

Experiment Investigation of SiO₂ Containing Amino Groups as a Kinetic Promoter for CO₂ Hydrates

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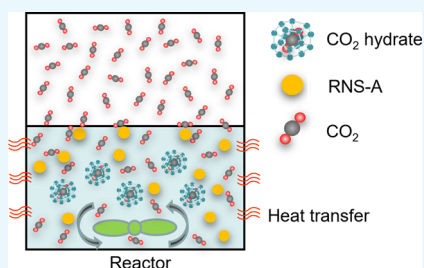
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ABSTRACT: To diminish the greenhouse effect by reducing CO₂ emission into the air based on a capture and sequestration method through hydrates, the thermodynamic and kinetic effects of additives on CO₂ hydrate formation under 1.5 MPa in the presence of 5, 6, 8, 10, and 20 wt % RNS-A (reactive SiO₂ containing amino groups) were studied, and the stirrer speed was set to 800 rpm. This paper calculated the gas consumption and explained the possible mechanisms of RNS-A on CO₂ hydrates. The results showed that RNS-A was a kinetic additive instead of a thermodynamic one. It was found that 5–10 wt % RNS-A all shortened the induction time of hydrates, but only 5 and 6 wt % RNS-A increased the gas consumption of CO₂ hydrates. Although we observed the shortest induction time at a 10 wt % RNS-A system, the lowest gas consumption indicated its weak CO₂ capture and storage ability. In addition, when the concentration was 6 wt %, RNS-A had the highest gas consumption and its reaction time was relatively short. Considering the induction time and gas consumption, 6 wt % RNS-A was the optimal RNS-A concentration for CO₂ capture and sequestration, which was the most suitable for practical applications.



1. INTRODUCTION

Gas hydrates are a new method to capture and store CO₂. They are an environmentally friendly and highly reusable technology with a simple operation, relatively higher storage density, safer process, and lower cost. However, the slow hydrate formation rate and gas storage capacity are the current bottlenecks for industrial application in gas capture, separation, and storage.

Chemical additives are considered to be more attractive to change the thermodynamic or kinetic characteristics of hydrate formation. Thermodynamic additives can change the temperature and pressure conditions for hydrate formation.¹ Thermodynamic additives (e.g., TBAB, THF, CP, and acetone) usually require a large amount of dosage, leading to a high cost.^{2–5} Kinetic additives can affect the surface tension and gas solubility of hydrates and ultimately affect the induction time and gas consumption of hydrate formation.⁶ Kinetic additives (e.g., SDS and SDBS) are harmful to the environment and are difficult to degrade naturally.⁷ In addition, the hydrates doped with surfactants will produce many foams during the separation process. Foams can cause blockages and equipment pollution, which have a negative effect on the practical application of gas hydrates.^{8–11} Rahimi Mofrad *et al.*¹² studied the effects of additives on the formation rate and stability of NGH (natural gas hydrate) in the presence of SL (sulfonated lignin) and SDS. The study found that both additives promote hydrate formation, but compared to SL, the hydrate formed in the SDS solution was less stable. In addition, the hydrate was loose and foamy, which brought some difficulties to the storage, transportation, and degassing of NGH.

A large part of hydrates in nature exists on the seafloor. The hydrate formation in the sandstone and the main component of sandstone is SiO₂. SiO₂ is a substance that exists widely in nature, which is convenient and easy to obtain, so it is chosen as an additive. Due to their small particle size and large specific surface area, nanoparticles can increase the contact area of gas–liquid, enhance heat dissipation, and promote the kinetics of gas hydrate formation.¹³ Nashed *et al.*¹⁴ reviewed the current research status of the effects of nanoparticles on hydrate formation, including carbon nanomaterials, metallic nanoparticles, and metallic and nonmetallic oxide nanomaterials. They stated that nanoparticles were kinetic additives with a weak impact on the equilibrium conditions of hydrates, and the concentration had little effect on thermodynamic effects. They also pointed out that nanoparticles are favorable for promoting hydrate formation of hydrophobic gases such as CH₄ rather than soluble gases such as CO₂. Wang *et al.*¹⁵ carried out experiments on CH₄ hydrate formation in the hydrophilic nano-SiO₂ system with the concentration range of 1–6 wt % under the initial experimental conditions of 278.15 K and 5 MPa. They stated that the hydrophilic nano-SiO₂ inhibited CH₄ hydrate formation. Increasing the concentration may

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enhance the inhibiting effect, while the inhibition may be weakened when the concentration was very high.

Nanoscale SiO₂ has been proven to promote hydrate formation in the previous literature, while microscale SiO₂ is mostly used as a porous medium to participate in hydrate formation. Ge *et al.*¹⁶ studied the formation and dissociation of CH₄ hydrates in a fixed bed of silica sand at 277.15 K and 8 MPa. Two sizes of silica sand (180–250 μm and 300–900 μm) were used in this experiment. They found that small-sized silica sand (180–250 μm) was more conducive to CH₄ hydrate formation and dissociation. Its hydrate formation amount, gas consumption, and CH₄ recovery rate were greater than those of large-sized silica sand. Mekala *et al.*¹⁷ studied the formation kinetics of CO₂ hydrates in pure water, seawater, and porous media at 276.15 K and 6 MPa. The silica sand used in this work included three particle sizes: 160, 460, and 920 μm. They found that the gas consumption and water conversion rate increased with the decrease in the silica sand particle size during the hydrate formation process. Kumar *et al.*¹⁸ studied the formation of CO₂ hydrates in three porous media silica gels (pore sizes of 60–120 mesh, 100–200 mesh, and 230–400 mesh, respectively) at 274 K and 3.55 MPa. They found that a larger surface area can consume more CO₂ during the hydrate formation process and shorten the induction time.

Many scholars had studied the effects of nano-SiO₂ and hundreds of micrometer-sized SiO₂ on hydrates, but no one had studied the effects of smaller micrometer-sized particles as an addition on hydrate formation. Also, Ren and Lu¹⁹ found that SiO₂ hydrophilic particles had better dispersibility in water than hydrophobic ones. Here, RNS-A (reactive SiO₂ with amino groups) and its concentration on the thermodynamic and kinetic hydrate process of CO₂ under 1.5 MPa will be investigated. Furthermore, the relationship between hydrate formation time and CO₂ consumption will also be studied by the grey correlation degree method. Also, the possible mechanisms of hydrophilicity and alkalinity of RNS-A on CO₂ hydrates would be explained. This basic thermodynamic and kinetic work will conduce to help develop the application of hydrates in the capture and storage of CO₂.

2. RESULTS AND DISCUSSION

2.1. Particle Size, Shape, and Element Analysis of RNS-A. The particle size of RNS-A was measured by the Laser Diffraction Particle Size Analyzer (see Figure 1). The average particle size of RNS-A is 5.89 μm. Among them, only about 3.5% of RNS-A has a particle size at the nanoscale.

The functional groups on the surface of RNS-A were characterized by FT-IR (Fourier transform infrared spectroscopy) (see Figure 2). The absorption peak of 1105 cm⁻¹ is caused by the asymmetric stretching vibration of Si–O–Si. The peaks at 797 and 467 cm⁻¹ correspond to the symmetric stretching and bending vibrations of Si–O, respectively. The peaks at 1637 and 1568 cm⁻¹ correspond to bending vibrations of N–H. The absorption peak of 3415 cm⁻¹ may correspond to the stretching vibration of N–H or O–H. It can be seen from the peak area that the content of amino groups is very small compared to SiO₂.

In addition, the micromorphology of RNS-A was observed by SEM (scanning tunneling microscopy), and the elements were analyzed by EDS (energy dispersive spectroscopy) (see Figure 3). It can be seen that the shape of RNS-A is spherical, and there is no agglomeration. Its elements include Si, O, C,

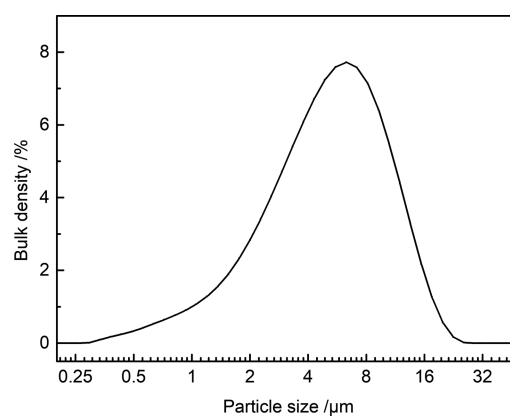


Figure 1. Particle size distribution of RNS-A.

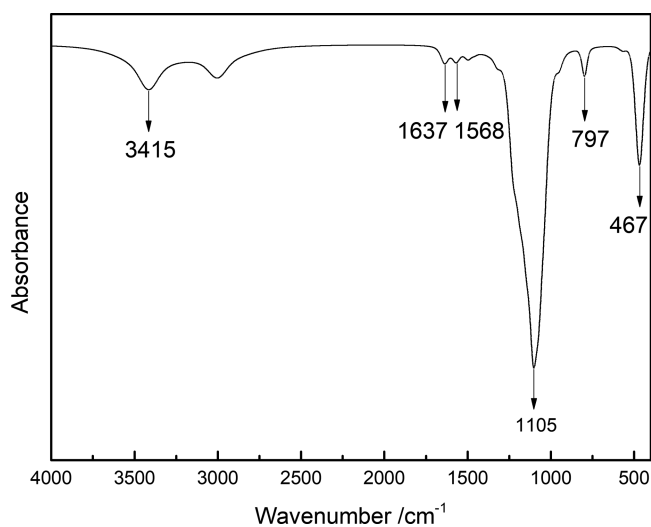


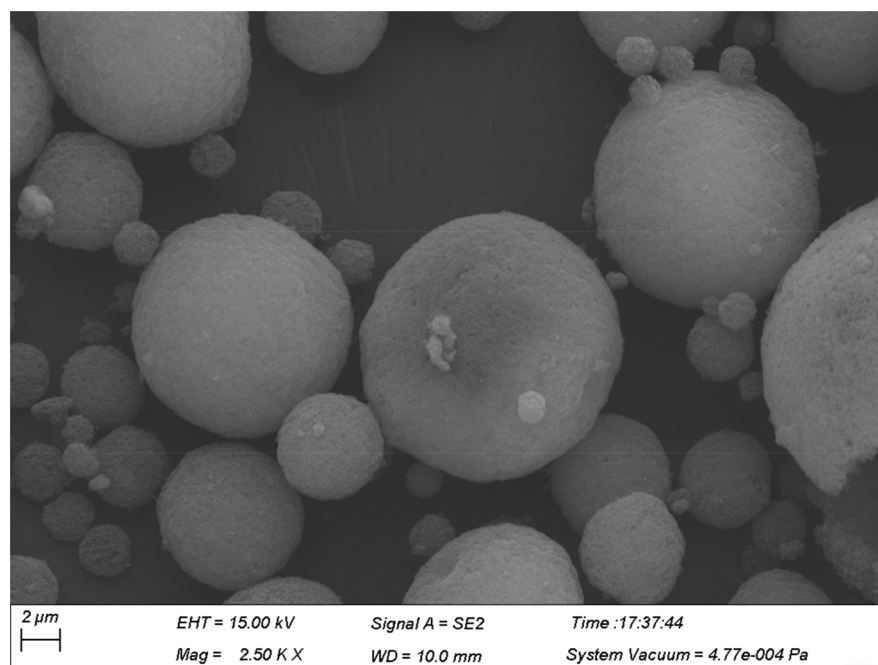
Figure 2. FT-IR images of RNS-A.

and N. The small content of N is observed in RNS-A, which comes from the amino functional group.

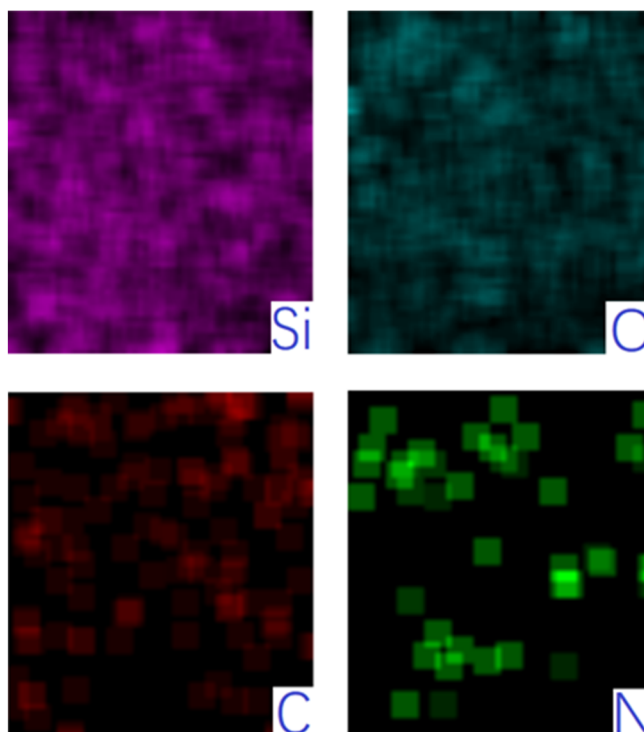
2.2. Reliability of the Experimental Equipment. To confirm the reliability of our experimental equipment, the phase equilibrium data of CO₂ in pure water were first tested and compared with the data from the reported literature (see Figure 4). It is seen that the phase equilibrium data are very close to the reported ones, proving that the experimental equipment used here is reliable.

2.3. Thermodynamic Analysis. Table 1 shows the phase equilibrium data of CO₂ hydrates in suspension in the presence of RNS-A with various concentrations. It is observed that all the equilibrium pressures and temperatures are very close to those of the pure water system. It indicates that the addition of RNS-A has little influence on the phase equilibrium condition of CO₂ hydrate formation, and the effect of RNS-A concentration is also very tiny. It can be inferred that RNS-A is not a thermodynamic additive for CO₂ hydrates.

2.4. Kinetic Analysis. Although RNS-A has a slight effect on hydrate equilibrium conditions, it can change the formation time and gas consumption during the hydrate process significantly. Figure 5 shows the formation time and gas consumption of CO₂ hydrates in all systems. Figure 5a shows the induction time, growth time, and total time. Figure 5b shows the gas consumption corresponding to each time.



(a)



(b)

Figure 3. (a) SEM images of RNS-A; (b) EDS images of RNS-A (face analysis).

2.4.1. Induction Time. It is observed that the induction time of hydrate formation is 32,556 s in the pure water system. In the presence of RNS-A concentration of 5–10 wt %, the induction time is shorter than that of the pure water system, indicating that RNS-A has a significant promoting effect on the nucleation of CO₂ hydrates. In addition, the induction time decreases with RNS-A concentration increasing from 5 wt % to 10 wt %. There are very slight differences in the induction time for systems with 5, 6, and 8 wt % RNS-A, which are about 78%

of the pure water system. When the RNS-A concentration is increased to 10 wt %, the induction time is reduced to 9198 s, which is much faster than that of the pure water system.

This is because the formation of CO₂ hydrates is an exothermic process that causes the temperature to rise, which reduces the driving force for hydrate formation. RNS-A has relatively good thermal conductivity and can remove heat more efficiently and faster.^{14,21} Second, RNS-A with smaller particles can provide a larger gas–liquid contact surface area.¹⁷ Third,

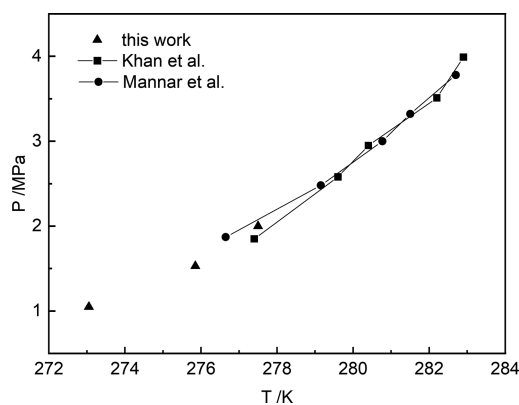


Figure 4. CO₂ phase equilibrium data in this work and other literature data: solid up-pointing triangle, this work; solid circle with a solid line, Mannar *et al.*; solid square with a solid line, Khan *et al.* Adapted from Mannar *et al.*⁵ and Khan *et al.*²⁰

Table 1. Phase Equilibrium Data of CO₂ Hydrates

system	P/MPa	T/K
pure water	1.53	275.75
5 wt % RNS-A	1.52	275.95
6 wt % RNS-A	1.50	275.40
8 wt % RNS-A	1.50	275.45
10 wt % RNS-A	1.51	275.55
20 wt % RNS-A	1.53	275.35

SiO₂ can also be the seeds of heterogeneous hydrate nucleation, providing nucleation sites, increasing the probability of nucleation, reducing the driving force required for the reaction, and shortening the induction time.^{17,22–26} In addition, the Brownian motion of RNS-A and the stirring of the magnetic stirrer intensify the collision between RNS-A and destroy the bound water molecules on the surface.²⁷ Moreover, Bai *et al.*²⁸ found that the water film on the particles with weak hydrophilicity is less structured, has a weak restriction on water molecules bound on the surface, and thus promotes the gas–liquid interaction.

However, in the very concentrated suspension with 20 wt % RNS-A, the induction time of CO₂ hydrates increases to 49,728 s, which is vastly longer than that of the pure water system, indicating that the very high concentration is not favorable for CO₂ hydrating nucleation. It may be ascribed to the fact that although RNS-A with smaller particles can provide a larger gas–liquid contact surface area, the high concentration increases the difficulty of gas–liquid contact.²⁹ The concentration of 20 wt % may cause agglomeration of RNS-A, which can reduce the effective specific surface area and block the heat dissipation, thereby delaying the CO₂ hydrate nucleation.³⁰

2.4.2. Gas Consumption. Gas consumption is another important index for selecting the optimal additives for hydrate formation. Large gas consumption implies an excellent ability for gas storage. Figure 5b shows the gas consumption corresponding to each time. We can find that except for the system of 20 wt %, Δn_g is less than Δn_h in other systems, indicating that a large number of hydrates are formed after nucleation. In the system with 20 wt % RNS-A, the growth of CO₂ hydrates is inhibited, with a very long induction time and thus relatively larger gas consumption during the inducing period. Figure 6 shows the change in gas consumption over time in each experimental system. Clearly, gas consumption

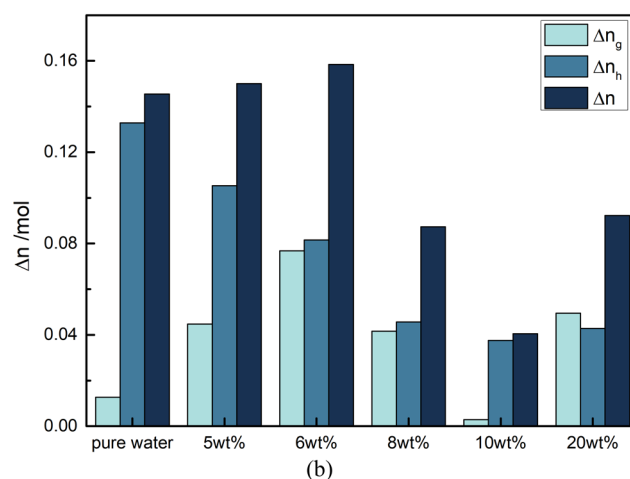
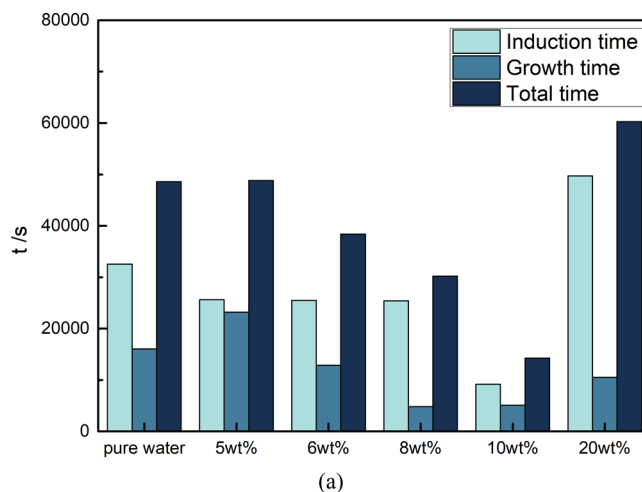


Figure 5. Relationship between formation time and gas consumption. (a) Induction time, growth time, and total time change with RNS-A concentration. (b) Gas consumption corresponding to each time (Δn_g is the gas consumption within the induction time; Δn_h is the gas consumption within hydrate growth time; Δn is the total gas consumption).

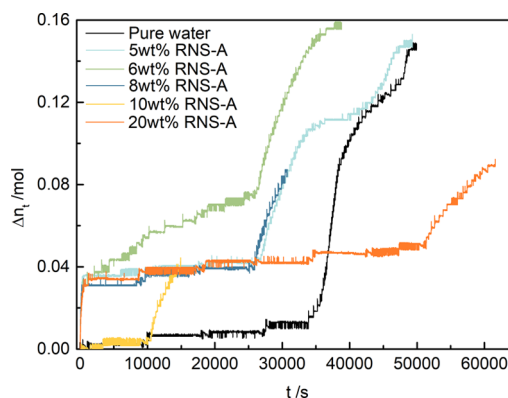


Figure 6. Change in gas consumption over time in different systems.

increases slowly at first and then sharply rises. The slow-growth phase causes gas dissolution and hydrate nucleation until the system reaches gas–liquid equilibrium, while the sharp-growth phase implies a large number of CO₂ hydrates being formed massively.

It is observed that within the range of 5–10 wt %, the overall trend is that CO₂ consumption decreases with increasing concentration of RNS-A. When the concentrations are 5 and 6 wt %, the gas consumption is slightly higher than that of the pure water system, wherein the gas consumption of 5 wt % RNS-A is slightly lower than that of 6 wt % RNS-A. It is because the lower concentration of amino groups has a relatively weaker effect on the inhibition of the activity of water molecules.²⁷ As the concentration of RNS-A increases, the water activity decreases with the increase in the number of amino groups. Furthermore, this is due to the Brownian motion of RNS-A, which reduces the membrane resistance at the gas–water interface and increases the gas consumption of CO₂ hydrates.^{14,31,32} In addition, the growth time of hydrates with 5 and 6 wt % RNS-A is relatively longer than those of other systems, providing sufficient hydrating time for more CO₂ into the liquid phase to become hydrates. It is also worthy to note that the amino groups can adsorb CO₂ chemically and thus enhance CO₂ consumption.³³

When the RNS-A concentration is increased to 8 wt %, CO₂ consumption is significantly reduced, reaching only 60% of the pure water system. When the concentration continues to increase to 10 wt %, CO₂ consumption is only half of 8 wt % RNS-A. Shorter growth periods are observed at 8 and 10 wt % compared to the pure water system, which possibly affects gas consumption. The gas–liquid interface is a suitable site for hydrate growth,³⁴ and the hydrate formed initially increases the resistance of CO₂ mass transfer from the gas phase into the liquid phase, which hinders the absorption of CO₂.³⁵ Therefore, the gas consumption of 8 and 10 wt % systems is significantly reduced.

When the RNS-A concentration is increased to 20 wt %, the gas consumption is 63% of the pure water system. The gas consumption at this time has a certain increase compared to 10 wt %, which is similar to the 8 wt % system. This is possible because of the fact that although the high-concentration RNS-A blocks CO₂ diffusion into the liquid phase, the increased hydrate formation time may provide an opportunity for more CO₂ to enter into the hydrate cages. Therefore, the gas consumption of the 20 wt % RNS-A system is higher than that of the 10 wt % RNS-A system.

The slope of Figure 6 represents the gas consumption rate during the hydrate formation process. It can be found that the hydrate is generated rapidly at first and then the rate becomes slower, which indicates that the formation of the initial hydrate hinders the mass transfer and slows down the rate of hydrate growth.

In summary, in the range of 5 to 10 wt %, RNS-A shortens the hydrate induction time. At 5 and 6 wt %, RNS-A increases the gas consumption of hydrates, wherein the gas consumption of 6 wt % RNS-A is slightly higher than that of 5 wt % RNS-A. In terms of induction time and gas consumption, 6 wt % RNS-A is the optimal concentration for CO₂ capture and storage. In addition, because RNS-A is also feasible economically, it can be taken into account to apply to industrial practice to capture, separate, and store CO₂.

2.5. Correlating Hydrating Time and Gas Consumption. Combining Figure 5a and Figure 5b, it is concluded that the hydrating time poses a direct impact on gas consumption, and usually, the rapid formation of hydrates will reduce the gas storage capacity of the hydrates. The critical task is to find a balance between hydrate formation time and gas consumption.

Based on the grey relational degree, the relationship between hydrate formation time and gas consumption was analyzed. The degree of relevance, r , ranges from 0 to 1, and is calculated and shown in Table 2. A larger r value indicates a higher correlation degree between time and gas consumption.

Table 2. Degree of Relevance, r , between Reaction Time and Gas Consumption

r	$\Delta n_g/\text{mol}$	$\Delta n_h/\text{mol}$	$\Delta n/\text{mol}$
induction time/s	0.6132	0.6276	0.7218
growth time/s	0.4635	0.7305	0.7715
total time/s	0.6308	0.6779	0.7882

It can be seen that the r between the total gas consumption and total time is the highest. Compared to the induction time we usually focus on, the r between growth time and gas consumption is more closely related. It is possible because gas consumption during the induction stage is much less, while a large amount of CO₂ would be consumed during the hydrate growth period. More gas consumption causes the growth period to affect the total gas consumption more than the induction period.

2.6. Mechanism of RNS-A on CO₂ Hydrates. In fact, the correlation between CO₂ hydrates and concentration is closely related to the property of RNS-A. As Qin *et al.*³³ reported, the amino group on the RNS-A surface, which is hydrophilic, has a certain inhibitory effect on the activity of water molecules. To a certain extent, this explains why 20 wt % RNS-A exhibits a strong inhibitory effect.

The possible specific inhibitory mechanism of RNS-A is shown in Figure 7. Figure 7a shows that RNS-A can bind the water molecules around RNS-A to form water films and reduce the activity of water molecules.²⁷ However, the hydrophilicity of amino groups is weaker, and the water film has less structured and weaker restrictions on water molecules bound on the surface, leading to a weaker inhibition effect on hydrate formation.²⁸ Figure 7b shows that the amino groups on the

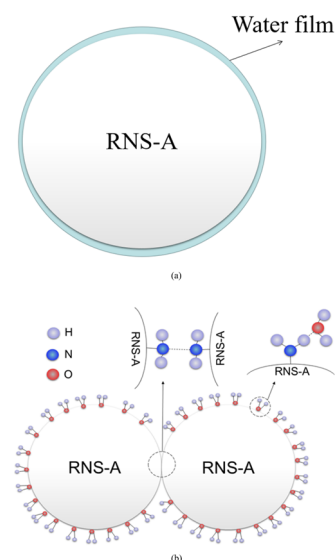


Figure 7. Possible inhibition mechanism of RNS-A on CO₂ hydrates. (a) Possible distribution of water films and CO₂ hydrates on the surface of RNS-A. (b) Surface characteristics of RNS-A: combination of amino groups with water molecules or adjacent amino groups.

surface of RNS-A form hydrogen bonds with water molecules against the influence of CO₂ molecules through weak Van der Waals force stably, competing with hydrates for host molecules. In addition, amino groups can form hydrogen bonds with adjacent amino groups. Hydrophilic groups can effectively destroy the structure of water, enhancing the inhibition effect of hydrates.³⁶

Nevertheless, the formation of gas hydrates is an exothermic process that causes temperatures to rise, thus reducing the driving force for hydrate formation. RNS-A has relatively good thermal conductivity and larger contact surface area, which can remove heat more efficiently and faster.^{14,17,21} In addition, RNS-A can act as seeds of heterogeneous hydrate nucleation, providing nucleation sites, increasing the probability of nucleation, and enhancing the driving force to promote hydrate formation.^{23–26} Meanwhile, the Brownian motion of RNS-A can reduce the film resistance at the gas–liquid interface and enhance the mass transfer effect.^{14,32} In addition, the Brownian motion of RNS-A and the stirring of the magnetic stirrer destroy the bound water molecules on the surface,²⁷ which promotes the hydrate formation.

Figure 8 shows the Brownian motion of RNS-A, and the stirring effect of the reactor aggravates the collision between

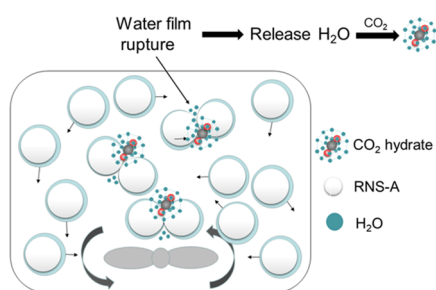


Figure 8. Possible promotion mechanism of RNS-A on CO₂ hydrates: the Brownian motion of RNS-A and water film rupture.

RNS-A. Due to the less structured water film on the surface of RNS-A, the restriction on water molecules is weaker, and thus, the water film around RNS-A is easier to break.²⁸ In addition,

due to the fact that amino groups can absorb CO₂ chemically, its surface can retain more CO₂ molecules, which results in concentrated CO₂ on the surface of RNS-A.³³ When the released water molecules are released and then form a cage, CO₂ molecules on the surface of RNS-A will enter the cage to create hydrates, which is a dynamic equilibrium process.

In cases of a low concentration of RNS-A, *i.e.*, 5 and 6 wt %, due to the fact that RNS-A has good thermal conductivity and large contact surface area, RNS-A can shorten the induction time and increase gas consumption, which acts as a kinetic promoter for CO₂ hydrates.

When the concentration continues to increase, *i.e.*, 8 and 10 wt %, the induction time continues to shorten. Due to the rapid formation of the initial hydrate, the diffusion of CO₂ from the gas phase to the liquid phase is hindered. Therefore, the gas consumption of the 8 and 10 wt % systems is reduced.

In the case of a very high concentration of RNS-A, *i.e.*, 20 wt % here, agglomeration of RNS-A will hinder the hydrate formation rate. In addition, too many RNS-A increases the resistance of Brownian motion³⁰ and limits CO₂ diffusion in solutions, consequently increasing the induction time and delaying hydrate growth. At this time, RNS-A acts as a kinetic inhibitor.

3. CONCLUSIONS

In this work, the effects of RNS-A on the phase equilibrium, induction time, and gas consumption of CO₂ hydrates were studied under 1.5 MPa. The experimental results showed that RNS-A had no significant effect on the phase equilibrium condition of CO₂ hydrates, whereas it could shorten the induction time, indicating that it was a kinetic promoter instead of a thermodynamic one. It was found that 5–10 wt % RNS-A all shortened the induction time of hydrates. Although 10 wt % RNS-A had the best effect on shortening the induction time, its gas consumption was inhibited, which was only 28.8% of that in the pure water system. In addition, only 5 and 6 wt % RNS-A increased the gas consumption of CO₂ hydrates. When the concentration increased to 20 wt %, RNS-A was transformed into the hydrate kinetic inhibitor completely, which shortened the induction time and reduced gas

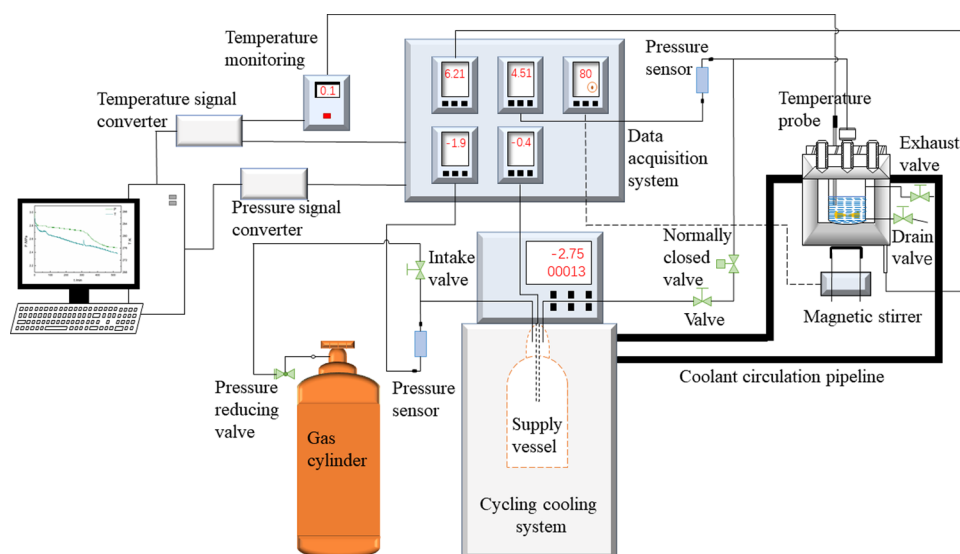


Figure 9. Schematic diagram of the hydrate kinetic experimental equipment.

consumption. A balance should be pursued between the induction time and gas storage capacity when considering appropriate promoters. Considering the induction time and gas consumption, 6 wt % RNS-A was the optimal RNS-A concentration for CO₂ capture and sequestration, which was the most suitable for practical applications.

In addition, RNS-A is only a kinetic promoter, and it can be taken into account to be compounded with a thermodynamic promoter to promote CO₂ hydrates in the future. This paper mainly studied the effect of amino groups on the formation of CO₂ hydrates, and we can also explore the effects of other functional groups on the formation of CO₂ hydrates and find better promoters for CO₂ capture and sequestration to be used in industrial practice as soon as possible.

4. EXPERIMENTAL SECTION

4.1. Experimental Materials. Deionized water, CO₂, and RNS-A were the main materials in this study. Deionized water was made in a laboratory. CO₂ (99.99%) was provided by Jining Xieli Special Gas Co., Ltd. RNS-A (reactive SiO₂ containing amino groups) was supplied by Henan Wangwu Nano Technology Co., Ltd.

4.2. Experimental Equipment. The schematic diagram of the hydrate kinetic experimental equipment is demonstrated in Figure 9. It consists of a high-pressure stirring reaction reactor, a cycling cooling system, and a data acquisition system. The high-pressure reactor is a cylindrical vessel with a volume of 100 mL, which can undertake pressures ranging from 0 to 20 MPa and temperatures from 253.15 to 423.15 K. The speed range of the stirring system (80YT25DV22) is 0–1500 rpm and its power is 25 W, which was supplied by JSCC Automation Co., Ltd. The cycling cooling system (CKDHX-1015 type) uses ethylene glycol as the coolant to maintain the low temperature. The temperature range of the cycling cooling system is 263.15–373.15 K, and the control precision is ±0.05 K. A Pt100 platinum resistance sensor was used to measure the temperature of the reactor and cycling cooling system with an accuracy of ±0.1 K. The pressure of this system was detected by a pressure sensor with an accuracy of ±5% (0–10 MPa). The temperature and pressure were monitored and recorded by a data acquisition system at an interval of 3 s.

4.3. Experimental Procedure. **4.3.1. Characterization of RNS-A.** The particle size of RNS-A was measured by the Laser Diffraction Particle Size Analyzer (Mastersizer 3000, Malvern Instruments Co., Ltd., England). The microscopic morphology of RNS-A was detected by SEM (Merlin Compact, Carl Zeiss NTS GmbH, Germany), and the component elements were analyzed with an attached facility OXFORD EDS. The functional groups on the surface of RNS-A were characterized by FT-IR (Tensor 37, NETZSCH-Gerätebau GmbH, Germany).

4.3.2. Hydrate Formation Experiments. Before starting the experiment, 5, 6, 8, 10, and 20 wt % RNS-A suspensions were prepared, which needed to be ultrasonicated for 30 min. The experimental pipelines, valves, reactor sealing caps, and instrument interfaces were then checked to avoid gas leakage.

After that, the prepared suspension (50 mL) was introduced into the reactor, and the reactor was sealed immediately. To avoid the influence of air in the reactor, CO₂ was used to flush the reactor three times.

The temperature of the reactor was cooled by the cycling cooling system with a speed of 0.2 K/min. When the temperature of the cooling system reached 279.15 K, the

temperature dropping rate was changed to be 0.5 K/h according to the report of Yan³⁷ to ensure the sufficient hydrate formation time and gas consumption. When the temperature of the reactor approached 285.15 K, CO₂ with a pressure of 1.5 MPa was introduced into the reactor, and the magnetic stirrer was turned on immediately. The stirring speed was set to 800 rpm according to Jiang and Yan.^{38,39} The temperature and pressure were collected by a data acquisition system. A sudden and sharp drop of pressure in the gas storage tank indicates that a large quantity of CO₂ was consumed due to the formation of CO₂ hydrates. When the pressure of the gas storage tank remained unchanged for 30 min, indicating that the hydrate process was finished, the data acquisition system was turned off, the stirring was stopped, and the gas inlet valve was closed.

4.4. Determination of Hydrate Induction Time.

Induction time refers to the time required from the first equilibrium of the system to a large number of visible crystals appearing in the system.³ The induction time was determined based on the macroscopic dynamic hydrating measurements. As shown in Figure 10, CO₂ dissolves in the solution and then

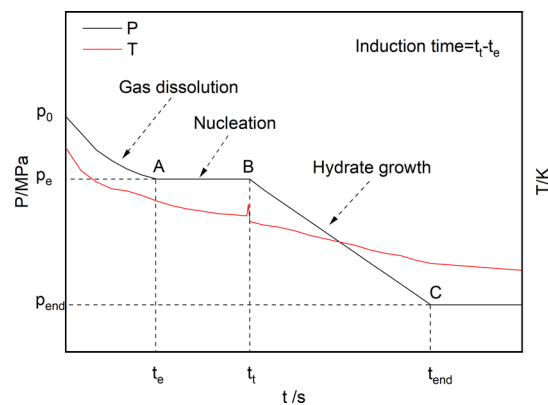


Figure 10. Determination of hydrate induction time.

reaches equilibrium at t_e .⁴⁰ After that, the pressure remains nearly unchanged between t_e and t_t until a large number of hydrates start to form at t_t when the pressure drops sharply. Here, the induction time is defined as the time gap between t_e and t_t .

4.5. Data-Processing Methods. **4.5.1. Kinetic Analyses.** Induction time, gas consumption, and gas consumption rate are the typical parameters obtained from hydrate formation kinetic measurements. According to the actual gas law, the amount of gas consumed in the hydrate formation process is calculated by eq 1,⁴¹

$$\Delta n_t = \frac{p_0 V}{Z_0 R T_0} - \frac{p_t V}{Z_t R T_t} \quad (1)$$

where Δn_t is the gas consumption from 0 to t (mol), V is the tank volume (mL), p_0 and p_t are pressures at the initial time and time t (MPa), T_0 and T_t are temperatures at the initial time and time t , respectively (K), Z_0 and Z_t are compression factors corresponding to T_0 and T_t , respectively, and R is the universal gas constant (8.3145 J/(mol·K)).

Among them, the compression factor can be calculated using the second virial coefficient, and the equations are shown in eqs 2 and 3,⁴²

$$B = 5.7400 \times 10 - 3.8829 \times 10^4/T + 4.2899 \times 10^5/T^2 - 1.4661 \times 10^9/T^3 \quad (2)$$

$$Z = 1 + \frac{Bp}{RT} \quad (3)$$

where B is the second virial coefficient (cm^3/mol).

4.5.2. Grey Relational Analysis. To reveal the relationship between the reaction time and gas consumption, grey relational analysis was applied to analyze the degree of correlation between the two.

According to the grey system theory,^{43–45} the parameters in the pure water system are taken as the reference sequence (X_0). The equation is shown in eq 4,

$$X_0(k) = \{X_0(1), X_0(2), X_0(3), \dots, X_0(n)\} \quad (4)$$

The comparison sequence (X_i) is constituted by the parameters of each system with RNS-A. The equation is shown in eq 5,

$$X_i(k) = \{X_i(1), X_i(2), X_i(3), \dots, X_i(n)\} \quad (5)$$

where $k = 1, 2, 3, \dots, n$, and n is the number of the experimental group, here, $n = 6$, denoting six concentrations of pure water, 5, 6, 8, 10, and 20 wt %; $i = 1, 2, 3, \dots, m$, and m is the number of comparison parameter, here, $m = 3$, denoting three kinds of times, induction time, growing time, and total time.

The original data are converted by a dimensionless method according to eq 6,

$$X'_i(k) = X_i(k)/X_0(k) \quad (6)$$

The absolute difference, $\Delta'_i(k)$, is calculated by the absolute gap between the comparison sequence X_i and the reference sequence X_0 at point k , as eq 7 shows.

$$\Delta'_i(k) = |X_0(k) - X_i(k)| \quad (7)$$

The correlation coefficient between the reference sequence X_0 and the comparison sequence X_i at point k is as shown in eq 8,

$$\varepsilon_i(k) = \frac{\min_i \min_k |X_0(k) - X_i(k)| + \rho \cdot \max_i \max_k |X_0(k) - X_i(k)|}{|X_0(k) - X_i(k)| + \rho \cdot \max_i \max_k |X_0(k) - X_i(k)|} \quad (8)$$

where ρ is the resolution coefficient (0.5).

Then, the degree of association, r , is calculated according to eq 9.

$$r_{0i} = \frac{1}{n} \sum_{k=1}^n \varepsilon_i(k) \quad (9)$$

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Notes

The authors declare no competing financial interest.

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