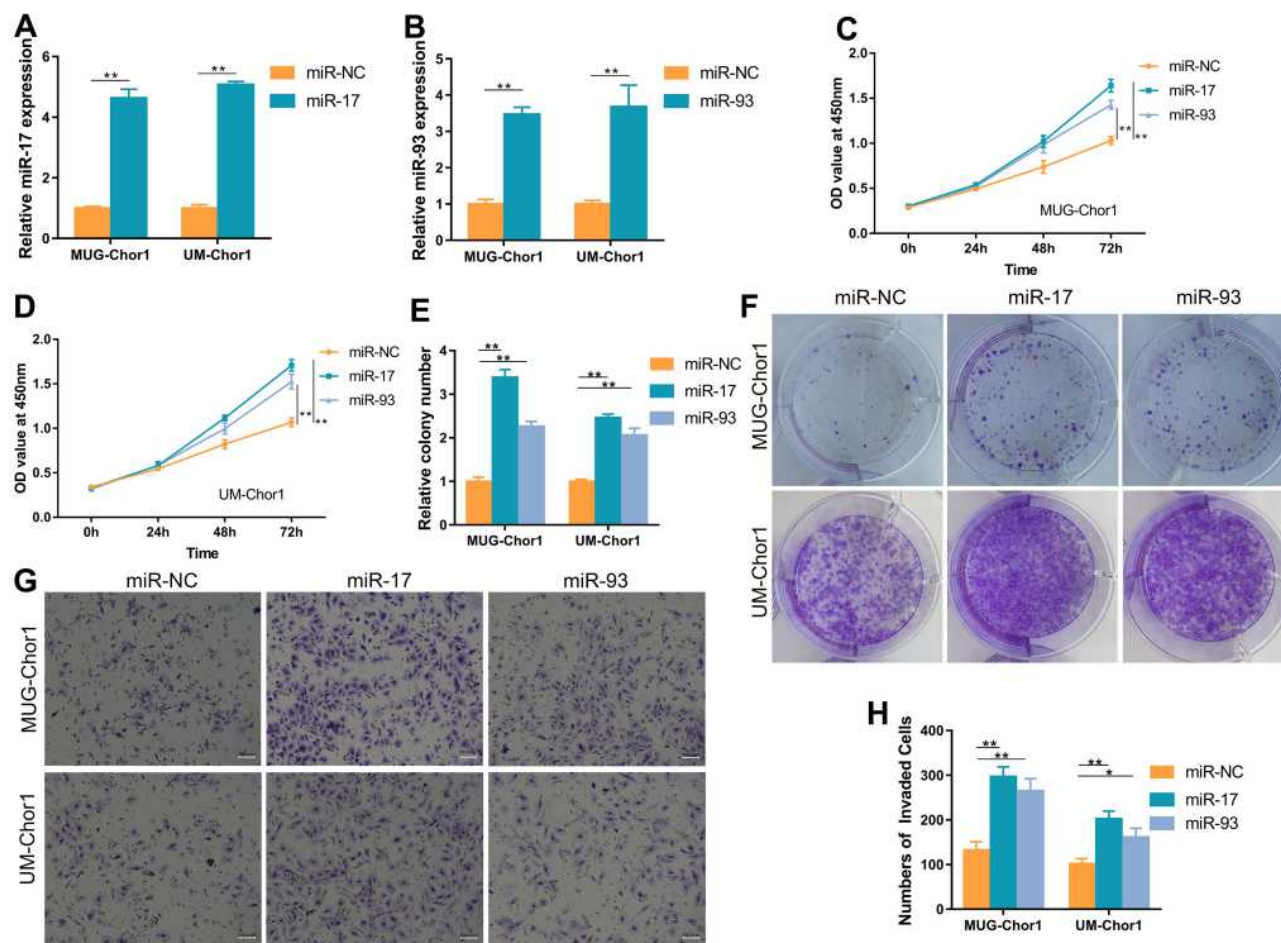


Figure 2 Overexpression of miR-17/miR-93 promotes MUG-Chor1 and UM-Chor1 cell proliferation, colony formation, and invasion in vitro. (A and B) Expression of miR-17/miR-93 was obviously increased by miRNA mimics. (C and D) CCK-8 assay showed that miR-17/miR-93 promotes cell proliferation compared to miR-NC. (E and F) Colony formation assay revealed that miR-17/miR-93 contribute to chordoma cell colony formation. (G and H) Transwell invasion assay indicated the invaded ability of chordoma cells increased after miR-17/miR-93 treated. Magnification, 100X. * $p < 0.05$, ** $p < 0.01$.

Overexpression of P21 Partially Abolished the Effect of MiR-17/93 in Chordoma Cells

We then performed the rescue assays to confirm the miR-17/93-p21 axis. MUG-Chor1 and UM-Chor1 cells transfected with miR-17/93 mimics were further co-transfected with p21 and vector plasmid. The mRNA level and protein expression of p21 were increased after transfected with p21 over-expressed plasmid (Figure 4A and B). The effect of miR-17/93 mimics on proliferation ability (Figure 4C) and the numbers of cloning cells (Figure 4D and E) was reversed by p21 overexpression. Moreover, the increased invasion ability of MUG-Chor1 and UM-Chor1 cells via the miR-17/93 mimics was recovered after the co-transfection of p21 (Figure 4F and G). Taken together, our findings revealed that miR-17/93 contributed to tumor progression via directly targeting p21 in chordoma.

Discussion

The essential role of miRNA in various diseases and tumor progression has been demonstrated in numerous studies of recent decades.^{8,13} In chordoma, several potential oncogenic/tumor suppressor miRNAs have been identified and showed potent predictive performance for outcome.^{14–16,29} However, to our knowledge, few studies analyze the role and function of miR-17/93 in chordoma. In the current study, we for the first time character the levels of miR-17/93 in chordoma patients and we found a significant up-regulation of miR-17/93 in chordoma. Further survival analysis showed that high expression of miR-17 or miR-93 was associated with unfavorable prognosis in chordoma, while low level of p21 was correlated with the shorter OS, and sequent multivariate Cox analysis identified that miR-17 could act as an independent indicator for patient outcome prediction. Functionally, the upregulation

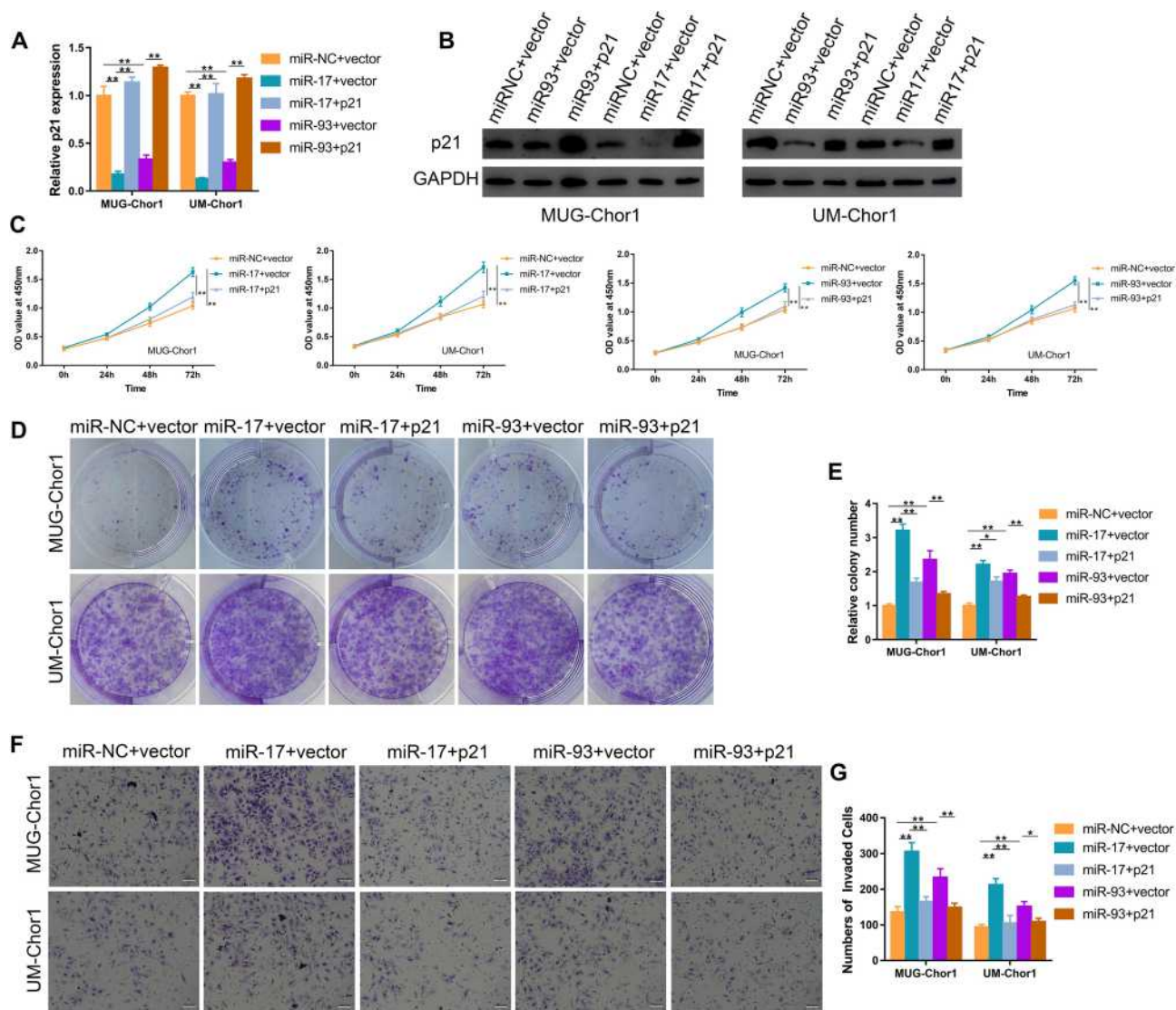


Figure 4 Overexpression of p21 counteracts the potential oncogenic effect of miR-17/93 on chordoma cells. (A and B) The qRT-PCR and Western blot were used to confirm the up-regulation of p21 in chordoma cells treated with p21 plasmid. (C) CCK-8 assay revealed that up-regulation of p21 reversed the miR-17/93 induced promotion of cell proliferation. (D and E) Colony formation assay showed the number of colony cells decreased after co-transfection with p21. (F and G) The miR-17/93 induced promotion of chordoma cell invasion was rescued after co-transfection with p21. Magnification, 100X. * $p < 0.05$, ** $p < 0.01$.

chordoma patients. Functional studies further confirmed the potential oncogenic role of miR-93 in chordoma cells. Importantly, our results revealed that miR-17 and miR-93 functioned via directly targeting p21 in chordoma, and identified the possible impact of the miR-17/93-p21 axis on chordoma progression.

Conclusions

Our results demonstrate that upregulation of miR-17/93 and decreased p21 expression is associated with poor outcomes, and miR-17 is an independent indicator of overall survival in chordoma. Our study reveals that targeting the

miR-17/93-p21 axis may be a potential therapy for chordoma patients. Further large-scale studies are wanted to validate our findings.

Abbreviations

miRNAs, micorRNAs; OS, overall survival; qRT-PCR, quantitative real-time polymerase chain reaction.

Data Sharing Statement

All data generated or analyzed during this study are included in this published article and its [Supplementary Information Files](#).

Ethics Approval and Informed Consent

The current study was approved by the ethics committee of Tangshan People's Hospital and carried out according to the Declaration of Helsinki. And informed consent was obtained from all participants.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis, and interpretation, or in all these areas; took part in drafting, revising, or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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Disclosure

The authors declare that they have no competing interests.

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