





Complete Genome Sequences of Two *Listeria* Phages of the Genus Pecentumvirus

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ABSTRACT Bacteriophages isolated from environmental sources can be used as a biocontrol against the foodborne pathogen Listeria monocytogenes. Here, we present the complete genomes of LP-039 and LP-066, two Pecentumvirus bacteriophages that infect L. monocytogenes. The genome sizes of LP-039 and LP-066 are 136.2 kb and 139.0 kb, respectively.

isteria monocytogenes nas causeu 1,131 miccions ... 2010 and 2018, with a 17% mortality rate (1), and was mainly associated with isteria monocytogenes has caused 1,151 infections in the United States between contaminated dairy or fruit products (2). Lytic Listeria phages, such as Pecentumvirus P100, are used to control L. monocytogenes in the food industry (3-7). Phages LP-039 and LP-066 are from a collection of Listeria phages previously isolated from silage samples obtained from New York dairy farms using L. monocytogenes strain MACK as previously described (8). These phages are of interest because they exhibit activity against two phage-resistant L. monocytogenes strains (9).

Phage DNA was isolated using a phenol-chloroform method (10), and libraries were prepared using Nextera XT kits. Samples were sequenced with an Illumina MiSeq v3 instrument (300-bp paired-end read chemistry; 275 cycles). Total read numbers of 55,642 and 160,826 were obtained for LP-039 and LP-066, respectively. The average read length was 251 bp. Reads were preprocessed with Trimmomatic v0.35 (ILLUMINACLIP: NexteraPE-PE.fa:2:30:10 LEADING:3 TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36) (11) and FastQC v0.11.7 (12). Single contigs were assembled using SPAdes v3.12.0 with the careful option (13). The redundant terminal region of A511 (14) mapped to the contigs at internal loci with higher read coverage (\sim 2× greater) than the rest of the contig. The contigs were reoriented so that these redundant terminal regions were correctly located at both ends. This was confirmed by mapping reads to the reoriented contigs to ensure that coverage across the redundant terminal regions and the newly formed contig junctions (where original contig ends were joined) was consistent with the rest of the assembly. Assembly statistics were determined using QUAST v4.6.3 (15), BBMap v38.88 (16), and SAMtools v0.1.8 (17). The genomes were annotated with RASTtk (customized pipeline, "annotate-proteins-phage" moved above "annotate-proteinskmer-v2") (18), and annotations of the redundant terminal regions were manually added. Relatedness to Pecentumvirus phages (ICTV Master Species List 2018b.v2 [https://talk.ictvonline.org/files/master-species-lists/m/msl/8266]) was determined with the JSpeciesWS average nucleotide identity MUMmer (ANIm) method (19). Variant analysis was performed with McCortex v0.0.3 (20) (k = 101; breakpoint caller; JOINT_CALLING=yes; USE_LINKS=yes) and SnpEff v4.3t (21).

LP-039 and LP-066 have terminally redundant linear genomes with large invariable, noncohesive ends. LP-039 had a total genome assembly length of 136,234 bp (including the 3,208-bp terminal redundancy) with $88\times$ average coverage and 35.9% G+C content. LP-066 had a total genome assembly length of 138,918 bp (including the 3,128-bp terminal redundancy) with 272× average coverage and 35.8% G+C content.

Citation Peters TL, Hudson LK, Song Y, Denes TG. 2019. Complete genome sequences of two Listeria phages of the genus Pecentumvirus. Microbiol Resour Announc 8:e01229-19. https://doi.org/10.1128/MRA.01229-19.

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Received 3 October 2019 Accepted 18 October 2019 Published 14 November 2019

TABLE 1 Results from JSpeciesWS for Pecentumvirus Listeria phages

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	Avg nucleotid	le identity (%)	Avg nucleotide identity (%) shared between phage pairs a	ר phage pairs a									
Phage	A511	AG20	List36	LMSP25	LMTA34	LMTA148	LP-048	LP-064	LP-083-2	P100 ⁶	WIL1	LP-039	LP-066
A511		97.14 (97.12)	97.09 (95.00) 96.39 (95.67)	96.39 (95.67)	(6.39 (95.67)	97.24 (95.51)	97.20 (96.02)	97.15 (97.57)	97.00 (97.84)	97.20 (95.13)	97.45 (98.52)	97.14 (96.03)	96.96 (97.84)
AG20	97.14 (98.43)		97.38 (96.29)	96.30 (96.46)	96.30 (96.46)	97.60 (96.36)	97.50 (98.15)	97.18 (98.39)	97.15 (98.36)	97.14 (96.39)	97.20 (98.00)	97.50 (98.16)	97.14 (98.36)
List36	97.09 (97.05)	97.41 (96.93)		96.56 (96.18)	96.56 (96.18)	97.60 (96.48)	97.41 (97.54)	97.23 (98.24)	97.40 (98.03)	97.16 (97.28)	97.26 (96.72)	97.40 (97.54)	97.40 (98.03)
LMSP25	96.39 (94.25)	96.30 (93.74)	96.56 (92.85)		100.00 (100.00)	96.78 (91.46)	96.42 (92.67)	96.64 (95.30)	96.57 (95.60)	96.71 (93.10)	96.40 (93.96)	96.41 (92.74)	96.59 (95.60)
LMTA34	96.39 (94.25)	96.30 (93.74)	96.56 (92.85)	100.00 (100.00)		96.78 (91.46)	96.42 (92.67)	96.64 (95.30)	96.57 (95.60)	96.71 (93.10)	96.40 (93.96)	96.41 (92.74)	96.59 (95.60)
LMTA148	97.24 (97.25)	97.60 (96.48)	97.60 (96.09)	96.78 (94.32)	96.78 (94.32)		97.69 (98.04)	97.15 (97.01)	97.28 (96.71)	97.28 (95.18)	97.17 (97.07)	97.67 (98.04)	97.26 (96.72)
LP-048	97.20 (97.78)	97.50 (98.13)	97.41 (96.83)	96.42 (95.36)	96.42 (95.36)	97.69 (97.87)		97.14 (98.29)	97.11 (98.30)	97.01 (96.71)	97.28 (97.58)	99.99 (100.00)	97.14 (98.30)
LP0-64	97.15 (97.22)	97.18 (96.76)	97.25 (95.88)	96.64 (96.49)	96.64 (96.49)	97.15 (95.22)	97.18 (96.73)		98.13 (99.05)	98.20 (97.22)	97.33 (97.14)	97.14 (96.74)	98.10 (99.06)
LP-083-2	97.00 (97.05)	97.15 (96.33)	97.37 (95.33)	96.57 (96.33)	96.57 (96.33)	97.28 (94.53)	97.11 (96.27)	98.13 (98.62)		98.32 (96.26)	97.35 (97.12)	97.11 (96.29)	99.98 (100.00)
P100 ⁶	97.20 (97.46)	97.14 (97.49)	97.16 (97.68)	96.71 (96.98)	96.71 (96.98)	97.20 (96.43)	97.01 (97.86)	98.19 (99.97)	98.32 (99.43)		97.34 (97.89)	97.01 (97.86)	98.33 (99.41)
WIL1	97.42 (98.63)	97.22 (96.89)	97.26 (95.11)	96.40 (95.50)	96.40 (95.50)	97.17 (95.73)	97.28 (96.41)	97.33 (97.78)	97.35 (98.07)	97.34 (95.73)		97.28 (96.41)	97.33 (98.07)
LP-039	97.14 (97.31)	97.50 (97.98)	97.40 (96.91)	96.41 (95.15)	96.41 (95.15)	97.67 (97.92)	(100.00)	97.14 (98.14)	97.11 (98.21)	97.01 (96.58)	97.28 (97.23)		97.14 (98.21)
LP-066	96.96 (97.03)	97.14 (96.46)	97.37 (95.48)	96.59 (96.34)	96.59 (96.34)	97.26 (94.60)	97.14 (96.29)	98.10 (98.68)	99.98 (100.00)	98.33 (96.38)	97.33 (97.23)	97.14 (96.31)	

 o Numbers represent the average nucleotide identity across aligned nucleotide percentage (aligned percentage is in brackets). b Type species of the genus p Percentumvirus.



Both genomes contained 193 to 198 coding sequences (each contained 9 duplicate coding sequences due to the terminal redundancy) and 17 tRNAs. LP-039 is closely related to LP-048 (Table 1). Variant analysis of LP-039 compared to LP-048 showed one mutation (a 3-nucleotide deletion) in LP048_062 (hypothetical protein) with potential upstream effects in tRNA genes. LP-066 is closely related to LP-083-2 (Table 1). Variant analysis of LP-066 compared to LP-083-2 showed four mutations. LP083-2_021 and LP083-2_130 (hypothetical proteins) each had one synonymous mutation with minimal predicted effects. One conservative nonsynonymous mutation (G > T) was found at position 1871 in gene LP083-2_152 (DNA polymerase I), producing a valine rather than a glycine. A 15-nucleotide in-frame insertion was found in gene LP083-2_101 (hypothetical protein) near genes involved in recombination. This mutation resulted in the duplication of the amino acid sequence K-E-E-P-K.

LP-039 and LP-066 were independently isolated from similar environmental sources as *Listeria* phages LP-048 and LP-083-2 (22) and are likely examples of recent evolutionary genetic divergence from a common ancestor under natural conditions. Genomic characterization of closely related phages such as the ones presented here will provide valuable information on genetic variation between wild phage strains and may help identify candidate phages for biocontrol applications.

Data availability. These phages are located under BioProject number PRJNA544516 (BioSample numbers SAMN12053438 and SAMN12053439). The raw reads have been deposited in the SRA (accession numbers SRR9597082 and SRR9597083), and the annotated genomes in GenBank (accession numbers MN172529 and MN128594).

ACKNOWLEDGMENTS

This work was supported by the University of Tennessee Institute of Agriculture (experimental startup package for T. G. Denes) and multistate project S1077: Enhancing Microbial Food Safety by Risk Analysis.

We acknowledge our appreciation to Veronica Brown of the UT Genomics Core and Daniel Bryan of the UT Department of Food Science for assistance with sequencing.

REFERENCES

- CDC. 2019. FoodNet fast. CDC, Atlanta, GA. http://wwwn.cdc.gov/foodnetfast. Accessed 30 May 2019.
- IFSAC. 2018. Foodborne illness source attribution estimates for 2016 for Salmonella, Escherichia coli O157, Listeria monocytogenes, and Campylobacter using multi-year outbreak surveillance data, United States. IFSAC, Lexington, KY. https://www.cdc.gov/foodsafety/ifsac/pdf/P19-2016 -report-TriAgency-508.pdf. Accessed 30 May 2019.
- Akhtar M, Viazis S, Christensen K, Kraemer P, Diez-Gonzalez F. 2017. Isolation, characterization and evaluation of virulent bacteriophages against *Listeria monocytogenes*. Food Control 75:108–115. https://doi.org/10.1016/j.foodcont.2016.12.035.
- Gray JA, Chandry PS, Kaur M, Kocharunchitt C, Bowman JP, Fox EM. 2018. Novel biocontrol methods for *Listeria monocytogenes* biofilms in food production facilities. Front Microbiol 9:605. https://doi.org/10.3389/fmicb.2018.00605.
- Strydom A, Witthuhn CR. 2015. Listeria monocytogenes: a target for bacteriophage biocontrol. Compr Rev Food Sci Food Saf 14:694–704. https://doi.org/10.1111/1541-4337.12153.
- Carlton RM, Noordman WH, Biswas B, de Meester ED, Loessner MJ. 2005. Bacteriophage P100 for control of *Listeria monocytogenes* in foods: genome sequence, bioinformatic analyses, oral toxicity study, and application. Regul Toxicol Pharmacol 43:301–312. https://doi.org/10.1016/i.vrtph.2005.08.005.
- Guenther S, Huwyler D, Richard S, Loessner MJ. 2009. Virulent bacteriophage for efficient biocontrol of *Listeria monocytogenes* in readyto-eat foods. Appl Environ Microbiol 75:93–100. https://doi.org/10.1128/AEM.01711-08.
- Vongkamjan K, Switt AM, den Bakker HC, Fortes ED, Wiedmann M. 2012. Silage collected on dairy farms harbors an abundance of listeriaphages with considerable host range and genome size diversity. Appl Environ Microbiol 78:8666–8675. https://doi.org/10.1128/ AEM.01859-12.

- Trudelle DM, Bryan DW, Hudson LK, Denes TG. 2019. Cross-resistance to phage infection in *Listeria monocytogenes* serotype 1/2a mutants. Food Microbiol 84:103239. https://doi.org/10.1016/j.fm.2019.06.003.
- Sambrook J, Russell DW. 2006. Extraction of bacteriophage λ DNA from large-scale cultures using proteinase k and sds. Cold Spring Harb Protoc 2006:pdb.prot3972. https://doi.org/10.1101/pdb.prot3972.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. https://www.bioinformatics.babraham.ac.uk/projects/fastqc/.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Klumpp J, Dorscht J, Lurz R, Bielmann R, Wieland M, Zimmer M, Calendar R, Loessner MJ. 2008. The terminally redundant, nonpermuted genome of *Listeria* bacteriophage a511: a model for the spo1-like myoviruses of Gram-positive bacteria. J Bacteriol 190:5753–5765. https://doi.org/10 .1128/JB.00461-08.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
- 16. Bushnell B. 2018. BBTools. https://jgi.doe.gov/data-and-tools/bbtools/.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. Bioinformatics 25:2078–2079. https://doi.org/10.1093/bioinformatics/btp352.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible

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- implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365–8365. https://doi.org/10.1038/srep08365.
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. Bioinformatics 32:929–931. https://doi.org/10.1093/ bioinformatics/btv681.
- 20. Cingolani P, Platts A, Wang LL, Coon M, Nguyen T, Wang L, Land SJ, Lu X, Ruden DM. 2012. A program for annotating and predicting the effects of
- single nucleotide polymorphisms. Fly (Austin) 6:80-92. https://doi.org/10.4161/fly.19695.
- Turner I, McVean G, Garimella KV, Iqbal Z. 2018. Integrating long-range connectivity information into de bruijn graphs. Bioinformatics 34: 2556–2565. https://doi.org/10.1093/bioinformatics/bty157.
- Denes T, Vongkamjan K, Ackermann H-W, Moreno Switt AI, Wiedmann M, den Bakker HC. 2014. Comparative genomic and morphological analyses of *Listeria* phages isolated from farm environments. Appl Environ Microbiol 80:4616–4625. https://doi.org/10.1128/AEM.00720-14.

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