

Recurrent Spontaneous Abortion (RSA) and Maternal *KIR* Genes: A Comprehensive Meta-Analysis

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ABSTRACT

Natural killer cells (NKs) are the most important cells in the fetomaternal immune tolerance induced through interaction of maternal killer-cell immunoglobulin-like receptors (KIR) and fetal human leucocyte antigens (HLA). Hence, we intend to perform a meta-analysis on the role of maternal *KIR* genes diversity in recurrent spontaneous abortion (RSA). The present paper is a meta-analysis of previous genetic association studies and our previous original study. The results showed that *KIR3DL1* was a significantly protecting factor for RSA ($p=0.044$; OR=0.833 [0.698-0.995]; fixed effect model). *KIR2DS2* ($p=0.034$; OR=1.195 [1.013-1.408]; fixed effect model) and *KIR2DS3* ($p=0.013$; OR=1.246 [1.047-1.483]; fixed effect model) were significantly risk factors for RSA. For *KIR2DS1* there was a high heterogeneity and publication bias. Briefly, the inhibitory gene *KIR3DL1* was a protecting factor, and the activating genes *KIR2DS2* and *KIR2DS3* were risk factors for RSA. However, the effect sizes were not suitable. We suggest further studies on different causes of pregnancy loss, to find the role of *KIR2DS1*.

Keywords: recurrent spontaneous abortion, killer-cell immunoglobulin-like receptor, human leukocyte antigen, meta-analysis

INTRODUCTION

Rationale

Recurrent spontaneous abortion (RSA) and pregnancy loss have different pathogeneses, consisting of genetic and chromosomal abnormalities (Hume & Chasen, 2015), environmental toxicities and oxidative stress (Gupta *et al.*, 2007), infectious agents (Ambühl *et al.*, 2016), hormonal causes, etc. Among them, immunological causes and their involving molecules are still controversial and unknown topics. The immune system is a fascinating system, one that does not normally reject the semi-allograft fetus. The immune system has two roles in implantation and pregnancy; preventing the formation of abnormal embryos, and protecting the fetomaternal interaction by releasing angiogenic factors, cytokines and adhesive molecules. The fascinating point is how a system can have two mutually exclusive features; protection and rejection. Indeed, the immune system is the bodyguard of the body through self- and non-self recognition. However, pregnancy is a semi-allograft transplantation. So the question is what the immune system does in this situation; rejection or protection (Akbari *et al.*, 2018; Würfel, 2016)?!

Immune tolerance is the best answer for the above question (Akbari *et al.*, 2018; Würfel, 2016). Natural killer cells (NKs), which name is self-explanatory, are one of the most important lymphocytes in immune tolerance. They identify self-cells through their killer-cell immunoglobulin-like receptors (KIRs) expressed on their surface. The KIRs interact with their ligands, the human leukocyte antigens (HLAs) - the identification cards of self-cells. These interactions usually result in immune tolerance under normal conditions. Both *KIR* and *HLA* genes in human genome have loci (not locus), inherited as haplotypes. In addition, each gene in their loci is polymorphic. Thus, interaction of different KIR molecules with different HLA molecules results in different outcomes consisting of inhibitory and activating responses. *KIR* gene cluster is located on chromosome 19. This cluster has two types of genes, including 8 inhibitory and 6 activating genes, and 2 pseudogenes. Some of these genes exist in all individuals, like the *KIR2DL4*. From the viewpoint of medical anthropology, different people from different ethnicities have different KIR-HLA interactions (Alecsandru *et al.*, 2014; Ashouri *et al.*, 2016; Middleton *et al.*, 2008; Norman *et al.*, 2016; Solgi *et al.*, 2011).

HLA has two classes, I and II, and the class I can be further divided into classical and non-classical HLA. *KIR2DL4* is an inhibitory KIR binding to the trophoblast HLA-G, which is a non-classical HLA. The combination *KIR2DL4*+HLA-G triggers the immune tolerance. Both *KIR2DL4* and *HLA-G* are polymorphic genes. Therefore, anthropological variations can contribute to implantation success and pregnancy maintenance. For example, HLA-G*01:03:01 is a risk factor for implantation failure; because its connection with *KIR2DL4* is not sufficient to trigger inhibitory signals (Nardi *et al.*, 2012).

NKs may have the CD16 marker, which is the weapon of antibody-dependent cell-mediated cytotoxicity (ADCC). Usually CD56^{dim} NKs are CD16⁺. So CD16⁺CD56^{dim} NKs are known as cytotoxic NKs, whereas CD16⁻CD56^{bright} NKs are known as immune-regulatory NKs (Ghafourian *et al.*, 2015). About 90% of uterine NKs (UNKs) are immune-regulatory. In conclusion, UNKs are not usually cytotoxic for the embryo (Ghafourian *et al.*, 2015; Sacks, 2015).

Objectives

As we mentioned above, KIR and HLA have different genes and interactions. KIR has 8 inhibitory (*2DL1*, *2DL2*, *2DL3*, *2DL4*, *2DL5*, *3DL1*, *3DL2* and *3DL3*) and 6 activating

genes (*2DS1*, *2DS2*, *2DS3*, *2DS4*, *2DS5* and *3DS1*). Since the involving NKs in implantation of embryo are maternal, we intend to perform a meta-analysis on the role of maternal *KIR* genes diversity in RSA. Previously, Perez *et al.* (2017) carried out a meta-analysis on different genes, including the *KIR*. Nevertheless, their studies were few and therefore our study can serve as an update for that meta-analysis.

MATERIALS AND METHODS

Study selection

For the present meta-analysis, we searched in scientific databases such as Web of Science, PubMed, Scopus, Google Scholar, etc. Our keywords were searched only among the titles. After exclusion of duplicates, all the eligible studies were used for qualitative systematic review.

Eligibility criteria

Among the studies imported for qualitative systematic review, only the studies with available and enough numerical data were imported for the quantitative meta-analysis. Our original paper on this topic was manually added (Table 1) (Akbari *et al.*, 2018). Performing *KIR* typing was the most important criterion.

Statistical analysis

To perform the present meta-analysis, we used the comprehensive meta-analysis version 2 software (Biostat, US). The analyses were carried out through a p value and individual sample size using fixed-effect and random-effect models. Since the p values were calculated using Yate's correction (or Fisher's exact test if necessary), the odds ratios (OR) (effect sizes) achieved from these p values were underestimated. This statistical protocol has been previously published (Anbari & Ahmadi, 2017).

Heterogeneity and publication bias

We used the I^2 scale and $I^2 < 50$ was considered as homogeneity. In the cases of heterogeneity, we used the random-effect model. In order to find publication bias, we used funnel plots. If a study were to be found outside the funnel, it meant that its effect size was outside the expected 95% confidence interval (CI). In other words, its difference with other studies is statistically significant at $p=0.05$. Hence, a publication bias does not have necessarily a negative connotation. In the present study, a funnel plot p value < 0.05 means that the mentioned individual study is outside the funnel of 95% CI.

Additional analyses

In order to cluster the studies for meta-analysis, we designed a dendrogram using the STATA14 software (StataCorp LLC, US). This cluster analysis involved the complete linkage of binary variables (Table 2, Figure 1).

RESULTS

Eligible studies

Table 1 depicts the findings from the selected studies, in addition to our original case-control study, this table includes 11 studies. The p values were analyzed through Yate's correction (or Fisher's exact test when necessary). Positive effect directions show each gene as a risk factor and negative effect directions show each gene as a protecting factor. Our cluster analysis showed that the study by Dambaeva *et al.* (2016) had a different design in comparison to other studies (Figure 1). Hence, it was excluded from the meta-analysis. At the end, 10 studies remained.

Meta-analysis

The role of *KIR2DL1* in RSA was not statistically significant ($p=0.051$; OR=0.849; fixed). Faridi *et al.* (2009) showed a significantly more protective effect of this gene in comparison to other studies (funnel plot p value < 0.05) (Figures 2 and 3).

The role of *KIR2DL2* in RSA was not statistically significant ($p=0.325$; OR=1.091; fixed). Hong *et al.* (2008) showed a significantly higher risk of this gene's effect in comparison to other studies (funnel plot p value < 0.05) (Figures 4 and 5). The role of *KIR2DL3* in RSA was not statistically significant ($p=0.448$; OR=1.062; fixed). No publication bias was found based on the funnel plot (Figures 6 and 7). The role of *KIR2DL5* in RSA was not statistically significant ($p=0.767$; OR=0.960; random). Hiby *et al.* (2008) showed a significantly more protective effect of this gene in comparison to other studies (funnel plot p value < 0.05) (Figures 8 and 9).

The role of *KIR3DL1* in RSA was statistically significant ($p=0.044^*$; OR=0.833; fixed). Faridi *et al.* (2009) showed a significantly more protective effect of this gene in comparison to other studies ($p < 0.05$; based on funnel plot) (Figures 10 and 11). The role of *KIR2DS1* in RSA was not statistically significant ($p=0.726$; OR=1.056; random). Inconclusive publication bias was found for this analysis based on the funnel plot (Figures 12 and 13). The role of *KIR2DS2* in RSA was statistically significant ($p=0.034^*$; OR=1.195; fixed). Faridi *et al.* (2009) study showed significantly more risk effect of this gene in comparison to other studies (funnel plot value < 0.05) (Figures 14 and 15). The role of *KIR2DS3* in RSA was statistically significant ($p=0.013^*$; OR=1.246; fixed). Faridi *et al.* (2009) showed significantly more risk effect of this gene in comparison to other studies (funnel plot p value < 0.05) (Figures 16 and 17).

The role of *KIR2DS4* in RSA was not statistically significant ($p=0.094$; OR=0.762; fixed). Faridi *et al.* (2009) showed significantly more protective effect of this gene in comparison to other studies (funnel plot p value < 0.05) (Figures 18 and 19). The role of *KIR2DS5* in RSA was not statistically significant ($p=0.642$; OR=1.042; fixed). Hiby *et al.* (2008) showed a significantly more protective effect of this gene in comparison to other studies (funnel plot p value < 0.05) (Figures 20 and 21). The role of *KIR3DS1* in RSA was not statistically significant ($p=0.851$; OR=1.037; random). Hiby *et al.* (2008) and Faridi *et al.* (2009) showed significantly more protective and risk effect of this gene in comparison to other studies, respectively (funnel plot p value < 0.05) (Figures 22 and 23).

DISCUSSION

Summary of evidence

NKs are lymphocytes that participate in the innate immune system. They have 2 subtypes: CD16⁺CD56^{dim} and CD16⁺CD56^{bright} that are called as cytotoxic and immune-regulatory NKs, respectively. In the implantation site, the NKs are mainly CD56^{bright}. Hence, the immune system has a positive and protecting role in implantation and early pregnancy. Embryo implantation and pregnancy are a type of transplantation called semi-allograft. Thus, we need immune tolerance to have a successful pregnancy. The NKs play their roles with their KIRs interacting with the HLAs expressed on trophoblasts (Würfel, 2016). Because of the important roles of NKs in the implantation process, this meta-analysis aimed to investigate the role of maternal *KIR* genes diversity in RSA.

Among the investigated genes, only the results of *3DL1*, *2DS2* and *2DS3* were statistically significant

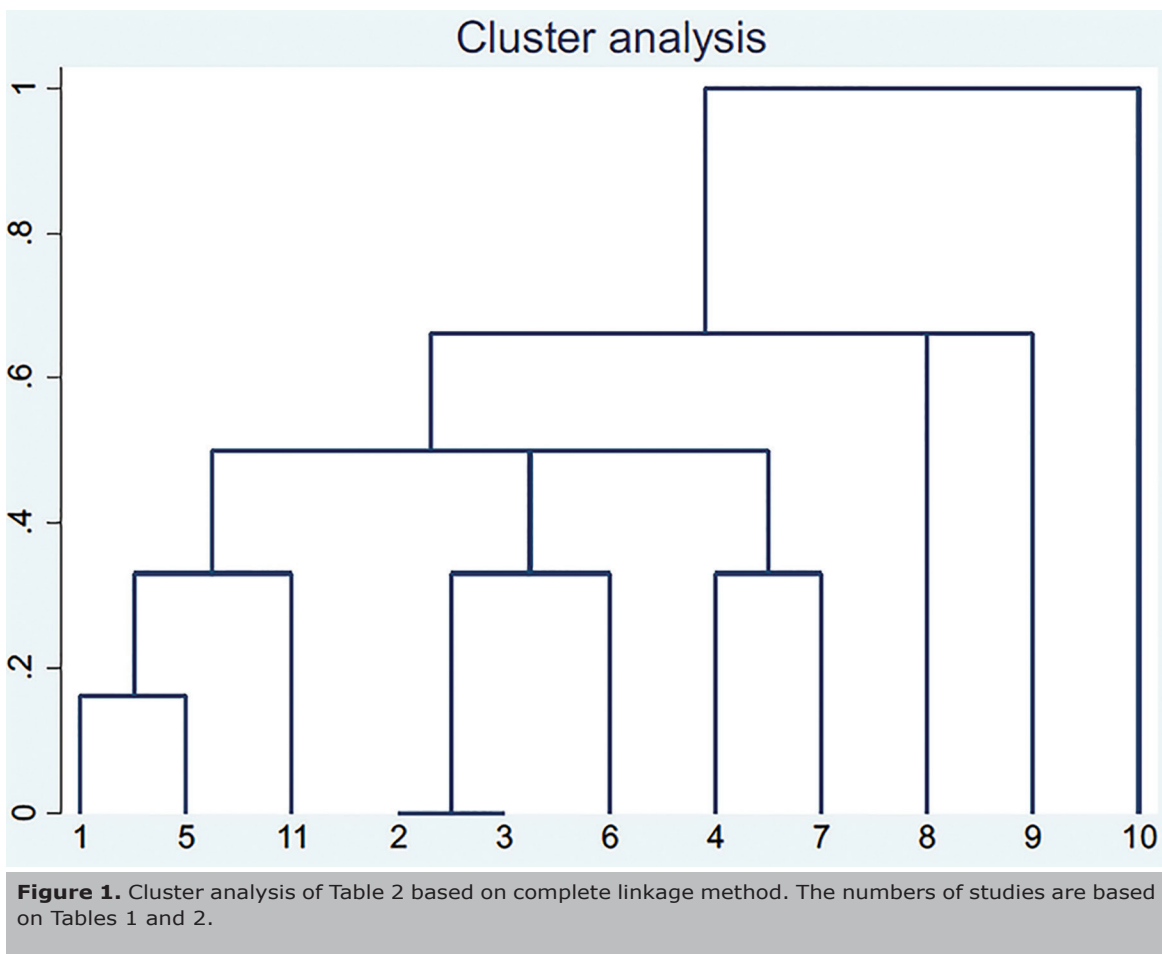
Table 1. Data summary of the found articles.

Study	Witt et al., 2004	Wang et al., 2007	Hong et al., 2008	Hiby et al., 2008	Vargas et al., 2009	Faridi et al., 2009	Khosravifar et al., 2011	Ozturk et al., 2012	Djulejic et al., 2015	Dambaeva et al., 2016	Our original study
Gene	RSA N=52	RSA N=73	RSA N=16	RSA N=95	RSA N=68	RSA N=205	RSA N=100	RSA N=40	RSA N=25	RSA N=139	RSA N=100
	Control N=55	Control N=68	Control N=41	Control N=269	Control N=68	Control N=224	Control N=100	Control N=90	Control N=122	Control N=195	Control N=100
2DL1	52	73	8	92	63	141	97	40	24	135	93
<i>p</i> value (ED) a	1 (FET) b	1 (FET)	0.841 (-)	0.769 (FET) (-)	0.999 (FET) (-)	0.0001 (-)	0.720 (+)	1 (FET)	1 (FET)	1 (FET)	0.764 (-)
2DL2	29	22	16	45	43	110	52	26	17	69	96
<i>p</i> value (ED)	0.211 (+)	0.777 (-)	0.002 (+)	0.361 (-)	0.383 (+)	0.446 (+)	0.475 (-)	0.632 (+)	0.537 (+)	1	
2DL3	47	72	6	88	58	169	87	37	24	124	172
<i>p</i> value (ED)	0.631 (+)	1 (FET)	0.887 (-)	0.806 (+)	1 (FET)	0.887 (-)	0.841 (+)	0.207 (+)	0.469 (FET) (+)	0.920 (+)	
2DL4											
2DL5	16	35	5	36	37	127	35	32	4	79	58
<i>p</i> value (ED)	0.680 (-)	0.521 (+)	1 (FET)	0.005 (-)	0.610 (+)	0.238 (+)	0.084 (-)	0.072 (+)	0.032 (-)	0.537 (+)	0.887 (-)
3DL1	50	73		88	64	120		36	24	125	93
<i>p</i> value (ED)	0.162 (FET) (+)	1 (FET)		0.502 (-)	0.999 (FET) (+)	0.0001 (-)		1 (FET)	1 (FET)	0.131 (-)	0.764 (-)
3DL2											
3DL3											
2DS1	21	44	1	24	32	92	48	21	8	63	73
<i>p</i> value (ED)	0.740 (-)	0.035 (+)	1 (FET)	0.001 (-)	0.386 (+)	0.283 (+)	0.084 (-)	0.005 (+)	0.228 (-)	0.182 (+)	0.254 (+)
2DS2	27	22	1	46	45	104	50	26	14	69	59
<i>p</i> value (ED)	0.777 (+)	0.764 (+)	1 (FET)	0.624 (-)	0.377 (+)	0.001 (+)	0.319 (-)	0.063 (+)	0.806 (+)	1	0.565 (+)
2DS3	16	25	2	22	22	94		17	11	44	38
<i>p</i> value (ED)	0.824 (+)	0.622 (+)	0.613 (+)	0.680 (-)	0.862 (-)	0.0007 (+)		0.350 (+)	0.399 (+)	0.577 (+)	0.654 (+)
2DS4	18	72	8	90	62	109		36	25	130	95
<i>p</i> value (ED)	0.862 (-)	0.352 (FET) (+)	0.777 (-)	1 (FET)	0.740 (-)	0.0001 (-)		1 (FET)	0.588 (FET) (+)	0.777 (-)	1
2DS5	10	38	4	23	30	122		22	6	53	35
<i>p</i> value (ED)	0.171 (-)	0.139 (+)	0.722 (FET) (+)	0.021 (-)	0.074 (+)	0.337 (+)		0.129 (+)	0.698 (-)	0.764 (+)	1
3DS1	17	38		24	34	162		16	7	62	41
<i>p</i> value (ED)	0.590 (-)	0.761 (+)		0.001 (-)	0.082 (+)	0.0001 (+)		0.920 (-)	0.488 (-)	0.409 (+)	1
2DP1											
3DP1											
Study design	Case-control	Case-control	Case-control	Case-control	Case-control	Case-control	Case-control	Case-control	Case-control	Cohort for KIR2DS1	Case-control
Genotyping method	PCR-SSP	PCR-SSP	PCR-SSP	PCR-SSP	PCR-SSO	PCR-SSP	PCR-SSP	PCR-SSO	PCR-SSP	PCR-SSO	PCR-SSP
RSA definition	3 spontaneous abortion	3 spontaneous abortion	3 spontaneous abortion	3 spontaneous abortion	3 spontaneous abortion	3 spontaneous abortion	3 spontaneous abortion	A history of miscarriage	Any fertility problem	2 spontaneous abortion	3 spontaneous abortion
Control definition	2 history of normal delivery	2 history of normal delivery	2 history of normal delivery	Any primiparous woman	2 history of normal delivery	2 history of normal delivery	1 history of normal delivery	2 history of normal delivery	Not mentioned	Not mentioned	2 history of normal delivery
Place	Brazil	China	China	London	Brazil	India	Iranian	Mediterranean	Albania	America	Iran
Ethnicity	Caucasian	Chinese	Chinese	Caucasian	Caucasian	Indian	Caucasian	Caucasian	Caucasian	Caucasian	Caucasian
Study number in dendrogram	1	2	3	4	5	6	7	8	9	10	11

a) ED stands for effect direction; the positive ones show risk factors and the negative ones show protecting factors. b) FET stands for Fisher's exact test.

Table 2. Dissimilarity matrix of studies' characteristics based on the below of Table 1

		Witt et al., 2004	Wang et al., 2007	Hong et al., 2008	Hiby et al., 2008	Vargas et al., 2009	Faridi et al., 2009	Khosravifar et al., 2011	Ozturk et al., 2012	Djulejic et al., 2015	Dambaeva et al. 2016	Our study
1	Witt et al., 2004	0	0.33	0.33	0.33	0.16	0.33	0.33	0.50	0.50	0.83	0.16
2	Wang et al., 2007	0.33	0	0	0.50	0.50	0.33	0.50	0.66	0.66	1	0.33
3	Hong et al., 2008	0.33	0	0	0.50	0.50	0.33	0.50	0.66	0.66	0.83	0.33
4	Hiby et al., 2008	0.33	0.50	0.50	0	0.50	0.50	0.33	0.66	0.50	0.83	0.33
5	Vargas et al., 2009	0.16	0.50	0.50	0.50	0	0.50	0.50	0.33	0.66	0.66	0.33
6	Faridi et al., 2009	0.33	0.33	0.33	0.50	0.50	0	0.50	0.66	0.66	1	0.33
7	Khosravifar et al., 2011	0.33	0.50	0.50	0.33	0.50	0.50	0	0.66	0.50	0.83	0.33
8	Ozturk et al., 2012	0.50	0.66	0.66	0.66	0.33	0.66	0.66	0	0.66	0.66	0.50
9	Djulejic et al., 2015	0.50	0.66	0.66	0.50	0.66	0.66	0.50	0.66	0	0.66	0.50
10	Dambaeva et al., 2016	0.83	1	0.83	0.83	0.66	1	0.83	0.66	0.66	0	0.83
11	Our original study	0.16	0.33	0.33	0.33	0.33	0.33	0.33	0.50	0.50	0.83	0



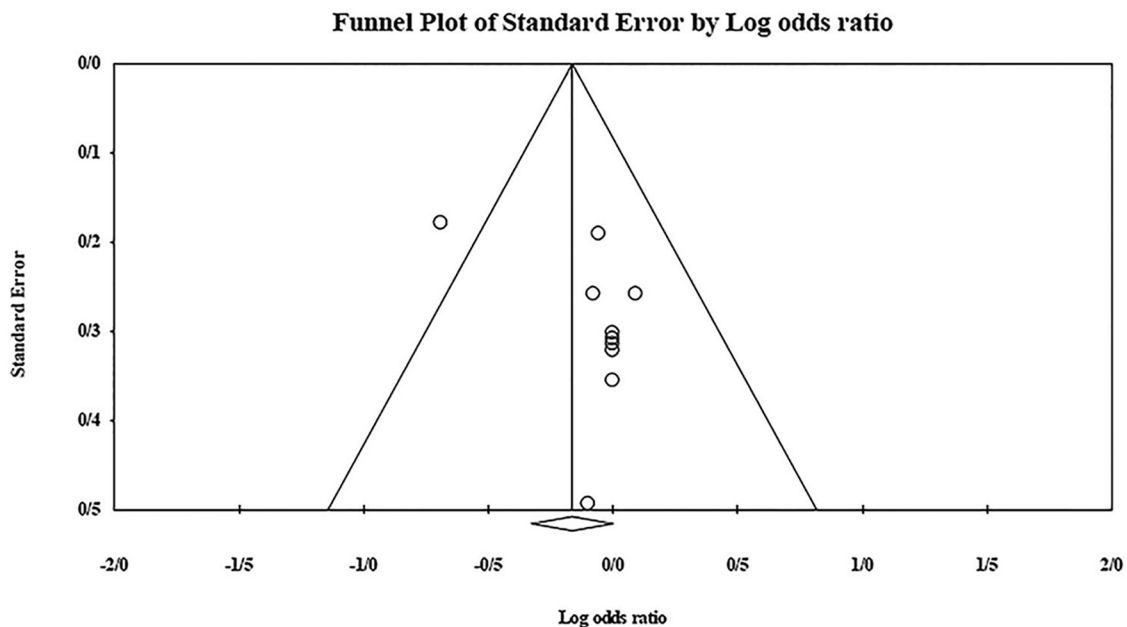


Figure 2. KIR2DL1 Funnel plot showing a significant bias for Faridi *et al.* (2009).

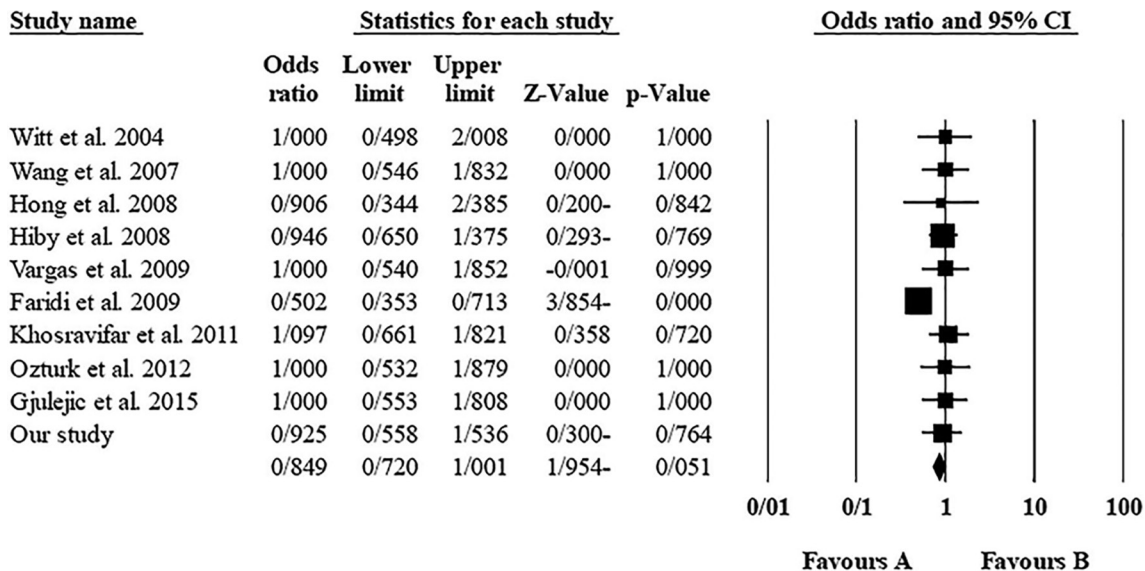


Figure 3. Forest plot of KIR2DL1 (fixed). Favours A shows protecting effect and favours B shows harmful effect (in all figures).

with protective, risk and risk effect impacts, respectively (Table 3). If we adjust multiple test correction for these findings, none of them would remain significant. It shows that there is no specific *KIR* gene predicting RSA. The funnel plot analyses showed that Faridi *et al.* (2009), in India, had more publication bias in comparison to the others. In our original study we showed that maternal KIR2DS1 in combination with

paternal HLA-C2 can be a risk factor (Akbari *et al.*, 2018).

Literature review

This concern in reproductive immunology dates back to 2004. Witt *et al.* (2004) found no significant association of maternal *KIR* genes with the risk of RSA in a Brazilian population. Lack of paternal or fetal

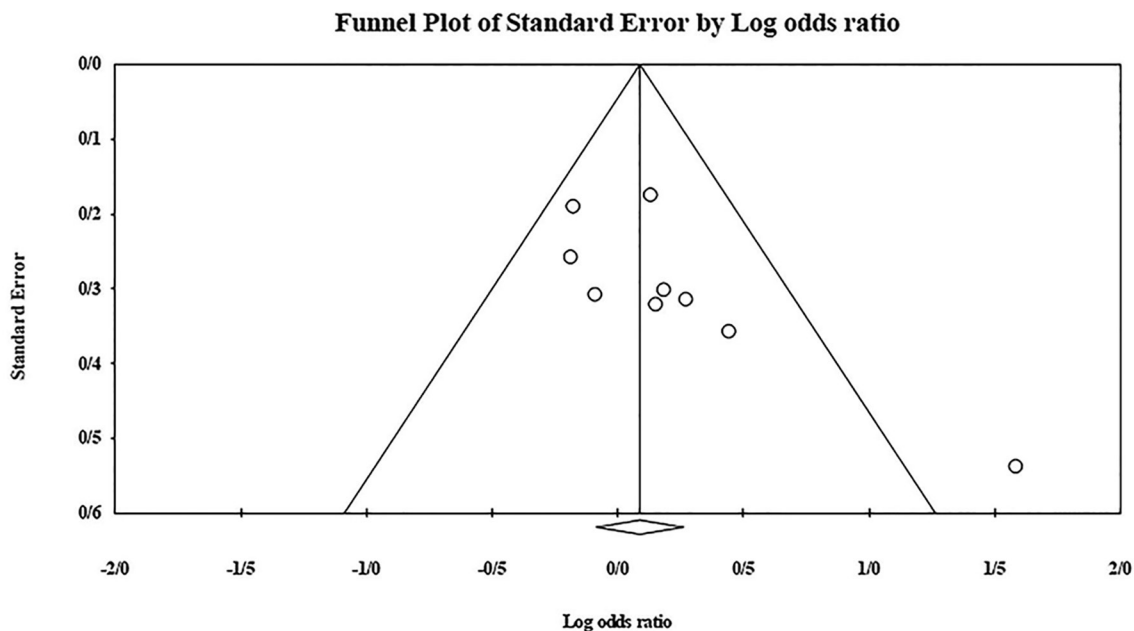


Figure 4. KIR2DL2 Funnel plot showing a significant bias for Hong *et al.* (2008).

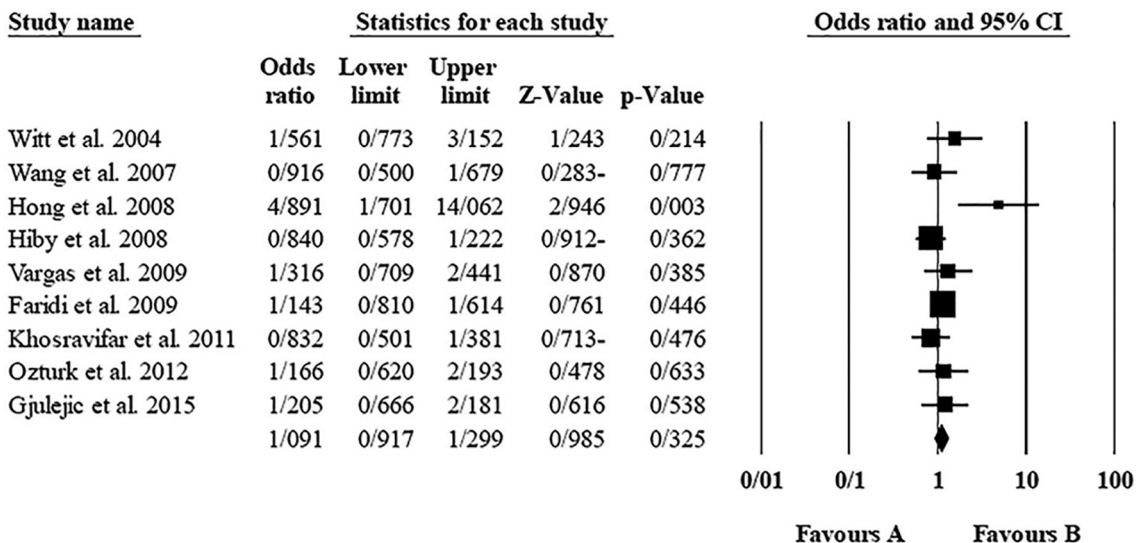


Figure 5. Forest plot of KIR2DL2 (fixed).

evaluation of *HLA-C* was their study limitation. Yamada *et al.* (2004) evaluated different immune markers such as CD94, CD158 (the very KIR) and CD161 through flow cytometry in 20 RSA women and 15 fertile controls. They found a lower level of CD158a (the very KIR2DL1) in the RSA group. Their low sample size was a limitation in their study (Yamada *et al.*, 2004). Because of their quantitative approach and different aims

and protocols, we excluded that study from our meta-analysis. Varla-Leftherioti *et al.* (2005) evaluated only *KIR2DL1*, *2DL2* and *2DL3* among the *KIR* genes in a small sample size. Wang *et al.* (2007) found a risk association for *KIR2DS1* in a Chinese population. They evaluated *HLA-C* in couples, similar to our original experience. Conversely, our original study and some studies before, e.g. Hiby *et al.* (2008), found a

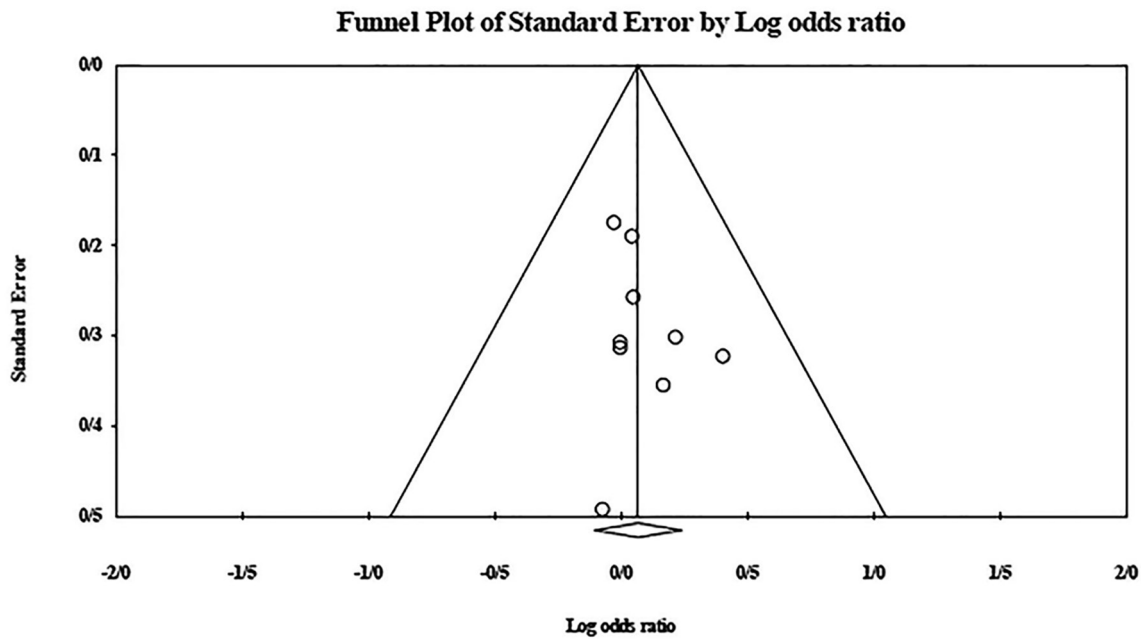


Figure 6. Funnel plot of KIR2DL3 shows no publication bias.

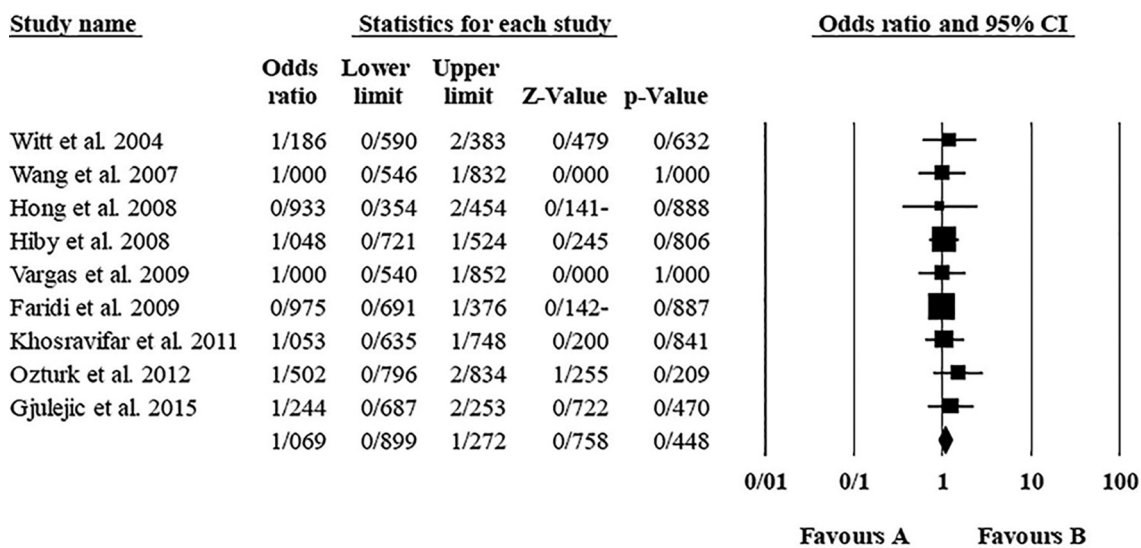


Figure 7. KIR2DL3 Forest plot (fixed).

strongly protecting association for *KIR2DS1* in a Caucasian population. However, since their control group criteria was to be a first-birth woman, this might be the reason of their publication bias. Vargas *et al.* (2009) found a risk association for the number of maternal activating *KIR* genes. Faridi *et al.* (2009) found that RSA was more associated with activating, and

more protected with inhibitory *KIR* genes. Nowak *et al.* (2009) found that RSA could be associated with *KIR* genotypes. Conversely, other studies found that RSA was more frequent in patients with genotypes bearing 6 inhibitory genes. Because we did not have access to the frequencies of *KIR* genes, we excluded this study from our meta-analysis. Nowak *et al.* (2011) found

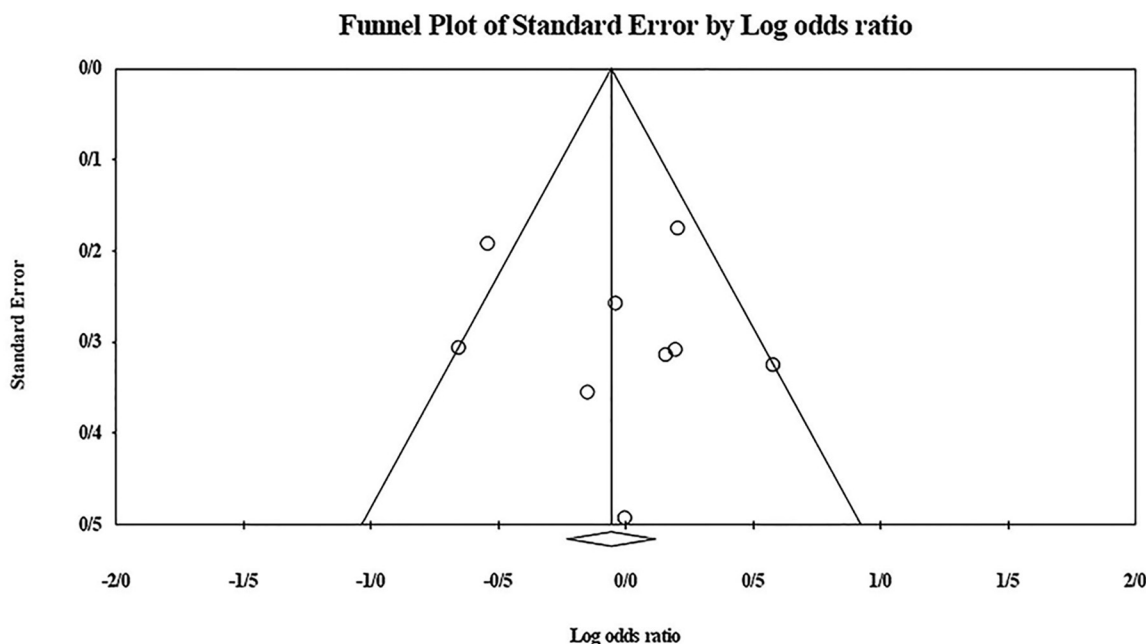


Figure 8. KIR2DL5 Forest plot showing a significant bias for Hiby et al. (2008) study.

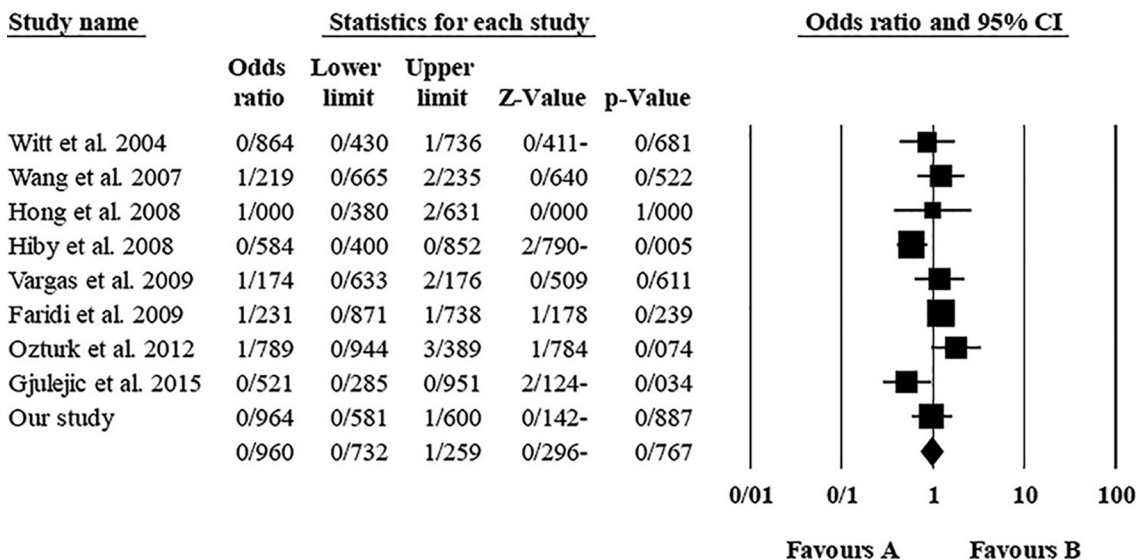


Figure 9. KIR2DL5 Forest plot (random).

that female heterozygosity for *HLA-C* in combination with AA KIR genotype could be a protecting factor for RSA. Khosravifar *et al.* (2011) investigated the role of maternal *KIR* and parental *HLA-C* in an Iranian population. They found that RSA was associated with maternal *HLA-C2*. Ozturk *et al.* (2012) found a protecting role for the KIR AA genotype. A small sample size and

one miscarriage episode in the RSA group were the negative points of their study. Alecsandru *et al.* (2014) found that maternal AA genotype was a risk factor affecting the success of double embryo transformation. Djulejic *et al.* (2015) evaluated the role of *KIR* genes on women with any fertility problem. Hence, we excluded it from our meta-analysis. Nowak *et al.* (2016)

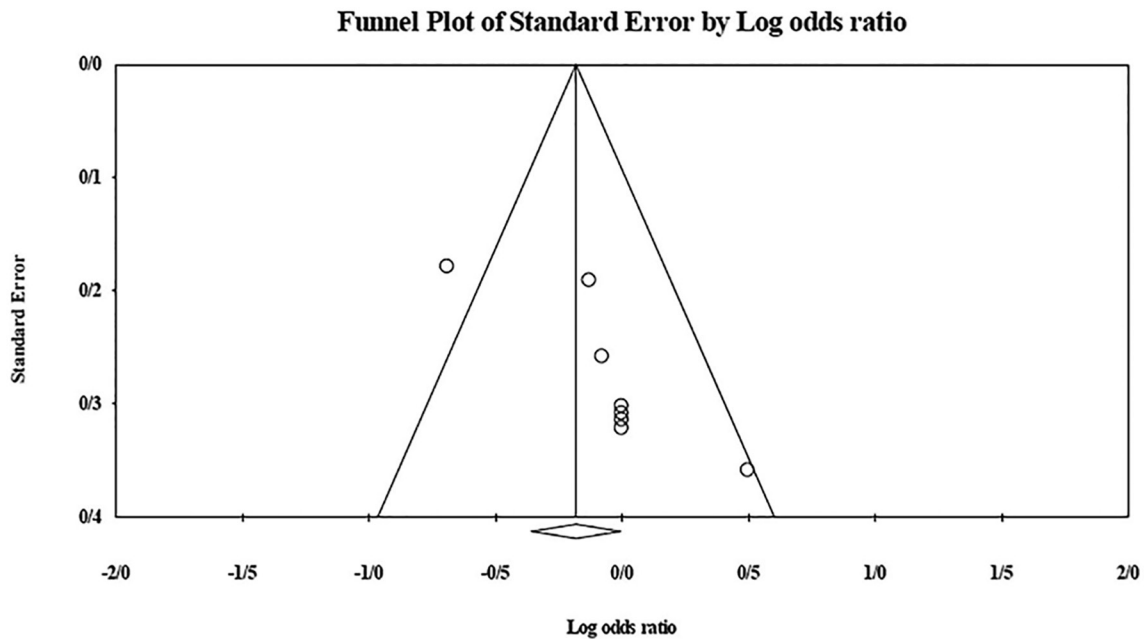


Figure 10. KIR3DL1 Funnel plot showing a significant bias for Faridi et al. (2009).

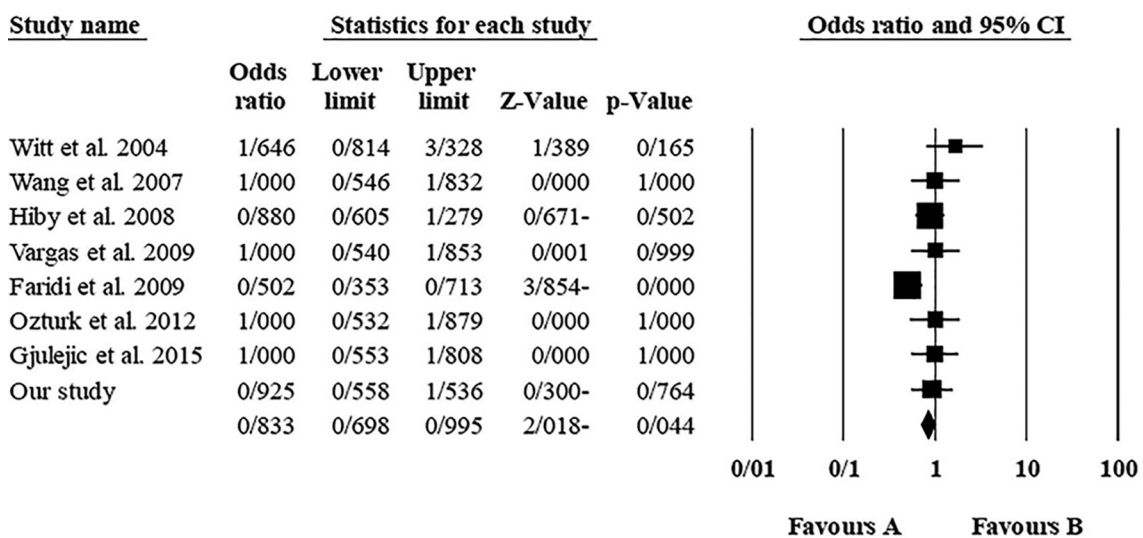


Figure 11. KIR3DL1 Forest plot (fixed).

investigated the role of *KIR2DL4* and *HLA-G* polymorphisms in RSA. Dambaeva *et al.* (2016) showed that maternal *KIR2DS1* is not a risk factor for RSA by itself, rather its combination with maternal *HLA-C2* could be associated.

Interpretation

As we observe above, there are many paradoxical findings for the role of maternal *KIR* genes in RSA. This

can be justified through reasons like different ethnicities, different sample sizes, different RSA group criteria, different control criteria, and so on. In all the studies in Table 1, the genotyping method used was polymerase chain reaction with sequencing specific primers (PCR-SSP), and PCR with sequence specific oligonucleotides (PCR-SSO). Therefore, the genotyping method cannot be a reason for such paradoxes. Other features likely

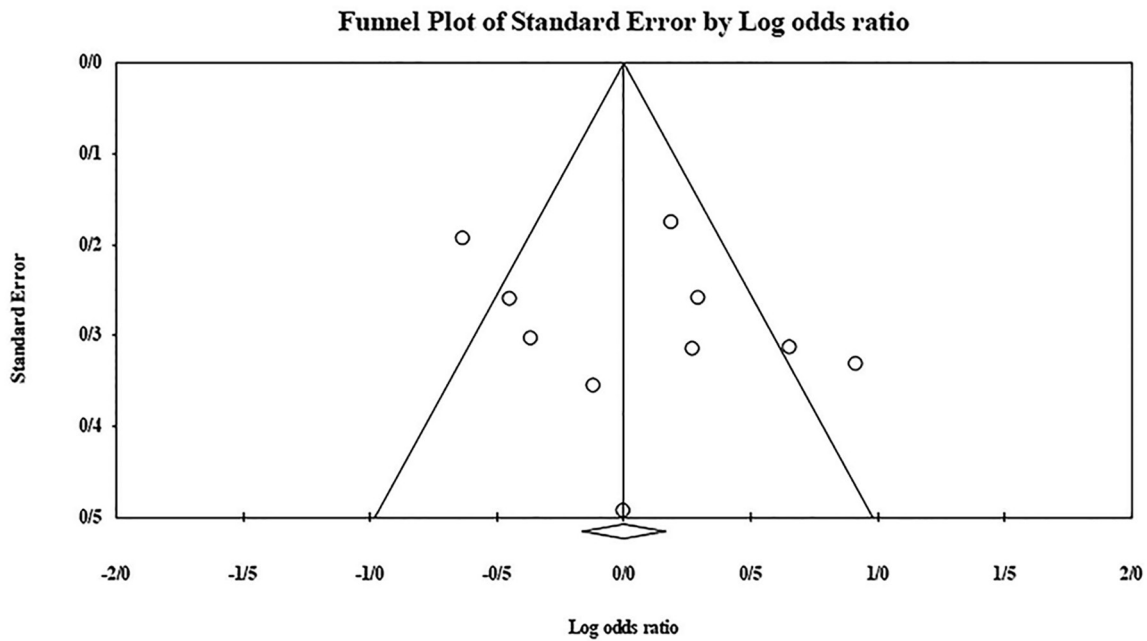


Figure 12. KIR2DS1 Funnel plot showing a huge publication bias which is inconclusive.

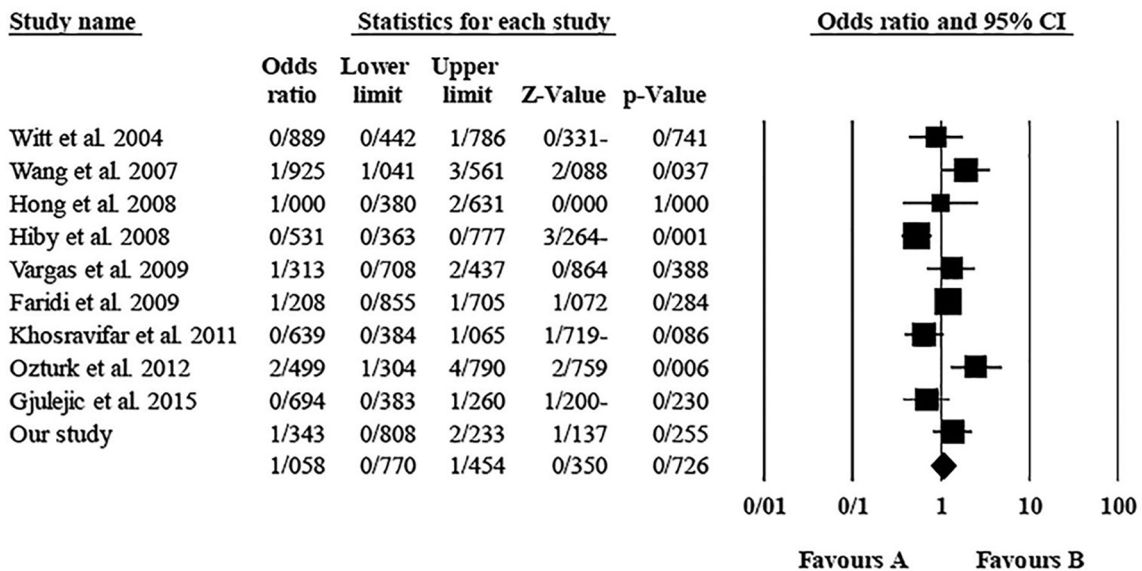


Figure 13. KIR2DS1 Forest plot (random).

to be involved with this paradox are shown as a cluster analysis (Tables 1 and 2, Figure 1).

The results of *KIR2DS1* had more publication bias based on funnel plots than the present meta-analysis. A paradoxical piece of evidence is that in early pregnancy *KIR2DS1* is a helping factor (contrary to some studies), because its activating role (especially in combination

with trophoblast HLA-C2) results in higher cytokine releasing of UNKs (Xiong *et al.*, 2013). Hence, it seems that this receptor has a protecting role for implantation and placentation, and is a risk factor for late pregnancy maintenance. For instance, Alesandru *et al.* (2014) found that maternal AA genotype was a risk factor for the success of assisted reproduction. AA is the most

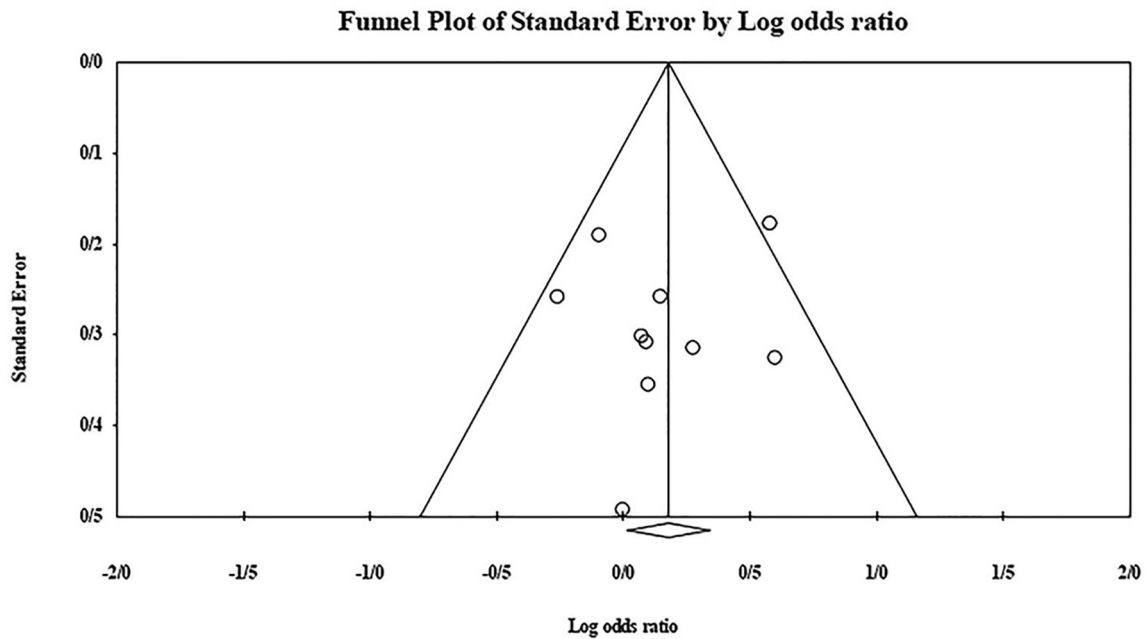


Figure 14. KIR2DS2 Funnel plot showing a significant bias for Faridi et al. (2009).

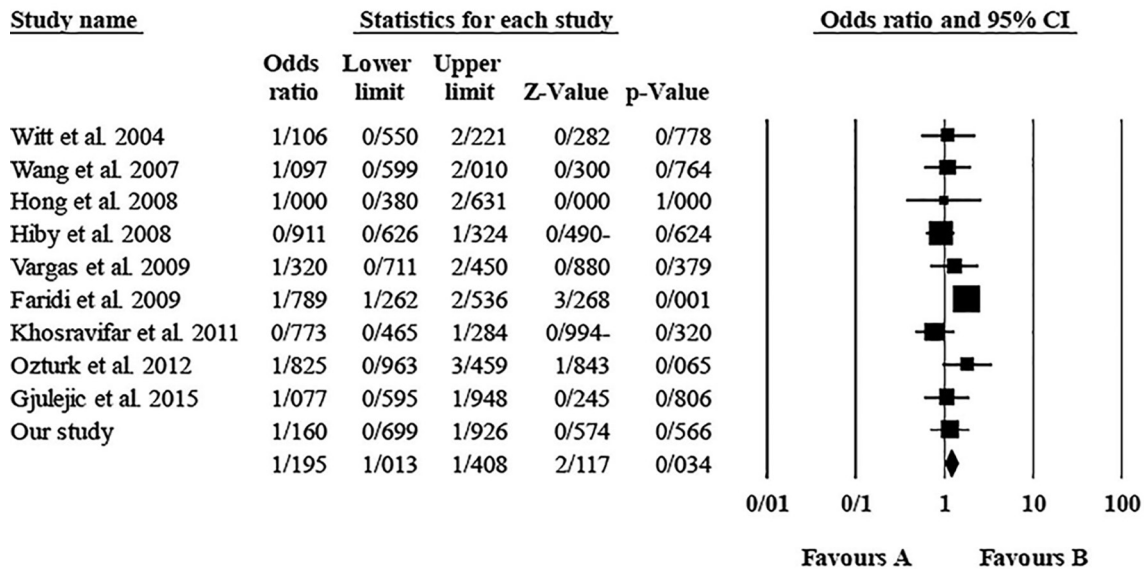


Figure 15. KIR2DS2 Forest plot (fixed).

inhibitory genotype and therefore it supports this hypothesis that NK activation is necessary in early pregnancy. Pregnancy loss has numerous causes, in particular embryo genetic and chromosomal abnormalities. Therefore, the immune system's theoretical role is to reject such malformed embryos. Therefore, this risky role of activating KIRs is in fact a protecting role! Of course, it is remarkable that the lack of genetic evalua-

tion of the lost embryos was a limitation for the studies imported to this meta-analysis. It is suggested that this variable should be adjusted in future studies.

Limitations

Although we found significant associations involving 3 genes in the meta-analysis (Table 3), but these findings would not be reliable, because, 1) the odds ratios

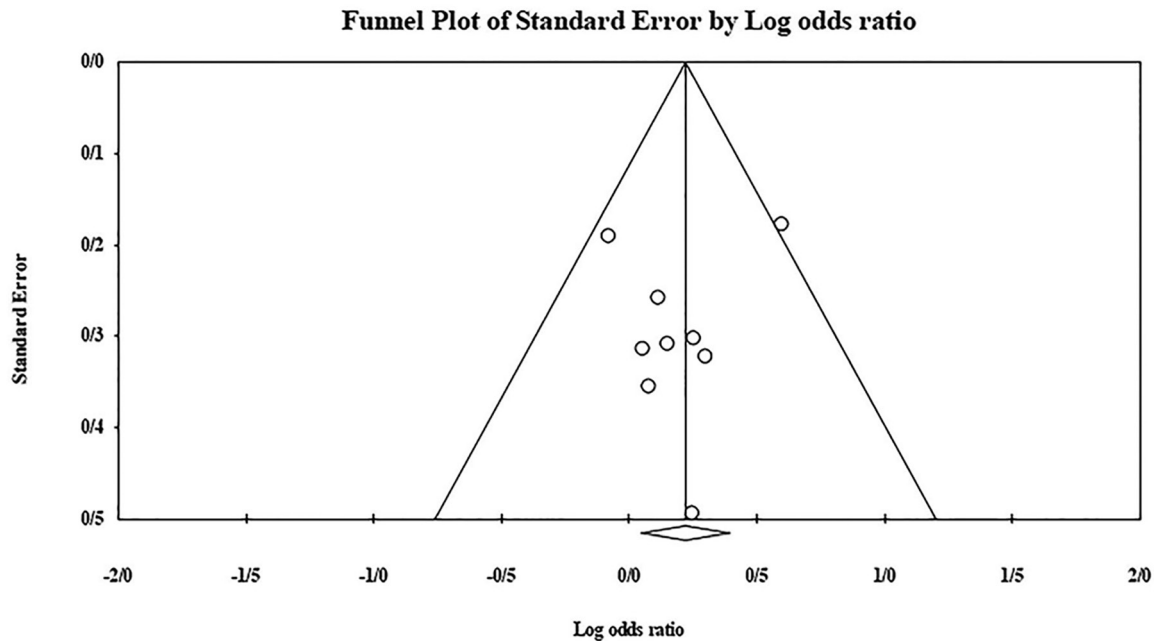


Figure 16. KIR2DS3 Funnel plot showing a rather significant bias for Faridi et al. (2009).

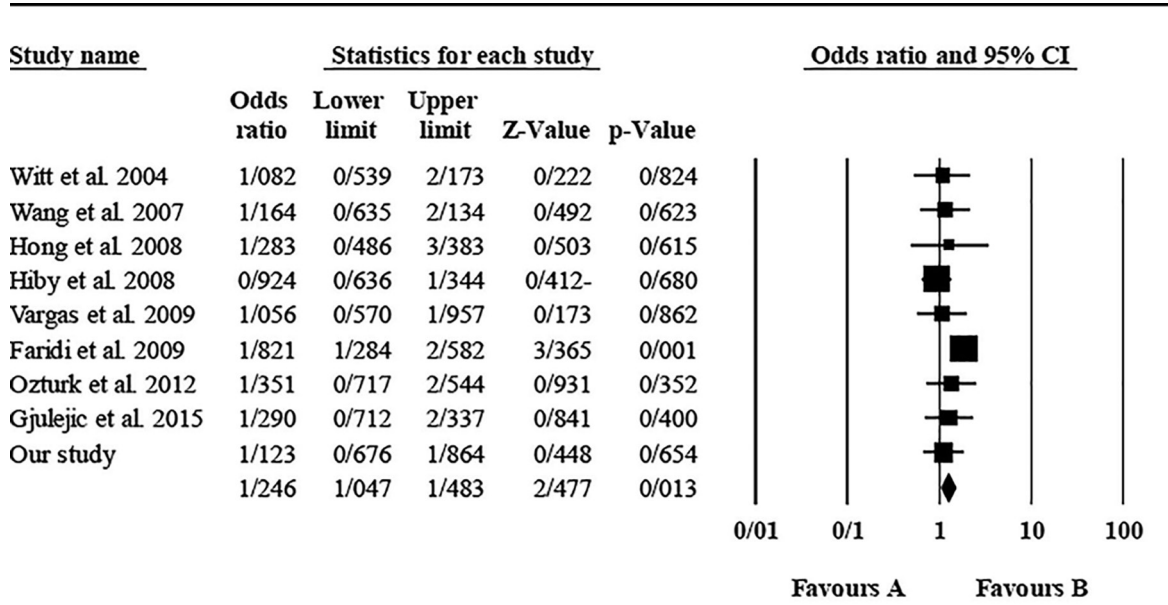


Figure 17. KIR2DS3 Forest plot (fixed).

are not large enough to show a remarkable effect size; 2) the paper selection and homogenizing process of meta-analyses are different and customized among researchers; 3) there were a lot of missed data even in the cited studies; 4) pregnancy loss has a number of

definitions such as abortion, stillbirth (Gold *et al.*, 2010) and assisted reproduction failure (Mitra & Boroujeni, 2015), and happens because due to conditions such as the anti-phospholipid syndrome (APS) (Rand *et al.*, 1997), and there might be confusion involving these

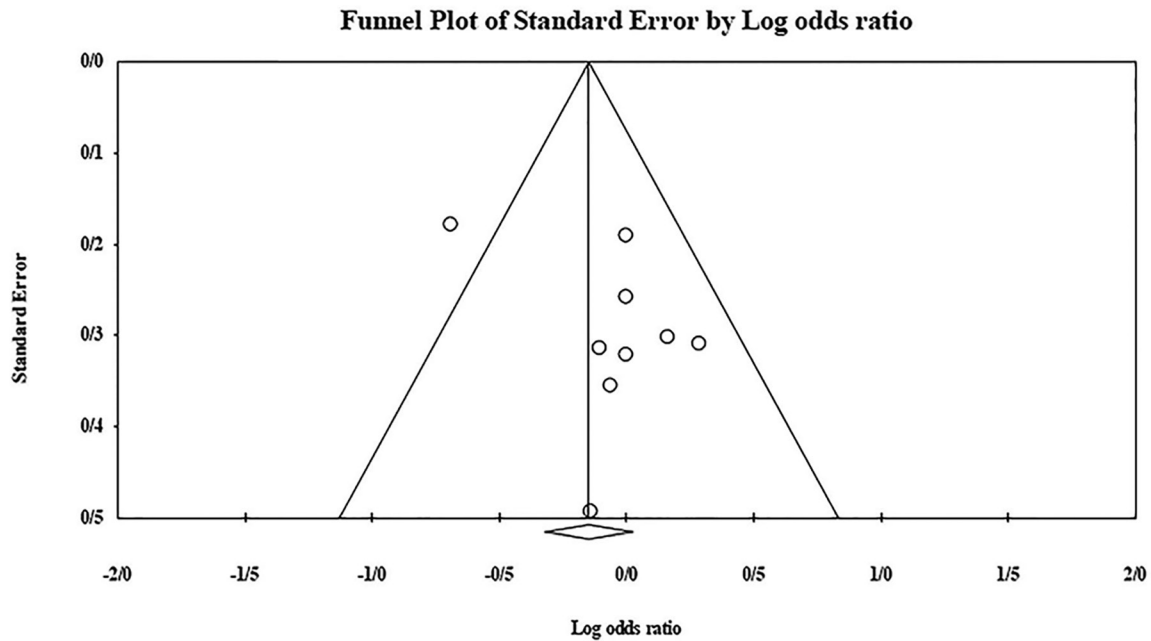


Figure 18. Funnel plot of KIR2DS4 shows a significant bias for Faridi et al. (2009).

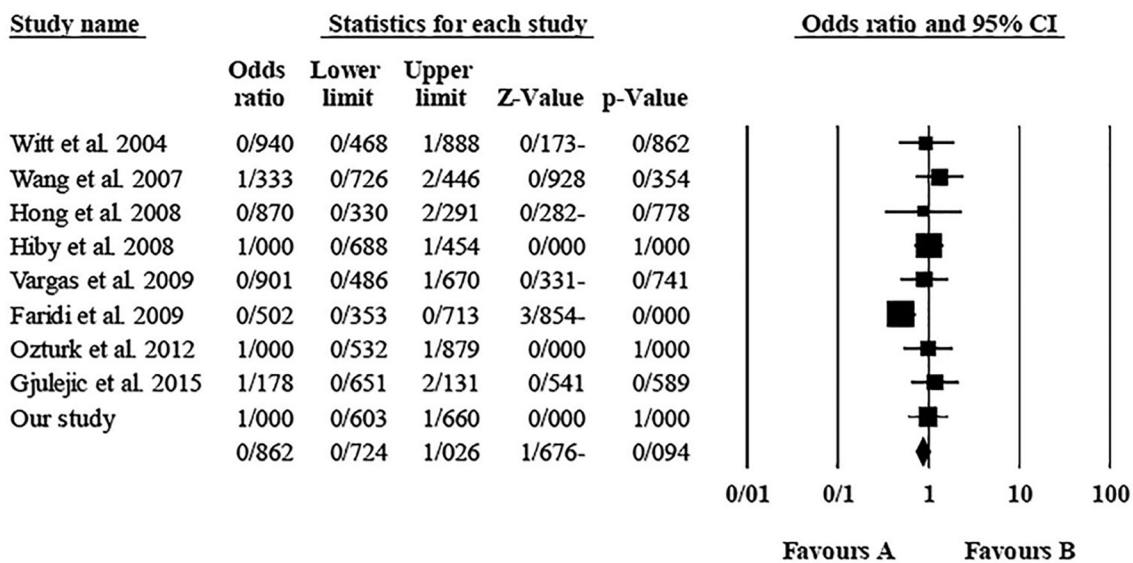


Figure 19. KIR2DS4 Forest plot (fixed).

concepts. Adjusting models in future studies help researchers solve these limitations.

CONCLUSION

The role of maternal *KIR* gene diversity in RSA is still unclear, although our meta-analysis showed 3 genes as associated factors. *KIR3DL1* was a protecting

factor, and *KIR2DS2* and *KIR2DS3*, which proved to be risk factors for RSA. For *KIR2DS1* there was a high heterogeneity. It seems that its role is different among different causes of pregnancy loss. Our previous case-control original investigation showed a significant relation with maternal *KIR2DS1* in combination with paternal *HLA-C2* as a risk factor. In order to clarify this

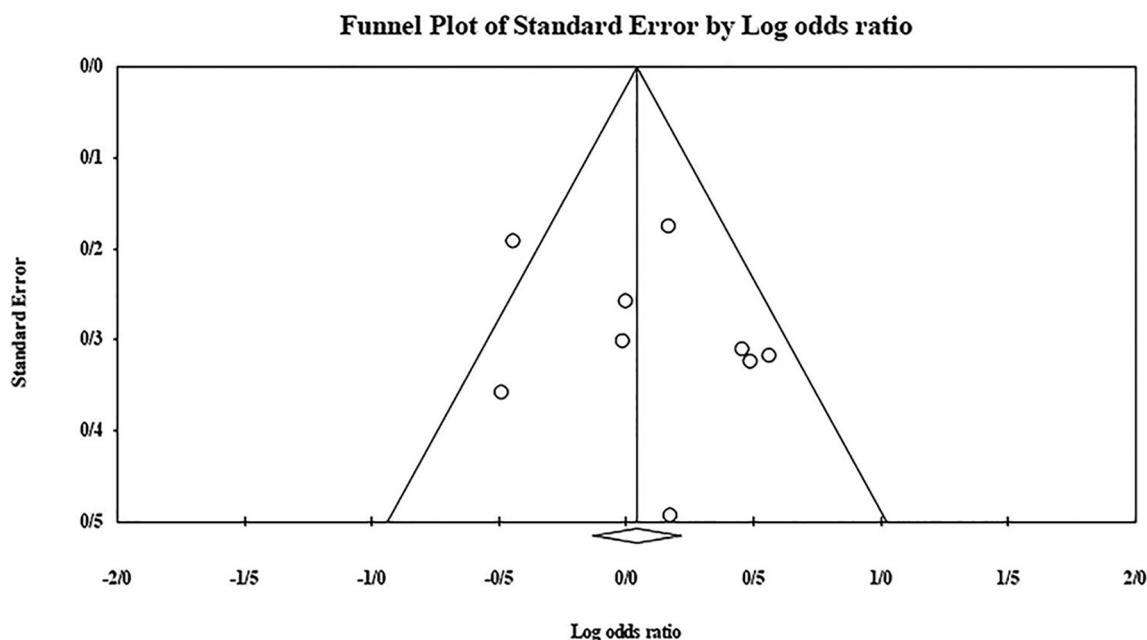


Figure 20. KIR2DS5 Funnel plot showing a significant bias for Hiby et al. (2008).

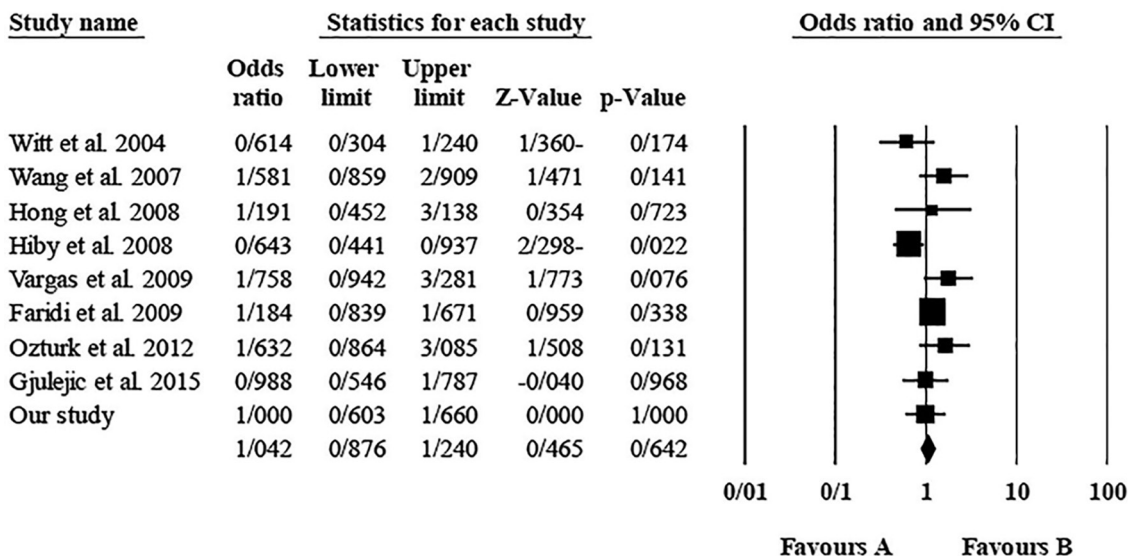


Figure 21. KIR2DS5 Forest plot (fixed).

role we have some suggestions for future studies, such as investigations of this combination concerning the success rate of assisted reproduction, for early first trimester abortions occurring after implantation and early placentation, for stillbirth groups, for abortions

secondary to APS, and for successful and unsuccessful pregnancies of malformed embryos and fetuses. We would also like to suggest adjusting models and cohort studies.

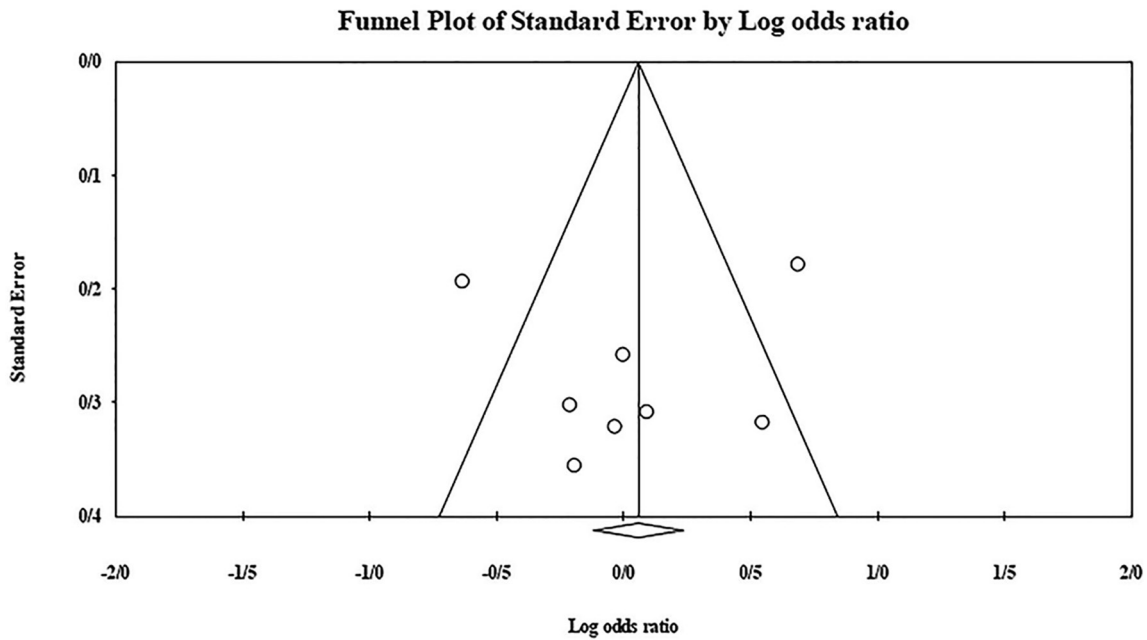


Figure 22. KIR3DS1 Funnel plot showing a significant bias for Hiby et al. (2008) and Faridi et al. (2009).

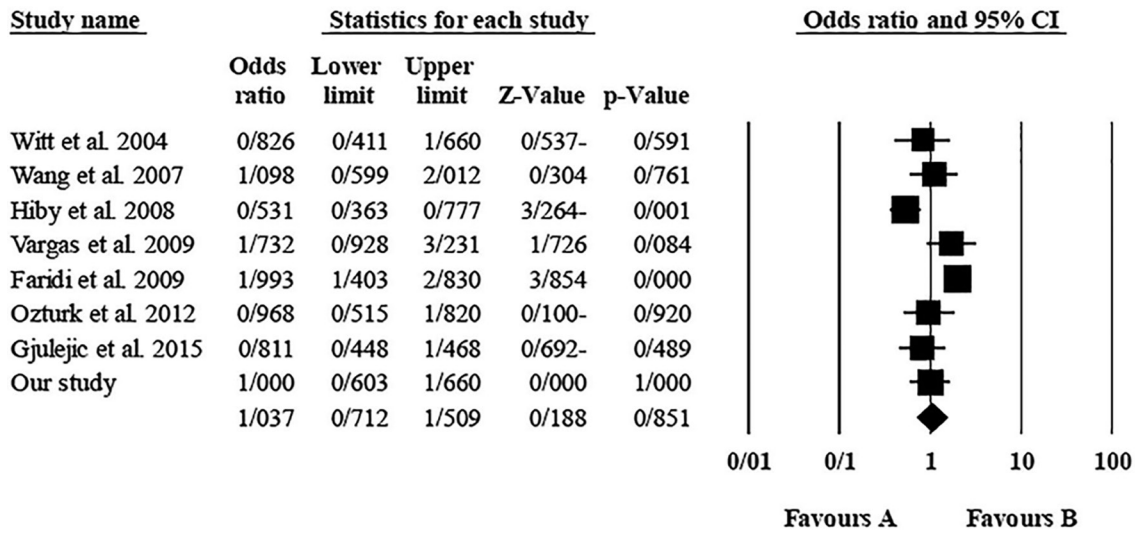


Figure 23. KIR3DS1 Forest plot (random).

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CONFLICT OF INTEREST

None to declare.

Table 3. The pooled results of the meta-analysis. In the cases I2>50 random effect model has also been performed.

Pooled	Fixed effect			Random effect		
	I2	p value	Odds ratio	I2	p value	Odds ratio
Gene						
2DL1	20.92	0.051	0.849	-	-	-
2DL2	36.59	0.325	1.091	-	-	-
2DL3	0.00	0.448	1.069	-	-	-
2DL4						
2DL5	53.79	0.521	0.945	0.00	0.767	0.960
3DL1	47.16	0.044*	0.833	-	-	-
3DL2						
3DL3						
2DS1	70.31	0.990	0.999	0.00	0.726	1.058
2DS2	25.97	0.034*	1.195	-	-	-
2DS3	0.00	0.013*	1.246	-	-	-
2DS4	40.36	0.094	0.862	-	-	-
2DS5	48.52	0.642	1.042	-	-	-
3DS1	75.82	0.525	1.059	0.00	0.851	1.037
2DP1						
3DP1						

* significant at 0.05.

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