



Draft Genome Sequences of *Enterobacter cloacae* Strains CAPREx E7 and CAPREx E2-2

Rita E. Monson, Joseph Honger,* Alison Rawlinson, George P. C. Salmond

Department of Biochemistry, University of Cambridge, Cambridge, United Kingdom

ABSTRACT Enterobacter cloacae strains CAPREx E7 and CAPREx E2-2 were isolated from Ghanaian yams at a London market. The draft genome sequences indicate that the two strains are similar, with genomes of 5,042,838 and 5,039,930 bp and 56.19% and 55.05% G+C content, respectively. Both strains encoded three different β -lactamases, including one of the AmpC family.

E *interobacter cloacae* is a Gram-negative rod-shaped member of the family *Enterobacteriaceae*. *E. cloacae* strains are often associated with human infections, and many strains of *E. cloacae* carry β -lactamases or carbapenemases that confer antibiotic resistance (1). *E. cloacae* strains have also been isolated from plants, soil, and animals (2–5). Here, we report the genome sequences of two new strains of *E. cloacae*, CAPREx E7 and CAPREx E2-2, isolated from Ghanaian yams purchased from a London produce market. These strains were isolated from slices of yams that were grown on crystal violet pectate agar (6). Both strains were also able to grow at 30°C and 37°C on LB agar.

The draft genomes of both strains were generated by sequencing genomic DNA on Illumina MiSeq and HiSeq platforms using 2 × 250-bp paired-end reads. These reads were trimmed using Trimmomatic (7), assembled using SPAdes (8), and annotated using Prokka (9). The quality of the reads and assembly was also assessed using BWA-mem (10). The final assembly of CAPREx E7 contained 5,042,810 bp, with 56.19% G+C content, 4,712 open reading frames (ORFs), 78 tRNAs, and a mean coverage of 89.9×. The final assembly of CAPREx E2-2 contained 5,039,930 bp, with 55.05% G+C content, 4720 ORFs, 78 tRNAs, and a mean coverage of 137.07×.

E. cloacae strains have been isolated from many sources, and a recent source of interest has been their resistance to antibiotics and presence in persistent bacterial infections. Both CAPREx E2-2 and CAPREx E7 contained a single copy of *ampC*, a gene encoding a clinically important β -lactamase in the cephalosporinase family (1, 11). Two class B metallo- β -lactamases (MBLs) were also identified within each genome. The first, E7_03710 (CAPREx E7) or E2_03468 (CAPREx E2-2), showed high levels of similarity to a class B MBL in *Brenneria goodwinii*, a bacterium associated with oak trees (12), and *Erwinia toletana*, a bacterium associated with olive tree knots (13). The second class B MBL (E7_04150 in CAPREx E7 or E2_00247 in CAPREx E2-2) has previously been identified in many other *E. cloacae* strains but not in other plant-associated organisms.

We were also able to identify putative machinery for several secretion systems, including types I, II, IV, and VI. However, we were unable to identify any type III secretion machinery, although previous work in clinical *E. cloacae* samples found that only 27% contained type III secretion machinery (14). Secondary metabolite production can also be important in virulence. Both sequences strains contain genetic clusters predicted to encode the production machinery for the siderophore aerobactin (15, 16) and an arylpolyene similar to that produced by *Escherichia coli* (17).

The draft genomes of these two strains contain antibiotic resistance genes characteristic of other *E. cloacae* strains. However, some of their secondary metabolite clusters Received 18 April 2017 Accepted 20 April 2017 Published 8 June 2017

Citation Monson RE, Honger J, Rawlinson A, Salmond GPC. 2017. Draft genome sequences of *Enterobacter cloacae* strains CAPREx E7 and CAPREx E2-2. Genome Announc 5:e00488-17. https://doi.org/10.1128/genomeA.00488-17.

Copyright © 2017 Monson et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to George P. C. Salmond, gpcs2@cam.ac.uk.

* Present address: Joseph Honger, Soil and Irrigation Research Centre, University of Ghana, Legon, Accra, Ghana. are most similar to those in plant-associated bacterial strains. Our hope is that these genomes will provide further help in elucidating the plant-associated life cycle of some *Enterbacter cloacae* strains.

Accession number(s). The draft genomes of these two strains have been deposited in GenBank under accession no. MWME00000000 (CAPREx E2-2) and MWMD00000000 (CAPREx E7).

ACKNOWLEDGMENTS

Work in the Salmond Lab is supported by the BBSRC. J.H. was supported by a Cambridge in Africa (CAPREx) fellowship and by the Cambridge-Africa Alborada fund.

We thank Sarah Barker for technical support. The sequencing of this strain was undertaken at MicrobesNG.

REFERENCES

- 1. Mezzatesta ML, Gona F, Stefani S. 2012. *Enterobacter cloacae* complex: clinical impact and emerging antibiotic resistance. Future Microbiol 7:887–902. https://doi.org/10.2217/fmb.12.61.
- 2. Yan Y, Zhao CW, Zhang YZ, Zhang ZH, Pan GL, Liu WW, Ma QY, Hou R, Tan XM. 2012. Draft genome sequence of *Enterobacter cloacae* subsp. *cloacae* strain 08XA1, a fecal bacterium of giant pandas. J Bacteriol 194:6928–6929. https://doi.org/10.1128/JB.01790-12.
- 3. Osei Sekyere J. 2016. Current state of resistance to antibiotics of lastresort in South Africa: a review from a public health perspective. Front Public Health 4:209.
- Liu WY, Wong CF, Chung KM-k, Jiang JW, Leung FC-C. 2013. Comparative genome analysis of *Enterobacter cloacae*. PLoS One 8:e74487. https://doi .org/10.1371/journal.pone.0074487.
- Humann JL, Wildung M, Cheng CH, Lee T, Stewart JE, Drew JC, Triplett EW, Main D, Schroeder BK. 2011. Complete genome of the onion pathogen *Enterobacter cloacae* EcWSU1. Stand Genomic Sci 5:279–286. https://doi.org/10.4056/sigs.2174950.
- Perombelon MCM, Van Der Wolf JM. 2002. Methods for the detection and quantification of *Erwinia carotovora* subsp. *atroseptica (Pectobacterium carotovorum* subsp. *atrosepticum)* on potatoes: a laboratory manual. Scottish Crop Research Institute, Dundee, Scotland. http://www.scri .ac.uk/files/files/Staff/ErwiniaManual.pdf.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.

- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv arXiv:1303.3997.
- Jacoby GA. 2009. AmpC β-lactamases. Clin Microbiol Rev 22:161–182. https://doi.org/10.1128/CMR.00036-08.
- Denman S, Brady C, Kirk S, Cleenwerck I, Venter S, Coutinho T, De Vos P. 2012. *Brenneria goodwinii* sp. nov., associated with acute oak decline in the UK. Int J Syst Evol Microbiol 62:2451–2456. https://doi.org/10.1099/ ijs.0.037879-0.
- Rojas AM, de los Rios JEG, Fischer-Le Saux M, Jimenez P, Reche P, Bonneau S, Sutra L, Mathieu-Daudé F, McClelland M. 2004. *Erwinia toletana* sp. nov., associated with *Pseudomonas savastanoi*-induced tree knots. Int J Syst Evol Microbiol 54:2217–2222. https://doi.org/10.1099/ijs .0.02924-0.
- Krzymińska S, Mokracka J, Koczura R, Kaznowski A. 2009. Cytotoxic activity of *Enterobacter cloacae* human isolates. FEMS Immunol Med Microbiol 56:248–252. https://doi.org/10.1111/j.1574-695X.2009.00572.x.
- Keller R, Pedroso MZ, Ritchmann R, Silva RM. 1998. Occurrence of virulence-associated properties in *Enterobacter cloacae*. Infect Immun 66:645–649.
- Van Tiel-Menkveld GJ, Mentjox-Vervuurt JM, Oudega B, de Graaf FK. 1982. Siderophore production by *Enterobacter cloacae* and a common receptor protein for the uptake of aerobactin and cloacin DF13. J Bacteriol 150:490–497.
- Cimermancic P, Medema MH, Claesen J, Kurita K, Wieland Brown LC, Mavrommatis K, Pati A, Godfrey PA, Koehrsen M, Clardy J, Birren BW, Takano E, Sali A, Linington RG, Fischbach MA. 2014. Insights into secondary metabolism from a global analysis of prokaryotic biosynthetic gene clusters. Cell 158:412–421.