

Treatment patterns and outcomes of 2310 patients with secondary acute myeloid leukemia: a PETHEMA registry study

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Key Points

- The large PETHEMA registry shows that secondary AML represents 27% of AML cases and confirms its independent adverse prognostic value.
- Subcategories of secondary AML were analyzed, including MSD/MPN and therapy-related cases, with different features and outcomes.

Secondary acute myeloid leukemia (sAML) comprises a heterogeneous group of patients and is associated with poor overall survival (OS). We analyze the characteristics, treatment patterns, and outcomes of adult patients with sAML in the Programa Español de Tratamientos en Hematología (PETHEMA) registry. Overall, 6211 (72.9%) were de novo and 2310 (27.1%) had sAML, divided into myelodysplastic syndrome AML (MDS-AML, 44%), MDS/myeloproliferative AML (MDS/MPN-AML, 10%), MPN-AML (11%), therapy-related AML (*t*-AML, 25%), and antecedent neoplasia without prior chemotherapy/radiotherapy (neo-AML, 9%). Compared with de novo, patients with sAML were older (median age, 69 years), had more Eastern Cooperative Oncology Group ≥ 2 (35%) or high-risk cytogenetics (40%), less FMS-like tyrosine kinase 3 internal tandem duplication (11%), and nucleophosmin 1 (*NPM1*) mutations (21%) and received less intensive chemotherapy regimens (38%) (all $P < .001$). Median OS was higher for de novo than sAML (10.9 vs 5.6 months; $P < .001$) and shorter in sAML after hematologic disorder (MDS, MDS/MPN, or MPN) compared with *t*-AML and neo-AML (5.3 vs 6.1 vs 5.7 months, respectively; $P = .04$). After intensive chemotherapy, median OS was better among patients with de novo and neo-AML (17.2 and 14.6 months, respectively). No OS differences were observed after hypomethylating agents according to type of AML. sAML was an independent adverse prognostic factor for OS. We confirmed high prevalence and adverse features of sAML and established its independent adverse prognostic value. This trial was registered at www.clinicaltrials.gov as #NCT02607059.

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Requests for data sharing should be sent to Pau Montesinos (montesinos_pau@gva.es).

The full-text version of this article contains a data supplement.

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Introduction

Secondary acute myeloid leukemia (sAML) includes therapy-related AML (*t*-AML) and antecedent hematologic disorders.^{1,2} The reported incidence of sAML ranges from 18% to 28% of all AML cases and is associated with poor outcomes, including lower complete remission (CR), overall survival (OS), and relapse-free survival rates than de novo AML.^{1,3-6} Consequently, the diagnosis of sAML has been considered a risk factor for early death and OS in some predictive models.^{7,8} Nevertheless, its independent prognostic value has been questioned due to its association with older age, frequent comorbidities/organ dysfunction, and unfavorable cytogenetic and molecular profile.⁹ On the other hand, whether patients with AML with antecedent neoplasia who did not receive chemotherapy or radiotherapy for the first malignancy (neo-AML) should be considered as having sAML remains an unresolved issue. To our knowledge, it is unknown if these patients have differential characteristics and outcomes compared with other sAMLs.

This study aims to analyze the incidence, clinical and biological characteristics, and outcome in a large series of patients with sAML reported to the Programa Español de Tratamientos en Hematología (PETHEMA) registry. These characteristics and outcomes will be compared with those of patients with de novo AML as well as between the different types of sAML and according to therapeutic approaches.

Methods

PETHEMA registry

The PETHEMA AML registry (NCT02607059) includes patients diagnosed with AML, regardless of the treatment administered. Patient and disease characteristics, in addition to treatment approaches and outcomes, were collected retrospectively, including demographics (age and gender), cytomorphological subtype of AML, cytogenetics, details on treatment schedule, response assessment, and disease follow-up (relapse or death). Optional forms were also collected, including baseline physical and laboratory examination; prior neoplastic/hematologic diseases or exposure to radiation, chemotherapy, or immunosuppressive drugs; molecular assessment (according to the site's routine practice); treatment-related toxicities; and consolidation and post-consolidation schedules (eg, transplant and maintenance). Patient and disease baseline characteristics were captured at the time of diagnosis (before starting treatment).

Eligibility

Adult patients (age ≥ 18 years) reported to the multinational PETHEMA AML registry and diagnosed in Spanish and Portuguese institutions from 1 January 1990 until 31 December 2019 were included in the study. Patients with acute promyelocytic leukemia were excluded, just like those without information related to antecedents of prior malignancies, preleukemic hematological disorders, and leukemogenic or immunosuppressive therapies. The study was approved by Hospital Universitari i Politècnic La Fe's Research Ethics Board according to the Declaration of Helsinki, and informed consent was a requisite for patients alive at the time of the analyses (15 September 2020).

Treatment schedules

Patients were classified into five groups according to intensity of therapy: intensive chemotherapy (IC), nonintensive chemotherapy (non-IC), hypomethylating agents (HMA), clinical trial, and best supportive care (BSC). IC schedules included the following regimens: 3 + 7 (idarubicin or daunorubicin and Ara-C), mitoxantrone plus Ara-C, FLAG-Ida (fludarabine, idarubicin, and Ara-C), FLAT (fludarabine, Ara-C, and topotecan), or ICE (idarubicin, Ara-C, and etoposide). HMA comprised decitabine or azacitidine at low doses. The non-IC group included FLUGA (fludarabine and Ara-C), FLAGIDA-Lite (fludarabine, Ara-C, and idarubicin), or LDAC. Regardless of the regimen intensity, all clinical trials were grouped in a separate treatment category. BSC included patients receiving transfusions and other supportive measures, including oral agents to control the white blood cell (WBC) counts (ie, hydroxyurea, melphalan, mercaptopurine, or thioguanine). Treatment schedules are detailed in supplemental Table 1.

Study definitions and endpoints

sAML comprised patients derived from myelodysplastic syndrome (MDS-AML), myeloproliferative neoplasm (MPN-AML), myelodysplastic/myeloproliferative neoplasm (MDS/MPN-AML), and *t*-AML. For the purposes of this study, we also included the aforementioned patients with neo-AML. The remaining patients were classified as de novo AML. Latency period was the time between diagnosis of the primary disease or previous cytotoxic therapy and sAML. Cytogenetics was classified according to the Medical Research Council (MRC) classification.¹⁰

According to the revised criteria by Cheson et al,¹¹ CR required $<5\%$ of blast cells in bone marrow (BM) and absence of them in peripheral blood (PB), with neutrophil and platelet counts $>1 \times 10^9/L$ and $>100 \times 10^9/L$, respectively. If patients did not achieve these values of neutrophil and platelet counts in PB, response was classified as CR with incomplete recovery (CRi). Both CR and CRi had to be lacking in extramedullary disease. BM blasts between 5% and 25% and reduction $>50\%$ compared with the basal value was considered partial remission (PR). Independent of the therapeutic approach, response in patients who died before having been assessed was classified as induction death. Patients not meeting the previous criteria were considered resistant. After achieving CR/CRi, reappearance of disease in PB, blast cells in BM $\geq 5\%$, and development of extramedullary disease were criteria for relapse.

The primary end point was to compare baseline characteristics, CR/CRi, OS, and event-free survival (EFS) in all patients with reported sAML and according to therapeutic approaches. Other secondary end points were to analyze the differences in baseline characteristics and OS between patients with AML de novo and sAML and between different sAML subsets.

Statistical analysis

To analyze the relationship between the characteristics of the patients' subgroups, χ -square with Yates' correction, analysis of variance, Wilcoxon test, Student *t* test, and Kruskal-Wallis test were performed depending on the variable.

The Kaplan-Meier estimator¹² was used to calculate unadjusted time-to-event variables and log-rank test¹³ to compare them. The corrected method of Kalbfleisch and Prentice was used for the

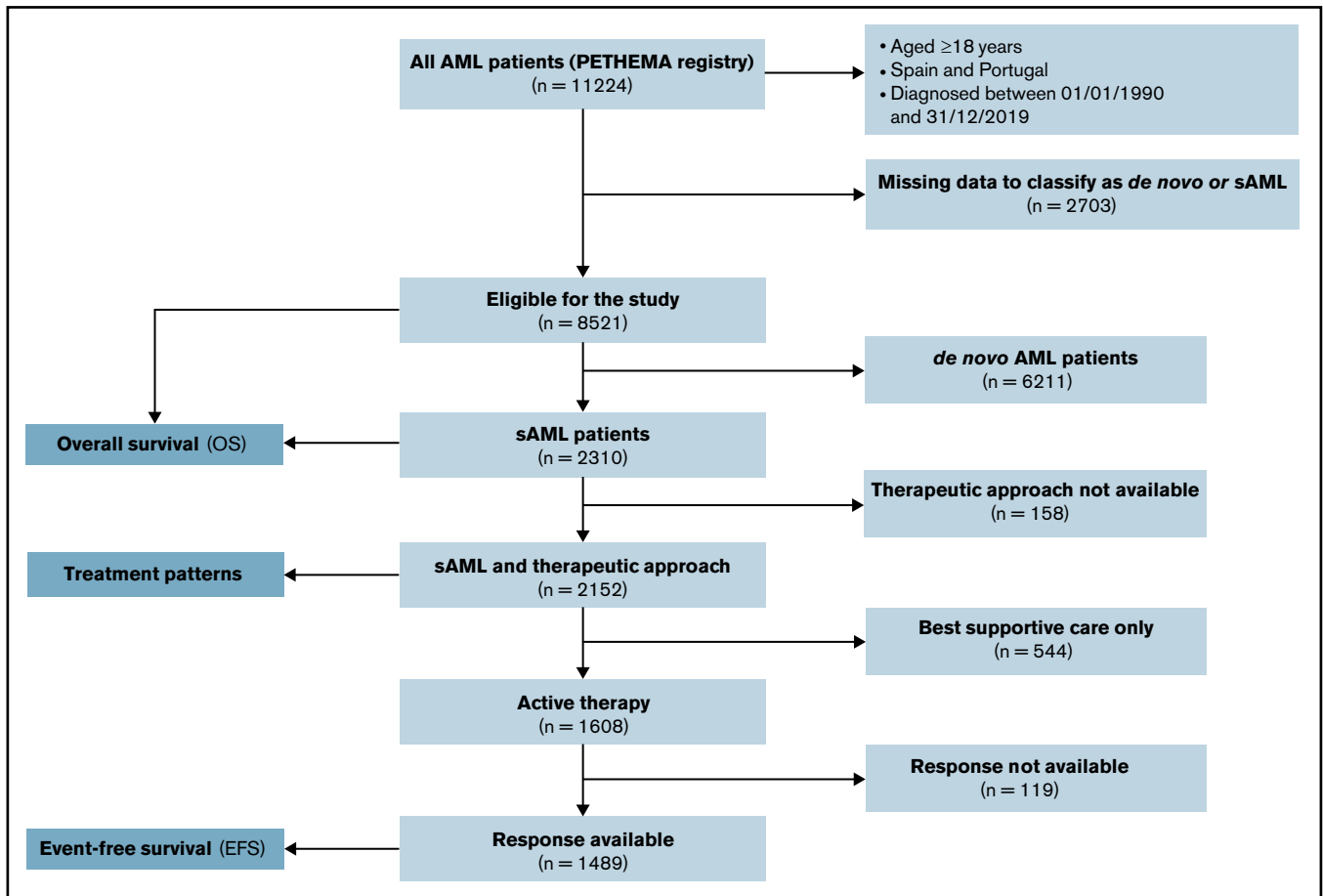


Figure 1. Consolidated Standards of Reporting Trials (CONSORT) diagram for secondary AML adult patients.

estimation of medians and 95% confidence intervals (CI).¹⁴ OS was analyzed from the date of diagnosis of sAML until death and EFS from the date of diagnosis to death due to any cause, treatment failure, or relapse, whichever occurred first. Multivariate analysis for OS was performed through a Cox proportional hazards model and through multinomial logistic regression for CR/CRi. Characteristics with significant association in univariate analysis ($P < .1$) and those available variables with a possible relationship in previous studies were included in the multivariate analysis. Living patients at the time of the analysis were censored at the last known living day. Patient follow-up was updated on 31 December 2019. All P values reported are 2-sided. Computations were executed using the R 2.14.0 software package.

Results

Characteristics and treatment (sAML vs de novo AML)

Between January 1990 and December 2019, 11 224 patients with AML were registered from several institutions in Spain and Portugal. Of these patients, 8521 (75.9%) could be classified as de novo AML ($n = 6211$, 72.9%) or sAML ($n = 2310$, 27.1%). A Consolidated Standards of Reporting Trials (CONSORT) diagram is shown in Figure 1. Characteristics and treatment patterns of de novo vs sAML patients are shown in Table 1. Compared with patients with

de novo AML, patients with sAML were older (median age in years, 69 vs 64; $P < .001$), were more frequently male (60% vs 53%; $P < .001$), had performance status Eastern Cooperative Oncology Group (ECOG) ≥ 2 (35% vs 28%; $P < .001$), had lower median WBC count in PB and blast cells percentage in BM ($6.6 \times 10^9/L$ and 42% vs $10.4 \times 10^9/L$ and 64%, respectively, both $P < .001$), were more often classified with high-risk cytogenetics according to MRC (40% vs 24%; $P < .001$), and had less FMS-like tyrosine kinase 3 (*FLT3*) internal tandem duplication (ITD) and nucleophosmin 1 (*NPM1*) mutations (11% and 16% vs 21% and 33%, respectively; $P < .001$). Patients with sAML were less commonly treated up front with IC ($P < .001$).

Characteristics of sAML according to treatment approach

Table 2 shows patient and disease features according to treatment approach in sAML. Among 2152 patients with sAML with available front-line treatment, 876 (38%) received IC, 328 (14%) received HMA, 325 (14%) received non-IC, 79 (3%) were in a clinical trial, and 544 (24%) received BSC. Patients who received BSC were older and had worse ECOG, higher uric acid, lactate dehydrogenase (LDH), creatinine, and alkaline phosphatase serum levels, lower albumin levels, and higher WBC in PB and BM blast percentage. Patients who received IC were younger, with more frequent *t*-AML or neo-AML, higher BM blast cells percentage, and more

Table 1. Demographic and baseline characteristics of the study population (de novo vs sAML, n = 8521)

Characteristic	Overall		De novo		Secondary AML		P value
	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	
Total		8521 (100)		6211 (100)		2310 (100)	
Age, y	66 (18-104)	8507 (100)	64 (18-99)	6201 (100)	69 (18-104)	2306 (100)	<.001*
18-29		389 (5)		359 (6)		30 (1)	<.001
30-39		549 (6)		480 (8)		69 (3)	
40-49		823 (10)		706 (11)		117 (5)	
50-59		1285 (15)		998 (16)		287 (12)	
60-69		2087 (25)		1421 (23)		666 (29)	
70-79		2333 (27)		1535 (25)		798 (35)	
80-89		967 (11)		649 (10)		318 (14)	
≥90		74 (1)		53 (1)		21 (1)	
Gender		8432 (100)		6127 (100)		2304 (100)	
Male		4643 (55)		3266 (53)		1377 (60)	<.001
Female		3788 (45)		2861 (47)		927 (40)	
ECOG	1 (0-4)	6640 (100)	1 (0-4)	4819 (100)	1 (0-4)	1821 (100)	<.001*
0		1949 (29)		1481 (31)		468 (26)	<.001
1		2740 (41)		2028 (42)		712 (39)	
2		1239 (19)		852 (18)		387 (21)	
3		538 (8)		336 (7)		202 (11)	
4		174 (3)		122 (3)		52 (3)	
FAB subtype		8521 (100)		6211 (100)		2310 (100)	
M0/M6/M7		872 (10)		638 (10)		234 (10)	<.001
M1/M2		2301 (27)		1814 (29)		487 (21)	
M4/M5		2156 (37)		1764 (28)		392 (17)	
Not available		3192 (37)		1995 (32)		1197 (52)	
Hepatosplenomegaly		4380 (100)		3263 (100)		1117 (100)	.63
Yes		962 (22)		723 (22)		239 (21)	
No		3418 (78)		2540 (78)		878 (79)	
WBC, ×10⁹/L	9.2 (0-669)	7801 (100)	10.4 (0-669)	5762 (100)	6.6 (0.1-312)	2039 (100)	<.001*
≤5		3135 (40)		2199 (38)		936 (46)	<.001
5-10		883 (11)		651 (11)		232 (11)	
10-50		2137 (27)		1574 (27)		563 (28)	
>50		1646 (21)		1338 (23)		308 (15)	
Hemoglobin, g/dL	9 (1.3-18.7)	7204 (100)	9 (1.3-18.7)	5353 (100)	9 (3.3-17.2)	1851 (100)	.71*
Platelet count, ×10 ⁹ /L	56 (0-1645)	7225 (100)	57 (0-1419)	5368 (100)	51 (1-1645)	1857 (100)	<.001*
BM blasts, %	60 (0-100)	6612 (100)	64 (0-100)	4937 (100)	42 (0-100)	1675 (100)	<.001*
Creatinine, mg/dL	0.9 (0.1-10)	4439 (100)	0.88 (0.1-10)	3022 (100)	0.94 (0.1-9.2)	1417 (100)	<.001*
Urea, mg/dL	34 (2-267)	3931 (100)	33 (4-241)	2650 (100)	38 (2-267)	1281 (100)	<.001*
Uric acid, mg/dL	4.9 (0-45)	3649 (100)	4.8 (0-45)	2443 (100)	5.3 (0-21)	1206 (100)	<.001*
Bilirubin, mg/dL	0.6 (0.07-21)	4011 (100)	0.6 (0.1-21)	2692 (100)	0.67 (0.07-9.94)	1319 (100)	<.001*
AST, U/L	22 (2-1085)	3869 (100)	22 (2-1085)	2599 (100)	22 (5-496)	1270 (100)	.61*
ALT, U/L	20 (1-743)	3987 (100)	20 (1-743)	2682 (100)	19 (2-623)	1305 (100)	.004*
AP, U/L	73 (2-3571)	3570 (100)	71 (4-1474)	2385 (100)	78 (2-3571)	1185 (100)	<.001
Albumin, g/dL	3.6 (0.5-8.5)	3520 (100)	3.6 (0.5-8.5)	2388 (100)	3.7 (1.7-7)	1132 (100)	.03*
LDH, U/L	480 (12-42 630)	4827 (100)	486 (16-42 630)	3342 (100)	460 (12-13 420)	1485 (100)	.38*
Fibrinogen, mg/dL	426 (1-1500)	3686 (100)	423 (1-1150)	2610 (100)	430 (1-1500)	1076 (100)	.22*

ALT, alanine transaminase; AP, alkaline phosphatase; AST, aspartate transaminase; FAB, French-American-British.

*P compare continuous variables.

Table 1. (continued)

Characteristic	Overall		De novo		Secondary AML		P value
	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	
Cytogenetics		8521 (100)		6211 (100)		2310 (100)	
Normal		2892 (34)		2299 (37)		593 (26)	<.001
Abnormal		3192 (37)		2201 (35)		991 (43)	
No metaphases		592 (7)		436 (7)		156 (7)	
Not available		1845 (22)		1275 (21)		570 (22)	
MRC cytogenetic risk		6099 (100)		4501 (100)		1598 (100)	
Favorable		474 (8)		420 (9)		54 (3)	<.001
Intermediate		3919 (64)		3014 (67)		905 (57)	
Adverse		1706 (28)		1067 (24)		639 (40)	
FLT3-ITD		4291 (100)		3301 (100)		990 (100)	
Positive		789 (18)		677 (21)		112 (11)	<.001
Negative		3502 (82)		2624 (79)		878 (89)	
NPM1		4024 (100)		3097 (100)		927 (100)	
Positive		1173 (29)		1023 (33)		150 (16)	<.001
Negative		2851 (71)		2074 (67)		777 (84)	
Therapeutic approach		8520 (100)		6210 (100)		2310 (100)	
Intensive		4620 (54)		3744 (60)		876 (38)	<.001
Nonintensive		974 (11)		649 (10)		325 (14)	
HMA		780 (9)		452 (7)		328 (14)	
Clinical trial		192 (2)		113 (2)		79 (3)	
BSC		1467 (17)		923 (15)		544 (24)	
Unknown		487 (6)		329 (5)		158 (7)	

ALT, alanine transaminase; AP, alkaline phosphatase; AST, aspartate transaminase; FAB, French–American–British.
*P compare continuous variables.

favorable cytogenetics, and they showed a trend to have more *NPM1* mutations.

Disease characteristics and treatment according to type of sAML

Information related to type of sAML was available in 2000 patients; 884 (44%) had MDS-AML, 209 (10%) had MDS/MPN-AML, 226 (11%) had MPN-AML, 502 (25%) had *t*-AML, and 179 (9%) had neo-AML. Refractory cytopenia with excess blasts was the most frequent condition preceding MDS-AML (Figure 2A), chronic myelomonocytic leukemia for MDS/MPN-AML (Figure 2B), and essential thrombocythemia for MPN-AML (Figure 2C). Hematologic malignancies and breast cancer were the most frequent conditions preceding *t*-AML (Figure 2D), and 65% of patients with *t*-AML were previously treated with chemotherapy with or without radiotherapy (Figure 2E). Lung, prostate, and gynecologic cancer were the most frequent antecedents for patients with neo-AML (Figure 2F). The median latency period in patients with sAML was 34.1 months (range, 0.07-471.7), 71.8 months for MPN-AML (range, 2.2-360.8), 60.4 months for *t*-AML (range, 1.5-451.7), 59.2 months for neo-AML (range, 0.1-392.1), 20 months for MDS/MPN-AML (range, 0.1-272.6), and 16.2 months for MDS-AML (range, 0.2-327.6) ($P < .001$).

Table 3 shows characteristics and treatment according to the type of sAML. Patients with neo-AML were older, and only among *t*-AML was female gender more frequent. Patients with MDS-AML and *t*-AML

had better ECOG. Patients with MDS/MPN-AML had more frequent hepatosplenomegaly, French–American–British M4/M5, and higher WBC counts in PB, creatinine, uric acid, and urea serum levels. Patients with MPN-AML showed more hepatosplenomegaly and higher LDH and fibrinogen levels. *t*-AML and neo-AML showed higher blast percentage in BM, and more frequent favorable cytogenetics, *NPM1* and *FLT3*-ITD mutations. An adverse cytogenetic reaction was more frequent among MPN-AML and *t*-AML subgroups. IC was the main therapeutic choice for patients with *t*-AML and neo-AML (52% and 41%, respectively), whereas patients with MDS-, MDS/MPN-, and MPN-AML were more commonly treated with non-IC or HMA.

Response after front-line therapy in sAML

Response assessment after front-line therapy was available in 1486 (93%) patients with sAML actively treated (Figure 1). CR/CRi was achieved in 619 (41%) patients, 117 (8%) showed PR, 561 (38%) were resistant, and 189 (13%) died during induction (Table 4). CR/CRi rate was higher after IC (55%) followed by clinical trial (31%), non-IC (30%), and HMA (16%). Lower induction mortality was observed after HMA (9%). Among 619 patients who achieved a first CR/CRi, information about hematopoietic stem cell transplantation (HSCT) was available in 582: 51 (9%) received autologous HSCT (49 after IC and 2 after non-IC), 152 (26%) received an allogeneic HSCT (131 after IC, 18 after non-IC, and 3 after clinical trial), and no HSCT was performed in the remaining 379 (65%) patients.

Table 2. Characteristics of patients with sAML as per front-line treatment (n = 2152)

Characteristic	IC		Non-IC		HMA		Clinical trial		BSC		P value
	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	
Total	876 (100)	325 (100)	325 (100)	325 (100)	73 (29-88)	79 (100)	72 (31-89)	77 (23-104)	544 (100)	<.001*	
Age, y	61 (18-85)	875 (100)	72 (24-91)	325 (100)	73 (29-88)	327 (100)	72 (31-89)	77 (23-104)	542 (100)	<.001*	
Gender											
Male	870 (100)	325 (100)	325 (100)	325 (100)	73 (29-88)	79 (100)	72 (31-89)	77 (23-104)	544 (100)		
Female	518 (59)	199 (61)	199 (61)	209 (64)	73 (29-88)	52 (66)	72 (31-89)	77 (23-104)	305 (56)	.15	
	352 (41)	126 (39)	126 (39)	119 (36)	73 (29-88)	27 (34)	72 (31-89)	77 (23-104)	239 (44)		
ECOG											
0	1 (0-4)	726 (100)	1 (0-4)	256 (100)	1 (0-4)	273 (100)	1 (0-3)	2 (0-4)	433 (100)	<.001*	
1		252 (34)		62 (24)		50 (18)			57 (13)	<.001	
2		320 (44)		101 (40)		134 (49)			99 (23)		
3		113 (16)		59 (23)		70 (26)			126 (29)		
4		35 (5)		26 (10)		17 (6)			115 (27)		
		6 (1)		8 (3)		2 (1)			36 (8)		
Type of sAML											
MDS-AML, MDS/MPN, and MPN-AML		764 (100)		302 (100)		258 (100)			481 (100)	<.001	
t-AML		429 (56)		209 (69)		189 (73)			354 (74)		
Neo-AML		261 (34)		68 (23)		51 (20)			83 (17)		
		74 (10)		25 (8)		18 (7)			44 (9)		
FAB subtype											
M0/M6/M7		658 (100)		252 (100)		189 (100)			341 (100)	<.001	
M1/M2		100 (15)		37 (15)		26 (14)			59 (17)		
M4/M5		233 (35)		86 (34)		40 (21)			103 (30)		
Not available		196 (30)		56 (22)		28 (15)			92 (27)		
		129 (20)		73 (29)		95 (60)			87 (26)		
WBC, ×10 ⁹ /L	6.7 (0.1-289)	800 (100)	6.3 (0.4-312)	296 (100)	4.3 (0.5-214)	300 (100)	4.7 (0.5-158)	10.2 (0.2-310)	484 (100)	<.001*	
Hemoglobin, g/dL	9.1 (3.3-17.2)	730 (100)	8.8 (4.1-13.9)	287 (100)	8.9 (4.8-14.9)	293 (100)	9.0 (3.2-14.3)	8.9 (3.4-16.4)	393 (100)	.02*	
Platelet count, ×10 ⁹ /L	49 (2-1442)	731 (100)	50 (1-1645)	287 (100)	59 (3-870)	294 (100)	40 (7-350)	58 (1-1372)	396 (100)	.15*	
BM blasts, %	50 (0-100)	676 (100)	39 (3-100)	270 (100)	32 (0-100)	286 (100)	34 (16-94)	47 (3-100)	324 (100)	<.001*	
Creatinine, mg/dL	0.87 (0.07-7.2)	613 (100)	0.99 (0.39-3.63)	220 (100)	0.99 (0.47-7)	185 (100)	0.96 (0.42-1.84)	1.12 (0.25-9.2)	281 (100)	<.001*	
≤1.3		534 (87)		172 (78)		149 (81)			182 (65)	<.001	
>1.3		79 (13)		48 (22)		36 (19)			99 (35)		
Urea, mg/dL	32 (4-255)	547 (100)	42 (1.9-162)	205 (100)	41 (12-195)	170 (100)	38 (12-117)	47 (10-267)	246 (100)	<.001*	
Uric acid, mg/dL	4.7 (0-14.4)	540 (100)	5.6 (0-17.1)	187 (100)	5.7 (1.7-13.6)	158 (100)	5.3 (0.1-11)	6.1 (0.6-21)	218 (100)	<.001*	

*P compare continuous variables.

Table 2. (continued)

Characteristic	IC		Non-IC		HMA		Clinical trial		BSC		P value
	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	
Bilirubin, mg/dL	0.6 (0.1-8.1)	582 (100)	0.7 (0.1-9.9)	207 (100)	0.7 (0.2-5.3)	172 (100)	0.7 (0.2-3.9)	65 (100)	0.7 (0.2-8.1)	248 (100)	.01*
AST, U/L	22 (5-213)	565 (100)	22 (8-154)	204 (100)	20 (7-88)	163 (100)	22 (11-99)	56 (100)	23 (7-141)	238 (100)	.18*
ALT, U/L	20 (3-498)	577 (100)	19 (2-233)	206 (100)	16 (3-91)	170 (100)	18 (4-623)	59 (100)	19 (3-168)	247 (100)	.02*
AP, U/L	76 (2-1252)	533 (100)	75 (18-812)	186 (100)	73 (19-894)	159 (100)	75 (42-416)	55 (100)	95 (23-3571)	208 (100)	<.001*
Albumin, g/dL	3.7 (2-5.8)	508 (100)	3.7 (2-5.2)	187 (100)	3.8 (2.7-5.9)	143 (100)	4.1 (2-5)	61 (100)	3.4 (1.7-7)	197 (100)	<.001*
≤3.5		212 (42)		86 (46)		53 (37)		18 (30)		107 (54)	.001
> 3.5		296 (58)		101 (54)		90 (63)		43 (70)		90 (46)	
LDH, U/L	471 (12-9540)	602 (100)	437 (57-12 460)	253 (100)	385 (61-4266)	220 (100)	390 (144-3557)	63 (100)	617 (102-13 420)	302 (100)	<.001*
Cytogenetics											
Normal		876 (100)		325 (100)		328 (100)		79 (100)		544 (100)	
Abnormal		255 (29)		93 (29)		107 (33)		29 (37)		90 (17)	<.001
No metaphases		422 (48)		161 (50)		152 (48)		41 (52)		192 (35)	
Not available		76 (9)		27 (8)		19 (6)		2 (3)		31 (6)	
		123 (14)		44 (14)		50 (15)		7 (9)		231 (42)	
MRC cytogenetic risk											
Favorable		690 (100)		256 (100)		263 (100)		71 (100)		277 (100)	
Intermediate		40 (6)		8 (3)		2 (1)		0 (0)		4 (2)	<.001
Adverse		386 (56)		147 (57)		150 (57)		46 (65)		148 (53)	
		264 (38)		101 (40)		111 (42)		25 (35)		125 (45)	
FLT3-ITD											
Positive		467 (100)		174 (100)		118 (100)		70 (100)		122 (100)	
Negative		54 (12)		20 (11)		10 (8)		10 (14)		12 (10)	.76
		413 (88)		154 (89)		10108 (92)		60 (85)		110 (90)	
NPM1											
Positive		427 (100)		169 (100)		113 (100)		67 (100)		113 (100)	
Negative		85 (20)		25 (15)		16 (14)		8 (12)		11 (10)	.06
		342 (80)		144 (85)		97 (86)		59 (88)		102 (90)	
Period											
1990-1994		876 (100)		325 (100)		328 (100)		79 (100)		544 (100)	
1995-1999		33 (4)		7 (2)		0 (0)		0 (0)		13 (2)	<.001
2000-2004		68 (8)		2 (1)		0 (0)		0 (0)		28 (5)	
2005-2009		92 (11)		2 (1)		3 (1)		0 (0)		66 (12)	
2010-2014		168 (19)		23 (7)		51 (16)		0 (0)		164 (30)	
2015-2019		290 (33)		171 (53)		132 (40)		23 (29)		192 (35)	
		225 (26)		120 (37)		142 (43)		56 (71)		81 (15)	

*P compare continuous variables.

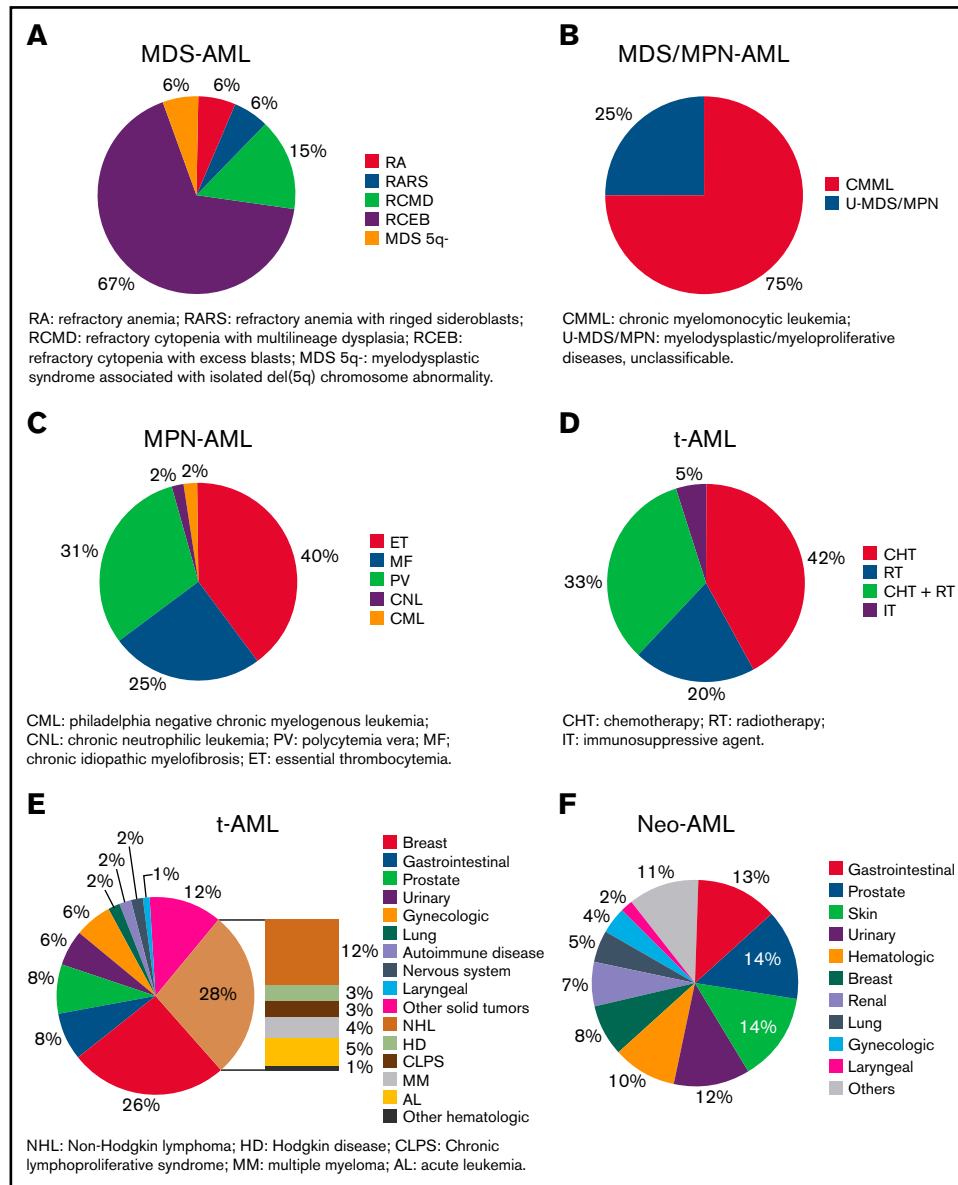


Figure 2. Description of disorders diagnosed before AML. (A) Type of previous MDS in patients included in MDS-AML group. Patients with nonspecified and unclassifiable MDS were not included in this figure. (B) Type of previous MDS/MPN in patients included in MDS/MPN-AML group. (C) Type of previous MPN in patients included in MPN-AML group. (D) Type of previous therapy in patients included in *t*-AML group. (E) Type of previous neoplasm in treated patients included in *t*-AML group. (F) Type of previous solid neoplasm in patients with cancer antecedents without prior treatment.

CR/CRi rate after IC was superior among patients with *t*-AML and neo-AML (64% and 63%, respectively), as compared with MDS-MDS/MPN- and MPN-AML (52%, 44%, and 45%, respectively) ($P < .001$). Response after other therapies and HSCT rates were similar among all sAML subgroups (supplemental Table 2).

Multivariate analyses for complete remission

When all patients receiving IC were included (de novo and sAML) in a multinomial logistic regression, sAML ($P < .001$), age ($P < .001$), ECOG ($P < .001$), WBC count ($P < .001$), and creatinine level >1.2 mg/dL ($P = .003$) were independent adverse prognostic factors for CR/CRi, whereas presence of NPM1 mutation and

favorable and intermediate cytogenetic risk were independent favorable prognostic factors for CR/CRi (all $P < .001$).

Among patients with sAML, age ($P < .001$), ECOG ($P = .004$), WBC count ($P = .002$), and bilirubin level >1.2 mg/dL ($P = .03$) were independent prognostic factors for CR/CRi. NPM1 mutation ($P < .001$), favorable and intermediate cytogenetic risk (both $P < .001$), *t*-AML ($P < .001$), and neo-AML ($P = .01$) were independent favorable prognostic factors for CR/CRi.

OS

The median OS of the 8521 patients included in the study was 9.1 months (95% CI, 8.7-9.6). Median OS was higher in patients with

Table 3. Disease and patient characteristics according to the type of sAML (n = 2000)

Characteristic	MDS-AML		MDS/MPN-AML		MPN-AML		t-AML		neo-AML		P value
	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	
Global	884 (100)	209 (100)	226 (100)	502 (100)	179 (100)	<.001*					
Age, y	70 (19-97)	883 (100)	69 (31-92)	208 (100)	68 (31-92)	225 (100)	67 (18-104)	502 (100)	73 (26-93)	179 (100)	<.001*
Gender											
Male	882 (100)	209 (100)	226 (100)	502 (100)	179 (100)	<.001					
Female	548 (62)	148 (71)	129 (57)	248 (49)	121 (68)	<.001					
	334 (38)	61 (29)	97 (43)	254 (51)	58 (32)						
ECOG											
0	1 (0-4)	670 (100)	1 (0-4)	188 (100)	1 (0-4)	435 (100)	1 (0-4)	169 (100)	1 (0-4)	169 (100)	<.001*
1	185 (28)	34 (21)	39 (21)	130 (30)	40 (24)	<.001					
2	268 (40)	62 (38)	73 (39)	178 (41)	64 (38)						
3	139 (21)	36 (22)	35 (19)	80 (18)	44 (26)						
4	67 (10)	25 (15)	27 (14)	38 (9)	18 (11)						
	11 (2)	8 (5)	14 (7)	9 (2)	3 (2)						
FAB subtype											
M0/M6/M7	884 (100)	209 (100)	226 (100)	502 (100)	179 (100)	<.001					
M1/M2	80 (9)	4 (2)	42 (19)	65 (13)	22 (12)	<.001					
M4/M5	230 (26)	18 (9)	30 (13)	118 (24)	51 (28)						
Not available	100 (11)	79 (38)	15 (7)	121 (24)	54 (30)						
	474 (54)	108 (52)	139 (61)	198 (39)	52 (29)						
Hepatosplenomegaly											
Yes	419 (100)	114 (100)	112 (100)	290 (100)	142 (100)	<.001					
No	77 (18)	37 (32)	44 (39)	52 (18)	24 (17)						
	342 (82)	77 (68)	68 (61)	238 (82)	118 (83)						
WBC, ×10 ⁹ /L	4.4 (0.1-310.4)	786 (100)	23.5 (0.1-300.2)	169 (100)	10.8 (0.3-312)	198 (100)	5.1 (0.1-289)	466 (100)	10.7 (0.2-230.7)	173 (100)	<.001*
Hemoglobin, g/dL	8.9 (3.3-17.2)	690 (100)	9.1 (4-15.1)	154 (100)	8.7 (4.7-15)	173 (100)	9.1 (3.2-15.2)	452 (100)	9.1 (4.4-14.4)	170 (100)	.23*
Platelet count, ×10 ⁹ /L	45 (1-1645)	691 (100)	55 (6-459)	155 (100)	74 (1-1372)	173 (100)	47 (2-1442)	456 (100)	58 (5-462)	171 (100)	<.001*
BM blasts, %	36 (2-100)	653 (100)	40 (1-100)	136 (100)	35 (0-94)	148 (100)	55 (0-100)	408 (100)	58 (10-100)	160 (100)	<.001*
Creatinine, mg/dL	0.97 (0.32-9.2)	500 (100)	1.07 (0.29-7.2)	133 (100)	0.95 (0.25-5.09)	141 (100)	0.9 (0.07-5.48)	416 (100)	0.97 (0.4-8.5)	163 (100)	<.001*
Urea, mg/dL	38 (4-192)	456 (100)	44 (11-255)	121 (100)	39 (8-242)	126 (100)	35 (2-243)	391 (100)	41 (11-267)	146 (100)	<.001*
Uric acid, mg/dL	5 (0.1-19)	413 (100)	6.5 (0.1-17.1)	119 (100)	5.8 (0-16.2)	121 (100)	4.9 (0-14.4)	366 (100)	5.3 (0-13)	143 (100)	<.001*
Bilirubin, mg/dL	0.7 (0.14-7.5)	449 (100)	0.75 (0.22-7)	127 (100)	0.67 (0.2-5.5)	137 (100)	0.64 (0.07-9.94)	392 (100)	0.62 (0.1-4.57)	151 (100)	.06*
AST, U/L	22 (7-171)	451 (100)	22 (6-496)	119 (100)	23 (8-151)	124 (100)	22 (5-213)	383 (100)	21 (8-338)	150 (100)	.81*
ALT, U/L	18 (2-272)	461 (100)	16 (3-515)	121 (100)	18 (3-160)	131 (100)	20 (3-623)	395 (100)	19 (5-328)	152 (100)	.24*

The type of sAML was not available in 310 patients.

*P compare continuous variables.

Table 3. (continued)

Characteristic	MDS-AML		MDS/MPN-AML		MPN-AML		t-AML		neo-AML		P value
	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	Median (range)	n (%)	
AP, U/L	78 (29-702)	412 (100)	82 (14-1158)	115 (100)	80 (26-1525)	120 (100)	77 (19-3571)	354 (100)	75 (2-3402)	143 (100)	.41*
Albumin, g/dL	3.7 (2.1-7)	389 (100)	3.7 (1.7-5.8)	113 (100)	3.7 (1.9-5.2)	117 (100)	3.7 (1.8-5.8)	340 (100)	3.7 (2.6-4.9)	136 (100)	.95*
LDH, U/L	416 (13-9168)	571 (100)	612 (96-11380)	138 (100)	768 (102-10560)	147 (100)	436 (12-9540)	398 (100)	561 (112-13420)	149 (100)	<.001*
Fibrinogen, mg/dL	423 (1-1033)	358 (100)	410 (1-940)	111 (100)	467 (3-889)	115 (100)	422 (3-983)	328 (100)	444 (2-1500)	137 (100)	.04*
Cytogenetics	884 (100)	209 (100)									
Normal	247 (28)	61 (29)				226 (100)		502 (100)		179 (100)	
Abnormal	361 (41)	72 (34)			49 (22)	49 (22)		107 (21)		60 (34)	<.001
No metaphases	65 (7)	10 (5)			102 (45)	102 (45)		266 (53)		74 (41)	
Not available	211 (24)	66 (32)			12 (5)	12 (5)		45 (9)		14 (8)	
MRC cytogenetic risk	612 (100)	134 (100)									
Favorable	4 (1)	1 (1)				152 (100)		382 (100)		137 (100)	
Intermediate	377 (62)	94 (70)				2 (1)		31 (8)		11 (8)	<.001
Adverse	231 (38)	38 (28)				64 (42)		172 (45)		88 (64)	
						86 (57)		179 (47)		38 (28)	
FLT3-ITD	340 (100)	104 (100)									
Positive	35 (10)	11 (11)				86 (100)		288 (100)		114 (100)	
Negative	305 (90)	93 (89)				3 (3)		26 (9)		24 (21)	<.001
						83 (97)		262 (91)		90 (79)	
NPM1	312 (100)	103 (100)									
Positive	34 (11)	13 (13)				85 (100)		265 (100)		105 (100)	
Negative	278 (89)	90 (87)				7 (8)		50 (19)		28 (27)	<.001
						78 (92)		215 (81)		77 (73)	
Therapeutic approach	884 (100)	209 (100)									
IC	294 (33)	66 (32)				226 (100)		502 (100)		179 (100)	
Non-IC	149 (17)	35 (17)				69 (31)		261 (52)		74 (41)	<.001
HMA	127 (14)	27 (13)				25 (11)		68 (14)		25 (14)	
Clinical trial	35 (4)	11 (5)				35 (15)		51 (10)		18 (10)	
BSC	234 (26)	50 (24)				9 (4)		14 (3)		7 (4)	
Not available	45 (5)	20 (10)				70 (31)		83 (17)		44 (25)	
						18 (8)		25 (5)		11 (6)	

The type of sAML was not available in 310 patients.
*P compare continuous variables.

Table 4. Response and HSCT rates, OS, and EFS according to therapeutic approach in patients with sAML

Variable	Overall	IC	Non-IC	HMA	Clinical trial	BSC	P value
Response, n (%)	1489 (100)	844 (100)	303 (100)	274 (100)	68 (100)		
ORR (CR + CRi)	620 (41)	467 (55)	89 (30)	43 (16)	21 (31)		<.001
CR	559 (37)	437 (52)	71 (23)	33 (12)	18 (26)		
Cri	61 (4)	30 (4)	18 (6)	10 (4)	3 (4)		
PR	118 (8)	54 (6)	30 (10)	21 (8)	13 (19)		
Resistance	562 (38)	220 (26)	138 (46)	184 (67)	20 (30)		
Death	189 (13)	103 (12)	46 (15)	26 (9)	14 (21)		
HSCT in first CR/CRi, n (%)*	584 (100)	439 (100)	83 (100)	43 (100)	19 (100)		
Allogeneic HSCT rate, n (%)	151 (26)	130 (30)	18 (22)	0 (0)	3 (16)		<.001
Autologous HSCT rate, n (%)	51 (9)	49 (11)	2 (2)	0 (0)	0 (0)		
No HSCT, n (%)	382 (65)	260 (59)	63 (76)	43 (100)	16 (84)		
OS, n (%)	2310 (100)*	876 (100)	325 (100)	328 (100)	79 (100)	544 (100)	
Median (CI 95%), mo	5.6 (5.2-6.3)	9.9 (8.9-11.4)	5.5 (4.6-7.4)	9.0 (7.6-10.6)	7.7 (6.4-10.6)	1.2 (1.0-1.5)	<.001
1 y (CI 95%)	31.4 (29.3-33.4)	44.4 (41.1-48.0)	29.7 (24.9-35.5)	40.3 (35.0-46.3)	32.9 (22.8-47.5)	6.5 (4.7-9.0)	
5 y (CI 95%)	8.2 (6.8-9.6)	15.7 (13.0-18.9)	6.4 (3.7-11.0)	3.7 (1.8-7.6)	7.1 (2.1-23.7)	NA (NA-NA)	
EFS, n (%)	1394 (100)	810 (100)	288 (100)	230 (100)	66 (100)	NA	<.001
Median (CI 95%), mo	3.6 (3.2-4.0)	4.1 (3.5-5.7)	1.7 (1.5-2.1)	4.8 (4.0-6.0)	3.3 (1.9-5.9)	NA	
1 y (CI 95%)	22.2 (20.0-24.5)	28.3 (25.3-31.7)	13.7 (10.2-18.4)	23.0 (18.2-29.2)	10.3 (4.7-22.6)	NA	
5 y (CI 95%)	8.4 (6.8-10.0)	11.3 (9.1-14.1)	6.0 (3.6-9.7)	2.5 (1.0-6.1)	6.2 (2.1-17.9)	NA	

NA, not applicable; ORR, overall response rate;

*Data from patients in CR/CRi after front-line therapy. Global OS results includes all patients with sAML (2310 patients), but in 162 patients the therapeutic approach was not available. Including only patients with therapeutic approach available (2148 patients), the median OS was 8.9 mo (8.0-9.6), 1 y OS was 40% (38-43), and 5 y OS was 11% (9-13).

de novo than sAML (10.9 [95% CI, 10.3-11.5] vs 5.6 months [95% CI, 5.2-6.3], respectively; $P < .001$), with 1- and 5-year OS of 47.8% (95% CI, 46.5-49.1) vs 31.4% (95% CI, 29.3-33.4), and 23.4% (95% CI, 22.2-24.7) vs 8.3% (95% CI, 6.7-9.7), respectively ($P < .001$) (Figure 3A).

Patients with sAML who received IC or HMA had better median OS than others (9.9 and 9.0 months, respectively), but the 5-year OS was better among the IC group (15.7%) (Figure 3B; Table 4). Favorable cytogenetic risk (Figure 3C) and *NPM1* mutations (Figure 3D) were associated with prolonged OS, whereas *FLT3*-ITD mutations did not affect OS (Figure 3E). OS was significantly lower in patients with sAML after hematologic disorders (together MDS-AML, MDS/MPN-AML, and MPN-AML) as compared with *t*-AML and neo-AML (median OS 5.3 [95% CI, 5.0-6.1] vs 6.1 [95% CI, 5.1-7.8] vs 5.7 [95% CI, 4.1-7.8] months, respectively; $P = .04$) (Figure 3F). Other risk factors for OS were identified in univariate analyses (ie, age, ECOG, WBC count, or alkaline phosphatase) (supplemental Table 3).

Patients who achieved CR/CRi had higher median OS as compared with patients with PR or resistance (19.5 [95% CI, 16.7-23.8], 15.5 [95% CI, 14.6-18.9], 10.8 [95% CI, 7.7-13.3], and 5.8 [95% CI, 5.3-6.5] months, respectively; $P < .001$). Patients in first CR/CRi allocated to allogeneic HSCT had longer median OS than those assigned to autologous therapy or chemotherapy exclusively (44.9 [95% CI, 30.2-NA] vs 31.5 [95% CI, 23.1-59.8] vs 14.9 [13.7-16.9] months, respectively; $P < .001$) (supplemental Figure 1A-B).

After IC front-line, median OS was significantly better among patients with neo-AML as compared with other sAML (supplemental

Table 4), but no differences were observed between neo-AML and de novo AML (14.6 [95% CI, 10.3-42.4] vs 17.2 [95% CI, 16.0-18.6], respectively; $P = .63$) (supplemental Figure 2A). No differences were observed among patients treated with HMA (supplemental Figure 2B) or non-IC according to the type of sAML although patients with nonsignificant, neo-AML aged <60 years and those with intermediate cytogenetics had better 5-year OS compared with other sAML subsets (supplemental Table 4).

Among 666 patients with MDS-AML, MDS/MPN-AML, and MPN-AML with available information of prior HMAs exposure, OS was significantly lower among those treated with HMAs before AML transformation ($P < .001$) (supplemental Figure 3A). We also explored the prognosis of 101 patients with *t*-AML who received only prior radiotherapy as compared with other sAML types and de novo AML (supplemental Figure 3B). After including this new category in the multivariate analysis, prior administration of radiotherapy exclusively was not an independent prognostic factor: hazard ratio (HR) was 1.06 (95% CI, 0.84-1.35; $P = .61$).

Multivariate analyses for OS

Among patients with sAML with an age between 30 and 59 years, neo-AML, platelet count $>20 \times 10^9/L$, albumin >3.5 g/dL, favorable and intermediate cytogenetic risk, and *NPM1* mutation had a positive impact on OS. ECOG ≥ 2 , WBC count $\geq 10 \times 10^9/L$, blast cells in BM $>70\%$, creatinine >1.3 mg/dL, and alkaline phosphatase >150 U/L were independent adverse prognostic factors for OS (supplemental Table 5).

When all patients were included (de novo and sAML) in a multivariate Cox regression, sAML was an independent adverse prognostic

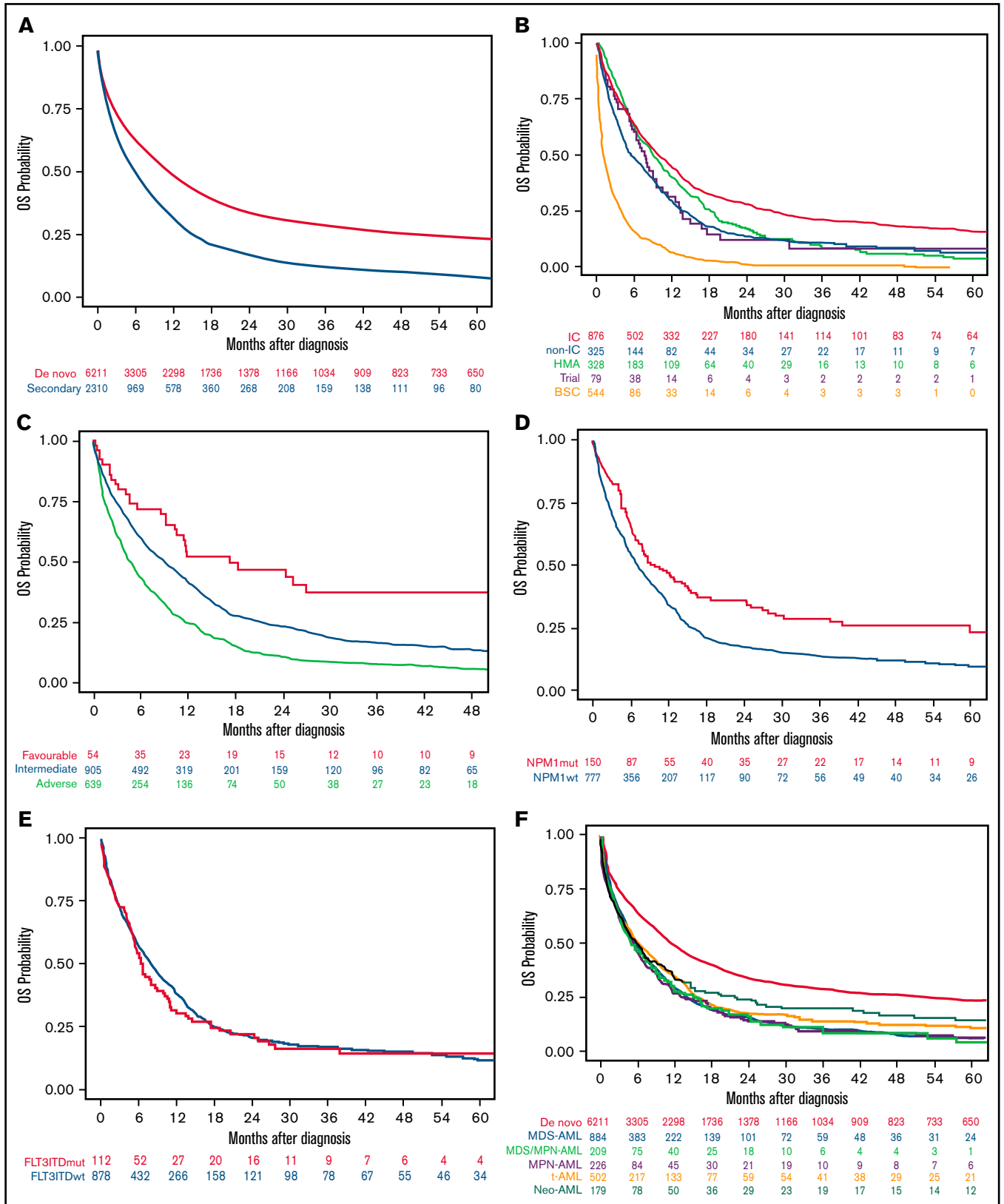


Figure 3. OS of the AML patients included in the study according to different disease characteristics or therapeutic approach. (A) OS in the entire cohort of 8521 patients with AML according to type of AML ($P < .001$). (B) OS in secondary AML according to therapeutic approach ($P < .001$). (C) OS in secondary AML according to cytogenetic risk ($P < .001$). (D) OS in secondary AML according to NPM1 mutation ($P < .001$). (E) OS in secondary AML according to FLT3-ITD mutation ($P = .45$). (F) OS in secondary AML according to the different subgroups ($P = .04$).

factor for OS ($P = .01$; HR 1.08 [95% CI, 1.02-1.15]). Because neo-AML had different prognosis than other sAML types, we performed a multivariate analysis, including neo-AML in the de novo AML category, showing that sAML (without neo-AML) was a significant predictor for OS ($P = .008$; HR 1.09 [95% CI, 1.02-1.16]). Other independent prognostic factors are detailed in supplemental Table 6. We analyzed the effect of cytogenetics with type of AML interaction, showing that in all MRC risk subgroups, the presence of sAML was accompanied by lower OS (supplemental Figure 4A).

We also analyzed the trend in OS across the study period, showing that OS improved gradually (90s vs 00s vs 10s decade) among patients with de novo AML, but such improvement was not observed among patients with sAML (supplementary Figure 4B).

EFS

The median EFS of 1394 evaluable patients with sAML was 3.6 months (95% CI, 3.2-4.0), with 1- and 5-year EFS of 22.2% (95% CI, 20.0-24.5) and 8.4% (95% CI, 6.8-10.0), respectively (supplemental Figure 5A). Higher median EFS was observed after IC, HMA, and clinical trials compared with non-IC regimens (4.1 months [95% CI, 3.5-5.7], 4.8 months [95% CI, 4.0-6.0], 3.3 months [95% CI, 1.9-5.9], and 1.7 months [95% CI, 1.5-2.1] respectively; $P < .001$) (supplemental Figure 5B).

Discussion

In this study we reported the largest cohort of patients with sAML to date and dissected their characteristics, treatment patterns, and main outcomes. We showed in a real-life setting that sAML represents roughly 25% of all AML cases, with worse prognosis and different baseline characteristics and patterns of care as compared with de novo AML (especially for those arising after MDS, MDS/MPN, and MPN). Nevertheless, we highlighted that patients with neo-AML have similar outcomes and biology than patients with de novo AML. It should be noted that therapeutic results were equally poor for unfit patients with de novo and sAML; thus, the adverse prognostic impact of sAML is mainly applicable to the eligible for IC setting.

In our registry, 27.1% of patients with AML were classified as sAML, which is superior to a median prevalence of 22.0% (range: 5.7% to 42.1%) previously reported.^{3-5,10,15-27} However, we have also considered as sAML those patients with AML following prior neoplasia without exposure to leukemogenic agents (namely neo-AML, 2.1%), which is uncommon in prior series.¹⁶ Regarding well-recognized sAML subsets, our frequencies are in line with prior studies showing 13.1% (range: 5.7% to 26.5%) MDS-AML, 7.1% (range: 1.6% to 8.3%) MPN-AML, and 6.2% (range: 1.9% to 13.7%) *t*-AML.^{3,4,15-31} As expected, we observed shorter median latency in MDS- and MDS/MPN-AML (<2 years),⁴ whereas leukemic transformation occurred over a 5-year period in *t*-AML^{4,25} and even longer in MPN-AML.³² The latency period could depend on the cumulative dose, dose intensity, and type of preceding chemotherapy and/or radiation therapy,^{33,34} but those factors were not analyzed here.

We confirm that sAML is closely associated with older age, comorbidities (ie, renal and liver dysfunction), worse performance status, and unfavorable genetic features.^{4,7-9} We also show that male gender was more predominant among patients with sAML compared with de novo AML. The exception was *t*-AML, as previously

reported,²⁵ in which breast cancer history was present in 25%. Surprisingly, 28% of *t*-AML cases evolved from prior hematological malignancies (eg, lymphoma), highlighting an increased risk of developing sAML in this setting. Another interesting finding was that patients with MDS- and *t*-AML had less WBC and BM blasts at diagnosis because they are usually less proliferative and can show signs and symptoms of hematopoietic insufficiency due to prior anti-neoplastic therapies.³⁵ On the contrary, MPN- and MDS/MPN-AML showed signs and symptoms related to the previous disorder, such as increased WBC, M4/M5 morphology, hepatomegaly, and splenomegaly.³⁶ We found that low-risk karyotype was uncommon, especially for MDS-, MDS/MPN-, and MPN-AML,⁴ whereas high-risk karyotype was frequent, especially in *t*-AML^{4,24} and MPN-AML. Regarding molecular findings, we could only analyze *NPM1* and *FLT3*-ITD mutations, confirming previous studies showing that these lesions are less prevalent among sAMLs.³⁷⁻³⁹ Interestingly, although neo-AML and *t*-AML shared 8% core-binding factor (CBF) rearrangements, only neo-AML cases showed a prevalence for *NPM1* and *FLT3*-ITD comparable to de novo AML, supporting that they should not be considered sAML.

The optimal treatment options for patients with sAML are not yet established. This quandary comes from the lack of studies in this subset of patients because they are generally excluded from trials and protocols.^{22,40} As expected, this study shows that few patients with sAML received up-front IC regimens (38%) compared with de novo (60%),^{3-5,16,21,24,27,28} curative approaches being the preferred option only in a subset of favorable risk sAML subjects (ie, younger, CBF, and *NPM1*). Only 8 sAML registry studies provided data regarding CR/CRi after IC,^{3-5,17,22,24,27} obtaining a median of 50.7% (range: 21% to 63%), slightly lower than in our series (55%). As in other series, we found better CR/CRi rates among patients with *t*-AML (63%) followed by MDS-AML (52%), and the worst rates were obtained among patients with MDS/MPN- and MPN-AML.^{3,4,41} Of note, 3 + 7 and FLAG-Ida regimens were administered in most IC patients in our historical sAML cohort, whereas only 6 patients received CPX-351. Our CR/CRi rate after IC was >48% in the CPX-351 and 33% in the 3 + 7 arm of a recently published phase 3 study.⁴² However, we should avoid comparisons because this clinical trial included patients ≥ 65 years and excluded those with MPN-AML. Scarce studies have analyzed the CR/CRi rates after HMA in sAML, showing 12% to 36% CR/CRi, in line with 16% herein reported.^{43,44} Interestingly, we show 30% CR/CRi after non-IC for unfit patients (ie, FLUGA regimen or similar), but this was not accompanied by improved survival compared with HMA.⁴⁵

We reproduced significantly poorer median OS in sAML (5.6 months) as compared with de novo AML (10.9 months).^{3-5,16,19,21,24-28} We can affirm that median OS in different studies is affected by the proportion of IC-treated patients in each series. As an example, two registries reported a median OS of 7 and 8.6 months,^{17,27} with percentage of IC treatment of 56% and 57%, respectively, whereas median OS was 5.8 months in a study with only 28% IC²⁴ (similar to our registry, in which 38% received IC). We confirmed that HMA could lead to comparable median OS than IC strategies, but long-term survival might be improved after IC approaches.⁴⁴ We hypothesized that sAML patients achieving a CR/CRi should be considered for allogeneic HSCT to prolong survival, but few data have been published comparing patients with or without HSCT after induction therapy in this setting.⁴¹ The population-based study in sAML by Nilsson et al reported improved 5-year OS after allogeneic HSCT (18% to

32%) compared with non-HSCT (2% to 10%). Here we show improved median OS in patients undergoing allogeneic or autologous HSCT in first CR/CRi (45 and 31 months, respectively) compared with 65% who did not receive transplant (15 months). We should cautiously interpret these findings, acknowledging the inherent selection bias and the fact that analyses were not time dependent.

The dilemma about considering sAML an independent prognostic factor remains unsolved because published manuscripts revealed discrepant results.^{5,21,22,46} In this large study, we showed that sAML is an independent risk factor. Nevertheless, in line with a German cohort of patients with *t*-MN,²¹ significantly worse OS was observed only among patients receiving IC strategies, as compared with de novo AML, whereas therapeutic outcomes were as bad among all unfit patients. Some studies have shown a different prognosis depending on the type of sAML; patients with MPN who develop a leukemic transformation could have dismal outcomes.^{43,47} The population-based study by Hulegardh et al reported 1-year OS of 10% in MPN-AML, 20% in *t*-AML, and 43% in MDS-AML.³ However, in our cohort, we observed lower OS among patients with MDS/MPN-, MDS-, and MPN-AML compared with *t*-AML. As previously published, patients with sAML with CBF had a longer OS than those with intermediate and adverse genetic risks.⁴⁸ However, the prognostic impact of some well-established gene mutations in sAML remains unclear (eg, *FLT3* and *NPM1*). Our study supports the favorable OS impact of *NPM1* suggested by Stölzel et al,⁴⁹ but we were unable to demonstrate any prognostic role of *FLT3*-ITD mutations in sAML. Our multivariable model confirmed previous risk factors for OS in sAML as age, ECOG, adverse cytogenetic risk, and lower platelet count,^{10,16,21,22,24,49-52} but we also found new independent variables such as higher WBC count and BM blast percentage, low albumin levels, creatinine >1.3 mg/dL, and increased alkaline phosphatase. We were not able to confirm an independent favorable risk of patients with *t*-AML who received only prior radiotherapy, as suggested by Nardi et al.⁵³

Our study has some limitations: (1) This is a real-life analysis, but our registry was not capturing, by far, most patients with AML in Spain and Portugal. (2) Molecular data (eg, *P53* and *IDH*) were not analyzed. (3) The study period started in 1990, but most cases of sAML were included after 2010 (Table 2). (4) For some variables, the proportion of missing data precluded granular analyses (eg, impact of cumulative doses of cytotoxic agents).

In conclusion, this large study confirms that the frequency of adverse features, such as older age, worse performance status, and adverse karyotype, is by far higher in sAML than in de novo AML. In addition, sAML itself should be considered an independent risk factor, especially for patients treated with IC approaches. Although the best therapeutic results are obtained after IC followed by an allogeneic HSCT, this strategy is only accomplished in a minority of patients. Given the challenging condition that they represent, obtaining improvements in sAML should be a priority, warranting that this field become an active area of basic and clinical research in the forthcoming years.

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Authorship

Contribution: D.M.-C. and P.M. conceived the study; D.M.-C., J.E.M.-V., and P.M. analyzed and interpreted the data and wrote the paper; D.M.-C. and P.M. performed the statistical analyses; D.M.-C., J.E.M.-V., J.S., P.M.-S., E.R.-A., C.G., E.A., J.B., J.L.L.-L., T.B., A.E., M.C., C.R.-M., M.L.-P., M.T., L.A., M.L.A., M.J.S., J.L., J.I.R.-G., C.B., L.C.-B., R.G.-B., E.L.-R., S.V., P.H., D.G.-B., M.M.H., G.V.E., M.I.G.-R., A.C., G.B., A.B., J.M., B.B., M.A.S., and P.M. included data of patients treated in their institutions, reviewed the manuscript, and contributed to the final draft.

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A complete list of the institutions and clinicians participating in the PETHEMA epidemiologic registry of acute myeloid leukemia and acute promyelocytic leukemia appears in "Appendix."

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Appendix

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References

1. Leone G, Mele L, Pulsoni A, Equitani F, Pagano L. The incidence of secondary leukemias. *Haematologica*. 1999;84(10):937-945.
2. Arber DA, Orazi A, Hasserjian R, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood*. 2016;127(20):2391-2405.
3. Granfeldt Østgård LS, Medeiros BC, Sengeløv H, et al. Epidemiology and clinical significance of secondary and therapy-related acute myeloid leukemia: a national population-based cohort study. *J Clin Oncol*. 2015;33(31):3641-3649.
4. Hulegårdh E, Nilsson C, Lazarevic V, et al. Characterization and prognostic features of secondary acute myeloid leukemia in a population-based setting: a report from the Swedish Acute Leukemia Registry. *Am J Hematol*. 2015;90(3):208-214.
5. Sztokowski T, Rohon P, Zapletalova L, Sicova K, Hubacek J, Indrak K. Secondary acute myeloid leukemia: a single center experience. *Neoplasma*. 2010;57(2):170-178.

6. Xu X-Q, Wang J-M, Gao L, et al. Characteristics of acute myeloid leukemia with myelodysplasia-related changes: a retrospective analysis in a cohort of Chinese patients. *Am J Hematol*. 2014;89(9):874-881.
7. Krug U, Röllig C, Koschmieder A, et al; Study Alliance Leukemia Investigators. Complete remission and early death after intensive chemotherapy in patients aged 60 years or older with acute myeloid leukaemia: a web-based application for prediction of outcomes. *Lancet*. 2010;376(9757):2000-2008.
8. Walter RB, Othus M, Borthakur G, et al. Prediction of early death after induction therapy for newly diagnosed acute myeloid leukemia with pretreatment risk scores: a novel paradigm for treatment assignment. *J Clin Oncol*. 2011;29(33):4417-4423.
9. Larson RA. Is secondary leukemia an independent poor prognostic factor in acute myeloid leukemia? *Best Pract Res Clin Haematol*. 2007;20(1):29-37.
10. Grimwade D, Walker H, Harrison G, et al; Medical Research Council Adult Leukemia Working Party. The predictive value of hierarchical cytogenetic classification in older adults with acute myeloid leukemia (AML): analysis of 1065 patients entered into the United Kingdom Medical Research Council AML11 trial. *Blood*. 2001;98(5):1312-1320.
11. Cheson BD, Bennett JM, Kopecky KJ, et al; International Working Group for Diagnosis, Standardization of Response Criteria, Treatment Outcomes, and Reporting Standards for Therapeutic Trials in Acute Myeloid Leukemia. Revised recommendations of the International Working Group for diagnosis, standardization of response criteria, treatment outcomes, and reporting standards for therapeutic trials in acute myeloid leukemia. *J Clin Oncol*. 2003;21(24):4642-4649.
12. Kaplan EL, Meier P. Nonparametric estimation from incomplete observations. *J Am Stat Assoc*. 1958;53(282):457-481.
13. Mantel N. Evaluation of survival data and two new rank order statistics arising in its consideration. *Cancer Chemother Rep*. 1966;50(3):163-170.
14. Kalbfleisch JD, Prentice RL. *The Statistical Analysis of Failure Time Data*. New York, NY: John Wiley & Sons; 1980.
15. Grimwade D, Walker H, Oliver F, et al; The Medical Research Council Adult and Children's Leukaemia Working Parties. The importance of diagnostic cytogenetics on outcome in AML: analysis of 1,612 patients entered into the MRC AML 10 trial. *Blood*. 1998;92(7):2322-2333.
16. Aström M, Bodin L, Nilsson I, Tidefelt U. Treatment, long-term outcome and prognostic variables in 214 unselected AML patients in Sweden. *Br J Cancer*. 2000;82(8):1387-1392.
17. Pagano L, Pulsoni A, Tosti ME, et al; Gruppo Italiano Malattie Ematologiche Maligne dell'Adulto. Clinical and biological features of acute myeloid leukaemia occurring as second malignancy: GIMEMA archive of adult acute leukaemia. *Br J Haematol*. 2001;112(1):109-117.
18. Mauritzson N, Albin M, Rylander L, et al. Pooled analysis of clinical and cytogenetic features in treatment-related and de novo adult acute myeloid leukemia and myelodysplastic syndromes based on a consecutive series of 761 patients analyzed 1976-1993 and on 5098 unselected cases reported in the literature 1974-2001. *Leukemia*. 2002;16(12):2366-2378.
19. Schoch C, Schnittger S, Klaus M, Kern W, Hiddemann W, Haferlach T. AML with 11q23/MLL abnormalities as defined by the WHO classification: incidence, partner chromosomes, FAB subtype, age distribution, and prognostic impact in an unselected series of 1897 cytogenetically analyzed AML cases. *Blood*. 2003;102(7):2395-2402.
20. Schoch C, Kern W, Schnittger S, Hiddemann W, Haferlach T. Karyotype is an independent prognostic parameter in therapy-related acute myeloid leukemia (t-AML): an analysis of 93 patients with t-AML in comparison to 1091 patients with de novo AML. *Leukemia*. 2004;18(1):120-125.
21. Wheatley K, Brookes CL, Howman AJ, et al; United Kingdom National Cancer Research Institute Haematological Oncology Clinical Studies Group and Acute Myeloid Leukaemia Subgroup. Prognostic factor analysis of the survival of elderly patients with AML in the MRC AML11 and LRF AML14 trials. *Br J Haematol*. 2009;145(5):598-605.
22. Juliusson G, Antunovic P, Derolf A, et al. Age and acute myeloid leukemia: real world data on decision to treat and outcomes from the Swedish Acute Leukemia Registry. *Blood*. 2009;113(18):4179-4187.
23. Preiss BS, Bergmann OJ, Friis LS, et al; AML Study Group of Southern Denmark. Cytogenetic findings in adult secondary acute myeloid leukemia (AML): frequency of favorable and adverse chromosomal aberrations do not differ from adult de novo AML. *Cancer Genet Cytogenet*. 2010;202(2):108-122.
24. Ostgård LSG, Kjeldsen E, Holm MS, et al. Reasons for treating secondary AML as de novo AML. *Eur J Haematol*. 2010;85(3):217-226.
25. Kayser S, Döhner K, Krauter J, et al; German-Austrian AMLSG. The impact of therapy-related acute myeloid leukemia (AML) on outcome in 2853 adult patients with newly diagnosed AML. *Blood*. 2011;117(7):2137-2145.
26. Oran B, Weisdorf DJ. Survival for older patients with acute myeloid leukemia: a population-based study. *Haematologica*. 2012;97(12):1916-1924.
27. Gangatharan SA, Grove CS, P'ng S, et al. Acute myeloid leukaemia in Western Australia 1991-2005: a retrospective population-based study of 898 patients regarding epidemiology, cytogenetics, treatment and outcome. *Intern Med J*. 2013;43(8):903-911.
28. Medeiros BC, Satram-Hoang S, Hurst D, Hoang KQ, Momin F, Reyes C. Big data analysis of treatment patterns and outcomes among elderly acute myeloid leukemia patients in the United States. *Ann Hematol*. 2015;94(7):1127-1138.
29. Nagel G, Weber D, Fromm E, et al; German-Austrian AML Study Group (AMLSG). Epidemiological, genetic, and clinical characterization by age of newly diagnosed acute myeloid leukemia based on an academic population-based registry study (AMLSG BiO). *Ann Hematol*. 2017;96(12):1993-2003.
30. Bertoli S, Tavitian S, Huynh A, et al. Improved outcome for AML patients over the years 2000-2014. *Blood Cancer J*. 2017;7(12):635.
31. Döhner H, Estey E, Grimwade D, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood*. 2017;129(4):424-447.

32. Cervantes F, Tassies D, Salgado C, Rovira M, Pereira A, Rozman C. Acute transformation in nonleukemic chronic myeloproliferative disorders: actuarial probability and main characteristics in a series of 218 patients. *Acta Haematol.* 1991;85(3):124-127.
33. Godley LA, Larson RA. Therapy-related myeloid leukemia. *Semin Oncol.* 2008;35(4):418-429.
34. Borthakur G, Estey AE. Therapy-related acute myelogenous leukemia and myelodysplastic syndrome. *Curr Oncol Rep.* 2007;9(5):373-377.
35. Appelbaum FR, Gundacker H, Head DR, et al. Age and acute myeloid leukemia. *Blood.* 2006;107(9):3481-3485.
36. Campbell PJ, Baxter EJ, Beer PA, et al. Mutation of JAK2 in the myeloproliferative disorders: timing, clonality studies, cytogenetic associations, and role in leukemic transformation. *Blood.* 2006;108(10):3548-3555.
37. Ok CY, Patel KP, Garcia-Manero G, et al. Mutational profiling of therapy-related myelodysplastic syndromes and acute myeloid leukemia by next generation sequencing, a comparison with de novo diseases. *Leuk Res.* 2015;39(3):348-354.
38. Fröhling S, Schlenk RF, Breitnick J, et al; AML Study Group Ulm. Prognostic significance of activating FLT3 mutations in younger adults (16 to 60 years) with acute myeloid leukemia and normal cytogenetics: a study of the AML Study Group Ulm. *Blood.* 2002;100(13):4372-4380.
39. Stone RM, Manley PW, Larson RA, Capdeville R. Midostaurin: its odyssey from discovery to approval for treating acute myeloid leukemia and advanced systemic mastocytosis. *Blood Adv.* 2018;2(4):444-453.
40. Mengis C, Aebi S, Tobler A, Dähler W, Fey MF. Assessment of differences in patient populations selected for excluded from participation in clinical phase III acute myelogenous leukemia trials. *J Clin Oncol.* 2003;21(21):3933-3939.
41. Nilsson C, Hulegårdh E, Garelius H, et al. Secondary acute myeloid leukemia and the role of allogeneic stem cell transplantation in a population-based setting. *Biol Blood Marrow Transplant.* 2019;25(9):1770-1778.
42. Lancet JE, Uy GL, Cortes JE, et al. Cpx-351 (cytarabine and daunorubicin) liposome for injection versus conventional cytarabine plus daunorubicin in older patients with newly diagnosed secondary acute myeloid leukemia. *J Clin Oncol.* 2018;36(26):2684-2692.
43. Thepot S, Itzykson R, Seegers V, et al; Groupe Francophone des Myelodysplasies (GFM). Treatment of progression of Philadelphia-negative myeloproliferative neoplasms to myelodysplastic syndrome or acute myeloid leukemia by azacitidine: a report on 54 cases on the behalf of the Groupe Francophone des Myelodysplasies (GFM). *Blood.* 2010;116(19):3735-3742.
44. Dumas P-Y, Bertoli S, Bérard E, et al. Azacitidine or intensive chemotherapy for older patients with secondary or therapy-related acute myeloid leukemia. *Oncotarget.* 2017;8(45):79126-79136.
45. Vives S, Martínez-Cuadrón D, Bergua Burgues J, et al. A phase 3 trial of azacitidine versus a semi-intensive fludarabine and cytarabine schedule in older patients with untreated acute myeloid leukemia. *Cancer.* 2021;127(12):2003-2014.
46. Fröhling S, Schlenk RF, Kayser S, et al; German-Austrian AML Study Group. Cytogenetics and age are major determinants of outcome in intensively treated acute myeloid leukemia patients older than 60 years: results from AMLSG trial AML HD98-B. *Blood.* 2006;108(10):3280-3288.
47. Mesa RA, Li CY, Ketterling RP, Schroeder GS, Knudson RA, Tefferi A. Leukemic transformation in myelofibrosis with myeloid metaplasia: a single-institution experience with 91 cases. *Blood.* 2005;105(3):973-977.
48. Borthakur G, Lin E, Jain N, et al. Survival is poorer in patients with secondary core-binding factor acute myelogenous leukemia compared with de novo core-binding factor leukemia. *Cancer.* 2009;115(14):3217-3221.
49. Stölzel F, Pfirrmann M, Aulitzky WE, et al; Study Alliance Leukemia. Risk stratification using a new prognostic score for patients with secondary acute myeloid leukemia: results of the prospective AML96 trial. *Leukemia.* 2011;25(3):420-428.
50. Zeichner SB, Arellano ML. Secondary adult acute myeloid leukemia: a review of our evolving understanding of a complex disease process. *Curr Treat Options Oncol.* 2015;16(8):37.
51. Christiansen DH, Andersen MK, Pedersen-Bjergaard J. Mutations with loss of heterozygosity of p53 are common in therapy-related myelodysplasia and acute myeloid leukemia after exposure to alkylating agents and significantly associated with deletion or loss of 5q, a complex karyotype, and a poor prognosis. *J Clin Oncol.* 2001;19(5):1405-1413.
52. Andersen MK, Christiansen DH, Kirchoff M, Pedersen-Bjergaard J. Duplication or amplification of chromosome band 11q23, including the unrearranged MLL gene, is a recurrent abnormality in therapy-related MDS and AML, and is closely related to mutation of the TP53 gene and to previous therapy with alkylating agents. *Genes Chromosomes Cancer.* 2001;31(1):33-41.
53. Nardi V, Winkfield KM, Ok CY, et al. Acute myeloid leukemia and myelodysplastic syndromes after radiation therapy are similar to de novo disease and differ from other therapy-related myeloid neoplasms. *J Clin Oncol.* 2012;30(19):2340-2347.