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Article

# Synthesis and Antibacterial Activity of Some Heterocyclic Chalcone Analogues Alone and in Combination with Antibiotics

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Abstract: A series of simple heterocyclic chalcone analogues have been synthesized by Claisen Schmidt condensation reactions between substituted benzaldehydes and heteroaryl methyl ketones and evaluated for their antibacterial activity. The structures of the synthesized chalcones were established by IR and <sup>1</sup>H-NMR analysis. The biological data shows that compounds  $\mathbf{p}_5$ ,  $\mathbf{f}_6$  and  $\mathbf{t}_5$  had strong activities against both susceptible and resistant *Staphylococcus aureus* strains, but not activity against a vancomycin and methicillin resistant *Staphylococcus aureus* isolated from a human sample. The structure and activity relationships confirmed that compounds  $\mathbf{f}_5$ ,  $\mathbf{f}_6$  and  $\mathbf{t}_5$  are potential candidates for future drug discovery and development.

**Keywords:** chalcone; heterocyclic chalcone analogues; synthesis; antibacterial; synergitic activity; antibiotics combination; MRSA

#### 1. Introduction

In the past, many decades since penicillin was discovered and introduced as a powerful antibacterial agent, antibiotics have become critical in the fight against infectious diseases caused by bacteria and other microbes. However, widespread antibiotic use has promoted the emergence of antibiotic-resistant pathogens, including multidrug resistant strains [1–3]. At present, the appearance of more and more pathogenetic bacterial species resistant to conventional antibiotics has resulted in either high expenses or failure in the treatment of infectious diseases. An alarming increase in resistance of bacteria that cause community acquired infections has also been documented, especially in Staphylococci and Pneumococci, which are prevalent causes of disease and mortality. In addition, the risk of opportunistic fungal infections increases rapidly accompanied with AIDS disease, and as an obviously consequence invasive infections represent a major cause of mortality for these patients [4]. With the emergence of new microbial strains resistant to many conventional available antibiotics there is growing interest in the discovery of new antibacterial agents [1,2]. According to the known structure and activity relationships, it is considered that certain small heterocyclic molecules act as highly functionalized scaffolds and are known pharmacophores of a number of biologically active and medicinally useful molecules [5,6].

Chalcones (1,3-diphenyl-propene-1-one) belonging to the flavonoid family, are natural and synthetic products that have been reviewed for their wide range of biological activities as antibacterial [7,8], anti-tumor [9–11], anti-inflammatory [12–14] and antioxidant agents [15–18], *etc.* Although studies on the bioavailability of heterocyclic chalcones from natural sources are limited, they have been reported as having a wide range of biological activities, especially antibacterial [19–28], and antifungal activities [29]. In an effort to diversify the biological activities of conventional chalcones, a series of heterocyclic chalcone analogues in which an electron rich nitrogen or oxygen as well as thiophene heterocyclic chalcone analogues (Figure 1) using a conventional base catalyzed Claisen Schmidt condensation reaction and their possible antibacterial activity alone and in combination with antibiotics.

**Figure 1.** Structure of heterocyclic chalcones obtained from the Claisen Schmidt condensation between heteroaryl methyl ketones and substituted benzaldehydes.



#### 2. Results and Discussion

#### 2.1. Chemistry

A number of known and novel heterocyclic chalcone analogues were prepared via Claisen Schmidt condensation reactions [14,30] between appropriate benzaldehydes and heteroaryl methyl ketones like pyridine-2-yl methyl ketone, thiophene-2-yl methyl ketone and furan-2-yl methyl ketone (Scheme 1).

The purity of the products and progress of the reaction was checked by TLC on silica gel plates. The condensation step was carried out over 6 h to 36 h. After recrystallization from methanol all corresponding chalcones were obtained in 43–63% yields.

Scheme 1. Claisen–Schmidt condensation.



The structures of the synthesized compounds were confirmed by IR, <sup>1</sup>H-NMR and mass spectrometry measurements. The <sup>1</sup>H-NMR spectrum of the synthesized chalcones displayed two doublets at  $\delta$  7.2–8.4 ppm with a characteristic coupling constant (*J*) of 15–16 Hz, which confirms the formation of chalcones (possessing a  $\alpha,\beta$ -unsaturated ketone). This higher coupling constant value indicates all synthetic compounds were geometrically pure and were exclusively *trans* (*E*) isomers. This condensation produced three series of heterocyclic chalcones, which differ in the A ring like pyridine-2-yl-chalcones (**p**<sub>1-7</sub>), furan-2-yl-chalcones (**f**<sub>1-7</sub>) and thiophene-2-yl-chalcone (**t**<sub>1-7</sub>), as listed in Table 1.

#### 2.2. Antimicrobial Activity

The antimicrobial assays were carried out by the dilution method using the following bacterial strains: methicillin-sensitive *Staphylococcus aureus* ATCC 29213 (SA or MSSA), methicillin-resistant *Staphylococcus aureus* ATCC 43300 (MRSA) and a vancomycin and methicillin resistant *Staphylococcus aureus* isolated from a human sample. Table 2 summarizes the results obtained for the MICs of the heterocyclic chalcones against the three bacterial strains. Of the 21 heterocyclic chalcones, five compounds had strong activity against the tested *S. aureus*, including two compounds pertaining to the pyridine-2-yl-chalcone group ( $\mathbf{p}_5$  and  $\mathbf{p}_6$ ), two compounds pertaining to the furan-2-yl-chalcone group ( $\mathbf{t}_5$ ).

Results of determining the antibacterial coordination ability of the test substances and antibiotics are shown in Table 3. Interestingly, while the five abovementioned chalcones ( $\mathbf{p}_5$ ,  $\mathbf{p}_6$ ,  $\mathbf{f}_5$ ,  $\mathbf{f}_6$ ,  $\mathbf{t}_5$ ) showed moderate antibacterial effects, they might increase dramatically the activity of some combined antibiotics. The chalcones in these combinations also exhibited an antibacterial effect better than that seen when they were used alone (Table 4). In the checkerboard technique, results of quantitatively determining antibacterial coordination ability proved that combinations between antibiotics and chalcone derivatives  $\mathbf{f}_5$  and  $\mathbf{p}_6$  had synergistic effects against MRSA ATCC 43300. Such combinations are mixtures of compound  $\mathbf{f}_5$  and oxacillin; compound  $\mathbf{p}_6$  and vancomycin. However, no synergistic effect of the combinations of the synthesized heterocylic chalcones and test antibiotics could be found on the local hospital isolated MRSA.

	3
	$2 \longrightarrow 4$
R	5
	6

Common da	A ring		Substitu	tion in B rin		Isolated yield	
Compounds	(R)	C2	<b>C3</b>	C4	C5	Formula	(%)
p1		Н	$NO_2$	Н	Н	$C_{14}H_{10}N_2O_3$	45
թ2		Н	Н	$N(CH_3)_2$	Н	$C_{16}H_{16}N_2O$	62
ր3		Н	Н	OCH <sub>3</sub>	OCH <sub>3</sub>	$C_{16}H_{15}NO_3$	58
p4	Ľ N	Н	$OCH_3$	OCH <sub>3</sub>	OCH <sub>3</sub>	$C_{17}H_{17}NO_4$	51
p5		OH	Н	Н	Η	$C_{14}H_{11}NO_2$	51
р6		Н	OH	Н	Н	$C_{14}H_{11}NO_2$	50
p7		Н	Н	OH	Η	$C_{14}H_{11}NO_2$	58
f1		Н	$NO_2$	Н	Η	$C_{13}H_9NO_4$	43
f2		Η	Н	$N(CH_3)_2$	Η	$C_{15}H_{15}O_2$	61
f3		Н	Н	OCH <sub>3</sub>	$OCH_3$	$C_{15}H_{14}O_4$	55
f4	0	Н	$OCH_3$	OCH <sub>3</sub>	$OCH_3$	$C_{16}H_{16}O_3$	51
f5		OH	Н	Н	Н	$C_{13}H_{10}O_5$	54
f6		Н	OH	Н	Н	$C_{13}H_{10}O_{3}$	58
f7		Н	Н	OH	Н	$C_{13}H_{10}O_{3}$	62
t1		Н	$NO_2$	Н	Η	$C_{13}H_9NO_3S$	46
t2		Н	Н	$N(CH_3)_2$	Н	$C_{15}H_{15}NOS$	63
t3		Н	Н	OCH <sub>3</sub>	$OCH_3$	$C_{15}H_{14}O_3S$	50
t4	`S´ `	Н	$OCH_3$	OCH <sub>3</sub>	$OCH_3$	$C_{16}H_{16}O_4S$	53
t5		OH	Н	Н	Η	$C_{13}H_{10}O_2S$	55
t6		Н	OH	Н	Н	$C_{13}H_{10}O_2S$	52
t7		Н	Н	OH	Н	$C_{13}H_{10}O_2S$	58

Table 2. Anti-*Staphylococcus aureus* activity of the synthetic compounds (minimum inhibitory concentration in  $\mu$ g/mL).

Chalcones <sup>a</sup>	Pyridi	n-2yl-ch	alcones	Furan	-2yl-cha	lcones	Thiophen-2yl-chalcones		
Bacteria	<b>p</b> 5	<b>p</b> 6	<b>p</b> 6	<b>f</b> 5	<b>f</b> <sub>6</sub>	<b>f</b> <sub>7</sub>	t5	t <sub>6</sub>	
MSSA ATCC 25923	128	64	512	64	128	256	32	256	
MRSA ATCC 44330	128	128	512	64	128	256	64	128	
MRSA-I (isolated) <sup>b</sup>	-	64	-	32	512	-	256	-	

<sup>a</sup>: the remaining compounds ( $p_{1-4}$ ;  $f_{1-4}$ ;  $t_{1-4}$  and  $t_7$ ) were inactive at a concentration of 512 µg/mL;

<sup>b</sup>: isolated MRSA (MICs: oxacillin 256; vancomycin 64  $\mu$ g/mL); (-): inactive at a concentration of 512  $\mu$ g/mL; ND: not determined.

	P <sub>6</sub>			$\mathbf{f}_5$			$\mathbf{f}_6$				t <sub>5</sub>		
Antibiotics	SA	MRSA	Isolated MRSA	SA	MRSA	Isolated MRSA	SA	MRSA	Isolated MRSA	SA	MRSA	Isolated MRSA	
Vancomycin	-	+	-	-	+	-	-	+	-	+	+	-	
Doxycycline	-	-	-	-	-	-	-	-	-	-	-	-	
Ciprofloxacin	+	-	-	-	-	-	+	-	-	-	-	-	
Gentamicin	-	-	-	-	-	-	-	-	-	-	-	-	
Chloramphenicol	-	-	-	-	-	-	-	-	-	-	-	-	
Oxacicllin	-	-	-	-	+	-	-	+	-	-	+	-	
Erythromycin	-	-	-	-	-	-	-	-	-	+	-	-	

 Table 3. Antibacterial coordination ability results.

(SA): MSSA ATCC 25923; (MRSA): MRSA ATCC 43300; (isolated MRSA): MRSA with MICs of oxacicllin 256;  $64 \mu g/mL$  and vancomycin  $64 \mu g/mL$ .

	MSSA ATCC 25923						MRSA ATCC 43300					
Miy <sup>a</sup>	MICs				Increasing	MICs				Increasing		
IVIIX	(μg/	(µg/mL)		Interpretation	rate	(µg/	(µg/mL)		Interpretation	rate		
	Alone	Mix <sup>a</sup>			(fold) <sup>b</sup>	Alone	Mix <sup>a</sup>			(fold) <sup>b</sup>		
<b>p</b> 6 Cipro	64 0.5	32 0.125	0.75	Additive		ND	ND	ND	ND	ND		
<b>p</b> 6 Vanco	ND	ND	ND	ND	ND	128 1	32 0.0625	0.31	Synergistic	4 16		
<b>f</b> 5 Oxa	ND	ND	ND	ND	ND	64 2	16 0.125	0.31	Synergistic	4 16		
<b>f</b> <sub>6</sub> Cipro	128 0.5	64 0.125	0.75	Additive	2 4	ND	ND	ND	ND	ND		
f <sub>6</sub> Vanco	ND	ND	ND	ND	ND	128 1	64 0.125	0.625	Additive	2 8		
f <sub>6</sub> Oxa	ND	ND	ND	ND	ND	128 2	64 0.125	0.56	Additive	2 16		
<b>t</b> 5 Ery	32 0.5	32 0.03	1.06	Indifferent	1 1.67	ND	ND	ND	ND	ND		
<b>t</b> 5 Cipro	32 0.5	16 0.25	1	Additive	2 2	ND	ND	ND	ND	ND		
t <sub>5</sub> Vanco	ND	ND	ND	ND	ND	64 1	32 0.5	1.0	Additive	2 2		
t <sub>5</sub> Oxa	ND	ND	ND	ND	ND	64 2	32 0.125	0.56	Additive	2 16		

<sup>(a)</sup> Mix = Combination; <sup>(b)</sup> Rate in increasing antibacterial activity of antibiotic/chalcone in combination compared to that of in alone (fold); ND = not dertemined. Vanco: vancomycin; Cipro: ciprofloxacin; Ery: erythromycin; Oxa: oxacillin.

Some synthesized heterocyclic chalcones exhibit strong antibacterial activities and may act by damaging the cell wall of SA, which is clearly similar to the observed mechanism of a well-known cell membrane permeabilizer, polymyxin B [27]. However, the molecular mechanism of action is not yet

clearly understood. Some preliminary remarks on structure–activity relationship can be drawn from the bioactivity results as follows:

(*i*) A free hydroxyl group in position 2/3 in the B ring (phenyl moiety) appears to be very important for anti-*Staphylococcus aureus* activity ( $\mathbf{p}_5$ ,  $\mathbf{p}_6$ ,  $\mathbf{f}_5$ ,  $\mathbf{f}_6$ ,  $\mathbf{t}_5$ ). In case of substitution of a hydroxyl group in position 4 ( $\mathbf{p}_7$ ,  $\mathbf{f}_7$ ,  $\mathbf{t}_7$ ), chalcones were inactive on the tested *Staphylococcus aureus* strains.

(*ii*) Substitution in the B ring with a nitro group in position 3 (as in  $\mathbf{p}_1$ ,  $\mathbf{f}_1$ ,  $\mathbf{t}_1$ ) or with two/three methoxyl groups in positions 3/4/5 (as in  $\mathbf{p}_3$ ,  $\mathbf{p}_4$ ,  $\mathbf{f}_3$ ,  $\mathbf{f}_4$ ,  $\mathbf{t}_3$  and  $\mathbf{t}_4$ ) might also be responsible for the decrease in the anti-*Staphylococcus aureus* activity;

(*iii*) The chalcones possessing two/three methoxy groups in B ring were inactive, regardless of their A ring structures. This means that methoxy groups seem to abolish the hydrophilic property of the phenol hydroxy moiety which can affect penetration of antibiotics through bacterial cell walls.

(*iv*) The chalcones possessing a hydroxyl group in B ring ( $\mathbf{p}_6$ ,  $\mathbf{f}_6$  and  $\mathbf{t}_5$ ) demonstrated strongly positive interactions with antibiotic such as ciprofloxacin on methicillin-sensitive-*Staphylococcus aureus*. These chalcones increased significantly the activity of combined antibiotics like vancomycin and oxacillin up to sixteen-fold on methicillin -resistant-*Staphylococcus aureus*. These combinations could lead to develop new treatments for MRSA infectious diseases;

(v) The analysis of structural influence of A ring on anti-*Staphylococcus aureus* activity suggested that a furan-2-yl moiety may be more important than the thiophene-2-yl or pyridine-2yl-moieties.

## 3. Experimental

## 3.1. General

Melting points (mp) were taken in open capillary tubes using a Gallenkamp apparatus and are uncorrected. The IR spectra were recorded on a Shimadzu FTIR 8201 PC spectrophotometer. <sup>1</sup>H-NMR spectra were determined on a Bruker Ultrashield 500 spectrometer using tetramethylsilane (TMS) as an internal reference. All the starting materials are commercially available.

## 3.2. General Procedure for the Synthesis of Chalcones [14,30]

A solution of heteroaryl methyl ketone (5 mM) and aromatic aldehyde (5 mM) in methanol (15 mL) was cooled to 5–10 °C in an ice bath. The cooled solution was treated with a small portion of pulverized KOH (10 mM). The reaction mixture was magnetically stirred for 60 min and then left overnight or longer, as monitored by thin layer chromatography using *n*-hexane–acetone (5:1) as developing solvent. The resulting dark solution was diluted with ice water and carefully acidified using dilute hydrochloric acid. The chalcones, which separated as a yellow solid, were collected by filtration after washing with water and further purified by crystallization from methanol. When needed, the crude product was further purified by silica gel column chromatography.

Characterization data of the synthesized heterocyclic chalcone analogues:

*I-(Pyridine-2-yl)-3-(3-nitrophenyl)-2-propene-1-one* (**p**<sub>1</sub>). Mp: 180 °C; IR (vcm<sup>-1</sup>, KBr): 1674 (υ<sub>C=O</sub>) and 1616 (υ<sub>C=C</sub>). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>, δ ppm): 8.83 (d, J = 4.5 Hz, 1H, H<sub>6</sub>'); 8.63 (s, 1H, H<sub>2</sub>); 8.40 (d, J = 16 Hz, 1H, H<sub>β</sub>); 8.31 (d, J = 7.5 Hz, 1H, H<sub>4</sub>); 8.29 (d, J = 2 Hz, 1H, H<sub>3</sub>'); 8.14 (d, J = 8 Hz, 1H,

H<sub>6</sub>); 8.10 (m, 1H, H<sub>4</sub>); 7.98 (d, J = 16 Hz, 1H, H<sub> $\alpha$ </sub>); 7.77 (t, 1H, H<sub>5</sub>); 7.74 (m, 1H, H<sub>5</sub>). MS (ESI, positive scan): m/z 277.23 [M+Na]<sup>+</sup>.

*1-(Pyridine-2-yl)-3-(4-dimethylaminophenyl)-2-propene-1-one* (**p**<sub>2</sub>). Mp: 138 °C; IR: (vcm<sup>-1</sup>, KBr): 1656 ( $\upsilon_{C=O}$ ), 1604 ( $\upsilon_{C=C}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 8.78 (d, *J* = 4.5 Hz, 1H, H<sub>6</sub>); 8.08 (d, *J* = 8 Hz, 1H, H<sub>3'</sub>); 8.02 (d, 1H, H<sub>4'</sub>); 8.01 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 7.79 (d, *J* = 16 Hz, 1H, H<sub>a</sub>); 7.67–7.66 (m, 1H, H<sub>5'</sub>); 7.65 (d, *J* = 9 Hz, 2H, H<sub>2</sub> and H<sub>6</sub>); 6.77 (d, *J* = 9 Hz, 2H, H<sub>3</sub> and H<sub>4</sub>); 3.02 (s, 6H, 2 × CH<sub>3</sub>). MS (ESI, positive scan): *m/z* 255.35 [M+H]<sup>+</sup>.

*1-(Pyridine-2-yl)-3-(3,4-dimethoxyphenyl)-2-propene-1-one* (**p**<sub>3</sub>). Mp: 105 °C; IR (vcm<sup>-1</sup>, KBr): 1666 ( $\upsilon_{C=O}$ ); 1596 ( $\upsilon_{C=C}$ ); 1271 ( $\upsilon_{C-O}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 8.80 (m, 1H, H<sub>6</sub>'); 8.13 (d, *J* = 16 Hz; 1H, H<sub>β</sub>); 8.09 (d, *J* = 7.5 Hz, 1H, H<sub>3</sub>'); 8.04 (m, 1H, H<sub>4</sub>'); 7.82 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.68 (m, 1H, H<sub>5</sub>'); 7.40 (d, *J* = 8 Hz, 1H, H<sub>2</sub>); 7.37 (d, *J* = 1.5 Hz, 1H, H<sub>6</sub>); 7.04 (d, *J* = 8 Hz, 1H, H<sub>5</sub>); 3.83 (s, 6H, 2 × -OCH<sub>3</sub>). MS (ESI, positive scan): *m/z* 292.34 [M+Na]<sup>+</sup>.

*1-(Pyridine-2-yl)-3-(3,4,5-trimethoxyphenyl)-2-propene-1-one* ( $\mathbf{p}_4$ ). Mp: 160 °C; IR (vcm<sup>-1</sup>, KBr): 1668 ( $\upsilon_{C=O}$ ); 1610 ( $\upsilon_{C=C}$ ); 1124 ( $\upsilon_{C-O}$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>,  $\delta$  ppm): 8.76 (m, 1H, H<sub>6</sub>'); 8.20 (s, 1H, H<sub>3</sub>'); 8.17 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 7.89 (t, 1H, H<sub>4</sub>'); 7.87 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.51 (m, 1H, H<sub>5</sub>'); 6.96 (s, 2H, H<sub>2</sub> and H<sub>6</sub>); 3.94 (s, 6H, 2 × -OCH<sub>3</sub>); 3.91 (s, 3H, -OCH<sub>3</sub>). MS (ESI, positive scan): *m/z* 322.14 [M+H]<sup>+</sup>.

*1-(Pyridine-2-yl)-3-(2-hydroxyphenyl)-2-propene-1-one* (**p**<sub>5</sub>). Mp: 140 °C; IR (vcm<sup>-1</sup>, KBr): 1660 ( $\upsilon_{C=O}$ ); 1591 ( $\upsilon_{C=C}$ ); 3375 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$ ppm): 10.32 (s, 1H, -OH); 8.79 (m, 1H, H<sub>6</sub>'); 8.30 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 8.10 (d, *J* = 1 Hz, 6.5 Hz, 1H, H<sub>3</sub>'); 8.08 (m, 1H, H<sub>4</sub>'); 8.04 (m, 1H, H<sub>5</sub>'); 7.70 (d, *J* = 1 Hz, 7.5 Hz, 1H, H<sub>3</sub>); 7.67 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.29 (m, 1H, H<sub>4</sub>); 6.96 (d, *J* = 1 Hz, 8 Hz, 1H, H<sub>6</sub>); 6.90 (t, 1H, H<sub>5</sub>). MS (ESI, positive scan): *m/z* 248.07 [M+Na]<sup>+</sup>.

*1-(Pyridine-2-yl)-3-(3-hydroxyphenyl)-2-propen-1-one* (**p**<sub>6</sub>). Mp: 131 °C; IR (vcm<sup>-1</sup>, KBr): 1660 ( $\upsilon_{C=O}$ ); 1566 ( $\upsilon_{C=C}$ ); 3359 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 9.70 (s, 1H, OH); 8.80 (d, *J* = 4.5 Hz, 1H, H<sub>6</sub>'); 8.19 (d, *J* = 16.5 Hz, 1H, H<sub>β</sub>); 8.10 (d, *J* = 8 Hz, 1H, H<sub>3</sub>'); 8.05 (t, 1H, H<sub>4</sub>'); 7.75 (d, *J* = 16 Hz, 1H, H<sub>a</sub>); 7.69 (m, 1H, H<sub>5</sub>'); 7.27 (t, 1H, H<sub>5</sub>); 7.23 (d, *J* = 8 Hz, 1H, H<sub>6</sub>); 7.18 (s, 1H, H<sub>2</sub>); 6.88 (d, *J* = 1 Hz, 7.5 Hz, 1H, H<sub>4</sub>). MS (ESI, positive scan): *m/z* 248.07 [M+Na]<sup>+</sup>.

*l-(pyridine-2-yl)-3-(4-hydroxyphenyl)-2-propene-1-one* ( $\mathbf{p}_7$ ). Mp: 137 °C; IR (vcm<sup>-1</sup>, KBr): 1664 ( $\upsilon_{C=O}$ ); 1579 ( $\upsilon_{C=C}$ ); 3100 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 10.12 (s, 1H, -OH); 8.77 (d, *J* = 5.5 Hz, 1H, H<sub>6</sub>); 8.08 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 8.03 (m, 2H, H<sub>3</sub>', H<sub>4</sub>'); 7.77 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.71 (m, 1H, H<sub>5</sub>'); 7.69 (d, 2H, H<sub>2</sub>, H<sub>6</sub>); 6.84 (d, 2H, H<sub>3</sub>, H<sub>5</sub>). MS (ESI, positive scan): *m/z* 248.07 [M+Na]<sup>+</sup>.

*1-(Furan-2-yl)-3-(3-nitrophenyl)-2-propene-1-one* ( $\mathbf{f}_1$ ). Mp: 185 °C; IR (vcm<sup>-1</sup>, KBr): 1656 ( $\upsilon_{C=0}$ ); 1604 ( $\upsilon_{C=C}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 8.73 (s, 1H, H<sub>5</sub><sup>-</sup>); 8.31 (d, *J* = 8 Hz, 1H, H<sub>4</sub>); 8.27 (d, *J* = 2 Hz, 8 Hz, 1H, H<sub>6</sub>); 8.10 (t, 1H, H<sub>2</sub>); 7.94 (d, *J* = 3.5 Hz, 1H, H<sub>3</sub><sup>-</sup>); 7.91 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 7.85 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.76 (t, 1H, H<sub>5</sub>); 6.82 (m, 1H, H<sub>4</sub><sup>-</sup>). MS (ESI, positive scan): *m/z* 266.2 [M+Na]<sup>+</sup>.

*1-(Furan-2-yl)-3-(4-dimethylaminophenyl)-2-propene-1-one* (**f**<sub>2</sub>). Mp: 104 °C; IR (vcm<sup>-1</sup>, KBr): 1643 ( $\nu_{C=O}$ ); 1612 ( $\nu_{C=C}$ ). <sup>1</sup>H-NMR (DMSO-<sub>*d*6</sub>,  $\delta$  ppm): 8.00 (d, *J* = 1 Hz, 1H, H<sub>5</sub><sup>-</sup>); 7.67 (d, *J* = 16 Hz, 1H,

H<sub>β</sub>); 7.66 (d, J = 9 Hz, 3H, H<sub>2</sub>, H<sub>6</sub> and H<sub>3'</sub>); 7.40 (d, J = 16 Hz, 1H, H<sub>α</sub>); 6.76 (m, 3H, H<sub>3</sub>, H<sub>5</sub> and H<sub>4'</sub>); 3.00 (s, 6H, 2 × -CH<sub>3</sub>). MS (ESI, positive scan): m/z 242.2 [M+H]<sup>+</sup>.

*l-(Furan-2-yl)-3-(3,4-dimethoxyphenyl)-2-propene-1-one* (**f**<sub>3</sub>). Mp: 105 °C; IR (vcm<sup>-1</sup>, KBr): 1651 ( $\upsilon_{C=O}$ ); 1595 ( $\upsilon_{C=C}$ ); 1263( $\upsilon_{C-O}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 8.04 (d, *J* = 1 Hz, 1H, H<sub>5</sub>'); 7.77 (d, *J* = 3 Hz; 1H, H<sub>3</sub>'); 7.70 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 7.56 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.47 (s, 1H, H<sub>2</sub>); 7.37 (m, 1H, H<sub>6</sub>); 7.02 (d, *J* = 8.5 Hz, 1H, H<sub>5</sub>); 6.78 (m, 1H, H<sub>4</sub>'); 3.83 (s, 6H, 2 × -OCH<sub>3</sub>). MS (ESI, positive scan): *m/z* 281.4 [M+Na]<sup>+</sup>.

*1-(Furan-2-yl)-3-(3,4,5-trimethoxyphenyl)-2-propene-1-one* (**f**<sub>4</sub>). Mp: 158 °C; IR (vcm<sup>-1</sup>, KBr): 1654 ( $\upsilon_{C=O}$ ); 1604 ( $\upsilon_{C=C}$ ); 1122 ( $\upsilon_{C-O}$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>,  $\delta$  ppm): 7.80 (d, *J* = 16 Hz, 1H, H<sub> $\beta$ </sub>); 7.66 (d, *J* = 1 Hz, 1H, H<sub>5</sub><sup>-</sup>); 7.34 (d, *J* = 16 Hz, 1H, H<sub> $\alpha$ </sub>); 7.34 (d, *J* = 3.5 Hz, 1H, H<sub>3</sub><sup>-</sup>); 6.88 (s, 2H, H<sub>2</sub> and H<sub>6</sub>); 6.61 (m, 1H, H<sub>4</sub><sup>-</sup>); 3.93 (s, 6H, 2 × -OCH<sub>3</sub>); 3.90 (s, 3H, -OCH<sub>3</sub>). MS (ESI, positive scan): *m/z* 311.06 [M+Na]<sup>+</sup>.

*1-(Furan-2-yl)-3-(2-hydroxyphenyl)-2-propene-1-one* (**f**<sub>5</sub>). Mp: 138 °C; IR (vcm<sup>-1</sup>, KBr): 1643 ( $\upsilon_{C=0}$ ), 1577 ( $\upsilon_{C=C}$ ); 3124 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 10.24 (s, 1H, -OH); 8.03 (d, *J* = 16 Hz, 1H, H<sub>\beta</sub>); 8.03 (d, *J* = 1 Hz, 7.5 Hz, 1H, H<sub>5'</sub>); 7.78 (d, *J* = 1.5 Hz, 6 Hz, 1H, H<sub>\beta</sub>'); 7.67 (d, *J* = 3.5 Hz, 1H, H<sub>\beta</sub>'); 7.64 (d, *J* = 16 Hz, 1H, H<sub>\alpha</sub>); 7.27 (m, 1H, H<sub>\alpha</sub>); 6.93 (d, *J* = 0.5 Hz, 8Hz, 1H, H<sub>\alpha</sub>); 6.87 (t, 1H, H<sub>\alpha</sub>'); 6.76 (m, 1H, H<sub>\beta</sub>). MS (ESI, positive scan): *m/z* 248.07 [M+Na]<sup>+</sup>.

*1-(Furan-2-yl)-3-(3-hydroxyphenyl)-2-propene-1-one* (**f**<sub>6</sub>). Mp: 141 °C; IR (vcm<sup>-1</sup>, KBr): 1643 ( $\upsilon_{C=0}$ ); 1577 ( $\upsilon_{C=C}$ ); 3309 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 9.61 (s, 1H, -OH); 8.05 (d, 1H, H<sub>5</sub>'); 7.79 (d, 1H, H<sub>3</sub>'); 7.65 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 7.58 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.25 (t, 1H, H<sub>4</sub>'); 7.25 (d, 1H, H<sub>6</sub>); 7.19 (s, 1H, H<sub>2</sub>); 6.87 (m, 1H, H<sub>5</sub>); 6.78 (m, 1H, H<sub>4</sub>). MS (ESI, positive scan): *m/z* 248.07 [M+Na]<sup>+</sup>.

*1-(Furan-2-yl)-3-(4-hydroxyphenyl)-2-propene-1-one* ( $\mathbf{f}_7$ ). Mp: 142 °C; IR (vcm<sup>-1</sup>, KBr): 1647 ( $\upsilon_{C=0}$ ); 1610 ( $\upsilon_{C=C}$ ); 3200 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 10.09 (s, 1H, -OH); 8.02 (s, 1H, H<sub>5'</sub>); 7.72 (s, 1H, H<sub>3'</sub>); 7.71(d, J = 7.5 Hz, 2H, H<sub>2</sub> and H<sub>6</sub>); 7.66 (d, J = 17 Hz, 1H, H<sub>β</sub>); 7.48 (d, J = 16.5 Hz, 1H, H<sub>α</sub>); 6.83 (m, 2H, H<sub>3</sub> and H<sub>5</sub>); 6.76 (m, 1H, H<sub>4'</sub>). MS (ESI, positive scan): *m/z* 248.06 [M+Na]<sup>+</sup>.

*1-(Thiophene-2-yl)-3-(3-nitrophenyl)-2-propene-1-one* (t<sub>1</sub>). Mp: 159 °C; IR (vcm<sup>-1</sup>, KBr): 1651 ( $\upsilon_{C=O}$ ); 1600 ( $\upsilon_{C=C}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 8.77 (d, J = 1.5 Hz, 1H, H<sub>5</sub>'); 8.43 (d, J = 5 Hz, 1H, H<sub>3</sub>'); 8.33 (d, J = 8 Hz, 1H, H<sub>4</sub>); 8.27 (d, J = 2 Hz, 8 Hz, 1H, H<sub>6</sub>); 8.09 (d, J = 16 Hz, 1H, H<sub>β</sub>); 8.10 (d, J = 1 Hz, 1H, H<sub>2</sub>); 7.85 (d, J = 15.5 Hz, 1H, H<sub>α</sub>); 7.77 (t, 1H, H<sub>5</sub>); 7.54 (m, 1H, H<sub>4</sub>'). MS (ESI, positive scan): m/z 282.02 [M+Na]<sup>+</sup>.

*1-(Thiophene-2-yl)-3-(4-dimethylaminophenyl)-2-propene-1-one* (t<sub>2</sub>). Mp: 131 °C; IR (vcm<sup>-1</sup>, KBr): 1633 ( $\upsilon_{C=0}$ ); 1612 ( $\upsilon_{C=C}$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>,  $\delta$  ppm): 7.83 (d, *J* = 15.5 Hz, 1H, H<sub> $\beta$ </sub>); 7.83 (d, *J* = 9 Hz, 1H, H<sub>5</sub>); 7.62 (d, *J* = 6 Hz, 1H, H<sub>3</sub>); 7.55 (d, *J* = 9 Hz, 2H, H<sub>2</sub> and H<sub>6</sub>); 7.23 (d, *J* = 15.5 Hz, 1H, H<sub> $\alpha$ </sub>); 7.16 (t, 1H, H<sub>4</sub>); 6.70 (d, *J* = 8.5 Hz, 2H, H<sub>3</sub> and H<sub>5</sub>); 3.04 (s, 6H, 2 × -CH<sub>3</sub>). MS (ESI, positive scan): *m/z* 258.08 [M+H]<sup>+</sup>.

*1-(Thiophene-2-yl)-3-(3,4-dimethoxyphenyl)-2-propene-1-one* ( $t_3$ ). Mp: 99 °C; IR (vcm<sup>-1</sup>, KBr): 1639 ( $v_{C=0}$ ); 1573 ( $v_{C=C}$ ); 1265 ( $v_{C-0}$ ). <sup>1</sup>H-NMR (DMSO- $d_6$ ,  $\delta$  ppm): 8.32 (m, 1H, H<sub>5'</sub>); 8.03 (m, 1H, H<sub>3'</sub>); 7.75

(d, J = 15.5 Hz, 1H, H<sub> $\beta$ </sub>); 7.68 (d, J = 15.5 Hz, 1H, H<sub> $\alpha$ </sub>); 7.52 (s, 1H, H<sub>2</sub>); 7.40 (m, 1H, H<sub>6</sub>); 7.31 (m, 1H, H<sub>4</sub>·); 7.02 (d, J = 8.5 Hz, 1H, H<sub>5</sub>); 3.83 (s, 6H, 2 × -OCH<sub>3</sub>). MS (ESI, positive scan): m/z 297.02 [M+Na]<sup>+</sup>.

*1-(Thiophene-2-yl)-3-(3,4,5-trimethoxyphenyl)-2-propene-1-one* (**t**<sub>4</sub>). Mp: 156 °C; IR (vcm<sup>-1</sup>, KBr): 1645 ( $\upsilon_{C=O}$ ); 1596 ( $\upsilon_{C=C}$ ); 1124 ( $\upsilon_{C-O}$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>,  $\delta$  ppm): 7.88 (m, 1H, H<sub>5</sub>'); 7.77 (d, *J* = 15.5 Hz, 1H, H<sub> $\beta$ </sub>); 7.69 (d, *J* = 1 Hz, 5 Hz, 1H, H<sub>3</sub>'); 7.30 (d, *J* = 15.5 Hz, 1H, H<sub> $\alpha$ </sub>); 7.19 (t, 1H, H<sub>4</sub>'); 6.87 (s, 2H, H<sub>2</sub> and H<sub>6</sub>); 3.93 (s, 6H, 2 × -OCH<sub>3</sub>); 3.91 (s, 3H, -OCH<sub>3</sub>). MS (ESI, positive scan): *m/z* 327.04 [M+Na]<sup>+</sup>.

*1-(thiophene-2-yl)-3-(2-hydroxyphenyl)-2-propene-1-one* (t<sub>5</sub>). Mp: 158 °C; IR (vcm<sup>-1</sup>, KBr): 1637 ( $\nu_{C=O}$ ); 1560 ( $\nu_{C=C}$ ); 3332 ( $\nu_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 10.30 (s, 1H, -OH); 8.20 (d, *J* = 1 Hz, 3 Hz, 1H, H<sub>5</sub>'); 8.04 (d, *J* = 16 Hz, 1H, H<sub>β</sub>); 8.03 (d, 1H, H<sub>6</sub>); 7.86 (d, *J* = 1.5 Hz, 8 Hz, 1H, H<sub>3</sub>); 7.77 (d, *J* = 15.5 Hz, 1H, H<sub>α</sub>); 7.26–7.30 (m, 2H, H<sub>4</sub>, H<sub>5</sub>); 6.94 (d, *J* = 1 Hz, 4Hz, H<sub>3</sub>); 6.88 (t, 1H, H<sub>4'</sub>). MS (ESI, positive scan): *m/z* 253. [M+Na]<sup>+</sup>.

*1-(Thiophene-2-yl)-3-(3-hydroxyphenyl)-2-propene-1-one* (**t**<sub>6</sub>). Mp: 130 °C; IR (vcm<sup>-1</sup>, KBr): 1643 ( $\upsilon_{C=O}$ ); 1577 ( $\upsilon_{C=C}$ ); 3334 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$ ppm): 9.62 (s, 1H, -OH); 8.31 (d, *J* = 4 Hz, 1H, H<sub>5'</sub>); 8.05 (d, *J* = 5 Hz, 1H, H<sub>3'</sub>); 7.77 (d, *J* = 15.5 Hz, 1H, H<sub>β</sub>); 7.63 (d, *J* = 16 Hz, 1H, H<sub>α</sub>); 7.31 (t, 1H, H<sub>4'</sub>); 7.30 (d, 1H, H<sub>6</sub>); 7.27 (t, 1H, H<sub>5</sub>); 7.23 (s, 1H, H<sub>2</sub>); 6.87 (m, 1H, H<sub>4</sub>). MS (ESI, positive scan): *m/z* 253. [M+Na]<sup>+</sup>.

*1-(Thiophene-2-yl)-3-(4-hydroxyphenyl)-2-propene-1-one* ( $t_7$ ). Mp: 151 °C; IR (vcm<sup>-1</sup>, KBr): 1643 ( $\upsilon_{C=O}$ ); 1610 ( $\upsilon_{C=C}$ ); 3172 ( $\upsilon_{O-H}$ ). <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>,  $\delta$  ppm): 10.09 (s, 1H, -OH); 8.25 (m, 1H, H<sub>5'</sub>); 8.01 (m, 1H, H<sub>3'</sub>); 7.73 (m, 2H, H<sub>2</sub> and H<sub>6</sub>); 7.65 (s, 2H, H<sub>β</sub> and H<sub>α</sub>); 6.83 (m, 2H, H<sub>3</sub> and H<sub>5</sub>). MS (ESI, positive scan): *m/z* 253. [M+Na]<sup>+</sup>.

# 3.3. Antibacterial Activity

## 3.3.1. Antibiotics and Antibacterial Agents

Standard powder forms of vancomycin and ciprofloxacin (Sigma Chemical Co., Ltd.), chloramphenicol, doxycycline and gentamicin (Shanghai Fine Chemicals Co., Ltd.), were stored at 2 to 8 °C until use. Antibiotics discs like gentamycin (10  $\mu$ g), cloramphenicol (30  $\mu$ g), vancomycin (30  $\mu$ g), clindamycin (2  $\mu$ g), doxycycline (30  $\mu$ g), ciprofloxacin (5  $\mu$ g) and erythromycin (15  $\mu$ g) were supplied by Nam Khoa Co., Ltd., Vietnam.

Three standard strains of bacteria: *Staphylococcus aureus* ATCC 25923 and methicillin-resistant *Staphylococcus aureus* ATCC 43300 and isolated MDR MRSA preserved and activated atour Department of Microbiology were used in the study. The bacterial culture media were purchased from Merck. Tryptic Soy Agar (TSA) and Tryptic Soy Broth (TSB) were used for bacteria culture and Mueller-Hinton Agar (MHA) was used for testing antibacterial activity.

# 3.3.2. Antibacterial Susceptibility Testing

The disc diffusion method was carried out for the antibacterial tests [31]. Briefly a suspension of bacterial strains (100  $\mu$ L) containing 10<sup>6</sup> cfu/mL of bacteria was spread on Mueller Hinton agar

(MHA) medium. The disc (6 mm in diameter), impregnated with 10  $\mu$ L of the test compound at the concentration of 102.4 mg/mL in DMSO were placed on the inoculated agar. Negative control was prepared using the same solvent (DMSO), which was employed to dissolve the test compounds. Antibiotics (2–30  $\mu$ g/disc, 6 mm in diameter) were used as references. The inoculated plates were incubated at 35–37 °C for 24 h. The diameter of inhibitory zones (mm) was measured (Table 2).

#### 3.2.3. Qualitative Determination of the Interaction between the Chalcones and Antibiotics

A routine Kirby-Bauer antibiotics susceptibility test [32,33] was performed to determine the inhibitory zone of selected compounds. Briefly, sterile MHA medium is melted and poured into Petri dishes to get an agar layer about 3–4 millimeters thick. A sterile cotton stick is dipped into the prepared bacterial suspension (prepared to a final concentration of approximately  $10^8$  CFU/mL), then pressed on the tube wall to dry, then the bacteria are spread on the agar surface evenly. Opened boxes are put in the incubator in 3–5 min to drain. Wells (6 mm in diameter) were punched in the agar. The distances from the hole (synthesized chalcone analogues) and disc (antibiotics) were measured so that margins of two antibiotic inhibitory zones would meet or slightly overlap. The chalcone samples prepared in solutions at rational concentration (1,024 µg/mL) in DMSO were filled into the wells. After allowing the test compounds to diffuse into agar (15 min at 37 °C) the plates were further incubated at 37 °C for a period of 18 h (for MSSA) or 24 h (for MRSA). A positive interaction between a specific antibiotic and a test substance was reported when the inhibitory zone of each compound extended towards the other (Table 3).

#### 3.2.4. Measurement of MIC Values

The MICs of antibiotics and selected active chalcones were determined by the microdilution method as described by The National Committee for Clinical Laboratoty Standard (NCCLS) [34,35]. Each test compound was run in duplicate. The test plates were incubated at 35–37 °C for 24 h. The MIC was taken as the minimum concentration of the dilutions that inhibited the growth of the test microorganism. The concentration of the solvents used in the following assays was maintained at less than 2% so that no inhibition of organisms or interference occurred.

#### 3.2.5. Evaluation of Combined Activity

Antibacterial interactions were determined using the checkerboard assay as previously described [36]. The range of tested compounds concentration used in the checkerboard analysis was such that the dilution range encompassed the MIC for each drug used in the analysis. The first compound of the combination was serially diluted along the ordinate, while the second compound was diluted along the abscissa. Broth micro-dilution plates were inoculated with each *S. aureus* strain to yield ~5 × 10<sup>5</sup> CFU/mL in a 100  $\mu$ L final volume, and incubated for 18 h–24 h at 37 °C. Synergy has been defined as requiring a fourfold reduction in the MIC of both antibiotics in combination, compared with each used alone, measuring the fractional inhibitory concentration index (FICI). The FICI was calculated for each combination (A and B as tested compounds) using the following formula [36]:

$$FICI = \frac{MIC_{A}(combination)}{MIC_{A}(alone)} + \frac{MIC_{B}(combination)}{MIC_{B}(alone)}$$

The FICI was interpreted as follows: synergistic, FICI  $\leq 0.5$ ; additive,  $0.5 < \text{FICI} \leq 1$ ; indifferent,  $1 < \text{FICI} \leq 2$ ; antagonistic, FICI > 2.

# 4. Conclusions

The study has proved that the synthesized heterocyclic chalcone analogues have some anti-*Staphyloccus aureus* effects. The results, based on the potentially active chalcone skeleton, have pointed out the importance of the positions of the phenolic hydroxy groups in the B ring for obtaining antibacterial activity. In addition, the furan-2-yl moiety is more active than either the thiophene-2-yl or pyridine-2-yl one in anti-*Staphylococcus aureus* activity. Compounds  $\mathbf{p}_5$ ,  $\mathbf{f}_6$  and  $\mathbf{t}_5$  exhibited potent inhibitory activity against methicillin-resistant *Staphylococcus aureus* in combination with vancomycin and oxacillin. They may provide a template to design new combinations of antibiotics and non-antibiotics to treatment of *Staphylococcus aureus* infections.

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## **Conflict of Interest**

The authors declare no conflict of interest.

# **References and Notes**

- 1. Neu, H.C. The crisis in antibiotic resistance. *Science* 1992, 257, 1064–1073.
- 2. Michel, M.; Gutmann, L. Methicillin-resistant Staphyllococus aureus and vancomycin-resistan enterococci: therapeut realities and possibilities. *Lancet* **1997**, *349*, 1901–1906.
- 3. Normark, B.H.; Normark, S. Evolution and spread of antibiotic resistance. *J. Intern. Med.* 2002, *252*, 91–106.
- Shekarchi, M.; Pirali-Hamedani, M.; Navidpour, L.; Adib, N.; Shafiee, A. Synthesis, Antibacterial and Antifungal Activities of 3-Aryl-5-(pyridin-3-yl)-4,5-dihydropyrazole-1-carbothioamide Derivatives. J. Iran. Chem. Soc. 2008, 5, 150–158.
- 5. Silverman, R.B. *Organic Chemistry of Drug Design and Drug Action*; Academic Press: San Diego, CA, USA, 1992.
- Thompson, L.A.; Ellman, J.A. Synthesis and applications of small molecule libraries. *Chem. Rev.* 1996, 96, 555–600.
- 7. Alcaraz, L.E.; Blanco, S.E.; Puig, O.N.; Tomas, F.; Ferretti, F.H. Antibacterial activity of Flavonoids against methicillin- resistant *Staphylococcus aureus* strains. *J. Theor. Biol.* **2000**, *205*, 231–240.

- 8. Baviskar, B.A.; Baviskar, B.; Shiradkar, M.R.; Deokate, U.A.; Khadabadi, S.S. Antimicrobial activity of some novel. Benzimidazolyl chalcones. *E-J. Chem.* **2009**, *6*, 196–200.
- 9. Echeverria, C.; Santibañez, J.F.; Donoso-Tauda, O.; Escobar, C.A.; Ramirez-Tagle, R. Structural antitumoral activity relationships of synthetic chalcones. *Int. J. Mol. Sci.* **2009**, *10*, 221–231.
- 10. Modzelewska, A.; Pettit, C.; Achanta, G.; Davidson, N.E.; Huang, P.; Khan S.R. Anticancer activities of novel chalcone and bis-chalcone derivatives. *Bioorg. Med. Chem.* **2006**, *14*, 3491–3495.
- 11. Vogel, S.; Heilmann, J. Synthesis, cytotoxicity, and antioxidative activity of minor prenylated chalcones from Humulus lupulus. *J. Nat. Prod.* **2008**, *71*, 1237–1241.
- Babasaheb, P.B.; Sachin, A.P.; Rajesh, N.G. Synthesis and biological evaluation of nitrogencontaining chalcones as possible anti-inflammatory and antioxidant agents. *Bioorg. Med. Chem. Lett.* 2010, 20, 730–733.
- 13. Vogel, S.; Barbic, M.; Jürgenliemk, G.; Heilmann, J. Synthesis, cytotoxicity, anti-oxidative and anti-inflammatory activity of chalcones and influence of A-ring modifications on the pharmacological effect. *Eur. J. Med. Chem.* **2010**, *45*, 2206–2213.
- Tran, T.-D.; Park, H.; Kim, H.P.; Ecker, G.F.; Thai, K.-M. Inhibitory activity of prostaglandin E2 production by the synthetic 2'-hydroxychalcone analogues: Synthesis and SAR study. *Bioorg. Med. Chem. Lett.* 2009, 19, 1650–1653.
- Beom-Tae, K.; Kwang-Joong, O.; Jae-Chul, C.; Ki-Jun, H. Synthesis of dihydroxylated chalcone derivatives with diverse substitution patterns and their radical scavenging ability toward DPPH free radicals. *Bull. Korean Chem. Soc.* 2008, 29, 1125–1130.
- 16. Doan, T.N.; Tran, T.-D. Synthesis, antioxidant and antimicrobial activities of a novel series of chalcones, pyrazolic chalcones, and allylic chalcones. *Pharmacol. Pharm.* **2011**, *2*, 282–288.
- 17. Sivakumar, P.M.; Prabhakar, P.K.; Doble, M. Synthesis, antioxidant evaluation, and quantitative structure–activity relationship studies of chalcones. *Med. Chem. Res.* **2011**, *20*, 482–492.
- 18. Vogel, S.; Ohmayer, S.; Brunner, G.; Heilmann, J. Natural and non-natural prenylated chalcones: synthesis, cytotoxicity and anti-oxidative activity. *Bioorg. Med. Chem.* **2008**, *16*, 4286–4293.
- 19. Munawar, M.A.; Azad, M.; Siddiqui, H.L. Synthesis and antimicrobial studies of some quinolinylpyrimidine derivatives. *J. Chin. Soc.* **2008**, *55*, 394–400.
- 20. Azad, M.; Munawar, M.A.; Siddiqui, H.L. Antimicrobial activity and synthesis of quinoline-base chalcones. *J. Appl. Sci.* **2007**, *7*, 2485–2489.
- 21. Kalirajan, R.; Sivakumar, S.U.; Jubie, S.; Gowramma, B.; Suresh, B. Synthesis and biological evaluation of some heterocyclic derivatives of chalcones. *Int. J. ChemTech Res.* **2009**, *1*, 27–34.
- Talia, J.M.; Debattista, N.B.; Pappano, N.B. New antimicrobial combinations: Substituted chalcones- oxacillin against methicillin resistant Staphylococcus aureus. *Braz. J Microbiol.* 2011, 42, 470–475.
- 23. Eumkeb, G.; Siriwong, S.; Phitaktim, S.; Rojtinnakorn, N.; Sakdarat, S. Synergistic activity and mode of action of flavonoids isolated from smaller galangal and amoxicillin combinations against amoxicillin-resistant Escherichia coli. *J. Appl. Microbiol.* **2012**, *112*, 55–64.
- Sato, Y.; Shibata, H.; Arakaki, N.; Higuti, T. 6,7-dihydroxyflavone dramatically intensifies the susceptibility of methicillin-resistant or -sensitive Staphylococcus aureus to beta-lactams. *Antimicrob. Agents Chemother.* 2004, 48, 1357–1360.

- Liu, M.H.; Otsuka, N.; Noyori, K.; Shiota, S.; Ogawa, W.; Kuroda, T.; Hatano, T.; Tsuchiya, T. Synergistic effect of kaempferol glycosides purified from Laurus nobilis and fluoroquinolones on methicillin-resistant Staphylococcus aureus. *Biol. Pharm. Bull.* 2009, *32*, 489–492.
- 26. Tiwari, B.; Pratapwar, A.S.; Tapas, A.R.; Butle, S.R.; Vatkar, B.S. Synthesis and antimicrobial activity of some chalcone derivatives. *Int. J. ChemTech Res.* **2010**, *2*, 499–503.
- Sivakumar, P.M.; Priya, S.; Doble, M. Synthesis, biological evaluation, mechanism of action and quantitative structure-activity relationship studies of chalcones as antibacterial agents. *Chem. Biol. Drug Des.* 2009, 73, 403–415.
- Zheng, C.J.; Jiang, S.M.; Chen, Z.H.; Ye, B.J.; Piao, H.R. Synthesis and anti-bacterial activity of some heterocyclic chalcone derivatives bearing thiofuran, furan, and quinoline moieties. *Arch. Pharm. Chem. Life Sci.* 2011, 344, 689–695.
- Lopez, S.N.; Castelli, M.V.; Zacchino, S.A.; Domnguez, J.N.; Lobo, G.; Charris-Charris, J.; Cortes, J.C.G.; Ribas, J.C.; Devia, C.; Rodrguez, A.M.; *et al. In Vitro* Antifungal evaluation and structure–activity relationships of a new series of chalcone derivatives and synthetic analogues, with inhibitory properties against polymers of the fungal cell wall. *Bioorg. Med. Chem.* 2001, *9*, 1999–2013.
- 30. Funiss, B.S.; Hannford, A.J.; Smith, P.W.G.; Tatchell, A.R. *Vogel's Textbook of Practical Organic Chemistry*, 5th ed.; Prentice Hall: Upper Saddle River: NJ, USA, 2004; pp. 1032–1035.
- National Committee for Clinical Laboratory Standards. *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts*. Approved Standard M27–A2, 3rd ed.; NCCLS: Wayne, PA, USA, 2002; Volume 22, No. 15, pp. 1–30.
- 32. Bauer, A.W.; Kirby, W.M.M.; Sherris, J.C.; Turck, M. Antibiotic Susceptibility Testing by a Standardized Single Disk Method. *Am. J. Clin. Pathol.* **1966**, *36*, 493–496.
- Jorgensen, J.H.; Turnidge, J.D. *Manual of Clinical Microbiology*, 9th ed.; Murray, P.R., Baron, E.J., Jorgensen, J.H., Landry, M.L., Pfaller, M.A., Eds.; ASM Press: Washington, DC, USA, 2007; pp. 1152–1172.
- National Committee for Clinical Laboratory Standards. *Methods for Dilution Antimicrobial* Susceptibility Tests for Bacteria that Grow Aerobically. Approved standard M7-A6, 6th ed.; NCCLS: Wayne, PA, USA, 2003.
- 35. National Committee for Clinical Laboratory Standards. *Performance Standards for Antimicrobial Susceptibility Testing*. Informational supplement M100-S13, 12th ed.; NCCLS: Wayne, PA, USA, 2003.
- Bajaksouzian, S.; Visalli, M.A.; Jacobs, MR.; Appelbaum, PC. Activities of levofloxacin, ofloxacin, and ciprofloxacin, alone and in combination with amikacin, against acinetobacters as determined by checkerboard and time-kill studies. *Antimicrob. Agents Chemother.* 1997, 41, 1073–1076.

Sample Availability: Samples of the compounds  $p_{1-7}$ ,  $f_{1-7}$  and  $t_{1-7}$  are available from the authors.

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