TransLnc: a comprehensive resource for translatable IncRNAs extends immunopeptidome

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Received August 08, 2021; Revised September 05, 2021; Editorial Decision September 10, 2021; Accepted September 10, 2021

ABSTRACT

LncRNAs are not only well-known as non-coding elements, but also serve as templates for peptide translation, playing important roles in fundamental cellular processes and diseases. Here, we describe a database, TransLnc (http://bio-bigdata.hrbmu.edu. cn/TransLnc/), which aims to provide comprehensive experimentally supported and predicted IncRNA peptides in multiple species. TransLnc currently documents approximate 583 840 peptides encoded by 33 094 IncRNAs. Six types of direct and indirect evidences supporting the coding potential of IncR-NAs were integrated, and 65.28% peptides entries were with at least one type of evidence. Considering the strong tissue-specific expression of IncR-NAs, TransLnc allows users to access IncRNA peptides in any of the 34 tissues involved in. In addition, both the unique characteristic and homology relationship were also predicted and provided. Importantly. TransLnc provides computationally predicted tumour neoantigens from peptides encoded by IncR-NAs, which would provide novel insights into cancer immunotherapy. There were 220 791 and 237 915 candidate neoantigens binding by major histocompatibility complex (MHC) class I or II molecules, respectively. Several flexible tools were developed to aid retrieve and analyse, particularly IncRNAs tissue expression patterns, clinical relevance across cancer types. TransLnc will serve as a valuable resource for investigating the translation capacity of IncRNAs and greatly extends the cancer immunopeptidome.

INTRODUCTION

Long non-coding RNAs (lncRNAs) have recently been demonstrated as an abundant class of RNA molecules that play important roles in various types of cellular processes (1,2). Advances in transcriptome have identified a large number of lncRNAs, and mis-regulation of lncRNA expression programs has been found to cause a broad range of human diseases (3-5). Increasing lncRNAs have been identified as oncogenic drivers or tumour suppressors in a variety of cancers (6-8).

LncRNAs are not only well-known as 'non-coding' elements, but also serve as templates for protein translation. Functional peptides encoded by lncRNAs have drawn increasing attention from researchers due to their emerging important roles in fundamental cellular processes and diseases (9,10). The existence of functional peptide products of lncRNAs emphasizes the need to thoroughly separate the RNA functions and peptide functions of lncRNAs. Several mechanisms and evidences for lncRNA translation have been developed. For example, many short sequences, such as N-6-methyladenosine (m6A) sites, have been reported as internal ribosome entry sites (IRESs)-like elements to drive ncRNAs translation (11,12). Rapid developments of genome-wide translation profiling, ribosome profiling and mass spectrometry further provide a chance that has never existed before for genome-wide identification of potential lncRNA peptides (13). Advances in ribosome profiling aid to the systematically identification of small open reading frames (sORFs) from lncRNAs (14). In addition, it is possible to identify translatable lncRNAs based on proteomic data (15). Currently, large number of lncRNAs have been identified based on transcriptome sequencing, and several databases have been developed for describing their genomic annotation (16), tissue-specific expression (17), regulation (5) or diseases (18). However, a specialized and comprehensive resource for lncRNA peptides is still lacking.

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Moreover, peptides that are presented on surface-bound MHC proteins provide valuable targets for cancer immunotherapy (19). Neoantigens have been identified extensively based on the detection of cancer-specific mutations (20,21). Besides genetic mutations, several lines of evidences have suggested other potential sources of neoantigens, such as alternative splicing, RNA editing and peptides generated from noncoding regions (22–24). All these peptides thus expand the pool of MHC immunopeptidomes, targetable by immunotherapies. In particular, lncRNAs have been demonstrated to play an important role in immune regulation (25,26), and their encoded peptides provided a rich resource for tumour neoantigen. However, the potential neoantigen from lncRNAs have yet to be systematically predicted.

To meet this need, we proposed TransLnc for providing or visualizing both computationally predicted and experimentally supported lncRNA peptides in multiple species (human, mouse and rat). By integrating several direct and indirect evidences, we performed an integrative analysis to identify functional peptides encoded by lncRNAs. Importantly, we have computationally predicted the neoantigens from the peptides encoded by lncRNAs, which would provide novel insights into the immunotherapy of cancer. Several flexible tools have been developed to aid retrieval and analysis of the data, particularly the tissue expression patterns of lncRNAs across tissues, the clinical relevance of lncRNAs across cancer types, and the BLAST alignment for finding the best matched lncRNAs of the user queried sequences. Collectively, we expect the TransLnc database will be an important resource for providing novel insights into cancer immunotherapy. All the data and tools are freely available at http://bio-bigdata.hrbmu.edu.cn/TransLnc/.

MATERIALS AND METHODS

Prediction of lncRNAs translation

Both the genomic coordinates and transcript sequences of human and mouse lncRNAs were downloaded from GEN-CODE (Human V32 and Mouse VM23). The corresponding lncRNA information in rat was downloaded from Ensemble database.

To identify all putative peptides encoded by lncRNAs, we used three-frame translation similar as previous studies (11,15,27). All sequences of lncRNA transcripts were first translated into amino acids by the function 'translate' in sequinr package in R program (Figure 1A). ORFs are the nucleic acids sequences between start codons (AUG) and stop codons (UAA, UAG and UGA). Only ORFs that produced peptides longer than 10 amino acids were retained. The selected minimum threshold for ORF lengths was based on the lengths of lncRNAs known to encode peptides in previous studies (27,28). The coding ability of an lncRNAs transcript is calculated as the proportion of RNA sequences that can encode peptides.

Multi-omics evidence for lncRNAs translation

With the development of high-throughput experimental technologies, multi-omics evidence for lncRNAs translation have been revealed. For each computationally predicted

lncRNA peptide, TransLnc integrates six types of direct and indirect evidences to support the coding potential of lncR-NAs (Figure 1A).

Collection of experimentally verified translated lncRNAs. We collected the lncRNAs encoded peptides from four manually curated databases, cncRNAdb (29), EVlncRNAs (30), ncEP (31) and FuncPEP (32). In total, 1185, 18, 13 and 41 lncRNAs that can encode peptides were obtained from four databases. The experimentally supported peptides were aligned with predicted peptides. We identified all lncRNAs and their encoded peptides that covered the experimentally supported ones.

Ribosome-associated lncRNAs. We collected publicly available 2115 Ribo-seq samples from human, mouse and rat. To identify ribosome-associated lncRNAs, the Ribo-seq datasets were processed and reanalysed by a pipeline similar as one of previous studies (33). Briefly, SRA-Toolkit was used to convert sra files to fastaq format. Quality control was performed based on FastQC. We next removed adapter and low quality sequences using Cutadapt (34) and Sickle. The sequences from tRNAs and rRNAs were removed and then aligned to the reference genome by STAR (35). The read count and reads per kilobase per million mapped reads (RPKM) were estimated by featureCounts (36). LncRNAs with RPKM >1 in at least one sample were identified.

Prediction of lncORFs. RibORF was used to systematically detect the actively translated regions in the Ribo-seq datasets (37). Moreover, the predicted ORFs from Meta-mORF (38) and OpenProt (39) were obtained. The ORFs predicted by at least two methods were remained for further analyses. The predicted ORFs were compared with lncR-NAs encoded peptides. LncRNA encoded peptides overlapped with the predicted ORFs were defined as supported by lncORFs.

IRES-supported lncRNAs translation. To identify IRES elements in lncRNAs, we first extracted experimentally validated IRES sequences from IRESbase (40). BLASTN was used to align the IRES sequences to lncRNAs peptides (41). The parameters '-evalue 1e-5 –outfmt 6 –perc_identity 80 – num_alignments 30' were used in our analyses. All lncRNAs encoded peptides with IRES elements were identified.

m6A sites in lncRNAs. The m6A modification peaks across cell lines or tissues were extracted from REPIC (42), which is a database for exploring the m6A methylome. Three computational tools, including exomePeak (43), MeTPeak (44) and MACS2 v2.1.1 (45) were used to detect peaks. The m6A peaks were aligned to the genomic regions of lncRNAs encoded peptides by BEDTools (46).

Detection of lncRNAs-encoded peptides by mass spectrometry. Mass spectrometry-based proteomic data from were used for detection peptides derived from lncRNAs. We first extracted the peptides from PepTransDB resource across nine cancer types in human (15). In addition, we detected the peptides from collected additional MS data of 31 human

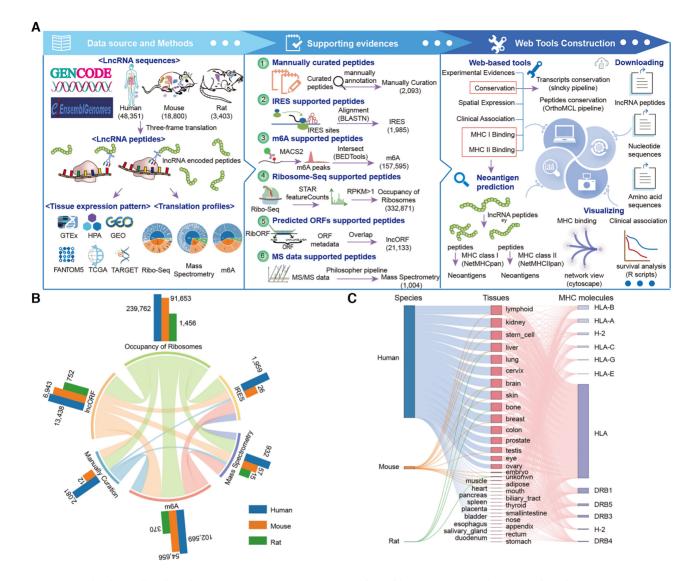


Figure 1. Schematic of overall design of TransLnc. (A) Data content, supporting evidences and web tools construction of TransLnc. (B) Numbers of lncRNAs-encoded peptides with diverse types of supporting evidences. (C) Numbers of potential neoantigens encoded by lncRNAs across different tissues in three species.

normal tissues, as well as mouse and rat tissues. All the MS datasets were obtained from the PRIDE proteomics identifications database (47). There were 41 samples of 31 tissues for human, 9 samples of 3 tissues for mouse and 3 samples of 1 tissue for rat. The similar pipeline in PepTransDB was used to search lncRNA peptides, where MS/MS spectra search was performed using the MSFragger search engine (48) and PSMs were processed using PeptideProphet as implemented in the Philosopher pipeline (49).

Cross-species conservation of translatable lncRNAs

The conservation of both lncRNA transcripts and peptides were calculated. For lncRNA transcripts, we used *slncky* pipeline for identifying orthologous lncRNAs between two species (50), which is a commonly used sensitive method for orthologous lncRNAs alignment. The parameters were the same as one previous study (51). On the other hand, we used reciprocal best hits between pairs of species to cluster lncR- NAs peptides into homologous families with OrthoMCL (52). The parameters were used similar as the study of Sarropoulos *et al.* (2).

Detection of potential neoantigens encoded by lncRNAs

The binding of antigenic peptides to the major histocompatibility complex (MHC) molecules represents a critical step for cellular immunity. MHC molecules come in two main variants: MHC Class I (MHC-I) and MHC Class II (MHC-II). MHC-I binds peptides from intracellular proteins after these undergo proteasomal degradation while MHC-II binds peptides generated by protease-digestion of extracellular proteins. We thus used NetMHCpan-4.1 and NetMHCIIpan-4.0, two commonly used web servers to predict binding between lncRNA peptides and MHC-I or MHC-II, respectively (53). We used the %Rank values for detecting strong MHC binder (SB) or a weak MHC binder (WB). The %Rank < 0.5% and %Rank < 2% thresholds are considered for SBs and WBs for class I, and %Rank <2% and %Rank <10% for SBs and WBs for class II.

Tissue-specificity analysis

Besides Ribo-Seq and MS data with known tissue information, the other data were first mapped to corresponding tissues according the annotations in Cancer Cell Line Encyclopedia (CCLE) (54). Then the remaining cell types were manually mapped to corresponding tissues. The expressions of lncRNAs across various types of human tissues were collected from LncSpA (17), which is a comprehensive resource that provides the spatial expression patterns of lncRNAs. In addition, expression levels of mouse and rat lncRNAs across multiple-tissues were also integrated into TransLnc (55,56).

Clinical relevance

Differentially expressed lncRNAs in cancer with more than five adjacent tissues were identified. We compared the expression of lncRNAs between cancer and normal samples by *t*-test and Fisher's exact test. LncRNAs that were with *P*-value <0.05 were defined as differentially expressed ones. In addition, we used univariate and multivariate cox regression to evaluate the association between survival and expression level of each lncRNA. Patients were ranked based on expression of lncRNA and the difference in survival between low- and high-expression groups was evaluated by the Kaplan-Meier method.

Database implementation

The backend of TransLnc is powered by MySQL (v5.5.48) and accessed via the Java Server Pages with Tomcat container (v 6.0) as the middleware. The front-end of TransLnc is a multi-page web application built based on HTML, JavaScript, and CSS code that consists of jQuery (v3.3.1), Datatable (1.10.25) and ECharts (V5.5.1) plugin. All statistical analyses were performed using the R framework (V3.6.3). TransLnc has been tested on several popular web browsers, including Google Chrome (preferred), Firefox or Apple Safari browsers.

DATABASE CONTENT

Based on data collection and integrative analysis, there were 380,630 (33.58%), 153,926 (13.58%) and 15,160 (1.34%) peptides under the 10 aa length in Human, Mouse and Rat, respectively. As number of studies focused on longer peptides (15,37), we thus excluded the peptides <10 aa in our analysis. In total, the current version of TransLnc contains a total of 583 840 computationally predicted peptides for 33 094 translatable lncRNAs across three different species (Human, Mouse, Rat). In addition, 383 105 lncRNA peptides were supported by at least one of six types of direct or indirect evidences across 34 tissues (Figure 1B). Next, we compared the general characterizations of the predicted peptides versus experimentally confirmed. We found that the predicted peptides were with similar length and chromosome localization (Supplementary Figure S1), but different in amino acid compositions (Supplementary Figure S2), with experimentally confirmed ones. Importantly, there are 220 791 candidate neoantigens binding by MHC class I proteins, and 237 915 candidates binding by MHC II proteins (Figure 1C), extending the cancer immunopeptidome. We found that there were 67,288 peptides can be bound by both molecules, which account for 30.48% and 28.28% of MHC-I and MHC-II binding peptides. Moreover, we compared the characterizations of lncRNAs that encoded peptides binding to MHCs versus no binding. We found that the lncRNAs that encode peptides binding to MHCs were more likely to be implicated in cancer (Supplementary Figure S3). In addition, we obtained the immune-related lncR-NAs from ImmLnc (6), and we found that lncRNAs that encode peptides binding to MHCs were more likely to be involved in immune regulation (Supplementary Figure S4).

To provide an efficient way for users to investigate the function of lncRNAs of interest, TransLnc also provided the useful tools for investigating the function of lncRNAs that encoded peptides. Considering the strong tissue-specificity of lncRNAs, TransLnc allows users to access lncRNA peptides in tissues of interest, multiple visualization results were provided for facilitate understanding their roles in cancer immunity. Several flexible tools have been developed to aid retrieval and analysis of the data, particularly the tissue expression patterns of lncRNAs across tissues, the clinical relevance of lncRNAs across cancer types, and the BLAST alignment for finding the best matched lncRNAs of the user queried sequences.

WEB INTERFACE

TransLnc allows users to explore the peptides encoded by lncRNAs and identify the potential peptides binding by MHC molecules. The easy-to-use interface allows for querving, browsing, visualising and downloading data (Figure 2). In the 'Search' page, TransLnc allows users to query lncRNAs or peptides of interest. TransLnc provides a direct BLAST tool for customized sequence query that allows identifying relevant lncRNAs with associated peptides (Figure 2A). In the 'Browse' page, the users can browse IncRNA encoded peptides by species, tissues, or by experimental evidences, or by peptides bound by MHC I or II (Figure 2B). In addition, six types of interactive tools (experimental evidences, conservation, MHC I or II binding, spatial expression and clinical relevance) were provided in TransLnc to retrieve detail information of lncRNA encoded peptides (Figure 2C). In the 'Result' page, lncRNA and corresponding encoded peptide information, and supporting evidences were shown (Figure 2D).

Once in the detailed page for each lncRNA peptide, the users can get extensive information of sequences and supporting evidences for lncRNA translation (Figure 2E). First, the basic information for lncRNA encoded peptides was provided in a table. In addition, the conservations of lncRNAs and peptides between species were analysed, including the identified novel homologous families. Detailed information for supporting evidences were also provided in the corresponding pages. Moreover, the binding between peptides and MHC molecules were shown as a network. Considering the strong tissue-specificity of lncRNAs, TransLnc allows users to access lncRNA expressions across

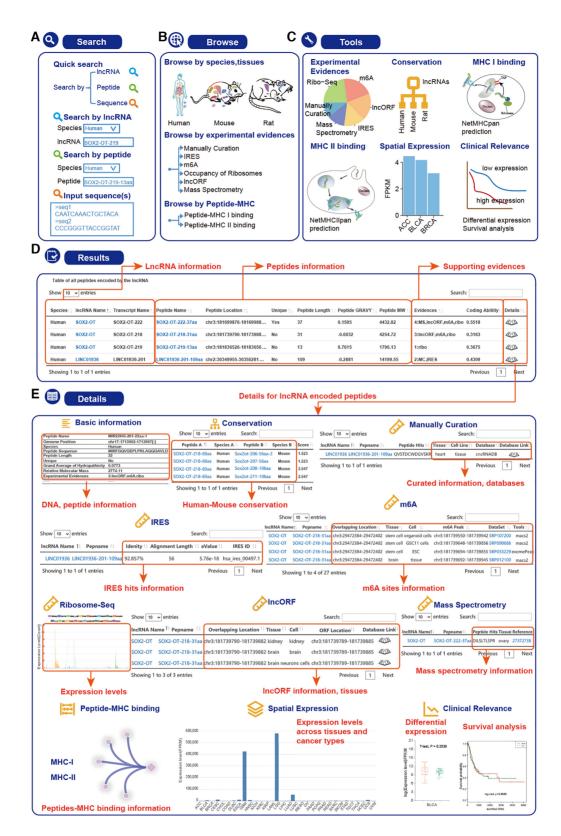


Figure 2. User interface and workflow of using TransLnc. (A) Search page for lncRNAs or peptides of interests. (B) Browse page for lncRNAs or peptides. (C) Interactive tools provided in TransLnc. (D) The result list for lncRNAs and peptides. (E) Detailed information for lncRNA encoded peptides with diverse supporting evidences. Conservation, spatial expression and clinical relevance in diverse cancer types are also provided in TransLnc resource.

tissues or diverse cancer types. Multiple visualization results were provided for facilitate understanding their roles in cancer. TransLnc provides the download of predicted and experimentally supported lncRNA peptides in the 'Download' page and a detail tutorial was provided in the 'Help' page for users showing how to use TransLnc.

CASE STUDY

The lncRNAs HOXB-AS3 is a tumor suppressor and substantially down-regulated in several types of cancer (27). When querying the HOXB-AS3 by 'IncRNA Search' or 'Peptide Search' in TransLnc (Supplementary Figure S5A-B), we obtained 36 entries suggesting that this lncRNAs can encode peptides (Supplementary Figure S5C). In particular, we found that a peptide with 53 aa length was supported by four types of evidence (Supplementary Figure S5C), including manually curation, lncORF, m6A and ribosome-seq. Indeed, previous studies have demonstrated that HOXB-AS3 encoded a conserved 53-aa peptide, than can suppress cancer cell growth, colony formation and migration (27,57). Moreover, based on the expression of lncR-NAs across cancer types, we found that high expression of lncRNAs was associated with better survival of cancer patients. This is consistent with the observation that high HOXB-AS3 peptide levels indicated a better prognosis for colorectal cancer patients (57). Another example is the lncRNA LINC00689, which has been found to play important roles in glioma development and progression (58). Moreover, we found that this lncRNAs can encode 73 potential peptides (Supplementary Figure S6). The lncRNAencoded peptides were supported by multiple lines of evidence. Indeed, previous study has found that linc00689 can encode a micropeptide, STORM (Stress- and TNF-αactivated ORF Micropeptide) (59). Expression analysis indicated that LINC00689 was highly expressed in glioma patients, suggesting the oncogenic roles in cancer (Supplementary Figure S6).

CONCLUSIONS AND FUTURE DEVELOPMENT

With the development of high-throughput technology and experimental validation methods, increasing evidence have suggested that lncRNA-encoded proteins play a critical role in cancer development and progression. However, identification of translatable lncRNAs and analysis of their clinical relevance is lagging. Here we reported an interactive database where multiple-omics evidences were integrated to identify lncRNA-encoded peptides. The database is easy to navigate with interactive tools were integrated, and all data can be downloaded for additional analysis. Importantly, we have computationally predicted the novel neoantigens from the peptides encoded by lncRNAs, which would provide new insights into the immunotherapy of cancer.

Although several database have been developed for describing lncRNAs genomic annotation (16), tissue-specific expression (17), regulation (5) or association with diseases (18), TransLnc is the first resource that focus on lncR-NAs encoded peptides. Moreover, TransLnc also integrated useful tools for investigating the tissue expression patterns and clinical association for lncRNAs of interest. Thus,

TransLnc provides one-stop service for users to explore the function of lncRNAs in cancer. Moreover, there is still room for improvement of TransLnc. In the future, we will continue to update and integrate data content in TransLnc by: (i) continuous literature mining to add more experimentally validated lncRNA peptides, (ii) expanding the available Ribo-seq, m6A and IRES datasets to support the peptides, (iii) extending collection of proteomics data to further enhance the identification of ORF-encoded peptides. These additions are anticipated to enhance the efficiency of TransLnc and it will be an important resource for investigating the translation capacity of lncRNAs and the potential IncRNA encoded peptides. More rigorous investigation of lncRNAs and the small peptides hidden within them would lead to a profound understanding and provide new insights into cancer immunotherapy.

DATA AVAILABILITY

TransLnc is an open comprehensive database for translatable lncRNAs, which is freely available at http://biobigdata.hrbmu.edu.cn/TransLnc/.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

FUNDING

National Natural Science Foundation of China [31871338, 31970646, 61873075, 32060152, 32070673, 32170676]; project supported by Hainan Province Clinical Medical Center, Hainan Provincial Natural Science Foundation of China [820MS053]; Key R&D projects in Hainan Province [ZDYF2021SHFZ051]; Major Science and Technology Program of Hainan Province [ZDKJ202003]; National Key R&D Program of China [2018YFC2000100]; Natural Science Foundation for Distinguished Young Scholars of Heilongjiang Province [JQ2019C004]; Heilongjiang Touyan Innovation Team Program. Funding for open access charge: National Natural Science Foundation of China.

Conflict of interest statement. None declared.

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