Review Article The Role of Matrix Metalloproteinases in Diabetic Wound Healing in relation to Photobiomodulation

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The integration of several cellular responses initiates the process of wound healing. Matrix Metalloproteinases (MMPs) play an integral role in wound healing. Their main function is degradation, by removal of damaged extracellular matrix (ECM) during the inflammatory phase, breakdown of the capillary basement membrane for angiogenesis and cell migration during the proliferation phase, and contraction and remodelling of tissue in the remodelling phase. For effective healing to occur, all wounds require a certain amount of these enzymes, which on the contrary could be very damaging at high concentrations causing excessive degradation and impaired wound healing. The imbalance in MMPs may increase the chronicity of a wound, a familiar problem seen in diabetic patients. The association of diabetes with impaired wound healing and other vascular complications is a serious public health issue. These may eventually lead to chronic foot ulcers and amputation. Low intensity laser irradiation (LILI) or photobiomodulation (PBM) is known to stimulate several wound healing processes; however, its role in matrix proteins and diabetic wound healing has not been fully investigated. This review focuses on the role of MMPs in diabetic wound healing and their interaction in PBM.

1. Introduction

Wound healing is a highly coordinated and carefully orchestrated process to promote proper healing [1, 2]. This process forms the ECM that enhances cellular processes in the wound, including cell adhesion, migration, and tissue remodelling. Matrix Metalloproteinases (MMPs) are zinc endopeptidases capable of degrading all components of the extracellular matrix (ECM) [3]. They participate in all the phases of wound healing by removing damaged proteins and temporary ECM during the inflammatory phase, breaking down the capillary basement membrane in the proliferation phase for angiogenesis and cell migration, as well as contracting and remodelling tissue in the remodelling phase. In addition, MMPs interplay the activity of a few growth factors and polymorphonuclear cells (PMNs) [4–10].

The MMP family comprises enzymes based on their substrate specificity, collagenases (MMP-1, MMP-8, MMP-13, and MMP-18), gelatinases (MMP-2 and MMP-9), stromelysins (MMP-3, MMP-10, and MMP-11), membrane type (MT) metalloproteinases (MMP-14, MMP-15, MMP-16, MMP-17, MMP-24, and MMP-25), matrilysins (MMP-7 and MMP-26), and others (MMP-12, MMP-19, MMP-20, MMP-21, MMP-23, MMP-27, and MMP-28) [11, 12]. They are comprised of three histidines conserved on the zinc site as well as a conserved methionine below the active site. Gelatinases (MMP-2 and MMP-9) and collagenases (MMP-1 and MMP-8) are particularly involved in the wound healing process. Tissue inhibitor of metalloproteinase (TIMP), of which there are four different subtypes (TIMP-1, TIMP-2, TIMP-3, and TIMP-4), interrupts the action of MMPs and thus regulates their action. TIMPs attach to alternate or active sites of MMPs and are capable of inhibiting all MMPs, although their affinity levels may differ [13]. Even though the importance of MMPs in wound healing cannot be underestimated, they can negatively affect healing if not present in the correct amount [14, 15]. Under normal conditions, the interaction of MMPs with growth factors and receptors is capable of maintaining the balance of the ECM. In such circumstances, the amount of MMP is decreased. However, predictors of healing will depend on the ratio between MMPs and TIMPs [16].

Within the ECM, an imbalance of these matrix proteins and their inhibitors may cause degradation of the matrix [17] and is associated with proteolytic enzymes distributed in several groups involved in disease development [18]. The interaction lowers growth factor and TIMP levels. It also increases the level of proteases and MMPs, especially MMP-1, MMP-2, MMP-8, and MMP-9, and proinflammatory cytokines such as interleukin-1 (IL-1) and tumour necrosis factor-alpha (TNF- α) released by neutrophils and macrophages. The effect is higher in chronic wounds as opposed to acute wounds. The imbalance is associated with abnormal wound healing conditions such as those in diabetes mellitus (DM) and other vascular complications [19, 20].

2. Mechanism of Matrix Metalloproteinases in Delayed Wound Healing

MMPs control the composition of ECM in various vascular and nonvascular inflammatory conditions [4, 5, 7-9, 21]. The ECM supports the vascular walls through the action of MMPs and their inhibitors. When an injury occurs, in the vascular system, it initiates an inflammatory reaction. The interaction of MMPs in disease pathogenesis is very critical. Some of the events include inflammation, activation of PMNs, and release of MMPs. Inflammatory cytokines such as IL1- α , IL1- β , IL-2, IL-17, C-reactive protein (CRP), insulin-like growth factor-1 (ILGF-1), and TGF- α stimulate the production of neutrophil gelatinase-associated lipocalin (NGAL). NGAL then activates MMP activities especially MMP-9 to form MMP-9/NGAL complex. This process occurs during the onset and healing phases of nonhealing venous ulcers [9, 22, 23]. This serves as a contributing factor to delayed wound healing even though it is not clear how many biochemical factors are involved.

Several studies have supported the central role played by NGAL in the activation of MMPs, especially MMP-9 in delayed wound healing [7-10, 22, 24]. Amato et al. [7] observed that MMP-1 and MMP-8 are pivotal in normal wound healing; however, overexpression was seen in patients with nonhealing chronic venous ulcers (CVUs) as compared to patients with healing ulcers undergoing normal and physical treatment. Similarly, Serra et al. [8] also found MMP and NGAL to be pivotal in arterial aneurysms due to high levels seen in patient's plasma. They concluded that the use of MMPs as molecular markers to prevent aneurysmal rupture was vital. In contrast, Karlsson et al. [25] did not find any correlation between MMP, NGAL, and inflammatory cytokines in their investigation. Most authors concluded and linked the association of MMP/NGAL to the pathophysiology of chronic wounds during the inflammatory and proliferative phases, whereby proteases appear to be significantly high, resulting in repeated migration of neutrophils, PMNs, and macrophages to the wound site. They concluded that overexpression of these proteins could have prolonged the inflammatory phase due to excessive collagenolytic properties and neutrophils resulting in delayed wound healing [8-10, 22, 24].

Furthermore, the production of MMPs and endotoxins, provoked by high metabolic activity of bacteria due to replication, has influenced the wound progression at different stages negatively. Infections are predominant with improper regulation of the immune defence system and a factor favouring bacterial growth, which may cause the wound to remain chronic, except for the eradication of the causative agent. Many studies have confirmed the major role of MMPs in infectious diseases [10, 26–28]. During unfavourable conditions, MMP degrades the ECM causing immune cells to migrate to initiate inflammatory response or proteolysis to eradicate the pathogen [29, 30]. The bacterial colonisation depends on several factors and patients with persistent infections are more predisposed to prolonged healing and care [31, 32].

3. Interaction of Matrix Metalloproteinases (MMPs) in Diabetic Wound Healing

Diabetes is a chronic metabolic disorder affecting millions of people globally [33]. Hyperglycaemia induces the majority of the micro- and macrovascular complications associated with DM [20] and increases MMP activity directly or indirectly through oxidative stress or advanced glycation end products (AGEs) (Figure 1) [14, 20, 34]. There are significantly higher levels of MMP in patients with metabolic syndrome as compared with normal individuals. This indicates that such patients may have high tendencies of developing other physiological problems [13]. The process of wound healing necessitates ECM degradation to be controlled; thus an imbalance between ECM formation and the degradation process could lead to the development of chronic ulcers or fibrosis [35]. Cellular and biochemical imbalances, tissue damage, or other disease conditions may present varied effects in the healing process. This also upsets the proteases, cytokines, and growth factors leading to an absence or delay of wound closure preventing successful skin repair [14, 15, 36]. There is a serious concern in the health sector regarding the unresolved problems associated with venous leg ulcers (VLU) and diabetes, which may not only be associated with pain, depreciated life, and/or eventual death, but also pose a severe economic burden [37-39]. Diabetic foot ulcer is common among any age group [40] and is predominantly seen in those over the age of 65 years with an occurrence of more than 3% [41] and an incidence of more than 1% of the world population [42].

Enzyme activity affected by hyperglycaemia disrupts the expression of MMPs in diabetes. An increased proteolytic environment provoked by an alteration in MMPs and TIMPs affects patients with diabetic ulcers [43]. Recent indications have shown the association of MMPs and their inhibitors with diabetic development and progression to be poor [43, 44]. Increased activities and levels of MMP-1, MMP-8, and MMP-9 have been identified in slow-to-heal wounds, with relatively low TIMP levels. Neutrophil derived MMP-8 may be up to 50-100 times higher as compared to normal wounds due to chronic inflammation [15, 45–47]. MMP-9 degrades fibronectin into fragments, which further activates MMP, cell migration, and proliferation. This provokes white blood cell infiltration, tissue damage, and continuous inflammation [48]. MMP-1, MMP-8, and MMP-9 are highly expressed in venous wounds in the absence of TIMPs [5, 7, 23]. Pirilä et al. [49] studied the expression of MMP-8 and MMP-26 in human cutaneous wounds and concluded that MMP-8 and



FIGURE 1: Mechanism of MMPs in chronic wounds. The development of chronic wounds results from delayed inflammation producing high levels of proteinases that destroy the essential elements of wound healing, growth factors, various receptors, and ECM proteins leading to increased MMPs, inflammatory cytokines, and reduced growth factors. The destruction of prohealing factors and ECM also reduces the ability for cells to migrate and proliferate. However as the wound begins to heal through various therapies, there would be increased mitogenic activity, reduced protease activity, and reduction of inflammatory cytokines.

MMP-26 could be associated with both acute and chronic wounds. Khattri and colleagues [13], after examining 388 subjects, found an overexpression of MMP-2 and MMP-9 in the serum of patients with metabolic syndrome as compared to controls. They concluded that altered expression of MMPs may provoke pathogenesis in several tissues [13, 50, 51]. Singh et al. [52] elucidated altered gene expression in MMP-9 as a cause of nonhealing diabetic ulcers, whereby diabetic wounds as compared to the controls had significant unmethylated MMP-9 gene promoter status that could be a result of impaired MMP expression in such wounds. According to Lobmann et al. [53], MMP-1, MMP-2, MMP-8, and MMP-9 were highly expressed in normal and chronic diabetic wounds with a decrease in TIMP-2. They concluded that this could be due to high proteolytic surroundings promoting poor healing in diabetes. Similarly, there was an overexpression of MMP-1 and MMP-9, as well as TIMP-1, in keratinocytes derived from foot ulcers in diabetic type 1 patients. This study supports the upregulation of certain MMPs and TIMPs in diabetic foot ulcers [54]. In addition, Menghini and colleagues [43] found MMP-9 to be increased and TIMP-3 to be decreased in diabetic type 2 patients. They suggested that the progression of ulcers could be due to a poor proteolytic environment. They concluded that new therapeutic strategies should focus on increasing the expression levels of TIMP-3. Dinh and colleagues [55] found increased expression of MMP-9, TNF- α , and other growth factors in diabetic foot ulcers and concluded that they could be linked with slow-toheal ulcers in diabetics and therefore a target for new therapeutic management. These studies have all ascertained that MMPs and TIMPs are elevated in chronic wounds; however, they may also play a role in determining the level of chronicity

[13]. Uccioli et al. [56] elucidated that chronicity is associated with an increase mainly in MMP-9 and MMP-8 and elastase activity that may eventually alter collagen synthesis and the release of growth factor and cytokines into the site of injury.

An increased activity of MMPs may initiate the development of diabetic peripheral arterial disease. Death and colleagues [57] found that hyperglycaemia affected the regulation of MMP/TIMP and increased the activities of MMP-1, MMP-2, and MMP-9 in vascular cells, stimulating the degradation of the ECM and causing an imbalance in diabetes. Chung and colleagues [34] elucidated that an increase in expression of MMP-2 and MMP-9 as well as protein expression of TIMP-1 may be a resulting factor in impaired wound healing and might provide an explanation for human arterial vasculature in type 2 DM.

4. Matrix Metalloproteinases (MMPs) and Current Therapeutic Strategies

Some of the pathophysiological events responsible for initiating CVUs are inflammation, PMN activation, and the release of proteases, especially MMPs. Many different compounds incorporated into conventional treatment have improved the quality of life in patients suffering from difficult-to-heal wounds. Compounds such as bioflavonoids, nutraceuticals, and glycoaminoglycans such as sulodexide (SDX) and cilostazol, together with conventional treatments, have improved healing and clinical conditions of patients [4, 5, 10]. In a recent study done by Serra and colleagues [10] on the effects of new nutraceutical substances, the researchers found that there was an overexpression of MMPs, NGAL, and inflammatory



FIGURE 2: The interaction of photobiomodulation (PBM) and the extracellular matrix (ECM). Light absorbed by photons in the electron transport chain within the mitochondria generates the production of Adenosine Triphosphate (ATP), Reactive Oxygen Species (ROS), and Nitric Oxide (NO). This evokes the release of transcription factors leading to the transcription of genes, which develop and enhance various cellular processes. Matrix Metalloproteinases (MMPs) are associated with changes that occur in the ECM. Under disease conditions, excess MMP is expressed which leads to the enhanced degradation of the ECM. When cells are exposed to PBM, there is an enhancement in the release of various cells including fibroblasts, growth factors, and adhesion molecules into the basement membrane leading to ECM remodelling.

cytokines in CVUs on treating patients conventionally. They introduced a nutraceutical substance together with compression therapy and surgical correction for 8 months and found a decrease in inflammatory cytokines with improved wound healing. Their findings explained that there is a correlation between the pathophysiology and impaired wound healing probably due to the overexpression of MMPs, NGAL, and inflammatory cytokines. Their findings are consistent with other studies using glycoaminoglycans such as sulodexide (SDX) and cilostazol [4, 5]. The authors concluded that inhibition could be a possible therapeutic intervention. In addition to these compounds, the use of biomarkers would be vital when clinical signs are misleading [58, 59]. However, mimicking the action of MMP inhibitors could possibly assist in managing infected wounds or wounds activated through MMP or cytokine release [4, 5, 10, 60].

5. Matrix Metalloproteinases (MMPs), Wound Healing, and Photobiomodulation (PBM)

PBM is a noninvasive form of light therapy for wound healing, whereby several biological, chemical, and cellular processes are stimulated to speed up healing (Figure 2). The adequacy of this therapy is specific to a spectral range of 500–1,100 nm and power output ranging within 10–200 mW [61]. Investigations carried out found PBM to enhance wound healing [62–64]. Studies have also established its effect on wounded cell models *in vitro* using various wavelengths and dosages [65–67]. In addition, studies performed on mice, rats, and humans showed a stimulatory effect of PBM in diabetic wound healing [62, 63, 68–71].

The application of PBM in altering the gene expression in cell cultures is evident [72, 73]. PBM is able to alter the expression of MMPs in diabetic wounds and enhance collagen production [64, 74, 75]. However, experiments done on human skin fibroblasts demonstrated variations in gene expression [75, 76]. Ayuk and colleagues [75] found that PBM altered 49 genes involved in the ECM in vitro, with genes in a diabetic wounded cell model mostly downregulated, among which were MMP-1, MMP-2, MMP-8, MMP-12, MMP-14, and MMP-16. PBM in another study (1 or 3 J/cm² with 810 nm) was shown to reduce the gene expression of MMP-3, MMP-9, and MMP-13 in Achilles tendons of rats as well as prostaglandin, implying that it was able to reduce tendon inflammation and preserve elasticity and resistance [77]. Casalechi and colleagues [78] investigated a nonconventional treatment for tendinopathies and found that PBM (780 nm; 107 mW/cm²; 7.5 J/cm²) alongside various inflammatory treatments modulated mRNA gene expression of MMP-1, MMP-13, and IL-10 and vascular endothelial growth factor (VGEF) in a Wistar rat model. Cury et al. [79] conducted an experiment using sixty male Wistar rats which were irradiated at 660 and 780 nm with a fluence of 30 and 40 J/cm² and found a decrease in MMP-2 activity and increased expression in angiogenesis markers such as VEGF. Therefore, PBM was associated with promoting healing of skin flaps in a dose dependent manner. PBM also plays a role in regulating inflammatory cytokines. In a study conducted by Laraia and colleagues [80] to analyze the role of PBM (660 nm; 100 mW) on protein expression levels in rat Achilles tendons with their main focus on the inflammatory mediators, they found that the group treated with lasers showed positive responses on IL-6 and IL-10 after 6, 24, and 72 h compared to the control. They concluded that PBM plays a significant role in regulating inflammatory cytokines after injury.

Ye et al. [81] used three different dosages (0, 0.6, 1.5, and 2.5 J/cm^2) at 1064 nm on the dorsal skin of Sprague-Dawley rats and showed an increase in collagen content, as well as the upregulation of the gene expression of collagen type 1 (Col-1) and collagen type 3 (Col-3) and TIMP-1 and TIMP-2, and a significant decrease in MMP-2 and MMP-3. They concluded in their study that irradiation at 1064 nm could markedly increase collagen production and prevent degradation and activate the Erk1/2 and JNK-MAPK pathway. In addition, Lee et al. [82] found that long-pulsed Nd:YAG laser (1064 nm) decreased the expression of MMP-1 at low energy dosages; meanwhile there was an increase in dermal collagen and transforming growth factor-beta (TGF- β) with no significant difference in the expression of TIMP-1 in hairless mice. A similar study using cultured human skin fibroblasts after two successive irradiations with 1.5 J/cm^2 at wavelengths of 1064 and 532 nm significantly increased Col-1 and Col-3 at 24 and 48 h after irradiation. This study also found an increase in the expression of TIMP-1 and TIMP-2 and a decrease in MMP-1 and MMP-2. It ascertained that 1064 nm was more effective as compared to 532 nm in promoting molecular activities [83]. Kim et al. [84] found PBM (780 nm; 1.75 W/cm²; 5 J/cm²) to modulate the gene expression of five MMPs (MMP-1, MMP-2, MMP-8, MMP-9, and MMP-13) in the periodontal ligament of Sprague-Dawley rats and inhibited immunoreactivity of TIMP-1. Similarly, PBM (780 nm; 125 mW/cm²; 0,

0.5, 1.0, 2.5, 5.0, and 20 J/cm²) with different energy densities modulated the gene expression of MMP-2 and MMP-9 as well as the oxidative metabolism of the masseter muscle in rats, indicating possible ECM remodelling [85].

The biostimulatory effect of PBM in the near infrared (NIR) range modulates wound healing events in various cells. Gavish et al. [86] found that PBM (780 nm, 1 or 2 J/cm²) increased collagen activity twofold, increased MMP-2 activity, upregulated MMP-1 and TIMP-2 expression, and downregulated MMP-2 and IL-1 β . Their findings may be of therapeutic importance in situations with depleted smooth cells, weakened ECM, and increased proinflammatory markers as major pathological components. Migliario et al. [87] observed a decrease in MMP-9 in normal keratinocytes exposed to laser irradiation at 635.2 nm and a fluence of 82.5 or 112.4 J/cm², a very high dosage, which could influence the production of enzymes during wound healing and probably explain the decrease in cell proliferation. In addition, irradiation using a diode laser prototype (780±3 nm, 40 mW) at different doses of 0.5, 1.5, 3.5, and 7 J/cm² over 24 h showed that a dose range of 0.5–3 J/cm² promotes a biostimulatory effect on keratinocytes [88]. Luo and colleagues [89] found that intense pulse light (IPL) irradiation had a time dependent effect on mouse skin at 2 weeks up to 8 weeks with MMP-1 and MMP-2 showing a decrease in mRNA expression after irradiation, and the authors concluded that irradiation with IPL modulated collagen synthesis as well as ECM degradation.

After treating patients with laser therapy (660 nm; power output 25 mW; dose 6.2 J/cm²), Oton-Leite and colleagues [90] found that there was a trend in the reduction levels of IL-1 β , IL-10, TNF- α , TGF- β , MMP-2/TIMP-2, and MMP-9/TIMP-2 as compared to the untreated group, even though there was no significant difference. They concluded that it was effective in reducing pain, inflammation, and repair. Silva and colleagues [91] performed a randomized preliminary study to evaluate PBM using saliva and blood from haematopoietic stem cell transplantation patients and found a positive response in MMP-2 in the saliva and IL-10 in the blood. They found out that PBM is clinically effective in reducing chemotherapy induced oral mucositis; however, there was no mechanism linked with inflammatory cytokines, growth factors, and MMPs. Furthermore, some researchers performed a controlled randomized clinical trial and found that after treatment for chronic periodontitis there was a significant difference in the levels of MMP-1, MMP-8, and TIMP-1 between the various groups as compared with the controls. They found a significant decrease in the total amount of IL-1 β , IL-6, MMP-1, MMP-8, and TIMP-1 with MMP-8 significantly affected. The researchers concluded that MMP-8 has the most effective impact for use as an adjunct laser treatment on nonsurgical periodontal therapy [92]. In addition, Qadri et al. [93] used two wavelengths, 636 and 830 nm, on patients with moderate gingival periodontitis and their results showed a decrease in the total amount of MMP-8 as compared to the placebo. From their study, they found PBM to ameliorate periodontal gingival inflammation. In a similar study, Qadri and colleagues [94] found that scale and root application together with Nd: YAG laser irradiation significantly reduced IL-1 β and MMP-8 and therefore improved clinical signs linked with periodontal inflammation as compared to the treatment alone.

PBM is known for its stimulatory effects and promotes MMP activity and gene expression; hence maintaining a dynamic balance between the proteolytic activity and degradation could be a target for therapeutic advancement. However, its effect on various matrix proteins still needs to be further understood. Suggestions of further research in TIMPs and MMPs would be very important for the selection of these as indicators of early disease, in order to deduce standard protocols and improve clinical regimen [95].

6. Conclusion

This review covers the relationship between MMPs and their inhibitors in diabetic wound healing in relation to PBM and the ECM in both human and animal models, *in vivo* and *in vitro*. MMPs play a significant role in tissue remodelling. Given the fact that MMPs are known for degradation of the ECM and have been shown to be upregulated in most pathologies, they have been recognised as potential targets for new therapeutic measures. PBM can significantly restore the balance between the ECM and degradation enzymes. However, it is necessary to further elucidate the relationship between PBM, diabetic wound healing, and MMPs.

Competing Interests

The authors have no competing financial interests.

References

- S. Maxson, E. A. Lopez, D. Yoo, A. Danilkovitch-Miagkova, and M. A. LeRoux, "Concise review: role of mesenchymal stem cells in wound repair," *Stem Cells Translational Medicine*, vol. 1, no. 2, pp. 142–149, 2012.
- [2] J. M. Reinke and H. Sorg, "Wound repair and regeneration," *European Surgical Research*, vol. 49, no. 1, pp. 35–43, 2012.
- [3] K. Vitlianova, J. Georgieva, M. Milanova, and S. Tzonev, "Blood pressure control predicts plasma matrix metalloproteinase-9 in diabetes mellitus type II," *Archives of Medical Science*, vol. 11, no. 1, pp. 85–91, 2015.
- [4] S. de Franciscis, L. Gallelli, L. Battaglia et al., "Cilostazol prevents foot ulcers in diabetic patients with peripheral vascular disease," *International Wound Journal*, vol. 12, no. 3, pp. 250– 253, 2015.
- [5] R. Serra, G. Buffone, G. Costanzo et al., "Altered metalloproteinase-9 expression as least common denominator between varicocele, inguinal hernia, and chronic venous disorders," *Annals of Vascular Surgery*, vol. 28, no. 3, pp. 705–709, 2014.
- [6] F. Sabino and U. A. D. Keller, "Matrix metalloproteinases in impaired wound healing," *Journal of Metalloproteinases In Medicine*, vol. 2, pp. 1–87, 2015.
- [7] B. Amato, G. Coretti, R. Compagna et al., "Role of matrix metalloproteinases in non-healing venous ulcers," *International Wound Journal*, vol. 12, no. 6, pp. 641–645, 2015.
- [8] R. Serra, G. Buffone, V. Molinari et al., "Low molecular weight heparin improves healing of chronic venous ulcers especially in

the elderly," *International Wound Journal*, vol. 12, no. 2, pp. 150–153, 2015.

- [9] R. Serra, L. Gallelli, R. Grande et al., "Hemorrhoids and matrix metalloproteinases: a multicenter study on the predictive role of biomarkers," *Surgery*, vol. 159, no. 2, pp. 487–494, 2016.
- [10] R. Serra, R. Grande, G. Buffone et al., "Extracellular matrix assessment of infected chronic venous leg ulcers: role of metalloproteinases and inflammatory cytokines," *International Wound Journal*, vol. 13, no. 1, pp. 53–58, 2016.
- [11] C. M. Overall, "Matrix metalloproteinase substrate binding domains, modules and exosites. Overview and experimental strategies," *Methods in Molecular Biology*, vol. 151, pp. 79–120, 2001.
- [12] R. Visse and H. Nagase, "Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry," *Circulation Research*, vol. 92, no. 8, pp. 827–839, 2003.
- [13] S. Khattri, R. Mandal, K. Pant et al., "Significance of impaired serum gelatinases activities in metabolic syndrome," *Toxicology International*, vol. 21, no. 1, pp. 107–111, 2014.
- [14] V. L. Martins, M. Caley, and E. A. O'Toole, "Matrix metalloproteinases and epidermal wound repair," *Cell and Tissue Research*, vol. 351, no. 2, pp. 255–268, 2013.
- [15] M. N. Menke, N. B. Menke, C. H. Boardman, and R. F. Diegelmann, "Biologic therapeutics and molecular profiling to optimize wound healing," *Gynecologic Oncology*, vol. 111, no. 2, supplement, pp. S87–S91, 2008.
- [16] M. Muller, C. Trocme, B. Lardy, F. Morel, S. Halimi, and P. Y. Benhamou, "Matrix metalloproteinases and diabetic foot ulcers: the ratio of MMP-1 to TIMP-1 is a predictor of wound healing," *Diabetic Medicine*, vol. 25, no. 4, pp. 419–426, 2008.
- [17] C. M. Overall and C. López-Otín, "Strategies for MMP inhibition in cancer: innovations for the post-trial era," *Nature Reviews Cancer*, vol. 2, no. 9, pp. 657–672, 2002.
- [18] O. Lenz, S. J. Elliot, and W. G. Stetler-Stevenson, "Matrix metalloproteinases in renal development and disease," *Journal* of the American Society of Nephrology, vol. 11, no. 3, pp. 574–581, 2000.
- [19] S. Gharagozlian, K. Svennevig, H.-J. Bangstad, J.-O. Winberg, and S. O. Kolset, "Matrix metalloproteinases in subjects with type 1 diabetes," *BMC Clinical Pathology*, vol. 9, article 7, 2009.
- [20] S. Guo and L. A. DiPietro, "Factors affecting wound healing," *Journal of Dental Research*, vol. 89, no. 3, pp. 219–229, 2010.
- [21] R. Serra, G. Buffone, A. de Franciscis et al., "Skin grafting followed by low-molecular-weight heparin long-term therapy in chronic venous leg ulcers," *Annals of Vascular Surgery*, vol. 26, no. 2, pp. 190–197, 2012.
- [22] S. de Franciscis, P. Mastroroberto, L. Gallelli, G. Buffone, R. Montemurro, and R. Serra, "Increased plasma levels of metalloproteinase-9 and neutrophil gelatinase-associated lipocalin in a rare case of multiple artery aneurysm," *Annals of Vascular Surgery*, vol. 27, no. 8, pp. 1185.e5–1185.e7, 2013.
- [23] R. Serra, G. Buffone, D. Falcone et al., "Chronic venous leg ulcers are associated with high levels of metalloproteinases-9 and neutrophil gelatinase-associated lipocalin," *Wound Repair and Regeneration*, vol. 21, no. 3, pp. 395–401, 2013.
- [24] M. T. Busceti, R. Grande, B. Amato et al., "Pulmonary embolism, metalloproteinases and neutrophil gelatinase associated lipocalin," *Acta Phlebologica*, vol. 14, no. 3, pp. 115–121, 2013.

- [25] L. Karlsson, D. Bergqvist, J. Lindbäck, and H. Pärsson, "Expansion of small-diameter abdominal aortic aneurysms is not reflected by the release of inflammatory mediators IL-6, MMP-9 and CRP in plasma," *European Journal of Vascular and Endovascular Surgery*, vol. 37, no. 4, pp. 420–424, 2009.
- [26] S.-I. Miyoshi and S. Shinoda, "Microbial metalloproteases and pathogenesis," *Microbes and Infection*, vol. 2, no. 1, pp. 91–98, 2000.
- [27] Z. Cao, C. Li, and G. Zhu, "Inhibitory effects of baicalin on IL-1induced MMP-1/TIMP-1 and its stimulated effect on Collagen-I production in human periodontal ligament cells," *European Journal of Pharmacology*, vol. 641, no. 1, pp. 1–6, 2010.
- [28] N. Geurts, G. Opdenakker, and P. E. Van den Steen, "Matrix metalloproteinases as therapeutic targets in protozoan parasitic infections," *Pharmacology and Therapeutics*, vol. 133, no. 3, pp. 257–279, 2012.
- [29] G. Jantet, "Chronic venous insufficiency: worldwide results of the RELIEF study. Reflux assessment and quality of life improvement with micronized Flavonoids," *Angiology*, vol. 53, no. 3, pp. 245–256, 2002.
- [30] S. L. Dholakiya and K. E. Benzeroual, "Protective effect of diosmin on LPS-induced apoptosis in PC12 cells and inhibition of TNF-α expression," *Toxicology in Vitro*, vol. 25, no. 5, pp. 1039–1044, 2011.
- [31] P. Gloviczki, A. J. Comerota, M. C. Dalsing et al., "The care of patients with varicose veins and associated chronic venous diseases: clinical practice guidelines of the Society for Vascular Surgery and the American Venous Forum," *Journal of Vascular Surgery*, vol. 53, no. 5, supplement, pp. 2S–48S, 2011.
- [32] P. Gloviczki and M. L. Gloviczki, "Guidelines for the management of varicose veins," *Phlebology*, vol. 27, supplement 1, pp. 2–9, 2012.
- [33] International Diabetes Federation, *Diabetes Atlas*, International Diabetes Federation, Brussels, Belgium, 6th edition, 2014, http://www.idf.org/diabetesatlas.
- [34] A. W. Y. Chung, Y. N. Hsiang, L. A. Matzke, B. M. McManus, C. Van Breemen, and E. B. Okon, "Reduced expression of vascular endothelial growth factor paralleled with the increased angiostatin expression resulting from the upregulated activities of matrix metalloproteinase-2 and -9 in human type 2 diabetic arterial vasculature," *Circulation Research*, vol. 99, no. 2, pp. 140– 148, 2006.
- [35] Z. Li, S. Guo, F. Yao, Y. Zhang, and T. Li, "Increased ratio of serum matrix metalloproteinase-9 against TIMP-1 predicts poor wound healing in diabetic foot ulcers," *Journal of Diabetes and Its Complications*, vol. 27, no. 4, pp. 380–382, 2013.
- [36] S. M. McCarty and S. L. Percival, "Proteases and delayed wound healing," *Advances in Wound Care*, vol. 2, no. 8, pp. 438–447, 2013.
- [37] P. Price, K. Fogh, C. Glynn, D. L. Krasner, J. Osterbrink, and R. G. Sibbald, "Managing painful chronic wounds: the wound pain management model," *International Wound Journal*, vol. 4, supplement 1, pp. 4–15, 2007.
- [38] S. Ryan, C. Eager, and R. G. Sibbald, "Venous leg ulcer pain," Ostomy/Wound Management, vol. 49, no. 4, pp. 16–23, 2003.
- [39] K. Soon and C. Acton, "Pain-induced stress: a barrier to wound healing," *Wounds UK*, vol. 2, no. 4, pp. 92–101, 2006.
- [40] J. Kantor and D. J. Margolis, "Epidemiology: leg ulcers," in A Problem-Based Learning Approach, M. J. Morison, C. J. Moffatt, and P. J. Franks, Eds., pp. 65–73, Mosby, London, UK, 2007.

- [41] J. R. Mekkes, M. A. M. Loots, A. C. Van Der Wal, and J. D. Bos, "Causes, investigation and treatment of leg ulceration," *British Journal of Dermatology*, vol. 148, no. 3, pp. 388–401, 2003.
- [42] I. Anderson, "Aetiology, assessment and management of leg ulcers," Wound Essentials, vol. 1, pp. 20–36, 2006.
- [43] R. Menghini, L. Uccioli, E. Vainieri et al., "Expression of tissue inhibitor of metalloprotease 3 is reduced in ischemic but not neuropathic ulcers from patients with type 2 diabetes mellitus," *Acta Diabetologica*, vol. 50, no. 6, pp. 907–910, 2013.
- [44] E. Hopps, R. Lo Presti, M. Montana et al., "Gelatinases and their tissue inhibitors in a group of subjects with metabolic syndrome," *Journal of Investigative Medicine*, vol. 61, no. 6, pp. 978–983, 2013.
- [45] D. G. Armstrong and E. B. Jude, "The role of matrix metalloproteinases in wound healing," *Journal of the American Podiatric Medical Association*, vol. 92, no. 1, pp. 12–18, 2002.
- [46] V. Falanga, "The chronic wound: impaired healing and solutions in the context of wound bed preparation," *Blood Cells*, *Molecules, and Diseases*, vol. 32, no. 1, pp. 88–94, 2004.
- [47] D. J. Gibson and G. S. Schultz, "Molecular wound assessments: matrix metalloproteinases," *Advances in Wound Care*, vol. 2, no. 1, pp. 18–23, 2013.
- [48] A. D. Widgerow, "Chronic wound fluid-thinking outside the box," Wound Repair and Regeneration, vol. 19, no. 3, pp. 287–291, 2011.
- [49] E. Pirilä, J. T. Korpi, T. Korkiamäki et al., "Collagenase-2 (MMP-8) and matrilysin-2 (MMP-26) expression in human wounds of different etiologies," *Wound Repair and Regeneration*, vol. 15, no. 1, pp. 47–57, 2007.
- [50] J. W. Burrow, J. A. Koch, H.-H. Chuang, W. Zhong, D. D. Dean, and V. L. Sylvia, "Nitric oxide donors selectively reduce the expression of matrix metalloproteinases-8 and -9 by human diabetic skin fibroblasts," *Journal of Surgical Research*, vol. 140, no. 1, pp. 90–98, 2007.
- [51] S. Das and A. Maiti, "Matrix metalloproteinases in subjects with type 2 diabetes mellitus: pattern of MMP-2 and MMP-9 profile in diabetes mellitus Type-2 patients," *American International Journal of Research in Formal, Applied and Natural Sciences*, vol. 3, no. 1, pp. 57–60, 2013.
- [52] K. Singh, N. K. Agrawal, S. K. Gupta, G. Mohan, S. Chaturvedi, and K. Singh, "Differential expression of matrix metalloproteinase-9 gene in wounds of type 2 diabetes mellitus cases with susceptible –1562C > T genotypes and wound severity," *International Journal of Lower Extremity Wounds*, vol. 13, no. 2, pp. 94–102, 2014.
- [53] R. Lobmann, A. Ambrosch, G. Schultz, K. Waldmann, S. Schiweck, and H. Lehnert, "Expression of matrixmetalloproteinases and their inhibitors in the wounds of diabetic and non-diabetic patients," *Diabetologia*, vol. 45, no. 7, pp. 1011–1016, 2002.
- [54] N. López-López, I. González-Curiel, M. B. Treviño-Santa Cruz et al., "Expression and vitamin D-mediated regulation of matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMPs) in healthy skin and in diabetic foot ulcers," *Archives of Dermatological Research*, vol. 306, no. 9, pp. 809–821, 2014.
- [55] T. Dinh, F. Tecilazich, A. Kafanas et al., "Mechanisms involved in the development and healing of diabetic foot ulceration," *Diabetes*, vol. 61, no. 11, pp. 2937–2947, 2012.
- [56] L. Uccioli, V. Izzo, M. Meloni, E. Vainieri, V. Ruotolo, and L. Giurato, "Non-healing foot ulcers in diabetic patients: general

and local interfering conditions and management options with advanced wound dressings," *Journal of Wound Care*, vol. 24, supplement 4, pp. 35–42, 2015.

- [57] A. K. Death, E. J. Fisher, K. C. Y. McGrath, and D. K. Yue, "High glucose alters matrix metalloproteinase expression in two key vascular cells: potential impact on atherosclerosis in diabetes," *Atherosclerosis*, vol. 168, no. 2, pp. 263–269, 2003.
- [58] M. Massara, G. De Caridi, R. Serra et al., "The role of procalcitonin as a marker of diabetic foot ulcer infection," *International Wound Journal*, 2015.
- [59] N. J. Jafari, M. S. Firouzabadi, M. Izadi, M. S. S. Firouzabadi, and A. Saburi, "Can procalcitonin be an accurate diagnostic marker for the classification of diabetic foot ulcers?" *International Journal of Endocrinology and Metabolism*, vol. 12, no. 1, Article ID e13376, 2014.
- [60] S. de Franciscis, G. De Sarro, P. Longo et al., "Hyperhomocysteinaemia and chronic venous ulcers," *International Wound Journal*, vol. 12, no. 1, pp. 22–26, 2015.
- [61] D. H. Evans and H. Abrahamse, "A review of laboratory-based methods to investigate second messengers in low-level laser therapy (LLLT)," *Medical Laser Application*, vol. 24, no. 3, pp. 201–215, 2009.
- [62] T.-Y. Chung, P. V. Peplow, and G. D. Baxter, "Laser photobiomodulation of wound healing in diabetic and non-diabetic mice: effects in splinted and unsplinted wounds," *Photomedicine and Laser Surgery*, vol. 28, no. 2, pp. 251–261, 2010.
- [63] P. V. Peplow, T.-Y. Chung, B. Ryan, and G. D. Baxter, "Laser photobiostimulation of wound healing: reciprocity of irradiance and exposure time on energy density for splinted wounds in diabetic mice," *Lasers in Surgery and Medicine*, vol. 43, no. 8, pp. 843–850, 2011.
- [64] F. L. Primo, L. B. de Paula, M. P. de Siqueira-Moura, and A. C. Tedesco, "Photobiostimulation on wound healing treatment by ClAlPc-nanoemulsion from a multiple-wavelength portable light source on a 3D-human stem cell dermal equivalent," *Current Medicinal Chemistry*, vol. 19, no. 30, pp. 5157–5163, 2012.
- [65] S. M. Ayuk, N. N. Houreld, and H. Abrahamse, "Collagen production in diabetic wounded fibroblasts in response to lowintensity laser irradiation at 660 nm," *Diabetes Technology and Therapeutics*, vol. 14, no. 12, pp. 1110–1117, 2012.
- [66] M. S. Dawood and S. D. Salman, "Low level diode laser accelerates wound healing," *Lasers in Medical Science*, vol. 28, no. 3, pp. 941–945, 2013.
- [67] N. Houreld and H. Abrahamse, "Low-intensity laser irradiation stimulates wound healing in diabetic wounded fibroblast cells (WS1)," *Diabetes Technology and Therapeutics*, vol. 12, no. 12, pp. 971–978, 2010.
- [68] P. Lau, N. Bidin, G. Krishnan et al., "Photobiostimulation effect on diabetic wound at different power density of near infrared laser," *Journal of Photochemistry and Photobiology B: Biology*, vol. 151, pp. 201–207, 2015.
- [69] R. Fekrazad, A. Mirmoezzi, K. A. Kalhori, and P. Arany, "The effect of red, green and blue lasers on healing of oral wounds in diabetic rats," *Journal of Photochemistry and Photobiology B: Biology*, vol. 148, pp. 242–245, 2015.
- [70] P. V. Peplow and G. D. Baxter, "Defining a therapeutic window for laser irradiation (810 nm) applied to the inguinal region to ameliorate diabetes in diabetic mice," *Photomedicine and Laser Surgery*, vol. 32, no. 9, pp. 500–504, 2014.
- [71] M. C. Feitosa, A. F. Carvalho, V. C. Feitosa, I. M. Coelho, R. A. Oliveira, and E. Â. Arisawa, "Effects of the Low-Level Laser

Therapy (LLLT) in the process of healing diabetic foot ulcers," *Acta Cirurgica Brasileira*, vol. 30, no. 12, pp. 852–857, 2015.

- [72] N. N. Houreld, S. M. Ayuk, and H. Abrahamse, "Expression of genes in normal fibroblast cells (WS1) in response to irradiation at 660 nm," *Journal of Photochemistry and Photobiology B: Biology*, vol. 130, pp. 146–152, 2014.
- [73] N. C. Rodrigues, R. Brunelli, H. S. S. De Araújo, N. A. Parizotto, and A. C. M. Renno, "Low-level laser therapy (LLLT) (660 nm) alters gene expression during muscle healing in rats," *Journal of Photochemistry and Photobiology B: Biology*, vol. 120, pp. 29–35, 2013.
- [74] A. Aparecida Da Silva, E. C. P. Leal-Junior, A. C. A. Alves et al., "Wound-healing effects of low-level laser therapy in diabetic rats involve the modulation of MMP-2 and MMP-9 and the redistribution of collagen types I and III," *Journal of Cosmetic and Laser Therapy*, vol. 15, no. 4, pp. 210–216, 2013.
- [75] S. M. Ayuk, N. N. Houreld, and H. Abrahamse, "Laser irradiation alters the expression profile of genes involved in the extracellular matrix in vitro," *International Journal of Photoenergy*, vol. 2014, Article ID 604518, 17 pages, 2014.
- [76] H. Y. Chang, J.-T. Chi, S. Dudoit et al., "Diversity, topographic differentiation, and positional memory in human fibroblasts," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 20, pp. 12877–12882, 2002.
- [77] R. L. Marcos, E. C. P. Leal-Junior, G. Arnold et al., "Low-level laser therapy in collagenase-induced Achilles tendinitis in rats: analyses of biochemical and biomechanical aspects," *Journal of Orthopaedic Research*, vol. 30, no. 12, pp. 1945–1951, 2012.
- [78] H. L. Casalechi, E. C. P. Leal-Junior, M. Xavier et al., "Low-level laser therapy in experimental model of collagenase-induced tendinitis in rats: effects in acute and chronic inflammatory phases," *Lasers in Medical Science*, vol. 28, no. 3, pp. 989–995, 2013.
- [79] V. Cury, A. I. S. Moretti, L. Assis et al., "Low level laser therapy increases angiogenesis in a model of ischemic skin flap in rats mediated by VEGF, HIF-1α and MMP-2," *Journal of Photochemistry and Photobiology B: Biology*, vol. 125, pp. 164– 170, 2013.
- [80] E. M. S. Laraia, I. S. Silva, D. M. Pereira et al., "Effect of lowlevel laser therapy (660 nm) on acute inflammation induced by tenotomy of achilles tendon in rats," *Photochemistry and Photobiology*, vol. 88, no. 6, pp. 1546–1550, 2012.
- [81] X. Ye, L. Wang, Y. Dang, B. Liu, and D. Zhao, "Investigation of the 1064 nm Q-switched Nd:YAG laser on collagen expression in an animal model," *Photomedicine and Laser Surgery*, vol. 30, no. 10, pp. 604–609, 2012.
- [82] Y. B. Lee, N. H. Kang, Y. S. Eun et al., "Effects of long-pulsed 1,064-nm neodymium-doped yttrium aluminum garnet laser on dermal collagen remodeling in hairless mice," *Dermatologic Surgery*, vol. 38, no. 7, part 1, pp. 985–992, 2012.
- [83] Y. Dang, X. Ye, Y. Weng, Z. Tong, and Q. Ren, "Effects of the 532-nm and 1,064-nm Q-switched Nd:YAG lasers on collagen turnover of cultured human skin fibroblasts: a comparative study," *Lasers in Medical Science*, vol. 25, no. 5, pp. 719–726, 2010.
- [84] S.-J. Kim, Y.-G. Kang, J.-H. Park, E.-C. Kim, and Y.-G. Park, "Effects of low-intensity laser therapy on periodontal tissue remodeling during relapse and retention of orthodontically moved teeth," *Lasers in Medical Science*, vol. 28, no. 1, pp. 325– 333, 2013.
- [85] F. J. Diasé, J. P. M. Issa, F. T. M. de Carvalho Vicentini et al., "Effects of low-level laser therapy on the oxidative metabolism"

and matrix proteins in the rat masseter muscle," *Photomedicine and Laser Surgery*, vol. 29, no. 10, pp. 677–684, 2011.

- [86] L. Gavish, L. Perez, and S. D. Gertz, "Low-level laser irradiation modulates matrix metalloproteinase activity and gene expression in porcine aortic smooth muscle cells," *Lasers in Surgery and Medicine*, vol. 38, no. 8, pp. 779–786, 2006.
- [87] M. Migliario, M. Rizzi, V. Rocchetti, M. Cannas, and F. Renò, "In vitro toxicity of photodynamic antimicrobial chemotherapy on human keratinocytes proliferation," *Lasers in Medical Science*, vol. 28, no. 2, pp. 565–569, 2013.
- [88] F. G. Basso, C. F. Oliveira, C. Kurachi, J. Hebling, and C. A. D. S. Costa, "Biostimulatory effect of low-level laser therapy on keratinocytes in vitro," *Lasers in Medical Science*, vol. 28, no. 2, pp. 367–374, 2013.
- [89] D. Luo, Y. Cao, D. Wu, Y. Xu, B. Chen, and Z. Xue, "Impact of intense pulse light irradiation on BALB/c mouse skin-in vivo study on collagens, matrix metalloproteinases and vascular endothelial growth factor," *Lasers in Medical Science*, vol. 24, no. 1, pp. 101–108, 2009.
- [90] A. F. Oton-Leite, G. B. L. Silva, M. O. Morais et al., "Effect of low-level laser therapy on chemoradiotherapy-induced oral mucositis and salivary inflammatory mediators in head and neck cancer patients," *Lasers in Surgery and Medicine*, vol. 47, no. 4, pp. 296–305, 2015.
- [91] G. B. L. Silva, N. T. Sacono, A. F. Othon-Leite et al., "Effect of low-level laser therapy on inflammatory mediator release during chemotherapy-induced oral mucositis: a randomized preliminary study," *Lasers in Medical Science*, vol. 30, no. 1, pp. 117–126, 2015.
- [92] M. Saglam, A. Kantarci, N. Dundar, and S. S. Hakki, "Clinical and biochemical effects of diode laser as an adjunct to nonsurgical treatment of chronic periodontitis: a randomized, controlled clinical trial," *Lasers in Medical Science*, vol. 29, no. 1, pp. 37–46, 2014.
- [93] T. Qadri, L. Miranda, J. Tunér, and A. Gustafsson, "The shortterm effects of low-level lasers as adjunct therapy in the treatment of periodontal inflammation," *Journal of Clinical Periodontology*, vol. 32, no. 7, pp. 714–719, 2005.
- [94] T. Qadri, P. Poddani, F. Javed, J. Tuner, and A. Gustafsson, "A short-term evaluation of Nd:YAG laser as an adjunct to scaling and root planing in the treatment of periodontal inflammation," *Journal of Periodontology*, vol. 81, no. 8, pp. 1161–1166, 2010.
- [95] C. Symeonidis, E. Papakonstantinou, A. Galli et al., "Matrix metalloproteinase (MMP-2, -9) and tissue inhibitor (TIMP-1, -2) activity in tear samples of pediatric type 1 diabetic patients: MMPs in tear samples from type 1 diabetes," *Graefe's Archive for Clinical and Experimental Ophthalmology*, vol. 251, no. 3, pp. 741–749, 2013.