

Article

Modular Synthesis and Biological Investigation of 5-Hydroxymethyl Dibenzyl Butyrolactones and **Related Lignans**

Samuel J. Davidson¹, Lisa I. Pilkington¹, Nina C. Dempsey-Hibbert², Mohamed El-Mohtadi², Shiying Tang², Thomas Wainwright², Kathryn A. Whitehead² and David Barker^{1,3,*}

- School of Chemical Sciences, University of Auckland, Auckland 1010, New Zealand; sdav134@aucklanduni.ac.nz (S.J.D.); lisa.pilkington@auckland.ac.nz (L.I.P.)
- 2 School of Healthcare Science, Manchester Metropolitan University, Manchester M1 5GD, UK; N.Dempsey-Hibbert@mmu.ac.uk (N.C.D.-H.); MOHAMED.EL-MOHTADI@stu.mmu.ac.uk (M.E.-M.); SHIYING.TANG@stu.mmu.ac.uk (S.T.); thomas.wainwright@stu.mmu.ac.uk (T.W.); K.A.Whitehead@mmu.ac.uk (K.A.W.)
- 3 The MacDiarmid Institute for Advanced Materials and Nanotechnology, Wellington 6140, New Zealand
- Correspondence: d.barker@auckland.ac.nz; Tel.: +64-9-373-7599

Received: 12 November 2018; Accepted: 21 November 2018; Published: 22 November 2018



Abstract: Dibenzyl butyrolactone lignans are well known for their excellent biological properties, particularly for their notable anti-proliferative activities. Herein we report a novel, efficient, convergent synthesis of dibenzyl butyrolactone lignans utilizing the acyl-Claisen rearrangement to stereoselectively prepare a key intermediate. The reported synthetic route enables the modification of these lignans to give rise to 5-hydroxymethyl derivatives of these lignans. The biological activities of these analogues were assessed, with derivatives showing an excellent cytotoxic profile which resulted in programmed cell death of Jurkat T-leukemia cells with less than 2% of the incubated cells entering a necrotic cell death pathway.

Keywords: lignans; dibenzyl butyrolactones; anti-proliferative; acyl-Claisen; stereoselective synthesis

1. Introduction

Dibenzyl butyrolactone lignans 1 are a class of lignans which have been reported to exhibit a range of biological activities, including, but not limited to neuroprotective [1], anti-cancer [2,3], anti-inflammatory [2,4], and anti-aging effects (see Figure 1) [5]. Perhaps the most notable of these biological properties is their reported potent anti-proliferative activities; examples of this class include (-)-matairesinol 2 and (-)-arctigenin 3 which, along with their synthesized derivatives, have been shown to exhibit excellent activity against various cancer cell lines, including pancreatic, breast, endometrial, colorectal, lung, and bladder cancers [6–12].

Owing to their anti-cancer properties and their classification as drug-like compounds [13] extensive work has gone into the study of these compounds and their related analogues to explore and establish structure-activity relationships and the possible use of these lignans as lead compounds for therapeutics. Whilst previous work has explored the synthesis of these lignans and analogues thereof [14–16], mainly focusing on changing the substituents on the aryl rings [17], one area that has not been extensively investigated is the synthesis of C-5 substituted analogues of these butyrolactone lignans, represented by 4.



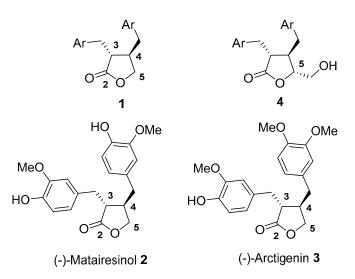


Figure 1. General structures of butyrolactone lignan **1**, natural dibenzylbutyrolactone lignans, (–)-matairesinol **2** and (–)-artigenin **3**, and 5-hydroxymethyl analogues **4**.

We have previously shown that the acyl-Claisen rearrangement can be used to prepare disubstituted morpholine pentenamides **5** with high diastereoselectivity at the C-3 and C-4 positions which correspond to the benzyl groups in the lactone scaffold (Figure 2) [18–22]. Furthermore, in our efforts to a prepare a number of different lignan scaffolds [18–36], we have used amides such as **5** to prepare compounds including tetrahydrofuran lignans (e.g., galbelgin **6**), aryltetralins (e.g., ovafolinin **7**) and aryl dihydronaphthalene lignans (e.g., (–)-pycananthuligene B **8**).

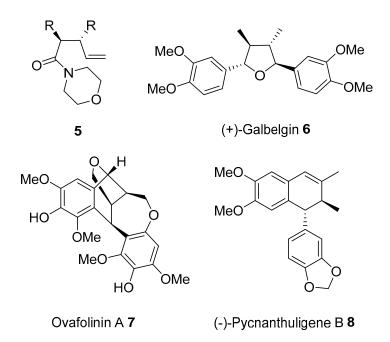
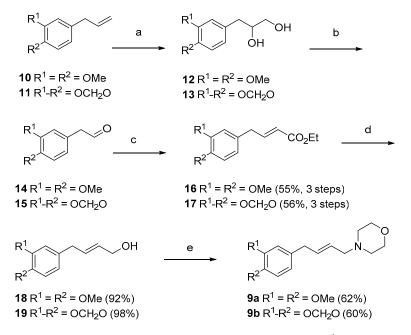


Figure 2. Use of amide **5**, the product of an acyl-Claisen rearrangement to access a number of lignan scaffolds and natural products **6–8**.

We wished to explore the usage of this methodology to synthesise butyrolactone lignans, as well as probe the effect of adding a substituent at the C-5 position on the biological activity. The route would be convergent and modular, allowing for simple modification of aromatic groups resulting in the synthesis of a number of analogues.

2. Results and Discussion

In order to utilise the acyl-Claisen rearrangement to prepare the desired lactones, the corresponding allylic morpholines and acid chlorides first needed to be synthesised. Allylic morpholines **9a** and **9b** were synthesised in five steps from 4-allyl-1,2-dimethoxybenzene **10** and safrole **11** (Scheme 1), respectively. Firstly, allylic benzenes **10** and **11** were dihydroxylated using catalytic osmium tetroxide giving **12** and **13**, followed by periodate cleavage to give aldehydes **14** and **15**. Aldehydes **14** and **15** were immediately used in a Wittig reaction with (carbethoxymethylene)-triphenylphosphorane to exclusively give the *E*-isomer of α , β -unsaturated esters **16** and **17**, in 55% and 56% yields, respectively, over three steps. The esters **16** and **17** were then reduced to allylic alcohols **18** and **19** using di-*iso*-butyl aluminium hydride (DIBAL-H) in excellent yields. Alcohols **18** and **19** were then converted to the corresponding allylic morpholines **9a** and **9b**, by first generating a mesylate in situ, which then underwent substitution to give allylic morpholines **9a** and **9b**.

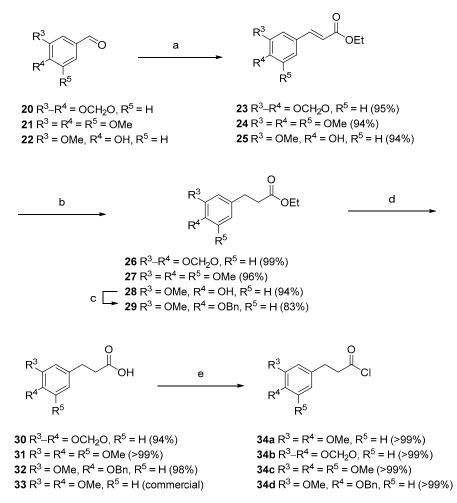


Scheme 1. (a) OsO₄ (0.1−0.3 mol%), *N*-methylmorpholine-*N*-oxide (3 eq.), ^{*t*}BuOH/H₂O (1:1), 4 days; (b) NaIO₄ (1.2 eq.), MeOH/H₂O (3:1), 0.5−2 h; (c) Ph₃PCHCO₂Et (1.1 eq.), CH₂Cl₂, 16 h; (d) 18: DIBAL-H (3 eq.), CH₂Cl₂, −78 °C, 10 min, 19: DIBAL-H (2.2 eq.), toluene, −10 °C, 10 min; (e) Et₃N (3 eq.), MsCl (1.2 eq.), morpholine (1.5 eq.), CH₂Cl₂, 0 °C, 2−18 h.

The required acid chlorides were then synthesised in four or five steps from commercially available benzaldehydes—piperonal **20**, 3,4,5-trimethoxybenzaldehyde **21** and vanillin **22** (Scheme 2). Benzaldehydes **20–22** first underwent a Wittig reaction with (carbethoxymethylene)triphenylphosphorane to give α , β -unsaturated esters **23–25** which were then hydrogenated using Pd on Carbon (10% w/w), giving saturated esters **26–28** in 88–94% yield over two steps. The phenol in **28** was protected as the benzyl ether, **29**, in 83% yield. Esters **26**, **27**, and **29** were hydrolysed using NaOH in methanol/water to the corresponding carboxylic acids **30**, **31**, and **32**, respectively, in 94–99% yields. Finally, chlorination of acids **30–32**, along with commercially available 3,4-dimethoxyphenyl propionic acid **33**, using oxalyl chloride gave acid chlorides **34a–d** in quantitative yields.

Acyl-Claisen rearrangements were undertaken using two allylic morpholines **9a** and **9b** which were reacted individually with the four acid chlorides **34a–d**, using TiCl₄·2THF as the Lewis acid, providing eight morpholine amides **35aa–bd** in 42–95% yields. All amides **35aa–bd** were obtained as single diastereomers with a *syn*-configuration between the C-2 and C-3 substituents (Scheme 3).

All amides **35aa–bd** then underwent dihydroxylation using osmium tetroxide and *N*-methylporpholine *N*-oxide (NMO) to give cyclized 5-hydroxymethyllactones **4aa–bd**.



Scheme 2. (a) Ph₃PCHCO₂Et (1.1 eq.), CH₂Cl₂, 3–20 h; (b) H₂, Pd/C (10% *w/w*), ethyl acetate, 1–2 h; (c) BnBr, K₂CO₃, CH₃CN, 65 h; (d) NaOH (4 eq.), MeOH/H₂O, 2.5 h; (e) (COCl)₂ (2 eq.), CH₂Cl₂, 1.5–4 h.

In all cases it was observed that only the 3,4-*trans*-4,5-*trans*-lactone was obtained. This configuration was confirmed through NOESY NMR analysis, depicted in Figure 3 with **4bb**. We propose that only this isomer was obtained due to the preferential cyclisation of the 3,4-*anti* diol **36**, leaving the polar uncyclised 3,4-*syn* diols **37** which were difficult to isolate. Upon dihydroxylation of amide **35bb** at a larger scale and following isolation of lactone **4bb** by column chromatography, a small sample of the corresponding uncyclised diol **37** was able to be isolated. This diol **37** was subsequently cyclised using 2 M H₂SO₄ in methanol to give the corresponding C-5 epimer, *epi*-**4bb**, confirming this hypothesis (Scheme 4).

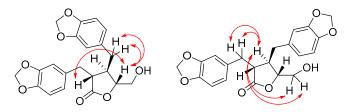
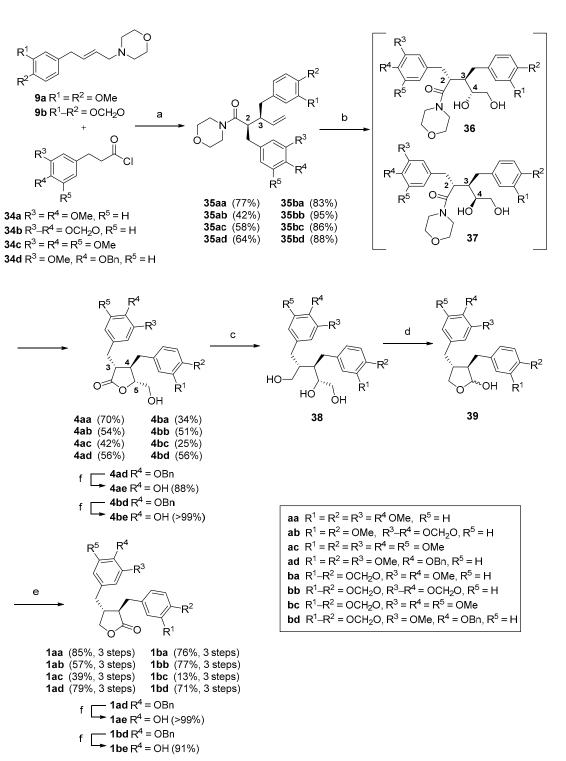


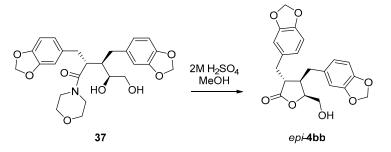
Figure 3. Selected NOESY correlations showing *trans,trans*-relationship of hydroxymethyl lactone lignan analogue **4bb**.



Scheme 3. (a) TiCl₄·2THF (100 mol%), ^{*i*}Pr₂NEt (1.5 eq.), acid chloride (1.2 eq.), CH₂Cl₂, 18–24 h; (b) OsO₄ (8 mol %), NMO (3 eq.), ^{*t*}BuOH/H₂O (1:1), 3–7 days; (c) LiAlH₄ (1.5 eq.), THF, 0.5–2 h; (d) NaIO₄ (1.2 eq.), MeOH/H₂O (3:1), 0.25–1 h; (e) Ag₂CO₃/Celite (2 eq.), toluene, reflux, 2–3 h; (f) H₂, Pd/C (10% *w/w*), MeOH, 10 min.

Finally, to deprotect the benzyl-protected lactones **4ad** and **4bd** to their respective alcohols, they were subjected to hydrogenolysis to give **4ae** and **4be** in excellent yields. Transformation of C-5 hydroxymethyl analogues **4** into dibenzylbutryolactone lignans **1** was achieved via reduction using LiAlH₄, to the corresponding triols **38aa–bd**, followed by periodate cleavage, forming lactols **39aa–bd**. These lactols **39aa–bd** were then oxidised using Fetizon's reagent [37,38] to give racemic samples of

dibenzyl butyrolactone lignans **1aa–bd**, including known natural products arcitin **1aa**, bursehernin **1ab**, (3*R**,4*R**)-3-(3″,4″-dimethoxybenzyl)-4-(3′,4′,5′-trimethoxybenzyl)dihydrofuran-2(3*H*)-one **1ac**, kusunokinin **1ba**, hinokinin **1bb**, and isoyatein **1bc**. Additionally, phenolic lignans, buplerol **1ae**, and haplomyrfolin **1be** were produced by the debenzylation of **1ad** and **1bd**, respectively.



Scheme 4. Synthesis of *epi*-4bb.

Several of the synthesised compounds were then tested for their anti-microbial and cytotoxic activities. All tested compounds were found to be inactive against *Staphlycoccus aureus* and *Escherichia*. coli, showing no to little antimicrobial activity, while the compounds were shown to exhibit antiproliferative effects against Jurkat T-leukaemia cells, while also showing effects on cell cycle progression (Figure 4). While the synthesised naturally-occurring dibenzyl butyrolactones, arcitin 1aa, bursehernin **1ab**, and (3*R**,4*R**)-3-(3",4"-dimethoxybenzyl)-4-(3',4',5'-trimethoxybenzyl)dihydrofuran-2(3H)-one **1ac**, boasted the best activities, 5-hydroxymethyl analogue **4bb** had similar potency. Compound 4bb was shown to have the best activity of all of the 5-hydroxymethyl analogues tested, inducing apoptosis, evidenced by the presence of cells in the early and predominantly in the late apoptotic cell cycle (Figure 4). Additionally the compounds demonstrated an effect on cell cycle progression. A significantly greater number of 4N cells were present following treatment with compound **4bb** in particular causing a significant increase in 4N cells (Figure 4D,E). During the cell cycle, DNA is replicated in the S-phase, going from 2N in G_1 , to 4N by the end of this phase. The DNA content in cells then remains at 4N during G2 and M phases, before cytokinesis at the M-phase. The observation that there was in increase in 4N cells indicates that it is likely these cells have arrested in G_2/M and will not re-enter next G_1 -phase after this mitotic slippage. This is in-line with published cell cycle data following treatment with other lignans [39,40]. Furthermore, our compounds showed minimal levels of necrosis, less than 2% (except 4ba with 7%), suggesting that the cells are in fact entering programmed cell death cycles, which is considered the most effective and non-inflammatory mechanism of cancer-cell death.

In conclusion, the synthesis of dibenzyl butyrolactone lignans utilising the acyl-Claisen rearrangement has been accomplished and represent a new, modular, and convergent method towards the synthesis of this class of natural products. Furthermore, this route gives rise to the previously-unexplored 5-hydroxymethyl derivatives **4** of these natural products. The biological activities of this new set of derivatives were assessed, with one derivative in particular, **4bb**, showing a superior cytotoxic profile and resulting in cell cycle arrest and programmed cell death of Jurkat T-leukaemia cells with less than 2% of the incubated cells entering a necrotic cell death pathway.

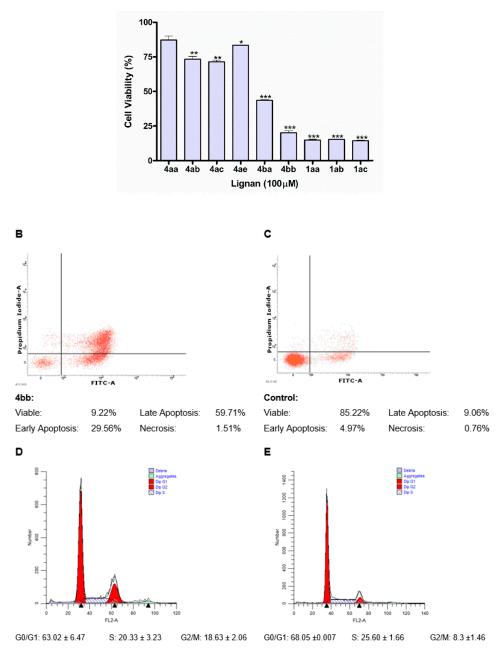


Figure 4. (**A**) Cell survival (by a measure of metabolic activity) of Jurkat T-cell leukaemia cells incubated with 100 μ M of lignans and lignan analogues for 48 h. The data represents means of triplicate experiments and is shown as means \pm SEM (*n* = 3). The positive control (not shown) had a growth of 100%. Significance of the compound activity compared to the control is expressed: (*) *p*-value <0.05; (**) *p*-value <0.01; (***) *p*-value <0.001. (**B**) Dotplot showing the viability of Jurkat T-leukaemia cells after incubation with 100 μ M **4bb** for 24 h followed by labelling with annexin V/propidium iodide and analysis using flow cytometry. Cells in the bottom-left quadrant represent viable cells, bottom-right quadrant are positive for annexin V and are in early apoptosis, top-right quadrant are double positive for propidium iodide and are undergoing necrosis. (**C**) Negative control showing the viability of unsynchronized cells incubated in the presence of 100 μ M **4bb** or **E**: vehicle for 24 h. DNA content of the cells was determined by flow cytometry. Percentage of cells in each stage of the cell cycle (average of three replicates \pm SD is reported).

3. Experimental Section

3.1. General Methods

All reactions were carried out with oven-dried glassware and under a nitrogen atmosphere in dry, freshly distilled solvents unless otherwise noted. Diisopropylethylamine was distilled from CaH₂ and stored over activated 4Å molecular sieves. All melting points for solid compounds, given in degrees Celsius (°C), were measured using a Reicher–Kofler block and are uncorrected. Infrared (IR) spectra were recorded using a Perkin Elmer Spectrum1000 FT-IR spectrometer. The NMR spectra were recorded on a 400 MHz spectrometer. Chemical shifts are reported relative to the solvent peak of chloroform (δ 7.26 for ¹H and δ 77.16 ± 0.06 for ¹³C). The ¹H-NMR data was reported as position (δ), relative integral, multiplicity (s, singlet; d, doublet; dd, doublet of doublets; dd, doublet of doublets; dt, doublet of triplets; dq, doublet of quartets; t, triplet; td, triplet of doublets; q, quartet; m, multiplet), coupling constant (*J*, Hz), and the assignment of the atom. The ¹³C-NMR data were reported as position (δ) and assignment of the atom. The NMR assignments were performed using COSY, HSQC and HMBC experiments. High-resolution mass spectroscopy (HRMS) was carried out by electrospray ionization (ESI) on a MicroTOF-Q mass spectrometer. Fetizon's reagent was prepared following a literature procedure [41]. Unless noted, chemical reagents were used as purchased.

3.2. Synthetic Methods

3.2.1. General Procedure A: Acyl-Claisen

To a stirred suspension of TiCl₄·2THF (1 mmol) in CH₂Cl₂ (5 mL), under an atmosphere of nitrogen, was added a solution of allylic morpholine (1 mmol) in CH₂Cl₂ (2.5 mL) followed by dropwise addition of ^{*i*}Pr₂NEt (1.5 mmol). After stirring for 10 min a solution of acid chloride (1.2 mmol) in CH₂Cl₂ (2.5 mL) was added dropwise and the resultant mixture stirred for the specified time. The reaction mixture was quenched with aqueous NaOH (12 mL, 1 M) and the aqueous phase extracted with CH₂Cl₂ (3 × 10 mL). The combined organic extracts were washed with brine (6 mL), dried (MgSO₄), the solvent removed in vacuo and the crude product purified by column chromatography.

3.2.2. General Procedure B: Dihydroxylation

To a stirred solution of morpholine pentenamide (1 mmol) in ${}^{t}BuOH/H_2O$ (1:1, 20 mL) or ${}^{t}BuOH/H_2O/THF$ (1:1:1, 30 mL) was added NMO (3 mmol). A solution of OsO₄ (0.08 mmol, 2.5% w/v in ${}^{t}BuOH$) was then added dropwise and the resultant mixture stirred for the specified time. The mixture was quenched with saturated aqueous Na₂SO₃ (30 mL) and stirred for a further 1 h. The aqueous phase was extracted with ethyl acetate (3 × 20 mL), the combined organic extracts washed with aqueous KOH (5 mL, 1 M), dried (MgSO₄), the solvent removed in vacuo and the crude product purified by column chromatography.

3.2.3. General Procedure C: Lithium Aluminum Hydride Reduction

To a stirred suspension of LiAlH₄ (1.4 mmol) in THF (10 mL), under an atmosphere of nitrogen at 0 °C, was added a solution of lactone (1 mmol) in THF (10 mL) and the mixture stirred for the specified time. After warming to room temperature, the mixture was quenched with the addition of water (30 mL) and the aqueous phase extracted with ethyl acetate (3 × 40 mL). The combined organic extracts were washed with brine (25 mL), dried (MgSO₄), and the solvent removed in vacuo.

3.2.4. General Procedure D: Periodate Cleavage

To a stirred solution of triol (1 mmol) in MeOH/H₂O (3:1, 50 mL) was added NaIO₄ (1.2 mmol) and the resultant mixture stirred for the specified time. The reaction mixture was quenched with brine (40 mL) and extracted with ethyl acetate (3×80 mL). The organic layers were combined, washed with

water (2 \times 40 mL), dried (MgSO₄), and solvent removed in vacuo to give the crude product which was purified by column chromatography if necessary.

3.2.5. General Procedure E: Fétizon's Oxidation

To a stirred solution of lactol (1 mmol) in toluene (60 mL), under an atmosphere of nitrogen, was added Fétizon's reagent (2 mmol) and heated at reflux for the specified time. The reaction mixture was allowed to cool and filtered, the solvent removed in vacuo and the crude product purified by column chromatography.

3.2.6. General Procedure F: Benzyl Deprotection

To a stirred solution of benzyl ether (1 mmol) in MeOH (30 mL) was added 10% palladium on carbon (20% w/w) and the resultant mixture stirred under and atmosphere of hydrogen for the specified time. The reaction mixture was filtered through celite, washed with methanol (3×20 mL), the solvent removed in vacuo and the crude product purified by column chromatography if necessary (The ¹H and ¹³C-NMR spectra of compounds in the Supplemental Materials).

(E)-Ethyl 4-(3',4'-dimethoxyphenyl)but-2-enoate (16). To a stirred solution of NMO (7.9 g, 67.3 mmol) in H₂O/^tBuOH (1:1, 80 mL) was added 4-allyl-1,2-dimethoxybenzene 10 (3.86 mL, 22.4 mmol). A solution of OsO₄ (0.6 mL, 0.059 mmol, 2.5% w/v in ^tBuOH) was then added dropwise and the resulting mixture stirred at room temperature for 4 days. The mixture was then quenched with saturated aqueous Na_2SO_3 (100 mL) and stirred for 1 h. The mixture was extracted with ethyl acetate (3 \times 50 mL), the organic layers combined, washed with aqueous KOH (1 M, 20 mL), and dried (MgSO₄). Solvent was removed in vacuo to give 12 (4.8 g, quant.) as a white solid which was used without further purification. To a stirred solution of diol 12 (4.8 g, 22.8 mmol) in methanol/H₂O (3:1, 100 mL) was added NaIO₄ (5.9 g, 27.4 mmol) and stirred for 30 min. The reaction mixture was then quenched with addition of brine (50 mL) and extracted with ethyl acetate (3 \times 40 mL). The organic extracts were combined, washed with water (2×20 mL), and dried (MgSO₄). Solvent was removed in vacuo to give 14 (2.68 g, 65%) as a pale-yellow oil which was used without further purification. To a stirred solution of 2-(3,4-dimethoxyphenyl)acetaldehyde 14 (2.68 g, 14.8 mmol) in CH₂Cl₂ (100 mL), under an atmosphere of nitrogen, was added (carbethoxymethylene)triphenylphosphorane (5.7 g, 16.3 mmol) and the resulting mixture stirred for 16 h. Solvent was removed in vacuo and the crude product purified by column chromatography (3:1, hexanes, ethyl acetate) to give the title compound 16 (3.13 g, 84%) as a colourless oil. $R_f = 0.56$ (2:1 hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 1.27 (3H, t, *J* = 7.2 Hz, 1-OCH₂CH₃), 3.45 (2H, dd, *J* = 1.5, 6.7 Hz, 4-H), 3.86 (6H, s, 3', 4'-H), 4.17 (2H, q, *J* = 7.2 Hz, 1-OCH₂CH₃), 5.80 (1H, td, *J* = 1.6, 15.5 Hz, 2-H), 6.67 (1H, d, *J* = 1.9 Hz, 2'-H), 6.71 (1H, dd, *J* = 1.9, 8.1 Hz, 6'-H), 6.81 (1H, d, J = 8.1 Hz, 5'-H), 7.07 (1H, td, J = 6.7, 15.5 Hz, 3-H). δ_{C} (100 MHz; CDCl₃) 14.3 (1-OCH₂CH₃), 38.1 (C-4), 55.9, 56.0 (3', 4'-OCH₃), 60.3 (1-OCH₂CH₃), 111.5 (C-5'), 112.1 (C-2'), 120.8 (C-6'), 122.2 (C-2), 130.2 (C-1'), 147.6 (C-3), 147.9 (C-4'), 149.1 (C-3'), 166.6 (C-1). Values are in agreement with literature data [42].

(E)-4-(3',4'-Dimethoxyphenyl)but-2-en-1-ol (**18**). To a stirred solution of ester **16** (1.0 g, 4.0 mmol) in CH₂Cl₂ (20 mL), under an atmosphere of nitrogen at -78 °C, was added DIBAL (12 mL, 1 M in cyclohexane) and the resulting mixture stirred for 10 min. The reaction mixture was quenched with addition of 2 M HCl until gas evolution ceased, the organic phase separated and the aqueous phase further extracted with CH₂Cl₂ (3 × 10 mL). The organic layers were combined then washed with water (10 mL) and dried (MgSO₄). Solvent was removed in vacuo and the crude product purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound **18** (0.76 g, 92%) as a colourless oil. R_f = 0.18 (2:1, hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 3.30 (2H, d, *J* = 6.6 Hz, 4-H), 3.82 (3H, s, 4'-OCH₃), 3.83 (3H, s, 3'-OCH₃), 4.08 (2H, d, *J* = 5.6 Hz, 1-H), 5.64–5.69 (1H, m, 2-H), 5.78–5.83 (1H, m, 3-H), 6.68 (1H, s, 2'-H), 6.69 (1H, d, *J* = 8.0 Hz, 6'-H), 6.77 (1H, d, *J* = 8.0 Hz, 5'-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 38.2 (C-4), 55.8 and 55.9 (3' and 4'-OCH₃), 63.3 (C-1), 111.4 (C-5'), 112.0 (C-2'), 120.4 (C-6'), 130.2 (C-2),

131.6 (C-3), 132.7 (C-1'), 147.4 (C-4'), 148.9 (C-3'). IR: ν_{MAX} (film)/cm⁻¹; 3391 (broad), 2933, 2835, 1591, 1512, 1463, 1417, 1258, 1232, 1137, 1025, 971, 852, 806, 762. HRMS (ESI⁺) Found [M + Na]⁺ 231.0995; C₁₂H₁₆NaO₃ requires 231.0992.

(*E*)-4-(4-(3',4'-Dimethoxyphenyl)but-2-en-1-yl)morpholine (**9a**). To a stirred solution of alcohol **18** (0.73 g, 3.5 mmol) in CH₂Cl₂ (20 mL), under an atmosphere of nitrogen at 0 °C, was added Et₃N (1.5 mL, 10.5 mmol) and stirred for 5 min. MsCl (0.48 mL, 4.2 mmol) was added and stirred for 10 min. Morpholine (0.50 mL, 5.3 mmol) was added and the mixture brought to room temperature and stirred for 2 h. Saturated aqueous NaHCO₃ (20 mL) and water (4 mL) was then added and the aqueous layer further extracted with CH₂Cl₂ (3 × 20 mL). The organic layers were then combined, dried (MgSO₄) and the solvent removed in vacuo. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound 9a (0.60 g, 62%) as a colourless oil. R_f = 0.31 (1:2 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.41–2.44 (4H, m, O(CH₂CH₂)₂N), 2.96 (2H, d, *J* = 6.8 Hz, 1-H), 3.30 (2H, d, *J* = 6.7 Hz, 4-H), 3.68–3.71 (4H, m, O(CH₂CH₂)₂N), 3.83 (6H, s, 3', 4'-OCH₃), 5.52–5.57 (1H, m, 3-H), 5.71–5.78 (1H, m, 2-H), 6.67–6.70 (2H, m, 2' and 6'-H), 6.78 (1H, d, *J* = 7.9 Hz, 5'-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 38.5 (C-4), 53.6 (O(CH₂CH₂)₂N), 55.8, 56.0 (3', 4'-OCH₃), 61.1 (C-1), 67.0 (O(CH₂CH₂)₂N), 111.4 (C-5'), 111.9 (C-2'), 120.3 (C-6'), 127.1 (C-3), 132.8 (C-1'), 133.8 (C-2), 147.5 (C-4'), 149.0 (C-3'). IR: $\nu_{\rm MAX}$ (film)/cm⁻¹; 2934, 2851, 1591, 1453, 1260, 1138, 1028, 976, 864, 805, 763. HRMS (ESI⁺) Found [M + H]⁺ 278.1762; C₁₆H₂₄NO₃ requires 278.1751.

(E)-Ethyl 4-(3',4'-methylenedioxyphenyl)but-2-enoate (17). To a stirred solution of NMO (8.67 g, 74.0 mmol) in $H_2O/^tBuOH$ (1:1, 80 mL) was added safrole 11 (4.0 mL, 27 mmol). A solution of OsO₄ (0.75 mL, 0.074 mmol, 2.5% w/v in ^tBuOH) was added dropwise and the resultant mixture stirred at room temperature for 17 h. The reaction mixture was quenched with saturated aqueous Na₂SO₃ (100 mL) and stirred for 1 h. The mixture was extracted with ethyl acetate (3×50 mL), the organic layers were combined, washed with aqueous KOH (1 M, 20 mL) and dried (MgSO₄). Solvent was removed in vacuo to give diol 13 (5.2 g, quant.) as a white solid which was used without further purification. To a stirred solution of diol 13 (5.2 g, 27 mmol) in methanol/H₂O (3:1, 100 mL) was added NaIO₄ (6.8 g, 32 mmol) and stirred for 2 h. The mixture was then quenched with addition of brine (50 mL) and extracted with ethyl acetate (3 \times 50 mL). The organic extracts were combined, washed with water $(2 \times 20 \text{ mL})$, brine (10 mL), and dried (MgSO₄). Solvent was removed in vacuo to give aldehyde 15 (4.4 g, quant.) as a yellow oil which was used without further purification. To a stirred solution of 2-(3,4-methylenedioxyphenyl)acetaldehyde 15 (4.4 g, 27 mmol) in CH₂Cl₂ (50 mL), under an atmosphere of nitrogen, was added (carbethoxymethylene)triphenylphosphorane (10.4 g, 30 mmol) and the resulting mixture stirred for 16 h. Solvent was removed in vacuo and the crude product purified by column chromatography (19:1, hexanes, ethyl acetate) to give the title compound 17 (3.54 g, 56%) as a colourless oil. $R_f = 0.73$ (2:1 hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 1.27 (3H, t, *J* = 7.2 Hz, 1-OCH₂CH₃), 3.42 (2H, dd, *J* = 6.6, 1.6 Hz, 4-H), 4.17 (2H, q, *J* = 7.2 Hz, 1-OCH₂CH₃), 5.78 (1H, dt, J = 15.5, 1.6 Hz, 2-H), 5.93 (2H, s, OCH₂O), 6.61 (1H, dd, J = 8.0, 2.0 Hz, 6'-H), 6.64 (1H, d, J = 2.0 Hz, 2'-H), 6.74 (1H, d, J = 8.0 Hz, 5'-H), 7.04 (1H, dt, J = 15.5, 6.6 Hz, 3-H). δ_C (100 MHz; CDCl₃) 14.4 (1-OCH₂CH₃), 38.2 (C-4), 60.4 (1-OCH₂CH₃), 101.1 (OCH₂O), 108.5 (C-5'), 109.4 (C-2'), 121.9 (C-6'), 122.4 (C-2), 131.5 (C-1'), 146.5 (C-4'), 147.5 (C-3), 148.0 (C-3'), 166.6 (C-1). Values are in agreement with literature data [43].

(*E*)-4-(3',4'-*Methylenedioxyphenyl)but*-2-*en*-1-*ol* (**19**). To a stirred solution of ester **17** (3.2 g, 13.7 mmol) in toluene (100 mL), under an atmosphere of nitrogen at -10 °C, was added DIBAL (30 mL, 1 M in toluene) and the resultant mixture stirred for 10 min. The reaction mixture was quenched with addition of 2 M HCl until gas evolution ceased, the organic layer was separated and the aqueous phase further extracted with CH₂Cl₂ (3 × 50 mL). The organic layers were combined, washed with brine (30 mL) and dried (MgSO₄). Solvent was removed in vacuo and the crude product purified by column chromatography (3:1 hexanes, ethyl acetate) to give the title compound **19** (2.59 g, 98%) as a pale yellow oil. R_f = 0.42 (hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.41 (1H, br s, 1-OH), 3.30 (2H, d, *J* =

6.6 Hz, 4-H), 4.12 (2H, br d, *J* = 4.5 Hz, 1-H), 5.64–5.72 (1H, m, 2-H), 5.77–5.85 (1H, m, 3-H), 5.92 (2H, s, OCH₂O), 6.63 (1H, dd, *J* = 7.9, 1.9 Hz, 6'-H), 6.67 (1H, d, *J* = 1.9 Hz, 2'-H), 6.73 (1H, d, 7.9 Hz, 5'-H). δ_{C} (100 MHz; CDCl₃) 38.4 (C-4), 63.6 (C-1), 101.0 (OCH₂O), 108.3 (C-5'), 109.2 (C-2'), 121.4 (C-6'), 130.4, 131.8 (C-2, 3), 133.9 (C-1'), 146.0, 147.8 (C-3', 4'). Values are in agreement with literature data [43].

(*E*)-4-(4-(3',4'-*Methylenedioxyphenyl*)*but*-2-*en*-1-*yl*)*morpholine* (**9b**). To a stirred solution of alcohol **19** (1.66 g, 8.6 mmol) in CH₂Cl₂ (15 mL), under an atmosphere of nitrogen at 0 °C, was added Et₃N (3.6 mL, 25.9 mmol) and stirred for 5 min. MsCl (1.2 mL, 10.4 mmol) was added and stirred for 10 min. Morpholine (1.3 mL, 13.8 mmol) was added and the mixture brought to room temperature and stirred for 18 h. Saturated aqueous NaHCO₃ (25 mL) and water (5 mL) was added and the aqueous layer further extracted with CH₂Cl₂ (3 × 30 mL). The organic layers were combined, dried (MgSO₄) and the solvent removed in vacuo. The crude product was purified by column chromatography (2:1 hexanes, ethyl acetate) to give the title compound **9b** (1.4 g, 60%) as a pale yellow oil. R_f = 0.39 (1:2 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.43 (4H, br t, *J* = 4.7 Hz, NCH₂CH₂O), 2.96 (2H, d, *J* = 6.5 Hz, 1-H), 3.28 (2H, d, *J* = 7.0 Hz, 4-H), 3.71 (4H, t, *J* = 4.7 Hz, NCH₂CH₂O), 5.49–5.56 (1H, m, 2-H), 5.69–5.76 (1H, m, 3-H), 5.91 (2H, d, *J* = 2.0 Hz, OCH₂O), 6.61 (1H, dd, *J* = 7.5, 2.0 Hz, 6'-H), 6.65 (1H, d, *J* = 2.0 Hz, 2'-H), 6.72 (1H, d, *J* = 7.5 Hz, 5'-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 38.7 (C-4), 53.7 (NCH₂CH₂O), 61.2 (C-1), 67.1 (NCH₂CH₂O), 100.9 (OCH₂O), 108.3 (C-5'), 109.1 (C-2'), 121.4 (C-6'), 127.4 (C-2), 133.7 (C-3), 134.1 (C-1'), 146.0 (C-4'), 147.8 (C-3'). IR: $\nu_{\rm MAX}$ (film)/cm⁻¹; 2855, 1739, 1488, 1242, 1115, 1036, 926, 864, 736. HRMS (ESI⁺) Found [M + H]⁺ 262.1428; C₁₅H₂₀NO₃ requires 262.1438.

(*E*)-*Ethyl*-3-(3',4'-*methylenedioxyphenyl*)*prop*-2-*enoate* (23). To a stirred solution of piperonal **20** (5.0 g, 33 mmol) in CH₂Cl₂ (100 mL), under an atmosphere of nitrogen, was added (carbethoxymethylene)triphenylphosphorane (12.8 g, 37.0 mmol) and the resulting mixture stirred for 20 h. Solvent was then removed in vacuo and the crude product purified by column chromatography (3:1, hexanes, ethyl acetate) to give the title compound **23** (6.97 g, 95%) as a white solid. R_f = 0.68 (2:1 hexanes, ethyl acetate). Melting point: 62–64 °C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.32 (3H, t, *J* = 7.2 Hz, 1-OCH₂CH₃), 4.25 (2H, q, *J* = 7.2 Hz, 1-OCH₂CH₃), 6.00 (2H, s, -OCH₂O-), 6.25 (1H, d, *J* = 15.9 Hz, 2-H), 6.80 (1H, d, *J* = 8.0 Hz, 5'-H), 7.00 (1H, dd, *J* = 1.4, 8.0 Hz, 6'-H), 7.02 (1H, d, *J* = 1.4 Hz, 6'-H), 7.58 (1H, d, *J* = 15.9 Hz, 3-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 14.5 (1-OCH₂CH₃), 60.5 (1-OCH₂CH₃), 101.7 (-OCH₂O-), 106.6 (C-5'), 108.7 (C-2'), 116.4 (C-2), 124.5 (C-6'), 129.1 (C-1'), 144.4 (C-3), 148.5 (C-4'), 149.7 (C-3'), 167.3 (C-1). Values are in agreement with literature data [44].

(*E*)-*Ethyl*-3-(3',4',5'-*trimethoxyphenyl*)*prop*-2-*enoate* (24). To a stirred solution of 3,4,5trimethoxybenzaldehyde 21 (3.0 g, 15.3 mmol) in CH₂Cl₂ (100 mL), under an atmosphere of nitrogen, was added (carbethoxymethylene)triphenylphosphorane (5.9 g, 16.8 mmol) and the resulting mixture stirred for 3 h. Solvent was then removed in vacuo and the crude product purified by column chromatography (3:1, hexanes, ethyl acetate) to give the title compound 24 (4.0 g, 94%) as a white solid. R_f = 0.52 (2:1 hexanes, ethyl acetate). Melting point: 64–66 °C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.34 (3H, t, *J* = 7.2 Hz, 1-OCH₂CH₃), 3.87 (3H, s, 4'-OCH₃), 3.88 (6H, s, 3'-OCH₃), 4.26 (2H, q, *J* = 7.2 Hz, 1-OCH₂CH₃), 6.34 (1H, d, *J* = 15.9 Hz, 2-H), 6.75 (2H, s, 2'-H), 7.60 (1H, d, *J* = 15.9 Hz, 3-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 14.5 (1-OCH₂CH₃), 56.3 (3'-OCH₃), 60.6 (1-OCH₂CH₃), 61.1 (4'-OCH₃), 105.3 (C-2'), 117.7 (C-2), 130.1 (C-1'), 140.2 (C-4'), 144.7 (C-3), 153.6 (C-3'), 167.1 (C-1). Values are in agreement with literature data [45].

3-(3',4',5'-Trimethoxyphenyl) propionic acid (31). To a stirred solution of 24 (5.4 g, 19.4 mmol) in ethyl acetate (30 mL) was added 10% palladium on activated carbon (0.54 g, 10% w/w). The solution was flushed with an atmosphere of hydrogen and stirred for 2 h. The reaction mixture was then filtered through a plug of celite and washed with ethyl acetate, solvent was then removed in vacuo to give saturated ester 27 (5.23 g, 96%) which was then used without further purification.

To a stirred solution of ester 27 (5.1 g, 17.9 mmol) in methanol (30 mL) was added aqueous NaOH (72 mL, 1 M, 4 eq.) and stirred for 20 min. The mixture was then extracted with CH_2Cl_2 (10 mL) and

the aqueous layer acidified with aqueous 2 M HCl. The aqueous phase was then extracted with ethyl acetate (3 × 50 mL), dried (MgSO₄) and solvent removed in vacuo to give the title compound **31** (4.6 g, quant.) as a white solid. $R_f = 0.15$ (2:1 hexanes, ethyl acetate). Melting point: 104–105 °C. δ_H (400 MHz; CDCl₃) 2.68 (2H, t, *J* = 7.8 Hz, 2-H), 2.90 (2H, t, *J* = 7.8 Hz, 3-H), 3.82 (3H, s, 4'-OCH₃), 3.84 (6H, s, 3'-OCH₃), 6.43 (2H, s, 2'-H). δ_C (100 MHz; CDCl₃) 31.1 (C-2), 35.8 (C-3), 56.2 (3'-OCH₃), 61.0 (4'-OCH₃), 105.4 (C-2'), 136.0 (C-1'), 136.7 (C-4'), 153.4 (C-3'), 178.8 (C-1). Values are in agreement with literature data [46].

3-(3',4'-Methylenedioxyphenyl) propionic acid (**30**). To a stirred solution of **23** (6.92 g, 31.4 mmol) in ethyl acetate (30 mL) was added 10% palladium on activated carbon (0.69 g, 10% *w/w*). The solution was flushed with an atmosphere of hydrogen and stirred for 1 h. The reaction mixture was then filtered through a plug of celite and washed with ethyl acetate, solvent was then removed in vacuo to give saturated ester **26** (6.9 g, 99%) which was then used without further purification.

To a stirred solution of ester **26** (6.74 g, 30.0 mmol) in methanol (30 mL) was added aqueous NaOH (121 mL, 1 M, 4 eq.) and stirred for 2.5 h. The mixture was then extracted with ethyl acetate (10 mL) and the aqueous layer acidified with aqueous 2 M HCl. The aqueous phase was then extracted with ethyl acetate (3×50 mL), dried (MgSO₄) and solvent removed in vacuo to give the title compound **30** (5.5 g, 94%) as a white solid. R_f = 0.44 (2:1 hexanes, ethyl acetate). Melting point: 80–82°C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.64 (2H, t, *J* = 7.7 Hz, 2-H), 2.88 (2H, t, *J* = 7.7 Hz, 3-H), 5.93 (2H, s, -OCH₂O-), 6.66 (1H, dd, *J* = 7.9, 1.4 Hz, 6'-H), 6.70 (1H, d, *J* = 1.4 Hz, 2'-H), 6.74 (1H, d, *J* = 7.9 Hz, 5'-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 30.5 (C-2), 36.1 (C-3), 101.0 (-OCH₂O-), 108.4 (C-2'), 108.9 (C-5'), 121.2 (C-6'), 134.1 (C-1'), 146.2 (C-3'), 147.8 (C-4'), 179.1 (C-1). Values are in agreement with literature data [47].

3-(3'-Methoxy-4'-benzyloxyphenyl)propionic acid (32). To a stirred solution of vanillin 22 (3.0 g, 19.7 mmol) in CH₂Cl₂ (100 mL), under an atmosphere of nitrogen, was added (carbethoxymethylene)triphenylphosphorane (7.56 g, 21.7 mmol) and the resulting mixture stirred for 18 h. Solvent was then removed in vacuo and the crude product purified by column chromatography (2:1, hexanes, ethyl acetate) to give a 2:1 mixture of E and Z isomers of unsaturated ester 25 (4.13 g, 94%) as a yellow oil which was used immediately.

To a stirred solution of unsaturated ester 25 (4.13 g, 18.6 mmol) in ethyl acetate (30 mL) was added 10% palladium on activated carbon (0.4 g, 10% w/w). The solution was flushed with an atmosphere of hydrogen and stirred for 2 h. The reaction mixture was then filtered through a plug of celite and washed with ethyl acetate, solvent was then removed in vacuo to give saturated ester 28 (3.9 g, 94%) as a yellow oil which was then used without further purification. To a stirred solution of phenol 28 (3.75 g, 16.7 mmol) in acetonitrile (40 mL), under an atmosphere of nitrogen, was added K₂CO₃ (6.9 g, 50.0 mmol) and stirred for 10 min. Benzyl bromide (6.0 mL, 50.0 mmol) was then added and the resulting mixture allowed to stir for 65 h. The reaction mixture was then quenched with addition of water (50 mL) and extracted with CH_2Cl_2 (3 × 30 mL). The organic phases were combined, washed with water $(2 \times 10 \text{ mL})$ and dried (MgSO₄). Solvent was then removed in vacuo and the crude product purified by column chromatography (9:1 hexanes, ethyl acetate) to give benzyl ether 29 (4.38 g, 83%) as a colourless oil which was used immediately. To a stirred solution of ester 29 (4.3 g, 13.7 mmol) in methanol (30 mL) was added aqueous NaOH (55 mL, 1 M, 4 eq.) and stirred for 2.5 h. The mixture was then acidified with aqueous 2 M HCl, extracted with ethyl acetate (3×50 mL), dried (MgSO₄) and solvent removed in vacuo to give the title compound **32** (3.85 g, 98%) as a white solid. $R_f = 0.30$ (2:1 hexanes, ethyl acetate). Melting point: 99–100°C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.66 (2H, t, J = 7.7 Hz, 2-H), 2.90 (2H, t, J = 7.7 Hz, 3-H), 3.88 (3H, s, 3'-OCH₃), 5.13 (2H, s, 7'-H), 6.68 (1H, dd, J = 8.2, 2.0 Hz, 6'-H), 6.76 (1H, d, J = 2.0 Hz, 2'-H), 6.81 (1H, d, J = 8.2 Hz, 5'-H), 7.27–7.32 (1H, m, 11'-H), 7.34–7.39 (2H, m, 10'-H), 7.41–7.45 (2H, m, 9'-H). δ_C (100 MHz; CDCl₃) 30.4 (C-2), 35.9 (C-3), 56.1 (3'-OCH₃), 71.3 (C-7'), 112.4 (C-2'), 114.5 (C-5'), 120.3 (C-6'), 127.4 (C-9'), 127.9 (C-11'), 128.7 (C-10'), 133.5 (C-1'), 137.4 (C-8'), 146.9 (C-4'), 149.8 (C-3'), 178.8 (C-1). Values are in agreement with literature data [48].

3-(3',4'-Methylenedioxyphenyl)propanoyl chloride (**34b**). To a stirred solution of carboxylic acid **30** (0.22 g, 1.2 mmol) in CH₂Cl₂ (3 mL), under an atmosphere of nitrogen, was added oxalyl chloride (0.2 mL, 2.3 mmol) dropwise and the mixture stirred for 4 h. The solvent was removed in vacuo to give the title compound **34b** (0.24 g, quant.) as a green oil, which was placed under nitrogen and used without further purification.

3-(3',4'-Dimethoxyphenyl)propanoyl chloride (**34a**). To a stirred solution of carboxylic acid **33** (0.24 g, 1.2 mmol) in CH₂Cl₂ (5 mL), under an atmosphere of nitrogen, was added oxalyl chloride (0.2 mL, 2.3 mmol) dropwise and the mixture stirred for 2.5 h. The solvent was removed in vacuo to give the title compound **34a** (0.26 g, quant.) as a yellow oil, which was placed under nitrogen and used without further purification.

3-(3',4',5'-Trimethoxyphenyl) propanoyl chloride (**34c**). To a stirred solution of carboxylic acid **31** (0.25 g, 1.2 mmol) in CH₂Cl₂ (3 mL), under an atmosphere of nitrogen, was added oxalyl chloride (0.2 mL, 2.3 mmol) dropwise and the mixture stirred for 1.5 h. The solvent removed in vacuo to give the title compound **34c** (0.27 g, quant.) as a green crystalline solid, which was placed under nitrogen and used without further purification.

3-(3',4'-Methylenedioxyphenyl)propanoyl chloride (34d). To a stirred solution of carboxylic acid 32 (0.33 g, 1.2 mmol) in CH₂Cl₂ (3 mL), under an atmosphere of nitrogen, was added oxalyl chloride (0.2 mL, 2.3 mmol) dropwise and the mixture stirred for 4 h. The solvent was removed in vacuo to give the title compound 34d (0.35 g, quant.) as a yellow oil, which was placed under nitrogen and used without further purification.

 $(2R^*, 3S^*)$ -2-(3', 4'-Methylenedioxybenzyl)-3-(3'', 4''-dimethoxybenzyl)-1-morpholinopent-4-en-1-one (35ab). Using general procedure A: Morpholine 9a (0.57 g, 2.06 mmol), acid chloride 34b (0.52 g, 2.47 mmol) and reaction time of 24 h. The crude product was purified by column chromatography (2:1 hexanes, ethyl acetate) to give the title compound **35ab** (0.39 g, 42%) as a pale-yellow amorphous solid. $R_f =$ 0.58 (1:3, hexanes, ethyl acetate). Melting point: 114–116 °C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.57 (1H, dd, J = 13.6, 9.0 Hz, 7"-H_A), 2.66–2.73 (1H, m, 3-H), 2.77–2.85 (2H, m, 7'-H_A, OCH_ACH₂N), 2.85–2.94 (4H, m, 2-H, 7'-H_B, 7"-H_B, OCH₂CH_AN), 3.06 (1H, ddd, J = 13.3, 7.9, 3.3 Hz, OCH₂CH_BN), 3.27–3.41 (3H, m, OCH_CCH_CN, OCH_BCH₂N), 3.53–3.60 (1H, m, OH_DCH₂N), 3.67–3.75 (1H, m, OCH₂CH_DN), 3.85 (3H, s, 4["]-OCH₃), 3.86 (3H, s, 3["]-OCH₃), 4.88 (1H, dd, *J* = 16.9, 1.8 Hz, 5-H_A), 4.98 (1H, dd, *J* = 10.3, 1.8 Hz, 5-H_B), 5.85 (1H, ddd, *J* = 16.9, 10.3, 9.5 Hz, 4-H), 5.90 (1H, d, *J* = 1.3 Hz, OCH_AO), 5.91 (1H, d, J = 1.3 Hz, OCH_BO), 6.60 (1H, dd, J = 7.8, 1.6 Hz, 6'-H), 6.64 (1H, d, J = 1.6 Hz, 2'-H), 6.65–6.68 (2H, m, 2", 6"-H), 6.70 (1H, d, J = 7.8 Hz, 5'-H), 6.77 (1H, d, J = 8.7 Hz, 5"-H). δ_C (100 MHz; CDCl₃) 37.4 (C-7'), 38.3 (C-7"), 42.0 (OCH₂CH_{CD}N), 46.4 (OCH₂CH_{AB}N), 46.6 (C-2), 48.5 (C-3), 56.0 (3', 4'-OCH₃), 66.4 (OCH_{AB}CH₂N), 67.0 (OCH_{CD}CH₂N), 101.0 (OCH₂O), 108.4 (C-5'), 109.6 (C-2'), 111.1 (C-5''), 112.4 (C-2"), 116.8 (C-5), 121.3 (C-6"), 122.0 (C-6'), 132.3 (C-1"), 133.6 (C-1'), 139.3 (C-4), 146.2 (C-4'), 147.5 (C-4"), 147.7 (C-3'), 148.9 (C-3"), 172.6 (C-1). IR: v_{MAX} (film)/cm⁻¹; 2963, 1631, 1515, 1488, 1442, 1236, 1031, 925, 807, 730. HRMS (ESI⁺) Found [M + H]⁺ 454.2241; C₂₆H₃₂NO₆ requires 454.2224.

 $(2R^*,3S^*)-2-(3',4',5'-Trimethoxybenzyl)-3-(3'',4''-dimethoxybenzyl)-1-morpholinopent-4-en-1-one$ (35ac). Using general procedure A: Morpholine 9a (0.47 g, 1.7 mmol), acid chloride 34c (0.53 g, 2.0 mmol) and a reaction time of 19 h. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound 35ac (0.50 g, 58%) as a yellow oil. R_f = 0.38 (1:3 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.59 (1H, dd, J = 13.6, 9.2 Hz, $7''-H_{\rm A}$), 2.67–2.74 (1H, m, 3-H), 2.78 (1H, ddd, J = 11.4, 7.8, 3.0 Hz, NCH₂CH₄O), 2.82–2.96 (5H, m, 2-H, 7'-H, 7''-H_B, NCH₄CH₂O), 3.06 (1H, ddd, J = 13.2, 7.8, 3.0 Hz, NCH₂CH₂O), 3.25–3.40 (3H, m, NCH_BCH₂O, NCH_CCH_CO), 3.54–3.61 (1H, m, NCH_DCH₂O), 3.67–3.73 (1H, m, NCH₂CH_DO), 3.80 (3H, s, 4'-OCH₃), 3.82 (6H, s, 3'-OCH₃), 4.90 (1H, dd, J = 17.0, 1.8 Hz, 5-H_A), 5.00 (1H, dd, J = 17.0, 10.2, 9.1 Hz, 4-H), 6.37 (2H, s, 2'-H), 6.66–6.70 (2H, m, 2'', 6''-H), 6.78 (1H, d, J = 8.7 Hz, 5''-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 38.1 (C-7'), 38.3 (C-7''), 42.0 (NCH_CCH₂O),

46.4 (NCH_{AB}CH₂O), 46.5 (C-2), 48.7 (C-3), 56.0 (3", 4"-OCH₃), 56.3 (3'-OCH₃), 61.0 (4'-OCH₃), 66.4 (NCH₂CH_{AB}O), 66.9 (NCH₂CH_{CD}O), 106.2 (C-2'), 111.1 (C-5"), 112.5 (C-2"), 116.8 (C-5), 121.2 (C-6"), 132.3 (C-1"), 135.6 (C-1'), 136.8 (C-4'), 139.2 (C-4), 147.5 (C-4"), 148.8 (C-3"), 153.3 (C-3'), 172.6 (C-1). IR: ν_{MAX} (film)/cm⁻¹; 2940, 1632, 1589, 1459, 1236, 1123, 1028, 913, 735. HRMS (ESI⁺) Found [M + Na]⁺ 522.2474; C₂₈H₃₇NNaO₇ requires 522.2462.

 $(2R^*,3S^*)$ -2-(3',4'-Dimethoxybenzyl)-3-(3'',4''-dimethoxybenzyl)-1-morpholinopent-4-en-1-one (**35aa**). Using general procedure A: Morpholine **9a** (0.53 g, 1.91 mmol), acid chloride **34a** (0.52 g, 2.29 mmol) and a reaction time of 24 h. The crude product was purified by flash chromatography (1:3 hexanes, ethyl acetate) to give the title compound **35aa** (0.63 g, 77% yield) as a pale-yellow amorphous solid. R_f = 0.42 (19:1 CH₂Cl₂, methanol). Melting point: 98–101 °C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.55–2.63 (1H, m, 7''-H_A), 2.85–2.93 (1H, m, 7''-H_B), 2.67–2.85 (3H, m, 3-H, OCH₂CH_{AB}N), 3.29-3.37 (4H, m, OCH₂CH_{CD}N, OCH_{AB}CH₂N), 2.85–3.06 (3H, m, 2-H, 7'-H), 3.50–3.67 (2H, m, OCH_{CD}CH₂N), 3.83, 3.84, 3.85, 3.86 (12H, s, 3', 4', 3'', 4''-OCH₃), 4.89 (1H, dd, *J* = 17.1, 1.7 Hz, 5-H), 4.99 (1H, dd, *J* = 10.3, 1.9 Hz, 5-H), 5.82–5.91 (1H, m, 4-H), 6.67–6.69 (4H, m, 2', 6', 2'', 6''-H), 6.75–6.78 (2H, m, 5', 5''-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 37.2 (C-2), 38.2 (C-7''), 41.9, 46.5 (OCH₂CH₂N), 46.2 (C-7'), 48.5 (C-3), 55.8, 55.9 (3', 4', 3'', 4''-OCH₃), 66.3, 66.8 (OCH₂CH₂N), 111.0, 111.3 (C-5', 5''), 112.4, 112.6 (C-2', 2''), 116.6 (C-5), 120.9, 121.2 (C-6', 6''), 132.2, 132.3 (C-1', 1''), 139.2 (C-4), 147.3, 147.6 (4', 4''-OCH₃), 148.7, 148.8 (3', 3''-OCH₃), 172.6 (C-1). IR: v_{MAX} (film)/cm⁻¹; 2935, 1628, 1591, 1462, 1260, 1155, 1027, 912, 857, 765. HRMS (ESI⁺) Found [M + H]⁺ 470.2537; C₂₇H₃₆NO₆ requires 470.2537

(2*R**,3*S**)-2-(3'-*Methoxy*-4'-*benzyloxybenzyl*)-3-(3",4"-*dimethoxybenzyl*)-1-*morpholino-pent*-4-*en*-1-*one* (**35ad**). Using general procedure A: Morpholine **9a** (0.47 g, 1.7 mmol), acid chloride **34d** (0.62 g, 2.0 mmol) and a reaction time of 22 h. The crude product was purified by column chromatography (2:1 hexanes, ethyl acetate) to give the title compound **35ad** (0.59 g, 64%) as a yellow oil.

R_f = 0.58 (1:3, hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.57 (1H, dd, *J* = 13.5, 9.0 Hz, 7"-H_A), 2.62–2.68 (1H, m, 3-H), 2.68–2.74 (1H, m, OCH_ACH₂N), 2.75–2.82 (1H, m, OCH₂CH_AN), 2.83–2.92 (4H, m, 2-H, 7'-H, 7"-H_B), 2.99 (1H, ddd, *J* = 13.3, 7.6, 3.2 Hz, OCH₂CH_BN), 3.20–3.32 (3H, m, OCH_BCH₂N, OCH_CCH_CN), 3.50–3.55 (1H, m, OCH_DCH₂N), 3.61–3.67 (1H, m, OCH₂CH_DN), 3.84 (3H, s, 3'-OCH₃), 3.85 (3H, s, 4"-OCH₃), 3.85 (3H, s, 3"-OCH₃), 4.88 (1H, dd, *J* = 17.1, 1.9 Hz, 5-H_A), 4.97 (1H, dd, *J* = 10.3, 1.9 Hz, 5-H_B), 5.13 (1H, s, 7^{'''}-H), 5.85 (1H, ddd, *J* = 17.1, 10.3, 9.0 Hz, 4-H), 6.59 (1H, dd, *J* = 8.1, 1.9 Hz, 6'-H), 6.65–6.68 (2H, m, 2"-H), 6.69 (1H, d, *J* = 1.9 Hz, 2'-H), 6.74 (1H, d, *J* = 8.1 Hz, 5'-H), 6.77 (1H, d, *J* = 8.5 Hz, 5"-H), 7.25–7.30 (1H, m, 4^{'''}-H), 7.32–7.37 (2H, m, 3^{'''}-H), 7.38–7.42 (2H, m, 2^{'''}-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 37.4 (C-7'), 38.3 (C-7''), 41.9 (OCH₂CH_{CD}N), 46.3 (OCH₂CH_{AB}N), 46.5 (C-2), 48.6 (C-3), 56.0, 56.2 (3', 3", 4^{''}-OCH₃), 66.4 (OCH_{AB}CH₂N), 66.9 (OCH_{CD}CH₂N), 71.2 (C-7^{'''}), 111.1 (C-5^{''}), 112.4 (C-2^{''}), 113.3 (C-2'), 114.6 (C-5'), 116.7 (C-5), 120.9 (C-6'), 121.3 (C-6''), 127.3 (C-4''), 148.8 (C-3''), 149.7 (C-3''), 172.7 (C-1). IR: ν_{MAX} (film)/cm⁻¹; 2936, 1736, 1633, 1513, 1454, 1261, 1140, 1028, 915, 733. HRMS (ESI⁺) Found [M + Na]⁺ 568.2671; C₃₃H₃₉NNaO₆ requires 568.2670.

(2*R**,3*S**)-2-(3',4'-Dimethoxybenzyl)-3-(3'',4''-methylenedioxybenzyl)-1-morpholinopent-4-en-1-one (**35ba**). Using general procedure A: Morpholine **9b** (0.25 g, 0.96 mmol), acid chloride **34a** (0.26 g, 1.2 mmol) and a reaction time of 21 h. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound **35ba** (0.36 g, 83%) as a yellow oil.

 R_f = 0.50 (1:3 hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 2.56 (1H, dd, *J* = 13.4, 9.0 Hz, 7"-H_A), 2.62–2.70 (1H, m, 3-H), 2.75–2.94 (6H, m, 2-H, 7'-H, 7"-H_B, NCH_ACH_AO), 3.05 (1H, ddd, *J* = 13.6, 7.9, 3.1 Hz, NCH_BCH₂O), 3.28–3.41 (3H, m, NCH₂CH_BO, NCH_CCH_CO), 3.51–3.57 (1H, m, NCH₂CH_DO), 3.58–3.64 (1H, m, NCH_DCH₂O), 3.83 (3H, s, 3'-H), 3.84 (3H, s, 4'-H), 4.89 (1H, dd, *J* = 17.2, 1.9 Hz, 5-H_A), 4.99 (1H, dd, *J* = 10.2, 1.9 Hz, 5-H_B), 5.86 (1H, ddd, *J* = 17.2, 10.2, 9.1 Hz, 4-H), 5.92 (1H, d, *J* = 1.4 Hz, OCH_AO), 5.92 (1H, d, *J* = 1.4 Hz, OCH_BO), 6.58 (1H, dd, *J* = 7.9, 1.6 Hz, 6"-H), 6.64 (1H, d, *J* = 1.6 Hz, 2"-H), 6.66–6.70 (2H, m, 2', 6'-H), 6.71 (1H, d, *J* = 7.9 Hz, 5"-H), 6.76 (1H, d, *J* = 8.1 Hz, 5'-H).

$$\begin{split} &\delta_{C} \ (100 \ \text{MHz; CDCl}_3) \ 37.3 \ (\text{C-7'}), \ 38.5 \ (\text{C-7''}), \ 42.0 \ (\text{NCH}_{AB}\text{CH}_2\text{O}), \ 46.3 \ (\text{NCH}_{CD}\text{CH}_2\text{O}), \ 46.5 \ (\text{C-2}), \\ &48.8 \ (\text{C-3}), \ 56.1 \ (3', \ 4'-\text{OCH}_3), \ 66.4 \ (\text{NCH}_2\text{CH}_{AB}\text{O}), \ 66.9 \ (\text{NCH}_2\text{CH}_{CD}\text{O}), \ 101.0 \ (\text{OCH}_2\text{O}), \ 108.1 \ (\text{C-5''}), \\ &109.6 \ (\text{C-2''}), \ 111.4 \ (\text{C-5'}), \ 112.7 \ (\text{C-2'}), \ 116.8 \ (\text{C-5}), \ 121.0 \ (\text{C-6''}), \ 122.1 \ (\text{C-6''}), \ 132.4 \ (\text{C-1'}), \ 133.7 \ (\text{C-1''}), \\ &139.2 \ (\text{C-4}), \ 145.9 \ (\text{C-4''}), \ 147.8 \ (\text{C-3''}), \ 149.0 \ (\text{C-3'}), \ 172.7 \ (\text{C-1}). \ \text{IR: } \nu_{\text{MAX}} \ (\text{film})/\text{cm}^{-1}; \ 2908, \\ &1740, \ 1630, \ 1515, \ 1441, \ 1237, \ 1029, \ 923, \ 730. \ \text{HRMS} \ (\text{ESI}^+) \ \text{Found} \ [\text{M} + \text{Na}]^+ \ 476.2042; \ \text{C}_{26}\text{H}_{31}\text{NNaO}_6 \\ &\text{requires} \ 476.2044. \end{split}$$

 $(2R^*, 3S^*)$ -2-(3', 4'-Methylenedioxybenzyl)-3-(3'', 4''-methylenedioxybenzyl)-1-morpholinopent-4-en-1-one (35bb). Using general procedure A: Morpholine 9b (0.5 g, 1.91 mmol), acid chloride 34b (0.49 g, 2.30 mmol) and a reaction time of 30 min. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound **35bb** (0.798 g, 95%) as a pale-yellow solid. $R_f =$ 0.68 (1:3 hexanes, ethyl acetate). Melting point: 131–133 °C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.54 (1H, dd, J = 13.5, 8.9 Hz, 7"-H_A), 2.61–2.69 (1H, m, 3-H), 2.78–2.93 (6H, m, 2-H, 7'-H, 7"-H_B, NCH_ACH_AO), 3.06 (1H, ddd, J = 13.2, 7.8, 3.1 Hz, NCH_BCH₂O), 3.29–3.41 (3H, m, NCH₂CH_BO, NCH_CCH_CO), 3.53–3.61 (1H, m, NCH₂CH_DO), 3.66–3.74 (1H, m, NCH_DCH₂O), 4.89 (1H, dd, J = 17.0, 1.9 Hz, 5-H_A), 4.99 (1H, dd, J = 10.2, 1.9 Hz, 5-H_B), 5.85 (1H, ddd, J = 17.0, 10.2, 9.1 Hz, 4-H), 5.90 (1H, d, J = 1.4 Hz, 3'-OCH_AO), 5.91 (1H, d, I = 1.4 Hz, 3'-OCH_BO), 5.92 (1H, d, I = 1.5 Hz, 3"-OCH_AO), 5.93 (1H, d, $J = 1.5 \text{ Hz}, 3''-\text{OCH}_{B}\text{O}$), 6.55–6.61 (2H, m, 6', 6''-H), 6.62–6.64 (2H, m, 2', 2''-H), 6.70, 6.71 (2 × 1H, $2 \times d_{I} = 8.0 \text{ Hz}, 5', 5''-\text{H}$). δ_{C} (100 MHz; CDCl₃) 37.4 (C-7'), 38.5 (C-7''), 42.0 (NCH_{CD}CH₂O), 46.4 (NCH_{AB}CH₂O), 46.5 (C-2), 48.8 (C-3), 66.4 (NCH₂CH_{AB}O), 67.0 (NCH₂CH_{CD}O), 101.0 (2 × OCH₂O), 108.2, 108.4 (C-5', 5"), 109.6 (C-2', 2"), 116.8 (C-5), 122.1 (C-6', 6"), 133.6 (C-1', 1"), 139.2 (C-4), 145.9, 146.2 (C-4', 4"), 147.6, 147.7 (C-3', 3"), 172.6 (C-1). IR: v_{MAX} (film)/cm⁻¹; 2897, 1630, 1487, 1440, 1244, 1036, 925, 808, 730. HRMS (ESI⁺) Found [M + Na]⁺ 460.1722; C₂₅H₂₇NNaO₆ requires 460.1731.

 $(2R^*,3S^*)-2-(3',4',5'-Trimethoxybenzyl)-3-(3'',4''-methylenedioxybenzyl)-1-morpholinopent-4-en-1-one (35bc).$ Using general procedure A: Morpholine 9b (0.25 g, 0.96 mmol), acid chloride 34c (0.27 g, 1.2 mmol) and a reaction time of 18 h. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound **35bc** (0.40 g, 86%) as a pale-yellow solid. $R_f = 0.55$ (1:3 hexanes, ethyl acetate). Melting point: 104–106 °C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.56 (1H, dd, J = 13.4, 9.0 Hz, 7"-H_A), 2.62–2.70 (1H, m, 3-H), 2.75–2.95 (6H, m, 2-H, 7'-H, 7"-H_B, NCH_ACH_AO), 3.06 (1H, ddd, J = 13.2, 7.7, 3.0 Hz, NCH_BCH₂O), 3.25–3.40 (3H, m, NCH₂CH_BO, NCH_CCH_CO), 3.54–3.60 (1H, m, NCH₂CH_DO), 3.65–6.71 (1H, m, NCH_DCH₂O), 3.80 (3H, s, 4'-OCH₃), 3.82 (6H, s, 3'-OCH₃), 4.90 (1H, dd, J = 17.2, 1.9 Hz, 5-H_A), 5.00 (1H, dd, J = 10.2, 1.9 Hz, 5-H_B), 5.85 (1H, ddd, J = 17.2, 10.2, 9.0 Hz, 4-H), 5.92 (1H, d, J = 1.4 Hz, OCH_AO), 5.93 (1H, d, J = 1.4 Hz, OCH_BO), 6.36 (2H, s, 2'-H), 6.59 (1H, dd, J = 7.9, 1.6 Hz, 6"-H), 6.65 (1H, d, J = 1.6 Hz, 2"-H), 6.72 (1H, d, J = 7.9 Hz, 5"-H). δ_{C} (100 MHz; CDCl₃) 38.1 (C-7'), 38.5 (C-7"), 42.0 (NCH_{CD}CH₂O), 46.4 (C-2, NCH_{AB}CH₂O), 48.9 (C-3), 56.4 (3'-OCH₃), 61.1 (4'-OCH₃), 66.4 (NCH₂CH_{AB}O), 67.0 (NCH₂CH_{CD}O), 101.0 (OCH₂O), 106.2 (C-2'), 108.2 (C-5"), 109.6 (C-2"), 116.9 9 (C-5), 122.1 (C-6"), 133.6 (C-1"), 135.6 (C-1'), 136.9 (C-4'), 139.1 (C-4), 145.9 (C-4"), 147.7 (C-3"), 153.3 (C-3'), 172.6 (C-1). IR: v_{MAX} (film)/cm⁻¹; 2922, 1632, 1589, 1490, 1240, 1120, 1036, 925, 730. HRMS (ESI⁺) Found [M + Na]⁺ 506.2145; C₂₇H₃₃NNaO₇ requires 506.2149.

(2*R**,3*S**)-2-(3'-*Methoxy*-4'-*benzyloxybenzyl*)-3-(3",4"-*methylenedioxybenzyl*)-1-*morpholinopent*-4-*en*-1-*one* (**35bd**). Using general procedure A: Morpholine **9b** (0.25 g, 0.96 mmol), acid chloride **34d** (0.35 g, 1.2 mmol) and a reaction time of 18 h. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound **35bd** (0.45 g, 88%) as a yellow oil.

R_f = 0.67 (1:3 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.54 (1H, dd, *J* = 13.5, 8.9 Hz, 7"-H_A), 2.61–2.70 (2H, m, 3-H, NCH₂CH_AO), 2.73–2.91 (5H, m, 2-H, 7'-H, 7"-H_B, NCH_ACH₂O), 2.99 (1H, ddd, *J* = 13.2, 7.7, 3.0 Hz, NCH_BCH₂O), 3.20–3.35 (3H, m, NCH₂CH_BO, NCH_CCH_CO), 3.53 (1H, ddd, *J* = 11.0, 5.5, 2.5 Hz, NCH₂CH_DO), 3.62 (1H, ddd, *J* = 13.0, 5.5, 2.5 Hz, NCH_DCH₂O), 3.84 (3H, s, 3'-OCH₃), 4.88 (1H, dd, *J* = 17.0, 1.9 Hz, 5-H_A), 4.98 (1H, dd, *J* = 10.2, 1.9 Hz, 5-H_B), 5.13 (2H, s, 7"'-H), 5.84 (1H, ddd, *J* = 17.0, 10.2, 9.1 Hz, 4-H), 5.91 (1H, d, *J* = 1.4 Hz, OCH_AO), 5.92 (1H, d, *J* = 1.4 Hz,

OCH_BO), 6.57 (1H, dd, *J* = 8.0, 1.9 Hz, 6″-H), 6.59 (1H, dd, *J* = 8.2, 1.8 Hz, 6′-H), 6.64 (1H, d, *J* = 1.8 Hz, 2″-H), 6.69 (1H, d, *J* = 1.9 Hz, 2″-H), 6.71 (1H, d, *J* = 8.0 Hz, 5″-H), 6.75 (1H, d, *J* = 8.2 Hz, 5′-H), 7.25–7.30 (1H, m, 4‴-H), 7.32–7.37 (2H, m, 3‴-H), 7.38–7.43 (2H, m, 2‴-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 37.4 (C-7′), 38.5 (C-7″), 41.9 (NCH_{CD}CH₂O), 46.3 (NCH_{AB}CH₂O), 46.4 (C-2), 48.8 (C-3), 56.2 (3′-OCH₃), 66.3 (NCH₂CH_{AB}O), 66.9 (NCH₂CH_{CD}O), 71.2 (C-7‴), 100.9 (OCH₂O), 108.1 (C-5″), 109.6 (C-2″), 113.2 (C-2′), 114.5 (C-5′), 116.7 (C-5), 121.0 (C-6′), 122.1 (C-6″), 127.3 (C-2″'), 127.9 (C-4″'), 128.6 (C-3″'), 133.1 (C-1′) 133.6 (C-1″), 137.3 (C-1‴), 139.2 (C-4), 145.9 (C-4″), 146.7 (C-4′), 147.6 (C-3″), 149.7 (C-3′), 172.6 (C-1). IR: ν_{MAX} (film)/cm⁻¹; 2920, 1630, 1489, 1231, 1114, 1034, 913, 729. HRMS (ESI⁺) Found [M + Na]⁺ 552.2354; C₃₂H₃₅NNaO₆ requires 552.2357.

 $(3R^*,4R^*)$ -3-(3',4'-Methylenedioxybenzyl)-4-(3'',4''-dimethoxybenzyl)-5-(hydroxymethyl)dihydrofuran-2(3H)-*one* (**4ab**). Using general procedure B: Amide **35ab** (0.38 g, 0.84 mmol) in ^tBuOH/H₂O and a reaction
time of 3 days. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate)
to give the title compound **4ab** (180 mg, 54%) as a white foam. R_f = 0.50 (19:1 CH₂Cl₂, methanol). $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.79 (1H, t, *J* = 6.4 Hz, 6-OH), 2.36–2.44 (1H, m, 4-H), 2.51 (1H, dd, *J* = 13.7, 7.9 Hz,
7''-H_A), 2.58 (1H, dd, *J* = 13.7, 6.6 Hz, 7''-H_B), 2.68 (1H, ddd, *J* = 9.3, 7.0, 5.5 Hz, 3-H), 2.85 (1H, dd, *J* =
14.0, 7.0 Hz, 7'-H_A), 2.92 (1H, dd, *J* = 14.0, 5.5 Hz, 7'-H_B), 3.15 (1H, ddd, *J* = 12.5, 6.4, 5.1 Hz, 6-H_A), 3.54
(1H, ddd, *J* = 12.5, 6.4, 2.5 Hz, 6-H_B), 3.83 (3H, s, 3''-OCH₃), 3.85 (3H, s, 4''-OCH₃), 4.19 (1H, ddd, *J* =
8.0, 5.1, 2.5 Hz, 5-H), 5.92 (1H, d, *J* = 1.5 Hz, OCH_AH_BO), 5.93 (1H, d, *J* = 1.5 Hz, OCH_AH_BO), 6.47 (1H,
d, *J* = 2.0 Hz, 2''-H), 6.57–6.60 (2H, m, 6' and 6''-H), 6.61 (1H, d, *J* = 1.5 Hz, OCH_AH_BO), 6.47 (1H,
d, *J* = 2.0 Hz, 2''-H), 6.57–6.60 (2H, m, 6' and 6''-H), 6.61 (1H, d, *J* = 1.5 Hz, OC+A_AH_BO), 6.47 (1H,
d, *J* = 2.0 Hz, 2''-H), 6.57–6.60 (2H, m, 6' and 6''-H), 6.61 (1H, d, *J* = 1.5 Hz, OC+A_AH_BO), 6.47 (1H,
d, *J* = 8.1 Hz, 5''-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 35.3 (C-7'), 38.7 (C-7''), 41.6 (C-4), 47.6 (C-3),
56.0 (3''-OCH₃, 4''-OCH₃), 63.2 (C-6), 84.1 (C-5), 101.2 (OCH₂O), 108.3 (C-5'), 109.7 (C-2'), 111.4 (C-5''),
112.0 (C-2''), 121.0 (C-6''), 122.5 (C-6'), 130.3 (C-1''), 131.6 (C-1'), 146.6 (C-4'), 148.0 (C-4''), 148.1 (C-3'),
149.3 (C-3''), 177.7 (C-2). IR: $\nu_{\rm MAX}$ (film)/cm⁻¹; 3496 (broad), 2936, 2254, 1760, 1515, 1489, 1442, 1239,
1025, 909, 809, 766. HRMS (ESI⁺) Found [M + Na]⁺ 423.1427; C₂₂H₂₄NaO₇ requires 423.1414.

 $(3R^*,4R^*)$ -3,4-*bis*(3',4'-*Dimethoxybenzyl*)-5-(*hydroxymethyl*)*dihydrofuran*-2(3*H*)-*one* (**4aa**). Using general procedure B: Amide **35aa** (0.29 g, 0.61 mmol), in ^tBuOH/H₂O and a reaction time of 6 days. The crude product was purified by flash chromatography (1:1 hexanes, ethyl acetate) to give the title compound **4aa** (0.18 g, 70%) as a colourless oil. R_f = 0.32 (19:1 CH₂Cl₂, methanol). δ_{H} (400 MHz; CDCl₃) 2.39–2.44 (1H, m, 4-H), 2.53 (1H, dd, *J* = 13.7, 7.3 Hz, 7''-H_A), 2.58 (1H, dd, *J* = 13.7, 6.5 Hz, 7''-H_B), 2.64 (1H, br s, 6-OH), 2.71 (1H, ddd, *J* = 9.3, 6.7, 5.7 Hz, 3-H), 2.88 (1H, dd, *J* = 14.0, 6.7 Hz, 7'-H_A), 2.94 (1H, dd, *J* = 14.0, 5.5 Hz, 7'-H_B), 3.16 (1H, dd, *J* = 12.6, 4.9 Hz, 6-H_A), 3.53 (1H, dd, *J* = 12.6, 2.4 Hz, 6-H_B), 3.81, 3.83, 3.84 (12H, s, 3', 4', 3'', 4''-OCH₃), 4.15 (1H, ddd, *J* = 8.0, 4.9, 2.4 Hz, 5-H), 6.49 (1H, d, *J* = 1.9 Hz, 2''-H), 6.57 (1H, dd, *J* = 8.1, 1.9 Hz, 6''-H), 6.66–6.68 (2H, m, 2', 6'-H), 6.73–6.80 (2H, m, 5', 5''-H). δ_{C} (100 MHz; CDCl₃) 35.0 (C-7'), 38.5 (C-7''), 41.6 (C-4), 47.5 (C-3), 55.8 (3', 4', 3'', 4''-OCH₃), 62.9 (C-6), 84.0 (C-5), 111.2, 111.4 (C-5', 5''), 112.1 (C-2''), 112.6 (C-2'), 120.9 (C-6''), 121.4 (C-6'), 130.4 (C-1', 1''), 147.9 (C-4', 4''), 149.0 (C-3', 3''), 178.0 (C-2). IR: ν_{MAX} (film)/cm⁻¹; 3505 (br), 2938, 1761, 1591, 1514, 1465, 1259, 1156, 1025, 910, 808, 766, 647. HRMS (ESI⁺) Found [M + H]⁺ 417.1909; C₂₃H₂₉O₇ requires 417.1908.

 $(3R^*,4R^*)$ -3-(3',4',5'-Trimethoxybenzyl)-4-(3'',4''-dimethoxybenzyl)-5-(hydroxymethyl)dihydrofuran-2(3H)-one (4ac). Using general procedure B: Amide 35ac (0.45 g, 0.90 mmol) in ^tBuOH/H₂O/THF and a reaction time of 3 days. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound 4ac (0.17 g, 42%) as a pale-yellow solid.

R_f = 0.31 (19:1 CH₂Cl₂, methanol). Melting point: 141–142 °C. $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.68 (1H, t, *J* = 6.5 Hz, 6-OH), 2.38–2.46 (1H, m, 4-H), 2.55 (1H, dd, *J* = 13.8, 8.2 Hz, 7"-H_A), 2.65 (1H, dd, *J* = 13.8, 5.9 Hz, 7"-H_B), 2.72 (1H, ddd, *J* = 9.7, 6.3, 5.7 Hz, 3-H), 2.90 (1H, dd, *J* = 14.0, 6.3 Hz, 7'-H_A), 2.95 (1H, dd, *J* = 14.0, 5.7 Hz, 7'-H_B), 3.15 (1H, ddd, *J* = 12.4, 5.1, 5.4 Hz, 6-H_A), 3.54 (1H, ddd, *J* = 12.4, 6.5, 2.5 Hz, 6-H_B), 3.82 (6H, s, 4', 3"-OCH₃), 3.83 (6H, s, 3'-OCH₃), 3.85 (3H, s, 4"-OCH₃), 4.20 (1H, ddd, *J* = 8.2, 5.1, 2.5 Hz, 5-H), 6.38 (2H, s, 2'-H), 6.49 (1H, d, *J* = 2.0 Hz, 2"-H), 6.58 (1H, dd, *J* = 8.1, 2.0 Hz, 6"-H), 6.76 (1H, d, *J* = 8.1 Hz, 5"-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 35.7 (C-7'), 38.6 (C-7"), 41.8 (C-4), 47.7 (C-3), 56.0,

56.1 (3", 4"-OCH₃), 56.3 (3'-OCH₃), 61.0 (4'-OCH₃), 63.2 (C-6), 83.9 (C-5), 106.5 (C-2'), 111.5 (C-5"), 112.2 (C-2"), 121.0 (C-6"), 130.3 (C-1"), 133.7 (C-1'), 137.2 (C-4'), 148.2 (C-4"), 149.3 (C-3"), 153.5 (C-3'), 177.7 (C-2). IR: ν_{MAX} (film)/cm⁻¹; 3527 (br), 2938, 1761, 1590, 1514, 1237, 1126, 1026, 735. HRMS (ESI⁺) Found [M + Na]⁺ 469.1839; C₂₄H₃₀NaO₈ requires 469.1833.

2(3H)-one (4ad). Using general procedure B: Amide 35ad (0.59 g, 1.1 mmol) in ^tBuOH/H₂O and a reaction time of 7 days. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound 4ad (0.30 g, 56%) as a cloudy oil. $R_f = 0.27$ (19:1 CH₂Cl₂, methanol). δ_H (400 MHz; CDCl₃) 1.57 (1H, t, *J* = 6.5 Hz, 6-OH), 2.34–2.42 (1H, m, 4-H), 2.50 (1H, dd, *J* = 13.5, 8.0 Hz, 7''-H_A), 2.59 (1H, dd, J = 13.5, 6.0 Hz, 7''-H_B), 2.70 (1H, ddd, J = 9.7, 6.2, 5.6 Hz, 3-H), 2.90 $(1H, dd, J = 14.1, 6.2 Hz, 7'-H_A), 2.94 (1H, dd, J = 14.1, 5.6 Hz, 7'-H_B), 3.10 (1H, ddd, J = 12.5, 6.5, 5.2)$ Hz, 6-H_A), 3.48 (1H, ddd, *J* = 12.5, 6.5, 2.7 Hz, 6-H_B), 3.80 (3H, s, 3"-OCH₃), 3.86 (6H, s, 3', 4"-OCH₃), 4.18 (1H, ddd, J = 8.3, 5.2, 2.7 Hz, 5-H), 5.12 (2H, s, 7^{'''}-H), 6.46 (1H, d, J = 2.0 Hz, 2^{''}-H), 6.56 (1H, dd, *J* = 8.0, 2.0 Hz, 6^{*II*}-H), 6.61 (1H, dd, *J* = 8.1, 2.0 Hz, 6^{*I*}-H), 6.72 (1H, d, *J* = 2.0 Hz, 2^{*I*}-H), 6.75 (1H, d, *J* = 8.0 Hz, 5"-H), 6.79 (1H, d, J = 8.1 Hz, 5'-H), 7.25–7.30 (1H, m, 4"'-H), 7.31–7.36 (2H, m, 3"'-H), 7.39–7.42 (2H, m, 2^{///}-H). δ_C (100 MHz; CDCl₃) 35.1 (C-7[/]), 38.6 (C-7^{//}), 41.7 (C-4), 47.6 (C-3), 56.0 (3^{//}, 4^{//}-OCH₃), 56.2 (3'-OCH₃), 63.3 (C-6), 71.3 (C-7^{'''}), 84.0 (C-5), 111.5 (C-5^{''}), 112.1 (C-2^{''}), 113.3 (C-2[']), 114.3 (C-5[']), 121.0 (C-6"), 121.6 (C-6'), 127.4 (C-2""), 128.0 (C-4""), 128.7 (C-3""), 130.3 (C-1"), 131.1 (C-1'), 137.2 (C-1'''), 147.2 (C-4'), 148.2 (C-4''), 149.3 (C-3''), 150.0 (C-3'), 177.8 (C-2). IR: v_{MAX} (film)/cm⁻¹; 3523 (br), 2935, 1761, 1514, 1261, 1025, 911, 730. HRMS (ESI⁺) Found [M + Na]⁺ 515.2023; C₂₉H₃₂NaO₇ requires 515.2040.

(3*R**,4*R**,5*S**)-4-(3",4"-Dimethoxybenzyl)-3-(4'-hydroxy-3'-methoxybenzyl)-5-(hydroxymethyl)dihydrofuran-2(3*H*)-one (**4ae**). Using general procedure F: Benzyl ether **4ad** (0.27 g, 0.55 mmol) gave the title compound **4ae** (0.19 g, 88%) as a yellow solid. R_f = 0.43 (19:1 CH₂Cl₂, methanol). Melting point: 183–185 °C. δ_H (400 MHz; CDCl₃) 1.63 (1H, t, *J* = 6.5 Hz, 6-OH), 2.34–2.43 (1H, m, 4-H), 2.53 (1H, dd, *J* = 13.8, 8.1 Hz, 7"-H_A), 2.62 (1H, dd, *J* = 13.8, 6.1 Hz, 7"-H_B), 2.69 (1H, dt, *J* = 9.5, 6.0 Hz, 3-H), 2.92 (2H, d, *J* = 6.0 Hz, 7'-H), 3.13 (1H, ddd, *J* = 12.5, 6.5, 5.3 Hz, 6-H_A), 3.51 (1H, ddd, *J* = 12.5, 6.5, 2.5 Hz, 6-H_B), 3.82 (3H, s, 3'-OCH₃), 3.84 (3H, s, 3"-OCH₃), 3.85 (3H, s, 4"-OCH₃), 4.19 (1H, ddd, *J* = 8.0, 5.3, 2.5 Hz, 5-H), 5.52 (1H, s, 4'-OH), 6.46 (1H, d, *J* = 2.0 Hz, 2"-H), 6.76 (1H, dd, *J* = 8.1 Hz, 5"-H), 6.83 (1H, dd, *J* = 8.0 Hz, 5'-H), 6.6C (100 MHz; CDCl₃) 35.1 (C-7'), 38.6 (C-7"), 41.6 (C-4), 47.7 (C-3), 56.0, 56.1 (3', 3", 4"-OCH₃), 63.4 (C-6), 84.1 (C-5), 111.5 (C-5"), 111.9 (C-2'), 112.1 (C-2"), 114.4 (C-5'), 121.0 (C-6"), 122.3 (C-6'), 129.7 (C-1'), 130.4 (C-1"), 144.7 (C-4'), 146.8 (C-3'), 148.2 (C-4"), 149.3 (C-3"), 177.8 (C-2). IR: ν_{MAX} (film)/cm⁻¹; 3438 (br), 2937, 1755, 1514, 1236, 1155, 1025, 907, 723. HRMS (ESI⁺) Found [M + Na]⁺ 425.1564; C₂₂H₂₆NaO₇ requires 425.1571.

 $(3R^*,4R^*)$ -3,4-bis(3',4'-Methylenedioxybenzyl)-5-(hydroxymethyl)dihydrofuran-2(3H)-one (**4bb**). Using general procedure B: Morpholine amide **35bb** (0.322 g, 0.74 mmol) in ^tBuOH/H₂O/THF and a reaction time of 5 days. The crude product was then purified by column chromatography (2:1 hexanes, ethyl acetate) to give the title compound **4bb** (0.145 g, 51%) as a pale-yellow oil.

 $\begin{array}{l} R_{\rm f} = 0.59 \; (1:3 \; {\rm hexanes, ethyl acetate}). \; \delta_{\rm H} \; (400 \; {\rm MHz; CDCl_3}) \; 1.72 \; (1{\rm H, br, 6-OH}), 2.32-2.41 \; (1{\rm H, m, 4-H}), \\ 2.47 \; (1{\rm H, dd}, J = 13.7, 8.1 \; {\rm Hz}, 7''-{\rm H_A}), 2.56 \; (1{\rm H, dd}, J = 13.7, 6.2 \; {\rm Hz}, 7''-{\rm H_B}), 2.65 \; (1{\rm H, ddd}, J = 9.0, 7.5, \\ 5.3 \; {\rm Hz}, 3-{\rm H}), 2.85 \; (1{\rm H, dd}, J = 14.0, 7.5 \; {\rm Hz}, 7'-{\rm H_A}), 2.96 \; (1{\rm H, dd}, J = 14.0, 5.3 \; {\rm Hz}, 7''-{\rm H_B}), 3.15 \; (1{\rm H, dd}, J = 12.6, 4.9 \; {\rm Hz}, 6-{\rm H_A}), 3.54 \; (1{\rm H, dd}, J = 12.6, 2.5 \; {\rm Hz}, 6-{\rm H_B}), 4.18 \; (1{\rm H, ddd}, J = 7.7, 4.9, 2.5 \; {\rm Hz}, 5-{\rm H}), \\ 5.93-5.95 \; (4{\rm H, m}, 2 \times {\rm OCH}_2{\rm O}), 6.45-6.49 \; (2{\rm H, m}, 2'', 6''-{\rm H}), 6.60 \; (1{\rm H, dd}, J = 7.8, 1.7 \; {\rm Hz}, 6'-{\rm H}), 6.63 \; (1{\rm H}, d, J = 1.7 \; {\rm Hz}, 2'-{\rm H}), 6.70 \; (1{\rm H, d}, J = 7.8 \; {\rm Hz}, 5''-{\rm H}), 6.73 \; (1{\rm H, d}, J = 7.8 \; {\rm Hz}, 5'-{\rm H}), \delta_{\rm C} \; (100 \; {\rm MHz}; {\rm CDCl}_3) \\ 35.4 \; ({\rm C-7'}), \; 38.9 \; ({\rm C-7''}), 41.8 \; ({\rm C-4}), 47.6 \; ({\rm C-3}), 63.3 \; ({\rm C-6}), 83.9 \; ({\rm C-5}), 101.1, 101.2 \; (2 \times {\rm OCH}_2{\rm O}), 108.4 \; ({\rm C-5'}), 108.6 \; ({\rm C-5''}), 109.2 \; ({\rm C-2''}), 109.6 \; ({\rm C-2'}), 121.9 \; ({\rm C-6''}), 122.4 \; ({\rm C-6'}), 131.5 \; ({\rm C-1'}, 1''), 146.6 \; ({\rm C-4'}, 4''), \\ \end{array}$

148.0, 148.1 (C-3', 3"), 177.6 (C-2). IR: ν_{MAX} (film)/cm⁻¹; 3432 (br), 2922, 1760, 1503, 1490, 1444, 1247, 1038, 927, 811. HRMS (ESI⁺) Found [M + H]⁺ 385.1279; C₂₁H₂₁O₇ requires 385.1282.

 $(3R^*,4R^*)$ -3-(3',4'-Dimethoxybenzyl)-4-(3'',4''-methylenedioxybenzyl)-5-(hydroxymethyl)dihydrofuran-2(3H)one (**4ba**). Using general procedure B: Morpholine amide **35ba** (0.336 g, 0.74 mmol) in ^tBuOH/H₂O/ THF and a reaction time of 4 days. The crude product was then purified by column chromatography (1:3 hexanes, ethyl acetate) to give the title compound **4ba** (0.103 g, 34%) as a pale yellow oil.

R_f = 0.48 (1:3 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.68 (1H, t, *J* = 6.6 Hz, 6-OH), 2.33–2.42 (1H, m, 4-H), 2.48 (1H, dd, *J* = 13.7, 7.9 Hz, 7"-H_A), 2.56 (1H, dd, *J* = 13.7, 6.3 Hz, 7"-H_B), 2.68 (1H, ddd, *J* = 9.3, 6.9, 5.4 Hz, 3-H), 2.89 (1H, dd, *J* = 14.0, 6.9 Hz, 7'-H_A), 2.96 (1H, dd, *J* = 14.0, 5.4 Hz, 7'-H_B), 3.15 (1H, ddd, *J* = 12.5, 6.6, 5.2 Hz, 6-H_A), 3.52 (1H, ddd, *J* = 12.5, 6.6, 2.6 Hz, 6-H_B), 3.85 (3H, s, 3'-OCH₃), 3.86 (3H, s, 4'-OCH₃), 4.18 (1H, ddd, *J* = 7.9, 5.2, 2.6 Hz, 5-H), 5.93 (1H, d, *J* = 1.4 Hz, OCH_AO), 5.94 (1H, d, *J* = 1.4 Hz, OCH_BO), 6.44 (1H, d, *J* = 1.6 Hz, 2"-H), 6.47 (1H, dd, *J* = 7.8, 1.6 Hz, 6"-H), 6.67 (1H, d, *J* = 2.2 Hz, 2'-H), 6.68–6.72 (2H, m, 6', 5"-H), 6.79 (1H, d, *J* = 8.0 Hz, 5'-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 35.2 (C-7'), 38.8 (C-7''), 41.7 (C-4), 47.6 (C-3), 56.0 (3', 4'-OCH₃), 63.4 (C-6), 83.8 (C-5), 101.3 (OCH₂O), 108.5 (C-5'), 109.2 (C-2''), 111.3 (C-5''), 112.5 (C-2'), 121.6 (C-6'), 121.9 (C-6''), 130.3 (C-1'), 131.5 (C-1''), 146.7 (C-4''), 148.1 (C-4', 3''), 149.2 (C-3'), 177.7 (C-2). IR: v_{MAX} (film)/cm⁻¹; 3472 (br), 2933, 1760, 1516, 1490, 1242, 1157, 1028, 925, 810, 730. HRMS (ESI⁺) Found [M + Na]⁺ 423.1423; C₂₂H₂₄NaO₇ requires 423.1414.

 $(3R^*,4R^*)$ -3-(3',4',5'-Trimethoxybenzyl)-4-(3'',4''-methylenedioxybenzyl)-5-(hydroxymethyl)dihydrofuran-2(3H)one (**4bc**). Using general procedure B: Morpholine amide **35bc** (0.372 g, 0.77 mmol) in ^tBuOH/H₂O/THF and a reaction time of 4 days. The crude product was then purified by column chromatography (1:3 hexanes, ethyl acetate) to give the title compound **4bc** (0.084 g, 25%) as a pale-yellow oil.

R_f = 0.38 (1:3 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.69 (1H, t, *J* = 6.6 Hz, 6-OH), 2.36–2.45 (1H, m, 4-H), 2.53 (1H, dd, *J* = 13.8, 7.6 Hz, 7"-H_A), 2.59 (1H, dd, *J* = 13.8, 6.8 Hz, 7"-H_B), 2.70 (1H, ddd, *J* = 9.5, 6.7, 5.4 Hz, 3-H), 2.87 (1H, dd, *J* = 14.0, 6.7 Hz, 7'-H_A), 2.93 (1H, dd, *J* = 14.0, 5.4 Hz, 7'-H_B), 3.22 (1H, ddd, *J* = 12.7, 6.6, 5.0 Hz, 6-H_A), 3.58 (1H, ddd, *J* = 12.7, 6.6, 2.5 Hz, 6-H_B), 3.82 (3H, s, 4'-OCH₃), 3.84 (6H, s, 3'-OCH₃), 3.85 (3H, s, 4"-OCH₃), 4.19 (1H, ddd, *J* = 7.9, 5.0, 2.5 Hz, 5-H), 5.94 (1H, d, *J* = 1.4 Hz, OCH_BO), 6.37 (2H, s, 2'-H), 6.46 (1H, d, *J* = 1.8 Hz, 2"-H), 6.48 (1H, dd, *J* = 7.9, 1.8 Hz, 6"-H), 6.70 (1H, d, *J* = 7.9 Hz, 5"-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 36.0 (C-7'), 38.8 (C-7"), 41.9 (C-4), 47.6 (C-3), 56.3 (3'-OCH₃), 61.1 (4'-OCH₃), 63.3 (C-6), 83.8 (C-5), 101.3 (OCH₂O), 106.5 (C-2'), 108.5 (C-5"), 109.2 (C-2"), 121.9 (C-6"), 131.4 (C-1"), 133.6 (C-1'), 137.1 (C-4'), 146.7 (C-4"), 148.2 (C-3"), 153.5 (C-3'), 177.7 (C-2). IR: $\nu_{\rm MAX}$ (film)/cm⁻¹; 3475 (br), 2941, 1760, 1591, 1490, 1445, 1244, 1127, 1036, 926. HRMS (ESI⁺) Found [M + Na]⁺ 453.1519; C₂₃H₂₆NaO₈ requires 453.1520.

 $(3R^*,4R^*)$ -3-(3'-Methoxy-4'-benzyloxybenzyl)-4-(3'',4''-methylenedioxybenzyl)-5-(hydroxymethyl)dihydrofuran-2(3H)-one (**4bd**). Using general procedure B: Morpholine amide **35bd** (0.405 g, 0.77 mmol) in ^tBuOH/H₂O/THF and a reaction time of 5 days. The crude product was then purified by column chromatography (1:3 hexanes, ethyl acetate) to give the title compound **4bd** (0.205 g, 56%) as a pale-yellow oil.

R_f = 0.58 (1:3 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.64 (1H, t, *J* = 6.6 Hz, 6-OH), 2.31–2.40 (1H, m, 4-H), 2.46 (1H, dd, *J* = 13.7, 7.9 Hz, 7''-H_A), 2.53 (1H, dd, *J* = 13.7, 6.3 Hz, 7''-H_B), 2.67 (1H, ddd, *J* = 9.2, 7.2, 5.3 Hz, 3-H), 2.87 (1H, dd, *J* = 14.0, 7.2 Hz, 7'-H_A), 2.95 (1H, dd, *J* = 14.0, 5.3 Hz, 7'-H_B), 3.13 (1H, ddd, *J* = 12.6, 6.6, 5.1 Hz, 6-H_A), 3.48 (1H, ddd, *J* = 12.6, 6.6, 2.6 Hz, 6-H_B), 3.86 (3H, s, 3'-OCH₃), 4.17 (1H, ddd, *J* = 7.8, 5.1, 2.6 Hz, 5-H), 5.13 (2H, s, 7'''-H), 5.93 (1H, d, *J* = 1.4 Hz, OCH_AO), 5.94 (1H, d, *J* = 1.4 Hz, OCH_BO), 6.43 (1H, d, *J* = 1.6 Hz, 2''-H), 6.45 (1H, dd, *J* = 7.9, 1.6 Hz, 6''-H), 6.64 (1H, dd, *J* = 8.2, 2.0 Hz, 6'-H), 6.68–6.70 (2H, m, 2', 5''-H), 6.81 (1H, d, *J* = 8.2 Hz, 5'-H), 7.27–7.30 (1H, m, 4'''-H), 7.32–7.36 (2H, m, 3'''-H), 7.40–7.44 (2H, m, 2'''-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 35.3 (C-7'), 38.8 (C-7''), 41.8 (C-4), 47.6 (C-3), 56.1 (3'-OCH₃), 63.4 (C-6), 71.3 (C-7'''), 83.8 (C-5), 101.3 (OCH₂O), 108.5 (C-5''), 109.2 (C-2''), 113.0 (C-2'), 114.4 (C-5'), 121.5 (C-6'), 121.9 (C-6''), 127.5 (C-2'''), 128.0 (C-4'''), 128.7 (C-3'''), 131.0 (C-1'), 131.5 (C-1''), 137.3 (C-1'''), 146.7 (C-4''), 147.2 (C-4'), 148.1 (C-3''), 150.0 (C-3'), 177.7 (C-2).

IR: ν_{MAX} (film)/cm⁻¹; 3471 (br), 2940, 1743, 1504, 1490, 1366, 1230, 1036, 926, 735. HRMS (ESI⁺) Found [M + Na]⁺ 499.1729; C₂₈H₂₈NaO₇ requires 499.1727.

 $(3R^*,4R^*,5S^*)-4-(3'',4''-Methylenedioxybenzyl)-3-(4'-hydroxy-3'-methoxybenzyl)-5-(hydroxymethyl)$ dihydrofuran-2(3H)-one (**4be**). Using general procedure F: Benzyl ether **4bd** (0.02 g, 0.04 mmol) and a reaction time of 1 h. The crude product was then purified by column chromatography (1:3 hexanes, ethyl acetate) to give the title compound **4be** (0.017 g, quant.) as a colourless oil. R_f = 0.52 (1:3 hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.74 (1H, br, 6-OH), 2.33–2.42 (1H, m, 4-H), 2.48 (1H, dd, *J* = 13.7, 8.0 Hz, 7''-H_A), 2.57 (1H, dd, *J* = 13.7, 6.2 Hz, 7''-H_B), 2.67 (1H, ddd, *J* = 9.4, 6.9, 5.5 Hz, 3-H), 2.88 (1H, dd, *J* = 14.0, 6.9 Hz, 7'-H_A), 2.94 (1H, dd, *J* = 14.0, 5.5 Hz, 7'-H_B), 3.15 (1H, br d, *J* = 12.6 Hz, 6-H_A), 3.52 (1H, br d, *J* = 12.6 Hz, 6-H_B), 3.86 (3H, s, 3'-OCH₃), 4.18 (1H, ddd, *J* = 8.0, 5.0, 2.5 Hz, 5-H), 5.54 (1H, s, 4'-OH), 5.93 (1H, d, *J* = 1.4 Hz, OCH_AO), 5.94 (1H, dd, *J* = 8.0, 1.9 Hz, 6'-H), 6.67 (1H, d, *J* = 1.9 Hz, 2''-H), 6.47 (1H, dd, *J* = 7.7, 1.9 Hz, 6''-H), 6.63 (1H, dd, *J* = 8.0, 1.9 Hz, 6'-H), 6.67 (1H, d, *J* = 1.9 Hz, 2''-H), 6.70 (1H, d, *J* = 7.7 Hz, 5''-H), 6.84 (1H, d, *J* = 8.0 Hz, 5'-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 35.3 (C-7'), 38.8 (C-7''), 41.7 (C-4), 47.7 (C-3), 56.1 (3'-OCH₃), 63.4 (C-6), 83.9 (C-5), 101.3 (OCH₂O), 108.6 (C-5''), 109.2 (C-2''), 111.8 (C-2'), 114.5 (C-5'), 121.9 (C-6''), 122.3 (C-6'), 129.6 (C-1'), 131.5 (C-1''), 144.7 (C-4'), 146.7 (C-3'), 146.8 (C-4''), 148.1 (C-3''), 177.8 (C-2). IR: v_{MAX} (film)/cm⁻¹; 3449 (br), 2933, 1754, 1516, 1490, 1246, 1036, 926, 812. HRMS (ESI⁺) Found [M + Na]⁺ 409.1246; C₂₁H₂₂NaO₇ requires 409.1258.

(±)-Arcitin (1aa). Using general procedure C: Lactone 4aa (0.16 g, 0.39 mmol) and a reaction time of 2 h to give triol **38aa** (0.17 g, quant.) as a colourless oil. Then using general procedure D: Triol **38aa** (0.16 g, 0.37 mmol) and a reaction time of 2.5 h to give lactol **39aa** (0.14 g, 97%) which was used without further purification. Then using general procedure E: Lactol 39aa (0.054 g, 0.14 mmol) and a reaction time of 3 h. The crude product was purified by column chromatography (1:1, hexanes, ethyl acetate) to give the title compound **1aa** (0.05 g, 88%) as a pale yellow amorphous solid. $R_f = 0.45$ (19:1, CH₂Cl₂, methanol). Melting point: 114–116 °C [lit. [49] 113 °C]. δ_H (400 MHz; CDCl₃) 2.45–2.68 (4H, m, 8, 7', 8'-H), 2.92 (1H, dd, J = 14.3, 6.8 Hz, 7-H_A), 2.97 (1H, dd, J = 14.3, 5.5 Hz, 7-H_B), 3.82 (3H, s, 3'-OCH₃), 3.83 (3H, s, 3-OCH₃), 3.85–3.90 (7H, m, 4, 4'-OCH₃, 9'-H_A), 4.13 (1H, t, J = 7.0 Hz, 9'-H_B), 6.49 (1H, d, *J* = 1.9 Hz, 2'-H), 6.55 (1H, dd, *J* = 8.1, 1.9 Hz, 6'-H), 6.66 (1H, dd, *J* = 8.1, 1.9 Hz, 6-H), 6.69 (1H, d, J = 1.9 Hz, 2-H), 6.75 (1H, d, J = 8.1 Hz, 5-H), 6.77 (1H, d, J = 8.1 Hz, 5'-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 34.5 (C-7), 38.2 (C-7'), 41.1 (C-8'), 46.6 (C-8), 55.8, 55.9 (3, 4, 3', 4'-OCH₃), 71.2 (C-9'), 111.1 (C-5), 111.4 (C-5'), 111.9 (C-2'), 112.4 (C-2), 120.6 (C-6'), 121.4 (C-6), 130.2 (C-1), 130.5 (C-1'), 147.9 (C-4'), 148.0 (C-4), 149.1 (C-3, 3'), 178.7 (C-9). IR: v_{MAX} (film)/cm⁻¹; 2956, 1753, 1588, 1513, 1257, 1236, 1153, 1137, 1019, 825, 764. HRMS (ESI⁺) Found [M + H]⁺ 387.1806; C₂₂H₂₇O₆ requires 387.1802. Values are in agreement with literature data [50].

(±)-*Bursehernin* (1a). Using general procedure C: Lactone 4ab (0.114 g, 0.28 mmol) and a reaction time of 30 min to give triol 38ab (0.111 g, 97%) as a cloudy oil. Then using general procedure D: Triol 38ab (0.111 g, 0.27 mmol) and a reaction time of 1 h to give lactol 39ab (0.093 g, 91%) which was used without further purification. Then using general procedure E: Lactol 39ab (0.093 g, 0.25 mmol) and a reaction time of 2 h. The crude product was purified by column chromatography (1:1, hexanes, ethyl acetate) to give the title compound 1ab (0.06 g, 65%) as a pale-yellow oil. $R_f = 0.66$ (19:1, CH_2Cl_2 , methanol). δ_H (400 MHz; CDCl₃) 2.41–2.62 (4H, m, 8, 7', 8'-H), 2.88 (1H, dd, J = 14.0, 6.9 Hz, 7-H_A), 2.96 (1H, dd, J = 14.0, 5.1 Hz, 7-H_B), 3.82 (3H, s, 3-OCH₃), 3.83–3.86 (4H, m, 4-OCH₃, 9'-H_A), 4.10 (1H, dd, J = 9.1, 6.9 Hz, 9'-H_B), 5.91 (1H, d, J = 1.4 Hz, OCH_AO), 5.92 (1H, d, J = 1.4 Hz, OCH_BO), 6.42 (1H, d, J = 1.5 Hz, 2'-H), 6.44 (1H, dd, J = 7.9, 1.5 Hz, 6'-H), 6.66 (1H, d, J = 1.9 Hz, 2-H), 6.67–6.70 (2H, m, 6, 5'-H), 6.78 (1H, d, J = 8.0 Hz, 5-H). δ_C (100 MHz; CDCl₃) 34.7 (C-7), 38.4 (C-7'), 41.2 (C-8'), 46.6 (C-8), 55.9 (3, 4-OCH₃), 71.2 (C-9'), 101.1 (OCH₂O), 108.4 (C-5'), 108.8 (C-2'), 111.2 (C-5), 112.3 (C-2), 121.4 (C-6), 121.6 (C-6'), 130.2 (C-1), 131.7 (C-1'), 146.4 (C-4'), 148.0 (C-3'), 2.38, 2.3

(±)-4-O-Methyl traxillagenin (1ac). Using general procedure C: Lactone 4ac (0.119 g, 0.27 mmol) and a reaction time of 45 min to give triol 38ac (0.11 g, 90%) as a cloudy oil. The using general procedure D: Triol **38ac** (0.11 g, 0.24 mmol) and a reaction time of 15 min. The crude product was purified by column chromatography (1:2 hexanes, ethyl acetate) to give lactol **39ac** (0.06 g, 60%) as a colourless oil. Then using general procedure E: Lactol **39ac** (0.06 g, 0.15 mmol) and a reaction time of 3 h. The crude product purified by column chromatography (1:1, hexanes, ethyl acetate) to give the title compound **1ac** (0.044 g, 73%) as a white solid. $R_f = 0.61$ (19:1, CH₂Cl₂, methanol). Melting point: 126 °C. δ_H (400 MHz; CDCl₃) 2.44–2.66 (4H, m, 8, 7', 8'-H), 2.91 (1H, dd, J = 14.1, 6.6 Hz, 7-H_A), 2.98 (1H, dd, J = 14.1, 5.4 Hz, 7-H_B), 3.79 (6H, s, 3'-OCH₃), 3.80 (6H, s, 4'-OCH₃), 3.83 (3H, s, 3-OCH₃), 3.84 (3H, s, 4-OCH₃), 3.87 (1H, dd, J = 9.2, 7.3 Hz, 9'-H_A), 4.14 (1H, dd, J = 9.2, 7.0 Hz, 9'-H_B), 6.19 (2H, s, 2'-H), 6.63 $(1H, dd, J = 8.0, 2.0 Hz, 6-H), 6.70 (1H, d, J = 2.0 Hz, 2-H), 6.75 (1H, d, J = 8.0 Hz, 5-H). \delta_{C} (100 MHz;$ CDCl₃) 34.6 (C-7), 39.0 (C-7'), 41.2 (C-8'), 46.7 (C-8), 56.0 (3, 4-OCH₃), 56.2 (3'-OCH₃), 60.9 (4'-OCH₃), 71.3 (C-9'), 105.7 (C-2'), 111.2 (C-5), 112.6 (C-2), 121.4 (C-6), 130.3 (C-1), 133.8 (C-1'), 137.0 (C-4'), 148.1 (C-4), 149.2 (C-3), 153.5 (C-3'), 178.7 (C-9). IR: ν_{MAX} (film)/cm⁻¹; 2938, 1764, 1590, 1509, 1460, 1237, 1123, 1014, 731. HRMS (ESI⁺) Found [M + Na]⁺ 439.1716; C₂₃H₂₈NaO₇ requires 439.1727. Values are in agreement with literature data [52].

 (\pm) -4'-O-Benzyl buplerol (1ad). Using general procedure C: Lactone 4ad (0.505 g, 1.02 mmol) and a reaction time of 3 h to give the triol **38ad** (0.472 g, 93%) as a cloudy oil. Then using general procedure D: Triol 38ad (0.472 g, 0.95 mmol) and a reaction time of 30 min to give lactol 39ad (0.416 g, 94%) as a white solid which was used without further purification. Then using general procedure E: Lactol 39ad (0.416 g, 0.90 mmol) and a reaction time of 1.5 h. The crude product was purified by column chromatography (1:1, hexanes, ethyl acetate) to give the title compound **1ad** (0.374 g, 90%) as a pale-yellow oil. $R_f = 0.52$ (1:1, hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 2.42–2.66 (4H, m, 8, 7', 8'-H), 2.91 (1H, dd, J = 14.1, 6.2 Hz, 7-H_A), 2.95 (1H, dd, J = 14.1, 5.7 Hz, 7-H_B), 3.827, 3.829 (6H, 2 × s, 3, 3'-OCH₃), 3.85 (3H, s, 4-OCH₃), 3.83–3.88 (1H, m, 9'-H_A), 4.11 (1H, dd, J = 8.7, 7.0 Hz, 9'-H_B), 5.12 (2H, s, Ph-CH₂), 6.48 (1H, dd, J = 8.0, 2.0 Hz, 6'-H), 6.51 (1H, d, J = 2.0 Hz, 2'-H), 6.64 (1H, dd, J = 8.2, 2.0 Hz, 6-H), 6.68 (1H, d, J = 2.0 Hz, 2-H), 6.76 (1H, d, J = 8.2 Hz, 5-H), 6.77 (1H, d, J = 8.0 Hz, 5'-H), 7.27–7.32 (1H, m, Ph-*p*-H), 7.33–7.38 (2H, m, Ph-*m*-H), 7.40–7.44 (2H, m, Ph-*o*-H). δ_C (100 MHz; CDCl₃) 34.6 (C-7), 38.3 (C-7'), 41.2 (C-8'), 46.7 (C-8), 56.0 (3, 3'-OCH₃), 56.1 (4-OCH₃), 71.3, 71.4 (C-9', Ph-CH₂), 111.3 (C-5), 112.5 (C-2), 112.6 (C-5'), 114.5 (C-5'), 120.7 (C-6'), 121.5 (C-6), 127.4 (Ph-o-C), 128.0 (Ph-p-C), 128.7 (Ph-m-C), 130.3 (C-1), 131.3 (C-1'), 137.3 (Ph-i-C), 147.2 (C-4'), 148.1 (C-4), 149.2 (C-3), 149.9 (C-3'), 178.8 (C-9). IR: v_{MAX} (film)/cm⁻¹; 2935, 1763, 1512, 1260, 1233, 1140, 1014, 736, 697. HRMS (ESI⁺) Found [M + Na]⁺ 485.1934; C₂₈H₃₀NaO₆ requires 485.1935.

(±)-*Buplerol* (**1ae**). Using general procedure F: Lactone **1ad** (0.336 g, 0.73 mmol) and a reaction time of 3.5 h to give the title compound **1ae** (0.271 g, quant.) as a white solid. $R_f = 0.33$ (1:1, hexanes, ethyl acetate). Melting point: 101–103 °C. δ_H (400 MHz; CDCl₃) 2.42–2.66 (4H, m, 8, 7', 8'-H), 2.90 (1H, dd, J = 14.1, 6.8 Hz, 7-H_A), 2.97 (1H, dd, J = 14.1, 5.3 Hz, 7-H_B), 3.81 (3H, s, 3-OCH₃), 3.83 (3H, s, 3'-OCH₃), 3.86 (4H, m, 4-OCH₃), 3.87 (1H, dd, J = 8.9, 7.1 Hz, 9'-H_A), 4.13 (1H, dd, J = 9.3, 7.1 Hz, 9'-H_B), 5.51 (1H, s, 4'-OH), 6.43 (1H, d, J = 1.9 Hz, 2'-H), 6.52 (1H, dd, J = 8.0, 1.9 Hz, 6'-H), 6.64–6.67 (2H, m, 2, 6-H), 6.77 (1H, d, J = 8.6 Hz, 5-H), 6.80 (1H, d, J = 8.0 Hz, 5'-H). δ_C (100 MHz; CDCl₃) 34.7 (C-7), 38.5 (C-7'), 41.3 (C-8'), 46.7 (C-8), 55.9, 56.0 (3, 3', 4-OCH₃), 71.4 (C-9'), 111.1, 111.2 (C-5, 5'), 112.5 (C-2), 114.6 (C-2'), 121.5 (C-6, 6'), 129.9 (C-1'), 130.4 (C-1), 144.6 (C-4'), 146.7 (C-3'), 148.1 (C-4), 149.2 (C-3), 178.9 (C-9). IR: ν_{MAX} (film)/cm⁻¹; 3417, 2938, 1760, 1513, 1236, 1148, 1023, 812, 795. HRMS (ESI⁺) Found [M + Na]⁺ 395.1462; C₂₁H₂₄NaO₆ requires 395.1465. Values are in agreement with literature data [53].

 (\pm) -*Kusunokinin* (**1ba**). Using general procedure C: Lactone **4ba** (0.082 g, 0.20 mmol) and a reaction time of 1 h to give the triol **38ba** (0.083 g, quant.) as a cloudy oil. Then using general procedure D: Triol **38ba** (0.083 g, 0.20 mmol) and a reaction time of 15 min to give lactol **39ba** (0.064 g, 84%) which was used without further purification. Then using general procedure E: Lactol **39ba** (0.056 g, 0.15 mmol) and a reaction time of 1 h. The crude product was purified by column chromatography (2:1, hexanes,

ethyl acetate) to give the title compound **1ba** (0.051 g, 91%) as a colourless oil. $R_f = 0.48$ (1:1, hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 2.44–2.65 (4H, m, 8, 7', 8'-H), 2.84 (1H, dd, J = 14.1, 7.0 Hz, 7-H_A), 2.95 (1H, dd, J = 14.1, 5.1 Hz, 7-H_B), 3.82 (3H, s, 3'-OCH₃), 3.85 (3H, s, 4'-OCH₃), 3.87 (1H, dd, J = 9.2, 7.2 Hz, 9'-H_A), 4.14 (1H, dd, J = 9.2, 7.0 Hz, 9'-H_B), 5.92 (1H, d, J = 1.4 Hz, OCH_AO), 5.93 (1H, d, J = 1.4 Hz, OCH_BO), 6.48 (1H, d, J = 2.0 Hz, 2'-H), 6.55–6.60 (3H, m, 2, 6, 6'-H), 6.71 (1H, d, J = 7.7 Hz, 5-H), 6.76 (1H, d, J = 8.2 Hz, 5'-H). δ_C (100 MHz; CDCl₃) 34.9 (C-7), 38.4 (C-7'), 41.3 (C-8'), 46.6 (C-8), 55.9 (3'-OCH₃), 56.0 (4'-OCH₃), 71.4 (C-9'), 101.1 (OCH₂O), 108.3 (C-5), 109.6 (C-2), 111.4 (C-5'), 111.8 (C-2'), 120.8 (C-6'), 122.4 (C-6), 130.6 (C-1'), 131.5 (C-1), 146.6 (C-4), 148.0 (C-3, 4'), 149.2 (C-3'), 178.6 (C-9). IR: ν_{MAX} (film)/cm⁻¹; 2908, 1764, 1515, 1489, 1442, 1242, 1024, 912, 809, 729. HRMS (ESI⁺) Found [M + Na]⁺ 393.1301; C₂₁H₂₂NaO₆ requires 393.1309. Values are in agreement with literature data [50].

(±)-*Hinokinin* (**1bb**). Using general procedure C: Lactone **4bb** (0.12 g, 0.31 mmol) and a reaction time of 30 min to give the triol **38bb** (0.12 g, quant.) as a cloudy oil. Then using general procedure D: Triol **38bb** (0.121 g, 0.31 mmol) and a reaction time of 10 min to give lactol **39bb** (0.096 g, 86%) which was used without further purification. Then using general procedure E: Lactol **39bb** (0.089 g, 0.25 mmol) and a reaction time of 1 h. The crude product was purified by column chromatography (1:1, hexanes, ethyl acetate) to give the title compound **1bb** (0.08 g, 90%) as a pale-yellow oil. $R_f = 0.73$ (1:1, hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 2.41–2.62 (4H, m, 8, 7', 8'-H), 2.83 (1H, dd, *J* = 14.1, 7.2 Hz, 7-H_A), 2.98 (1H, dd, *J* = 14.1, 5.0 Hz, 7-H_B), 3.85 (1H, dd, *J* = 9.2, 7.1 Hz, 9'-H_A), 4.12 (1H, dd, *J* = 9.2, 6.9 Hz, 9'-H_B), 5.91–5.94 (4H, m, 2 × OCH₂O), 6.44–6.47 (2H, m, 2', 6'-H), 6.59 (1H, dd, *J* = 7.9, 1.8 Hz, 6-H), 6.62 (1H, d, *J* = 1.8 Hz, 2-H), 6.69 (1H, d, *J* = 8.4 Hz, 5'-H), 6.72 (1H, d, *J* = 7.9 Hz, 5-H). δ_C (100 MHz; CDCl₃) 34.9 (C-7), 38.4 (C-7'), 41.4 (C-8'), 46.6 (C-8), 71.2 (C-9'), 101.1 (2 × OCH₂O), 108.4 (C-4'), 148.0 (C-3, 3'), 178.5 (C-9). IR: ν_{MAX} (film)/cm⁻¹; 2901, 1764, 1488, 1441, 1242, 1015, 924, 808, 728. HRMS (ESI⁺) Found [M + Na]⁺ 377.0986; C₂₀H₁₈NaO₆ requires 377.0996. Values are in agreement with literature data [54].

(\pm)-*Isoyatein* (**1bc**). Using general procedure C: Lactone **4bc** (0.076 g, 0.18 mmol) and a reaction time of 1 h to give the triol **38bc** (0.077 g, >99%) as a cloudy oil. Then using general procedure D: Triol **38bc** (0.077 g, 0.18 mmol) and a reaction time of 1 h to give lactol **39bc** (0.057 g, 80%) which was used without further purification. Then using general procedure E: Lactol **39bc** (0.05 g, 0.12 mmol) and a reaction time of 3 h. The crude product was purified by column chromatography (1:1, hexanes, ethyl acetate) to give the title compound **1bc** (0.8 mg, 16%) as a pale-yellow oil. R_f = 0.55 (1:1, hexanes, ethyl acetate). $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.46–2.64 (4H, m, 8, 7', 8'-H), 2.86 (1H, dd, *J* = 14.1, 7.0 Hz, 7-H_A), 2.98 (1H, dd, *J* = 14.1, 5.1 Hz, 7-H_B), 3.81 (6H, s, 3'-OCH₃), 3.82 (3H, s, 4'-OCH₃), 3.89 (1H, dd, *J* = 9.2, 7.0 Hz, 9'-H_A), 4.19 (1H, dd, *J* = 9.2, 6.8 Hz, 9'-H_B), 5.93 (1H, d, *J* = 1.5 Hz, OCH_AO), 5.94 (1H, d, *J* = 1.5 Hz, OCH_BO), 6.20 (2H, s, 2'-H), 6.58 (1H, dd, *J* = 7.9, 1.8 Hz, 6-H), 6.61 (1H, d, *J* = 1.8 Hz, 2-H), 6.71 (1H, d, *J* = 7.9 Hz, 5-H). $\delta_{\rm C}$ (100 MHz; CDCl₃) 34.9 (C-7), 39.2 (C-7'), 41.4 (C-8'), 46.6 (C-8), 56.2 (3'-OCH₃), 61.0 (4'-OCH₃), 71.4 (C-9'), 101.2 (OCH₂O), 105.7 (C-2'), 108.3 (C-5), 109.6 (C-2), 122.4 (C-6), 131.5 (C-1), 133.8 (C-1'), 137.0 (C-4'), 146.7 (C-4), 148.1 (C-3), 153.5 (C-3'), 178.5 (C-9). IR: v_{MAX} (film)/cm⁻¹; 2938, 1763, 1590, 1489, 1443, 1241, 1122, 1011, 927, 813, 732. HRMS (ESI⁺) Found [M + Na]⁺ 423.1400; C₂₂H₂₄NaO₇ requires 423.1414. Values are in agreement with literature data [55].

(±)-4'-O-Benzyl haplomyrfolin (1bd). Using general procedure C: Lactone 4bd (0.18 g, 0.38 mmol) and a reaction time of 20 min to give the triol **38bd** (0.18 g, quant.) as a cloudy oil. Then using general procedure D: Triol **38bd** (0.18 g, 0.38 mmol) and a reaction time of 20 min to give lactol **39bd** (0.13 g, 76%) as a white solid which was used without further purification. Then using general procedure E: Lactol **39bd** (0.13 g, 0.28 mmol) and a reaction time of 2 h. The crude product was purified by column chromatography (3:1, hexanes, ethyl acetate) to give the title compound **1bd** (0.12 g, 94%) as a colourless oil. $R_f = 0.65$ (1:1, hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 2.43–2.64 (4H, m, 8, 7', 8'-H), 2.84 (1H, dd, J = 14.1, 7.0 Hz, 7-H_A), 2.94 (1H, dd, J = 14.1, 5.1 Hz, 7-H_B), 3.83 (3H, s, 3'-OCH₃), 3.87 (1H, dd, J = 9.1, 7.2 Hz, 9'-H_A), 4.14 (1H, dd, J = 9.1, 7.0 Hz, 9'-H_B), 5.12 (2H, s, 7''-H), 5.91 (1H, d,

 $J = 1.4 \text{ Hz}, \text{OCH}_{A}\text{O}, 5.93 (1\text{H}, \text{d}, J = 1.4 \text{ Hz}, \text{OCH}_{B}\text{O}), 6.49-6.52 (2\text{H}, \text{m}, 2', 6'-\text{H}), 6.57 (1\text{H}, \text{dd}, J = 7.9, 1.8 \text{ Hz}, 6-\text{H}), 6.59 (1\text{H}, \text{d}, J = 1.8 \text{ Hz}, 2-\text{H}), 6.70 (1\text{H}, \text{d}, J = 7.9 \text{ Hz}, 5-\text{H}), 6.78 (1\text{H}, \text{d}, J = 8.5 \text{ Hz}, 5'-\text{H}), 7.27-7.32 (1\text{H}, \text{m}, 4''-\text{H}), 7.33-7.38 (2\text{H}, \text{m}, 3''-\text{H}), 7.41-7.45 (2\text{H}, \text{m}, 2''-\text{H}). \delta_{C} (100 \text{ MHz}; \text{CDCl}_{3}) 34.8 (C-7), 38.4 (C-7'), 41.3 (C-8'), 46.5 (C-8), 56.0 (3'-\text{OCH}_{3}), 71.2 (C-7''), 71.3 (C-9'), 101.1 (\text{OCH}_{2}\text{O}), 108.3 (C-5), 109.6 (C-2), 112.4 (C-2'), 114.4 (C-5'), 120.7 (C-6'), 122.4 (C-6), 127.4 (C-2''), 128.0 (C-4''), 128.6 (C-3''), 131.2 (C-1), 131.5 (C-1'), 137.2 (C-1''), 146.6 (C-4), 147.1 (C-4'), 148.0 (C-3), 149.9 (C-3'), 178.6 (C-9). IR: <math>\nu_{MAX}$ (film)/cm⁻¹; 2907, 1765, 1504, 1489, 1443, 1244, 1140, 1034, 911, 809, 730. HRMS (ESI⁺) Found [M + Na]⁺ 469.1612; C₂₇H₂₆NaO₆ requires 469.1622.

(±)-*Haplomyrfolin* (**1be**). Using general procedure F: Lactone **1bd** (0.119 g, 0.27 mmol) and a reaction time of 1.5 h. The crude product was purified by column chromatography (1:1 hexanes, ethyl acetate) to give the title compound **1be** (0.086 g, 91%) as a colourless oil. $R_f = 0.47$ (1:1 hexanes, ethyl acetate). δ_H (400 MHz; CDCl₃) 2.43–2.63 (4H, m, 8, 7', 8'-H), 2.84 (1H, dd, *J* = 14.1, 7.0 Hz, 7-H_A), 2.95 (1H, dd, *J* = 14.1, 5.2 Hz, 7-H_B), 3.83 (3H, s, 3'-OCH₃), 3.86 (1H, dd, *J* = 9.1, 7.2 Hz, 9'-H_A), 4.13 (1H, dd, *J* = 9.1, 7.0 Hz, 9'-H_B), 5.63 (1H, s, 4'-OH), 5.91 (1H, d, *J* = 1.4 Hz, OCH_AO), 5.92 (1H, dd, *J* = 1.4 Hz, OCH_BO), 6.46 (1H, d, *J* = 1.9 Hz, 2'-H), 6.51 (1H, dd, *J* = 8.0, 1.9 Hz, 6'-H), 6.58 (1H, dd, *J* = 7.8, 1.7 Hz, 6-H), 6.60 (1H, d, *J* = 1.7 Hz, 2-H), 6.70 (1H, d, *J* = 7.8 Hz, 5-H), 6.80 (1H, d, *J* = 8.0 Hz, 5'-H). δ_C (100 MHz; CDCl₃) 34.8 (C-7), 38.3 (C-7'), 41.4 (C-8'), 46.5 (C-8), 55.9 (3'-OCH₃), 71.3 (C-9'), 101.1 (OCH₂O), 108.3 (C-5), 109.6 (C-2), 111.2 (C-2'), 114.6 (C-5'), 121.4 (C-6'), 122.4 (C-6), 129.9 (C-1'), 131.5 (C-1), 144.5 (C-4'), 146.5 (C-4), 146.7 (C-3'), 147.9 (C-3), 178.7 (C-9). IR: ν_{MAX} (film)/cm⁻¹; 3468, 2921, 1762, 1515, 1489, 1443, 1243, 1035, 907, 725. HRMS (ESI⁺) Found [M + Na]⁺ 379.1151; C₂₀H₂₀NaO₆ requires 379.1152. Values are in agreement with literature data [56].

4. Biological Assay Methods

4.1. Cell Culture

Jurkat E61 cells (ECACC) were maintained at 37 °C in RMPI media (Lonza) supplemented with 10% Foetal Bovine Serum (FBS) (Lonza) (10% RPMI) in a humidified environment of 5% CO₂ in air. Cells were routinely passaged to maintain a cell density of between 1×10^5 and 1×10^6 /mL.

4.2. Drug Treatments

Lignans were diluted to stock concentrations of 30 mM in DMSO and further diluted to the working concentration in 10% RPMI. The DMSO diluted to the appropriate concentration was used as the vehicle-control. Cells were seeded at the relevant density per well depending upon the assay to be performed, in 100 μ L volume of fresh 10% RPMI. Trypan blue exclusion method was used to assess viability prior to experiments and cell viability was always >95%. Lignans were added at 100 μ L/well to the relevant wells. Cells were incubated at 37 °C in a humidified environment of 5% CO₂ in air for the indicated times. Dead cell controls were included in subsequent viability assays by treating cells with 50 μ L/well EtOH (final concentration 50%) for 48 h. Apoptotic controls were included in subsequent apoptosis assays by exposing cells to a heat shock at 43 °C for 2 h. Positive controls for cell cycle analysis were included by treating cells with 0.5 μ M camptothecin for 4 h to induce cell cycle arrest.

4.3. MTS Assay

Following treatments at a cell density of 1×10^5 cells/well, the samples were centrifuged at 500 g for 5 min and the supernatant was removed. A 100 µL/well volume of fresh 10 % RPMI was added. A 20 µL volume of MTS solution (Promega, G1112) was added to each well and the plate was incubated in the dark for 1 h at 37 °C. The absorbance was detected at 490 nm on a Synergy HT plate reader.

for 5 min and the supernatant was removed. Cells were washed in 500 µL DPBS before addition of 100 μ L of 1 \times Annexin V binding buffer (BD Biosciences). A 5 μ L volume of FITC-conjugated Annexin V (BD Biosciences) and 10 µL Propidium Iodide (BD Biosciences) was added and the cells were incubated in the dark for 20 min. Samples were diluted by addition of 400 μ L 1 \times Annexin V binding buffer before immediate analysis on an Accuri C6 Flow Cytometer (Becton Dickinson, Oxford, UK).

4.5. Cell Cycle Analysis

Following treatments at a cell density of 5×10^6 /well, cells were centrifuged at 500 g for 5 min and the supernatant was removed. The remaining cell pellet was vortexed while simultaneously adding 500 μ L of 70% ethanol dropwise, fixing the cells and minimising clumping. The samples were incubated at 4 °C for 30 min, and then centrifuged at 1000 g for 5 min. The supernatant was discarded, and the pellet was re-suspended in 500 μ L DPBS. The samples were centrifuged again at 1000 g for 5 min, and the supernatant was removed a final time. The pellet was resuspended in 50 μ L RNase A (100 µg/mL stock; Roche, UK) and 200 µL PI (50 µg/mL stock; Sigma, UK). The samples were analyzed on an Accuri C6 flow cytometer (Becton Dickinson) and data was modelled and interpreted using ModFit Analysis Software, version 5.0 (Verity Software House).

Supplementary Materials: The following are available online.

Author Contributions: Conceptualization, D.B. and N.C.D.-H.; Methodology, S.J.D., M.E.-M., S.T. and T.W.; Formal Analysis, L.I.P., D.B. and N.C.D.-H.; Investigation, S.J.D., M.E.-M., S.T. and T.W.; Writing-Original Draft Preparation, S.J.D. and L.I.P.; Writing-Review & Editing, D.B., L.I.P., S.J.D., K.A.W. and N.C.D.-H.; Supervision, D.B., K.A.W. and N.C.D.-H.; Project Administration, D.B.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Jang, Y.P.; Kim, S.R.; Kim, Y.C. Neuroprotective dibenzylbutyrolactone lignans of Torreya nucifera. 1. Planta Med. 2001, 67, 470-472. [CrossRef] [PubMed]
- 2. Marcotullio, M.C.; Pelosi, A.; Curini, M. Hinokinin, an emerging bioactive lignan. Molecules 2014, 19, 14862-14878. [CrossRef] [PubMed]
- 3. Su, S.; Cheng, X.; Wink, M. Natural lignans from Arctium lappa modulate P-glycoprotein efflux function in multidrug resistant cancer cells. Phytomedicine 2015, 22, 301-307. [CrossRef] [PubMed]
- 4. Yang, Y.-N.; Huang, X.-Y.; Feng, Z.-M.; Jiang, J.-S.; Zhang, P.-C. New butyrolactone type lignans from Arctii fructus and their anti-inflammatory activities. J. Agric. Food Chem. 2015, 63, 7958–7966. [CrossRef] [PubMed]
- 5. Su, S.; Wink, M. Natural lignans from Arctium lappa as antiaging agents in Caenorhabditis elegans. Phytochemistry 2015, 117, 340–350. [CrossRef] [PubMed]
- 6. Chang, H.; Wang, Y.; Gao, X.; Song, Z.; Awale, S.; Han, N.; Liu, Z.; Yina, J. Lignans from the root of Wikstroemia indica and their cytotoxic activity against PANC-1 human pancreatic cancer cells. Fitoterpia **2017**, *121*, *31–37*. [CrossRef] [PubMed]
- 7. Maxwell, T.; Chun, S.Y.; Lee, K.S.; Kim, S.; Nam, K.S. The anti-metastatic effects of the phytoestrogen arctigenin on human breast cancer cell lines regardless of the status of ER expression. Int. J. Oncol. 2017, 50, 727-735. [CrossRef] [PubMed]
- Maxwell, T.; Lee, K.S.; Kim, S.; Nam, K.S. Arctigenin inhibits the activation of the mTOR pathway, resulting 8. in autophagic cell death and decreased ER expression in ER-positive human breast cancer cells. Int. J. Oncol. 2018, 52, 1339–1349. [CrossRef] [PubMed]
- 9. Huang, Q.; Qin, S.; Yuan, X.; Zhang, L.; Ji, J.; Liu, X.; Ma, W.; Zhang, Y.; Liu, P.; Sun, Z.; et al. Arctigenin inhibits triple-negative breast cancers by targeting CIP2A to reactivate protein phosphatase 2A. Oncol. Rep. 2017, 38, 598-606. [CrossRef] [PubMed]

- 10. Li, Q.C.; Liang, Y.; Tian, Y.; Hu, G.R. Arctigenin induces apoptosis in colon cancer cells through ROS/p38MAPK pathway. *J. Buon.* **2016**, *21*, 87–94. [PubMed]
- Han, Y.H.; Kee, J.Y.; Kim, D.S.; Mun, J.G.; Jeong, M.Y.; Park, S.H.; Choi, B.M.; Park, S.J.; Kim, H.J.; Um, J.Y.; et al. Arctigenin inhibits lung metastasis of colorectal cancer by regulating cell viability and metastatic phenotypes. *Molecules* 2016, *21*, 1135. [CrossRef] [PubMed]
- 12. Maimaitili, A.; Shu, Z.; Cheng, X.; Kaheerman, K.; Sikandeer, A.; Li, W. Arctigenin, a natural lignan compound, induces G0/G1 cell cycle arrest and apoptosis in human glioma cells. *Oncol. Lett.* **2017**, *13*, 1007–1013. [CrossRef] [PubMed]
- 13. Pilkington, L.I. Lignans: A chemometric analysis. Molecules 2018, 23, 1666. [CrossRef] [PubMed]
- Amancha, P.K.; Liu, H.-J.; Ly, T.W.; Shia, K.-S. General approach to 2,3-dibenzyl-γ-butyrolactone lignans: Application to the total synthesis of (±)-5'-methoxyyatein, (±)-5'-methoxyclusin, and (±)-4'-hydroxycubebinone. *Eur. J. Org. Chem.* 2010, 2010, 3473–3480. [CrossRef]
- 15. Isemori, Y.; Kobayashi, Y. An approach to β-substituted γ-butyrolactones and its application to the synthesis of lignans. *Synlett* **2004**, 1941–1944. [CrossRef]
- Ferrié, L.; Bouyssi, D.; Balme, G. Selective lewis acid catalyzed transformation (γ-butyrolactone versus cyclopropane) of 2-methoxy-4-benzyltetrahydrofuran derivatives. Efficient synthesis of lignan lactones. Org. Lett. 2005, 7, 3143–3146. [CrossRef] [PubMed]
- Duan, S.; Huang, S.; Gong, J.; Shen, Y.; Zeng, L.; Feng, Y.; Ren, W.; Leng, Y.; Hu, Y. Design and synthesis of novel arctigenin analogues for the amelioration of metabolic disorders. *ACS Med. Chem. Lett.* 2015, *6*, 386–391. [CrossRef] [PubMed]
- 18. Rye, C.; Barker, D. An acyl-Claisen approach to tetrasubstituted tetrahydrofuran lignans: Synthesis of fragransin A2, talaumidin, and lignan analogues. *Synlett* **2009**, 3315–3319. [CrossRef]
- 19. Barker, D.; Dickson, B.; Dittrich, N.; Rye, C.E. An acyl-Claisen approach to the synthesis of.lignans and substituted pyrroles. *Pure Appl. Chem.* **2012**, *84*, 1557–1565. [CrossRef]
- 20. Dickson, B.D.; Dittrich, N.; Barker, D. Synthesis of 2,3-syn-diarylpent-4-enamides via acyl-Claisen rearrangements of substituted cinnamyl morpholines: Application to the synthesis of magnosalicin. *Tetrahedron Lett.* **2012**, *53*, 4464–4468. [CrossRef]
- Duhamel, N.; Rye, C.E.; Barker, D. Total Synthesis of ent-hyperione A and ent-hyperione B. *Asian J. Org. Chem.* 2013, 2, 491–493. [CrossRef]
- 22. Rye, C.E.; Barker, D. Asymmetric synthesis of (+)-galbelgin, (–)-kadangustin J, (–)-cyclogalgravin and (–)-pycnanthulignenes A and B, three structurally distinct lignan classes, using a common chiral precursor. *J. Org. Chem.* **2011**, *76*, 6636–6648. [CrossRef] [PubMed]
- 23. Pilkington, L.I.; Wagoner, J.; Polyak, S.J.; Barker, D. Enantioselective synthesis, stereochemical correction, and biological investigation of the rodgersinine family of 1,4-benzodioxane neolignans. *Org. Lett.* **2015**, *17*, 1046–1049. [CrossRef] [PubMed]
- 24. Pilkington, L.I.; Barker, D. Synthesis and biology of 1,4-benzodioxane lignan natural products. *Nat. Prod. Rep.* **2015**, *32*, 1369–1388. [CrossRef] [PubMed]
- 25. Pilkington, L.I.; Barker, D. Asymmetric synthesis and CD investigation of the 1,4-benzodioxane lignans eusiderins A, B, C, G, L, and M. *J. Org. Chem.* **2012**, *77*, 8156–8166. [CrossRef] [PubMed]
- 26. Pilkington, L.I.; Barker, D. Total synthesis of (–)-isoamericanin A and (+)-isoamericanol A. *Eur. J. Org. Chem.* **2014**, 1037–1046. [CrossRef]
- 27. Jung, E.; Pilkington, L.I.; Barker, D. Enantioselective synthesis of 2,3-disubstituted benzomorpholines: Analogues of lignan natural products. *J. Org. Chem.* **2016**, *81*, 12012–12022. [CrossRef] [PubMed]
- 28. Jung, E.; Dittrich, N.; Pilkington, L.I.; Rye, C.E.; Leung, E.; Barker, D. Synthesis of aza-derivatives of tetrahydrofuran lignan natural products. *Tetrahedron* **2015**, *71*, 9439–9456. [CrossRef]
- 29. Paterson, D.L.; Barker, D. Synthesis of the furo[2,3-b]chromene ring system of hyperaspindols A and B. *Beilstein J. Org. Chem.* **2015**, *11*, 265–270. [CrossRef] [PubMed]
- 30. Davidson, S.J.; Barker, D. Synthesis of various lignans via the rearrangements of 1,4-diarylbutane-1,4-diols. *Tetrahedron Lett.* **2015**, *56*, 4549–4553. [CrossRef]
- 31. Pilkington, L.I.; Barker, D. Synthesis of 3-methylobovatol. Synlett 2015, 26, 2425–2428.
- 32. Rye, C.E.; Barker, D. Asymmetric synthesis and anti-protozoal activity of the 8,4'-oxyneolignans virolin, surinamensin and analogues. *Eur. J. Med. Chem.* **2013**, *60*, 240–248. [CrossRef] [PubMed]

- 33. Tran, H.; Dickson, B.; Barker, D. Unexpected O-alkylation and ester migration in phenolic 2,3-diaryl-2,3-dihydrobenzo[b]furans. *Tetrahedron Lett.* **2013**, *54*, 2093–2096. [CrossRef]
- 34. Pilkington, L.I.; Song, S.M.; Fedrizzi, B.; Barker, D. Efficient total synthesis of (±)-isoguaiacin and (±)-isogalbulin. *Synlett* **2017**, *28*, 1449–1452.
- 35. Davidson, S.J.; Barker, D. Total synthesis of ovafolinins A and B: Unique polycyclic benzoxepin lignans through a cascade cyclization. *Angew. Chem. Int. Ed.* **2017**, *56*, 9483–9486. [CrossRef] [PubMed]
- 36. Davidson, S.J.; Pearce, A.N.; Copp, B.R.; Barker, D. Total synthesis of (–)-bicubebin A, B, (+)-bicubebin C and structural reassignment of (–)-cis-cubebin. *Org. Lett.* **2017**, *19*, 5368–5371. [CrossRef] [PubMed]
- 37. Kakis, F.J.; Fetizon, M.; Douchkine, N.; Golfier, M.; Mourgues, P.; Prange, T. Mechanistic studies regarding the oxidation of alcohols by silver carbonate on celite. *J. Org. Chem.* **1974**, *39*, 523–533. [CrossRef]
- Fétizon, M.; Golfier, M.; Louis, J.-M. A new synthesis of lactones: Application to (±)-mevalonolactone. J. Chem. Soc. D 1969, 1118–1119. [CrossRef]
- Xin, H.; Kong, Y.; Wang, Y.; Zhou, Y.; Zhu, Y.; Li, D.; Tan, W. Lignans extracted from Vitex negundo possess cytotoxic activity by G2/M phase cell cycle arrest and apoptosis induction. *Phytomedicine* 2013, 20, 640–647. [CrossRef] [PubMed]
- Bose, J.S.; Gangan, V.; Prakash, R.; Kumar Jain, S.; Kumar Manna, S. A dihydrobenzofuran lignan induces cell death by modulating mitochondrial pathway and G2/M cell cycle arrest. *J. Med. Chem.* 2009, *52*, 3184–3190. [CrossRef] [PubMed]
- 41. Fetizon, M.; Balogh, V.; Golfier, M. Oxidations with silver carbonate/celite. V. Oxidations of phenols and related compounds. *J. Org. Chem.* **1971**, *36*, 1339–1341. [CrossRef]
- Reddy, R.S.; Prasad, P.K.; Ahuja, B.B.; Sudalai, A. CuCN-mediated cascade cyclization of 4-(2-bromophenyl)-2-butenoates: A high-yield synthesis of substituted naphthalene amino esters. *J. Org. Chem.* 2013, 78, 5045–5050. [CrossRef] [PubMed]
- 43. Sharma, P.; Ritson, D.J.; Burnley, J.; Moses, J.E. A synthetic approach to kingianin A based on biosynthetic speculation. *Chem. Commun.* **2011**, *47*, 10605–10607. [CrossRef] [PubMed]
- 44. Aldous, D.J.; Batsanov, A.S.; Yufit, D.S.; Dalençon, A.J.; Dutton, W.M.; Steel, P.G. The dihydrofuran template approach to furofuran synthesis. *Org. Biomol. Chem.* **2006**, *4*, 2912–2927. [CrossRef] [PubMed]
- Tanoguchi, M.; Kashima, T.; Saika, H.; Inoue, T.; Arimoto, M.; Yamaguchi, H. Studies on the constituents of the seeds of hernandia ovigera L. VII.: Syntheses of (±)-hernolactone and (±)-hernandin. *Chem. Pharm. Bull.* 1989, 37, 68–72. [CrossRef]
- Gomes, C.A.; Girão da Cruz, T.; Andrade, J.L.; Milhazes, N.; Borges, F.; Marques, M.P.M. Anticancer activity of phenolic acids of natural or synthetic origin: A structure-activity study. *J. Med. Chem.* 2003, *46*, 5395–5401. [CrossRef] [PubMed]
- Haga, Y.; Okazaki, M.; Shuto, Y. Systematic strategy for the synthesis of cyanobacterin and its stereoisomers.
 Asymmetric total synthesis of dechloro-cyanobacterin and its enantiomer. *Biosci. Biotechnol. Biochem.* 2003, 67, 2183–2193. [CrossRef] [PubMed]
- 48. Li, D.; Zhao, B.; Sim, S.-P.; Li, T.-K.; Liu, A.; Liu, L.F.; LaVoie, E.J. 8,9-Methylenedioxybenzo[i]phenanthridines: Topoisomerase I-targeting activity and cytotoxicity. *Bioorg. Med. Chem.* **2003**, *11*, 3795–3805. [CrossRef]
- 49. Takei, Y.; Mori, K.; Matsui, M. Synthesis of *dl*-matairesinol dimethyl ether, dehydrodimethylconidendrin and dehydrodimethylretrodendrin from ferulic acid. *Agric. Biol. Chem.* **1973**, *37*, 637–641. [CrossRef]
- 50. Brown, E.; Daugan, A. Lignames: 10. Preparation des (R)-(+) et (S)-(-)-β-piperonyl et β-veratryl-γbutyrolactones et leur utilisation dans la synthese totale de lignanes optiquement actifs. *Tetrahedron* **1989**, 45, 141–154. [CrossRef]
- 51. Baran, P.S.; DeMartino, M.P. Intermolecular oxidative enolate heterocoupling. *Angew. Chem. Int. Ed.* **2006**, 45, 7083–7086. [CrossRef] [PubMed]
- 52. Nishibe, S.; Okabe, K.; Hisada, S. Isolation of phenolic compounds and spectroscopic analysis of a new lignan from Trachelospermum asiaticum var. intermedium. *Chem. Pharm. Bull.* **1981**, *39*, 2078–2082. [CrossRef]
- 53. Gonzalez, A.G.; Estevez-Reyes, R.; Estevez-Braun, A.M. Buplerol and guayarol, new lignans from the seeds of bupleurum salicifolium. *J. Chem. Res.* **1990**, *21*, 220–221.
- 54. De Souza, V.A.; da Silva, R.; Pereira, A.C.; Royo, V.D.A.; Saraiva, J.; Montanheiro, M.; de Souza, G.H.B.; da Silva Filho, A.A.; Grando, M.D.; Donate, P.M.; et al. Trypanocidal activity of (–)-cubebin derivatives against free amastigote forms of Trypanosoma cruzi. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 303–307. [CrossRef] [PubMed]

- 55. Badheka, L.P.; Prabhu, B.R.; Mulchandani, N.B. Dibenzylbutyrolactone lignans from Piper cubeba.
- *Phytochemistry* **1986**, 25, 487–489. [CrossRef]
- 56. Evcim, U.; Gozler, B.; Freyer, A.J.; Shamma, M. Haplomyrtin and (–)-haplomyrfolin: Two lignans from haplophyllum myrtifolium. *Phytochemistry* **1986**, *25*, 1949–1951. [CrossRef]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).