REVIEW



Dermatologic Lesions with Eosinophilia in the Head and Neck

David T. Danielson¹ · Ian Lagerstrom¹ · Zachary Wary¹ · Aaron Auerbach² · David S. Cassarino³

Received: 1 December 2024 / Accepted: 23 January 2025

This is a U.S. Government work and not under copyright protection in the US; foreign copyright protection may apply 2025

Abstract

Background Dermatologic lesions with notable eosinophilic infiltration of the head and neck region represent a diverse group of conditions, ranging from benign to malignant proliferations.

Methods We performed a comprehensive literature review focusing on head and neck dermatologic conditions that commonly present with a prominent eosinophilic infiltrate.

Results This review provides an overview of common entities showing prominent associated eosinophilic inflammatory infiltrates in this region, including epithelioid hemangioma, eosinophilic cellulitis (Wells syndrome), eosinophilic folliculitis, eosinophilic granulomatosis with polyangiitis (Churg-Strauss syndrome), granuloma faciale, and Langerhans cell histiocytosis (LCH).

Conclusion Eosinophils play a key role in the pathogenesis of these disorders, although the exact mechanisms remain poorly understood. Accurate diagnosis is crucial for differentiating these conditions, as they can share similar histologic features. This review aims to enhance understanding of these eosinophilic dermatologic conditions, improving diagnostic accuracy and treatment strategies.

Keywords Eosinophilia · Dermatologic pathology · Head and neck pathology

Introduction

The head and neck region is frequently affected by dermatologic conditions characterized by eosinophilic infiltration. Each can present with similar histologic features but differ significantly in their clinical behaviors, ranging from benign and self-limiting to severe and life-threatening. The presence of eosinophils in these conditions provides a valuable diagnostic clue, yet challenges remain in distinguishing between the different entities based upon clinical and/or histopathologic findings alone. This review article aims to provide a comprehensive overview of the most common eosinophilic dermatologic lesions in the head and neck area, including epithelioid hemangioma (EH), eosinophilic cellulitis (EC), eosinophilic pustular folliculitis (EPF), eosinophilic

granulomatosis with polyangiitis (EGPA), granuloma faciale (GF), and Langerhans cell histiocytosis (LCH). By examining clinical presentations, histopathological features, and potential differential diagnoses, the authors aim to guide clinicians and pathologists in the accurate identification and management of these complex disorders. Furthermore, we explore the underlying immunologic and inflammatory mechanisms that drive eosinophil infiltration in these conditions and discuss emerging therapeutic strategies.

Epithelioid Hemangioma

EH, also known as histiocytoid hemangioma and angiolymphoid hyperplasia with eosinophilia, typically affects adults in their third to fifth decades of life with no clear sex predominance. Most lesions occur in the head and neck region with roughly one third of cases occurring around the ear [1]. EH presents as one or more red, pink, or brown papules/nodules that can coalesce to form a plaque and are typically less than one centimeter in greatest dimension [2, 3]. EH is often asymptomatic, although pruritus and bleeding may be

Published online: 25 February 2025



[☐] David T. Danielson david.t.danielson.mil@health.mil

Department of Pathology, Walter Reed National Military Medical Center, Bethesda, MD, USA

Joint Pathology Center, Silver Spring, MD, USA

Department of Pathology, Kaiser Permanente Los Angeles Medical Center, Los Angeles, CA, USA

27 Page 2 of 13 Head and Neck Pathology (2025) 19:27

present [4]. Peripheral eosinophilia is uncommon, but may be present in a small subset of cases [5].

Microscopically, EH typically appears as a well-circumscribed dermal (or deeper) proliferation of variably sized blood vessels in a distinctive lobular pattern with characteristic plump to epithelioid-appearing endothelial cells that often protrude into the lumen of vessels creating a "hobnailed" appearance (Fig. 1) [2, 3]. The vessels are surrounded by a dense mixed inflammatory infiltrate that is predominantly composed of lymphocytes and eosinophils [2, 3, 6, 7]. The lymphoid infiltrate can vary between cases and some cases will have lymphoid follicles. EH cases also vary over time with the lymphoid infiltrate typically becoming more prominent in the later stages of the disease [3]. Rare mitotic figures may be present in angiogenic foci but atypical mitotic figures should not be seen [2]. Fibrosis may be prominent in older lesions.

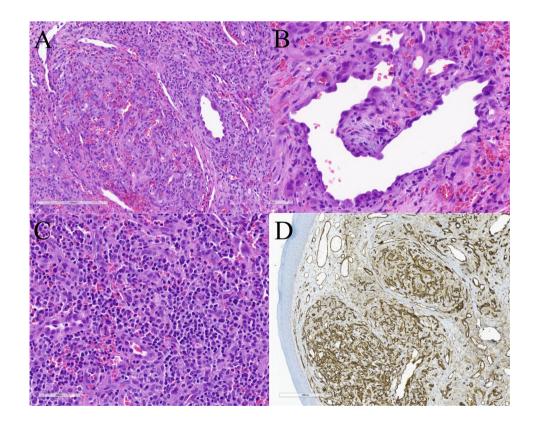
The pathogenesis of EH has not been fully elucidated, without consensus in the literature as to whether it is a reactive or neoplastic process. Many cases of EH have been observed to occur in close proximity to a damaged vessels, suggesting a reactive process that arises due to vascular damage and subsequent repair [8]. However, arguing against a reactive process, Wilms tumor 1 (WT1) immunoreactivity, which is seen in many vascular neoplasms, has also been seen in a majority of EH cases [9]. Moreover, endothelial cells of the tumor frequently show nuclear FOS and/or FOSB immunostaining pattern, and a subset of cases

have a *FOS* or *FOSB* gene rearrangement detectable by florescence in-situ hybridization (FISH) [10–13], findings suggestive of a neoplastic process. The exact mechanism of FOS and/or FOSB overexpression in EH cases that lack a detectable rearrangement remains unclear, but it may be the result of epigenetic modifications, point mutations, or histone modifications [13].

EH can have histologic overlap with both epithelioid angiosarcoma (EAS) and Kimura disease. EAS has increased mitotic activity and shows a higher degree of cytologic atypia compared to EH. Furthermore, necrosis and areas of anastomosing vessels or a sheet-like, confluent growth pattern are often present in EAS, but should not be present in EH [14]. EAS may be associated with a mild inflammatory infiltrate, but will not typically present with the brisk lymphocytic and eosinophil rich infiltrate seen in EH [15]. Although follicles may be present in EH, Kimura disease often has more prominent follicles with follicular hyperplasia and will demonstrate interfollicular eosinophilic abscesses [16]. Eosinophilic follicle lysis and hyaline material in germinal centers (due to IgE deposition) are often present [17]. Neoangiogenesis may be present in Kimura disease, but it should lack the plump to epithelioid endothelial cells seen in EH [18]. Elevated serum levels of IgE and peripheral blood eosinophilia are present in nearly all cases of Kimura disease, but are uncommon in EH [19].

Surgical resection is the most common treatment for EH, although other treatment options are available and include

Fig. 1 Epithelioid hemangioma. A Lobules of variably sized vessels lined by plump endothelial cells which range from slit like to large and dilated with a background predominantly lymphocytic inflammatory infiltrate (original magnification, ×200). B Vessel lined by plump endothelial cells that show a "hobnailed" appearance. The endothelial cells lack nuclear atypia and mitotic figures are not appreciated (original magnification, ×400). C The background inflammatory infiltrate is composed of predominantly small lymphocytes with scattered eosinophils (original magnification, ×400). D CD31 immunohistochemical staining highlights the endothelial cells and emphasizes the lobular architecture (original magnification, ×100)





Head and Neck Pathology (2025) 19:27 Page 3 of 13 27

corticosteroid injections, cryotherapy, pulsed dye laser therapy, and carbon-dioxide laser therapy [1, 20–22]. Regardless of the treatment method, local recurrence is common and occurs in the majority of cases. Of the described methods, local recurrence was lowest in cases treated with surgical excision, with a treatment failure rate of 40.8% and a mean disease-free survival of 4.2 years [1].

Eosinophilic Cellulitis

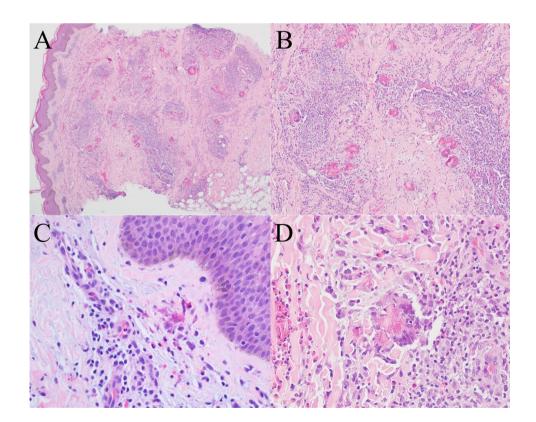
EC, also known as Wells syndrome, is a rare inflammatory dermatosis with a strikingly eosinophil-rich inflammatory infiltrate. This dermatosis was first described in 1971 as a recurrent granulomatous dermatitis with eosinophilia and later designated EC [23, 24]. There is no clear sex predominance, and it occurs at a mean age of 33.6. Clinically, it presents with large, erythematous plaques that can be either localized or diffuse. Systemic symptoms, including fever and malaise, may be present in a subset of cases [25]. Peripheral eosinophilia occurs in roughly 67% of cases, and leukocytosis is present in approximately 40% of cases [25, 26].

Histologically, EC presents with dermal edema and a diffuse eosinophilic infiltrate throughout both the superficial and deep dermis and occasionally the subcutis [24, 27, 28]. Epidermal spongiosis may be present in some cases and flame figures are often seen, which are formed by eosinophilic major basic protein from eosinophil granules

depositing on collagen fibers (Fig. 2) [27, 28]. Flame figures are a characteristic of EC, but they are not specific as they can be seen in other eosinophilic dermatoses [29]. The histologic appearance of the disease can change over time with later stages of the lesion showing a decreased eosinophilic infiltrate and the appearance of granulomatous inflammation surrounding the flame figures [27, 28].

The pathogenesis of EC is not fully understood. Cases of EC have been reported following triggering events including insect bites, viral infection, vaccination, parasitic infection, and mediation use [30-34]. EC cases have also been reported in association with an underlying disorder including hematologic and nonhematologic malignancies [35, 36], and a subset of cases are idiopathic with no identifiable triggering event or underlying disease [37–39]. The common relationship to a triggering event or underlying conditions hints that EC may be a hypersensitivity mediated reaction. CD4+T helper (Th) cells play a key role in hypersensitivity reactions and EC has been linked to an increase in IL-5 expressing Th2 cells [40]. Moreover, CD163+and CD206+M2 macrophages which promote the Th2 response have been shown to be present in the mixed inflammatory EC infiltrate [41, 42]. Cytokines secreted by Th2 cells, such as IL-5, are crucial for eosinophil development, expansion, and survival [43]. Eosinophils, when recruited, are thought to secrete cytokines IL-4 and IL-13, which promote and enhance the Th2 response [44]. Eosinophils in patients with hypereosinophilic syndromes such as EC have been shown

Fig. 2 Eosinophilic cellulitis. A Low power image demonstrating a superficial and deep perivascular and interstitial inflammatory infiltrate with extension into the subcutis (original magnification, ×40). B, Higher power image showing the inflammatory infiltrate that is made up of numerous eosinophils with admixed lymphocytes and histiocytes (original magnification, ×200). C and D Flame figures are appreciated in the dermis with surrounding eosinophils (C, original magnification, ×400) and surrounding palisaded histiocytes (D, original magnification, ×400)





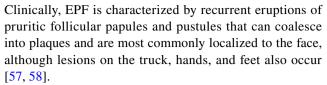
to express the alpha chain of the IL-2 receptor (CD25) and can be "primed" for granule release through binding of IL-2 (produced by activated T-cells) [45].

The differential diagnosis for EC includes other dermatologic conditions that may present with erythematous and indurated skin lesions. Bacterial cellulitis and the early stages of necrotizing fasciitis can have an overlapping clinical appearance with EC; however, the lesions will typically be painful, and the patient will have systemic symptoms such as fever. Necrotizing fasciitis will also show some response to antibiotic therapy or debridement, unlike EC [46]. Histologically, bacterial cellulitis and the early stages of necrotizing fasciitis have prominent dermal neutrophilic infiltrates, as opposed to the characteristic eosinophilic infiltrates, often with flame figures, seen in EC [47]. Another important item to consider in the differential is EGPA. The cutaneous manifestations of EGPA may have a similar appearance to EC; however, EGPA should have additional clinical findings, which can include asthma, paranasal sinusitis, pulmonary infiltrates, mononeuropathy, and/or polyneuropathy [48]. Microscopically, vasculitis should be present in EGPA and absent in EC [34]. EGPA will be discussed further in a later section.

The most common treatment option for EC is oral steroids, although topical steroids may be successful in the treatment of localized lesions [25, 49]. Cyclosporine is an immunosuppressant that suppresses Th cell function, a key contributor to eosinophil recruitment and activation, which has been used in some cases of EC [50, 51]. A variety of other treatment options have been reported in the literature as an alternative to steroids, which include dapsone, tacrolimus (oral or topical), antihistamines (often in combination with other treatment options), interferon alpha, and tumor necrosis factor (TNF)-alpha inhibitors [49]. This treatment list is not exhaustive.

Eosinophilic Pustular Folliculitis

EPF, previously known as Ofuji disease, is a noninfectious, recurrent dermatosis that was first described in 1970 by Ofuji et al. [52]. EPF has a predilection for males and most commonly occurs in the third and fourth decades of life [53]. There are three well-described variants of EPF depending on the demographic involved: immunosuppression-associated EPF, infancy-associated EPF, and classic EPF [54]. The immunosuppression-associated variant of EPF most commonly occurs in individuals with HIV, but can also occur in other immunosuppressive conditions such as malignancy [27]. Infancy-associated EPF presents at a mean age of 6.1 months of life and retains the male predilection seen in the other variants [55]. Classic EPF presents in otherwise healthy adults without evidence of immunosuppression [56].



Microscopically, EPF is characterized by an infiltration of many eosinophils into the hair follicles, frequently involving the infundibulum with intraluminal abscess formation (Fig. 3). Eosinophils may also be seen in the associated sebaceous glands and ducts [54]. Spongiosis is typically present and intraepithelial lymphocytes and eosinophils are often found in the surrounding epidermis [27, 54, 57, 58]. Follicular eosinophilic and neutrophilic abscesses may be present in a subset of cases [54, 58]. Cases of immunosuppression-associated EPF will have a nearly identical histologic appearance, with some reports of abundant perifollicular mast cells and a predominant CD8+T-cell population in the HIV-associated subtype [59, 60]. In infancy-associated EPF, true folliculitis is not always present, and the eosinophilic infiltrate may instead be mostly perifollicular and/or perivascular [61, 62]. A dense interstitial and/or perivascular dermal eosinophilic infiltrate is present in some cases of infancy-associated EPF with associated flame figures present in roughly a quarter of cases [55, 61, 62].

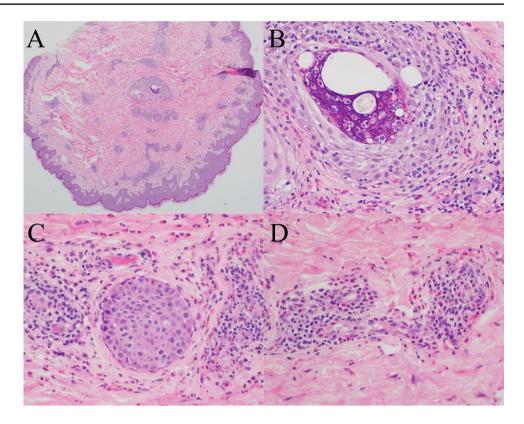
The exact etiopathogenesis of EPF is unknown, although it has been hypothesized that EPF may be a Th2 cytokine dependent condition. This is supported by a case of EPF that demonstrated a positive response to cyclosporin and a subsequent decrease in the mRNA expression levels of Th2 cytokines (IL-4, IL-5, IL-13, IFN-γ, and IL-17) in peripheral blood mononuclear cells (PBMC) following treatment [63]. Similarly, EPF lesions show a positive response to recombinant IFN-y, and IL-5 levels also decrease in PBMCs following therapy [64]. A Th2 dependent response is further supported by demonstration of an abundant CD163 positive M2 macrophage interstitial and perifollicular infiltrate and the presence of IL-10 and STAT6 positive infiltrating cells in EPF [65]. Interestingly, the infiltrating eosinophils in EPF produce prostaglandin D2 (PGD2), which in turn increases eotaxin-3 mRNA expression in human sebocytes [66]. Eotaxin is an eosinophil chemoattractant, and expression in sebocytes helps explain the predominant follicular and perifollicular eosinophilic infiltrate seen in EPF [66, 67].

The differential diagnosis for EPF can be challenging due to its overlapping clinical and histologic features with various other dermatologic conditions. Clinically, the pruritic papules and pustules in classic EPF may be confused with fungal folliculitis or demodex folliculitis, which both show folliculocentric inflammation, but can usually be differentiated microscopically by the identification of the microorganisms on routine hematoxylin and eosin or fungal stains and the absence of a prominent eosinophilic infiltrate [57, 68]. Additionally, acne, rosacea, seborrheic



Head and Neck Pathology (2025) 19:27 Page 5 of 13 27

Fig. 3 Eosinophilic pustular folliculitis. A Unremarkable epidermis with dermal perivascular inflammation and folliculitis with intrafollicular abscess (original magnification, ×40). B Hair follicle with significant eosinophilic infiltration and surrounding perifollicular mixed inflammation with eosinophils (original magnification, ×400). C Perifollicular mixed inflammation with eosinophils (original magnification, ×400). D Perivascular mixed inflammation with eosinophils, which alone is non-specific (original magnification, ×400)



dermatitis, folliculocentric drug hypersensitivity reactions, and folliculotropic T-cell lymphoproliferative disorders can share similar clinical manifestations to EPF, but these entities also lack the prominent follicular eosinophilic infiltrate of EPF [57, 69].

In infants and children, the differential diagnosis expands to include conditions like erythema toxicum neonatorum and transient neonatal pustular melanosis. These conditions are self-limiting neonatal dermatoses with eosinophilic and/or neutrophilic pustules that can mimic the pustules seen in infancy-associated EPF [54, 55]. Infantile acropustulosis can be clinically and microscopically indistinguishable for infantile-EPF, with some authors suggesting that they may be different presentations of the same disease [55, 70]. LCH, which may present with papules and pustules in young children, is also in the differential diagnosis. LCH can usually be differentiated microscopically and by immunohistochemistry due to the neoplastic histiocytic infiltrate [27, 55].

The first line therapy for EPF is typically systemic nonsteroidal anti-inflammatory drugs (NSAIDs), such as indomethacin, which is effective in greater than 70% of cases [56, 71]. Topical NSAIDs in combination with topical tacrolimus are also effective in most cases [71]. For cases that do not respond to NSAIDs, numerous other therapeutic options have been described, including ultraviolet phototherapy, interferon therapy, cyclosporine, corticosteroids (topical or oral), minocycline, isotretinoin, and dapsone [27, 58, 63, 64, 71].

Eosinophilic Granulomatosis with Polyangiitis

EGPA is a rare, anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis (AAV) affecting small-tomedium sized vessels. Previously known as Churg-Strauss syndrome, the clinical presentation is variable and multiple diagnostic criteria currently exist [48, 72–74]. It is characterized by adult-onset asthma, blood and tissue eosinophilia, and small-to-medium vessel necrotizing vasculitis [75]. Clinical criteria include obstructive airway disease, nasal polyps, and mononeuritis multiplex. Laboratory criteria include blood eosinophilia, extravascular eosinophilic predominant inflammation, p-ANCA or antiproteinase-3 antibodies, and hematuria [74]. Furthermore, EGPA can be subclassified based on the ANCA status (either ANCA-positive or ANCA-negative) [76]. This condition can manifest in the skin in approximately 40% of cases [77]. Skin manifestations include purpura, nodules, urticaria, livedo, and ulcers. EGPA most commonly affect the lower limbs; however, the head and neck can also be involved [78, 79].

The pathogenesis of EGPA is driven by both genetic and environmental factors. Genetically, ANCA-positive disease is associated with HLA-DQ, and ANCA-negative disease is associated with mutations in GPA33 and IL-5 [76]. Environmental factors that increase the risk of developing EGPA include exposure to silica, organic solvents, and farming, while smoking decreases the risk [80].

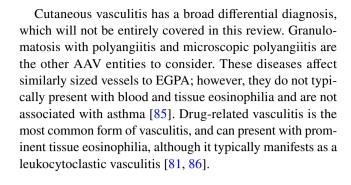


Eosinophils play an important role in the pathogenesis of EGPA, although the exact mechanisms of eosinophilmediated inflammation are not fully understood [81].

Recent advancements in the understanding of the pathogenesis of EGPA have led to new treatment options, including therapies that target B-cells, such as rituximab, and eosinophilic cytokines, such as IL-5 [82]. However, glucocorticoids remain an important component of therapy [76]. Treatment of EGPA was historically suggested to be dependent on ANCA status, but newer evidence did not reveal any significant difference in response to therapies between ANCA-positive and ANCA-negative groups [76, 83].

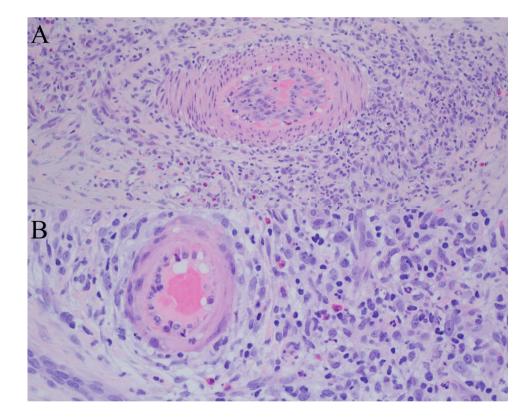
Biopsy is the gold standard for assessment of cutaneous vasculitis. Evaluation is based on the vessel size predominantly involved, the extent of involvement, the inflammatory cell type mainly mediating the damage, and incorporation of other relevant studies such as direct immunofluorescence [79]. Skin lesions in EGPA are characterized by necrotizing vasculitis of small to medium-sized vessels associated with granulomas and increased tissue eosinophils (Fig. 4) [78]. The deep dermal and subcutaneous vessels are typically involved, and direct immunofluorescence is often negative. Eosinophilic or 'red' extravascular granulomas may be present, which are palisaded neutrophilic granulomas with eosinophils [84]. These findings are not specific to EGPA, and correlation with the other clinical and laboratory findings is necessary to make the diagnosis.

Fig. 4 Eosinophilic granulomatosis with polyangiitis. A and B, Prominent perivascular and intravascular inflammation involving the dermal arterioles with inflammatory infiltrate that is composed of eosinophils, neutrophils, lymphocytes and histiocytes. In A, a central fibrin clot is appreciated (original magnification ×200 and ×400, respectively)



Granuloma Faciale

GF is a rare chronic inflammatory skin disease that typically presents as a brown–red plaque on the face, although multiple lesions or extra-facial lesions also occur [87]. Currently, the pathogenesis of GF is not well understood. It has been suggested that GF is a type of chronic leukocytoclastic vasculitis due to the presence of fibrinoid necrosis of small vessel walls identified in a small subset of cases [88]. A study utilizing direct immunofluorescence showed heavy deposition of IgG surrounding vessels in the dermis, which supports that the vessel injury seen in GF is due to the classical pathway activation of complement [89]. This would suggest that the neutrophils and eosinophils present within the lesion are responsible for the vessel injury, and vasculitis may not be part of the pathogenesis of this lesion. GF is





Head and Neck Pathology (2025) 19:27 Page 7 of 13 27

frequently unresponsive to therapy, but topical glucocorticoids and tacrolimus remain first line choices [90].

On histologic examination, GF is characterized by a mixed inflammatory infiltrate composed of many eosinophils, neutrophils, plasma cells, and lymphocytes that is typically separated from the overlying epidermis by a Grenz zone (Fig. 5) [87]. Interestingly, the Grenz zone was historically believed to be a unique feature of GF, but is now known to be a nonspecific finding [91]. As mentioned above, fibrinoid necrosis of vessels and other vascular changes might be seen. Histologic features of GF are on a broad spectrum due to the chronicity of the lesions and can present with both acute and chronic inflammatory patterns with variable amounts of fibrosis [87]. Patterned perivascular fibrosis in particular is often present in chronic lesions and may aid in making the diagnosis.

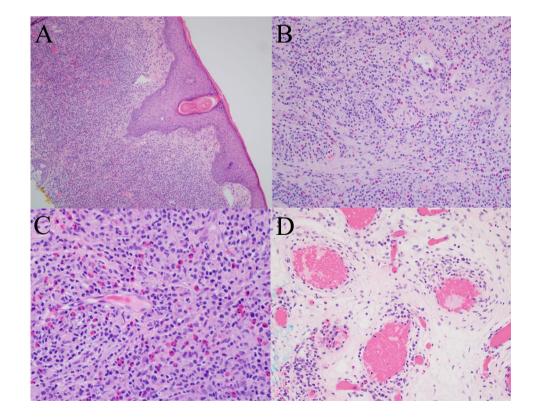
The differential diagnosis for GF includes other lesions with diffuse mixed inflammatory infiltrates. Erythema elevatum diutinum (EED) is also a chronic leukocytoclastic vasculitis that displays a predominately neutrophilic dermal infiltrate with fibrinoid necrosis of small vessel walls [92]. Similar to GF, EED presents as red-brown plaques, but they are more commonly located on extensor and acral surfaces [92, 93]. These lesions can be histologically indistinguishable from GF, but EED typically shows perivascular fibrosis and has greater numbers of neutrophils and fewer eosinophils and plasma cells [94]. Many consider EED and GF to be the same spectrum of disease, with the latter occurring

primarily on the face; although, this assertion is debated in the literature and further studies are needed to clarify the pathogenesis of these conditions. [87, 94].

IgG4-related sclerosing diseases (IgG4-RD) share similar histologic characteristics to GF and EED. Cutaneous lesions in IgG4-RD display small-vessel vasculitis, dermal fibrosis, and a prominent plasma cell infiltrate [95]. An eosinophilic component to the inflammatory infiltrate may also be present. Importantly, the IgG4/IgG ratio in the plasma cell infiltrate needs to be over 40% for the diagnosis of IgG4-RD. It has been suggested that GF and EED might represent localized forms of IgG4-RD due to the amount of histologic overlap [96]. Studies have shown conflicting results following immunohistochemical analysis of the IgG4/IgG ratio of GF cases [95, 96]. Further, cutaneous manifestations of IgG4-RD are rare, and an increased IgG4/IgG ratio alone is not sufficient to make the diagnosis as IgG4-RD as IgG-RD is a clinicopathologic diagnosis that is dependent on a combination of clinical, radiologic, pathologic, and/or laboratory findings [116]. Thus, there is currently not enough evidence to support GF as a cutaneous manifestation of IgG4-RD; however, it is an interesting consideration as further studies are conducted to elucidate the pathogenesis of GF.

Another important differential diagnosis is lymphocytoma cutis or pseudolymphoma, a heterogenous group of T- and B-cell lymphoproliferative processes that can mimic cutaneous lymphoma [97]. Histologically, they are often composed of a dermal infiltrate with many small, mature lymphocytes

Fig. 5 Granuloma Faciale. A The papillary dermis under the epidermis has significantly less inflammatory infiltrate compared to the deeper dermis, known as a Grenz zone (original magnification ×100). B Foci within the lesion that have a more dense, fibrotic stroma (original magnification ×200). C The mixed inflammatory infiltrate is composed of predominantly neutrophils and lymphocytes, with a few eosinophils, plasma cells, and histiocytes in this field. Numerous extravasated red blooe cells and focal leukocytoclastic (karyorrhectic) debris are also present, consistent with an LCV (original magnification ×400). D Chronic perivascular changes including organizing fibrosis are present (original magnifica $tion \times 400$)





27 Page 8 of 13 Head and Neck Pathology (2025) 19:27

admixed with other inflammatory cells. A Grenz zone is frequently present, but lymphocyte exocytosis can occur mimicking mycosis fungoides and other T-cell lymphomas [98]. Compared to GF, cutaneous pseudolymphoma lacks vasculitis and usually typically does not have a prominent neutrophilic or eosinophilic infiltrate. Additionally, germinal centers with tingible body macrophages may be present [98].

Langerhans Cell Histiocytosis

Langerhans cells (LC) are epidermal dendritic cells that play an important role in antigen presentation to T-cells [99, 100]. LCH is a clonal proliferation of cells with LC characteristics and its classification as a neoplasm was cemented after the identification of recurrent BRAF V600E mutations [101]. It is now known that BRAF V600E mutations are the most common and are correlated with high-risk disease and increased resistance to first-line therapy [102, 103]. Other mutations related to activation of the MAPK pathway have been identified, including MAP2K1 (second most common following BRAF V600E), ARAF, BRAF indel, BRAF fusion, and ERBB3 mutations [103]. The etiology remains poorly understood to date and investigations into the cell of origin are ongoing.

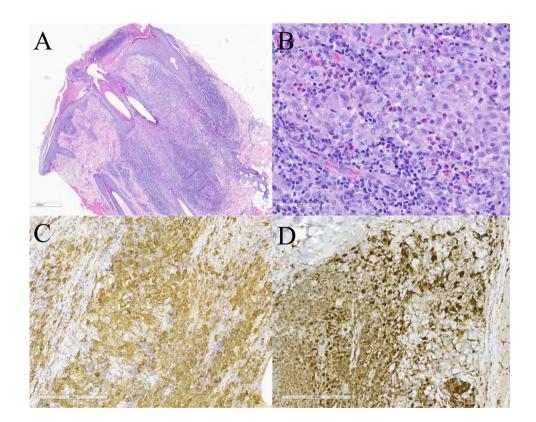
LCH is a disease predominately of childhood, but can present at any age [100, 104, 105]. It often presents with a mild clinical course, sometimes resolving spontaneously,

but in 20% of disseminated cases it can affect multiple organ systems and be fatal. Cutaneous involvement occurs in approximately 40% of cases and is typically a manifestation of multisystem disease [106, 107]. The skin is the second most commonly involved organ system following bone, but isolated skin involvement is rare [106]. When LCH involves the skin, it manifests as a seborrheic dermatitis-like or eczematous eruption most commonly on the scalp or trunk [106].

For unifocal LCH, observation or local therapies like surgery, intralesional steroids, or radiation are recommended unless it affects specific sites (e.g., nervous system, liver, spleen), where systemic treatment should be used [108]. Multifocal LCH treatments vary, with radiation or bisphosphonates typically used for osseous lesions, topical therapy or methotrexate for cutaneous disease, and chemotherapy for extensive multisystem or brain parenchyma involvement. Refractory cases may need targeted therapies like BRAF/MEK inhibitors [108].

LCH is characterized by the accumulation of large, round to oval histiocytes with complex nuclear contours with frequent nuclear grooves (Fig. 6). The inflammatory infiltrate typically includes many eosinophils, lymphocytes, and multinucleated histiocytes, with less frequent neutrophils and plasma cells. In active lesions, LCH cells are the predominant cell type, often accompanied by eosinophils. Fibrosis becomes more prominent as lesions

Fig. 6 Langerhans cell histiocytosis. A The epidermis is excoriated with overlying scale. A vaguely nodular lesion is appreciated underlying the dermis (original magnification ×40). **B** In the dermis, there are numerous medium to large epithelioid cells with abundant pale cytoplasm, irregular to round nuclei, and nuclear grooves. Scattered eosinophils and small lymphocytes are present (original magnification ×400). C and D The lesional cells stain with immunostains for CD1a and \$100, respectively (original magnification ×200)





Head and Neck Pathology (2025) 19:27 Page 9 of 13 2:

progress. By immunohistochemistry, S100, CD1a and CD207 (langerin) positivity is characteristic.

The main consideration in the differential diagnosis of LCH is the accumulation of reactive histocytes occurring in other conditions. Therefore, the differential diagnosis is dependent on the site. In the head and neck, juvenile xanthogranuloma (JXG) is a common non-LCH. It presents as a papule or nodule, often affecting children [109]. Histologically, the lesion is composed of a diffuse infiltrate of histocytes that lack the typical LC characteristics and are negative by CD1a immunohistochemistry. S100 is also notably negative. Eosinophils are usually present scattered throughout the lesion, but are less abundant compared to LCH. Touton giant cells, a type of multinucleated giant cell that have a central eosinophilic core surrounded by a wreath of nuclei, are a notable characteristic of JXG and can be useful in distinguishing it from other cutaneous histiocytoses. The prognosis of JXG is favorable, and it typically regresses spontaneously [110].

Rosai-Dorfman disease (RDD) is another non-LCH that commonly presents as painless bilateral cervical lymphade-nopathy that predominately affects children and young adults [111]. Cutaneous RDD is considered a separate disease entity and is more common in older individuals compared to the classic nodal form of RDD [112, 113]. Cutaneous RDD presents as papulonodular lesions, plaques, or eruptive xanthoma-like lesions without a predilection for a specific site [112].

Histologically, cutaneous lesions are remarkably similar to nodal disease and display nodules or sheets of histocytes with abundant eosinophilic cytoplasm associated with lymphocytes, neutrophils and plasma cells. Histocytes with emperipolesis are characteristic and eosinophils are usually absent. CD1a is negative by immunohistochemistry, which is helpful for distinguishing RDD from LCH. JXG is also in the differential diagnosis for cutaneous RDD, but RDD lacks the characteristic Touton giant cells of JXG, and the histiocytes in RDD are positive for S100 and OCT2 [112].

The pathogenesis of RDD is poorly understood, but recently recurrent mutations involving the activation of the MAPK/ERK pathway have been identified in nodal and extranodal RDD, but not in cutaneous RDD [114, 115]. This suggests that RDD is a clonal process, but further studies are needed. The clinical course of RDD is variable, with most of the cases following a benign course. A subset of cases may show a more aggressive clinical course, with involvement of multiple internal organ systems, and require systemic therapy [111].

Conclusions

In summary, eosinophilic lesions in the head and neck region encompass a wide range of conditions, from benign entities like EPF to neoplasms such as LCH. Tissue eosinophilia represents a valuable diagnostic clue for differentiating these conditions from other conditions of the head and neck skin. Continued research into the mechanisms underlying these conditions and the drivers of eosinophil proliferation and infiltration promises to enhance our ability to further refine diagnostic and therapeutic strategies, potentially benefiting patient outcomes in eosinophilic disorders of the head and neck.

Author Contribution D.D., I.L, Z. W., A.A., and D.C. wrote the manuscript text. D.D prepared the Figs. 1, 2, 3, 4, 5 and 6. All authors reviewed the manuscript.

Funding Open access funding provided by SCELC, Statewide California Electronic Library Consortium. This study was not supported by any funding.

Data Availability No datasets were generated or analysed during the current study.

Code Availability Not applicable.

Declarations

Conflict of interest No conflicting relationship exists for any author.

Ethics Approval This article does not contain any studies with human participants are animals performed by any of the authors.

Consent to Participate For this type of study informed consent is not required.

Consent for Publication For this type of study consent for publication is not required.

Disclaimer The views expressed herein are those of the authors and do not necessarily reflect the official policy or position of Walter Reed National Military Medical Center, Fort Belvoir Community Hospital, the U.S. Army Medical Department, the U.S. Army Office of the Surgeon General, the Department of the Air Force, the Department of the Army, Department of Defense, the Uniformed Services University of the Health Sciences, or any other agency of the U.S. Government. The identification of specific products or scientific instrumentation is considered an integral part of the scientific endeavor and does not constitute endorsement or implied endorsement on the part of the authors, DoD, or any component agency.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.



27 Page 10 of 13 Head and Neck Pathology (2025) 19:27

References

- 1. Adler BL, Krausz AE, Minuti A, Silverberg JI, Lev-Tov H (2016) Epidemiology and treatment of angiolymphoid hyperplasia with eosinophilia (ALHE): a systematic review. J Am Acad Dermatol 74(3):506-512.e11. https://doi.org/10.1016/j.jaad.2015.10.011
- Olsen TG, Helwig EB (1985) Angiolymphoid hyperplasia with eosinophilia. A clinicopathologic study of 116 patients. J Am Acad Dermatol 12(5 Pt 1):781–796. https://doi.org/10.1016/ s0190-9622(85)70098-9
- Guo R, Gavino ACP (2015) Angiolymphoid hyperplasia with eosinophilia. Arch Pathol Lab Med 139(5):683–686. https://doi. org/10.5858/arpa.2013-0334-RS
- Youssef A, Hasan AR, Youssef Y, Al-Soufi L, Elshimali Y, Alshehabi Z (2018) Angiolymphoid hyperplasia with eosinophilia: a case report. J Med Case Rep 12(1):89. https://doi.org/ 10.1186/s13256-018-1599-x
- Machado I, Chong A, Serrano A, Ugalde AMN, Pineda D, Savón L, Olivera E, Llombart-Bosch A (2016) Epithelioid Hemangioma (Angiolymphoid Hyperplasia With Eosinophilia) of the Heart With Peripheral Eosinophilia and Nephrotic Syndrome. Int J Surg Pathol. https://doi.org/10.1177/1066896915604254
- Mukherjee B, Kadaskar J, Priyadarshini O, Krishnakumar S, Biswas J (2015) Angiolymphoid hyperplasia with Eosinophilia of the orbit and adnexa. Ocul Oncol Pathol 2(1):40. https://doi. org/10.1159/000433545
- Zou A, Hu M, Niu B (2021) Comparison between Kimura's disease and angiolymphoid hyperplasia with eosinophilia: case reports and literature review. J Int Med Res 49(9):3000605211040976. https://doi.org/10.1177/03000605211040976
- Fetsch JF, Weiss SW (1991) Observations concerning the pathogenesis of epithelioid hemangioma (angiolymphoid hyperplasia). Mod Pathol 4(4):449–455
- Tokat F, Lehman JS, Sezer E, Cetin ED, Ince U, Durmaz EO (2018) Immunoreactivity of Wilms tumor 1 (WT1) as an additional evidence supporting hemangiomatous rather than inflammatory origin in the etiopathogenesis of angiolymphoid hyperplasia with eosinophilia. Dermatol Pract Concept 8(1):28–32. https://doi.org/10.5826/dpc.0801a06
- Huang S-C et al (2015) Frequent FOS gene rearrangements in epithelioid hemangioma: a molecular study of 58 cases with morphologic reappraisal. Am J Surg Pathol 39(10):1313–1321. https://doi.org/10.1097/PAS.0000000000000469
- Antonescu CR et al (2014) ZFP36-FOSB fusion defines a subset of epithelioid hemangioma with atypical features. Genes Chromosom Cancer. https://doi.org/10.1002/gcc.22206
- Ortins-Pina A, Llamas-Velasco M, Turpin S, Soares-de-Almeida L, Filipe P, Kutzner H (2018) FOSB immunoreactivity in endothelia of epithelioid hemangioma (angiolymphoid hyperplasia with eosinophilia). J Cutan Pathol 45(6):395–402. https:// doi.org/10.1111/cup.13141
- Goto K et al (2022) Categorization of cutaneous epithelioid angiomatous nodule as epithelioid hemangioma or angiolymphoid hyperplasia with eosinophilia: clinicopathologic, immunohistochemical, and molecular analyses of seven lesions. J Cutan Pathol 49(9):765–771. https://doi.org/10.1111/cup.14246
- Hart J, Mandavilli S (2011) Epithelioid angiosarcoma: a brief diagnostic review and differential diagnosis. Arch Pathol Lab Med 135(2):268–272. https://doi.org/10.5858/135.2.268
- 15. Suchak R, Thway K, Zelger B, Fisher C, Calonje E (2011) Primary cutaneous epithelioid angiosarcoma: a clinicopathologic study of 13 cases of a rare neoplasm occurring outside the setting of conventional angiosarcomas and with predilection for the

- limbs. Am J Surg Pathol 35(1):60–69. https://doi.org/10.1097/PAS.0b013e3181fee872
- Abuel-Haija M, Hurford MT (2007) Kimura disease. Arch Pathol Lab Med 131(4):650–651. https://doi.org/10.5858/ 2007-131-650-KD
- Sun Q-F et al (2008) Kimura disease: review of the literature. Intern Med J 38(8):668–672. https://doi.org/10.1111/j.1445-5994.2008.01711.x
- Fite-Trepat L, Martos-Fernandez M, Alberola-Ferranti M, Pablo-Garcia-Cuenca AD, Bescosatin C (2017) Angiolymphoid hyperplasia with eosinophilia involving the occipital artery: case report and review of literature. J Clin Diagn Res 11(3):ZD21–ZD23. https://doi.org/10.7860/JCDR/2017/23323.9569
- Lee C-C et al (2024) Kimura's disease: a clinicopathological study of 23 cases. Frontiers. https://doi.org/10.3389/fmed.2022. 1069102/full
- 21 Bahloul E et al (2017) Angiolymphoid hyperplasia with eosinophilia: report of nine cases. Int J Dermatol. https://doi.org/10. 1111/ijd.13800
- Sagi L, Halachmi S, Levi A, Amitai DB, Enk CD, Lapidoth M (2016) Combined pulsed dye and CO2 lasers in the treatment of angiolymphoid hyperplasia with eosinophilia. Lasers Med Sci 31(6):1093–1096. https://doi.org/10.1007/s10103-016-1954-3
- 23. Wells GC (1971) Recurrent granulomatous dermatitis with eosinophilia. Trans St Johns Hosp Dermatol Soc 57(1):46–56
- Wells GC, Smith NP (1979) Eosinophilic cellulitis. Br J Dermatol 100(1):101–109. https://doi.org/10.1111/j.1365-2133.1979. tb03574.x
- Sinno H et al (2012) Diagnosis and management of eosinophilic cellulitis (Wells' syndrome): a case series and literature review. Can J Plast Surg 20(2):91–97. https://doi.org/10.1177/22925 5031202000204
- Caputo R, Marzano AV, Vezzoli P, Lunardon L (2006) Wells syndrome in adults and children: a report of 19 cases. Arch Dermatol 142(9):1157–1161. https://doi.org/10.1001/archderm.142.9.1157
- Long H, Zhang G, Wang L, Lu Q (2016) Eosinophilic skin diseases: a comprehensive review. Clin Rev Allergy Immunol 50(2):189–213. https://doi.org/10.1007/s12016-015-8485-8
- 28. Moossavi M, Mehregan DR (2003) Wells' syndrome: a clinical and histopathologic review of seven cases. Int J Dermatol 42(1):62–67. https://doi.org/10.1046/j.1365-4362.2003.01705.x
- Qiao J, Sun C-E, Zhu W, Zhu D, Fang H (2013) Flame figures associated with eosinophilic dermatosis of hematologic malignancy: is it possible to distinguish the condition from eosinophilic cellulitis in patients with hematoproliferative disease? Int J Clin Exp Pathol 6(8):1683–1687
- Melski JW (1990) Wells' syndrome, insect bites, and eosinophils.
 Dermatol Clin 8(2):287–293
- Reichel M, Isseroff RR, Vogt PJ, Gandour-Edwards R (1991) Wells' syndrome in children: varicella infection as a precipitating event. Br J Dermatol 124(2):187–190. https://doi.org/10.1111/j. 1365-2133.1991.tb00431.x
- 32 Yu AM et al (2018) Pediatric Wells syndrome (eosinophilic cellulitis) after vaccination: a case report and review of the literature. Pediat Dermatol. https://doi.org/10.1111/pde.13532
- Boura P, Sarantopoulos A, Lefaki I, Skendros P, Papadopoulos P (2006) Eosinophilic cellulitis (Wells' syndrome) as a cutaneous reaction to the administration of adalimumab. Ann Rheum Dis 65(6):839–840. https://doi.org/10.1136/ard.2005.044685



Head and Neck Pathology (2025) 19:27 Page 11 of 13 2:

Hurni MA, Gerbig AW, Braathen LR, Hunziker T (1997) Toxocariasis and Wells' syndrome: a causal relationship? Dermatology 195(4):325–328. https://doi.org/10.1159/000245981

- 35. Spinelli M, Frigerio E, Cozzi A, Garutti C, Garavaglia MC, Altomare G (2008) Bullous Wells' syndrome associated with non-Hodgkin's lymphocytic lymphoma. Acta Derm Venereol 88(5):530–531. https://doi.org/10.2340/00015555-0492
- 36 Hirsch K et al (2005) Eosinophilic cellulitis (Wells' syndrome) associated with colon carcinoma. JDDG: Journal der Deutschen Dermatologischen Gesellschaft 3(7):530–531. https://doi.org/10.1111/j.1610-0387.2005.05726.x
- Lim CE, Ng SK, Thng ST (2017) Bullous pesentation of idiopathic wells syndrome (eosinophilic cellulitis). Ann Acad Med Singap 46(8):324–326
- Katoulis AC, Bozi E, Samara M, Kalogeromitros D, Panayiotides I, Stavrianeas NG (2009) Idiopathic bullous eosinophilic cellulitis (Wells' syndrome). Clin Exp Dermatol 34(7):e375-376. https://doi.org/10.1111/j.1365-2230.2009.03328.x
- Fujii K, Tanabe H, Kanno Y, Konishi K, Ohgou N (2003) Eosinophilic cellulitis as a cutaneous manifestation of idiopathic hypereosinophilic syndrome. J Am Acad Dermatol 49(6):1174–1177. https://doi.org/10.1016/s0190-9622(03)00466-3
- Yagi H, Tokura Y, Matsushita K, Hanaoka K, Furukawa F, Takigawa M (1997) Wells' syndrome: a pathogenic role for circulating CD4+CD7- T cells expressing interleukin-5 mRNA. Br J Dermatol 136(6):918–923
- 41. (2024) A possible mechanism in the recruitment of eosinophils and Th2 cells through CD163(+) M2 macrophages in the lesional skin of eosinophilic cellulitis. https://pubmed.ncbi.nlm.nih.gov/24721436/
- Barros MHM, Hauck F, Dreyer JH, Kempkes B, Niedobitek G (2013) Macrophage polarisation: an immunohistochemical approach for identifying M1 and M2 macrophages. PLoS ONE 8(11):e80908. https://doi.org/10.1371/journal.pone.0080908
- Kouro T, Takatsu K (2024) IL-5- and eosinophil-mediated inflammation: from discovery to therapy. Int Immunol. https:// academic.oup.com/intimm/article/21/12/1303/686327
- Spencer LA, Weller PF (2010) Eosinophils and Th2 immunity: contemporary insights. Immunol Cell Biol 88(3):250–256. https://doi.org/10.1038/icb.2009.115
- Simon H-U et al (2003) Interleukin-2 primes eosinophil degranulation in hypereosinophilia and Wells' syndrome. Eur J Immunol. https://doi.org/10.1002/eji.200323727
- Gandhi RK, Coloe J, Peters S, Zirwas M, Darabi K (2011) Wells syndrome (eosinophilic cellulitis): a clinical imitator of bacterial cellulitis. J Clin Aesthet Dermatol 4(7):55–57
- Cormerais M, Poizeau F, Darrieux L, Tisseau L, Safa G (2015)
 Wells' syndrome mimicking facial cellulitis: a report of two cases. Case Rep Dermatol 7(2):117–122. https://doi.org/10.1159/ 000432392
- 48. Masi AT et al (1990) The American College of Rheumatology 1990 criteria for the classification of churg-strauss syndrome (allergic granulomatosis and angiitis). Arthritis Rheum 33(8):1094–1100. https://doi.org/10.1002/art.1780330806
- Räßler F, Lukács J, Elsner P (2016) Treatment of eosinophilic cellulitis (Wells syndrome)—a systematic review. J Eur Acad Dermatol Venereol 30(9):1465–1479. https://doi.org/10.1111/jdv.13706
- Herr H, Koh JK (2001) Eosinophilic cellulitis (Wells' syndrome) successfully treated with low-dose cyclosporine. J Korean Med Sci 16(5):664–668. https://doi.org/10.3346/jkms.2001.16.5.664
- Kim SH, Kwon JE, Kim H-B (2013) Successful treatment of steroid-dependent eosinophilic cellulitis with cyclosporine. Allergy Asthma Immunol Res 5(1):62–64. https://doi.org/10.4168/aair. 2013.5.1.62

- Ofuji S, Ogino A, Horio T, Oseko T, Uehara M (1970) Eosinophilic pustular folliculitis. Acta Derm Venereol 50(3):195–203
- Ellis E, Scheinfeld N (2004) Eosinophilic pustular folliculitis: a comprehensive review of treatment options. Am J Clin Dermatol 5(3):189–197. https://doi.org/10.2165/00128071-20040 5030-00007
- Nervi SJ, Schwartz RA, Dmochowski M (2006) Eosinophilic pustular folliculitis: a 40 year retrospect. J Am Acad Dermatol 55(2):285–289. https://doi.org/10.1016/j.jaad.2006.02.034
- Hernández-Martín Á, Nuño-González A, Colmenero I, Torrelo A (2013) Eosinophilic pustular folliculitis of infancy: a series of 15 cases and review of the literature. J Am Acad Dermatol 68(1):150–155. https://doi.org/10.1016/j.jaad.2012.05.025
- Katoh M, Nomura T, Miyachi Y, Kabashima K (2013) Eosinophilic pustular folliculitis: a review of the Japanese published works. J Dermatol 40(1):15–20. https://doi.org/10.1111/1346-8138.12008
- "Clinical and histopathological differential diagnosis of eosinophilic pustular folliculitis - Fujiyama - 2013 - The Journal of Dermatology - Wiley Online Library.", 2024. https://doi.org/ 10.1111/1346-8138.12125
- (2024) Clinical and pathological analysis of 10 cases of eosinophilic pustular folliculitis. https://pubmed.ncbi.nlm.nih.gov/ 37719931/
- Holmes RB, Martins C, Horn T (2002) The histopathology of folliculitis in HIV-infected patients. J Cutan Pathol 29(2):93–95. https://doi.org/10.1034/j.1600-0560.2002.290205.x
- Afonso JPJM, Tomimori J, Michalany NS, Nonogaki S, Porro AM (2012) Pruritic papular eruption and eosinophilic folliculitis associated with human immunodeficiency virus (HIV) infection: a histopathological and immunohistochemical comparative study. J Am Acad Dermatol 67(2):269–275. https://doi.org/10.1016/j. jaad.2011.11.923
- Takei S et al (2024) Two cases of infancy associated eosinophilic pustular folliculitis (I-EPF) comparing the profile of infiltrating cells with classic EPF by immunohistochemical study. J Dermatol. https://doi.org/10.1111/1346-8138.16985
- Fertitta L, Bodemer C, Molina T, Frassati-Biaggi A, Fraitag S, Leclerc-Mercier S (2022) Eosinophilic pustular folliculitis of infancy: a histologic assessment of 43 cases. Am J Dermatopathol 44(6):395–403. https://doi.org/10.1097/DAD.0000000000 002006
- Otsuka A, Doi H, Miyachi Y, Kabashima K (2010) Treatment of eosinophilic pustular folliculitis with ciclosporin: suppression of mRNA expression of IL-4 and IL-13. J Eur Acad Dermatol Venereol 24(12):1489–1491. https://doi.org/10.1111/j.1468-3083.2010.03674.x
- 64. Fushimi M et al (1996) Eosinophilic pustular folliculitis effectively treated with recombinant interferon-gamma: suppression of mRNA expression of interleukin 5 in peripheral blood mononuclear cells. Br J Dermatol 134(4):766–772
- 65. Hagiwara A, Fujimura T, Furudate S, Kambayashi Y, Kagatani S, Aiba S (2014) Induction of CD163(+) M2 macrophages in the lesional skin of eosinophilic pustular folliculitis. Acta Derm Venereol 94(1):104–106. https://doi.org/10.2340/00015555-1612
- Nakahigashi K et al (2012) PGD2 induces eotaxin-3 via PPARγ from sebocytes: a possible pathogenesis of eosinophilic pustular folliculitis. J Allergy Clin Immunol 129(2):536–543. https://doi. org/10.1016/j.jaci.2011.11.034
- 67. Rankin SM, Conroy DM, Williams TJ (2000) Eotaxin and eosinophil recruitment: implications for human disease. Mol Med Today 6(1):20–27. https://doi.org/10.1016/S1357-4310(99) 01635-4
- 68 Haupt HM, Stern JB, Weber CB (1990) Eosinophilic pustular folliculitis: Fungal folliculitis? J Am Acad Dermatol 23(5):1012– 1014. https://doi.org/10.1016/0190-9622(90)70325-C



27 Page 12 of 13 Head and Neck Pathology (2025) 19:27

- 69 Two AM, Wu W, Gallo RL, Hata TR (2015) Rosacea: part I. Introduction, categorization, histology, pathogenesis, and risk factors. J Am Acad Dermatol 72(5):749–758. https://doi.org/10. 1016/j.jaad.2014.08.028
- 70 Vicente J et al (1996) Are eosinophilic pustular folliculitis of infancy and infantile acropustulosis the same entity? Brit J Dermatol. https://doi.org/10.1046/j.1365-2133.1996.d01-1085.x
- Nomura T, Katoh M, Yamamoto Y, Miyachi Y, Kabashima K (2016) Eosinophilic pustular folliculitis: a published work-based comprehensive analysis of therapeutic responsiveness. J Dermatol 43(8):919–927. https://doi.org/10.1111/1346-8138.13287
- Jennette JC et al (1994) Nomenclature of systemic vasculitides. Arthritis Rheum 37(2):187–192. https://doi.org/10.1002/art. 1780370206
- Lanham JG, Elkon KB, Pusey CD, Hughes GR (1984) Systemic vasculitis with asthma and eosinophilia: a clinical approach to the churg-strauss syndrome. Medicine 63(2):65
- 74. Grayson PC et al (2022) 2022 American college of rheumatology/european alliance of associations for rheumatology classification criteria for eosinophilic granulomatosis with polyangiitis. Ann Rheum Dis 81(3):309–314. https://doi.org/10.1136/annrheumdis-2021-221794
- White J, Dubey S (2023) Eosinophilic granulomatosis with polyangiitis: a review. Autoimmun Rev 22(1):103219. https:// doi.org/10.1016/j.autrev.2022.103219
- Emmi G et al (2023) Evidence-based guideline for the diagnosis and management of eosinophilic granulomatosis with polyangiitis. Nat Rev Rheumatol 19(6):378–393. https://doi.org/10.1038/s41584-023-00958-w
- Comarmond C et al (2013) Eosinophilic granulomatosis with polyangiitis (Churg–Strauss): clinical characteristics and longterm followup of the 383 patients enrolled in the French Vasculitis Study Group cohort. Arthritis Rheum 65(1):270–281. https://doi.org/10.1002/art.37721
- Vaglio A, Buzio C, Zwerina J (2013) Eosinophilic granulomatosis with polyangiitis (Churg-Strauss): state of the art. Allergy 68(3):261–273. https://doi.org/10.1111/all.12088
- Carlson JA, Cavaliere LF, Grant-Kels JM (2006) Cutaneous vasculitis: diagnosis and management. Clin Dermatol 24(5):414–429. https://doi.org/10.1016/j.clindermatol.2006.07.007
- Maritati F et al (2021) Occupational exposures and smoking in eosinophilic granulomatosis with polyangiitis: a case-control study. Arthritis Rheumatol 73(9):1694–1702. https://doi.org/ 10.1002/art.41722
- Khoury P, Grayson PC, Klion AD (2014) Eosinophils in vasculitis: characteristics and roles in pathogenesis. Nat Rev Rheumatol 10(8):474–483. https://doi.org/10.1038/nrrheum.2014.98
- 82 Trivioli G, Terrier B, Vaglio A (2020) Eosinophilic granulomatosis with polyangiitis: understanding the disease and its management. Rheumatology (Oxford) 59(Suppl 3):iii84–iii94. https://doi.org/10.1093/rheumatology/kez570
- Moiseev S et al (2020) International consensus on antineutrophil cytoplasm antibodies testing in eosinophilic granulomatosis with polyangiitis. Am J Respir Crit Care Med 202(10):1360–1372. https://doi.org/10.1164/rccm.202005-1628SO
- Carlson JA (2010) The histological assessment of cutaneous vasculitis. Histopathology 56(1):3–23. https://doi.org/10.1111/j. 1365-2559.2009.03443.x
- Tabb ES, Duncan LM, Nazarian RM (2021) Eosinophilic granulomatosis with polyangiitis: cutaneous clinical and histopathologic differential diagnosis. J Cutan Pathol 48(11):1379–1386. https://doi.org/10.1111/cup.14065
- Radić M, Martinović Kaliterna D, Radić J (2012) Drug-induced vasculitis: a clinical and pathological review. Neth J Med 70(1):12–17

- Ortonne N, Wechsler J, Bagot M, Grosshans E, Cribier B (2005) Granuloma faciale: a clinicopathologic study of 66 patients. J Am Acad Dermatol 53(6):1002–1009. https://doi.org/10.1016/j.jaad. 2005.08.021
- Marcoval J, Moreno A, Peyr J (2004) Granuloma faciale: a clinicopathological study of 11 cases. J Am Acad Dermatol 51(2):269–273. https://doi.org/10.1016/j.jaad.2003.11.071
- Barnadas MA, Curell R, Alomar A (2006) Direct immunofluorescence in granuloma faciale: a case report and review of literature.
 J Cutan Pathol 33(7):508–511. https://doi.org/10.1111/j.1600-0560.2006.00451.x
- Lindhaus C, Elsner P (2018) Granuloma faciale treatment: a systematic review. Acta Derm Venereol 98(1):14–18. https://doi.org/10.2340/00015555-2784
- Abbas O, Mahalingam M (2013) The grenz zone. Am J Dermatopathol 35(1):83. https://doi.org/10.1097/DAD.0b013e3182 4feb4e
- 92 Yiannias JA, El-Azhary RA, Gibson LE (1992) Erythema elevatum diutinum: a clinical and histopathologic study of 13 patients. J Am Acad Dermatol 26(1):38–44. https://doi.org/10.1016/0190-9622(92)70003-x
- 93. Sardiña LA, Jour G, Piliang MP, Bergfeld WF (2019) Erythema elevatum diutinum a rare and poorly understood cutaneous vasculitis: a single institution experience. J Cutan Pathol 46(2):97–101. https://doi.org/10.1111/cup.13378
- 94. Ziemer M, Koehler MJ, Weyers W (2011) Erythema elevatum diutinum—a chronic leukocytoclastic vasculitis microscopically indistinguishable from granuloma faciale? J Cutan Pathol 38(11):876–883. https://doi.org/10.1111/j.1600-0560.2011.
- Kavand S, Lehman JS, Gibson LE (2016) Granuloma faciale and erythema elevatum diutinum in relation to immunoglobulin G4-related disease: an appraisal of 32 cases. Am J Clin Pathol 145(3):401–406. https://doi.org/10.1093/ajcp/aqw004
- Cesinaro AM, Lonardi S, Facchetti F (2013) Granuloma faciale: a cutaneous lesion sharing features with IgG4-associated sclerosing diseases. Am J Surg Pathol 37(1):66–73. https://doi.org/10.1097/PAS.0b013e318271efd0
- 97 Ploysangam T, Breneman DL, Mutasim DF (1998) Cutaneous pseudolymphomas. J Am Acad Dermatol 38(6 Pt 1):877–895. https://doi.org/10.1016/s0190-9622(98)70154-9
- 98. Villalobos-Ayala RA et al (2022) Lymphocytoma cutis (cutaneous B-cell pseudolymphoma): study of 102 cases with emphasis on the histological characteristics and immunohistochemistry of the miliarial type. Int J Dermatol 61(3):316–323. https://doi.org/10.1111/jjd.15909
- Bechan GI, Egeler RM, Arceci RJ (2006) Biology of langerhans cells and langerhans cell histiocytosis. Int Rev Cytol 254:1–43. https://doi.org/10.1016/S0074-7696(06)54001-X
- Filipovich A, McClain K, Grom A (2010) Histiocytic disorders: recent insights into pathophysiology and practical guidelines. Biol Blood Marrow Transplant 16(1 Suppl):S82-89. https://doi. org/10.1016/j.bbmt.2009.11.014
- 101. Badalian-Very G et al (2010) Recurrent BRAF mutations in Langerhans cell histiocytosis. Blood 116(11):1919–1923. https://doi.org/10.1182/blood-2010-04-279083
- Héritier S et al (2016) BRAF mutation correlates with high-risk langerhans cell histiocytosis and increased resistance to first-line therapy. J Clin Oncol 34(25):3023–3030. https://doi.org/10.1200/ JCO.2015.65.9508
- Allen CE, Merad M, McClain KL (2018) Langerhans-cell histiocytosis. N Engl J Med 379(9):856–868. https://doi.org/10.1056/ NEJMra1607548
- Salotti JA, Nanduri V, Pearce MS, Parker L, Lynn R, Windebank KP (2009) Incidence and clinical features of Langerhans cell



Head and Neck Pathology (2025) 19:27 Page 13 of 13 27

histiocytosis in the UK and Ireland. Arch Dis Child 94(5):376–380. https://doi.org/10.1136/adc.2008.144527

- Stålemark H, Laurencikas E, Karis J, Gavhed D, Fadeel B, Henter J-I (2008) Incidence of Langerhans cell histiocytosis in children: a population-based study. Pediatr Blood Cancer 51(1):76–81. https://doi.org/10.1002/pbc.21504
- Krooks J, Minkov M, Weatherall AG (2018) Langerhans cell histiocytosis in children: history, classification, pathobiology, clinical manifestations, and prognosis. J Am Acad Dermatol 78(6):1035–1044. https://doi.org/10.1016/j.jaad.2017.05.059
- Allen CE, Ladisch S, McClain KL (2015) How I treat Langerhans cell histiocytosis. Blood 126(1):26–35. https://doi.org/10.1182/ blood-2014-12-569301
- Goyal G et al (2022) International expert consensus recommendations for the diagnosis and treatment of Langerhans cell histiccytosis in adults. Blood 139(17):2601–2621. https://doi.org/10.1182/blood.2021014343
- Szczerkowska-Dobosz A, Kozicka D, Purzycka-Bohdan D, Biernat W, Stawczyk M, Nowicki R (2014) Juvenile xanthogranuloma: a rare benign histiocytic disorder. Adv Dermatol Allergol Postępy Dermatologii i Alergologii 31(3):197. https://doi.org/10.5114/pdia.2014.40918
- Cohen BA, Hood A (1989) Xanthogranuloma: report on clinical and histologic findings in 64 patients. Pediatr Dermatol 6(4):262–266. https://doi.org/10.1111/j.1525-1470.1989.tb009 09.x

- Abla O et al (2018) Consensus recommendations for the diagnosis and clinical management of Rosai–Dorfman–Destombes disease. Blood 131(26):2877–2890. https://doi.org/10.1182/blood-2018-03-839753
- Bruce-Brand C, Schneider JW, Schubert P (2020) Rosai–Dorfman disease: an overview. J Clin Pathol 73(11):697–705. https://doi.org/10.1136/jclinpath-2020-206733
- Brenn T et al (2002) Cutaneous rosai-dorfman disease is a distinct clinical entity. Am J Dermatopathol 24(5):385–391. https://doi.org/10.1097/00000372-200210000-00001
- Goyal G et al (2020) Clinicopathological features, treatment approaches, and outcomes in Rosai–Dorfman disease. Haematologica 105(2):348–357. https://doi.org/10.3324/haematol.2019. 219626
- 115. Garces S et al (2017) Mutually exclusive recurrent KRAS and MAP2K1 mutations in Rosai–Dorfman disease. Mod Pathol 30(10):1367–1377. https://doi.org/10.1038/modpathol.2017.55

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

