



Opinion Association between Molecular Mechanisms and Tooth Eruption in Children with Obesity

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Abstract: Different works have reported earlier permanent teething in obese/overweight children compared to control ones. In contrast, others have reported a delayed permanent teething in undernutrition/underweight children compared to control one. It has been reported that becoming overweight or suffering from obesity can increase gingival pro-inflammatory drive and can affect orthodontic treatment (among other complications). In this sense, little is known about the molecular mechanisms affecting dental eruption timing. Leptin and adiponectin are adipocytokines signaling molecules released in overweight and underweight conditions, respectively. These adipocytokines can modulate osteocyte, odontoblast, and cementoblast activity, even regulating dental lamina initiation. The present review focuses on the molecular approach wherein leptin and adiponectin act as modulators of Runt-related transcription factor 2 (Runx 2) gene regulating dental eruption timing.

Keywords: pediatric obesity; tooth eruption; dentition; permanent; leptin



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1. Introduction

According to the World Health Organization (WHO) about 39 million children under the age of five suffer from being obese/overweight. On the other side, undernutrition or malnourishment is present in about 149 million children under the age of five. This paradox represents a significant burden affecting several individual health problems and psychosocial aspects among others.

Focusing on oral related hygiene, one's nutritional state can affect different dental aspects from caries to malocclusion [1]. Some research works conclude that one's nutritional state can also modify dental eruption timing. It has been shown that children who are obese and overweight can experience early permanent tooth eruption [2–5]. In contrast, malnourishment is associated with delayed teething [6–9]. Apart from genetic and ethnic differences, a lack of essential nutrients and vitamins seems to be directly related to the delayed teething observed in underweight kids. On the other side, the role of leptins as potential causes of early teething in obese kids is part of a weak rationale for these observed phenomena. In spite of the amount of data confirming these two dental alterations, little biological arguments have been singled out as potentially involved mechanisms. Herein, our proposal strongly suggests the involvement of leptin/adiponectin as pivotal elements of this phenomenon.

2. Dental Development and Permanent Dentition

Teeth development results from a complicated interaction of the odontogenic epithelium and the ectomesenchyme coming from the neural crest in the jaw/maxilla [10,11]. Initially, primary dentition includes incisors, canines, and molars. These are accompanied by a successional lamina that leads the permanent teeth development. However, the secondary molar dentition is developed by serial addition produced by the extension of the dental lamina in the first molar [12]. Even the dental laminae of permanent teeth can be already found at embryonic stages and can last up to 12 years [13]. During these years the dental successional lamina is normally in a resting state and is activated according to deciduous teeth lost [14]. During this process, some genes and molecules are orchestrated. Among them, Wnt, fibroblast growth factor (FGF) and Hedgehog signaling pathways have been demonstrated to regulate, beyond the bud stage, permanent tooth initiation [15–17]. More recently it has been demonstrated that permanent tooth initiation is promoted by mechanical stress release. This mechanical stress inhibits permanent tooth initiation due to the Runt-related transcription factor Wnt (RUNX-Wnt) pathway [18]. Briefly, the pressure exerted by primary teeth activates Runx2, inhibiting successional dental lamina. This blocks permanent tooth initiation. In contrast, the progressive relief of mechanical pressure during deciduous teeth loss decreases Runx2 and increases Wnt expression, leading to permanent tooth initiation.

More recently it has been demonstrated that permanent tooth initiation is promoted by mechanical stress release. This mechanical stress inhibits permanent tooth initiation due to the RUNX-Wnt pathway [18]. Briefly, the biomechanical stress of the primary teeth activates Runx2 inhibiting successional dental lamina. This blocks permanent tooth initiation. In contrast, the progressive relief of mechanical pressure during deciduous teeth loss decreases Runx2 and increases Wnt expression leading to permanent tooth initiation. In line with this, Li et al. [19] pointed out that Runx must be inhibited to promote odontoblast maturation and dentin formation.

3. Runx and Energy State

Runt-related transcription factor (RUNX) is a family with three related transcription factor tors, runt-related transcription factor 1 (RUNX1), RUNX2, and runt-related transcription factor 3 (RUNX3). It has a high conserved sequence of 128 amino acid DNA binding/protein-protein domains, known as the Runt-homology domain [20,21]. RUNX2 determines osteoblastosteocyte differentiation and regulates chondrocyte division-differentiation during endochondral bone development [22]. The Runx2 pathway is connected to integrin β 1 on the cell membrane. Once integrins are stimulated, it promotes extracellular signal-regulated kinase1 (ERK1) activation [23], resulting in Runx 2 transcription and phosphorylation [24]. Fosfatidilinositol 3 kinasa/Akt signalling pathway activation promotes Runt-related transcription factor 2 deoxyribonucleic acid (Runx2-DNA) binding and Runx2 transcription in murine osteoblasts [25].

Although falling outside of the scope of this review, Runx has been widely demonstrated to be related to tumor development, cancer progression, and metastasis in different organs [26,27].

AMP-activated kinase (AMPK) is considered a cellular energy sensor that guides signalling mechanisms leading to homeostatic balance via anabolic or catabolic pathways [28]. RUNX2 is a substrate of AMPK, which directly phosphorylates at serine 118 residue in the DNA-binding domain of RUNX2 [29]. It has been proven that high glucose levels reduced AMPK activity and this fact promotes adipogenesis vs. osteogenesis [30]. Fitting with this, high fat levels also decrease Runx2 expression [31] and even adiponectin increases Runx-2 activity [32]. Adiponectin and leptin are adipocytokines secreted by adipose tissue modulating several functions from cardiovascular modulation to bone metabolism [33,34]. It is generally accepted that obesity is associated to high leptin and low adiponectin levels and inversely for undernutrition [35].

Leptin secretion is increased with feeding and overnutrition when adipocyte number and size is increased [35]. Leptins can affect different pathways, the most relevant of which is the β -oxidation of fatty acids by activating AMP-dependent kinase [36]. It seems that adipose tissue overgrowth results in hypoxia by low vascularization increasing hypoxia inducible factor 1 alpha (HIF1 α) and then raising leptin production [37]. However, undernutrition and physical activity both increased adiponectin secretion decreasing adipose tissue volume due to lipolytic activity [38] (Figure 1).



Figure 1. Under obesity/overweight conditions, early permanent teeth eruption would be explained by increased leptin levels that decrease runt-related transcription factor 2 (Runx 2) expression increasing Wnt gene expression, receptor activator of NF kappa-b (RANK-L), and TNF- α . In underweight conditions, delayed permanent tooth eruption would be explained by increased adiponectin levels or decreased adiponectin. This promotes Runx 2 expression resulting in Wnt reduced expression and decreased pro-inflammatory RANK-L and TNF- α .

Kapur et al. [39] indicated that leptin receptors (LEPR) negatively modulate bone mechanosensitivity and that genetic variation in LEPR signaling causes a low osteogenic response to loading force. According to Um et al. [40] leptins can promote cementoblast/odontoblast differentiation in dental mesenchymal cells. Periodontal ligament fibroblasts over expressed pro-inflammatory factors such as the receptor activator of Nf-Kappa b (RANKL) in the presence of leptin under mechanical strain [41]. Leptin levels can be found and detected in gingival crevicular grooves in healthy subjects compared to those with periodontal disease [42]. Even more, during orthodontic treatment, leptin levels are increased one day after intervention and decreased one week after [43]. Despite the fact that most clinicians suggest a relationship between obesity and tooth movement, after reviewing several studies, it cannot be confirmed that obesity affects tooth movement [44]. Saloom et al. [45] published a prospective clinical cohort study with 55 teenagers (obese vs. normal-weight) and observed significant higher tooth movement in the obese group. Even more, they also found significantly higher levels of leptin and RANKL in this group. These findings support again the potential role of leptins on periodontal and dental evolution and revive the discussion of obesity and its role in orthodontic movement.

Secreted by fat cells and salivary gland epithelial cells [46]. Adiponectin can be bound to adiponectin receptor 1 (AdipoR1) and adiponectin receptor 2 (AdipoR2) receptor types activating the adenosine monophosphate-activated protein kinase pathway (among others) [47,48]. Adiponectin increases fatty acid oxidation, glucose uptake, insulin sensitivity, and can also present anti-inflammatory effects [34,49,50]. Both adiponectin receptors are found in periodontal ligament fibroblasts and osteoblasts [51,52]. According to Marjan Nokhbehsaim et al. [53] adiponectin promotes beneficial effects on periodontal ligament cells by increasing growth factor production and self-promoting adiponectin production. It has been shown that high adiponectin levels increase Runx-2 in cementoblasts, as well as promoting osteoblast differentiation and migration [32,54–56]. In this work, Yong et al. [32] also reported that high adiponectin level exposure increases alkaline phosphatase, osteocalcin, bone sialoprotein, osteocalcin and osteoprotegerin nucleic messenger (mRNA) levels. In contrast, the use of physiological adiponectin concentrations did not result in as significant a response. This means that adiponectin potentially modulates many periodontal-related factors (albeit without playing an exclusive role).

On the other hand, adiponectin decreases pro-inflammatory factors (e.g., tumor necrosis factor alpha). In this sense, Kraus et al. [57] pointed out that low levels of adiponectin in obese/overweight individuals could be related to periodontal inflammation and destruction.

Adiponectin is also involved in cell homeostasis by regulating the mitogen activated protein kinase pathway (MAPK) [58]. Luo et al. [59] suggest that adiponectin receptor-JNK pathway regulates osteoblast proliferation and that adiponectin receptor-P38 modulates differentiation. Previously, Kadowaki et al. [60] indicated that adiponectin stimulated osteogenesis involving adiponectin receptor 2 (P38-AdipoR1) and by increasing Runx-2.

In the literature we also find several environmental factors that affect the tooth eruption. As is well known, tooth eruption is a long lasting process (it often lasts years) and there are many factors that can modify this process (Figure 2). One can see how an overweight child or an obese child may have an advanced tooth eruption process [3–5,8,61–65].



Figure 2. Relationship between obesity and non-molecular factors [3–5,8,61–68].

4. Conclusions

In spite of the amount of data indicating that being obese/overweight can promote early permanent tooth eruption, on the contrary undernutrition leads to delayed permanent teething. There is a considerable lack of knowledge and rationale about the molecular signals giving response to these phenomena.

This review investigates the potential relation of leptin and adiponectin as molecular modulators of dental development. It has been demonstrated that leptins are present in the crevicular fluid of healthy subjects and that leptin levels can be altered during orthodontic mechanical strain. Adiponectin and leptin can also promote osteoblast and odontoblast differentiation. We consider Runx 2 as a potential regulator on this phenomenon since it is a substrate of AMPK. AMPK works as energy sensor and regulates Runx 2. Runx 2 acts as a bone and dental development regulator [18,19]. Both adiponectin and leptin can also affect Runx 2 activity, and this has been proposed as an approach to explain early or delayed dental eruption for both overweight and underweight children.

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References

- 1. Arid, J.; Vitiello, C.; Da Silva, R.A.; Da Silva, L.A.; Mussolino, A.; Calvano, E.; Nelso-Filho, P. Nutritional Status Is Associated with Permanent Tooth Eruption Chronology. *Braz. J. Oral Sci.* 2017, *16*, 1–7. [CrossRef]
- 2. Mohamedhussein, N.; Busuttil-Naudi, A.; Mohammed, H.; UlHaq, A. Association of obesity with the eruption of first and second permanent molars in children: A systematic review. *Eur. Arch. Paediatr. Dent.* **2020**, *21*, 13–23. [CrossRef] [PubMed]
- 3. Nicholas, C.L.; Kadavy, K.; Holton, N.E.; Marshall, T.; Richter, A.; Southard, T. Childhood body mass index is associated with early dental development and eruption in a longitudinal sample from the Iowa Facial Growth Study. *Am. J. Orthod. Dentofac. Orthop.* **2018**, *154*, 72–81. [CrossRef]
- Sánchez-Pérez, L.; Irigoyen, M.; Zepeda, M. Dental Caries, Tooth Eruption Timing and Obesity: A Longitudinal Study in A Group of Mexican Schoolchildren. Acta Odontol. Scand. 2009, 68, 57–64. [CrossRef] [PubMed]
- Traver-Ferrando, C.; Barcia-González, J. Early Permanent Dental Eruption in Obese/Overweigh Schoolchildren. J. Clin. Exp. Dent. 2022, 14, 199–204. [CrossRef]
- Dimaisip-Nabuab, J.; Duijster, D.; Benzian, H.; Heinrich-Weltzien, R.; Homsavath, A.; Monse, B.; Sithan, H.; Stauf, N.; Susilawati, S.; Kromeyer-Hauschild, K. Nutritional Status, Dental Caries and Tooth Eruption in Children: A Longitudinal Study in Cambodia, Indonesia and Lao PDR. *BMC Pediatrics* 2018, 18, 300. [CrossRef]
- Psoter, W.; Gebrian, B.; Prophete, S.; Reid, B.; Katz, R. Effect of Early Childhood Malnutrition on Tooth Eruption in Haitian Adolescents. *Community Dent. Oral Epidemiol.* 2008, 36, 179–189. [CrossRef] [PubMed]
- Evangelista, S.; Regina, K.; Vasconcelos, F.; Xavier, A.; Oliveira, S.; Luiz, A.; Nelso-Filho, P.; Da Silva, L.A.; Da Silva, R.A.; Bezerra, R.A.; et al. Timing of Permanent Tooth Emergence is Associated with Overweight/Obesity in Children from the Amazon Region. *Braz. Dent. J.* 2018, 29, 465–468. [CrossRef] [PubMed]
- Reis, C.L.B.; Barbosa, M.C.F.; Henklein, S.; Madalena, I.R.; de Lima, D.C.; Oliveira, M.A.H.M.; Küchler, E.C.; de Oliveira, D.S.B. Nutritional Status is Associated with Permanent Tooth Eruption in a Group of Brazilian School Children. *Glob. Pediatr. Health* 2021, *8*, 1–6. [CrossRef]
- Jernvall, J.; Thesleff, I. Reiterative Signaling and Patterning During Mammalian Tooth Morphogenesis. *Mech. Dev.* 2000, 92, 19–29. [CrossRef]
- Tucker, A.S.; Fraser, G.J. Evolution and Developmental Diversity of Tooth Regeneration. Semin. Cell Dev. Biol. 2014, 25, 71–80. [CrossRef]
- 12. Juuri, E.; Jussila, M.; Seidel, K.; Holmes, S.; Wu, P.; Richman, J.; Heikinheimo, K.; Chuong, C.M.; Arnold, K.; Hochedlinger, K. Sox2 Marks Epithelial Competence to Generate Teeth in Mammals and Reptiles. *Development* **2013**, *140*, 1424–1432. [CrossRef]
- 13. Pansky, B. Development of the Teeth. In Medical Embryology; Embryome Sciences, Inc.: Alameda, CA, USA, 1982; Volume 6, p. 77.
- Fraser, G.J.; Graham, A.; Smith, M.M. Developmental and evolutionary origins of the vertebrate dentition: Molecular controls for spatio-temporal organisation of tooth sites in osteichthyans. J. Exp. Zool. Part B Mol. Dev. Evol. 2006, 306, 183–203. [CrossRef] [PubMed]
- Klein, O.D.; Minowada, G.; Peterkova, R.; Kangas, A.; Yu, B.D.; Lesot, H.; Peterka, M.; Jernvall, J.; Martin, G.R. Sprouty, Genes Control Diastema Tooth Development Via Bidirectional Antagonism of Epithelial-Mesenchymal FGF Signaling. *Dev. Cell* 2006, 11, 181–190. [CrossRef]

- Ohazama, A.; Haycraft, C.J.; Seppala, M.; Blackburn, J.; Ghafoor, S.; Cobourne, M.; Martinelli, D.; Fan, C.M.; Peterkova, R.; Lesot, H.; et al. Primary Cilia Regulate Shh Activity in the Control of Molar Tooth Number. *Development* 2009, 136, 897–903. [CrossRef] [PubMed]
- 17. Ahn, Y.; Sanderson, B.W.; Klein, O.D.; Krumlauf, R. Inhibition of Wnt Signaling by Wise (Sostdc1) And Negative Feedback from Shh Controls Tooth Number and Patterning. *Development* **2010**, *137*, 3221–3231. [CrossRef]
- Wu, X.; Hu, J.; Li, G.; Li, Y.; Li, Y.; Zhang, J.; Wang, F.; Li, A.; Hu, L.; Fan, Z.; et al. Biomechanical Stress Regulates Mammalian Tooth Replacement Via the Integrin B1-RUNX2-Wnt Pathway. *EMBO J.* 2020, *39*, e102374. [CrossRef] [PubMed]
- Li, S.; Kong, H.; Yao, N.; Yu, Q.; Wang, P.; Lin, Y.; Wang, J.; Kuang, R.; Zhao, X.; Xu, J.; et al. The Role of Runt-Related Transcription Factor 2 (Runx2) in the Late Stage of Odontoblast Differentiation and Dentin Formation. *Biochem. Biophys. Res. Commun.* 2011, 410, 698–704. [CrossRef]
- Blyth, K.; Vaillant, F.; Jenkins, A.; McDonald, L.; Pringle, M.A.; Huser, C.; Stein, T.; Neil, J.; Cameron, E.R. Runx2 in Normal Tissues and Cancer Cells: A Developing Story. *Blood Cells Mol. Dis.* 2010, 45, 117–123. [CrossRef]
- Sun, S.S.; Zhang, L.; Yang, J.; Zhou, X. Role of Runt-Related Transcription Factor 2 In Signal Network of Tumors as an Inter-Mediator. *Cancer Lett.* 2015, 361, 1–7. [CrossRef]
- Chen, H.; Ghori-Javed, F.Y.; Rashid, H.; Adhami, M.D.; Serra, R.; Gutierrez, S.E.; Javed, A. Runx2 regulates endochondral ossification through control of chondrocyte proliferation and differentiation. J. Bone Miner. Res. 2014, 29, 2653–2665. [CrossRef]
- 23. Schlaepfer, D.D.; Hanks, S.K.; Hunter, T.; van der Geer, P. Integrin-Mediated Signal Transduction Linked to Ras Pathway by GRB2 Binding to Focal Adhesion Kinase. *Nature* **1994**, *372*, 786–791. [CrossRef]
- Ren, D.; Wei, F.; Hu, L.; Yang, S.; Wang, C.; Yuan, X. Phosphorylation of Runx2, Induced by Cyclic Mechanical Tension Via ERK1/2 Pathway, Contributes to Osteodifferentiation of Human Periodontal Ligament Fibroblasts. J. Cell. Physiol. 2015, 230, 2426–2436. [CrossRef]
- Fujita, T.; Azuma, Y.; Fukuyama, R.; Hattori, Y.; Yoshida, C.; Koida, M.; Ogita, K.; Komori, T. Runx2 Induces Osteoblast and Chondrocyte Differentiation and Enhances Their Migration by Coupling with PI3K-Akt Signaling. J. Cell Biol. 2004, 166, 85–95. [CrossRef]
- Cohen-Solal, K.A.; Boregowda, R.K.; Lasfar, A. RUNX2 and the PI3K/AKT Axis Reciprocal Activation as a Driving Force for Tumor Progression. *Mol. Cancer* 2015, 14, 137. [CrossRef]
- 27. Chuang, L.S.H.; Ito, Y. The Multiple Interactions of RUNX with the Hippo-YAP Pathway. Cells 2021, 10, 2925. [CrossRef]
- Hardie, D.G.; Ross, F.A.; Hawley, S.A. AMPK: A Nutrient and Energy Sensor That Maintains Energy Homeostasis. Nat. Rev. Mol. Cell Biol. 2012, 13, 251–262. [CrossRef]
- 29. Chava, S.; Chennakesavulu, S.; Gayatri, B.M.; Reddy, A.B.M. A Novel Phosphorylation By AMP-Activated Kinase Regulates RUNX2 From Ubiquitination in Osteogenesis Over Adipogenesis. *Cell Death Dis.* **2018**, *9*, 754. [CrossRef]
- Garcia, D.; Shaw, R.J. AMPK: Mechanisms of Cellular Energy Sensing and Restoration of Metabolic Balance. *Mol. Cell.* 2017, 66, 789–800. [CrossRef]
- Wu, X.; Zhang, Y.; Xing, Y.; Zhao, B.; Zhou, C.; Wen, Y.; Xu, X. High-Fat and High-Glucose Microenvironment Decreases Runx2 and TAZ Expression and Inhibits Bone Regeneration in the Mouse. J. Orthop. Surg. Res. 2019, 14, 55. [CrossRef]
- 32. Yong, J.; Von Bremen, J.; Ruiz-Heiland, G.; Ruf, S. Adiponectin Interacts In-Vitro with Cementoblasts Influencing Cell Migration, proliferation and cementogenesis partly through the MAPK signaling pathway. *Front. Pharmacol.* **2020**, *11*, 585346. [CrossRef]
- Scherer, P.E.; Williams, S.; Fogliano, M.; Baldini, G.; Lodish, H.F. A Novel Serum Protein Similar to C1q, Produced Exclusively in Adipocytes. J. Biol. Chem. 1995, 270, 26746–26749. [CrossRef]
- 34. Fantuzzi, G. Adiponectin in Inflammatory and Immune-Mediated Diseases. Cytokine 2013, 64, 1–10. [CrossRef]
- 35. Unger, R.H. Lipotoxic diseases. Annu. Rev. Med. 2002, 53, 319–336. [CrossRef]
- Minokoshi, Y.; Kim, Y.B.; Peroni, O.D.; Fryer, L.G.; Müller, C.; Carling, D.; Kahn, B.B. Leptin Stimulates Fatty-Acid Oxidation by Activating AMP-Activated Protein Kinase. *Nature* 2002, 415, 339–343. [CrossRef]
- Sun, K.; Halberg, N.; Khan, M.; Magalang, U.J.; Scherer, P.E. Selective Inhibition of Hypoxia-Inducible Factor 1α Ameliorates Adipose Tissue Dysfunction. *Mol. Cell Biol.* 2013, 33, 904–917. [CrossRef]
- Miyazaki, S.; Izawa, T.; Ogasawara, J.E.; Sakurai, T.; Nomura, S.; Kizaki, T.; Ohno, H.; Komabayashi, T. Effect of Exercise Training on Adipocyte-Size-Dependent Expression of Leptin and Adiponectin. *Life Sci.* 2010, *86*, 691–698. [CrossRef]
- Kapur, S.; Amoui, M.; Kesavan, C.; Wang, X.; Mohan, S.; Baylink, D.J.; Lau, K.H. Leptin Receptor (Lepr) Is a Negative Modulator of Bone Mechanosensitivity and Genetic Variations in Lepr May Contribute to The Differential Osteogenic Response to Mechanical Stimulation in the C57BL/6J And C3H/Hej Pair of Mouse Strains. J. Biol. Chem. 2010, 285, 37607–37618. [CrossRef]
- 40. Um, S.; Choi, J.R.; Lee, J.H.; Zhang, Q.; Seo, B. Effect of Leptin on Differentiation of Human Dental Stem Cells. *Oral Dis.* **2011**, 17, 662–669. [CrossRef]
- 41. Schröder, A.; Meyer, A.; Spanier, G.; Damanaki, A.; Paddenberg, E.; Proof, P.; Kirschneck, C. Impact of Leptin on Periodontal Ligament Fibroblasts during Mechanical Strain. *Int. J. Mol. Sci.* **2021**, *22*, 6847. [CrossRef]
- 42. Johnson, R.B.; Serio, F.G. Leptin within Healthy and Diseased Human Gingiva. J. Periodontol. 2000, 72, 1254–1257. [CrossRef]
- Srinivasan, B.; Chitharanjan, A.; Kailasam, V.; Lavu, V.; Ganapathy, V. Evaluation of Leptin Concentration in Gingival Crevicular Fluid (GCF) During Orthodontic Tooth Movement and Its Correlation to the Rate of Tooth Movement. J. Orthod. Sci. 2019, 8, 6. [CrossRef]

- 44. Consolaro, A. Obesity and Orthodontic Treatment: Is There Any Direct Relationship? *Dent. Press J. Orthod.* 2017, 22, 21–25. [CrossRef]
- 45. Saloom, H.F.; Papageorgiou, S.N.; Carpenter, G.H.; Cobourne, M.T. Impact of Obesity on Orthodontic Tooth Movement in Adolescents: A Prospective Clinical Cohort Study. J. Dent. Res. 2017, 96, 547–554. [CrossRef]
- Katsiougiannis, S.; Kapsogeorgou, E.K.; Manoussakis, M.N.; Skopouli, F.N. Salivary Gland Epithelial Cells: A New Source of The Immunoregulatory Hormone Adiponectin. *Arthritis Rheum. Off. J. Am. Coll. Rheumatol.* 2006, 54, 2295–2299. [CrossRef]
- Cheng, K.K.; Lam, K.S.; Wang, B.; Xu, A. Signaling Mechanisms Underlying the Insulin-Sensitizing Effects of Adiponectin. Best Pract. Res. Clin. Endocrinol. Metab. 2014, 28, 3–13. [CrossRef]
- 48. Yamauchi, T.; Iwabu, M.; Okada-Iwabu, M.; Kadowaki, T. Adiponectin Receptors: A Review of Their Structure, Function and How They Work. *Best Pract. Res. Clin. Endocrinol. Metab.* **2014**, *28*, 15–23. [CrossRef]
- 49. Carbone, F.; La Rocca, C.; Matarese, G. Immunological Functions of Leptin and Adiponectin. *Biochimie* **2012**, *94*, 2082–2088. [CrossRef]
- 50. Villarreal-Molina, M.T.; Antuna-Puente, B. Adiponectin: Anti-Inflammatory and Cardioprotective Effects. *Biochimie* 2012, *94*, 2143–2149. [CrossRef]
- Iwayama, T.; Yanagita, M.; Mori, K.; Sawada, K.; Ozasa, M.; Kubota, M.; Miki, K.; Kojima, Y.; Takedachi, M.; Kitamura, M.; et al. Adiponectin regulates functions of gingival fibroblasts and periodontal ligament cells. *J. Periodontal Res.* 2012, 47, 563–571. [CrossRef]
- 52. Yamaguchi, N.; Hamachi, T.; Kamio, N.; Akifusa, S.; Masuda, K.; Nakamura, Y.; Nonaka, K.; Maeda, K.; Hanazawa, S.; Yamashita, Y. Expression levels of adiponectin receptors and periodontitis. *J. Periodontal Res.* **2010**, *45*, 296–300. [CrossRef]
- 53. Nokhbehsaim, M.; Keser, S.; Nogueira, A.V.B.; Cirelli, J.; Jepsen, S.; Jäger, A.; Eick, S.; Deschner, J. Beneficial Effects of Adiponectin on Periodontal Ligament Cells under Normal and Regenerative Conditions. *J. Diabetes Res.* 2014, 2014, 11. [CrossRef]
- 54. Bosshardt, D.D. Are Cementoblasts a Subpopulation of Osteoblasts or A Unique Phenotype? J. Dent. Res. 2005, 84, 390–406. [CrossRef]
- 55. Liu, T.M.; Lee, E.H. Transcriptional Regulatory Cascades in Runx2-Dependent Bone Development. *Tissue Eng. Part B Rev.* 2013, 19, 254–263. [CrossRef]
- 56. Hakki, S.S.; Bozkurt, S.B.; Türkay, E.; Dard, M.; Purali, N.; Götz, W. Recombinant Amelogenin Regulates the Bioactivity of Mouse Cementoblasts In Vitro. *Int. J. Oral Sci.* 2018, *10*, 15. [CrossRef]
- 57. Kraus, D.; Winter, J.; Jepsen, S.; Jäger, A.; Meyer, R.; Deschner, J. Interactions of Adiponectin and Lipopolysaccharide from Porphyromonas Gingivalis on Human Oral Epithelial Cells. *PLoS ONE* **2012**, *7*, e30716. [CrossRef]
- 58. Jernvall, J.; Thesleff, I. Tooth Shape Formation and Tooth Renewal: Evolving with The Same Signals. *Development* **2012**, *139*, 3487–3497. [CrossRef]
- 59. Luo, X.H.; Guo, L.J.; Yuan, L.Q.; Xie, H.; Zhou, H.D.; Wu, X.P.; Liao, E.Y. Adiponectin Stimulates Human Osteoblasts Proliferation and Differentiation Via the MAPK Signaling Pathway. *Exp. Cell Res.* **2005**, *309*, 99–109. [CrossRef]
- 60. Kadowaki, T.; Yamauchi, T.; Kubota, N.; Hara, K.; Ueki, K.; Tobe, K. Adiponectin and Adiponectin Receptors in Insulin Resistance, Diabetes, And the Metabolic Syndrome. *J. Clin. Investig.* **2006**, *116*, 1784–1792. [CrossRef]
- 61. Must, A.; Phillips, S.M.; Tybor, D.J.; Lividini, K.; Hayes, C. The Association between Childhood Obesity and Tooth Eruption. *Obesity* **2012**, *20*, 2070–2074. [CrossRef]
- 62. Shaweesh, A.I.; Alsoleihat, F.D. Association between Body Mass Index and Timing of Permanent Tooth Emergence in Jordanian Children and Adolescents. *Int. J. Stomatol. Occlusion Med.* **2013**, *6*, 50–58. [CrossRef]
- 63. Wong, H.M.; Peng, S.-M.; Yang, Y.; King, N.M.; McGrath, C.P.J. Tooth Eruption and Obesity in 12-Year-Old Children. J. Clin. Investig. 2016, 12, 126–132. [CrossRef]
- 64. Pahel, B.; Vann, W.J.; Divaris, K.; Rozier, R. A Contemporary Examination of First and Second Permanent Molar Emergence. J. Dent. Res. 2017, 96, 1115–1121. [CrossRef] [PubMed]
- ŠindeSindelářová, R.; Soukup, P.; Broukal, Z. The Relationship of Obesity to the Timing of Permanent Tooth Emergence in Czech Children. Acta Odontol. Scand. 2018, 76, 220–225. [CrossRef]
- 66. Khan, N. Eruption Time of Permanent Teeth in Pakistani Children. Iran. J. Public Heal. 2011, 40, 63–73.
- 67. Katmskar, R. Forensic Medicine and Toxicology: Theory, Oral & Practical, 5th ed.; Academic Publishers: Cambridge, MA, USA, 2015; p. 612.
- 68. Garmash, O. Dependence of Deciduous Tooth Eruption Terms and Tooth Growth Rate on the Weight-Height Index at Birth in Macrosomic Children over the First Year of Life. *ACTA MEDICA* **2019**, *62*, 62–68. [CrossRef]