



# Long-lasting rescue of schizophrenia-relevant cognitive impairments via risperidone-loaded microPlates

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## Abstract

Schizophrenia is a disorder characterized by cognitive impairment and psychotic symptoms that fluctuate over time and can only be mitigated with the chronic administration of antipsychotics. Here, we propose biodegradable microPlates made of PLGA for the sustained release of risperidone over several weeks. Two microPlate configurations – short:  $20 \times 20 \times 10 \mu\text{m}$ ; tall:  $20 \times 20 \times 20 \mu\text{m}$  – are engineered and compared to conventional  $\sim 10 \mu\text{m}$  PLGA microspheres in terms of risperidone loading and release. Tall microPlates realize the slowest release documenting a 35% risperidone delivery at 100 days with a residual rate of 30 ng/ml. Short microPlates and microspheres present similar release profiles with over 50% of the loaded risperidone delivered within the first 40 days. Then, the therapeutic efficacy of one single intraperitoneal injection of risperidone microPlates is compared to the daily administration of free risperidone in heterozygous knockout mice for dysbindin-1, a clinically relevant mouse model of cognitive and psychiatric liability. In temporal order object recognition tasks, mice treated with risperidone microPlates outperform those receiving free risperidone up to 2, 4, 8, and 12 weeks of observation. This suggests that the sustained release of antipsychotics from one-time microPlate deposition can rescue cognitive impairment in dysbindin mice for up to several weeks. Overall, these results demonstrate that risperidone-loaded microPlates are a promising platform for improving cognitive symptoms associated to schizophrenia. Moreover, the long-term efficacy with one single administration could be of clinical relevance in terms of patient's compliance and adherence to the treatment regimen.

**Keywords** Long-acting formulations · Microparticles · Risperidone · Schizophrenia · Drug delivery

## Introduction

Schizophrenia is a chronic and debilitating neuropsychiatric disorder affecting almost 1% of the population worldwide and representing a significant health care burden with

social and economic implications [1–3]. The annual costs associated with schizophrenia are elevated and connected to life-long chronic debilitations [2–4]. At present, there is no cure for schizophrenia, and current guidelines recommend life-long treatment with antipsychotic drugs in combination with psychological intervention to improve symptoms and prevent psychosis relapse [5–7]. However, due to the nature of the chronic treatments and patients phenotypes, a major limitation in the cure of schizophrenia is the non-adherence to the treatment regimen, which leads to an elevated risk of relapse, re-hospitalization, and in some cases suicide [5–7]. In addition, it is well-known that the long-term use of antipsychotics can lead to metabolic side effects as well as to the high risk of extrapyramidal side effects [8, 9].

One method to increase patient's adherence and compliance and reduce side effects relies on the use of long-acting injectable (LAI) formulations, which have shown huge improvements in preventing relapse, hospitalization, and mortality in schizophrenic patients [6]. Importantly, LAI

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formulations avoid plasma drug fluctuations, thus preventing side effects due to high plasma drug levels or no effects due to low plasma levels [9, 10]. A number of LAIs already marketed or in clinical trials are meant for the long-term release of risperidone (RSP), one of the currently most largely used antipsychotic drug, and one of the few to be suggested also during pediatric and adolescent age [11, 12]. RSP has been demonstrated to be effective in improving cognitive symptoms, which are key in the pathophysiology of schizophrenia [13, 14]. In this context, the first long-acting RSP (Risperdal Consta™) was approved by FDA in 2004 [15]. This is an aqueous suspension of poly(lactic-co-glycolic acid) (PLGA) microspheres to be injected deep intramuscularly and showing a size of about 100  $\mu\text{m}$  [16–18]. The RSP release profile from these large microspheres shows an initial release of ~2%, followed by a lag phase of approximately 3 weeks (lag phase), and finally by a continuous release lasting for about 30 days [17, 18]. In 2018, FDA approved another RSP-based LAI formulation, named the RBP-7000 (Perseris™), that would require only one subcutaneous injection every month [6, 15, 19].

Although LAIs represent a great improvement over the daily RSP administration, the aforementioned clinically approved formulations present various limitations. These include the initial lag phase in drug release requiring RSP oral supplementation; the fast release rates forcing for at least monthly injections; and the large particle size causing the use of large needles for intratissue injection [6]. Accordingly, in this work, a long-term drug delivery system based on PLGA microPlates ( $\mu\text{PL}$ ) is designed and tested for the controlled and continuous release of RSP over a period of 3 months, following one single intraperitoneal administration. These micron-sized particles are fabricated using a top-down approach and exhibit a square base of  $20 \times 20 \mu\text{m}$ . The  $\mu\text{PL}$  height can be precisely tailored, between 10 and 20  $\mu\text{m}$ , to modulate the release profile.  $\mu\text{PL}$  are characterized for their morphological, physico-chemical, and pharmacological features. To test their *in vivo* efficacy,  $\mu\text{PL}$  are intraperitoneally injected in heterozygous knockout mice for dysbindin-1 (*Dys* $\pm$ ), a clinically relevant mouse model of cognitive and psychiatric liability, with proved evidence of modulatory responses to antipsychotic drugs in mice and human patients [20–24].

## Materials and methods

**Materials** Polydimethylsiloxane (PDMS, Sylgard 184) and elastomer were purchased from Dow Corning (Midland, MI, USA). Poly(vinyl alcohol) (PVA, Mw 31–50 kDa), poly(D,L-lactide-co-glycolide) (PLGA, lactide:glycolide 50:50, Mw 38–54 kDa), risperidone (RSP, Mw 410.40 Da), acetonitrile (ACN), and chloroform ( $\text{CHCl}_3$ ) were purchased

from Sigma Aldrich (Saint Louis, MO, USA). Phosphate buffered saline (PBS) was purchased from Thermo Fisher Scientific (USA). All reagents and solvents were used as received.

### Fabrication of the $\mu\text{PL}$

RSP-loaded PLGA  $\mu\text{PL}$  were fabricated via a top-down approach slightly modified from previous works [25–27]. Briefly, the preparation process follows sequential replica molding steps as schematically shown in Fig. 1a. First, a master template was created via direct laser writing. The master template consists in a silicon substrate having arrays of square wells with an edge length of 20  $\mu\text{m}$  and a depth of either 10  $\mu\text{m}$  or 20  $\mu\text{m}$ , used to obtain short and tall  $\mu\text{PL}$ , respectively. Each array is separated by a gap of 3  $\mu\text{m}$ . Subsequently, the silicon template was replicated by covering it with a mixture of PDMS and curing agent that was allowed to polymerize for 8 h at 60 °C. Then, the PDMS template was peeled off the silicon master, and a PVA solution (5% w/v in deionized (DI) water) was deposited on the PDMS template. After water evaporation, the PVA template presented the same arrays of wells as the original silicon master mold. Finally, the wells on the PVA template were loaded with a solution of PLGA (20 or 60 mg in ACN for the short and tall  $\mu\text{PL}$ , respectively) and risperidone (250  $\mu\text{g}$  in  $\text{CHCl}_3$ ). After solvent evaporation, the PVA template was dissolved in water, and the solution was filtered using polycarbonate filters (40  $\mu\text{m}$  pore size). The final  $\mu\text{PL}$  were collected through sequential centrifugation steps (5000 rpm  $\times$  5 min) and stored at 4 °C until further use. Empty  $\mu\text{PL}$  were fabricated following the same approach, with the only difference that the PVA template was loaded with a solution containing PLGA only.

### Morphological and dimensional characterization of the $\mu\text{PL}$

The morphology and dimensions of  $\mu\text{PL}$  were characterized using different techniques. Specifically,  $\mu\text{PL}$  shape and size analysis were carried out via scanning electron microscopy (SEM, JEOL JSM-6490LA). Briefly, a drop of sample was deposited on a silicon support and uniformly sputtered with gold to increase the contrast and reduce sample damaging. An acceleration voltage of 10 keV was used for SEM image acquisition. Additionally,  $\mu\text{PL}$  average size and distribution were obtained via volume impedance measurements on a Multisizer 4 COULTER particle counter (Beckman Coulter, CA).  $\mu\text{PL}$  were resuspended in an electrolyte solution and analyzed using a 100  $\mu\text{m}$  capillary. Furthermore, topographical analysis was carried out to assess the average thickness of the particles using an optical profilometer.

## Degradation of the $\mu$ PL

The degradation of  $\mu$ PL over time was evaluated in vitro via SEM and Multisizer analysis. Briefly,  $\mu$ PL were incubated in PBS under mild mechanical stirring at 37 °C. At different time points, samples were analyzed to assess morphological and structural changes [27].

## Biopharmaceutical characterization of the $\mu$ PL

The RSP loading and encapsulation efficiency (LE and EE, respectively) into RSP- $\mu$ PL were evaluated by dissolving the  $\mu$ PL in ACN/H<sub>2</sub>O (1:1 v/v) and analyzing the solution via high performance liquid chromatography (HPLC) (Agilent 1260 Infinity, Germany), equipped with a 100  $\mu$ l loop. The mobile phase consisted in ACN + 0.1% TFA (v/v) and DI water + 0.1% TFA (v/v) at a ratio of 57/43 v/v and pumped at an isocratic flow rate of 0.3 ml/min. The analysis was performed by using a C18 column (2.1  $\times$  10 mm, 3.5  $\mu$ m particle size, Agilent Eclipse Plus, USA) at a detection wavelength of 280 nm. Two different standard calibration curves were created starting from standard aliquots of RSP in either ACN/DI water or PBS over the range of 0.6 to 10  $\mu$ g/ml. In addition, the limit of detection (LoD) and the limit of quantification (LoQ) of the HPLC method used to quantify RSP were evaluated starting from the standard calibration curves. Specifically, data of the standard curves (concentration –  $\mu$ g/ml, and signal – area) were analyzed by a linear regression algorithm, and LoD and LoQ evaluated based on the standard deviation of the response and the slope, according to the following equations [28]:

$$LoD = \frac{3.3\sigma}{S} \quad (1)$$

$$LoQ = \frac{10\sigma}{S} \quad (2)$$

where  $\sigma$  and  $S$  are the standard deviation of the response and the slope of the calibration curve, respectively. Values of LoD and LoQ are shown in supplementary Table S1.

The amount of RSP loaded and encapsulated into the  $\mu$ PL was calculated interpolating a standard calibration curve. The loading (%LE) and encapsulation efficiency (%EE) were quantified using the following equations [27, 29]:

$$LE(\%) = \frac{\text{RSP weight in particles}}{\text{Total weight of particles}} \times 100 \quad (3)$$

$$EE(\%) = \frac{\text{RSP weight in particles}}{\text{RSP initial feeding amount}} \times 100 \quad (4)$$

To study RSP release, short and tall  $\mu$ PL were incubated in 500  $\mu$ l PBS under mechanical stirring at 37 °C. At predetermined time points, the  $\mu$ PL were centrifuged at 500 RPM, the supernatant removed, and 500  $\mu$ l of fresh PBS added. To quantify the amount of RSP released over time, the supernatant was analyzed via HPLC using the same method described above. To interpret the RSP release profile, the experimental data were fitted to the two-phase Weibull model equation [30]:

$$\frac{(M_t)}{(M_\infty)} = 1 - \exp(-a \times t^b) \quad (5)$$

where  $M_t$  and  $M_\infty$  represent the amount of drug released at time  $t$  and infinite, respectively,  $a$  is a constant based on the system, and the exponent  $b$  is an indicator of the mechanism of transport through the polymer matrix. Values of  $b \leq 0.75$  indicate Fickian diffusion, while a combined mechanism is related to  $b$  values in the range of  $0.75 < b < 1$ . For values of  $b > 1$ , the drug transport follows a complex release mechanism [30].

## In vivo studies

### Animals

All procedures were approved by the Italian Ministry of Health (permits n. 230/2009-B and 107/2015-PR) and the local Animal Use Committee and were conducted in accordance with the Guide for the Care and Use of Laboratory Animals of the National Institute of Health and the European Community Council Directives.

Behavioral analyses were carried out on 3- to 7-month-old male dysbindin heterozygous mutant mice (Dys $\pm$ ) and their wild-type littermates (Dys+/+). The breeding scheme used to obtain the experimental mice involved mating Dys $\pm$  male mice with Dys+/+ females. Dys+/+ mice were used as female breeders in order to avoid altered maternal behavior. Animals were group-housed (3–5 per cage) and kept at constant room temperature (22  $\pm$  2 °C) and relative humidity (60%), with a 12 light/dark cycle (7 am–7 pm). Animals had free access to food and water. Distinct cohort of animals was used for each experiment. Mice were handled on alternate days during the week preceding the start of the treatment. All the experiments were carried out during the light phase. At least 1 h prior the beginning of the test, animals were allowed to acclimatize to the testing room.

### Drug and treatment

RSP was dissolved in 10  $\mu$ l of acetic acid, made up to volume with physiological saline (0.9% NaCl), and pH was adjusted to 6 with 0.1 M NaOH. Each mouse received an

intraperitoneal (IP) injection of 10 ml/kg, corresponding to a dose of 0.1 mg/kg [20, 21]. Animals were treated with RSP or saline once daily for 14 consecutive days before the behavioral analysis.

Mice treated with RSP- $\mu$ PL received a single injection of either short or tall  $\mu$ PL on day 0. The amount of injected  $\mu$ PL contained the same amount of RSP that was administered once daily for 14 days (0.1 mg/kg/14 days).  $\mu$ PL were resuspended in saline solution, and 150  $\mu$ l of the particle suspension was IP injected. Animals were treated with either short and tall RSP- $\mu$ PL or short and tall empty  $\mu$ PL, and behavioral analyses were carried out at predetermined time points.

### Temporal order object recognition test

The temporal order object recognition (TOR) test was performed as previously described [20, 31–33]. Briefly, the task was carried out in an evenly illuminated ( $25 \pm 5$  lx) open-field arena ( $40 \times 40 \times 40$  cm, Ugo Basile, Gemonio, Italy). To minimize stress-related behavior and neophobia, mice were habituated to the experimental apparatus for 4 days (10 min per day). The TOR test consists of two sample phases (SP1 and SP2) and a test phase. During SP1 and SP2, mice were allowed to explore two identical objects (different for each sample phase) for 5 min. During the test phase, mice were allowed to explore an identical copy of the objects presented in SP1 and SP2, respectively. All the object used were plastic Duplo blocks (LEGO©) different in shape, color, and size (ranging from  $9 \times 8 \times 7$  cm to  $12 \times 11 \times 10$  cm). The objects were placed at the corners of the arena, 8 cm from the sidewalls. To avoid olfactory cues, the objects were cleaned with 10% ethanol solution. The sample phases and the test phase were recorded using a camera (Sony videocam PJ330E). The time spent exploring each object was scored from the recorded videos, when mouse was facing the object 1 cm away. Climbing or rearing against the objects was not considered as an exploration. In addition, mice that spent less than 2 s exploring the objects were excluded from the analysis. The discrimination between the objects was evaluated using the discrimination index (DI) calculated as:

$$DI = \frac{T_i - T_r}{T_{tot}} \quad (6)$$

where  $T_i$  is the less recently presented object exploration time,  $T_r$  is the most recently presented object exploration time, and  $T_{tot}$  is the total exploration time.

If the TOR memory is intact, during the test trial, mice should spend more time exploring the less recently presented object. All the results are expressed as the mean  $\pm$  standard error of the mean (SEM).

### In vivo RSP concentration

Twelve weeks after the single injection of tall RSP- $\mu$ PL, mice were sacrificed, and the whole blood collected to investigate the presence of RSP, if any. The whole blood was stored at room temperature for at least 1 h and centrifuged at 1000 *g* for 15 min to collect the serum. Serum was then stored at  $-80$  °C. Subsequently, RSP was extracted from mouse serum by acetonitrile protein precipitation method. Serum samples were extracted using 100  $\mu$ l of acetonitrile. The extracts were then centrifuged for 15 min at 6.000 rpm at 4 °C. The supernatant was transferred in a glass vial and then injected into LC–MS/MS for analysis. Sample analysis was carried out using an Agilent 6410-A series triple quadrupole mass spectrometry with an electrospray ionization source (Agilent Technologies, Santa Clara, CA, USA). The analysis was performed using a Phenomenex Kinetex C8 100 Å analytical column ( $50 \times 2.1$  mm, 2.6  $\mu$ m). The mobile phase consisted in DI water + 0.5% acetic acid (v/v) and acetonitrile at a ratio of 50/50 v/v, and pumped at a flow rate of 0.2 ml/min. RSP was analyzed in positive ionization mode following multiple reaction monitoring (MRM) transitions: 411.3  $\rightarrow$  191.3 m/z. The amount of RSP in serum was calculated interpolating a calibration curve in the concentration range of 0.5–30.87 ng/ml.

### Statistical analysis

One-way ANOVA (analysis of variance) was performed to analyze the difference among the three particle configurations in terms of loading, encapsulation efficiency, and RSP released. One- or two-way ANOVA was performed to analyze the effects of the treatments in the cognitive performance during the TOR test. Newman-Keuls or Bonferroni post-hoc test was used for comparison among groups when the overall ANOVA showed statistically significant differences. The treatment effect induced by empty or RSP- $\mu$ PL in Dys  $\pm$  mice at 4-weeks post-injection was also analyzed by Student *t* test. The level of significance was set at  $p < 0.05$ . All the data were analyzed using GraphPad Prism 8.0.2 (GraphPad Software, La Jolla, CA, USA).

## Results

### Fabrication and morphological characterization of the $\mu$ PL

Two configurations of RSP-loaded  $\mu$ PL – short  $\mu$ PL and tall  $\mu$ PL – were fabricated following a multi-step, top-down approach and previously described by the authors [25, 26]. In brief,  $\mu$ PL are realized using a template-based fabrication strategy involving a silicon master template, whose wells'

geometry dictate the particle morphology, replicated in an intermediate PDMS template and eventually into a sacrificial PVA template (Fig. 1a). The wells in the PVA template are accurately filled with a polymeric paste (PLGA) dissolved in organic solvent (acetonitrile) carrying the therapeutic molecule (RSP). The PVA template is dissolved in water releasing the RSP- $\mu$ PL in solution.

The two  $\mu$ PL configurations were characterized via scanning electron microscopy (SEM), Multisizer Coulter counter, and profilometric analyses for their morphological properties. The SEM images of Fig. 1b, c show well-defined short and tall  $\mu$ PL with a common edge size length of 20  $\mu$ m and a height of 10 and 20  $\mu$ m, respectively. The different  $\mu$ PL heights can be readily appreciated in the lateral magnified pics of the microparticles laying on their sides. No porous structure was observed on the particle surface at this magnification level. Moreover, as expected, no significant difference in morphology was documented when comparing empty  $\mu$ PL and RSP- $\mu$ PL (Fig. S1a, b). The Multisizer analysis showed single peaks around 20  $\mu$ m and 30  $\mu$ m for the short and tall  $\mu$ PL, respectively, with again no appreciable difference between empty and RSP- $\mu$ PL (Fig. 1d, e). Additional topographical analyses carried out with an optical profilometer confirmed the  $\mu$ PL geometrical features as documented by the cross-section profiles of Fig. 1f, g and in the false-coloring 3D reconstruction in Fig. S1c–f, for both the short and tall  $\mu$ PL. Indeed, one major advantage of the template-based fabrication strategy is the accurate control in particle morphology, which cannot be generally achieved with bottom-up fabrication strategies.

### Degradation of the $\mu$ PL

To investigate the biodegradation of the PLGA polymer matrix, both short and tall  $\mu$ PL were incubated in PBS at 37 °C, and their morphology was assessed over time to document any possible alterations. Figure 2a, b show SEM images of the  $\mu$ PL acquired at different time points, from 0 up to 42 days, post-incubation.

On day 0, both short and tall  $\mu$ PL appeared as well-defined, squared particles as from Fig. 1b–g. On day 7, however, both short and tall  $\mu$ PL started to present some preliminary sign of biodegradation as manifested by the rounding of the originally sharp corners and the progressive transition from a square base to an overall rounded shape. At longer time points, the process started to affect also the core of the short  $\mu$ PL with a degeneration of their structure into a round microparticle already on day 42. The tall  $\mu$ PL followed overall the same behavior but at lower degradation rates, likely due to the larger amount of PLGA used to realize the tall versus short  $\mu$ PL. Nonetheless, even for the tall  $\mu$ PL, the microparticles tended to progressively lose their original squared shape to become more globular towards the

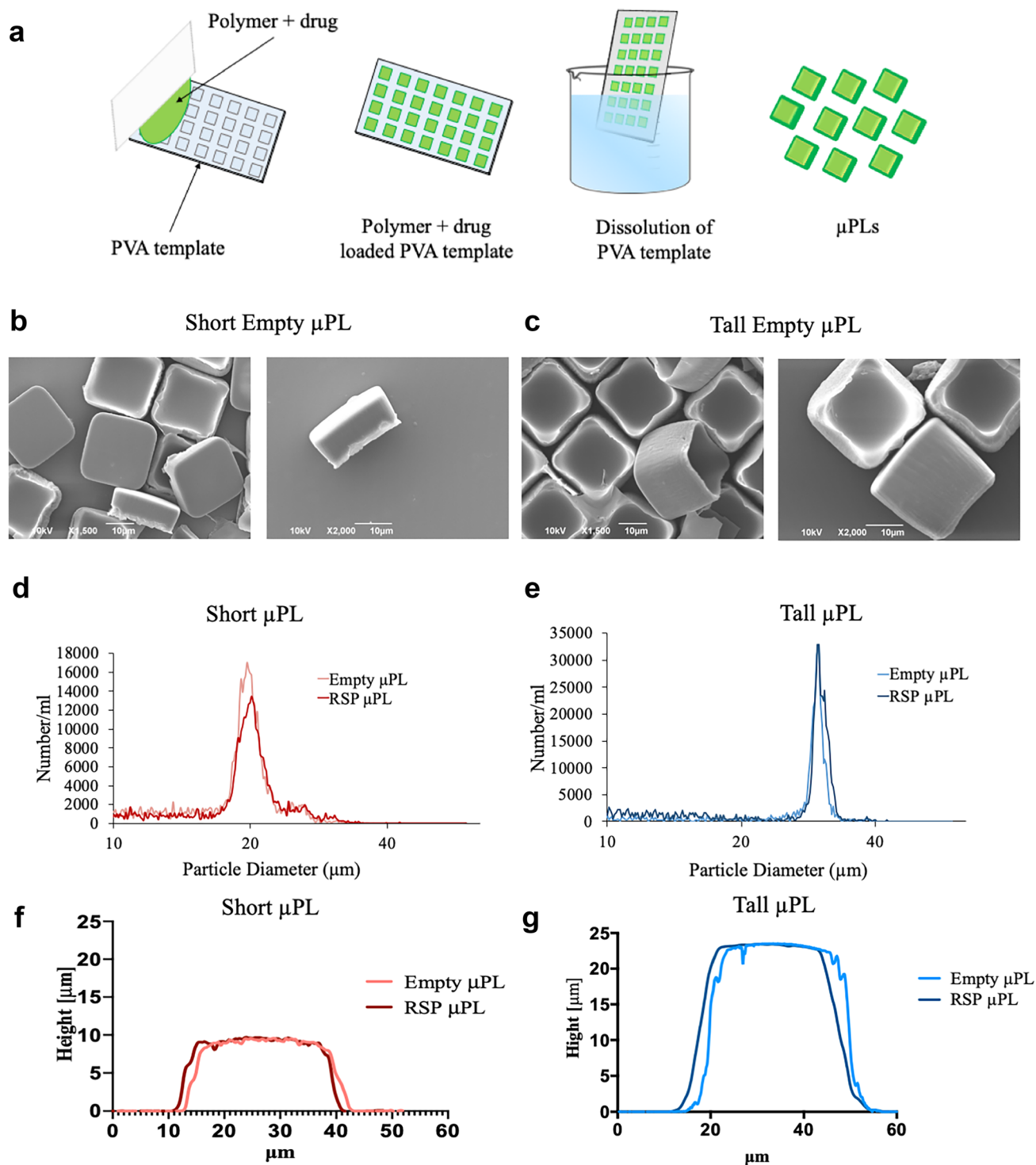
end of the observation period (Fig. S2). This behavior was quantified via a Multisizer Coulter counter analysis returning the overall size spectrum distribution for the short and tall  $\mu$ PL over time (Fig. 2c, d). This shows a progressive shift of the characteristic peak towards a larger average size and a reduction of the peak value and overall area under the curve. The shift of the peak has to be associated with the change in microparticle structure and possible coalescence with other particles or debris. The reduction in peak value is directly correlated with the number of particles. Multisizer data are referred to the degradation of particles from day 0 to day 21. Note that longer time point measurements were not possible due to the change in  $\mu$ PL morphology that made the sample unsuitable for the analysis.

### Biopharmaceutical characterization of the $\mu$ PL

Short and tall  $\mu$ PL loaded with RSP were characterized in terms of RSP loading (LE%), defined as the ratio between the loaded mass of RSP and the total particle mass; encapsulation efficiency (EE%), defined as the ratio between the loaded mass of RSP and the initial RSP input; as well as for their ability to control the release of RSP over time. A direct comparison with PLGA microspheres was also performed. Specifically,  $\sim$  10  $\mu$ m PLGA spherical particles ( $\mu$ S) were realized using a standard homogenization protocol [34, 35]. Figure S3 shows electron microscopy images, DLS, and Multisizer analysis for the  $\sim$  10  $\mu$ m microspheres.

No statistically significant differences were observed in terms of loading among the three different microparticles, with  $1.52 \pm 0.20\%$  for the short RSP- $\mu$ PL;  $1.73 \pm 0.59\%$  for the tall RSP- $\mu$ PL; and  $1.84 \pm 0.18\%$  for the  $\mu$ S, respectively (Fig. 3a – one-way ANOVA:  $p=0.685$  short  $\mu$ PL vs tall  $\mu$ PL,  $p=0.837$  short  $\mu$ PL vs  $\mu$ S,  $p=0.799$  tall  $\mu$ PL vs  $\mu$ S). On the other hand, significant differences were documented for the encapsulation efficiency. The drug encapsulated within the RSP- $\mu$ S ( $27.57 \pm 0.20\%$ ) was significantly higher than for the tall RSP- $\mu$ PL ( $6.55 \pm 0.13\%$ ) and the short RSP- $\mu$ PL ( $2.21 \pm 0.43\%$ ), respectively (Fig. 3b – one-way ANOVA,  $***p < 0.001$ ). Moreover, the encapsulation efficiency of the tall  $\mu$ PL was significantly higher than the encapsulation efficiency of the small  $\mu$ PL (Fig. 3b – one-way ANOVA:  $***p < 0.01$ ).

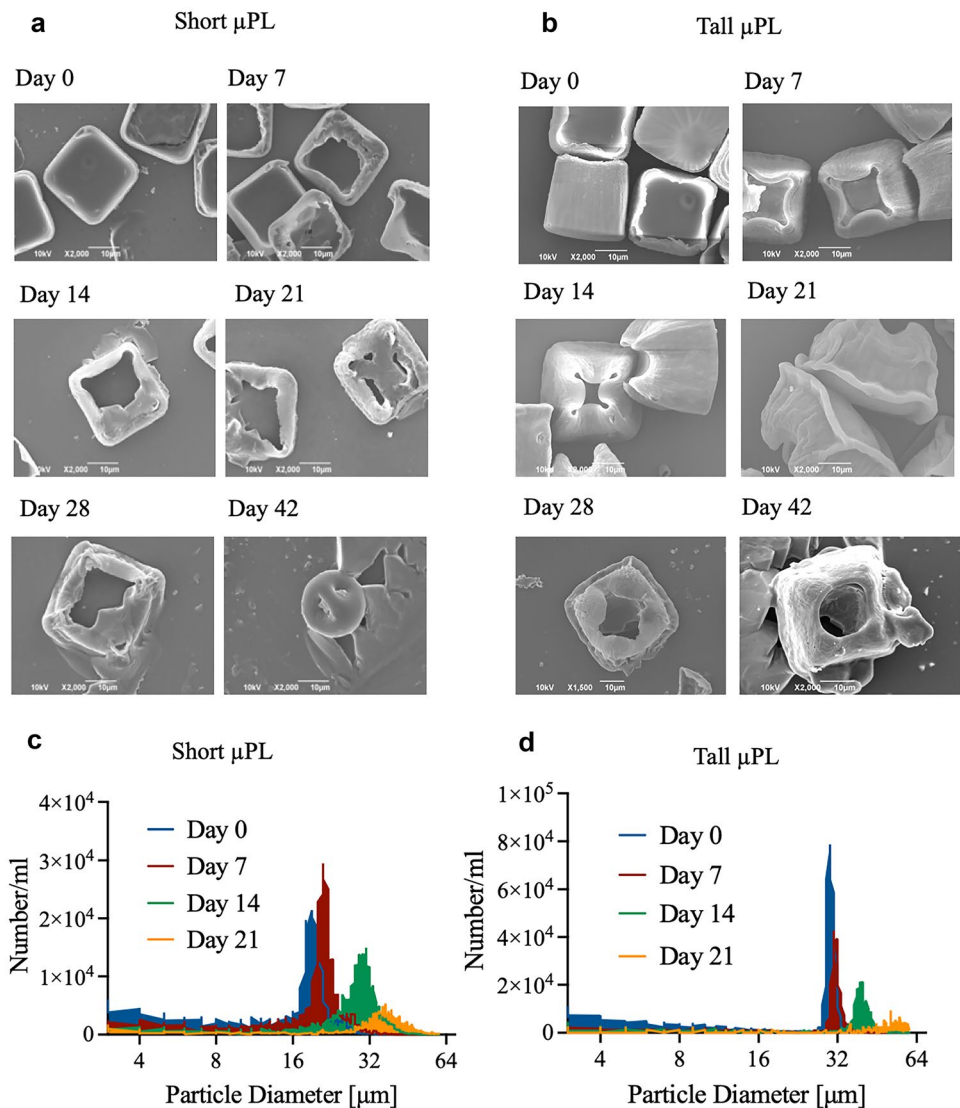
The RSP release kinetics was determined under physiological conditions (PBS, pH 7.4, 37 °C) in a 500  $\mu$ l volume to better mimic the confined space for intratissue deposition. The release profiles for the two  $\mu$ PL configurations and the  $\mu$ S are shown in Fig. 3c in terms of percentage of drug released over the first 40 days of observation. Tall RSP- $\mu$ PL showed the slowest release profile with less than 30% ( $26.85 \pm 1.50\%$ ) of the loaded drug being released within the first 5 weeks of observation. In the same time period, short RSP- $\mu$ PL and RSP- $\mu$ S released over 50% ( $57.81 \pm 0.85\%$  and



**Fig. 1**  $\mu$ PL fabrication process, morphological and dimensional characterization. **(a)** Sequential steps in the fabrication of  $20 \times 20 \times 10$  and  $20 \times 20 \times 20$   $\mu$ PL. A silicon master template is fabricated via direct laser writing and replicated into a PDMS template, whose pattern is then transferred into a sacrificial PVA template. This PVA template is loaded with the polymeric paste constituting the final  $\mu$ PL and enclosing the payload.  $\mu$ PL are released and collected upon dissolution in DI water of the sacrificial PVA template (from left to

right). **(b)** SEM micrographs of short and **(c)** tall  $\mu$ PL showing the characteristic dimensions of  $20 \times 20 \times 10$  and  $20 \times 20 \times 20$   $\mu$ m. (Scale bar: 10  $\mu$ m). **(d)** Size characterization of short and **(e)** tall  $\mu$ PL via Multisizer analyses. **(f)** Cross-section profile of short and **(g)** tall  $\mu$ PL acquired by optical profilometer showing the thickness of the  $\mu$ PL. No significant differences were highlighted between empty and RSP- $\mu$ PL

**Fig. 2** Degradation of  $\mu$ PL under physiological conditions. (a) SEM images representing the degradation of short and (b) tall  $\mu$ PL over time. (c) Degradation of short and (d) tall  $\mu$ PL analyzed via the Multisizer

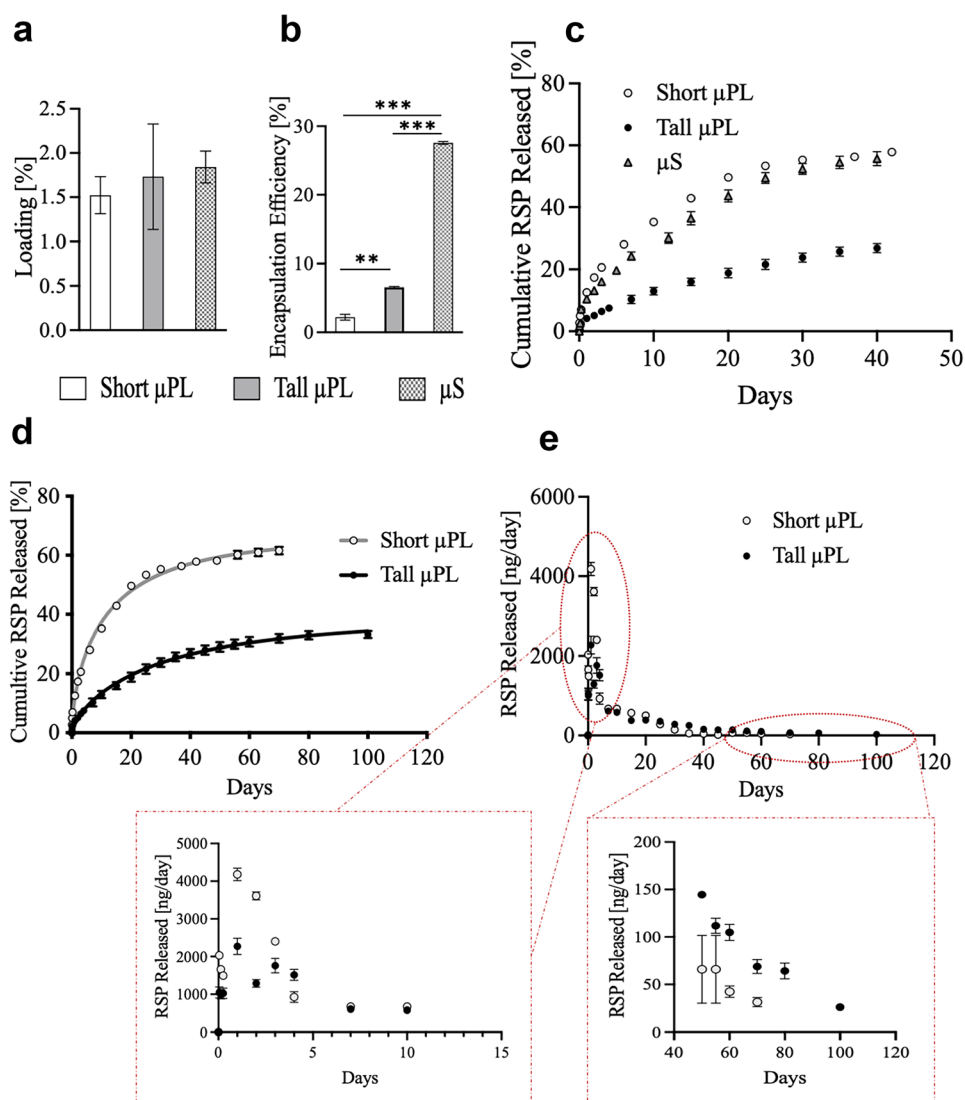


55.69 ± 2.28%, respectively) of the loaded drug (one-way ANOVA: # $p < 0.00001$  short  $\mu$ PL vs tall  $\mu$ PL, § $p < 0.0001$   $\mu$ S vs tall  $\mu$ PL). Given the similarity between the release profiles for the short RSP- $\mu$ PL and RSP- $\mu$ S ( $p = 0.21$  – no significant differences), more details are acquired for the short and tall RSP- $\mu$ PL in terms of cumulative and instantaneous drug released, as shown in Fig. 3d, e, respectively. Within the first 24 h of incubation, about 12% of RSP was released from the short  $\mu$ PL as opposed to 4% for the tall  $\mu$ PL (one-way ANOVA: \*\*\* $p < 0.001$ ), while at 50 days, about 60% and 29% of RSP was released from the short and tall  $\mu$ PL, respectively (one-way ANOVA: # $p < 0.00001$ ). For the tall  $\mu$ PL, the drug molecules were continuously delivered over time to reach just about 33% of release at 100 days (Fig. 3d). The daily release of RSP from short and tall  $\mu$ PL is shown in Fig. 3e. For the short RSP- $\mu$ PL, the drug is rapidly released within the first week reaching a peak of 4000 ng/ml at day 1 that drops rapidly to 1000 ng/ml at day 5 and continuously

decreases down to about 50 ng/ml around day 60. A similar trend is documented for the tall RSP- $\mu$ PL but with a more moderate variation in daily release rate. Specifically, a peak of only 2000 ng/ml is detected at day 1 followed by a drop to 1000 ng/ml at day 7 that stays quasi-constant till day 25. The daily release rate continuously decreases till 50 ng/ml at day 100.

The release kinetics data were interpolated using the Weibull equation to extrapolate the RSP release mechanisms for both short and tall  $\mu$ PL. The Weibull model parameters (a, b) and the corresponding fitting accuracy  $R^2$  for the in vitro release profiles are reported in Table 1. In this work, the fitting was successful for the entire set of data, with a  $R^2$  of 0.9960 and 0.9979 for short and tall  $\mu$ PL, respectively. The values of  $b$  obtained for the best fit were 0.6557 and 0.6766 for short and tall  $\mu$ PL, respectively. These values suggest that  $\mu$ PL provide a sustained release of RSP based on a diffusion-controlled mechanism through the PLGA matrix.

**Fig. 3** Pharmacological characterization of  $\mu$ PL. **(a)** RSP loading into short  $\mu$ PL, tall  $\mu$ PL, and  $\mu$ S. **(b)** RSP encapsulation into short  $\mu$ PL, tall  $\mu$ PL, and  $\mu$ Spheres. Statistical significance was determined by one-way ANOVA:  $**p < 0.01$  tall  $\mu$ PL vs short  $\mu$ PL,  $***p < 0.001$   $\mu$ S vs short  $\mu$ PL and  $\mu$ S vs tall  $\mu$ PL. **(c)** 40-day cumulative release profiles of RSP from short  $\mu$ PL, tall  $\mu$ PL, and  $\mu$ S (37 °C in PBS). **(d)** Cumulative and **(e)** daily release profiles of RSP from short and tall  $\mu$ PL (37 °C in PBS). Green and black lines in **(d)** represent the Weibull function fitting; 95% confident band. Data are expressed as the mean  $\pm$  standard deviation ( $n = 3$ ). Statistical significance was determined by one-way ANOVA:  $*p < 0.05$ ,  $***p < 0.001$ ,  $\$p < 0.0001$ ,  $\#p < 0.00001$ . Details of the statistical analysis are reported in Table S2



This implies that drug release from the  $\mu$ PL into the surrounding aqueous environment would depend on the molecular weight of the drug as well as on the size of the pores in the PLGA matrix rather than its biodegradation. These results are also in line with the release mechanism of other small molecules from similar microparticles [27]. In comparing the short and tall  $\mu$ PL, one can observe a doubling in volume ( $\mu$ PL height increasing from 10 to 20  $\mu$ m) and a three-time increase in PLGA mass (from 20 to 60 mg) used during the fabrication process. Therefore, it can be argued that the slower release kinetics associated with the tall  $\mu$ PL should be associated with their more compact structure.

## Temporal order object recognition (TOR) test

### Single injection of short RSP- $\mu$ PL

In order to investigate if a single injection of RSP- $\mu$ PL could ameliorate dysbindin-induced deficit, Dys $\pm$  mice received

either free RSP (or vehicle only) or RSP- $\mu$ PL (or empty  $\mu$ PL). The free drug or vehicle were administered intraperitoneally (0.1 mg/kg) on a daily basis for 2 weeks, whereas the RSP- $\mu$ PL or empty  $\mu$ PL were deposited i.p. once at the beginning of the experiment for a total equivalent RSP dose of 1.4 mg/kg, to match the total amount of free drug received in 2 weeks. TOR tests are conducted at 2- and 4-week post-treatment initiation, as described in Fig. 4a.

**Table 1** Weibull model parameters (a, b) and fitting accuracy  $R^2$  for RSP release from short and tall  $\mu$ PL

| $\mu$ PL           | a     | b     | $R^2$ |
|--------------------|-------|-------|-------|
| Short RSP $\mu$ PL | 0.201 | 0.655 | 0.99  |
| Tall RSP $\mu$ PL  | 0.112 | 0.677 | 0.99  |

$$\text{Weibull model equation: } \frac{(M_t)}{(M_\infty)} = 1 - \exp(-a \times t^b)$$



At all the time points, Dys  $\pm$  mice treated with short empty  $\mu$ PL were not able to discriminate between the two objects presented at different time, confirming previously found recency memory cognitive impairment (Fig. 4b, c). This suggests that the short  $\mu$ PL per se had no effect. At 2 weeks, the treatment with both free RSP and RSP- $\mu$ PL did not affect the total time ( $T_{tot}$ ) mice spent to explore the two objects ( $F_{(3,22)} = 1.276$ ,  $p = 0.3070$ ) as shown in Fig. S4a, but it is effective in selectively improving TOR cognitive performances ( $F_{(3,22)} = 7.943$ ,  $p = 0.0009$ ). Specifically, at 2 weeks, Dys  $\pm$  mice treated with daily injections of free RSP showed a significantly higher DI when compared to the vehicle-treated Dys  $\pm$  mice (\*\* $p < 0.01$ ) (light vs dark gray bars – Fig. 4b). Notably, also Dys  $\pm$  mice treated with a single injection of short RSP- $\mu$ PL improved their TOR performances, showing an increased DI that was significantly higher (\* $p < 0.05$ ) than that of the Dys  $\pm$  mice receiving a single injection of empty  $\mu$ PL (black vs white bars – Fig. 4b). Importantly, there were no significant differences between the TOR performance of Dys  $\pm$  mice treated with daily injections of free RSP and Dys  $\pm$  mice treated with a single injection of short RSP- $\mu$ PL (black vs light gray bars – Fig. 4b). This result suggests that the  $\mu$ PL-based system helped ameliorate the dysbindin-induced cognitive deficits

similarly to the free drug, with one single injection of particles releasing a lower amount of drug than the conventional dose. TOR was also performed at 4-week post-treatment initiation. Similar to the 2-week post-treatment, there was no significant difference in the time spent by the mice to explore the two objects ( $p = 0.5358$ ) (Fig. S4b). However, even if Dys  $\pm$  mice treated with short RSP- $\mu$ PL showed improvement in the TOR performances over Dys  $\pm$  mice treated with short empty  $\mu$ PL (Fig. 4c), the difference between the two groups was not significant ( $p = 0.6099$ ).

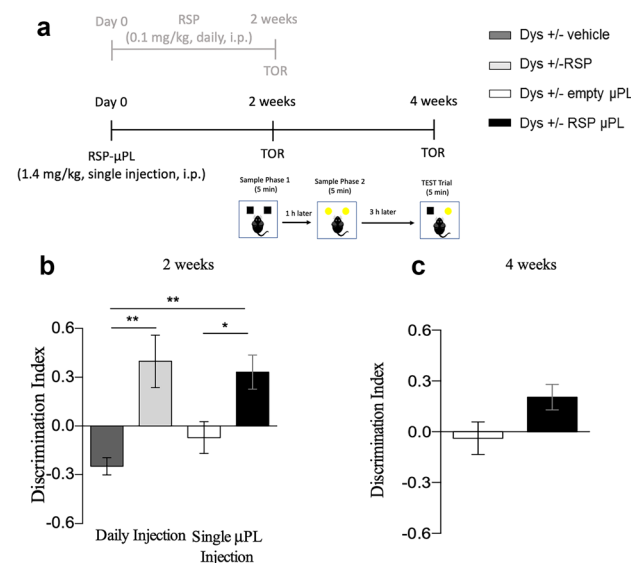
### Single injection of tall RSP- $\mu$ PL

In these studies, Dys  $\pm$  mice and their wild-type (Dys +/+ ) littermates are injected intraperitoneally on day 0 with tall empty  $\mu$ PL or tall RSP- $\mu$ PL (1.4 mg/kg drug equivalent dose), according to the timeline shown in Fig. 5a.

The mice were subjected to the TOR tests at 2-, 4-, 8-, and 12-week post-treatment initiation. At all the time points, treatment with tall RSP- $\mu$ PL did not lead to any difference in the total time spent by the mice to explore the two objects during the 5-min test trial (Fig. S5a–d), but it was effective in rescuing the cognitive impairment. Specifically, Dys  $\pm$  mice treated with the RSP- $\mu$ PL showed an increased DI when compared to Dys  $\pm$  mice treated with the empty  $\mu$ PL (Fig. 5b–e). However, the difference between the two was statistically significant at 2 (Newman-Keuls post-hoc: \*\* $p < 0.001$  vs empty  $\mu$ PL), 4 (Newman-Keuls post-hoc: \*\*\* $p < 0.0001$  vs empty  $\mu$ PL), and 8 weeks (Newman-Keuls post-hoc: \* $p < 0.05$  vs empty  $\mu$ PL) only, and not significant at 12 weeks. Second, the therapeutic effect of the tall RSP- $\mu$ PL on the Dys  $\pm$  mice was compared to the effect of both the tall empty  $\mu$ PL and the tall RSP- $\mu$ PL on the Dys +/+ mice. No significant difference was observed at all four time points (Fig. 5b–e), thus implying that the RSP- $\mu$ PL alone can rescue the animal behavior and that one single administration of RSP- $\mu$ PL at day 0 returns Dys  $\pm$  mice behaving just like the Dys +/+ mice up to 12 weeks.

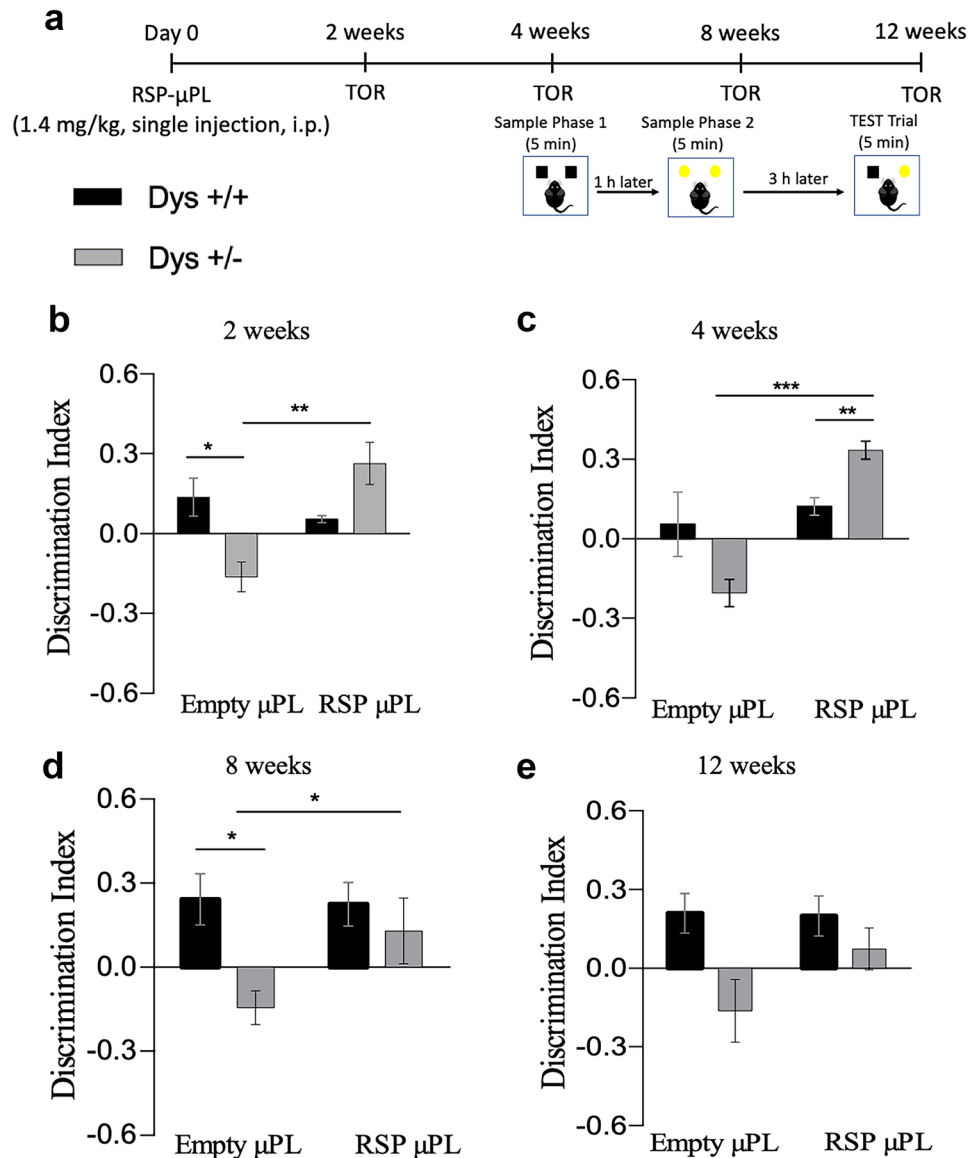
To assess if the tall RSP- $\mu$ PL would progressively lose their efficacy over time or if there was any kind of habituation of the animals to the TOR tasks, Dys  $\pm$  mice and Dys +/+ mice were injected with tall RSP- $\mu$ PL or empty  $\mu$ PL on day 0. Then, their behavioral performances are tested only at 8- and 12-week post-injection, according to the schematic of Fig. 6a.

As previously shown, treatment with tall RSP- $\mu$ PL did not lead to any difference in the total time spent by the mice to explore the two objects during the 5-min test trial (Fig. S6a, b). Importantly, a single injection of tall RSP- $\mu$ PL was effective in restoring the recency memory impairment in Dys  $\pm$  mice, with a DI that was significantly higher than that of Dys  $\pm$  receiving tall empty  $\mu$ PL, at both 8 (Bonferroni post-hoc: \*\*\* $p < 0.0001$  vs empty  $\mu$ PL) and



**Fig. 4** Single injection of short RSP- $\mu$ PL improves cognitive abilities of Dys  $\pm$  in mice in the TOR task. Each experimental group received free vehicle ( $n = 7$ , dark gray bars) or free risperidone ( $n = 7$ , light gray bars) for 14 days, and short empty ( $n = 6$ , white bars) or RSP- $\mu$ PL ( $n = 6$ , black bars) only once on day 0. (a) Experimental timeline. (b) Discrimination index showed by Dys  $\pm$  mice during the 5-min test phase 2 weeks post-treatment. (c) Discrimination index showed by Dys  $\pm$  receiving either empty or RSP- $\mu$ PL during the 5-min test phase 4 weeks post treatment. The values are the means  $\pm$  s.e.m. One-way ANOVA: treatment effect \* $p < 0.05$ , \*\* $p < 0.005$

**Fig. 5** Single injection of tall  $\mu$ PL improves the cognitive abilities of Dys  $\pm$  mice in the TOR task. Each experimental group ( $n=6$ ) received either empty or RSP- $\mu$ PL only once on day 0 and were tested in the TOR task at 2-, 4-, 8-, and 12-week post-injection. (a) Experimental timeline. (b) Discrimination index showed by mice during the 5-min test phase at 2, (c) 4, (d) 8, and (e) 12 weeks, respectively. The values are the means  $\pm$  s.e.m. Two-way ANOVA: treatment effect  $*p < 0.05$ ,  $**p < 0.01$ ,  $***p < 0.001$ . At 2- and 4-week post-injection, the discrimination performance was affected by the treatment (2 weeks – treatment effect:  $F_{(1,13)} = 7256$ ,  $p = 0.0184$ ; 4 weeks – treatment effect:  $F_{(1,13)} = 20.55$ ,  $p = 0.0006$ ), at 8- and 12-week post-injection the discrimination performance was impacted mainly by the genotype (8 weeks – genotype effect:  $F_{(1,13)} = 6.506$ ,  $p = 0.0242$ ; 12 weeks – genotype effect:  $F_{(1,13)} = 7.739$ ,  $p = 0.0156$ )

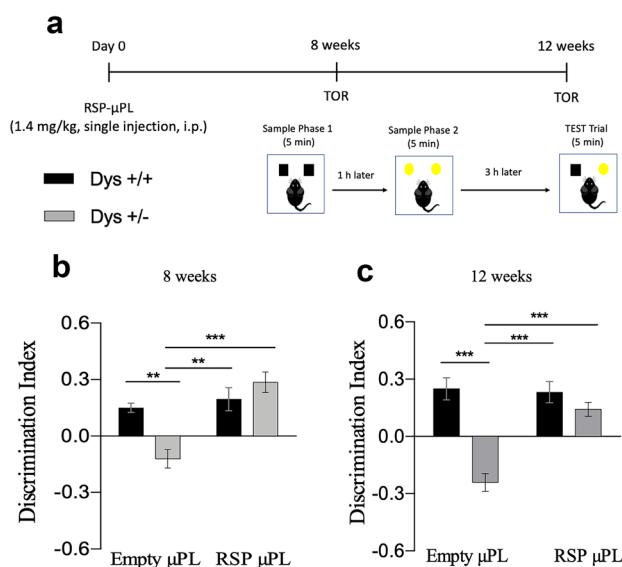


12 weeks (Bonferroni post-hoc:  $***p < 0.0001$  vs empty  $\mu$ PL) (Fig. 6b, c). Notably, at both time points, the cognitive performance was impacted by the treatment (8 weeks – treatment effect:  $F_{(1,28)} = 17.76$ ,  $p = 0.0002$ ; 12 weeks – treatment effect:  $F_{(1,28)} = 16.66$ ,  $p = 0.0003$ ) and by the interaction between treatment and genotype (8 weeks – treatment  $\times$  genotype effect:  $F_{(1,28)} = 11.29$ ,  $p = 0.0023$ ; 12 weeks – treatment  $\times$  genotype effect:  $F_{(1,28)} = 12.90$ ,  $p = 0.0009$ ). These results suggest that RSP- $\mu$ PL can have a prolonged effect up to 3 months upon single administration, leading to an overall improvement of the cognitive performances of mice carrying dysbindin-induced deficit.

It should be here highlighted that for all the TOR tests (Figs. 5 and 6), Dys  $\pm$  mice receiving a single injection of tall empty  $\mu$ PL showed no discrimination, indicating

the presence of a cognitive impairment and confirming that the tall  $\mu$ PL too per se had no effect. Notably, for all the TOR tests, the administration of tall empty  $\mu$ PL had no negative outcome on the cognitive performances of Dys +/+ mice, reinforcing the fact that the  $\mu$ PL per se did not lead to any side effect.

Finally, the RSP concentration was assessed through HPLC MS/MS quantitative determination 12 weeks after a single injection of tall RSP- $\mu$ PL. The analysis revealed a serum concentration of  $6.40 \pm 1.26$  ng/ml and  $4.90 \pm 0.77$  ng/mL in RSP- $\mu$ PL treated Dys +/+ and Dys  $\pm$  mice, respectively (Fig. S7), which appears to be comparable to the daily RSP release measured in vitro at 100 days (Fig. 3e). Non-detectable RSP concentration was confirmed for the empty  $\mu$ PL treated mice.



**Fig. 6** Single injection of tall  $\mu$ PL improves the cognitive abilities of Dys $\pm$  mice in the TOR task. Each experimental group received either empty (Dys +/+  $n=6$ , Dys  $\pm n=10$ ) or RSP- $\mu$ PL (Dys +/+  $n=6$ , Dys  $\pm n=10$ ) only once on day 0 and were tested in the TOR task at 8- and 12-week post-injection. (a) Experimental timeline. (b) Discrimination index showed by mice during the 5-min test phase at 8, (c) 12 weeks, respectively. The values are the means  $\pm$  s.e.m. Two-way ANOVA: treatment effect  $**p < 0.01$ ,  $***p < 0.001$

## Discussion

The current treatments for schizophrenia rely on the life-long administration of antipsychotic drugs, such as risperidone, in combination with psychological intervention. Indeed, given this premises, major limitations in the proper management of schizophrenia are the non-adherence to the chronic regimen and the prolonged use of antipsychotics, often leading to metabolic and extrapyramidal side effects. In this context, long-active injectable (LAI) formulations for the sustained release of small drug doses represent an interesting approach. However, most available LAI for RSP, suffer of discontinuous drug release, with extensive initial lag phases, and rapid drug release, with the majority of the drug dose being delivered within 30 days or less. Indeed, conventional microspheres in the size range of 100  $\mu$ m and larger show a typical cumulative RSP profile with an initially long lag phase (between 10 and 20 days) followed by an intermediate phase of sustained release (about 1 month) and a final phase of slow release of the residual amount of drug, which is typically insufficient for any in vivo activity [36]. As documented, this is also the case of the Risperdal Consta formulation [17, 18], which consequently requires the oral administration of RSP during the first 2–3-week post-microsphere injection to compensate for the lag phase. Several authors have tried to improve the release of RSP

from polymeric microspheres by reducing their particle size. It was demonstrated that PLGA microspheres in the order of 10  $\mu$ m in size have no lag phase but, given their small size, loose rapidly RSP so that the active release phase lasts 1–2 weeks [17, 18].

Differently, the release of RSP from both short and tall  $\mu$ PL follows a totally different kinetics. This is initially characterized by a relatively rapid release of the drug molecules mainly residing at the particles surface and is then followed by a prolonged and sustained release that is mainly diffusion driven. As expected, the increase in  $\mu$ PL thickness and polymer content for the tall over the short  $\mu$ PL reduced the initial burst effect and prolonged the release of RSP up to 100 days. Therefore, as opposed to microspheres, the tall  $\mu$ PL offers the unique advantage of providing the drug immediately from time zero (no lag phase) together with a sustained release for at least 3 months (long therapeutic action).

This unique release kinetic was the direct consequence of the exquisite control on  $\mu$ PL geometry and PLGA mass during the fabrication process. Differently from conventional microspheres, the  $\mu$ PL geometry can be finely tuned so to realize two  $\mu$ PL configurations: one is 10  $\mu$ m high (short  $\mu$ PL), and the second one is 20  $\mu$ m high (tall  $\mu$ PL). Both  $\mu$ PL have a square base with an edge length of 20  $\mu$ m. On the contrary, polymeric microspheres are often characterized by a significant size range, as well as porous/rough surface which clearly impacts on their release characteristics [16–18, 36, 37].

The difference in thickness and polymer content between the two  $\mu$ PL configurations clearly affected the RSP loading, encapsulation efficiency, and release kinetics. Indeed, the significantly higher EE of the tall  $\mu$ PL has to be ascribed to their larger height along with the increased polymer content (20  $\mu$ m, 60 mg), as opposed to the short  $\mu$ PL (10  $\mu$ m, 20 mg). On the other hand, the comparable loading was expected, since LE is defined as the ratio between the weight of drug loaded into the  $\mu$ PL (higher for the tall  $\mu$ PL) and the total weight of the particles, which grows with the amount of PLGA. Even though the LE and EE are relatively low, these values are in line with what has been already published for risperidone-loaded microspheres of comparable size. Indeed, the literature shows that microspheres with a diameter of a few tens of microns present an EE of about 30% and a loading of a few points percentage [17, 18]. Only much larger microspheres, with an average diameter in the range of 100  $\mu$ m and higher, have shown encapsulation efficiencies close to 90% and loading values in the range of 30–40% [16, 36, 37]. However, these larger particles require larger needle for the injection, and their higher loading does not necessarily translate in enhanced pharmacological performances, as described previously.

The  $\mu$ PL developed in this work, and in particular the tall particles, combine the benefits of the small PLGA microspheres (no lag phase) with those of the larger PLGA microspheres (prolonged release), thus extending the therapeutic window. This was clearly demonstrated by the blood concentration of RSP at 3-month post-administration, which was in the range of 4–6 ng/ml, with only 1.4 mg/kg initial administration. Other studies have demonstrated similar (or lower) plasma RSP concentrations at 3 months, but starting with an initial dose of 40 mg/kg [16]. Most importantly, the results of the behavioral studies demonstrated the efficacy of both short and tall  $\mu$ PL in improving the cognitive impairment characterizing *Dys*  $\pm$  mice. Disruption of recency memory is a core symptom for patients affected by psychiatric disorders, including schizophrenia [38]. In rodents, this ability is assessed using the temporal order object recognition task (TOR), which evaluates the ability to discriminate two objects presented at two different moments [31, 32, 39]. Using the TOR tests, we previously demonstrated that chronic daily treatment with RSP (0.1 mg/kg) rescued cognitive impairments in *Dys*  $\pm$  mice as similarly reported in human patients [20]. Within this context, the therapeutic efficacy and possible toxicity of one single injection of RSP- $\mu$ PL was tested in *Dys*  $\pm$  and their wild-type littermates (*Dys* +/+), subjected to TOR tests over a period of up to 3 months. By measuring the time spent to explore the two objects in the testing phase, the authors assessed if the treatment with RSP- $\mu$ PL was able to improve the cognitive impairment characterizing *Dys*  $\pm$  mice. Notably, treatments with short and tall  $\mu$ PL did not lead to any significant difference in the total time mice spent to explore the two objects during the 5-min test trial as documented by Figs. S4a, b, S5a, d, and S6a, b, suggesting that the  $\mu$ PL per se had no effect, thus giving important information on their safety profile. In addition, mice receiving empty  $\mu$ PL showed no discrimination, indicating the presence of a cognitive impairment and confirming that the particles per se have no effects. Notably, when  $\mu$ PL were administered to *Dys*  $\pm$  mice, they had no negative outcome on the cognitive performances, reinforcing the fact that  $\mu$ PL did not lead to any side effect. A single i.p. deposition of short RSP- $\mu$ PL was sufficient to rescue the cognitive deficit in *Dys*  $\pm$  mice for up to 14 days, similarly to the conventional daily administration of 0.1 mg/kg free RSP. However, at 4-week post-deposition, the rescuing effect of the short RSP- $\mu$ PL diminished, even if the trend was still visible. As documented by the biopharmaceutical analysis presented in Fig. 3, tall RSP- $\mu$ PL can load larger amounts of drug and release it over longer periods. Therefore, the following natural step was that of testing in vivo therapeutic efficacy over longer times. A single injection of tall  $\mu$ PL was effective in rescuing the cognitive impairment of *Dys*  $\pm$  mice up to 12 weeks, representing an advantage over the short RSP- $\mu$ PL, where the TOR

performances were improved only at 2-week post-injection. Indeed, this extended rescuing effect should be associated with the longer release of RSP from the tall over the short  $\mu$ PL. Importantly, a single injection of tall  $\mu$ PL not only helped restore the recency memory impairment, but led to a behavior of *Dys*  $\pm$  mice similar to those of *Dys* +/+ mice, thus confirming the efficacy of the treatment.

## Conclusions

In this study, square  $20 \times 20 \mu\text{m}$  PLGA  $\mu$ PL loaded with RSP were fabricated using a template-based fabrication approach. Specifically, two different  $\mu$ PL configurations were realized, characterized by a thickness of either 10  $\mu\text{m}$  (short  $\mu$ PL) or 20  $\mu\text{m}$  (tall  $\mu$ PL), and a polymer content of 20 mg and 60 mg, respectively. The difference in the height and polymer content of the  $\mu$ PL was reflected on the RSP encapsulation efficiency and release profile. Indeed, the higher PLGA mass and thickness of the tall  $\mu$ PL led to a larger drug encapsulation and longer sustained release as compared to short  $\mu$ PL. To test the in vivo efficacy,  $\mu$ PL were intraperitoneally injected in heterozygous knockout mice for *dysbindin-1*, a clinically relevant mouse model of cognitive and psychiatric liability, with proved evidence of modulatory responses to antipsychotic drugs in mice and human patients. At 2-, 4-, 8-, and 12-week post-treatment initiation, the ability of the animals to discriminate objects was assessed by performing temporal order object recognition tests. As for the in vitro pharmacological properties, the fine tuning of the  $\mu$ PL geometry and polymer content did affect the in vivo therapeutic performances. While short  $\mu$ PL were able to rescue the cognitive deficit of mice for up to 14 days, the tall  $\mu$ PL were effective up to 3 months. Finally, these findings suggest that RSP- $\mu$ PL can be used as a long-acting depot capable of releasing antipsychotic drugs in a controlled manner up to several weeks and ameliorating the *dysbindin*-induced deficits with one single administration for at least 12 weeks. Further studies of  $\mu$ PL will investigate alternative methods of fabrication to improve LE and EE, without altering the particle efficacy. This would lead to the possibility of injecting even lower amounts of polymer, thus improving the overall safety of the  $\mu$ PL.

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**Availability of data and material** The raw data required to reproduce these results are available from the corresponding author upon reasonable request.

**Code availability** Not applicable.

## Declarations

**Ethics approval** Not applicable.

**Consent to participate** Not applicable.

**Consent for publication** All authors have given their consent for publication.

**Conflict of interest** The authors declare no competing interests.

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