REVIEW

Inhibiting the kynurenine pathway in spinal cord injury: multiple therapeutic potentials?

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Abstract

Chronic induction of the kynurenine pathway (KP) contributes to neuroinflammation by producing the excitotoxin quinolinic acid (QUIN). This has led to significant interest in the development of inhibitors of this pathway, particularly in the context of neurodegenerative disease. However, acute spinal cord injury (SCI) also results in deleterious increases in QUIN, as secondary inflammatory processes mediated largely by infiltrating macrophages, become predominant. QUIN mediates significant neurotoxicity primarily by excitotoxic stimulation of the N-methyl-D-aspartate receptor, but other mechanisms of QUIN toxicity are known. More recent focus has assessed the contribution that neuroinflammation and modulations in the KP make in mood and psychiatric disorders with recent studies linking inflammation and modulations in the KP, to impaired cognitive performance and depressed mood in SCI patients. We hypothesize that these findings suggest that in SCI, inhibition of QUIN production and other metabolites, may have multiple therapeutic modalities and further studies investigating this are warranted. However, for central nervous system-based conditions, achieving good blood-brain-barrier permeability continues to be a limitation of current KP inhibitors.

Key Words: spinal cord injury; neuroinflammation; kynurenine pathway; activated microglia; infiltrating macrophages; quinolinic acid; neuropsychiatry; depression

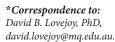
Neuroinflammation, Microglial Phenotype and the Kynurenine Pathway (KP)

The inflammatory response initially involves induction of various cytokines and chemokines to coordinate insult repair. However, in the context of the central nervous system (CNS), sustained neuroinflammation, results in a set of well-described neuroinflammatory events that exacerbate injury. Central to these processes is the switch from the neuroprotective M2 microglial/infiltrating macrophage phenotype to the neurotoxic M1 phenotype, mediated largely by interferon- γ (INF- γ) or lipopolysaccharide (LPS). It is well reported that activated M1 microglia and infiltrating macrophages are the prime generators of the increased quinolinic acid (QUIN) levels observed in all the major neurodegenerative conditions. In fact, macrophages stimulated by INF-y produced nearly 40-fold more QUIN relative to microglia (Guillemin et al., 2004). QUIN potently agonizes the N-methyl-D-aspartate (NMDA) receptor, resulting in excitotoxic stimulation that initiates a cascade of events involving neuronal Ca²⁺ influx, neuronal nitric oxide synthase (nNOS) activation that increases nitric oxide (NO) production and redox stress, which in-turn results in mitochondrial membrane destabilisation, activation of poly ADP-ribose polymerase (PARP), NAD⁺ depletion and eventually neuronal cell death (Jacobs et al., 2017; Figure 1). Accordingly, excessive QUIN production by the over-activated KP is viewed as a significant neuroinflammatory contributor and inhibition of QUIN production has received much attention as a potential therapeutic strategy for neurodegenerative disease. Another potentially neurotoxic metabolite resulting from KP activation is 3-hydroxykynurenine (3-HK), produced by the enzyme kynurenine 3-monooxygenase (KMO) following

hydroxylation of its substrate, kynurenine (KYN). Originally viewed as a neurotoxic metabolite that produces substantial redox activity, 3-HK is now thought to exert more pleiotropic effects on redox status (Jacobs et al., 2017). Similarly, like 3-HK, 3-hydroxyanthranilic acid (3-HAA) was shown to induce neuronal death by redox stress mechanisms, although it was also shown to exert anti-inflammatory effects by supressing cytokine and chemokine expression in activated glial cells (Krause et al., 2011). Other studies showed that while 3-HAA inhibited the Fenton reaction and scavenged reactive oxygen species (ROS) in a deoxyribose degradation assay, pro-oxidant effects were observed in an iron (II) autoxidation assay (Chobot et al., 2015). Interestingly, decreased 3-HAA expression is reported in Huntington's disease, stroke and major depression and as 3-HAA is readily oxidized by radical scavenging, lower 3-HAA concentrations are considered a sensitive biomarker of ROS-mediated disease processes (Darlington et al., 2010).

Spinal Cord Injury (SCI), Macrophages and the KP

SCIs fail to heal like skin or muscle injuries. One perspective suggests that this reflects desynchronization of the conventional sequential pattern of macrophage phenotypes, in which the neuroprotective M2 phenotype that promotes proliferative and remodelling phases of repair, becomes dominated by the pro-inflammatory M1 phenotype such that remodelling is not properly initiated. These authors draw parallels to the chronic inflammation present in non-healing wounds (Gensel and Zhang, 2015). Indeed, shifting the cytokine profile towards that of the neuroprotective M2 phenotype by temporal blockage of interleukin



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doi: 10.4103/1673-5374.241446

Received: 2018-07-06 **Accepted:** 2018-08-13





Jacobs KR, Lovejoy DB (2018) Inhibiting the kynurenine pathway in spinal cord injury: multiple therapeutic potentials? Neural Regen Res 13(12):2073-2076. doi:10.4103/1673-5374.241446

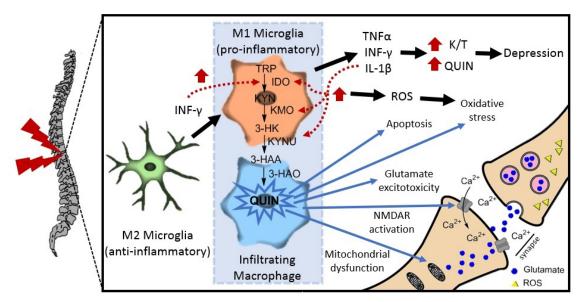


Figure 1 Kynurenine pathway (KP) involvement in spinal cord injury (SCI).

Activated microglia and infiltrating macrophages, cellular features reported in SCI, are the primary source of excitotoxic quinolinic acid (QUIN) in the central nervous system (CNS). Following chronic inflammation, pro-inflammatory cytokines including interferon- γ (INF- γ) induce a phenotypic change from anti-inflammatory M2 microglia/macrophages to pro-inflammatory M1 microglia/macrophages. Once activated, these immune cells produce additional pro-inflammatory cytokines including INF- γ , tumor necrosis factor- α (TNF α) and interleukin (IL)-1 β further inducing the M1 phenotype and upregulating the kynurenine 3-monooxygenase (KMO) branch of the KP, resulting in the production of excess QUIN. QUIN mediated neurotoxicity occurs by several mechanisms, primarily by activation of the N-methyl-D-aspartate receptor (NMDAR). Other mechanisms include increased glutamate release leading to Ca²⁺ influx, mitochondrial dysfunction, increased oxidative stress (reactive oxygen species, ROS) and induction of apoptosis. Neuropsychiatric sequelae are believed to result from the neurotoxic effects of QUIN in specific brain regions following induction of the KP as indicated by increased kynurenine/tryptophan (K/T) ratio (*i.e.*, increased production of kynurenine by the activity of indoleamine 2,3-dioxygenase, the first enzyme in the KP). 3-HAA: 3-Hydroxyanthranilic acid; 3-HAO: 3-hydroxyanthranilic acid oxygenase; 3-HK: 3-hydroxykynurenine; TDO: Indoleamine 2,3-dioxygenase; KYN: kynurenine; KYNU: kynureninase; TRP: tryptophan.

(IL)-6 improved locomotor Basso Mouse Scale (BMS) score following SCI, reduced levels of INF-y and tumor necrosis factor-a (TNF-a) and increased anti-inflammatory IL-4 and IL-13 at the site of the spinal lesion (Guerrero et al., 2012). Key proinflammatory drivers resulting from prolonged engagement of the M1 phenotype include TNF- α , IL-1 β , IFN- γ , NO and superoxide anion (O₂⁻), which are either known to activate the KMO branch of the KP (TNF- α , IL-1 β and IFN- γ) or are products of KP activation (NO and O_2^- ; Figure 1). Thus, KP induction, particularly when chronic, has the potential to engage a vicious-cycle that further fuels KP dysregulation and consequent damage. Considering the numerous observations of infiltrating macrophages in SCIs, the link to the KP becomes further compelling when it is appreciated that activated macrophages produce 204-fold more QUIN than non-activated macrophages and at levels identified as neurotoxic (Guillemin et al., 2004).

Hemeoxygenase-1 in SCI, Relationship to Nrf2 Signaling and the KP

Induction of the enzyme heme oxygensase-1 (HO-1) is frequently reported in animal models of SCI. For instance, in rat SCI, HO-1 was overexpressed by activated neutrophils and use of a HO-1 inhibitor delayed motor function recovery after SCI, suggesting that HO-1 coordinates significant anti-inflammatory and protective effects in damaged tissue (Liu et al., 2002). The nuclear factor erythroid 2-related factor 2/anti-oxidant response (Nrf-2/ARE) pathway is described as the master regulator of the cellular anti-oxidant defence network and is a known inducer of HO-1. Essentially, when exposed to ROS, Nrf-2 translocates to the nucleus, where it activates HO-1 transcription. Interestingly, the KP metabolites QUIN and 3-HK can both promote Nrf-2 nuclear translocation (Colin-Gonzalez et al., 2014a, b). This could imply an activity in opposition to their described deleterious roles in neurodegenerative disease. However, QUIN treated rat striatal slices showed increased Nrf-2 nuclear translocation after 1 hour and reduced lipid peroxidation after 3 hours (Colin-Gonzalez et al., 2014a). This temporal lag suggests that QUIN-induced nuclear translocation of Nrf-2 was not a direct effect, but rather a compensatory response to QUIN induced ROS production which forms part of the general anti-oxidant defence system. Although data is lacking, on balance, it seems likely that 3-HK also induces Nrf-2 translocation as a compensatory mechanism. However, unlike QUIN, 3-HAA seems to mediate an overall neuroprotective effect. In primary human fetal CNS cultures treated with IL-1 (with or without IFN- γ) or with Toll-like receptor ligands (as a model of the proinflammatory CNS environment), 3-HAA treatment suppressed glial cytokine and chemokine expression and reduced cytokine-induced neuronal death (Krause et al., 2011). 3-HAA also markedly increased the expression HO-1 by astrocytes and macrophages (Krause et al., 2011). After further in-vivo assessment, the authors concluded that 3-HAA showed promising activity as a potential neuroprotective compound.

Do Neuropsychiatric Symptoms Further Connect Macrophages and SCI to the KP?

There is also growing interest in the role that KP activation plays in major depression, suicidal ideation, autism spectrum disorder and schizophrenia (Steiner et al., 2011; Lim et al., 2016; Bryleva and Brundin, 2017; Plitman et al., 2017). Influential studies include those by Steiner et al. (2011), who showed that depressed patients had significantly increased immunoreactive QUIN-positive microglia in the subgenual anterior cingulate and cortex anterior midcingulate cortex compared to controls. In suicide attempters QUIN was significantly elevated approximately 2-fold in the cerebro-spinal fluid (CSF) relative to controls and was associated with increased CSF IL-6. QUIN CSF levels also correlated to scores on Suicide Intent Scale and significantly decreased 6 months post suicide attempt (Erhardt et al., 2013). Studies also link inflammation and KP modulation to mood disturbances in SCI patients. In a randomized, controlled clinical trial, 20 patients with varying severity of SCI and 20 control subjects received an anti-inflammatory diet for 3 months (Allison and Ditor, 2015). Significant group × time interactions were found for standardized score of depression, the ratio between tryptophan (TRP) and large neutral amino acids (LNAA; that reflects brain TRP concentration) and the composite score of pro-inflammatory cytokines. Changes in IL-1ß significantly correlated to standardized depression score and kynurenine/tryptophan (K/T) ratio (Pearson's r correlations). Changes in K/T ratio were also significantly correlated with depression scores, with further statistical analysis showing that this correlation was mediated by changes in IL-1 β (Allison and Ditor, 2015). Although a subsequent follow-up study showed that there were no significant links between consumption of an anti-inflammatory diet in SCI to improvements in cognitive function (Allison et al., 2017), presumably because participants had underlying hippocampal damage not tractable by reduced inflammation.

Targeting the KP in SCI

An early report showed that after a brief compression injury to the lower thoracic spinal cord (T13) of adult guinea pigs, QUIN levels were elevated greater than 10-fold at the injury site after 5 days, while the activity of the first enzyme in the KP indoleamine 2,3-dioxygenase (IDO-1) was increased approximately 2-fold. These changes were not found in adjacent undamaged regions (Blight et al., 1993). Other studies have demonstrated that injections of 4-chloro-3-hydroxyanthranilate (4-Cl-3-HAA) a 3-hydroxyanthranilic acid oxygenase (3-HAO) inhibitor, blocking the metabolism of 3-HAA into QUIN, every 12 hours for 12 days post SCI significantly reduced the accumulation of QUIN at the site of injury and reduced the severity of delayed functional deficits in adult guinea pigs (Blight et al., 1995; Yates et al., 2006). This coincided with a significant 100% increase in the area of surviving white matter at the injury site in this model (Yates et al., 2006). A single dose of 4-Cl-3-HAA did not produce the same sensory and motor improvements suggesting that the benefits of continued 4-Cl-3-HAA treatment result from a reduction in QUIN rather than any early anti-oxidant activity (Yates et al., 2014). However, the use of 4-Cl-3-HAA did not significantly increase upstream neuroprotective kynurenic acid (KYNA) which may offer additional benefits in SCI. For example, a KYNA derivative (glucosamine-kynurenic acid) administered *via* spinal catheter resulted in improved motor function recovery at 1–4 weeks post-injury in male adult Wistar rats (Korimová et al., 2012) and KYNA administered by stereotaxic microinjection resulted in significant dose dependent improvements in hind limb function and reduced overall functional deficits following SCI (Wrathall et al., 1992). Taken together this suggests that targeting the KP and in particular KMO in order to shift the KP away from QUIN and towards KYNA may have several beneficial effects in SCI.

Another therapeutic benefit of targeting KMO in SCI was shown by treatment of spinal cord damaged rats with gene transfer of human KAT-II (the KP enzyme that leads to KYNA synthesis) which improved overactive bladder problems that can result from SCI (Jia et al., 2014). Mechanically, this improvement resulted from increased KYNA expression that antagonises the NMDA receptor. Previous studies have shown that supressing NMDA receptors improves detrusor activity that leads to bladder overactivity in SCI. Additionally, ketamine and other NMDA receptor antagonists have shown significant promise for the treatment of neuropathic pain, a common complication following SCI (Niesters and Dahan, 2012; Hagen and Rekand, 2015). In a rat model of neuropathic pain, induced following chronic constriction injury (CCI) of the right sciatic nerve, mRNA levels of IDO2, KMO and 3-HAO were significantly increased 7 days post-injury. Repeated administration of KMO inhibitors UPF-648 and Ro 61-8048 reduced hypersensitivity and neuropathic pain (Rojewska et al., 2016, 2018), most likely by shifting the KP towards the NMDA antagonist KYNA.

Conclusions and Future Directions

The data discussed above forms a conceptual framework that supports investigating the therapeutic potential of KMO inhibitors in SCI to not only reduce the pathophysiological impact of chronic inflammation, but to potentially improve psychological disturbances and neuropathic pain that can result from chronic SCI. Since the demonstration by Zwilling et al. (2011), that KMO inhibition ameliorated neurodegeneration and produced other positive results in mouse models of Alzheimer's and Huntington's diseases, much effort has focused on the development of KMO inhibitors. KMO is viewed as a better target than IDO-1, as KMO is present at the first junction of the KP and its inhibition leads to reduced QUIN but also increased neuroprotective KYNA. However, as pointed out by Zwilling et al. (2011), the therapeutic effects of the KMO inhibitor used did not result from reduced brain levels of QUIN but reflected the peripheral increase in KYN that is able to cross the blood-brain-barrier (BBB) and leads to increased brain KYNA (Zwilling et al., 2011). Newer, alternate, KMO inhibitors similarly fail to achieve good CNS exposure (CHDI-246 and UPF-648) and a good case exists for the development of a BBB-permeable KMO inhibitor that would achieve both therapeutic goals, *i.e.*, reduced CNS QUIN with increased KYNA. In this vein, we recently commenced a KMO inhibitor discovery project. We utilized high-throughput screening (HTS), specifically using a lead-like structure library comprising a high proportion of leads likely to be CNS-permeable (Jacobs et al., 2018) and we hope to develop a BBB-permeable KMO inhibitor that could possibly be useful in SCI.

Acknowledgments: *The authors thank Prof Gilles Guillemin, Macquarie University for helpful discussions.*

Author contributions: *Manuscript conception: DBL; literature search and manuscript writing: KRJ and DBL.*

Conflicts of interest: None declared.

Financial support: None.

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Open peer reviewer: Hedong Li, Pennsylvania State University, USA. Additional file: Open peer review report 1.

References

- Allison DJ, Ditor DS (2015) Targeting inflammation to influence mood following spinal cord injury: a randomized clinical trial. J Neuroin-flammation 12:204.
- Allison DJ, Josse AR, Gabriel DA, Klentrou P, Ditor DS (2017) Targeting inflammation to influence cognitive function following spinal cord injury: a randomized clinical trial. Spinal cord 55:26-32.
- Blight AR, Saito K, Heyes MP (1993) Increased levels of the excitotoxin quinolinic acid in spinal cord following contusion injury. Brain Res 632:314-316.
- Blight AR, Cohen TI, Saito K, Heyes MP (1995) Quinolinic acid accumulation and functional deficits following experimental spinal cord injury. Brain 118:735-752.
- Bryleva EY, Brundin L (2017) Kynurenine pathway metabolites and suicidality. Neuropharmacology 112:324-330.
- Chobot V, Hadacek F, Weckwerth W, Kubicova L (2015) Iron chelation and redox chemistry of anthranilic acid and 3-hydroxyanthranilic acid: A comparison of two structurally related kynurenine pathway metabolites to obtain improved insights into their potential role in neurological disease development. J Organomet Chem 782:103-110.
- Colín-González AL, Maya-López M, Pedraza-Chaverrí J, Ali SF, Chavarría A, Santamaría A (2014b) The Janus faces of 3-hydroxykynurenine: Dual redox modulatory activity and lack of neurotoxicity in the rat striatum. Brain Res 1589:1-14.
- Colin-Gonzalez AL, Luna-Lopez A, Konigsberg M, Ali SF, Pedraza-Chaverri J, Santamaria A (2014a) Early modulation of the transcription factor Nrf2 in rodent striatal slices by quinolinic acid, a toxic metabolite of the kynurenine pathway. Neuroscience 260:130-139.
- Darlington LG, Forrest CM, Mackay GM, Smith RA, Smith AJ, Stoy N, Stone TW (2010) On the biological importance of the 3-hydroxyanthranilic acid: anthranilic acid ratio. Int J Tryptophan Res 3:51-59.
- Erhardt S, Lim CK, Linderholm KR, Janelidze S, Lindqvist D, Samuelsson M, Lundberg K, Postolache TT, Traskman-Bendz L, Guillemin GJ, Brundin L (2013) Connecting inflammation with glutamate agonism in suicidality. Neuropsychopharmacology 38:743-752.
- Gensel JC, Zhang B (2015) Macrophage activation and its role in repair and pathology after spinal cord injury. Brain Res 1619:1-11.
- Guerrero AR, Uchida K, Nakajima H, Watanabe S, Nakamura M, Johnson WEB, Baba H (2012) Blockade of interleukin-6 signaling inhibits the classic pathway and promotes an alternative pathway of macrophage activation after spinal cord injury in mice. J Neuroinflammation 9:40.

- Guillemin GJ, Smythe G, Takikawa O, Brew BJ (2004) Expression of indoleamine 2,3-dioxygenase and production of quinolinic acid by human microglia, astrocytes, and neurons. Glia 49:15-23.
- Hagen EM, Rekand T (2015) Management of neuropathic pain associated with spinal cord injury. Pain Ther 4:51-65.
- Jacobs KR, Guillemin GJ, Lovejoy DB (2018) Development of a rapid fluorescence-based high-throughput screening assay to identify novel kynurenine 3-monooxygenase inhibitor scaffolds. SLAS Discov 23:554-560.
- Jacobs KR, Castellano-Gonzalez G, Guillemin GJ, Lovejoy DB (2017) Major developments in the design of inhibitors along the kynurenine pathway. Curr Med Chem 24:2471-2495.
- Jia C, Yoshimura N, Liao L (2014) Herpes simplex virus vector-mediated gene transfer of kynurenine aminotransferase improves detrusor overactivity in spinal cord-injured rats. Gene Ther 21:484-489.
- Korimová A, Cížková D, Toldi J, Vécsei L, Vanický I (2012) Protective effects of glucosamine-kynurenic acid after compression-induced spinal cord injury in the rat. Cent Eur J Biol 7:996-1004.
- Krause D, Suh HS, Tarassishin L, Cui QL, Durafourt BA, Choi N, Bauman A, Cosenza-Nashat M, Antel JP, Zhao ML, Lee SC (2011) The tryptophan metabolite 3-hydroxyanthranilic acid plays anti-inflammatory and neuroprotective roles during inflammation: role of hemeoxygenase-1. Am J Pathol 179:1360-1372.
- Lim CK, Essa MM, de Paula Martins R, Lovejoy DB, Bilgin AA, Waly MI, Al-Farsi YM, Al-Sharbati M, Al-Shaffae MA, Guillemin GJ (2016) Altered kynurenine pathway metabolism in autism: Implication for immune-induced glutamatergic activity. Autism Res 9:621-631.
- Liu Y, Tachibana T, Dai Y, Kondo E, Fukuoka T, Yamanaka H, Noguchi K (2002) Heme oxygenase-1 expression after spinal cord injury: the induction in activated neutrophils. J Neurotrauma 19:479-490.
- Niesters M, Dahan A (2012) Pharmacokinetic and pharmacodynamic considerations for NMDA receptor antagonists in the treatment of chronic neuropathic pain. Expert Opin Drug Metab Toxicol 8:1409-1417.
- Plitman E, Iwata Y, Caravaggio F, Nakajima S, Chung JK, Gerretsen P, Kim J, Takeuchi H, Chakravarty MM, Remington G, Graff-Guerrero A (2017) Kynurenic acid in schizophrenia: a systematic review and meta-analysis. Schizophr Bull 43:764-777.
- Rojewska E, Piotrowska A, Makuch W, Przewłocka B, Mika J (2016) Pharmacological kynurenine 3-monooxygenase enzyme inhibition significantly reduces neuropathic pain in a rat model. Neuropharmacology 102:80-91.
- Rojewska E, Ciapala K, Piotrowska A, Makuch W, Mika J (2018) Pharmacological inhibition of indoleamine 2,3-dioxygenase-2 and kynurenine 3-monooxygenase, enzymes of the kynurenine pathway, significantly diminishes neuropathic pain in a rat model. Front Pharmacol 9:724.
- Steiner J, Walter M, Gos T, Guillemin GJ, Bernstein HG, Sarnyai Z, Mawrin C, Brisch R, Bielau H, zu Schwabedissen LM, Bogerts B, Myint AM (2011) Severe depression is associated with increased microglial quinolinic acid in subregions of the anterior cingulate gyrus: Evidence for an immune-modulated glutamatergic neurotransmission? J Neuroinflammation 8:94.
- Wrathall JR, Bouzoukis J, Choiniere D (1992) Effect of kynurenate on functional deficits resulting from traumatic spinal cord injury. Eur J Pharmacol 218:273-281.
- Yates JR, Heyes MP, Blight AR (2006) 4-Chloro-3-hydroxyanthranilate reduces local quinolinic acid synthesis, improves functional recovery, and preserves white matter after spinal cord injury. J Neurotrauma 23:866-881.
- Yates JR, Gay EA, Heyes MP, Blight AR (2014) Effects of methylprednisolone and 4-chloro-3-hydroxyanthranilic acid in experimental spinal cord injury in the guinea pig appear to be mediated by different and potentially complementary mechanisms. Spinal Cord 52:662-666.
- Zwilling D, Huang SY, Sathyasaikumar KV, Notarangelo FM, Guidetti P, Wu HQ, Lee J, Truong J, Andrews-Zwilling Y, Hsieh EW, Louie JY, Wu T, Scearce-Levie K, Patrick C, Adame A, Giorgini F, Moussaoui S, Laue G, Rassoulpour A, Flik G, et al. (2011) Kynurenine 3-monooxygenase inhibition in blood ameliorates neurodegeneration. Cell 145:863-874.

P-Reviewer: Li H; C-Editor: Zhao M; T-Editor: Liu XL