



Role of Mitochondria in the Redox Signaling Network and Its Outcomes in High Impact Inflammatory Syndromes

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Inflammation is associated with the release of soluble mediators that drive cellular activation and migration of inflammatory leukocytes to the site of injury, together with endothelial expression of adhesion molecules, and increased vascular permeability. It is a stepwise tightly regulated process that has been evolved to cope with a wide range of different inflammatory stimuli. However, under certain physiopathological conditions, the inflammatory response overwhelms local regulatory mechanisms and leads to systemic inflammation that, in turn, might affect metabolism in distant tissues and organs. In this sense, as mitochondria are able to perceive signals of inflammation is one of the first organelles to be affected by a dysregulation in the systemic inflammatory response, it has been associated with the progression of the physiopathological mechanisms. Mitochondria are also an important source of ROS (reactive oxygen species) within most mammalian cells and are therefore highly involved in oxidative stress. ROS production might contribute to mitochondrial damage in a range of pathologies and is also important in a complex redox signaling network from the organelle to the rest of the cell. Therefore, a role for ROS generated by mitochondria in regulating inflammatory signaling was postulated and mitochondria have been implicated in multiple aspects of the inflammatory response. An inflammatory condition that affects mitochondrial function in different organs is the exposure to air particulate matter (PM). Both after acute and chronic pollutants exposure, PM uptake by alveolar macrophages have been described to induce local cell activation and recruitment, cytokine release, and pulmonary inflammation. Afterwards, inflammatory mediators have been shown to be able to reach the bloodstream and induce a systemic response that affects metabolism in distant organs different from the lung. In this proinflammatory environment, impaired mitochondrial function that leads to bioenergetic dysfunction and enhanced production of oxidants have been shown to affect tissue homeostasis and organ function. In the present review, we aim to discuss the latest insights into the cellular and molecular mechanisms that link systemic inflammation and mitochondrial dysfunction in different organs, taking the exposure to air pollutants as a case model.

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INTRODUCTION

Mitochondria have been historically identified as the main source of cellular energy, by coupling the oxidation of fatty acids and pyruvate with the production of adenosine triphosphate (ATP) by the electron transport chain (1, 2). They are complex organelles that play a wide range of functions, including regulation of Ca²⁺ homeostasis, apoptosis, and differentiation (3, 4). Mitochondria are also an important source of ROS (reactive oxygen species) within most mammalian cells and are therefore highly involved in oxidative stress, where increased ROS production might contribute to mitochondrial damage in a range of pathologies (5, 6). They also play a significant role in a redox signaling system where an interplay is displayed from the organelle to the rest of the cell. Recently, new functions for the mitochondria were proposed, particularly linking the alterations in the mechanisms linked to ROS generation with the inflammatory responses involved in different pathological conditions (7-9). In this review, we will discuss the role of the mitochondria in the cellular and molecular mechanisms that link systemic inflammation and mitochondrial dysfunction in different organs, taking the exposure to air pollutants as a case model.

Reactive Oxygen Species and Oxidative Stress

The concept of ROS was originally presented to describe the luminol chemiluminescence of activated human monocytes in 1982 (10). The concept was rapidly adopted by the scientific community, despite the fact that the cited work did not include the definition of the involved chemical species. Originally, ROS comprised superoxide anion $(O_2^{\bullet-})$, hydrogen peroxide (H_2O_2) and hydroxyl radical (HO[•]), which derive from the partial reduction of molecular O_2 (11, 12). When molecular O_2 in its basal state accepts an electron, $O_2^{\bullet-}$ will be the product obtained, a reactive chemical species with only one unpaired electron. Adding a second electron will lead to the formation of the peroxide ion (O_2^{2-}) , from which H_2O_2 is its common form at physiological pH (13, 14). Since H₂O₂ does not have unpaired electrons is less reactive. However, this molecule is considered a reactive O₂ species because the O-O bond is relatively weak (bond energy: 138 kJ/mol). Therefore, it can decompose leading to HO[•] formation, whose reactivity is so high that it reacts very close to its site of formation (14, 15).

In biological systems, the reaction $O_2^{\bullet-}$ with H_2O_2 in the presence of transition metals such as Fe or Cu leads to the formation of HO[•] in a reaction postulated by F. Haber and J. Weiss and known as the Haber-Weiss reaction (16):

$$O_2^{\bullet-} + H_2 O_2 \xrightarrow{Fe^{2+}} O_2 + OH^- + HO^{\bullet}$$
(1)

This reaction proceeds in two consecutive steps: first, Fe^{3+} is reduced by the action of O₂,

$$\operatorname{Fe}^{3+} + \operatorname{O}_2^{\bullet-} \longrightarrow \operatorname{Fe}^{2+} + \operatorname{O}_2$$
 (2)

while in the second step H_2O_2 reacts with Fe^{2+} to produce hydroxyl radical, also known as the Fenton reaction.

$$Fe^{2+} + H_2O_2 \longrightarrow Fe^{3+} + OH^- + HO^{\bullet}$$
 (3)

The Fenton and Haber-Weiss reactions are responsible for the generation of HO^{\bullet} in biological systems and therefore are involved in the pathophysiological mechanisms of diseases where oxidative stress plays a significant role (17).

The concept of ROS was later extended and today it is accepted that these chemical species comprises $O_2^{\bullet-}$, H_2O_2 , and HO[•], some intermediates of the free radical-mediated lipid peroxidation, such as peroxyl radical(ROO•) and singlet oxygen (¹O₂) and also organic peroxides (ROOH) and peroxynitrite (ONOO⁻), the latter product of the reaction between and $O_2^{\bullet-}$ and nitric oxide (NO) (18, 19).

ROS are generated during normal intracellular metabolism in mitochondria and peroxisomes, as well as from a variety of cytosolic enzyme systems.

A complex antioxidant defense system comprising antioxidant enzymes and low molecular mass reductants counteracts and regulates overall ROS at physiological levels (20). The antioxidant enzymes act as a coordinated system and includes superoxide dismutase, catalase, glutathione peroxidase and peroxiredoxins; each one of them comprised by several isoforms with specific substrates and cellular locations (21, 22). Low molecular weight antioxidants are represented by glutathione, tocopherols, ascorbic acid, and carotenoids (23).

When an increased rate of ROS production occurs and the antioxidant system is overwhelmed, a disruption in the redox balance is observed and an oxidative stress situation can be characterized. At first, the adverse effects produced by increased ROS levels were believed to result in oxidative damage to proteins, lipids, and DNA. However, in addition to these effects, the increase over physiological ROS levels may also trigger diverse stress signals that can activate specific redox-sensitive signaling pathways. Once activated, the signaling pathways may have either deleterious effects or potentially adaptive functions (24, 25).

The definition of oxidative stress was first introduced by Sies (26) and recently updated to include the role of redox signaling. It is defined as an imbalance between oxidants and antioxidants in favor of the oxidants, leading to a disruption of redox signaling and control and/or molecular damage (27, 28). Based in this new description and given the enormous variety and range of ROS and related oxidant and antioxidant compounds, efforts were made to introduce a different scale that distinguishes basal (physiological) oxidative stress from a situation where cytotoxic responses are observed (29).

Figure 1 shows how intensity of oxidative stress can be classified into different grades that are related to specific responses and cellular outcomes. The different intensity response to oxidative stress allows biological systems to adequately react to these challenges in a dose-dependent manner (28).



Complex II has also been suggested as a mitochondrial $O_2^{\bullet-}$ production source through the reverse electron transfer, that can be regulated by ATP-sensitive potassium channels and mitochondrial ATP level (38, 39). Mitochondrial $O_2^{\bullet-}$ may also be produced by α -ketoglutarate dehydrogenase, pyruvate dehydrogenase, glycerol-3-phosphate dehydrogenase, fatty acid β-oxidation. Under pathophysiological conditions, changes in bioenergetic states may result in mitochondrial substrate availability alterations, affecting the mitochondrial ROS production source (38, 40). The highly reactive $O_2^{\bullet-}$ is considered as the stoichiometric precursor of mitochondrial H_2O_2 production as $O_2^{\bullet-}$ produce mainly on the matrix side is rapidly dismutated to H₂O₂ by mitochondrial manganese SOD (MnSOD). H₂O₂ is a neutral molecule that can easily diffuse through mitochondrial membranes regardless of the organelle energization. Given the variety of mitochondrial ROS and the bioenergetic conditions requirements, mitochondria are part of a dynamic networks within the cell that involves mitochondrial fission and fusion processes and subcellular trafficking in order to control subcellular location of ATP or ROS release to support specific cell functions (12, 41–43).

consumption (36-38). In complex I, $O_2^{\bullet-}$ is produced through the electron transfer to O₂ during the NADH-dehydrogenase

flavin mononucleotide semiquinone autoxidation, while in

complex III the electron is transferred by the ubiquinone. Under

normal conditions complex I contributes to one third and

complex III to two thirds of mitochondrial $O_2^{\bullet-}$ production.

The NOX represent a family of enzymes whose function is to mediate regulated cellular production of ROS by transferring electrons from NADPH, to reduce O_2 to $O_2^{\bullet-}$. NOX enzymes participate in important biological and pathophysiological processes. Although it is considered the defense against pathogens as the major NOX function, they also participate in inflammation response, cell signaling and regulation of cell growth, differentiation, and death (43). During inflammation NOX complex gets assembled and activated within the phagosomes to generate intraphagosomal ROS in order to kill ingested microorganismos by oxidative mechanisms (43-46). Up to now, seven NOX isoforms have been described: NOX1, NOX2, NOX3, NOX4, NOX5, and two higher molecular weight counterparts called dual oxidases, DUOX 1 and DUOX 2 (47, 48). Despite the fact that all isoforms structure shares the main functional domains, each NOX presents a difference in its regulation, activation and subcellular location. Regarding subcellular localization, different NOX catalytic and regulatory subunits have been detected in diverse cellular membranes and intracellular structures. Isoform NOX1, NOX2, NOX4, and NOX5 have been frequently informed to be found at the plasma membrane (45, 49-51). According to the reports, for NOX4, a constitutively active isoform enzyme, has been detected in the endoplasmic reticulum (52) as well as uniquely localizes to the mitochondria in various endothelial cell types (49, 53–55). Moreover, NOX isoforms also release different oxidant species. For example, NOX1 and NOX2 generate $O_2^{\bullet-}$ (43, 56), while NOX4 is responsible for the basal production of H_2O_2 (57), and NOX5 produces H_2O_2 as well but in a Ca²⁺-dependent fashion (58). The increased NOX activity that leads to an augmented

Mitochondria and NADPH Oxidases as Relevant Sources of ROS

In every eukaryotic cell ROS are generated under normal physiological conditions through enzymatic auto-oxidation reactions, which can involve both endogenous and xenobiotic compounds. The production sources can be localized at various specific subcellular systems such as plasma membrane, cytosol, peroxisomes, or organelle's membranes. Here we will be focused on NADPH oxidases (NOX) (30) and the mitochondrial electron transport chain (31) as they can be considered the predominant production sources in several pathophysiological situations.

Mitochondria are the main responsible for the cellular ATP synthesis by oxidative phosphorylation, as 36 ATP molecules are generated for each glucose molecule that is oxidized, as opposed to the two ATP molecules generated by glycolysis in the cytoplasm. It is based on transfer of electrons through the mitochondrial respiratory chain comprised of a series of integral proteins located on the inner membrane physically and functionally grouped into complexes (complex I, NADH dehydrogenase; complex II, succinate dehydrogenase; ubiquinolcytochrome c oxidoreductase complex III; and complex IV, cytochrome oxidase), allowing a sequential arrangement that facilitates the transfer of electrons between them, determining a high speed and efficiency of the system. The driving force that governs the electron transfer along the complexes is the standard reduction potential of each complex, which depends on the concentrations of the oxidized and reduced forms of each one at a certain pH and ends at the O₂ as the last electron acceptor, which is reduced to H_2O . This process leads to protons (H^+) pump from the matrix to the intermembrane space generating the proton motive force across the inner membrane. It is now known that not only ATP synthesis, but ROS production as well is a biological mitochondrial function, due to their potential key role in cell signaling to support mitochondrial integrity and adaptive responses that control homeostasis and promote health span (32-35). Electrons can leak to O_2 creating $O_2^{\bullet-}$ in different respiratory chain sites as complex I and III, the major site of cellular O₂

production of oxidizing species such as $O_2^{\bullet-}$ and consequently H₂O₂, has been largely associated with various pathological situations (59).

Regarding signaling functions, when ROS are released from NOX, in a regulated and deliberate fashion, they are able to activate tyrosine phosphatase. This enzyme is involved in numerous transcription factors phosphorylation required to modulate different cell proliferation, differentiation, or death pathways (39). All members of the NOX family are multitransmembrane proteins containing a flavocytochrome b558 (gp91 phox) that is associated with another transmembrane protein, p22phox. The gp91phox subunit contains the binding sites for NADPH and FAD and 2 heme groups necessary for transmembrane electrons transport from NADPH to O₂, generating $O_2^{\bullet-}$. Activation of the enzyme depends on the phosphorylation of 3 cytosolic regulatory proteins (p47phox, p67phox, p40phox) and together with GTPase and Rac1 they assemble to the transmembrane domains forming the functional NOX. In this sense, the enzyme goes from being at rest to being quickly activated against different cellular stimuli (47).

Redox Signaling and Crosstalk Between ROS Sources

Under normal physiological conditions, production of ROS is not only down-regulated by several mechanisms, but also highly restricted to specific subcellular sites (12, 39, 60). Compartmentalization of ROS production within cells is important not only in terms of target specificity and selectivity but also in elicit redox signaling or oxidative damage. However, when cells experience pathophysiological situations, an excessive ROS production that overwhelms the antioxidant defense systems, results in cellular dysfunction due to oxidative stress. The cellular redox status has been assessed through different approaches like measurements of GSH status, lipid, proteins and DNA oxidation showing these outcomes association with the development of several diseases such as atherosclerosis, heart failure, hypertension, ischemia/reperfusion injury, cancer, aging, and neurodegeneration (39, 43, 61).

An interplay between specific ROS sources has been recognized, where the consequences of the redox crosstalk, mainly between mitochondria and NOX, is of increased interest in the last years. In this scenario, various ROS sources interaction stimulates each other in a positive feedback fashion, resulting in a complex oxidative stress and redox signaling network (39, 62, 63). It is well-documented that all main ROS sources are, at the same time, regulated by oxidation. ROS production sources enhancement was first described for mitochondria-to-mitochondria communication, where ROS released from one organelle triggers ROS production by another organelle. Oxidative damage of the mitochondrial respiratory chain constituents leads to mild uncoupling resulting in an augmented ROS production (62). Also, inactivation of MnSOD due to oxidation or nitration, increase cytosolic $O_2^{\bullet-}$ levels (64). Therefore, mitochondria are efficient ROS amplifiers that may further feed this vicious cycle. Once ROS within mitochondria reach certain stationary state levels, the organelle is able to display specific mechanisms in order to interact with other mitochondria or ROS sources. Two different mechanisms were proposed to mediate the mitochondrial ROS production enhancement (40). In the first one, an increase of the mitochondrial respiratory chain- ROS release activates the mitochondrial permeability transition pore (mPTP) causing depolarization of the inner and outer membranes, which in turn yields a burst of ROS released to the cytosol (31). A second mechanism involves the direct opening of an inner mitochondrial membrane anion channel allowing ROS to enter the intermembrane space and then released into the cytosol via the voltage-dependent anion channel (40).

The mitochondrial ROS-induced ROS release concept was then widened to communication between different ROS sources like the described mitochondria to NOX crosstalk (63). In recent years, those mechanisms have been observed in different experimental models, namely aging, in response to nitroglycerin therapy, MnSOD deficiency, by angiotensin II, hypoxia or sepsis among others (30, 63, 65, 66). The link can either be triggered by NOX-released ROS to mitochondria or vice versa from mitochondrial site to NOX level, depending on the pathogenesis of the above-mentioned diseases. In both pathways, the opening of the mitochondrial ATP-sensitive potassium channels (mitoK_{ATP}) seems to play an important role and has been tested through the use of inhibitors or channel openers (63). It has been shown mitoKATP becomes activated by NOX-released ROS (67). Opening of the mitoKATP stimulates the potassium influx that shifts the mitochondrial matrix to an alkalization, initiating swelling, mild mitochondrial uncoupling, and ROS production. The opening of mitoKATP, triggers changes in the mitochondrial membrane potential, opening the mitochondrial permeability transition pore (mPTP) which leads to a subsequent mitochondrial ROS release to the cytosol resulting in additional NOX activation in a vicious circle (68). In a mechanism similar to the mitochondria to mitochondria interplay, once mPTP and mitoKATP channels are opened, mitochondrial ROS release into the cytoplasm activates protein kinase C (PKC) leading to NOX ensemble. This interaction was confirmed by inhibition of NOX enzymes, which prevented, for example, the mitochondrial dysfunction induced by angiotensin II (39, 63).

Given the complex cellular redox network depicted in **Figure 2**, better knowledge about the main ROS sources crosstalk potential mechanisms, along with understanding of the switch from redox signaling to oxidative damage will help in the searching for new therapeutic approaches and the development of more target specific antioxidants.

Inflammation and the Role of Mitochondria

Inflammation is a physiological response of the host against infection or tissue injury that occurs in order to eliminate the threat and restore tissue homeostasis, thus resolving the injury/infection and preventing damage progression (69–71). When the innate immune system detects tissue damage or senses a "danger" signal, activates the first line of defense and repair programs which initiate the inflammatory response triggering various mechanisms (72). The inflammation process is induced by a wide range of inflammatory stimuli, such usinfection, trauma, autoimmune disorders, ischemia, chemical, toxin



exposure, among others, and is recognized by a combination of three classical clinical signs: hyperthermia, vasodilation and edema. Despite the cause of damage, harmful stimuli initially activate an acute inflammatory response where tissue-resident macrophages play a key role, not only detecting the infection or injury but also influencing normal tissue homeostasis. This inflammatory response is initially localized at the site of injury and includes distinct cell types, such as neutrophils and macrophages, and leads to the production and release of a variety of soluble inflammatory mediators that will act on tissues and organs affecting their functionality and metabolic state (69, 72-74). These mediators include cytokines and chemokines, that serve to amplify the local response, driving cellular activation and migration of additional inflammatory cells to the damage site. Cytokines can also modulate the function of proximate cells, including secretion of more pro-inflammatory cytokines with synergistic effects (75, 76).

Also, these mediators change vascular permeability and endothelial function, increasing endothelial expression of members of the selectin family of adhesion molecules (L-selectin, E-selectin), that allow the rolling along the vascular endothelium, and the activation of integrins that bind to endothelial vascular adhesion molecules (ICAM-1 and VCAM-1) facilitating the immobilization and transmigration of leukocytes through the activated endothelium at the site of injury (69, 72, 73).

Taking into account the onset and progression of inflammation, it might be considered as either an acute or a chronic process. Physiological inflammation is frequently selflimited, showing a transient abnormal condition characterized by the production of pro-inflammatory cytokines that is followed closely and in a specific temporal manner by the production of anti-inflammatory cytokines with counter-regulating effects that attenuates or resolve the inflammatory process, thus contributing to the restoration of the homeostasis to the tissue and the eradication of the source of damage (75, 76). This counter-part inflammatory response is mediated mainly by tissue-resident and recruited macrophages (activated monocytes). However, when the local inflammatory stimuli persists or the mechanisms of repair fails, chronic inflammation ensues due to sustained pro-inflammatory mechanisms that may lead to a pathological state, usually seen in chronic infections and autoimmune diseases (69, 74).

Moreover, it can also be the case that the inflammatory response overwhelms local regulatory mechanisms and leads to systemic inflammation that, in turn, might affect metabolism in distant tissues and organs and eventually leads to the pathogenesis of inflammatory syndromes.

Chronic systemic inflammation occurs in a variety of severe diseases including cancer, diabetes, cardiovascular diseases and aging-related neurological diseases, and are associated with an imbalance of tissue homeostasis instead of the typical initiators of inflammation (infection, injury) (77, 78). When the inflammatory response is dysregulated and cannot be attenuated organ dysfunction could occur (70, 74).

The inflammatory response is typically initiated by pattern recognition receptors, as Toll-like receptors (TLRs) and NODlike receptors (NLRs), that are expressed on the surface of immune cells like neutrophils and macrophages. Upon recognition of pathogen-associated molecular pattern molecules (PAMPs) and damage-associated molecular pattern molecules (DAMPs) (such as lipopolysaccharide (LPS) of Gram-negative bacteria and nucleic acids from viruses, among others), these receptors activate inflammatory signaling pathways (45). The intracellular cascades triggered by TLRs activate downstream kinases like IkB and MAP kinases that regulate different transcription factors such as NF-kB and AP-1, that ultimately induce the expression of pro-inflammatory genes, such as cytokines (IL-1b, IL-6, TNF-a) and chemokines, that serve to recruit additional immune cells, and proteins like iNOS that generate nitric oxide (NO). There are also other types of receptors specialized in detecting intracellular PAMPs and DAMPs, as mentioned above, the NLRs. These receptors are involved in the formation of the multi-protein signaling complexes known as inflammasomes (79). Activation of NLRP3, the most fully characterized inflammasome due to its association with many inflammatory diseases, drives caspase-1 activation and maturation of pro-IL-1b, thus acting as sensors initiating innate inflammatory responses that are triggered by a variety of danger signals including metabolic stress (80–82). High levels of reactive oxygen species (ROS) showed to activate NLRP3 (71).

When the inflammatory response overwhelms local regulatory mechanisms leading to systemic inflammation, metabolism in distant tissues and organs may be affected. Interestingly, these structures might also show different degrees of sensitivity. In this sense, as mitochondria can perceive signals of inflammation is one of the first organelles to be affected by a dysregulation in the systemic inflammatory response and has been associated with the progression of physiopathological mechanisms. Therefore, there is a growing attention in the biology and medical research field related to mitochondrial modulation during inflammatory syndromes.

Besides the mitochondria major functions of synthesized ATP through the process of oxidative phosphorylation, and the other key cellular events through the generation of ROS, as was described above, the organelle also execute other important roles in regulating cellular apoptosis and modulation of calcium metabolism (83) that in turn regulates the metabolic state of the cell through different signaling pathways. Appropriate mitochondrial function is essential to supply energy requirements of immune cells showing an important role in the immunity regulation (84). In this sense, mitochondria bioenergetics is differentially regulated in activated macrophages (M1) and alternatively (M2) macrophages. M1 macrophages are important for clearance of pathogen infections, while M2 are involved in the termination inflammatory phase showing an anti-inflammatory phenotype (82).

There are several mechanisms by which mitochondria may lead to tissue dysfunction: (a) reduction in cellular high energy [understood as adenosine triphosphate (ATP)] levels due to impairment of mitochondrial metabolic pathways, (b) generation of active species, that can damage cell organelles directly (through the reaction with cellular components) or indirectly (by the activation of signaling pathways), (c) involvement in the intrinsic pathway of cellular apoptosis, and (d) impaired Ca^{2+} metabolism, that subsequently triggers an overproduction of reactive oxygen and nitrogen species (85).

The first sign of mitochondrial function lost upon inflammation is an altered oxygen consumption by the electron transport chain, which directly affects the oxidative phosphorylation leading to a decreased capacity to synthesize ATP. Moreover, increased production of ROS by damaged mitochondria could directly activate NLPR3 inflammasome (86) that may work together with NFkB signaling, perpetuating the inflammatory response and consequently conducting to an overstimulation of the inflammatory response (82). Therefore, inflammation induced by oxidative stress acts as a feedback system sustaining a harmful condition that could result in tissue damage and trigger chronic inflammation.

Mitochondria has been also linked to inflammation through another mechanism where mitochondrial DNA (mtDNA) may trigger innate immunity. Various mitochondrial stressors can lead to mtDNA leakage through the mitochondrial outer membrane permeabilization (MOMP) (87). Stressed mitochondria could become a relevant oxidized mtDNA release source (88). Moreover, incomplete degradation of damage mitochondria also causes subsequent cytoplasm accumulation (89). Once mtDNA reach the cytosol is a suitable ligand for the DNA sensing protein cGAS that catalyzes the production of the secondary messenger cGAMP (90-93). Afterwards, cGAMP binds the adaptor molecule STING an endoplasmic reticulum (ER)-resident protein that triggers innate immunity via activation of TBK1 kinase, responsible of IRF3 phosphorylation, the transcription factor initiates type I IFN response (89, 90, 94). The intrinsic function of the cGAS-STING pathway elicits inflammatory diseases regulation. Therefore, is a relevant pathway from the clinical point of view for future translational approach.

Finally, dysregulation of the systemic inflammatory response is associated with the induction of organ dysfunction and multiple organ failure (86, 95). The mechanisms underlying this deleterious effect could be a consequence of mitochondrial impaired bioenergetic processes impacting complex cellular and physiological functions. Unraveling the mechanisms that interconnect mitochondrial dysfunction, metabolism and systemic inflammation would significantly contribute to the better understanding of many chronic inflammatory diseases.

MITOCHONDRIAL FUNCTION IN INFLAMMATORY AND METABOLIC DISEASES ASSOCIATED WITH AIR POLLUTION EXPOSURE

Airborne PM Exposure Health Outcomes

According to the World Health Organization (WHO), 9 out of 10 people worldwide breathe low-quality air (96). Consequently, more than 9 million premature deaths occur every year due to the joint effects of household and ambient air pollution exposure (97). Recently, it has been estimated that breathing polluted air in urban environments reduces life expectancy by almost 3 years globally (98). In addition, model projections based on business-as-usual emission scenarios suggest that outdoor air pollution contribution to premature mortality could be doubled by 2050 (99). Human can be involuntary exposed to pollutants mainly through tissue or organs that directly interact with PM present in air pollution, as for example the respiratory tract by inhalation, penetration through the skin or eyes and ingestion via the gastrointestinal tract (100). Due to the adverse health effects associated with air pollution, increased incidence of respiratory diseases, such as pneumonia, chronic obstructive pulmonary disease (COPD), and lung cancer, has been observed (101). However, PM exposure on primary organs produce inflammation leading to systemic complications resulting in distant organs defects, such as heart (102, 103). Cardiovascular diseases largely account for the majority of the increase in morbidity and mortality rates (104). In fact, according to the Global Burden of Disease study, air pollution is responsible for one-fourth of the total death count from ischemic heart disease and stroke (105).

Air Pollution Composition and Classification

Air pollution is comprised of a mixture of gases (such as carbon monoxide, sulfur dioxide, nitrogen oxides, and ozone) and airborne PM (106). Besides the complex nature of air pollution and the coexistence of many compounds that may together contribute to the reported negative health impact, numerous epidemiological studies indicate that PM is the main responsible for the health outcomes (107). PM is a heterogeneous mixture of solid and liquid particles suspended in air that are broadly categorized by their size: PM with an aerodynamic diameter $<10\,\mu m$ (PM₁₀) can lodge in the upper respiratory airways and exert a local toxic effect. However, the most harmful particles are those with a diameter $< 2.5 \,\mu m$ (PM_{2.5}), as they can enter deeper into the lung and reach the alveoli (108) or penetrate the skin as hair follicles extend provide a route of penetration for particles from the dermis to the open surface of the skin (109, 110). While coarse particles (PM_{10}) usually arise from natural sources, fossil fuel combustion from transport, industry, and power generation mostly account for air pollution fine particles (PM_{2.5}) burden in urban environments (106). In addition, it has been suggested that nano-scale particles are even able to break through different epithelia and translocate into the bloodstream (111), being potentially able to induce direct damage to peripheral cells and tissues.

Smaller particles present a higher surface to volume ratio providing a larger area were different compounds could be adsorbed (112). Associations have been stablished between air pollution specific components and the toxic mechanisms elicited. Transition metals within the fine PM fraction which typically include iron (Fe), vanadium (V), nickel (Ni), chromium (Cr), copper (Cu),cadmium (Cd) and zinc (Zn), on the basis of their ability to generate ROS in biological tissues and produce an oxidative stress condition (113). Most of these metals have the ability of participate in Fenton-like reactions and contribute to an increase ROS release, initiating oxidative damage mechanisms (113, 114). Regarding organic compounds, polycyclic aromatic hydrocarbons (PAHs) can also be found coating particles. The cytotoxic mechanisms associated to PAHs involve also involve O₂-derived free radicals mainly from mitochondria (115).

PM Detrimental Effects on the Ocular Surface

In those organs constantly exposed to the environment, the epithelia act as the first physical barrier against pollutants becoming more vulnerable (100). For example, it has been shown that PM exacerbates irritation, burning, foreign body sensation, redness, itching in the eyes of people living in urban areas (116–121). Detrimental PM effects on ocular surface are

associated with oxidative stress and proinflammatory pathways (122-126). In this sense, both corneal and conjunctival epithelial cells exposed to different PM surrogates in vitro increase the inflammatory mediators' production (127-130). In turn, the cytokines release, such as IL-6, IL-8, TNF-a, IL-1β, and MCP-1, lead to morphological changes due to cellular hyperplasia affecting the refractive power of the cornea and the vision process (131, 132). Moreover, the eyes are highly vascularized, accordingly an important PM-induced mediators' source. It has been shown that conjunctival epithelial cells are able to uptake diesel exhaust particles (DEPs) where the polycyclic aromatic hydrocarbons (PAHs) compounds present in the particles, trigger the increased ROS released from mitochondria in early stages, and from NOX, in particular NOX4 later. In agreement with studies presenting a link between IL-6 and NOX expression and vice versa (133-136), DEP-induce NOX4 activation along with a proinflammatory response mediated by IL-6. In addition, ROS production induce a redox imbalance sufficient to initiate nuclear factor erythroid 2-related factor 2 (Nrf2) signaling that translocates to the nucleus to enhance the cellular antioxidant capacity (123). Interestingly, whole-body exposure to urban air models in mice showed similar results with macromolecular oxidative damage due to redox imbalance along with an inflammatory response modulated by the increase in IL-10 levels after 1 week of exposure, which early regulates the release of TNF- α and IL-6 (124).

Toxic Mechanisms of PM Exposure on the Skin

Skin is one of the main organs exposed to outdoor pollutants because it provides a major interface between the body and the environment. Similar to the ocular surface offers a biological barrier against air PM where the stratum corneum as is the upper layer represent the main PM-target (137). Inflammation in the skin and an altered redox homeostasis has been mentioned as relevant PM-induced mechanisms (110, 138-140) associated with the aggravation of skin diseases, including atopic dermatitis, acne, and psoriasis (141-143). It was reported that in keratinocytes altered ROS release may trigger mitogen-activated protein kinase (MAPK) signaling pathways resulting in the activation of redox-sensitive transcription factors NF-kB and AP-1. Once in the nucleus, those transcription factor promote the transcription of a variety of proinflammatory cytokines, including TNF- α , IL-1a, IL-6, and IL-8. IL-1 α and IL-1 β in keratinocytes (144). In addition, PM-induce ROS production through the NOX4 activation stimulates NF-κB translocation and increased transcript levels of cytokines (145). Assessment of a 3D skin model exposed to PM showed increased levels of oxidative damage markers resulting in activation of NF- κ B, increased levels of proinflammatory marker COX-2, release of IL-1 α , and DNA damage (146). Comparable to the mechanisms observed in eye and lung tissue exposed to airborne pollutants, skin can absorb particles induced tissue damage, suggesting a cascade of effects that are driven by inflammatory processes and oxidative damage, leading to systemic inflammation as a consequence (147) and also endothelial dysfunction (148, 149).

The Gastrointestinal Tract as a PM Target

The oral tracts along as well as the respiratory system and the skin are the common direct routes of exposure to outdoor pollutants. Particles can access the gut as a result of contaminated food ingestion or indirectly by inhalation (150, 151). Therefore, in recent years research aiming to evaluate the gastrointestinal (GI) tract toxic mechanisms after pollutants exposure have increased. Those studies are based on the premise that GI epithelium behave similar than other organs epithelial cells, like skin or lung tissue, in response to PM exposure (100). Thus, PM also triggers increased ROS generation, which initiate the activation of redox sensitive transcription factor NF-KB (152). In the GI cells NF-kB regulates transcription of myosin light chain kinase (MLCK) (153), affecting perijunctional actin, occludin, and ZO1 at tight junctions, resulting in alteration of gut permeability (152, 154). It has also been observed increased plasma levels of proinflammatory TNF-a and MCP-1 due to PM exposure (153). Take all those results into consideration presence of PM in the GI tract increased oxidative stress and inflammation, leading to structural tissue damage which results in mediators leaking. PMinitiated systemic inflammation may worsen several GI tracts issues, including Crohn's disease (155-157), inflammatory bowel disease (IBD) (158), appendicitis (159), and colorectal cancer (160, 161).

PM Toxicity Within the Brain

Epidemiological studies in humans have shown that high levels of PM are associated with cognitive function changes in children, adults, and the elderly (162). Alterations at the olfactory level, hearing deficits, symptoms of depression and other neuropsychological effects have also been reported (163). The main mechanisms of neurotoxicity produced by PM seem to be related to oxidative stress and neuroinflammation, which are also related to the pathophysiology of various neurodegenerative diseases (164). The way in which the PM reaches the brain and how the particles cause the damage has not been clarified yet. PM that circulates through the nasal compartment can cross the epithelial barrier into the bloodstream or translocate along the axon of the olfactory nerve and reach the central nervous system. Therefore, the adverse effects produced by the inhalation of particles could be the result of a direct effect of the particles via a route from the nasal mucosa to the axon of the olfactory nerve and from there to the olfactory bulb, or by an indirect route through of the systemic inflammatory reaction initiated by increased levels of proinflammatory mediators released into the bloodstream, and oxidative stress (165). Hence, alike GI tract, brain toxicity might begin through direct as well as indirect PM-initiated mechanism.

Cardiorespiratory Diseases Associated to PM Exposure

Following PM inhalation, the activation of oxidative stress and inflammatory pathways largely account for PM biological effects, both locally as well as systemically and in secondary organs, such as the heart (103, 166). In the lung, increased levels of pro-inflammatory cytokines, including Interleukin (IL)–1 β , Tumor Necrosis Factor (TNF) - α , IL-6, and Monocyte Chemoattractant

Protein (MCP) -1, are a frequent finding after PM exposure (167). Therefore, lung inflammatory cell recruitment is usually observed following PM exposure, both in humans (168) and in different animal models (169, 170). Increased plasma levels of these inflammatory mediators have been also associated with episodic elevations in PM in large-scale cohort studies (171), indicating that PM exposure triggers an inflammatory response that is not only confined to the lung, but is also systemic. As a result, metabolism is impaired in distant organs, such as in the heart (172).

Alveolar macrophages play a central role in maintaining lung homeostasis through the removal of exogenous materials and microorganisms from the respiratory surface by phagocytosis, including PM (173, 174). Evolution has refined alveolar macrophages ability for the recognition and clearance of pathogens. However, non-biological anthropogenic PM escapes from this machinery and overwhelms cell capacity for foreign material removal, leading to uncontrolled cell activation and ROS production, as well as an exaggerated inflammatory response and pro-inflammatory cytokine release (175, 176). Activation of the NLRP3 inflammasome following PM uptake seems to represent a central step in the cellular inflammatory response to PM in alveolar macrophages (177). Interestingly, PM has been also shown to accumulate inside mitochondria (174), suggesting a specific direct effect of PM over this organelle. Accordingly, PM exposure induces altered mitochondrial ultrastructure in alveolar macrophages, including swelling, cristae disorder, and organelle fragmentation at high doses, as well as modulation of mitochondrial fission/fusion gene expression (178). Moreover, quinones and polycyclic aromatic hydrocarbons (PAHs) in PM seem to drive mitochondrial depolarization and ROS production in vitro (179). We and others have recently showed that macrophage depletion by intranasal or intratracheal administration of clodronate liposomes reduce pulmonary pro-inflammatory cytokine release following PM exposure, this preventing enhanced thrombosis (180) as well as aggravated myocardial remodeling following experimental myocardial infarction in PM-exposed mice (181). These findings highlight the role of alveolar macrophages in the oxidative and inflammatory response following PM exposure, which negatively impact cardiorespiratory disease onset and progression. Moreover, the role of PM-induced mitochondrial ROS in the release of IL-6 and other pro-inflammatory cytokines in alveolar macrophages was confirmed by treatment with Metformin a complex I inhibitor which reduce mitochondrial complex III ROS production (182). Therefore, diminished mitochondrial ROS release might reduce the risk of PM-induced thrombosis (183).

We and others have also studied the role of the exposure to air pollution PM over lung and heart redox metabolism (184, 185), in which altered mitochondrial respiration together with enhanced NOX2 activity plays a central role as shown in **Figure 3** (186, 187). Interestingly, NOX2 seems to account for increased ROS production in the lung following PM exposure, while mitochondrial mild uncoupling, characterized as increased oxygen consumption rate and decreased inner membrane potential, together with decreased ATP production rate and lower



efficiency of the oxidative phosphorylation process (lower P/O ratio), may prevent further ROS release from this organelle (186). When increasing electron transport rate at the respiratory chain complexes, mitochondrial ROS production is attenuated by different mechanisms: First, mitochondria can significantly reduce O_2^- production by decreasing oxygen tension in the mitochondrial microenvironment; Second, by favoring more oxidized levels of respiratory chain intermediates; Third, by lowering NADH levels that could be used by mitochondrial matrix flavoenzymes; Forth, by preventing reverse electron transfer due to lower membrane potential (188).

Impaired cardiac mitochondrial function also arises as a central feature of air pollution PM toxicology. Mechanistically, an acute exposure to PM induces a decrease in active, but not rest, state oxygen consumption rate, together with inner membrane depolarization and reduced mitochondrial ATP production (96). Consequently, deficient contractile and lusitropic reserve is observed in PM-exposed mice, as the heart fails to properly increase cardiac contractility after a β -adrenergic stimulus with isoproterenol (187). Blunted mitochondrial ATP supply in mice breathing PM may account for this effect, as decreased ATP levels are a frequent finding in the failing heart (189, 190). Interestingly, this cardiac mitochondrial bioenergetic dysfunction seems to be

partially mediated by an inflammatory response triggered by PM exposure, since impaired mitochondrial respiration and cardiac contractility is attenuated by pretreatment with a chimeric anti-TNF- α antibody (Infliximab) in PM-exposed mice (172).

Given that heart perfusion with a low-calcium Krebs buffer also prevents impaired contractility in PM-exposed mice (172), calcium overload may also contribute with this scenario. In fact, altered calcium homeostasis has been recently reported in cardiomyocytes incubated with PM (191), which might be explained by hampered mitochondrial calcium uptake by aromatic chemicals in PM (179) and decreased activity of sarco/endoplasmic reticulum calcium-ATPase 2a (SERCA2a), a major regulator of cytosolic calcium concentration that couples ATP hydrolysis with calcium transport into the sarcoplasmic reticulum during relaxation (192). Moreover, as PM exposure impairs cardiac mitochondrial respiration, deficient mitochondrial ATP supply in PMexposed mice heart represent a plausible link between ROS production, altered calcium handling, and impaired myocardial contractile function and relaxation. In support of this concept, sarcomere shortening in cardiomyocytes incubated with PM was prevented by the mitochondrial targeted antioxidants Tiron and MitoTEMPOL (191).

Recently, updated epidemiological studies found a positive correlation between air pollution exposure and type 2 diabetes mellitus incidence (193) and mortality (194). Therefore, the role of air pollution PM exposure over metabolism, adipose tissue inflammation, and obesity gained crescent attention. Mechanistically, the exposure to PM in a mice model of diet induced obesity has been shown to aggravate insulin resistance and inflammation in white adipose tissue (195). Moreover, PM exposure induced increased ROS production and downregulation of uncoupling protein (UCP) -1 in brown adipose tissue (196). UCP-1 is a mitochondrial inner membrane protein that dissipates membrane potential to produce heat, and therefore modulates thermogenesis and protects against excessive ROS production (197). It remains unclear whether adipose tissue inflammation and altered mitochondrial function represent a cause or consequence of altered metabolism following PM exposure. In this context, the precise role of mitochondrial respiration, ROS release, and heat production needs to be further addressed in future studies.

Taken together, impaired mitochondrial respiration, enhanced ROS release, and deficient ATP supply (and maybe also uncoupling), play a central role in the adverse health effects reported after air pollution PM exposure. In this context, the modulation of mitochondrial function (e.g. by mitochondrial targeted antioxidants) arises as a potential therapeutic target to prevent excessive lung inflammatory response, as well as impaired cardiac contractility and metabolism in PM-exposed individuals at particular high risk.

REFERENCES

- Strappazzon F, Cecconi F. The multifaceted mitochondrion: an attractive candidate for therapeutic strategies. *Pharmacol Res.* (2015) 99:425– 33. doi: 10.1016/j.phrs.2015.03.007
- van der Bliek AM, Sedensky MM, Morgan PG. Cell biology of the mitochondrion. *Genetics*. (2017) 207:843– 71. doi: 10.1534/genetics.117.300262
- Orrenius S, Gogvadze V, Zhivotovsky B. Calcium and mitochondria in the regulation of cell death. *Biochem Biophys Res Commun.* (2015) 460:72– 81. doi: 10.1016/j.bbrc.2015.01.137
- Rossi A, Pizzo P, Filadi R. Calcium, mitochondria and cell metabolism: a functional triangle in bioenergetics. *Biochim Biophys Acta Mol Cell Res.* (2019) 1866:1068–78. doi: 10.1016/j.bbamcr.2018.10.016
- 5. Murphy MP. How mitochondria produce reactive oxygen species. *Biochem J.* (2009) 417:1–13. doi: 10.1042/BJ20081386
- Mailloux RJ, McBride SL, Harper ME. Unearthing the secrets of mitochondrial ROS and glutathione in bioenergetics. *Trends Biochem Sci.* (2013) 38:592–602. doi: 10.1016/j.tibs.2013.09.001
- Kamp DW, Shacter E, Weitzman SA. Chronic inflammation and cancer: the role of the mitochondria. Oncology (Williston Park). (2011) 25:400–13.
- West AP. Mitochondrial dysfunction as a trigger of innate immune responses and inflammation. *Toxicology.* (2017) 391:54–63. doi: 10.1016/j.tox.2017.07.016
- van Horssen J, van Schaik P, Witte M. Inflammation and mitochondrial dysfunction: a vicious circle in neurodegenerative disorders? *Neurosci Lett.* (2019) 710:132931. doi: 10.1016/j.neulet.2017.06.050
- Seim S. Production of reactive oxygen species and chemiluminescence by human monocytes during differentiation and lymphokine activation *in vitro. Acta Pathol Microbiol Immunol Scand C.* (1982) 90:179– 85. doi: 10.1111/j.1699-0463.1982.tb01436.x

CONCLUSIONS

The mitochondria-dependent mechanisms associated with inflammation are still poorly understood. A better understanding of the cellular pathways underlying this phenomenon would allow a more targeted approach to face the adverse effects linked to inflammatory syndromes. An interplay between mitochondria and NOX as specific ROS sources in inflammation has been recognized, where the redox crosstalk stimulates each other in a positive feedback fashion, resulting in a complex oxidative stress and redox signaling network. As a case study, inflammation and altered mitochondrial function represent a relevant mechanism of altered cell metabolism following PM exposure may contribute to the increased morbidity and mortality associated with polluted areas. The modulation of mitochondrial function by mitochondrial targeted antioxidants arises as a potential therapeutic target to prevent an excessive inflammatory response.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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- 11. Olson KR. Reactive oxygen species or reactive sulfur species: why we should consider the latter. *J Exp Biol.* (2020) 223:jeb196352. doi: 10.1242/jeb.196352
- Schieber M, Chandel NS. ROS function in redox signaling and oxidative stress. Curr Biol. (2014) 24:R453–62. doi: 10.1016/j.cub.2014.03.034
- Sies H. Role of metabolic H₂O₂ generation: redox signaling and oxidative stress. J Biol Chem. (2014) 289:8735–41. doi: 10.1074/jbc.R113.544635
- Green MJ, Hill HA. Chemistry of dioxygen. *Methods Enzymol.* (1984) 105:3– 22. doi: 10.1016/S0076-6879(84)05004-7
- Di Meo S, Reed TT, Venditti P, Victor VM. Role of ROS and RNS sources in physiological and pathological conditions. Oxid Med Cell Longev. (2016) 2016:1245049. doi: 10.1155/2016/1245049
- Halliwell B. Superoxide-dependent formation of hydroxyl radicals in the presence of iron chelates: is it a mechanism for hydroxyl radical production in biochemical systems? *FEBS Lett.* (1978) 92:321–6. doi: 10.1016/0014-5793(78)80779-0
- Liochev S, Fridovich I. The haber-weiss cycle–70 years later: an alternative view. *Redox Rep.* (2002) 7:55–57. doi: 10.1179/135100002125000190
- Azzi A, Davies KJ, Kelly F. Free radical biology terminology and critical thinking. FEBS Lett. (2004) 558:3–6. doi: 10.1016/S0014-5793(03)01526-6
- Nathan C, Ding A. SnapShot: reactive oxygen intermediates (ROI). Cell. (2010) 140:951.e2. doi: 10.1016/j.cell.2010.03.008
- Finkel T, Holbrook NJ. Oxidants, oxidative stress and the biology of ageing. Nature. (2000) 408:239–47. doi: 10.1038/35041687
- Couto N, Wood J, Barber J. The role of glutathione reductase and related enzymes on cellular redox homoeostasis network. *Free Radic Biol Med.* (2016) 95:27–42. doi: 10.1016/j.freeradbiomed.2016. 02.028
- 22 Rhee Kil Multiple SG. IS functions and regulation of mammalian peroxiredoxins. Annu Rev Biochem. (2017)86:749-75. doi: 10.1146/annurev-biochem-060815-0 14431

- Schmidt HH, Stocker R, Vollbracht C, Paulsen G, Riley D, Daiber A, et al. Antioxidants in translational medicine. *Antioxid Redox Signal*. (2015) 23:1130–43. doi: 10.1089/ars.2015.6393
- Liu H, Colavitti R, Rovira II, Finkel T. Redox-dependent transcriptional regulation. *Circ Res.* (2005) 97:967– 74. doi: 10.1161/01.RES.0000188210.72062.10
- Herrmann JM, Dick TP. Redox biology on the rise. *Biol Chem.* (2012) 393:999–1004. doi: 10.1515/hsz-2012-0111
- Sies H. Oxidative stress: introductory remarks. In: Sies H, editor. Oxidative Stress. London: Academic (1985). p. 1–8. doi: 10.1016/B978-0-12-642760-8.50005-3
- 27. Jones DP, Sies H. The redox code. Antioxid Redox Signal. (2015) 23:734–46. doi: 10.1089/ars.2015.6247
- Sies H, Berndt C, Jones DP. Oxidative stress. Annu Rev Biochem. (2017) 86:715–48. doi: 10.1146/annurev-biochem-061516-045037
- Lushchak VI. Free radicals, reactive oxygen species, oxidative stress and its classification. *Chem Biol Interact.* (2014) 224:164– 75. doi: 10.1016/j.cbi.2014.10.016
- Daiber A, Di Lisa F, Ferdinandy P. Pharmacology of oxidative stress: translational opportunities. Br J Pharmacol. (2017) 174:1511-13. doi: 10.1111/bph.13795
- Zorov DB, Filburn CR, Klotz LO, Zweier JL, Sollott SJ. Reactive oxygen species (ROS)-induced ROS release: a new phenomenon accompanying induction of the mitochondrial permeability transition in cardiac myocytes. *J Exp Med.* (2000) 192:1001–14. doi: 10.1084/jem.192.7.1001
- Ristow M. Unraveling the truth about antioxidants: mitohormesis explains ROS-induced health benefits. *Nat Med.* (2014) 20:709– 11. doi: 10.1038/nm.3624
- Sena LA, Chandel NS. Physiological roles of mitochondrial reactive oxygen species. *Mol Cell.* (2012) 48:158–67. doi: 10.1016/j.molcel.2012.09.025
- 34. Shadel GS, Horvath TL. Mitochondrial ROS signaling in organismal homeostasis. *Cell*. (2015) 163:560–9. doi: 10.1016/j.cell.2015.10.001
- 35. Topf U, Suppanz I, Samluk L, Wrobel L, Boser A, Sakowska P, et al. Quantitative proteomics identifies redox switches for global translation modulation by mitochondrially produced reactive oxygen species. *Nat Commun.* (2018) 9:324. doi: 10.1038/s41467-017-02694-8
- Drose S, Brandt U. Molecular mechanisms of superoxide production by the mitochondrial respiratory chain. Adv Exp Med Biol. (2012) 748:145– 69. doi: 10.1007/978-1-4614-3573-0_6
- Lenaz G. Mitochondria and reactive oxygen species. Which role in physiology and pathology? *Adv Exp Med Biol.* (2012) 942:93–136. doi: 10.1007/978-94-007-2869-1_5
- Wong HS, Dighe PA, Mezera V, Monternier PA, Brand MD. Production of superoxide and hydrogen peroxide from specific mitochondrial sites under different bioenergetic conditions. J Biol Chem. (2017) 292:16804– 9. doi: 10.1074/jbc.R117.789271
- Dikalov A. Cross talk between mitochondrial and NADPH oxidases. Free Radic Biol Med. (2011) 51:1289– 301. doi: 10.1016/j.freeradbiomed.2011.06.033
- Zandalinas SI, Mittler R. ROS-induced ROS release in plant and animal cells. *Free Radic Biol Med.* (2018) 122:21– 27. doi: 10.1016/j.freeradbiomed.2017.11.028
- López-Doménech G, Covill-Cooke C, Ivankovic D, Halff EF, Sheehan DF, Norkett R, et al. Miro proteins coordinate microtubule- and actindependent mitochondrial transport and distribution. *EMBO J.* (2018) 37:321–36. doi: 10.15252/embj.201696380
- Schuler MH, Lewandowska A, Caprio GD, Skillern W, Upadhyayula S, Kirchhausen T, et al. Miro1-mediated mitochondrial positioning shapes intracellular energy gradients required for cell migration. *Mol Biol Cell*. (2017) 28:2159–69. doi: 10.1091/mbc.e16-10-0741
- van der Vliet A, Danyal K, Heppner DE. Dual oxidase: a novel therapeutic target in allergic disease. Br J Pharmacol. (2018) 175:1401– 18. doi: 10.1111/bph.14158
- Babior BM, Lambeth JD, Nauseef W. The neutrophil NADPH oxidase. Arch Biochem Biophys. (2002) 397:342–4. doi: 10.1006/abbi.2001.2642
- Bedard K, Krause KH. The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology. *Physiol. Rev.* (2007) 87:245– 313. doi: 10.1152/physrev.00044.2005

- Winterbourn CC, Hampton MB, Livesey JH, Kettle AJ. Modeling the reactions of superoxide and myeloperoxidase in the neutrophil phagosome: implications for microbial killing. J Biol Chem. (2006) 281:39860– 9. doi: 10.1074/jbc.M605898200
- van der Vliet A. NADPH oxidases in lung biology and pathology: host defenses enzymes and more. *Free Radic Biol Med.* (2008) 44:938– 55. doi: 10.1016/j.freeradbiomed.2007.11.016
- Zhang Y, Murugesan P, Huang K, Cai H. NADPH oxidases and oxidase crosstalk in cardiovascular diseases: novel therapeutic targets. *Nat Rev Cardiol.* (2020) 17:170–94. doi: 10.1038/s41569-019-0260-8
- Block K, Gorin Y, Abboud HE. Subcellular localization of Nox4 and regulation in diabetes. *Proc Natl Acad Sci U S A.* (2009) 106:14385– 90. doi: 10.1073/pnas.0906805106
- Serrander L, Jaquet V, Bedard K, Plastre O, Hartley O, Arnaudeau S, et al. NOX5 is expressed at the plasma membrane and generates superoxide in response to protein kinase C activation. *Biochimie*. (2007) 89:1159– 67. doi: 10.1016/j.biochi.2007.05.004
- Paik YH, Kim J, Aoyama T, De Minicis S, Bataller R, Brenner DA. Role of NADPH oxidases in liver fibrosis. *Antioxid Redox Signal.* (2014) 20:2854– 72. doi: 10.1089/ars.2013.5619
- Van Buul JD, Fernandez-Borja M, Anthony EC, Hordijk PL. Expression and localization of NOX₂ and NOX₄ in primary human endothelial cells. *Antioxid Redox Signal.* (2005) 7:308–17. doi: 10.1089/ars.2005.7.308
- Shanmugasundaram K, Nayak BK, Friedrichs WE, Kaushik D, Rodriguez R, Block K. NOX₄ functions as a mitochondrial energetic sensor coupling cancer metabolic reprogramming to drug resistance. *Nat Commun.* (2017) 8:997. doi: 10.1038/s41467-017-01106-1
- Graham KA, Kulawiec M, Owens KM, Li X, Desouki MM, Chandra D, et al. NADPH oxidase 4 is an oncoprotein localized to mitochondria. *Cancer Biol Ther.* (2010) 10:223–31. doi: 10.4161/cbt.10.3.12207
- 55. Ago T, Kuroda J, Pain J, Fu C, Li H, Sadoshima J. Upregulation of Nox₄ by hypertrophic stimuli promotes apoptosis and mitochondrial dysfunction in cardiac myocytes. *Circ Res.* (2010) 106:1253–64. doi: 10.1161/CIRCRESAHA.109.213116
- Dikalov SI, Dikalov AE, Bikineyeva AT, Schmidt HH, Harrison DG, Griendling KK. Distinct roles of Nox1 and Nox4 in basal and angiotensin II-stimulated superoxide and hydrogen peroxide production. *Free Radic Biol Med.* (2008) 45:1340–51. doi: 10.1016/j.freeradbiomed.2008.08.013
- Takac I, Schroder K, Zhang L, Lardy B, Anilkumar N, Lambeth JD, et al. The E-loop is involved in hydrogen peroxide formation by the NADPH oxidase Nox₄. J Biol Chem. (2011) 286:13304–13. doi: 10.1074/jbc.M110.192138
- Guzik TJ, Chen W, Gongora MC, Guzik B, Lob HE, Mangalat D, et al. Calcium-dependent NOX₅ NADPH oxidase contributes to vascular oxidative stress in human coronary artery disease. J Am Coll Cardiol. (2008) 52:1803–9. doi: 10.1016/j.jacc.2008.07.063
- van der Vliet A. Nox enzymes in allergic airway inflammation. *Biochim Biophys Acta*. (2011) 1810:1035–44. doi: 10.1016/j.bbagen.2011.03.004
- Irrcher I, Ljubicic V, Hood DA. Interactions between ROS and AMP kinase activity in the regulation of PGC-1alpha transcription in skeletal muscle cells. *Am J Physiol Cell Physiol.* (2009) 296:C116–23. doi: 10.1152/ajpcell.00267.2007
- Fitzpatrick AM, Park Y, Brown LA, Jones DP. Children with severe asthma have unique oxidative stress-associated metabolomic profiles. J Allergy Clin Immunol. (2014) 133:258–61.E1–8. doi: 10.1016/j.jaci.2013. 10.012
- Daiber A. Redox signaling (cross-talk) from and to mitochondria involves mitochondrial pores and reactive oxygen species. *Biochim Biophys Acta*. (2010) 1797:897–906. doi: 10.1016/j.bbabio.2010.01.032
- Schulz E, Wenzel P, Munzel T, Daiber A. Mitochondrial redox signaling: interaction of mitochondrial reactive oxygen species with other sources of oxidative stress. *Antioxid Redox Signal.* (2014) 20:308–24. doi: 10.1089/ars.2012.4609
- Radi R, Cassina A, Hodara R, Quijano C, Castro L. Peroxynitrite reactions and formation in mitochondria. *Free Radic Biol Med.* (2002) 33:1451– 64. doi: 10.1016/S0891-5849(02)01111-5
- 65. Oelze M, Kroller-Schon S, Steven S, Lubos E, Doppler C, Hausding M, et al. Glutathione peroxidase-1 deficiency potentiates dysregulatory modifications of endothelial nitric oxide synthase

and vascular dysfunction in aging. *Hypertension.* (2014) 63:390-6. doi: 10.1161/HYPERTENSIONAHA.113.01602

- 66. Wenzel P, Mollnau H, Oelze M, Schulz E, Wickramanayake JM, Muller J, et al. First evidence for a crosstalk between mitochondrial and NADPH oxidase-derived reactive oxygen species in nitroglycerintriggered vascular dysfunction. *Antioxid Redox Signal.* (2008) 10:1435–47. doi: 10.1089/ars.2007.1969
- Queliconi BB, Wojtovich AP, Nadtochiy SM, Kowaltowski AJ, Brookes PS. Redox regulation of the mitochondrial K(ATP) channel in cardioprotection. *Biochim Biophys Acta.* (2011) 1813:1309–15. doi: 10.1016/j.bbamcr.2010.11.005
- Kroller-Schon S, Steven S, Kossmann S, Scholz A, Oelze M, Daub S, et al. Molecular mechanisms of the crosstalk between mitochondria and NADPH oxidase through reactive oxygen species-studies in white blood cells and in animal models. *Antioxid Redox Signal*. (2014) 20:247–66. doi: 10.1089/ars.2012.4953
- Medzhitov R. Origin and physiological roles of inflammation. *Nature*. (2008) 454:428–35. doi: 10.1038/nature07201
- López-Armada MJ, Riveiro-Naveira RR, Vaamonde-García C, Valcárcel-Ares MN. Mitochondrial dysfunction and the inflammatory response. *Mitochondrion.* (2013) 13:106–18. doi: 10.1016/j.mito.2013.01.003
- Tschopp J. Mitochondria: sovereign of inflammation? *Eur J Immunol.* (2011) 41:1196–202. doi: 10.1002/eji.201141436
- Bennet JM, Reeves G, Billman GE, Sturmberg JP. Inflammation-nature's way to efficiently respond to all types of challenges: implications for understanding and managing "the epidemic" of chronic diseases. *Front Med.* (2018) 5:316. doi: 10.3389/fmed.2018.00316
- Coussens LM, Werb Z. Inflammation and cancer. *Nature*. (2002) 420:860– 67. doi: 10.1038/nature01322
- Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH. Mechanisms underlying inflammation in neurodegeneration. *Cell.* (2010) 140:918– 34. doi: 10.1016/j.cell.2010.02.016
- Alvarez S, Vico T, Vanasco V. Cardiac dysfunction, mitochondrial architecture, energy production, and inflammatory pathways: interrelated aspects in endotoxemia and sepsis. *Int J Biochem Cell Biol.* (2016) 81(Pt B):307–14. doi: 10.1016/j.biocel.2016.07.032
- Barnig C, Bezema T, Calder PC, Charloux A, Frossard N, Garssen J, et al. Activation of resolution pathways to prevent and fight chronic inflammation: lessons from asthma and inflammatory bowel disease. *Front Immunol.* (2019) 10:1699. doi: 10.3389/fimmu.2019.01699
- Nathan C, Ding A. Nonresolving inflammation. Cell. (2010) 140:871– 82. doi: 10.1016/j.cell.2010.02.029
- Glaros TG, Chang S, Gilliam EA, Maitra U, Deng H, Li L. Causes and consequences of low grade endotoxemia and inflammatory diseases. *Front Biosci (Schol Ed).* (2013) 5:754–65. doi: 10.2741/S405
- Schroder K, Tschopp J. The inflammasomes. Cell. (2010) 140:821– 32. doi: 10.1016/j.cell.2010.01.040
- Barnes PJ. Inflammatory mechanisms in patients with chronic obstructive pulmonary disease. J Allergy Clin Immunol. (2016) 138:16–27. doi: 10.1016/j.jaci.2016.05.011
- Laleman W, Claria J, Van der Merwe S, Moreau R, Trebicka J. Systemic inflammation and acute-on-chronic liver failure: too much, not enough. *Can J Gastroenterol Hepatol.* (2018) 2018:1027152. doi: 10.1155/2018/1027152
- Mills EL, Kelly B, O'Neill LA. Mitochondria are the powerhouses of immunity. Nat Immunol. (2017) 18:488–98. doi: 10.1038/ni.3704
- Chen CC, Tsai SH, Lu CC, Hu ST, Wu TS, Huang TT, et al. Activation of an NLRP3 inflammasome restricts Mycobacterium kansasii infection. *PLoS* ONE. (2012) 7:e36292. doi: 10.1371/journal.pone.0036292
- Gkikas I, Palikaras K, Tavernarakis N. The role of mitophagy in innate immunity. *Front Immunol.* (2018) 9:1283. doi: 10.3389/fimmu.2018.01283
- Nunnari J, Suomalainen A. Mitochondria: in sickness and in health. Cell. (2012) 148:1145–59. doi: 10.1016/j.cell.2012.02.035
- Kozlov AV, Lancaster JR, Meszaros AT, Weidinger A. Mitochondriameditated pathways of organ failure upon inflammation. *Redox Biol.* (2017) 13:170–81. doi: 10.1016/j.redox.2017.05.017
- Riley JS, Quarato G, Cloix C, Lopez J, O'Prey J, Pearson M, et al. Mitochondrial inner membrane permeabilisation enables mtDNA release during apoptosis. *EMBO J.* (2018) 37:e99238. doi: 10.15252/embj.201899238

- West AP, Khoury-Hanold W, Staron M, Tal MC, Pineda CM, Lang SM, et al. Mitochondrial DNA stress primes the antiviral innate immune response. *Nature*. (2015) 520:553–7. doi: 10.1038/nature14156
- Wan D, Jiang W, Hao J. Research advances in how the cGAS-STING pathway controls the cellular inflammatory response. *Front Immunol.* (2020) 11:615. doi: 10.3389/fimmu.2020.00615
- Maekawa H, Inoue T, Ouchi H, Jao TM, Inoue R, Nishi H, et al. Mitochondrial damage causes inflammation via cGAS-STING signaling in acute kidney injury. *Cell Rep.* (2019) 29:1261–73.e6. doi: 10.1016/j.celrep.2019.09.050
- Ablasser A, Goldeck M, Cavlar T, Deimling T, Witte G, Röhl I, et al. cGAS produces a 20-50-linked cyclic dinucleotide second messenger that activates STING. *Nature*. (2013) 498:380–84. doi: 10.1038/nature12306
- Diner EJ, Burdette DL, Wilson SC, Monroe KM, Kellenberger CA, Hyodo M, et al. The innate immune DNA sensor cGAS produces a noncanonical cyclic dinucleotide that activates human STING. *Cell Rep.* (2013) 3:1355– 61. doi: 10.1016/j.celrep.2013.05.009
- Gao P, Ascano M, Wu Y, Barchet W, Gaffney BL, Zillinger T, et al. Cyclic [G(2',5')pA(3',5')p] is the metazoan second messenger produced by DNA-activated cyclic GMP-AMP synthase. *Cell.* (2013) 153:1094– 107. doi: 10.1016/j.cell.2013.04.046
- 94. Riley JS, Tait SW. Mitochondrial DNA in inflammation and immunity. *EMBO Rep.* (2020) 21:e49799. doi: 10.15252/embr.201949799
- Lee I, Hüttemann M. Energy crisis: the role of oxidative phosphorylation in acute inflammation and sepsis. *Biochim Biophys Acta*. (2014) 1842:1579– 86. doi: 10.1016/j.bbadis.2014.05.031
- 96. WHO. Burden of Disease From the Joint Effects of Household and Ambient Air Pollution for 2016. New York, NY: WHO Report. (2018).
- Landrigan PJ, Fuller R, Acosta NJR, Adeyi O, Arnold R, Basu NN, et al. The lancet commission on pollution and health. *Lancet.* (2018) 391:462– 512. doi: 10.1016/S0140-6736(17)32345-0
- Lelieveld J, Pozzer A, Pöschl U, Fnais M, Haines A, Münzel T. Loss of life expectancy from air pollution compared to other risk factors: a worldwide perspective [published online ahead of print, 2020 Mar 3]. *Cardiovasc Res.* (2020) 116:1910–17. doi: 10.1093/cvr/cva025
- Lelieveld J, Evans JS, Fnais M, Giannadaki D, Pozzer A. The contribution of outdoor air pollution sources to premature mortality on a global scale. *Nature*. (2015) 525:367–71. doi: 10.1038/nature15371
- Valacchi G, Magnani N, Woodby B, Ferreira SM, Evelson P. Particulate matter induces tissue oxinflammation: from mechanism to damage. *Antioxid Redox Signal.* (2020) 33:308–26. doi: 10.1089/ars.2019.8015
- 101. Kaiser J. Epidemiology. Mounting evidence indicts fine-particle pollution. Science. (2005) 307:1858–61. doi: 10.1126/science.307.5717.1858a
- 102. Brook RD, Franklin B, Cascio W, Hong Y, Howard G, Lipsett M, et al. Expert panel on population and prevention science of the american heart association. Air pollution and cardiovascular disease: a statement for healthcare professionals from the expert panel on population and prevention science of the American heart association. *Circulation*. (2004) 109:2655– 71. doi: 10.1161/01.CIR.0000128587.30041.C8
- 103. Marchini T, Zirlik A, Wolf D. Pathogenic role of air pollution particulate matter in cardiometabolic disease: evidence from mice and humans. *Antioxid Redox Signal.* (2020) 33:263–79. doi: 10.1089/ars.2020.8096
- 104. Brook RD, Rajagopalan S, Pope CA 3rd, Brook JR, Bhatnagar A, Diez-Roux AV, et al. Particulate matter air pollution and cardiovascular disease: an update to the scientific statement from the American heart association. *Circulation.* (2010) 121:2331–78. doi: 10.1161/CIR.0b013e3181dbece1
- 105. Fuller R, Rahona E, Fisher S, Caravanos J, Webb D, Kass D, et al. Pollution and non-communicable disease: time to end the neglect. *Lancet Planet Health.* (2018) 2:e96–e98. doi: 10.1016/S2542-5196(18)30020-2
- Brook RD. Cardiovascular effects of air pollution. Clin Sci (Lond). (2008) 115:175–87. doi: 10.1042/CS20070444
- Hamanaka RB, Mutlu GM. Particulate matter air pollution: effects on the cardiovascular system. Front Endocrinol (Lausanne). (2018) 9:680. doi: 10.3389/fendo.2018.0 0680
- Oberdörster G, Oberdörster E, Oberdörster J. Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect.* (2005) 113:823–39. doi: 10.1289/ehp.7339

- Lademann J, Schaefer H, Otberg N, Teichmann A, Blume-Peytavi U, Sterry W. Penetration of microparticles into human skin. *Hautarzt*. (2004) 55:1117–19. doi: 10.1007/s00105-004-0841-1
- 110. Jin SP, Li Z, Choi EK, Lee S, Kim YK, Seo EY, et al. Urban particulate matter in air pollution penetrates into the barrier-disrupted skin and produces ROSdependent cutaneous inflammatory response *in vivo. J Dermatol Sci.* (2018) 91:175–83. doi: 10.1016/j.jdermsci.2018.04.015
- 111. Nemmar A, Hoet PH, Vanquickenborne B, Dinsdale D, Thomeer M, Hoylaerts MF, et al. Passage of inhaled particles into the blood circulation in humans. *Circulation*. (2002) 105:411–4. doi: 10.1161/hc0402.104118
- Nel A, Xia T, M\u00e4dler L, Li N. Toxic potential of materials at the nanolevel. Science. (2006) 311:622–7. doi: 10.1126/science.1114397
- Chen LC, Lipmann M. Effects of metals within ambient air particulate matter (PM) on human health. *Inhal Toxicol.* (2009) 21:1–31. doi: 10.1080/08958370802105405
- 114. Magnani ND, Marchini T, Garcés M, Mebert A, Cáceres L, Diaz L, et al. Role of transition metals present in air particulate matter on lung oxygen metabolism. *Int J Biochem Cell Biol.* (2016) 81(Pt B):419– 26. doi: 10.1016/j.biocel.2016.10.009
- 115. Li N, Xia T, Nel AE. The role of oxidative stress in ambient particulate matter induced lung diseases and its implications in the toxicity of engineered nanoparticles. *Free Radic Biol Med.* (2008) 44:1689– 99. doi: 10.1016/j.freeradbiomed.2008.01.028
- Berra M, Galperin G, Dawidowski L, Tau J, Marquez I, Berra A. Impact of wildfire smoke in buenos aires, argentina, on ocular surface. *Arq Bras Oftalmol.* (2015) 78:110–14. doi: 10.5935/0004-2749.20150028
- 117. Fu Q, Mo Z, Lyu D, Zhang L, Qin Z, Tang Q, et al. Air pollution and outpatient visits for conjunctivitis: a casecrossover study in Hangzhou, China. *Environ Pollut.* (2017) 231:1344–50. doi: 10.1016/j.envpol.2017.08.109
- 118. Hwang SH, Choi YH, Paik HJ, Wee WR, Kim MK, Kim DH. Potential importance of ozone in the association between outdoor air pollution and dry eye disease in South Korea. *JAMA Ophthalmol.* (2016) 134:503– 10. doi: 10.1001/jamaophthalmol.2016.0139
- 119. Moss SE, Klein R, Klein BE. Prevalence of and risk factors for dry eye syndrome. Arch Ophthalmol. (2000) 118:1264– 8. doi: 10.1001/archopht.118.9.1264
- 120. Nucci P, Sacchi M, Pichi F, Allegri P, Serafino M, Dello Strologo M, et al. Pediatric conjunctivitis and air pollution exposure: a prospective observational study. *Semin Ophthalmol.* (2017) 32:407–11. doi: 10.3109/08820538.2015.1115088
- 121. Torricelli AAM, Matsuda M, Novaes P, Braga ALF, Saldiva PHN, Alves MR, et al. Effects of ambient levels of traffic-derived air pollution on the ocular surface: analysis of symptoms, conjunctival goblet cell count and mucin 5AC gene expression. *Environ Res.* (2014) 131:59–63. doi: 10.1016/j.envres.2014.02.014
- 122. Lasagni Vitar RM, Tau J, Reides CG, Berra A, Ferreira SM, Llesuy SF. Evaluation of oxidative stress markers in human conjunctival epithelial cells exposed to diesel exhaust particles (DEP). *Invest Ophthalmol Vis Sci.* (2015) 56:7058–66. doi: 10.1167/iovs.15-16864
- 123. Lasagni Vitar RM, Tau J, Janezic NS, Tesone AI, Hvozda Arana AG, Reides CG, et al. Diesel exhaust particles (DEP) induce an early redox imbalance followed by an IL-6 mediated inflammatory response on human conjunctival epithelial cells. *Exp Eye Res.* (2018) 171:37– 47. doi: 10.1016/j.exer.2018.03.005
- 124. Lasagni Vitar RM, Hvozda Arana AG, Janezic NS, Marchini T, Tau J, Martinefski M, et al. Urban air pollution induces redox imbalance and epithelium hyperplasia in mice cornea. *Toxicol Appl Pharmacol.* (2019) 384:114770. doi: 10.1016/j.taap.2019.114770
- 125. Tau J, Novaes P, Matsuda M, Tasat DR, Saldiva PH, Berra A. Diesel exhaust particles selectively induce both proinflammatory cytokines and mucin production in cornea and conjunctiva human cell lines. *Invest Ophthalmol Vis Sci.* (2013) 54:4759–65. doi: 10.1167/iovs.12-10541
- 126. Tesone AI, Lasagni Vitar RM, Tau J, Maglione GA, Llesuy S, Tasat DR, et al. Volcanic ash from puyehue-cordon caulle volcanic complex and calbuco promote a differential response of pro-inflammatory and oxidative stress mediators on human conjunctival epitelial cells. *Environ Res.* (2018) 167:87– 97. doi: 10.1016/j.envres.2018.07.013

- 127. Park EJ, Chae JB, Lyu J, Yoon C, Kim S, Yeom C, et al. Ambient fine particulate matters induce cell death and inflammatory response by influencing mitochondria function in human corneal epithelial cells. *Environ Res.* (2017) 159:595–605. doi: 10.1016/j.envres.2017.08.044
- Wu CC, Bao LJ, Tao S, Zeng EY. Dermal uptake from airborne organics as an important route of human exposure to E-waste combustion fumes. *Environ Sci Technol.* (2016) 50:6599–605. doi: 10.1021/acs.est.5b05952
- 129. Yang Q, Tang L, Shen M, Wang Y, Wei Y, Jeyalatha V, et al. Effects of diesel exhaust particles on the condition of mouse ocular surface. *Ecotoxicol Environ Saf.* (2018) 163:585–93. doi: 10.1016/j.ecoenv.2018.07.101
- 130. Yoon S, Han S, Jeon KJ, Kwon S. Effects of collected road dusts on cell viability, inflammatory response, and oxidative stress in cultured human corneal epithelial cells. *Toxicol Lett.* (2018) 284:152–60. doi: 10.1016/j.toxlet.2017.12.012
- Del Monte DW, Kim T. Anatomy and physiology of the cornea. J Cataract Refract. Surg. (2011) 37:588–98. doi: 10.1016/j.jcrs.2010.12.037
- Shoham A, Hadziahmetovic M, Dunaief JL, Mydlarski MB, Schipper HM. Oxidative stress in diseases of the human cornea. *Free Radic Biol Med.* (2008) 45:1047–55. doi: 10.1016/j.freeradbiomed.2008.07.021
- Behrens MM, Ali SS, Dugan LL. Interleukin-6 mediates the increase in NADPH-oxidase in the ketamine model of schizophrenia. *J Neurosci.* (2008) 28:13957–66. doi: 10.1523/JNEUROSCI.4457-08.2008
- 134. Didion SP. Cellular and oxidative mechanisms associated with interleukin-6 signaling in the vasculature. *Int J Mol Sci.* (2017) 18:2563. doi: 10.3390/ijms18122563
- 135. Park HS, Chun JN, Jung HY, Choi C, Bae YS. Role of NADPH oxidase 4 in lipopolysaccharide-induced proinflammatory responses by human aortic endothelial cells. *Cardiovasc Res.* (2006) 72:447–55. doi: 10.1016/j.cardiores.2006.09.012
- 136. Yu JH, Lim JW, Kim H, Kim KH. NADPH oxidase mediates interleukin-6 expression in cerulein-stimulated pancreatic acinar cells. *Int J Biochem Cell Biol.* (2005) 37:1458–69. doi: 10.1016/j.biocel.2005.02.004
- Valacchi G, Sticozzi C, Pecorelli A, Cervellati F, Cervellati C, Maioli E, et al. Cutaneous responses to environmental stressors. *Ann NY Acad Sci.* (2012) 1271:75–81. doi: 10.1111/j.1749-6632.2012.06724.x
- 138. Piao MJ, Ahn MJ, Kang KA, Ryu YS, Hyun YJ, Shilnikova K, et al. Particulate matter 2.5 damages skin cells by inducing oxidative stress, subcellular organelle dysfunction, and apoptosis. *Arch Toxicol.* (2018) 92:2077–91. doi: 10.1007/s00204-018-2197-9
- 139. Cervellati F, Benedusi M, Manarini F, Woodby B, Russo M, Valacchi G, et al. Proinflammatory properties and oxidative effects of atmospheric particle components in human keratinocytes. *Chemosphere.* (2020) 240:124746. doi: 10.1016/j.chemosphere.2019.124746
- 140. Ovrevik J, Refsnes M, Lag M, Holme JA, aSchwarze PE. Activation of proinflammatory responses in cells of the airway mucosa by particulate matter: oxidant- and non-oxidant-mediated triggering mechanisms. *Biomolecules*. (2015) 5:1399–440. doi: 10.3390/biom5031399
- 141. Kim KE, Cho D, Park HJ. Air pollution and skin diseases: adverse effects of airborne particulate matter on various skin diseases. *Life Sci.* (2016) 152:126–34. doi: 10.1016/j.lfs.2016.03.039
- 142. Marrot L. Pollution and sun exposure: a deleterious synergy. Mechanisms and opportunities for skin protection. *Curr Med Chem.* (2018) 25:5469– 86. doi: 10.2174/0929867324666170918123907
- 143. Vierkotter A, Schikowski T, Ranft U, Sugiri D, Matsui M, Kramer U, et al. Airborne particle exposure and extrinsic skin aging. J Invest Dermatol. (2010) 130:2719–26. doi: 10.1038/jid.20 10.204
- 144. Kim M, Kim JH, Jeong GJ, Park KY, Lee MK, Seo SJ. Particulate matter induces pro-inflammatory cytokines via phosphorylation of p38 MAPK possibly leading to dermal inflammaging. *Exp Dermatol.* (2019) 28:809– 15. doi: 10.1111/exd.13943
- 145. Wang Y, van Boxel-Dezaire AH, Cheon H, Yang J, Stark GR. STAT3 activation in response to IL-6 is prolonged by the binding of IL-6 receptor to EGF receptor. *Proc Natl Acad Sci U S A.* (2013) 110:16975– 80. doi: 10.1073/pnas.1315862110
- 146. Magnani ND, Muresan XM, Belmonte G, Cervellati F, Sticozzi C, Pecorelli A, et al. Skin damage mechanisms related to airborne particulatematter exposure. *Toxicol Sci.* (2016) 149:227–36. doi: 10.1093/toxsci/kfv230

- 147. Mehta NN, Azfar RS, Shin DB, Neimann AL, Troxel AB, Gelfand JM. Patients with severe psoriasis are at increased risk of cardiovascular mortality: cohort study using the general practice research database. *Eur Heart J.* (2010) 31:1000–6. doi: 10.1093/eurheartj/ehp567
- 148. Karbach S, Croxford AL, Oelze M, Schuler R, Minwegen D, Wegner J, et al. Interleukin 17 drives vascular inflammation, endothelial dysfunction, and arterial hypertension in psoriasis-like skin disease. *Arterioscler Thromb Vasc Biol.* (2014) 34:2658–68. doi: 10.1161/ATVBAHA.114.304108
- 149. Schuler R, Brand A, Klebow S, Wild J, Veras FP, Ullmann E, et al. Antagonization of IL-17A attenuates skin inflammation and vascular dysfunction in mouse models of psoriasis. *J Invest Dermatol.* (2019) 139:638– 47. doi: 10.1016/j.jid.2018.09.021
- Kreyling WG, Blanchard JD, Godleski JJ, Haeussermann S, Heyder J, Hutzler P, et al. Anatomic localization of 24- and 96-h particle retention in canine airways. J Appl Physiol. (1999) 87:269–84. doi: 10.1152/jappl.1999.87.1.269
- 151. Moller W, Haussinger K, Winkler-Heil R, Stahlhofen W, Meyer T, Hofmann W, et al. Mucociliary and longterm particle clearance in the airways of healthy nonsmoker subjects. *J Appl Physiol.* (1985) 97:2200– 6. doi: 10.1152/japplphysiol.00970.2003
- 152. Mutlu EA, Engen PA, Soberanes S, Urich D, Forsyth CB, Nigdelioglu R, et al. Particulate matter air pollution causes oxidant-mediated increase in gut permeability in mice. *Part Fibre Toxicol.* (2011) 8:19. doi: 10.1186/1743-8977-8-19
- 153. Al-Sadi R, Guo S, Ye D, Rawat M, Ma TY. TNFalpha modulation of intestinal tight junction permeability is mediated by NIK/IKK-alpha axis activation of the canonical NF-kappaB pathway. *Am J Pathol.* (2016) 186:1151– 65. doi: 10.1016/j.ajpath.2015.12.016
- Cunningham KE Turner JR. Myosin light chain kinase: pulling the strings of epithelial tight junction function. Ann N Y Acad Sci. (2012) 1258:34– 42. doi: 10.1111/j.1749-6632.2012.06526.x
- 155. Kaplan GG, Hubbard J, Korzenik J, Sands BE, Panaccione R, Ghosh S, et al. The inflammatory bowel diseases and ambient air pollution: a novel association. Am J Gastroenterol. (2010) 105:2412–19. doi: 10.1038/ajg.2010.252
- 156. Lomer MC, Hutchinson C, Volkert S, Greenfield SM, Catterall A, Thompson RP, et al. Dietary sources of inorganic microparticles and their intake in healthy subjects and patients with crohn's disease. *Br J Nutr.* (2004) 92:947– 55. doi: 10.1079/BJN20041276
- Lomer MC, Thompson RP, Powell JJ. Fine and ultrafine particles of the diet: influence on the mucosal immune response and association with crohn's disease. *Proc Nutr Soc.* (2002) 61:123–30. doi: 10.1079/PNS2001134
- Ananthakrishnan AN, McGinley EL, Binion DG, Saeian K. Ambient air pollution correlates with hospitalizations for inflammatory bowel disease: an ecologic analysis. *Inflamm Bowel Dis.* (2011) 17:1138– 45. doi: 10.1002/ibd.21455
- 159. Kaplan GG, Dixon E, Panaccione R, Fong A, Chen L, Szyszkowicz M, et al. Effect of ambient air pollution on the incidence of appendicitis. *CMAJ.* (2009) 181:591–7. doi: 10.1503/cmaj.082068
- Gerhardsson de Verdier M, Plato N, Steineck G, Peters JM. Occupational exposures and cancer of the colon and rectum. Am J Ind Med. (1992) 22:291–303. doi: 10.1002/ajim.4700220303
- 161. Guberan E, Usel M, Raymond L, Bolay J, Fioretta G, Puissant J. Increased risk for lung cancer and for cancer of the gastrointestinal tract among Geneva professional drivers. Br J Ind Med. (1992) 49:337– 44. doi: 10.1136/oem.49.5.337
- Chen JC, Schwartz J. Neurobehavioral effects of ambient air pollution on cognitive performance in US adults. *Neurotoxicology*. (2009) 30:231– 9. doi: 10.1016/j.neuro.2008.12.011
- 163. Calderón-Garcidueñas L, Engle R, Mora-Tiscareño A, Styner M, Gómez-Garza G, Zhu H, et al. Exposure to severe urban air pollution influences cognitive outcomes, brain volume and systemic inflammation in clinically healthy children. *Brain Cogn.* (2011) 77:345–55. doi: 10.1016/j.bandc.2011.09.006
- 164. Heusinkveld HJ, Wahle T, Campbell A, Westerink RHS, Tran L, Johnston H, et al. Neurodegenerative and neurological disorders by small inhaled particles. *Neurotoxicology.* (2016) 56:94–106. doi: 10.1016/j.neuro.2016.07.007

- 165. Costa LG, Cole TB, Coburn J, Chang YC, Dao K, Roqué PJ. Neurotoxicity of traffic-related air pollution. *Neurotoxicology*. (2017) 59:133–9. doi: 10.1016/j.neuro.2015.11.008
- 166. Kelly FJ, Fussell JC. Role of oxidative stress in cardiovascular disease outcomes following exposure to ambient air pollution. *Free Radic Biol Med.* (2017) 110:345–67. doi: 10.1016/j.freeradbiomed.2017.06.019
- 167. Miyata R, van Eeden SF. The innate and adaptive immune response induced by alveolar macrophages exposed to ambient particulate matter. *Toxicol Appl Pharmacol.* (2011) 257:209–26. doi: 10.1016/j.taap.2011.09.007
- 168. Ghio AJ, Soukup JM, Case M, Dailey LA, Richards J, Berntsen J, et al. Exposure to wood smoke particles produces inflammation in healthy volunteers. *Occup Environ Med.* (2012) 69:170– 75. doi: 10.1136/oem.2011.065276
- 169. Ghio AJ, Suliman HB, Carter JD, Abushamaa AM, Folz RJ. Overexpression of extracellular superoxide dismutase decreases lung injury after exposure to oil fly ash. Am J Physiol Lung Cell Mol Physiol. (2002) 283:L211– 8. doi: 10.1152/ajplung.00409.2001
- Marchini T, Magnani ND, Paz ML, Vanasco V, Tasat D, Gonzalez Maglio DH, et al. Time course of systemic oxidative stress and inflammatory response induced by an acute exposure to residual oil fly ash. *Toxicol Appl Pharmacol.* (2014) 274:274–82. doi: 10.1016/j.taap.2013.11.013
- 171. Pope CA 3rd, Bhatnagar A, McCracken JP, Abplanalp W, Conklin DJ, O'Toole T. Exposure to fine particulate air pollution is associated with endothelial injury and systemic inflammation. *Circ Res.* (2016) 119:1204– 14. doi: 10.1161/CIRCRESAHA.116.309279
- 172. Marchini T, D'Annunzio V, Paz ML, Cáceres L, Garcés M, Perez V, et al. Selective TNF-α targeting with infliximab attenuates impaired oxygen metabolism and contractile function induced by an acute exposure to air particulate matter. *Am J Physiol Heart Circ Physiol.* (2015) 309:H1621–8. doi: 10.1152/ajpheart.00359.2015
- 173. Alexis NE, Lay JC, Zeman KL, Geiser M, Kapp N, Bennett WD. In vivo particle uptake by airway macrophages in healthy volunteers. Am J Respir Cell Mol Biol. (2006) 34:305–13. doi: 10.1165/rcmb.2005-0373OC
- 174. Li N, Sioutas C, Cho A, Schmitz D, Misra C, Sempf J, et al. Ultrafine particulate pollutants induce oxidative stress and mitochondrial damage. *Environ Health Perspect.* (2003) 111:455–60. doi: 10.1289/ehp.6000
- 175. Hiraiwa K, van Eeden SF. Contribution of lung macrophages to the inflammatory responses induced by exposure to air pollutants. *Mediators Inflamm.* (2013) 2013:619523. doi: 10.1155/2013/619523
- 176. Zhao Q, Chen H, Yang T, Rui W, Liu F, Zhang F, et al. Direct effects of airborne PM2.5 exposure on macrophage polarizations. *Biochim Biophys Acta*. (2016) 1860:2835–43. doi: 10.1016/j.bbagen.2016.03.033
- 177. Muñoz-Planillo R, Kuffa P, Martínez-Colón G, Smith BL, Rajendiran TM, Núñez G. K⁺ efflux is the common trigger of NLRP3 inflammasome activation by bacterial toxins and particulate matter. *Immunity*. (2013) 38:1142–53. doi: 10.1016/j.immuni.2013.05.016
- 178. Li R, Kou X, Geng H, Xie J, Yang Z, Zhang Y, et al. Effect of ambient PM(2.5) on lung mitochondrial damage and fusion/fission gene expression in rats. *Chem Res Toxicol.* (2015) 28:408–18. doi: 10.1021/tx5003723
- 179. Xia T, Korge P, Weiss JN, Li N, Venkatesen MI, Sioutas C, et al. Quinones and aromatic chemical compounds in particulate matter induce mitochondrial dysfunction: implications for ultrafine particle toxicity. *Environ Health Perspect.* (2004) 112:1347–58. doi: 10.1289/ehp.7167
- 180. Mutlu GM, Green D, Bellmeyer A, Baker CM, Burgess Z, Rajamannan N, et al. Ambient particulate matter accelerates coagulation via an IL-6-dependent pathway. *J Clin Invest*. (2007) 117:2952–61. doi: 10.1172/JCI30639
- 181. Marchini T, Wolf D, Michel NA, Mauler M, Dufner B, Hoppe N, et al. Acute exposure to air pollution particulate matter aggravates experimental myocardial infarction in mice by potentiating cytokine secretion from lung macrophages. *Basic Res Cardiol.* (2016) 111:44. doi: 10.1007/s00395-016-0562-5
- 182. Soberanes S, Misharin AV, Jairaman A, Morales-Nebreda L, McQuattie-Pimentel AC, Cho T, et al. Metformin targets mitochondrial electron transport to reduce air-pollution-induced thrombosis. *Cell Metab.* (2019) 29:503. doi: 10.1016/j.cmet.2018.09.019
- 183. Chiarella SE, Budinger GR, Mutlu GM. β_2 -Agonist therapy may contribute to the air pollution and IL-6-associated risk of developing severe asthma

with dual-positive TH2/TH17 cells. J Allergy Clin Immunol. (2015) 135:290-1. doi: 10.1016/j.jaci.2014.09.037

- 184. Gurgueira SA, Lawrence J, Coull B, Murthy GG, González-Flecha B. Rapid increases in the steady-state concentration of reactive oxygen species in the lungs and heart after particulate air pollution inhalation. *Environ Health Perspect.* (2002) 110:749–55. doi: 10.1289/ehp.02110749
- Magnani ND, Marchini T, Tasat DR, Alvarez S, Evelson PA. Lung oxidative metabolism after exposure to ambient particles. *Biochem Biophys Res Commun.* (2011) 412:667–72. doi: 10.1016/j.bbrc.2011.08.021
- 186. Magnani ND, Marchini T, Vanasco V, Tasat DR, Alvarez S, Evelson P. Reactive oxygen species produced by NADPH oxidase and mitochondrial dysfunction in lung after an acute exposure to residual oil fly ashes. *Toxicol Appl Pharmacol.* (2013) 270:31–38. doi: 10.1016/j.taap.2013.04.002
- 187. Marchini T, Magnani N, D'Annunzio V, Tasat D, Gelpi RJ, Alvarez S, et al. Impaired cardiac mitochondrial function and contractile reserve following an acute exposure to environmental particulate matter. *Biochim Biophys Acta*. (2013) 1830:2545–52. doi: 10.1016/j.bbagen.2012.11.012
- Kowaltowski AJ, de Souza-Pinto NC, Castilho RF, Vercesi AE. Mitochondria and reactive oxygen species. *Free Radic Biol Med.* (2009) 47:333– 43. doi: 10.1016/j.freeradbiomed.2009.05.004
- 189. Beer M, Seyfarth T, Sandstede J, Landschütz W, Lipke C, Köstler H, et al. Absolute concentrations of high-energy phosphate metabolites in normal, hypertrophied, and failing human myocardium measured noninvasively with (31)P-SLOOP magnetic resonance spectroscopy. J Am Coll Cardiol. (2002) 40:1267–74. doi: 10.1016/S0735-1097(02)02160-5
- Neubauer S. The failing heart-an engine out of fuel. N Engl J Med. (2007) 356:1140–51. doi: 10.1056/NEJMra063052
- 191. Gorr MW, Youtz DJ, Eichenseer CM, Smith KE, Nelin TD, Cormet-Boyaka E, et al. *In vitro* particulate matter exposure causes direct and lung-mediated indirect effects on cardiomyocyte function. *Am J Physiol Heart Circ Physiol.* (2015) 309:H53–62. doi: 10.1152/ajpheart.00162. 2015

- 192. Wold LE, Ying Z, Hutchinson KR, Velten M, Gorr MW, Velten C, et al. Cardiovascular remodeling in response to long-term exposure to fine particulate matter air pollution. *Circ Heart Fail.* (2012) 5:452– 61. doi: 10.1161/CIRCHEARTFAILURE.112.966580
- 193. Pearson JF, Bachireddy C, Shyamprasad S, Goldfine AB, Brownstein JS. Association between fine particulate matter and diabetes prevalence in the U.S. Diabetes Care. (2010) 33:2196–201. doi: 10.2337/dc10-0698
- 194. Brook RD, Cakmak S, Turner MC, Brook JR, Crouse DL, Peters PA, et al. Long-term fine particulate matter exposure and mortality from diabetes in Canada. *Diabetes Care*. (2013) 36:3313–20. doi: 10.2337/dc12-2189
- 195. Sun Q, Yue P, Deiuliis JA, Lumeng CN, Kampfrath T, Mikolaj MB, et al. Ambient air pollution exaggerates adipose inflammation and insulin resistance in a mouse model of diet-induced obesity. *Circulation.* (2009) 119:538–46. doi: 10.1161/CIRCULATIONAHA.108.799015
- 196. Xu X, Liu C, Xu Z, Tzan K, Zhong M, Wang A, et al. Long-term exposure to ambient fine particulate pollution induces insulin resistance and mitochondrial alteration in adipose tissue. *Toxicol Sci.* (2011) 124:88– 98. doi: 10.1093/toxsci/kfr211
- Ricquier D. Respiration uncoupling and metabolism in the control of energy expenditure. *Proc Nutr Soc.* (2005) 64:47–52. doi: 10.1079/PNS2004408

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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