Note

Society of Appled Glycoxience

Purification and Characterization of α-Mannosidase from Onion, Allium cepa

(Received October 23, 2023; Accepted December 23, 2023)

Yui Narita,¹ Yota Tatara,² Shigeki Hamada,¹ Kaoru Kojima,¹ Shuai Li,¹ and Takashi Yoshida^{1,†}

¹ Department of Biochemistry and Molecular Biology, Faculty of Agriculture and Life Science, Hirosaki University (3 Bunkyo, Hirosaki, Aomori 036–8561, Japan)

² Department of Stress Response Science, Center for Advanced Medical Research, Hirosaki University Graduate School of Medicine (5 Zaifu-cho, Hirosaki, Aomori 036–8562, Japan)

Abstract: α -Mannosidase (ALMAN) extracted from onion (*Allium cepa*) was purified by column chromatography such as hydrophobic and gel filtration. ALMAN is an acidic α -mannosidase that exhibits maximum activity against *p*NP- α -Man at pH 4.0–5.0 at 50°C. Amino acid sequence analysis of ALMAN was consistent with α -mannosidase deduced from *Allium cepa* transcriptome analysis. The gene *alman* was amplified by PCR using mRNA extracted from onions, and a full-length gene of 3,054 bp encoding a protein of 1,018 amino acid residues was revealed. ALMAN is classified as Glycoside Hydrolase Family (GH) 38 and showed homology with other plant-derived α -mannosidases such as tomato and hot pepper.

Key words: α-Mannosidase, onion, Allium cepa

Plant α -mannosidases (EC 3.2.1.24) have been well characterized to date in several vegetables, such as bean, ginkgo, and tomato [1-3]. Genetic suppression of α-mannosidase in tomatoes and hot peppers is known to cause delayed ripening, suggesting an important role for this enzyme in plant physiology [4,5]. In our preliminary survey of vegetable glycosidases, we observed that α -mannosidase activity was particularly high in bulb vegetables such as onion and garlic. Vegetables are classified into climacteric types, which undergo additional ripening using ethylene after harvest, and non-climacteric types, which do not. Tomatoes are climacteric while the onions are non-climacteric. It has also been reported that onions have unique physiology such as ethylene suppressing sprout growth [6]. Regarding the source of enzyme isolation, plant GH38 a-mannosidases have been reported from seeds or fruits [1-5], while the nature and genes of the enzyme in bulbs are unknown. Onion (Allium cepa) is a popular bulb vegetable, but the enzymatic properties of α-mannosidase were unknown, and the gene had not been identified. In this report Allium cepa a-mannosidase (ALMAN) was characterized and the gene cloned.

Japanese yellow onion of the cultivar "Sapporo-ki" was peeled and homogenized by a cooking mixer in an equal volume (g/mL) of chilled water containing protease inhibi-

This is an open-access paper distributed under the terms of the Creative Commons Attribution Non-Commercial (by-nc) License (CC-BY-NC4.0: https://creativecommons.org/licenses/by-nc/4.0/). tors (1 mM PMSF, 1.5 µM pepstatin and 14 µM E-64). The homogenate was filtered through nonwoven paper and the filtrate was centrifuged. Proteins in the supernatants were precipitated by adding ammonium sulfate of 70 % saturation followed by centrifugation for 20 min at 8,000 rpm. The pellets were dissolved in 10 mM acetate buffer (pH 6.0) containing 1.5 M ammonium sulfate and the protease inhibitors, then applied to a column $(2.5 \times 18 \text{ cm})$ of Butyl Toyopearl 650M that was equilibrated in the same buffer. Proteins were eluted in a decreasing gradient of ammonium sulfate (1.5 to 0 M) in the buffer. Effluents containing the enzyme were pooled, concentrated by a centrifugal ultrafiltration tube (cut by 30 kDa, Macrosep®, Pall Co. USA, Port Washington, NY USA), then applied to a column (1.8×95) cm) of Toyopearl HW55F that was ran in the buffer containing 0.1 M NaCl. Eluate containing ALMAN was concentrated and used for further experiments.

Protein concentration was determined by a method developed by Bradford using bovine serum albumin as a standard [7]. PAGE in non-denaturing and denaturing conditions were performed according to the methods described by Davis and Laemmli, respectively [8, 9].

The activity of ALMAN was measured by incubating the enzyme and 0.8 mM *p*NP- α -Man (Fujifilm Wako Pure Chem. Co., Japan) in total 100 μ l of 20 mM sodium acetate buffer (pH 4.0) at 40°C for certain period. The reaction was stopped by adding 100 μ L of 1 M Na₂CO₃ then absorption at 405 nm was measured. Hydrolysis of mannobiose was determined according to the methods described in our previous paper [10].

Protein of ALMAN was reduced and carbamidomethylated in the gel after PAGE, then digested with trypsin. Internal amino acid sequences of the tryptic peptides were identified by LC-MS/MS (Triple TOF 6600, AB Sciex, Japan) with ProteinPilot Software (AB Sciex) for data processing. Since onion α -mannosidase was not found in existing protein databases, amino acid sequence of ALMAN

[†]Corresponding author (Tel. +81–172–39–3794, Email: ytakazyme@ gmail.com).

Abbreviations: CBB, Coomassie brilliant blue; DDBJ, DNA Data Bank of Japan; E-64, *N*-[*N*-(L-3-*trans*-carboxyirane-2-carbonyl)-L leucyl]-agmatine; LC-MS/MS, liquid chromatography triple quadrupole mass spectrometry; Man, D-mannose; ORF, open reading frame; PAGE, polyacrylamide gel electrophoresis; PCR, polymerase chain reaction; PMSF, phenylmethylsulfonyl fluoride; *pNP*, *p*-nitrophenyl; SDS, sodium dodecyl sulfate; SWATH : sequential window acquisition of all theoretical fragment ion spectra; TLC, thin-layer chromatography.

Steps	Total protein	Total activity	Specific activity	Yield	Purification
	(mg)	(U)	(U/mg)	(%)	(fold)
Crude extract (NH4)2SO4 precipitation Butyl Toyopearl Toyopearl HW55E	664 85.8 7.3	338 88.8 57.1 21.0	0.5 1.0 7.8	100 26 16	1 2 15

Table 1.Purification of ALMAN.

One unit (U) was defined as the amount of enzyme required to release 1 µmol of p-nitrophenol per min.

was determined by informatics works with the aid of *Allium cepa* transcriptome analysis (Bioproject Accession: PRJNA246669) [11]. By using the Prosite of ExPASy (http:// prosite.expasy.org/) the protein function was predicted from the amino acid sequence.

Onion RNA was extracted from an onion bulb by using RNAzol RT (Molecular Research Center, USA) and cDNA prepared by a reverse transcriptase. Gene *alman* was amplified by PCR using a set of primers AC-01 (TC<u>CATATG</u>GCGACTACTACTTATCCTC) and AC-02 (AG<u>CTCGAGA</u>GGAACGTCTACTTCAAG) those contained *Nde* I and *Xho* I restriction sites respectively (underlined), with the onion cDNA as the template. The PCR product was inserted



Fig. 1. PAGE of ALMAN in non-denatured (A) and SDS-denatured (B) conditions. Gels were stained with CBB.

to T vector pMD20 (Takara Bio Inc., Shiga, Japan) after adding dA and introduced to *E. coli* DH5 α . A plasmid DNA obtained from the transformants were used for sequencing.

ALMAN was purified by two steps of column chromatography (Table 1, Fig. S1: see J. Appl. Glycosci. Web site). It showed a single band on PAGE in non-denaturing condition, however five protein bands (71, 68, 53, 27, and 26 kDa) were observed on SDS-PAGE (Fig. 1). This suggests that there were proteolytic cleavages of the original protein. In tomato and chili pepper α -mannosidase, two polypeptides of approximately 70 and 50 kDa were reported [5,12]. In purification of ALMAN the low-molecular-weight polypeptides did not pass through a 30 K molecular filtration filter in their native state and were recovered on the high-molecular side, indicating that they are associated with the totality of α -mannosidase.

After ALMAN was subjected to SDS-PAGE, each of the five protein samples was subjected to amino acid sequencing by LC-MS/MS. As a result, all five samples contained a partial amino acid sequence of the same gene product, α -mannosidase (Allium cepa Transcriptome ID: Locus 2733.4) [11] (Table S1). Comprehensive detection and quantification of all compounds present in a sample (MS/MSALL) by SWATH Acquisition of proteins excised from undenatured PAGE gels revealed that the relative value of α-mannosidase (Transcriptome ID: Locus 2733.4) was the highest (as 100 %) among onion transcriptome, and another the sequence (Locus 50048.1) with the next highest value (27 %). The latter locus was thought to be the C-terminal sequence lacking in Locus 2733.4 because the two loci partially overlapped. There were no other candidates with the relative value of 10 % or more.

The optimum pH of ALMAN was on the acidic side between pH 4.0 and 5.0, and even at pH 2.0 it showed 70 % of its activity. The enzyme activity was highest at 50°C, while preheating at 60 °C for 30 min reduced the activity to less than 50 %, and above 70 °C there was no activity. Regarding



Fig. 2. TLC analysis of ALMAN reaction products.

The enzymes were reacted with 5.8 mM of each substrate in 20 mM acetate buffer (pH 4.0) at 40 $^{\circ}$ C for the times indicated. Sugars were developed in a solvent containing water, 1-butanol, and 2-propanol (4:3:12). M; α -D-Man as a standard. Mannobiose is shown as MM with its linkage.

181 271 Т Ρ М Α Т Т Y Н T. T. T. Τ. Т F Т 361 acttcagccaactacatcgcttac fatacatc tgcaacgatcgtctccggtaaactaaacgtccacgtggtcccccacactcatgacgacV T S A N Y I A Y N T S A T I V S G K **L N V H V V P H T H D D** gtgggatggctcaagaccatcgatcagtactacgtcggatca<u>aatatc</u>cattcagattgcttgtgtacagaatgtgctggattccttg 21 451 Y S G S N Ν I V Q N L 51 LK D O Υ 0 Ι A C G W Т Ι LD 541 VEQAF D K N R KF I Y F R W W 0 631 K 77 K GLVDSGQLEFVN GGMC MHDE AΑ Н $\texttt{TACGTTGATATGATTGACCAAACAACACTTGGGCACCGATTCATTAAACAAGAGTTTGGTCAGACTCCAAGAATTGGATGGCAAATTGAT$ 141 Y DMIDO TTLGHRF IKOEFGOTPR GW 0 р т т 811 Ρ OAYLLGAELGFDAL HSA Y F S G R D $\tt CGAATAAAACGTAAGAAGTTGAAGAATCTAGAGGTTGTATGGCGTGGTTCTAAGTCCCTTGGCTCTTCTGCAGATATTTTTACCGGCATA$ 901 EVVW Κ N L R G S SLGS S A D F I 991 $\tt TTTCCAAAGAATTATGAACCACCTCCTGGTGGATTTTACTATGAAATCAACGATGCCGACCCTGTTGTGCAGGATGACCCGCTTCTGTTT$ 231 F Ρ к NY E P PPG GFY Y E INDADP v v Q D D Ρ т. F GACTATAATGTTGAGGAGCGTGTAAATGATTTTGTTGCTGCAGCTCTAGAACAGGCGA<u>ACATTACT</u>AGAACAAATCACATGTTTACC 1081 VAA v ALEQAN т т 261 D Ν V E E R NDF I R T N H Ι MF Y ATGGGGACAGATTTCAAGTATCAATATGCACACCTCATGGTTCAGG CAGATGGATAAGTTTATTCATTATGTCAACCTGGATGGAAGGGTT QΥ F 291 м G D F ĸ Y AHSWF R 0 М D Κ I н Y N LD G R 1261 s P S ΙY TDAK А Κ Ε S W P L Κ ΤD F Ρ 321 ${\tt tatgctgacaatgctatgcgctattggacaggctattcactagtagacctgccctgaaaggctatgtcagaatgttgagtggctattat$ 1351 VRML NPNAYWTGYF т SRPALKGY Y 351 Y A D SG ${\tt ctggctgctaggcaattagaatttttcagaggaagaaatggtgatggcccaacaacagcagtttagcggatgctttggctattgcacaattgcaattgcacaatt$ 1441 381 L Α A RQLEF F RGRNGDGPT TDSLADAL Α I 0 caccatgatgcagttactggaactgagaaacagcatgtagccaatgattatgccaagagactggctataggttacacagaggctgagaaa411 HHD т G TEKOHVA N D YAKRLAI G Y т EAE K Α ${\tt ctagtagaagtttcacttgcttgtctaacagagtcagtttccaaatcaggttgcaggcaaaagacaaagtttgaacagtgtccacttctg}$ 1621 441 L E V S L A СЬ т E s V S K S G Q Κ Ε <u>AATATAAG</u>CTATTGTCCTCCAACAGAAGCGAAATTATCATCTGGAAATAGTTTGGTTGTTCTTGTCTACAATTCTCTGGGTGGAAAAGG 1711 N S Y P P E A K LSS ENSLVVLV Y N S т. R 471 т C Т GW ĸ 1801 501 E D R Ι Ρ v v SGDI v V H D SEGKE IES 0 L Ρ Ι Т 1891 ${\tt atggaagcttcccttaaattaagaagccgctatgtcaaggcatatttaggcacgtctccagaagtaacccccaaagtattggctagttttt$ Y т Ρ т Ρ Y W м R S R Κ Α L G s E ĸ L S L ĸ A L $\texttt{ccagtttctatacctcctccggtttcaatacgtacaccatctccacagcaaaaaagacaagtcaagcaa<u>acatgtca</u>actgtgttacaccacacaagacaagtcaagcaa<u>acatgtca</u>actgtgttacaccacagacaagacaagtcaagcaagacaagtcaagacaaagacaagacaagacaagacaagacaaagacaagacaagacaagacaagacaagacaagacaagac$ 1981 ŝ 37 T. 561 P P L GFNTY Т S А Κ КΤ S Q A N Н 2071 591 R R R ĸ G R Н Т E 77 G P G N L R T. S F D AK 0 G K Τ. H 2161 V 621 Ν S Т S K S Ε Т 0 0 S Y S Y Y ΤG DNGMG G D Ρ 0 R ${\tt gcatcgggagcatacatattccgcccagatgataagtttcccataaagcctaaatatcaggattcaacagtgtacagggatcgcttgta}$ Ρ D F v 0 0 651 2341 GATGAAGTGCATCAGCAGATAAATCCATGGATATATCAGATTACAAGAGCATACAAAGCAAAAGAACATGTTGAAGTTGAGTTTGTAGTT K A K E H V E V E F V v 681 D VН Q INPW I Y Q Т R 2431 gggccaatacctgtagaagatcgtatcgggaaagaagtagccaca caaataacaactgctatggttacca<u>acaaaaca</u>t TCTACACAGAT E DR G K EVA т тт TANV Т N тl F Y D G т P Т 0 K tcaagtggccgtgattttctaaaaaggattcgcgactacagatctgattgggaattggaagtccatcagcCagttggggaattattat 2521 741 D W E V Y S GR D T. K R R D Y R S L E Н 0 Ρ V Α G Ν S Т ${\tt cctatcaaccttgGaatttacattgaagatggcgcccaaggaactctcagtattggtagaccgctcagttgggggttccagtattgtagaccgctcagttgggggttccagtattgtagaccgctcagttgggggttccagtattgtagaccgctagtagacgctcagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtggggttccagttgtagaccgctagtggggttccagttgtagaccgctagtgggggttccagttgtagaccgctagtggggttccagttgtagaccgctagtggggttccagttgtagaccgctagtggggttccagttgtagaccgctagtggggttccagttgtagaccgctaggaggttccagttgtagaccgctaggggttccagttgtagaccgctaggggttccagttgtagaccgctaggaggttccagttgtaggagggttccagttgtaggaggttccagttgtaggaggttccagttgtaggaggttccagttgtaggaggttccagttgtaggaggttccagttgtaggaggttccagttgtaggaggttqcagttgtaggaggttccagttgtaggaggttgtaggaggttqcagttgtaggaggttgtaggaggttqcagttgtaggaggttccagttgtaggaggttgtaggaggttccagttgtaggggggttccagttgtagggggttccagttgtaggg$ 2611 Е D G AKE L s VLVD s G G s D L I R 801 G ELML H R R T. L V D D G R G V D E А T. D E DC D ${\tt gatgaatgtgacggcttgacagttaagggaaaggtttatcttagaattgatccaaaaggaggaggagcaaaatggcgtcgtcgtttggc}$ 2791 VKGKVY 831 DGLT LRIDPKGEGAKWR R S F G D E C 2881 VGSN 861 s Ρ L L I Α F SE 0 W A N S Н S Μ 0 E G K ${\tt gacccctcttatagcctggataatgttgcattgctcactcttcaggctcttgaagatggcagcaccactacttcgcttagcccacctttaggcccacctttagcccacctttagcccacctttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttaggcccactttagggcccccctttagggcccactttagggccccactttagggcccactttagggcccactttagggcc$ 2971 Ρ D Ν V A T L Q A LEDG S т LL R L 891 3061 TATGAGGTGGGGGAGGATAAAGATCTCTCAACAATGGCAAAGGTT GAACTTAAAAAGATGTTCCCTGGTAAAAAGATAAGCAAAATAACG V E 921 Е v GED ĸ D Τ. S M A Κ L Κ K M F Ρ G K Κ Т S K т 951 E т N LS A N QE R E КМЕ KKR L K W N V EGS Η S K E Ν ATTGTCAGAGGCGGCTTTATTGGTAGTTCAGATTTAGTAGTCGAGCTTGGTCCAATGAAATCAACATGGAAATTCGTACATTTGTCATTAGCTTTGATTAC 3241 981 R G G F Т G S S D T. E T. G Ρ ME R Т Т S Ε D D 3421 aacaatggcaaaggttgaacttaaaaagatgttccctggtaaaaaggtaacacaagcctcttccctttcatcatttaacaaaaggt 3511 ${\tt TTCTGTATTTTTCGACAACCAACCATAATCGGTATCCATCGATAAATGCTTTTATTCTATATAAAAGTGTCTGTAACCCAACTAATA$

3601 TTAGCATTT

Fig. 3. Nucleotide sequence of *Allium cepa* α -mannosidase (*alman*).

Amino acids residues identified by LC-MS/MS are shown in bold. The putative N-glycosylation sites are boxed. Horizontal bars indicate the positions of PCR primers. Initiation and termination of the ORF are shown in italics.

substrate specificity, ALMAN showed the highest activity towards *p*NP- α -Man with *K*m 1.69 mM and *V*max 8.2 × 10⁻² μ M/min, which was comparable to that of *Capsicum* α -Man [5]. When mannobioses (Sigma-Aldrich Co., USA) were the substrate, the activity was shown in the order of α 1-2 (3.3 %), α 1-3 (0.8 %), and α 1-6 (0.2 %) (numbers indicate relative activity to *p*NP- α -Man) (Fig. 2). Transglycosylation was not observed with mannobiose substrates. Activity towards methyl α -mannoside was negligible. ALMAN was inhibited by swainsonine with IC₅₀ of 0.2 μ M, while 1-deoxymannojirimycin, an inhibitor of 1,2- α -mannosidase [13], did not show effective inhibition.

PCR primers were designed based on the sequences of these two loci, and PCR was performed using mRNA extracted from onions as a template, resulting in a gene containing a 3,054 bp ORF (Fig. 3). All the amino acid sequences in Table S1 were identified in the ORF. The gene alman (DDBJ accession number LC779624) encoded a protein consisting of 1,018 amino acid residues with an estimated molecular mass of approximately 116 kDa. ALMAN is classified as Glycoside Hydrolase Family (GH) 38 and showed 64, 56, 58, and 65% identity with α -mannosidase from tomato [4], bean [14], rice [15], and chili pepper [5], respectively (Fig. S2: see J. Appl. Glycosci. Web site). The presence of mannose-related glycans or naturally occurring α -mannosides in onion is unknown, and the results obtained in this study will be used to investigate the relationship between ALMAN and onion growth in future studies.

CONFLICTS OF INTERESTS

The authors declare no conflict of interests.

ACKNOWLEDGMENTS

The authors are grateful to Miyu Miyazaki at the Center for Scientific Equipment Management, Hirosaki University Graduate School of Medicine, for help with LC-MS/MS analysis.

REFERENCES

- Snaith SM. Characterization of jack-bean α-D-mannosidase as zinc metalloenzyme. Biochem J. 1975; 147: 83–90.
- [2] Kwan KW, Miyazaki M, Hara S, Kimura M, Kimura Y. Purification and characterization of a Co(II)- sensitive α-mannosidase from *Ginkgo biloba* Seeds. Biosci Biotechnol Biochem. 2004; 68: 2547–56.
- [3] Hossain MA, Nakano R, Nakamura K, Hossain MT, Kimura Y. Molecular characterization of plant acidic α-mannosidase, a member of glycosyl hydrolase family

38, involved in the turnover of N-glycans during tomato fruit ripening. J. Biochem. 2010; 148: 603–16.

- [4] Meli VS, Ghosh S, Prabha TN, Chakraborty N, Chakraborty S, Datta A. Enhancement of fruit shelf life by suppressing N-glycan processing enzymes. Proc Natl Acad Sci USA. 2010; 107: 2413–8.
- [5] Ghosh S, Meli VS, Kumar KA, Thakur A, Chakraborty N, Chakraborty S, et al. The N-glycan processing enzymes α-mannosidase and β-D-N-acetylhexosaminidase are involved in ripening-associated softening in the non-climacteric fruits of capsicum. J Exp Bot. 2010; 62: 571–82.
- [6] Cools K, Chope GA, Hammond JP, Thompson AJ, Terry LA. Ethylene and 1-methylcyclopropene differentially regulate gene expression during onion sprout suppression. Plant Physiol. 2011; 156: 1639–52.
- [7] Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem. 1976; 72: 248– 254.
- [8] Davis BJ. Disc electrophoresis. ii. Method and application to human serum proteins. Ann NY Acad Sci. 1964; 121: 404– 27.
- [9] Laemmli UK. Cleavage of structural proteins during the assembly of the head of Bacteriophage T4. Nature. 1970; 227: 680-5.
- [10] Yoshida T, Inoue T, Ichishima E. 1,2-α-D-mannosidase from *Penicillium citrinum*: molecular and enzymic properties of two isoenzymes. Biochem J. 1993; 290: 349–54.
- [11] Kim S. Kim MS, Kim YM, Yeom SI, Cheong K, Kim KT, et al. Integrative structural annotation of de novo RNA-Seq provides an accurate reference gene set of the enormous genome of the onion (*Allium cepa* L.). DNA Res. 2015; 22: 19–27.
- [12] Hossain MA, Nakamura K, Kimura Y. α-Mannosidase involved in turnover of plant complex type N-glycans in tomato (*Lycopersicum esculentum*) fruits. Biosci Biotechnol Biochem. 2009; 73: 140–6.
- [13] Lobsanov YD, Vallée F, Imberty A, Yoshida T, Yip P, Herscovics A, Howell PL. Structure of *Penicillium citrinum* α 1,2-mannosidase reveals the basis for differences in specificity of the endoplasmic reticulum and Golgi class I enzymes. J Biol Chem. 2002; 277: 5620–30.
- [14] Kumar BSG, Pohlentz G, Schulte M, Mormann M, Kumar NS. Jack bean α-mannosidase: amino acid sequencing and N-glycosylation analysis of a valuable glycomics tool. Glycobiology. 2014; 24: 252–61.
- [15] Kawahara Y, Bastide M, Hamilton JP, Kanamori H, McCombie WR, Ouyang S, et al. Improvement of the *Oryza* sativa Nipponbare reference genome using next generation sequence and optical map data. Rice (NY). 2013; 6: 4.