Hindawi Minimally Invasive Surgery Volume 2021, Article ID 5564745, 9 pages https://doi.org/10.1155/2021/5564745

Review Article

The "Dark Side" of Pneumoperitoneum and Laparoscopy

Giuseppina Rosaria Umano, Giulia Delehaye, Carmine Noviello, and Alfonso Papparella 🕞

Department of Woman, Child, and General and Specialized Surgery, University of Campania Luigi Vanvitelli, Largo Madonna Delle Grazie N. 1, Naples 8013, Italy

Correspondence should be addressed to Alfonso Papparella; alfonso.papparella@unicampania.it

Received 8 January 2021; Revised 29 April 2021; Accepted 8 May 2021; Published 21 May 2021

Academic Editor: Peng Hui Wang

Copyright © 2021 Giuseppina Rosaria Umano et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Laparoscopic surgery has been one of the most common procedures for abdominal surgery at pediatric age during the last few decades as it has several advantages compared to laparotomy, such as shorter hospital stays, less pain, and better cosmetic results. However, it is associated with both local and systemic modifications. Recent evidence demonstrated that carbon dioxide pneumoperitoneum might be modulated in terms of pressure, duration, temperature, and humidity to mitigate and modulate these changes. The aim of this study is to review the current knowledge about animal and human models investigating pneumoperitoneum-related biological and histological impairment. In particular, pneumoperitoneum is associated with local and systemic inflammation, acidosis, oxidative stress, mesothelium lining abnormalities, and adhesion development. Animal studies reported that an increase in pressure and time and a decrease in humidity and temperature might enhance the rate of comorbidities. However, to date, few studies were conducted on humans; therefore, this research field should be further investigated to confirm in experimental models and humans how to improve laparoscopic procedures in the spirit of minimally invasive surgeries.

1. Introduction

Laparoscopy has progressively substituted abdominal open surgery for many procedures as it has several advantages compared to laparotomy, even in children. In particular, laparoscopy is associated with better cosmetic results, shorter hospital stays, lower postoperative pain, and a faster return to daily activities [1]. However, it is not free from short- and long-term comorbidities, such as light pain, abdominal discomfort, and adhesion development [2–4].

The pneumoperitoneum is essential for laparoscopic surgery, as it enables visibility and mobility in the working site. Nevertheless, the pneumoperitoneum modifies abdominal-cavity homeostasis and might promote metabolic changes through mechanical and biochemical effects [5–8].

Historically, several gases were used to induce a pneumoperitoneum with the aim of reducing the incidence of local and systemic adverse effects. Helium and argon are inert gases that are compromised by low solubility. Evidence

reported an increased rate of subcutaneous emphysema and venous gas embolism after helium insufflation [9]. Conversely, carbon dioxide (CO₂) has 50-fold higher solubility compared to helium. Argon, like helium, is almost insoluble in blood, so it is associated with a higher risk of embolism. In addition, it induces cardiac depression [10]. Nitrous oxide was the preferred gas during the 1970s, but it is not used in current practice because of the risk of explosion. In fact, nitrous oxide is highly flammable and causes explosion during electrocautery [5].

On the basis of this evidence, CO_2 has become a more suitable gas for pneumoperitoneum creation, as it is costeffective, noncombustible, and soluble, and it is rapidly excreted with respiration. However, recent studies suggest that the use of CO_2 during laparoscopic surgery is not free from adverse effects. The use of this gas might induce metabolic, immune, and structural modifications that can have clinical impact [11, 12].

Previous papers have shown that chemical, physical, and biological features of pneumoperitoneum might affect clinical outcomes, such as postoperative pain, length of stay, and recovery time [13–18].

These data suggest that chemical, physical, and biological effects of CO2 on peritoneum cause inflammation and tissue modifications [19]. In particular, we have seen that the grade of morphological changes is related to the level of intrabdominal pressure. CO2 pneumoperitoneum evokes an inflammatory response by modifying factors such as plasmatic and peritoneal chemokines levels [20].

To further investigate these aspects of laparoscopy, we reviewed the current research with regards to animal and human studies investigating peritoneal and systemic alterations during and after carbon dioxide pneumoperitoneum.

2. Materials and Methods

We performed a narrative search of Medline and PubMed databases with terms covering "laparoscopy", "pneumoperitoneum", "inflammation", "mesothelium", "ischemia–reperfusion", and "adherence" published between January 1993 and April 2021. These terms were used in different combinations. Only English language papers were included. Articles were evaluated for scientific relevance and pertinence to the effect of pneumoperitoneum on peritoneal homeostasis impairment. Relevant references from selected papers were included. Study selection was executed by two authors; if a disagreement occurred, it was solved through analysis with the other coauthors (Figure 1).

3. Results

The literature search revealed 621 articles. The majority of the selected papers investigated pneumoperitoneum effects in animal models: rats (n = 21), pigs (n = 4), rabbits (n = 1), and dogs (n = 1). Few papers were based on human studies (n = 20). Here, we report the results according to biological, metabolic, and histological alterations. Figure 2 summarizes the effects of pneumoperitoneum and its variables on the peritoneal environment.

3.1. Inflammation. Intentional or accidental peritoneal injury induces a local acute phase reaction, namely, inflammation [21]. Inflammatory response is characterized by cellular infiltration and the release of proinflammatory cytokines such as IL-6, IL-1, and TNF- α . Surgery acts as local trauma and leads to both local and systemic inflammation. Laparoscopy was shown to produce a less severe peritoneal reaction compared to that of an open approach [22, 23]. Therefore, mini-invasive surgery is associated with lower pain and postoperative hospital stay [24-26]. However, pneumoperitoneum parameters might be modulated during laparoscopy with the aim of reducing early changes related to CO2 insufflation. In 2007, Papparella et al. reported that peritoneal insufflation with CO2 in rats was associated with eosinophils, mastocytes, and macrophages 2h after laparoscopic surgery. The cellular response was also detectable at 24 h afterwards. The authors reported that peritoneal modification was lower in CO₂ compared to the room-air insufflation group; nevertheless, carbon

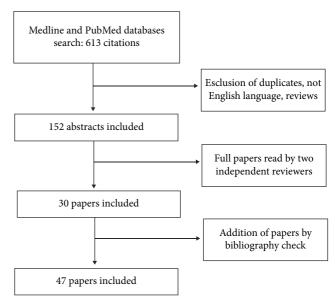


FIGURE 1: Flowchart of papers selection.

pneumoperitoneum resulted in an inflammatory response with respect to the control group [19]. Similar results were described by Ure et al. in pigs undergoing laparoscopy with room-air insufflation or CO_2 pneumoperitoneum. Carbon dioxide pneumoperitoneum induced a lower release of IL-6 compared to that of air, whereas no significant difference for IL-1 and TNF- α levels was found [27].

In addition to gas type, insufflation pressure was reported to affect peritoneal mucosa integrity and cellular infiltration. In a rat model, low pneumoperitoneum pressure was associated with less intense cellular inflammatory response in terms of eosinophil granulocyte, macrophage, mast-cell, and lymphocyte composition compared to high pneumoperitoneum pressure 24 h after surgery. Therefore, there is a clear relationship between the grade of mucosa inflammation and level of pressure [28]. Recently, similar findings were observed by Poerwosusanta et al. in rats exposed to 8, 10, and 12 mmHg pneumoperitoneum. The authors found a progressive and significant increase in mastcell peritoneal infiltration and degranulation with an increase in pressure 7 days after surgery [29]. Paralleling with local acute response, gas pressure was described as a factor influencing systemic inflammation. In fact, lower pneumoperitoneum pressure has been associated with less intense inflammatory cellular response after laparoscopy in subjects undergoing colorectal surgery [30]. Moreover, higher plasma TNF-α levels were found in 10 mm Hg pneumoperitoneum rats compared to those in 6 mm Hg and control groups. The three groups showed different IL-6 plasma levels, but the difference was not statistically significant [20]. Furthermore, the same authors showed systemic and local modifications of Rantes/CCL5 and MCP1/ CCL2 that could have a role in these changes. The authors postulated that modifying these factors could make laparoscopic surgery less invasive. To date, there are few available studies investigating the relation between pneumoperitoneum features and inflammatory response in human

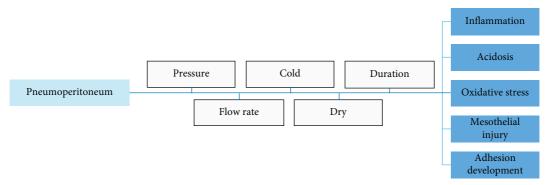


FIGURE 2: Pneumoperitoneum characteristics and related peritoneal effects.

models. Some evidence highlights lower cytokine release, stress-hormone levels, and plasma inflammatory markers after laparoscopy compared to laparotomy in both adults and children [31, 32]. Therefore, we need more studies to confirm the experimental findings about pneumoperitoneum influence in human inflammatory response.

3.2. Acidosis. Carbon dioxide induces the acidification of fluids, as it has the potential to induce carbonic acid reaction that dissociates in bicarbonate (HCO3-) and hydrogen ions (H+) [33]. This phenomenon is well-recognized to occur in blood and cerebrospinal fluid [33]; therefore, several experimental studies investigated the potential ability of CO₂ insufflation in inducing peritoneal acidosis. An experimental study in dogs reported that CO₂ abdominal insufflation induced acute peritoneal pH reduction after 10–20 minutes. This acidosis was also detectable in the blood, whereas laparotomy was not associated with any pH modification during and after surgery [34].

Wong et al. compared the insufflation effect of different gas types on pH modifications in pigs. They found that only CO₂ was associated with acute peritoneal acidosis, whereas nitrous oxide and helium were not. In addition, peritoneal acidification was more severe with higher-pressure insufflation (12–15 versus 5–8 mmHg) [35]. Similarly, Hanly et al. found no abdominal acidification after helium or air insufflation and concluded that peritoneal pH changes during laparoscopy are induced by the specific activity of CO2 and not by mechanical abdominal distention [36]. The insufflation effect of different gases in rats was also investigated by Kuntz and colleagues [37]. They evaluated pH modifications in the blood, subcutaneous fat tissue, and the abdominal cavity after insufflation of CO₂, air, and helium at different intra-abdominal pressure levels (0, 3, 6, and 9 mmHg). The authors reported that the CO2 effect differed according to site. In particular, CO₂ significantly decreased blood and subcutaneous pH compared to air and helium. The influence of pressure on pH reduction was more effective than that of pneumoperitoneum length [37].

Conversely, an experimental study in rats exposed to different pressure levels and insufflation rates failed to find significant effects on peritoneal ph. In fact, pH rapidly decreased in the site of manipulation independently of pressure and insufflation rates. Moreover, acidification was

only identified in areas directly exposed to the gas, whereas distal areas showed slight and not significant pH reduction [38].

Other physical CO₂ characteristics were shown to induce pH changes in the abdominal cavity. An experimental swine-model compared peritoneal pH in four different groups: standard laparoscopy, laparoscopy with heated and humidified gas, laparoscopy with the administration of heated humidified gas and bicarbonate, and laparotomy. All laparoscopy groups displayed peritoneal acidification, whereas laparotomy did not. The addition of bicarbonate was ineffective in contrasting CO₂ acidification. Conversely, the heated and humidified group showed lower pH reduction compared to that of standard laparoscopy. Heating and humidification could enhance local vascularization and subsequently CO₂ peritoneal uptake [39].

Nevertheless, the investigation of peritoneal acidosis during laparoscopy is an interesting field of research, as it might be associated with local changes of immune response that could benefit the patient. In fact, evidence reported changes of cytokine levels, namely, IL-10 increase and TNF- α decrease, in response to bacterial lipopolysaccharides after CO₂ insufflation [40]. Moreover, acidosis reduces macrophage activation [41]. Therefore, by modulating CO₂ insufflation, it is possible to influence the inflammatory response and peritoneal homeostasis.

3.3. Oxidative Stress. During laparoscopy, CO_2 insufflation in the abdominal cavity is necessary to visualize the working space; pressure and gas insufflation rate have a pivotal role in this phenomenon. However, increased intra-abdominal pressure might compromise splanchnic perfusion and capillary microcirculation and lead to reduced oxygen content and related ischemia–reperfusion injury with the consequent production of reactive oxygen species (ROS) [42–47].

ROS release, microcirculation impairment, and tissue hypoxia are the first steps of peritoneal injury, ranging from mesothelial denudation to inflammation and adhesion development [48]. Therefore, efforts in reducing oxidative-stress damage might be encouraged to prevent long-term laparoscopy-related comorbidities. Several animal and human studies indicated the role of pneumoperitoneal length and pressure in ischemia–reperfusion damage. Molinas et al.

reported that tissue hypoxia increased with longer pneumoperitoneal length and higher insufflation rate in a murine model. These parameters were associated with adhesion development, and the addition of 2%–3% oxygen to insufflation gas was effective in reducing peritoneal fibrosis [49]. Accordingly, low intra-abdominal pressure did not affect peritoneal-tissue oxygen tension during laparoscopy in rats compared to laparotomy, confirming that less pressure causes minor damage to capillary flow and consequently tissue injury [50]. In particular, other authors reported that pressure of at least 12 mmHg significantly reduces splanchnic blood flow in rats [51].

Contrasting results were reported by Giannotti and coworkers, who failed to find compromised gut partial tissue oxygen tension during laparoscopy for left colon resection versus laparotomy [52]. Laparoscopy was conversely associated with an increased tissue oxygen tension that was equally decreased after mesenterial traction in laparoscopy and laparotomy groups. However, the authors observed a rise in plasmatic markers of oxidative stress with 15 mmHg pneumoperitoneal pressure [52]. Previous studies reported similar findings about the increase in oxidative plasma markers after laparoscopy. In 1998, Taskin et al. reported changes in free radical scavengers, namely, glutathione, glutathione peroxidase, superoxide dismutase, and catalase in 28 women undergoing operative laparoscopy. Modifications were directly associated with CO2 exposure length and insufflation rate [53]. Similarly, Polat reported that 15 mmHg pressure was associated with a significant increase in plasmatic malondialdehyde, a maker of lipid peroxidation, compared to 10 mmHg pneumoperitoneum [54]. Moreover, recently, Poerwosusanta et al. showed that H₂O₂ and malondialdehyde serum levels, as a direct expression of oxidative stress, significantly increased with the rise of insufflation pressure in rats. In addition, the levels of catalase and superoxide dismutase decreased in terms of oxidativestress index. These changes were correlated with cellular inflammation and mesothelial-cell abnormalities [29].

In addition to minimizing intra-abdominal pressure, ischemic preconditioning was investigated as a method to reduce laparoscopy-induced oxidative stress. It consists of short ischemia–reperfusion cycles during early laparoscopy that might mitigate reoxygenation injury. Several experimental animal models ruled out the ability of preconditioning in reducing oxidative-stress injury in organs and plasma [55–58].

To date, there have been only two studies investigating ischemic preconditioning in humans. Kolbenschlag J et al. performed remote ischemic conditioning consisting of cycles of ischemia–reperfusion applied to the upper arm that were followed by improvement in skin microcirculation in the leg of 60 healthy volunteers [59]. More interesting are the results from the study of Veres and colleagues. They randomized 30 patients undergoing laparoscopic surgery to standard therapy and preconditioning and evaluated changes in plasmatic oxidative-stress markers. The ischemic preconditioning group showed significant improvement in glutathione levels and a significant reduction in pain according to the visual analog scale (VAS). Therefore, the

authors concluded that this modification of pneumoperitoneum induction is a promising tool in limiting oxidative stress during laparoscopy [60]. However, more studies are needed to confirm this extremely preliminary evidence.

3.4. Mesothelial Modifications. The peritoneum is constituted by a single layer of mesothelial cells with an underlying basal membrane and connective tissue containing immune cells, fibroblasts, collagen, and blood vessels [61, 62]. It is involved in the regulation of several biologic processes, such as inflammation, angiogenesis, fibrinolysis, and tissue remodeling. Any kind of mechanical and chemical insult activates inflammation and fibrotic tissue deposition [62]. Therefore, the preservation of the mesothelial lining is crucial for reducing postsurgical adhesions. Animal studies described mesothelium changes during and after a pneumoperitoneum with electron microscopy. Volz and coworkers found cellular retraction and bulging up, intercellular clefts, and the exposition of basal lamina from 60 to 120 minutes after pneumoperitoneum release. These alterations involved the entire abdominal cavity and worsened until 12 hours after surgery. Mesothelial integrity was restored 2 days after surgery [63]. Similar findings were proposed by Suematsu et al., who compared mesothelial characteristics after the insufflation of different gases, namely, helium, air, and carbon dioxide. CO₂ induced early cellular bulging up that reverted after 24 h. However, intercellular clefts and inflammatory infiltration were detectable at 24 h. Helium pneumoperitoneum was associated with more severe and persistent alterations [64]. Conversely, in another murine experimental model, peritoneal changes were not associated with gas type, whereas morphological peritoneal modifications were dependent on increased pneumoperitoneum period, insufflation flow, and pressure [65]. In addition, gas characteristics in terms of temperature and humidity were related to peritoneal injury. Standard pneumoperitoneum is induced with cold and dry gas [5, 6, 66] that induces dehydration and morphological alterations of the peritoneum. Therefore, the use of heated humidified gas was proven to prevent intercellular clefts [9], basal lamina exposure [67], and adhesion formation [68]. In contrast, in the study of Hazebroek et al., the effect on the mesothelial layer was independent from CO₂ temperature and humidity [69]. However, previous findings were confirmed by Sampurno et al. who performed randomized laparoscopic rectal resection in 16 pigs to receive dry-cold or humidified-heated CO₂. The dry-cold group showed significant early mesothelial modifications, such as cellular retraction and bulging, and microvillus alterations compared to the humidified-heated group [70]. Similarly, another experimental swine-model study reported reduced mesothelial thickness in pigs undergoing high-pressure pneumoperitoneum 7 days after laparoscopy. Damage was directly correlated with the increase in pressure and mastcell infiltration. Mast-cell activation and degranulation could lead to the release of metalloproteinases that mediate basal laminal damage and subsequent fibrosis [29].

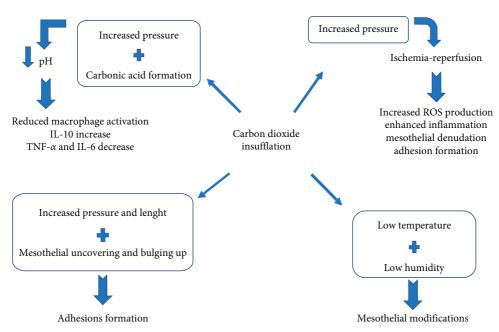


FIGURE 3: Summary of the mechanisms underlying peritoneal acidosis, oxidative stress, mesothelial alterations, and adhesions formation after CO₂ insufflation.

Few studies investigated mesothelial damage after pneumoperitoneum in humans [71, 72]. Tarhan et al. found mesothelial-cell apoptosis, cellular bulging up, intercellular clefts, and ultrastructural alterations in 20 biopsy specimens of the working area after laparoscopic cholecystectomy [72]. Therefore, this evidence highlights the role of different pneumoperitoneum parameters such as temperature, pressure, flow rate, and humidity in inducing mesothelial damage.

3.5. Adhesion Development. Peritoneal adhesions result in the abnormal repair of the peritoneum caused by surgical trauma, inflammation, and infection [62, 73, 74]. Consequently, the peritoneum promotes a healing process covering the damaged tissue with fibrin plug [75]. Normally, the fibrin deposit is entirely removed from the fibrinolytic system, and mesothelial regeneration is achieved in about 8 days [62,74]. Fibrinolytic activity can be compromised due to ischemic, inflammatory, or thermal insults causing fibroblast proliferation and adherence development [62,74].

During laparoscopy, the pneumoperitoneum can cause inflammatory phenomena that result in adhesions in relation to the type, temperature, and pressure of the insufflated gas and the length of pneumoperitoneum. Jacobi et al. observed a reduction in adhesions in mice undergoing laparoscopic colectomy using helium rather than CO₂ [76].

If we consider the influence of intrabdominal pressure and the perfusion injury, we may speculate that increased intra-abdominal oxygen tension could be beneficial to the patient. In fact, in 2004, Binda et al. noted that hypothermia in an animal model prevented the formation of adhesions through a reduction in cellular oxygen needs. Subsequently, better results were obtained when humidifying the

insufflated gas and adding 3% O₂ [77]. Molinas et al. gained similar results in a prospective randomized trial. They supposed that the pneumoperitoneum can increase adhesion formation promoting hypoxemia due to the compression of capillary flow. The authors showed that intensifying the insufflation of CO₂ or helium from 10 to 45 min increased the total adhesion score when adding 4% O2. No difference was observed between CO2 and helium [48]. In a randomized and controlled trial of 44 women undergoing deep endometriosis surgery, a decreased degree of adherence formation was found by inducing the pneumoperitoneum with a cooled and humidified gas composed of a mixture of 86% CO₂, 10% N₂O, and 4% O₂ [78]. These findings are in accordance with the clinical report of reduced pain complaint, length of stay, and analgesic use in subjects receiving heated humidified CO2 compared to those treated with dry

In 2007, Brokelman et al. reported that, in patients undergoing laparoscopic cholecystectomy using heated CO_2 , the level of plasminogen activator inhibitor type 1 (PAI-1) that hampered its activation was significantly reduced. Postoperative adhesions were thereby also reduced [80]. The same group observed a reduction in $TGF\beta1$, a PAI-1 promoter, when heating CO_2 during pneumoperitoneum [81].

In addition to temperature, experimental data suggested that insufflation pressure can modify adhesions development. In 2011, Matsuzaki et al. compared the impact of intraperitoneal pressure of the pneumoperitoneum [82] in human and animal models. They measured the ratio of tissue plasminogen activator (tPA) and plasminogen activator inhibitor type 1 (PAI-1) in patients undergoing laparoscopic hysterectomy and subdivided them into two groups: standard-IPP (12 mmHg) and low-IPP (8 mmHg). After 2 hours

of CO₂ pneumoperitoneum, the tPA/PAI-1 ratio was significantly decreased in the standard-IPP group. Similar results were found in animal models with mice exposed to 2 mmHg (low IPP) and 8 mmHg (high IPP). Results showed that low IPP and the shorter length of laparoscopic surgery might reduce the impact of fibrinolytic activity and prevent the surgical adhesions development in the peritoneal cavity [82].

4. Conclusions

In the era of minimally invasive surgery, knowledge of the basic physiological mechanisms that are modified by laparoscopy (Figure 3) and pneumoperitoneum is essential in making this surgical approach even less invasive. The actor responsible for these changes after laparoscopy is the lining that covers the abdominal cavity, the peritoneum, which maintains an extraordinary balance between all of its components and factors, such as chemokines, cytokines that play an important role in the activation and recruitment of leukocytes to inflammatory sites. Laparoscopy influences both peritoneal integrity and its biology, alters the immune system, and evokes peritoneal acidosis by CO2 insufflation and its influence on microcirculation. It is not clear how all these factors interact with each other and how they can be clinically translated, but knowledge of these mechanisms is the real challenge in truly minimally invasive surgery.

Conflicts of Interest

The authors declare no conflicts of interest.

Authors' Contributions

A. Papparella contributed to conceptualization and supervision. G. Delehaye and G. R. Umano contributed to methodology. G. Delehaye and C. Noviello wrote the original draft. A. Papparella and G. R. Umano reviewed and edited the article. All authors have read and agreed to the published version of the manuscript.

References

- T. Nechay, S. Titkova, A. Tyagunov, M. Anurov, and A. Sazhin, "Modified enhanced recovery after surgery protocol in patients with acute cholecystitis: efficacy, safety and feasibility. Multicenter randomized control study," *Updates in Surgery*, 2021.
- [2] J. López-Torres López, B. Cifuentes García, L. Fernández Ruipérez et al., "Predictive factors of admission in outpatient laparoscopic surgery," *Cirugía Española (English Edition)*, vol. 99, no. 2, pp. 140–146, 2021.
- [3] D. Garteiz-Martínez, E. Rodríguez-Ayala, A. Weber-Sánchez, C. Bravo-Torreblanca, and R. Carbó-Romano, "Pulmonary recruitment can reduce residual pneumoperitoneum and shoulder pain in conventional laparoscopic procedures: results of a randomized controlled trial," Surgical Endoscopy, 2020.
- [4] S. Wada, Y. Fukushi, M Nishimura et al., "Analysis of risk factors of postlaparoscopic shoulder pain," *The Journal of*

- Obstetrics and Gynaecology Research, vol. 46, no. 46, pp. 310-313, 2020.
- [5] D. E. Ott, "Shakespeare's view of the laparoscopic pneumoperitoneum," *Journal of the Society of Laparoendoscopic Surgeons*, vol. 15, no. 3, pp. 282–284, 2011.
- [6] D. E. Ott, "Abdominal compliance and laparoscopy: a review," JSLS, vol. 23, Article ID e2018, 2019.
- [7] G. Mazzinari, O. Diaz-Cambronero, A. Serpa Neto et al., "Modeling intra-abdominal volume and respiratory driving pressure during pneumoperitoneum insufflation-a patientlevel data meta-analysis," *Journal of Applied Physiology*, vol. 130, pp. 721–728, 1985.
- [8] P. Groene, U. Gündogar, K. Hofmann-Kiefer, and R. Ladurner, "Influence of insufflated carbon dioxide on abdominal temperature compared to oesophageal temperature during laparoscopic surgery," Surgical Endoscopy, 2020.
- [9] A. K. Davey, J. Hayward, J. K. Marshall, and A. E. Woods, "The effects of insufflation conditions on rat mesothelium," *International Journal of Inflammation*, vol. 2013, Article ID 816283, 2013.
- [10] C. Mann, G. Boccara, V. Grevy, F. Navarro, J. M. Fabre, and P. Colson, "Argon pneumoperitoneum is more dangerous than CO2 pneumoperitoneum during venous gas embolism," *Anesthesia & Analgesia*, vol. 85, no. 6, pp. 1367–1371, 1997.
- [11] S. J. Neuhaus, A. Gupta, and D. I. Watson, "Helium and other alternative insufflation gases for laparoscopy," *Surgical Endoscopy*, vol. 15, no. 6, pp. 553–560, 2001.
- [12] M. Rohloff, G. Peifer, J. Shakuri-Rad, and T. J. Maatman, "The impact of low pressure pneumoperitoneum in robotic assisted radical prostatectomy: a prospective, randomized, double blinded trial," World Journal of Urology, 2020.
- [13] T. S. Feng, G. Heulitt, A. Islam, and J. R. Porter, "Comparison of valve-less and standard insufflation on pneumoperitoneum-related complications in robotic partial nephrectomy: a prospective randomized trial," *Journal of Robotic Surgery*, 2020.
- [14] K. I. Albers, F. Polat, I. F. Panhuizen et al., "The effect of low-versus normal-pressure pneumoperitoneum during laparoscopic colorectal surgery on the early quality of recovery with perioperative care according to the enhanced recovery principles (RECOVER): study protocol for a randomized controlled study," *Trials*, vol. 21, p. 541, 2020.
- [15] C. E. Foley, E. Ryan, and J. Q. Huang, "Less is more: clinical impact of decreasing pneumoperitoneum pressures during robotic surgery," *Journal of Robotic Surgery*, vol. 15, no. 2, pp. 299–307, 2021.
- [16] A. D. Raval, S. Deshpande, M. Koufopoulou et al., "The impact of intra-abdominal pressure on perioperative outcomes in laparoscopic cholecystectomy: a systematic review and network meta-analysis of randomized controlled trials," *Surgical Endoscopy*, vol. 34, no. 7, pp. 2878–2890, 2020.
- [17] S. Celarier, S. Monziols, M. O. Francois et al., "Randomized trial comparing low-pressure versus standard-pressure pneumoperitoneum in laparoscopic colectomy: PAROS trial," *Trials*, vol. 21, p. 216, 2020.
- [18] K.-F. Hsu, C.-J. Chen, J.-C. Yu et al., "A novel strategy of laparoscopic insufflation rate improving shoulder pain: prospective randomized study," *Journal of Gastrointestinal Surgery*, vol. 23, no. 10, pp. 2049–2053, 2019.
- [19] A. Papparella, C. Noviello, M. Romano, P. Parmeggiani, O. Paciello, and S. Papparella, "Local and systemic impact of pnemoperitoneum on prepuberal rats," *Pediatric Surgery International*, vol. 23, no. 5, pp. 453–457, 2007.

- [20] A. Papparella, C. Noviello, S. Ranucci et al., "Pneumoperitoneum modifies serum and tissue CCL2–CCL5 expression in mice," *ISLS*, vol. 24, 2020.
- [21] K. A. Heel and J. C. Hall, "Peritoneal defences and peritoneum-associated lymphoid tissue," *The British Journal of Surgery*, vol. 83, no. 8, pp. 1031–1036, 1996.
- [22] F. P. K. Wu, C. Sietses, B. M. E. von Blomberg, P. A. M. van Leeuwen, S. Meijer, and M. A. Cuesta, "Systemic and peritoneal inflammatory response after laparoscopic or conventional colon resection in cancer patients," *Diseases of the Colon & Rectum*, vol. 46, no. 2, pp. 147–155, 2003.
- [23] K. L. Leung, P. B. S. Lai, R. L. K. Ho et al., "Systemic cytokine response after laparoscopic-assisted resection of rectosigmoid carcinoma," *Annals of Surgery*, vol. 231, no. 4, pp. 506–511, 2000.
- [24] E. Trondsen, O. Reiertsen, O. K. Andersen, and P. Kjaersgaard, "Laparoscopic and open cholecystectomy. A prospective, randomized study," *The European Journal of Surgery = Acta Chirurgica*, vol. 159, no. 159, pp. 217–221, 1993.
- [25] U. Berggren, T. Gordh, D. Grama, U. Haglund, J. Rastad, and D. Arvidsson, "Laparoscopic versus open cholecystectomy: hospitalization, sick leave, analgesia and trauma responses," *The British Journal of Surgery*, vol. 81, no. 81, pp. 1362–1365, 1994
- [26] G. C. Vander Velpen, S. M. Shimi, and A. Cuschieri, "Outcome after cholecystectomy for symptomatic gall stone disease and effect of surgical access: laparoscopic v open approach," *Gut*, vol. 34, no. 10, pp. 1448–1451, 1993.
- [27] B. M. Ure, T. A. Niewold, N. M. A. Bax, M. Ham, D. C. van der Zee, and G. J. Essen, "Peritoneal, systemic, and distant organ inflammatory responses are reduced by a laparoscopic approach and carbon dioxide vs. air," *Surgical Endoscopy and Other Interventional Techniques*, vol. 16, no. 5, pp. 836–842, 2002.
- [28] A. Papparella, F. Nino, S. Coppola, C. Noviello, O. Paciello, and S. Papparella, "Peritoneal morphological changes due to pneumoperitoneum: the effect of intra-abdominal pressure," *European Journal of Pediatric Surgery*, vol. 24, no. 24, pp. 322–327, 2014.
- [29] H. Poerwosusanta, N. Z. Gunadi, I. K Oktaviyanti et al., "The effect of laparoscopy on mast cell degranulation and mesothelium thickness in rats," *BMC Surg*, vol. 20, no. 1, p. 111, 2020
- [30] O. Díaz-Cambronero, G. Mazzinari, B. Flor Lorente et al., "Effect of an individualized versus standard pneumoperitoneum pressure strategy on postoperative recovery: a randomized clinical trial in laparoscopic colorectal surgery," *British Journal of Surgery*, vol. 107, no. 12, pp. 1605–1614, 2020.
- [31] J. M. Bellón, L. Manzano, L Bernardos et al., "Cytokine levels after open and laparoscopic cholecystectomy," *European Surgical Research. Europaische Chirurgische Forschung. Recherches Chirurgicales Europeennes*, vol. 29, no. 1, pp. 27–34, 1997.
- [32] M. Tylicka, E. Matuszczak, J. Kaminska et al., "Intraoperative peritoneal interleukin-6 concentration changes in relation to the high-mobility group protein B1 and heat shock protein 70 levels in children undergoing cholecystectomy," *Mediators Inflammatiom*, vol. 2020, p. 9613105, 2020.
- [33] E. P. Cummins, M. J. Strowitzki, and C. T. Taylor, "Mechanisms and consequences of oxygen and carbon dioxide sensing in mammals," *Physiological Reviews*, vol. 100, no. 1, pp. 463–488, 2020.

- [34] F. M. Duerr, D. C. Twedt, and E. Monnet, "Changes in pH of peritoneal fluid associated with carbon dioxide insufflation during laparoscopic surgery in dogs," *American Journal of Veterinary Research*, vol. 69, no. 2, pp. 298–301, 2008.
- [35] Y. T. Wong, P. C. Shah, D. H. Birkett, and D. M. Brams, "Peritoneal pH during laparoscopy is dependent on ambient gas environment helium and nitrous oxide do not cause peritoneal acidosis," *Surgical Endoscopy*, vol. 19, no. 1, pp. 60–64, 2005.
- [36] E. Hanly, A. Aurora, J. Fuentes et al., "Abdominal insufflation with CO causes peritoneal acidosis independent of systemic pH," *Journal of Gastrointestinal Surgery*, vol. 9, no. 9, pp. 1245–1252, 2005.
- [37] C. Kuntz, A. Wunsch, C. Bödeker et al., "Effect of pressure and gas type on intraabdominal, subcutaneous, and blood pH in laparoscopy," *Surgical Endoscopy*, vol. 14, no. 4, pp. 367–371, 2000
- [38] J. F. Kuebler, G. Vieten, A. Shimotakahara, M. L. Metzelder, N. K. Jesch, and B. M. Ure, "Acidification during carbon dioxide pneumoperitoneum is restricted to the gas-exposed peritoneal surface: effects of pressure, gas flow, and additional intraperitoneal fluids," *Journal of Laparoendoscopic & Advanced Surgical Techniques*, vol. 16, no. 6, pp. 654–658, 2006.
- [39] Y. T. Wong, P. C. Shah, D. H. Birkett, and D. M. Brams, "Carbon dioxide pneumoperitoneum causes severe peritoneal acidosis, unaltered by heating, humidification, or bicarbonate in a porcine model," *Surgical Endoscopy*, vol. 18, no. 10, pp. 1498–1503, 2004.
- [40] E. J. Hanly, A. A. Aurora, S. P. Shih et al., "Peritoneal acidosis mediates immunoprotection in laparoscopic surgery," *Surgery*, vol. 142, no. 3, pp. 357–364, 2007.
- [41] S. J. Neuhaus, D. I. Watson, T. Ellis, T. Lafullarde, G. G. Jamieson, and W. J. Russell, "Metabolic and immunologic consequences of laparoscopy with helium or carbon dioxide insufflation: a randomized clinical study," ANZ Journal of Surgery, vol. 71, no. 8, pp. 447–452, 2001.
- [42] M. Blobner, R. Bogdanski, E. Kochs, J. Henke, A. Findeis, and S. Jelen-Esselborn, "Effects of intraabdominanlly insufflated carbon dioxide and elevated intraabdominal pressure on splanchnic circulation," *Anesthesiology*, vol. 89, no. 2, pp. 475–482, 1998.
- [43] E. Eleftheriadis, K. Kotzampassi, D. Botsios, E. Tzartinoglou, H. Farmakis, and J. Dadoukis, "Splanchnic ischemia during laparoscopic cholecystectomy," *Surgical Endoscopy*, vol. 10, no. 3, pp. 324–326, 1996.
- [44] M. K. Schilling, C. Redaelli, L. Krähenbühl, C. Signer, and M. W. Büchler, "Splanchnic microcirculatory changes during CO2 laparoscopy," *Journal of the American College of Surgeons*, vol. 184, pp. 378–382, 1997.
- [45] K. I. Albers, F. Polat, T. Loonen et al., "Visualising improved peritoneal perfusion at lower intra-abdominal pressure by fluorescent imaging during laparoscopic surgery: a randomised controlled study," *International Journal of Surgery*, vol. 77, pp. 8–13, 2020.
- [46] N. Schukfeh, D. Huber, M. L. Metzelder et al., "Impact of capnoperitoneum on renal perfusion and urine production in infant and adolescent pigs: crystalloid versus colloid fluid resuscitation," *European Journal of Pediatric Surgery*, vol. 29, no. 6, pp. 539–544, 2019.
- [47] P. Neogi, P. Kumar, and S. Kumar, "Low-pressure pneumoperitoneum in laparoscopic cholecystectomy: a randomized controlled trial," *Surgical Laparoscopy, Endoscopy & Percutaneous Techniques*, vol. 30, no. 1, pp. 30–34, 2020.

- [48] C. R. Molinas and P. R. Koninckx, "Hypoxaemia induced by CO2 or helium pneumoperitoneum is a co-factor in adhesion formation in rabbits," *Human Reproduction*, vol. 15, no. 8, pp. 1758–1763, 2000.
- [49] C. R. Molinas, O. Mynbaev, A. Pauwels, P. Novak, and P. R. Koninckx, "Peritoneal mesothelial hypoxia during pneumoperitoneum is a cofactor in adhesion formation in a laparoscopic mouse model," *Fertility and Sterility*, vol. 76, no. 3, pp. 560–567, 2001.
- [50] N. Bourdel, S. Matsuzaki, J.-E. Bazin, J.-L. Pouly, G. Mage, and M. Canis, "Peritoneal tissue-oxygen tension during a carbon dioxide pneumoperitoneum in a mouse laparoscopic model with controlled respiratory support," *Human Reproduction*, vol. 22, no. 4, pp. 1149–1155, 2007.
- [51] C. N. Gutt and T. C. Schmandra, "Portal venous flow during CO2 pneumoperitoneum in the rat," *Surgical Endoscopy*, vol. 13, no. 9, pp. 902–905, 1999.
- [52] L. Gianotti, L. Nespoli, S. Rocchetti, A. Vignali, A. Nespoli, and M. Braga, "Gut oxygenation and oxidative damage during and after laparoscopic and open left-sided colon resection: a prospective, randomized, controlled clinical trial," *Surgical Endoscopy*, vol. 25, no. 6, pp. 1835–1843, 2011.
- [53] O. Taskin, A. Buhur, M. Birincioglu et al., "The effects of duration of CO2 insufflation and irrigation on peritoneal microcirculation assessed by free radical scavengers and total glutathion levels during operative laparoscopy," *The Journal of the American Association of Gynecologic Laparoscopists*, vol. 5, no. 2, pp. 129–133, 1998.
- [54] C. Polat, S. Yilmaz, M. Serteser, T. Koken, A. Kahraman, and O. N. Dilek, "The effect of different intraabdominal pressures on lipid peroxidation and protein oxidation status during laparoscopic cholecystectomy," *Surgical Endoscopy*, vol. 17, no. 11, pp. 1719–1722, 2003.
- [55] C. E. Murry, R. B. Jennings, and K. A. Reimer, "Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium," *Circulation*, vol. 74, no. 5, pp. 1124–1136, 1986.
- [56] S. Aksöyek, I. Cinel, D Avlan et al., "Intestinal ischemic preconditioning protects the intestine and reduces bacterial translocation," *Shock (Augusta, Ga.)*, vol. 18, no. 5, pp. 476–480, 2002.
- [57] D. A. Sahin, B. Haliloglu, F. K. Ahin et al., "Stepwise rising CO2Insufflation as an ischemic preconditioning method," *Journal of Laparoendoscopic & Advanced Surgical Techniques*, vol. 17, no. 6, pp. 723–730, 2007.
- [58] T. G. Veres, L. Petrovics, K Sárvári et al., "The effect of laparoscopic pre- and postconditioning on pneumoperitoneum induced injury of the peritoneum," *Clinical Hemorheology and Microcirculation*, vol. 73, no. 73, pp. 565–577, 2019.
- [59] J. Kolbenschlag, A. Sogorski, C. Timmermann et al., "Ten minutes of ischemia is superior to shorter intervals for the remote ischemic conditioning of human microcirculation," *Clinical Hemorheology and Microcirculation*, vol. 66, no. 3, pp. 239–248, 2017.
- [60] T. G. Veres, I. Takács, T. Nagy et al., "Pneumoperitoneum induced ischemia-reperfusion injury of the peritoneum preconditioning may reduce the negative side-effects caused by carbon-dioxide pneumoperitoneum - pilot study," *Clinical Hemorheology and Microcirculation*, vol. 69, no. 4, pp. 481– 488, 2018.
- [61] N. Chegini, "Peritoneal molecular environment adhesion formation and clinical implication," *Frontiers in Bioscience*, vol. 7, no. 5, pp. e91–115, 2002.

- [62] S. Yung and T. M. Chan, "Biology of the peritoneum in normal homeostasis and after surgical trauma," *Colorectal Disease*, vol. 9, no. 2, pp. 9–13, 2007.
- [63] J. Volz, S. Köster, Z. Spacek, and N. Paweletz, "Characteristic alterations of the peritoneum after carbon dioxide pneumoperitoneum," *Surgical Endoscopy*, vol. 13, no. 6, pp. 611–614, 1999.
- [64] T. Suematsu, Y. Hirabayashi, N. Shiraishi, Y. Adachi, H. Kitamura, and S. Kitano, "Morphology of the murine peritoneum after pneumoperitoneum vs laparotomy," Surgical Endoscopy, vol. 15, no. 9, pp. 954–958, 2001.
- [65] J. Du, P.-w. Yu, and B. Tang, "Application of stereology to study the effects of pneumoperitoneum on peritoneum," *Surgical Endoscopy*, vol. 25, no. 2, pp. 619–627, 2011.
- [66] S. S. Davis, D. J. Mikami, M. Newlin et al., "Heating and humidifying of carbon dioxide during pneumoperitoneum is not indicated," *Surgical Endoscopy*, vol. 20, no. 1, pp. 153–158, 2006.
- [67] M. Erikoglu, S. Yol, M. C. Avunduk, E. Erdemli, and A. Can, "Electron-microscopic alterations of the peritoneum after both cold and heated carbon dioxide pneumoperitoneum," *Journal of Surgical Research*, vol. 125, no. 1, pp. 73–77, 2005.
- [68] Y. Peng, M. Zheng, Q. Ye, X. Chen, B. Yu, and B. Liu, "Heated and humidified CO2 prevents hypothermia, peritoneal injury, and intra-abdominal adhesions during prolonged laparoscopic insufflations," *Journal of Surgical Research*, vol. 151, no. 1, pp. 40–47, 2009.
- [69] E. J. Hazebroek, M. A. Schreve, P. Visser, R. W. F. De Bruin, R. L. Marquet, and H. J. Bonjer, "Impact of temperature and humidity of carbon dioxide pneumoperitoneum on body temperature and peritoneal morphology," *Journal of Lapa*roendoscopic & Advanced Surgical Techniques, vol. 12, no. 5, pp. 355–364, 2002.
- [70] S. Sampurno, T. J. Chittleborough, S. Carpinteri et al., "Modes of carbon dioxide delivery during laparoscopy generate distinct differences in peritoneal damage and hypoxia in a porcine model," *Surgical Endoscopy*, vol. 34, no. 10, pp. 4395–4402, 2020.
- [71] Y. Liu and Q. X. Hou, "Effect of carbon dioxide pneumoperitoneum during laparoscopic surgery on morphology of peritoneum," *Zhonghua Yi Xue Za Zhi*, vol. 86, pp. 164–166, 2006
- [72] O. R. Tarhan, I. Barut, C. Ozogul, S. Bozkurt, B. Baykara, and M. Bulbul, "Structural deteriorations of the human peritoneum during laparoscopic cholecystectomy. A transmission electron microscopic study," *Surgical Endoscopy*, vol. 27, no. 8, pp. 2744–2750, 2013.
- [73] P. H. A. Morais, R. F. A. Silva, T. D. S. Ribeiro et al., "Does CO2 pneumoperitoneum in laparoscopy interfere with collagen deposition in abdominal surgical wounds?" Acta Cirúrgica Brasileira, vol. 35, Article ID e202000605, 2020.
- [74] W. Arung, M. Meurisse, and O. Detry, "Pathophysiology and prevention of postoperative peritoneal adhesions," World Journal of Gastroenterology, vol. 17, no. 41, pp. 4545–4553, 2011.
- [75] O. R. Tarhan, I. Barut, Y. Akdeniz, R. Sutcu, C. Cerci, and M. Bulbul, "Fibrinolytic responses of human peritoneal fluid in laparoscopic cholecystectomy: a prospective clinical study," *Surgical Endoscopy*, vol. 22, no. 4, pp. 1008–1013, 2008.
- [76] C. A. Jacobi, A. Sterzel, C. Braumann et al., "The impact of conventional and laparoscopic colon resection (CO2 or helium) on intraperitoneal adhesion formation in a rat peritonitis model," *Surgical Endoscopy*, vol. 15, no. 4, pp. 380–386, 2001.

- [77] M. M. Binda, C. R. Molinas, K. Mailova, and P. R. Koninckx, "Effect of temperature upon adhesion formation in a laparoscopic mouse model," *Human Reproduction*, vol. 19, no. 11, pp. 2626–2632, 2004.
- [78] P. R. Koninckx, R. Corona, D. Timmerman, J. Verguts, and L. Adamyan, "Peritoneal full-conditioning reduces postoperative adhesions and pain: a randomised controlled trial in deep endometriosis surgery," *Journal of Ovarian Research*, vol. 6, no. 1, p. 90, 2013.
- [79] R. Jiang, Y. Sun, H. Wang, M. Liang, and X. Xie, "Effect of different carbon dioxide (CO2) insufflation for laparoscopic colorectal surgery in elderly patients: a randomized controlled trial," *Medicine (Baltimore)*, vol. 98, Article ID e17520, 2019.
- [80] W. J. A. Brokelman, L. Holmdahl, M. Bergström, P. Falk, J. H. G. Klinkonbijl, and M. M. P. J. Reijnen, "Peritoneal transforming growth factor beta-1 expression during laparoscopic surgery: a clinical trial," *Surgical Endoscopy*, vol. 21, no. 9, pp. 1537–1541, 2007.
- [81] M. M. A. Lensvelt, M. L. Ivarsson, W. J. Brokelman, P. Falk, and M. M. Reijnen, "Decreased peritoneal expression of active transforming growth factor β1 during laparoscopic chole-cystectomy with heated carbon dioxide," *Archives of Surgery*, vol. 145, no. 10, pp. 968–972, 2010.
- [82] S. Matsuzaki, R. Botchorishvili, K. Jardon, E. Maleysson, M. Canis, and G. Mage, "Impact of intraperitoneal pressure and duration of surgery on levels of tissue plasminogen activator and plasminogen activator inhibitor-1 mRNA in peritoneal tissues during laparoscopic surgery," *Human Re*production, vol. 26, no. 5, pp. 1073–1081, 2011.