

OPEN

Association of Paraoxonase 1 Gene Polymorphisms With the Risk of Hepatitis B Virus-related Liver Diseases in a Guangxi Population

A Case-control Study

Xianjun Lao, MM, Xiaogang Wang, MM, Yanqiong Liu, MM, Yu Lu, PhD, Dongmei Yang, MM, Minyan Liu, MM, Xiaolian Zhang, MM, Chengzhi Rong, MM, Xue Qin, PhD, and Shan Li, PhD

Abstract: Paraoxonase 1 (PON1), a liver-induced glycoprotein enzyme responsible for antioxidant defense against reactive oxygen species and anti-inflammatory, has been linked to various cancers. The objective of this study was to explore the association of PON1 rs662 and rs705382 with the risk of chronic hepatitis B (CHB), hepatitis B virus-related liver cirrhosis (LC), and hepatocellular carcinoma (HCC) in patients living in the Guangxi region of southern China.

The PON1 rs662 and rs705382 single-nucleotide polymorphisms (SNPs) were genotyped by polymerase chain reaction–restriction fragment length polymorphism (PCR-RFLP) in 99 CHB patients, 84 LC patients, 258 HCC patients, and 221 healthy controls.

Significant associations with CHB risk were observed for the rs705382 SNP after adjusting for sex, age, ethnicity, smoking, alcohol consumption, and body mass index. When stratified by sex and age, this positive association was significantly strengthened among men and individuals over 40 years old. Moreover, a decreased risk of LC was associated with the rs705382 CG and the combined GG + CG genotypes among women, with borderline statistical significance. In haplotype analyses, the haplotype GA was associated with a 1.68-fold increase in the risk of HCC.

Our results showed that the PON1 rs705382 SNP might be a risk factor for CHB in Guangxi populations.

(*Medicine* 94(48):e2179)

Abbreviations: BMI = body mass index, CHB = chronic hepatitis B, CI = confidence interval, HBV = hepatitis B virus, HCC = hepatocellular carcinoma, LC = liver cirrhosis, LDL = low-density lipoprotein, NAFLD = nonalcoholic fatty liver disease, OR = odds ratio, PON1 = paraoxonase 1, ROS = reactive oxygen species, SNPs = single-nucleotide polymorphisms.

Editor: Raffaele Pezzilli.

Received: August 25, 2015; revised: October 30, 2015; accepted: November 4, 2015.

From the Department of Clinical Laboratory, First Affiliated Hospital of Guangxi Medical University, Nanning, Guangxi, China.

Correspondence: Shan Li, Department of Clinical Laboratory, First Affiliated Hospital of Guangxi Medical University, Nanning 530021, Guangxi, China (e-mail: lis8858@126.com).

Co-correspondence: Xue Qin, Department of Clinical Laboratory, First Affiliated Hospital of Guangxi Medical University, Nanning 530021, Guangxi, China (e-mail: qinxue919@126.com).

XL and XW contributed equally to this work.

The authors have no conflicts of interest to disclose.

Copyright © 2015 Wolters Kluwer Health, Inc. All rights reserved.

This is an open access article distributed under the Creative Commons Attribution License 4.0, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. ISSN: 0025-7974

DOI: 10.1097/MD.0000000000002179

INTRODUCTION

Hepatocellular carcinoma (HCC) is the fifth most commonly diagnosed cancer (782,000 new cancer cases) worldwide and the third most frequent cause of cancer-related death (746,000 cancer deaths) in 2012.¹ The great majority of HCC cases, regardless of etiology, usually develop in the context of liver cirrhosis (LC).² According to previous studies, an estimated 402,208 new liver cancer cases and 372,079 HCC-related deaths have occurred in China since 2008.³ Likewise, HCC cases greatly increased in the United States between 2007 and 2011.⁴ Thus, HCC remains a formidable challenge to medicine despite the great efforts made in its prevention and treatment.

As a multifactorial and complex process, the exact pathogenesis of HCC is still unclear. Hepatitis B virus (HBV) infection is globally the most important etiologic factor of HCC, accounting for an estimated 80% of cases worldwide.^{2,5} Previous cohort studies have indicated that being a chronic carrier of HBV was associated with a 7 to 98-fold higher risk for HCC development.⁶ The risk increases with progressive fibrosis, and is also high in LC patients. In addition to HBV, chronic infection with the hepatitis C virus (HCV), smoking, excessive alcohol consumption, and heavy aflatoxin exposure have also been proposed as risk factors for HCC.^{7–9} However, a large proportion of HCC cases develop among individuals without these known risk factors, suggesting that host genetic factors may play an important role in HCC etiology.⁹

In recent years, the effect of reactive oxygen species (ROS) on hepatic carcinogenesis has been well recognized. It has been suggested that excessive production of ROS may lead to oxidative stress and severe DNA damage, which has been implicated in abnormal cell functioning or even carcinogenesis.¹⁰ Paraoxonase 1 (PON1) is a liver-induced glycoprotein enzyme that is involved in the mechanism of antioxidant defense against ROS and anti-inflammatory.^{11,12} Mackness et al have reported that PON1 enzymatic activity is correlated with cardiovascular disease, primarily due to the effect of anti-low-density lipoprotein (LDL) oxidative modification.¹³ Additionally, the significant decline of serum PON1 activities has been found to serve as a strong predictor of acute viral hepatitis B, chronic alcoholic hepatitis, liver cirrhosis, and nonalcoholic fatty liver disease (NAFLD).^{14–16} Therefore, single-nucleotide polymorphisms (SNPs) in the PON1 gene may differentially affect the enzymatic activity, possibly contributing to the susceptibility to various diseases.¹⁷

The human PON1 gene, encoded by the chromosome 7p21–22, contains 9 exons and 8 introns. PON1 presents at least 2 polymorphisms (rs662 and rs854560) located in the coding region and 5 located in the promoter region (rs705379,

rs705380, rs705381, rs854571, and rs854572). Of these, rs705379 polymorphism contributed the most (22.8%) and rs854572 polymorphism contributed the least (<1%) of PON1 activity.¹⁸ The most studied mutation is rs662 (Q192R), which results in a glutamine (Q) to arginine (R) substitution at position 192. The 192Gln variant seems to induce higher PON1 hydrolytic activity towards paraoxon^{19,20} and to be more effective than 192Arg in the prevention of LDL oxidation.²¹ For, several studies have looked at the correlation between the rs662 SNP and the risk of various cancers, such as ovarian epithelial carcinoma,²² brain astrocytoma and meningioma,²³ lymphoma,²⁴ and lung cancer,²⁵ in various populations. To date, only 1 hospital-based case-control study has investigated the association of PON1 rs662 polymorphism with HCC susceptibility.²⁶ Another common polymorphism of PON1 is the rs705382 SNP, which is a C/G substitution at codon 1434 of the 5'-untranslated region (UTR). Previous work revealed that variant allele of rs705382-1434C>G was independently associated with higher levels of PON1 activity and expression.²⁷ However, the role of the rs705382 polymorphism in HCC is still unknown.

The aim of this study was to further explore the relationship between 2 previously studied PON1 polymorphisms (rs662 and rs705382) and CHB, HBC-related LC, and HCC susceptibility in Guangxi populations.

MATERIALS AND METHODS

Study Population

The present study included 441 HBV-related patients and 221 healthy controls. Patients were periodically recruited at the First Affiliated Hospital of Guangxi Medical University, from April through October of 2014, including 99 CHB patients, 84 HBV-related LC patients, and 258 HBV-related HCC patients. The inclusion and exclusion criteria have been previously described.²⁸ Eligible participants were confirmed to have a history of chronic HBV infection (≥ 6 months). CHB was diagnosed by elevation of the liver function markers alanine aminotransferase (ALT) or aspartate aminotransferase (AST) (>40 IU/mL), as well as by HBV-DNA levels $>1 \times 10^3$ copies/mL. HBV-LC was diagnosed pathologically or was based on the combination of laboratory tests and radiologic evidence. HBV-HCC was diagnosed based on positive findings obtained from ultrasonography, computed tomography (CT), and magnetic resonance imaging (MRI), and/or by pathological examination. A significant increase in serum α -fetoprotein (AFP) (>400 ng/mL) was also determined to indicate HBV-HCC. Patients were excluded from the current study if they had coinfection with other hepatitis virus (hepatitis A/C/D/E virus); alcoholic liver disease, autoimmune hepatitis, or primary biliary cirrhosis; any concomitant malignant cancers; or a family history of HCC, or a history of autoimmune diseases such as systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA). Controls were randomly selected from healthy individuals who attended the same hospital for a routine physical examination during the patient recruitment periods. The selection criteria for controls were absence of any malignancy or other hepatic illness, including HBV infection. All participants were from the Guangxi district. At recruitment, written informed consent was obtained from each participant. Additionally, blood samples and demographic characteristics, including age, sex, ethnicity, tobacco and alcohol use, and body mass index (BMI), were also collected by the interviewers. This study was

approved by the ethics committee of the First Affiliated Hospital of Guangxi Medical University.

DNA Extraction and PON1 Polymorphism Genotyping

Genomic DNA was extracted from 2-mL peripheral venous blood by a high-salting-out method using phenol-chloroform. The PON1 rs662 and rs705382 SNPs were genotyped by polymerase chain reaction–restriction fragment length polymorphism (PCR-RFLP), without knowledge of the case or control status. The PCR was carried out in 1.0 μ L of each primer, 12.5 μ L of Green PCR Master Mix (Shanghai Sangon Biotech Co., Ltd., Shanghai, China), 9.5 μ L of sterilized deionized water, and 2.0 μ L of template DNA per reaction. The forward and reverse primers used were rs662-F 5'-TATTGTTGCTGTGGGACCTGAG-3' and R 5'-CACGC-TAAACCCAAATACATCTC-3',²⁹ rs705382-F 5'-GAGAGG-GAAAGTGGTCAGCT-3' and R 5'-GAAGTGTGAGTTT GGGCAGG-3'. The PCR reaction for rs662 included a 5-min preincubation step at 95°C, followed by 40 cycles of 45 seconds at 95°C, 45 seconds at 59°C, 45 seconds at 72°C, and then a final 10-min extension step at 72°C. For rs705382, it included a 5-min preincubation step at 95°C, followed by 25 cycles of 39 seconds at 95°C, 30 seconds at 56°C, 30 seconds at 72°C, and then a final 10-min extension step at 72°C. After amplification, all products were separated on 2.5% agarose gel and subsequently stained with ethidium bromide to visualize the bands (Fig. 1). To control the quality of the PCR reaction, a negative control was also performed in each genotyping assay. In addition, a 10% random sample was sent to Sangon Biotech Company for genotyping in duplicates, and the reproducibility was 100%.

Statistical Analysis

The Hardy–Weinberg equilibrium (HWE) was assessed by a goodness-of-fit chi-square test for genotypes in the control groups. For baseline data, 1-way analysis of variance (ANOVA) tests were used for the categorical variables, and Student *t* tests were used for the continuous variables to compare the differences in demographic characteristics between patients and controls. Genotype and allele frequencies among different groups were compared using the chi-square test and Fisher exact test, when appropriate. Logistic regression analysis, adjusted for age, sex, tobacco smoking, alcohol consumption, and BMI, was utilized to calculate the adjusted odds ratios (ORs) and 95% confidence intervals (CIs). Further stratified analyses were conducted according to age and sex. The haplotype construction was performed using SHEsis software (<http://analysis.bio-x.cn/myAnalysis.php>).³⁰ A probability level of less than 0.05 was judged as statistically significant. All statistical analyses were performed using SPSS, version 13.0 (SPSS Inc., Chicago, IL).

RESULTS

Characteristics of the Study Population

The basic demographics of patients and controls are summarized in Table 1. Briefly, the CHB patients were, on average, significantly younger than the healthy control, LC, and HCC patients ($P < 0.001$). The HCC patients were predominantly men, tobacco smokers, and alcohol drinkers ($P < 0.001$). However, the healthy control group consisted largely of non-smokers and nondrinkers, which indicated that the controls

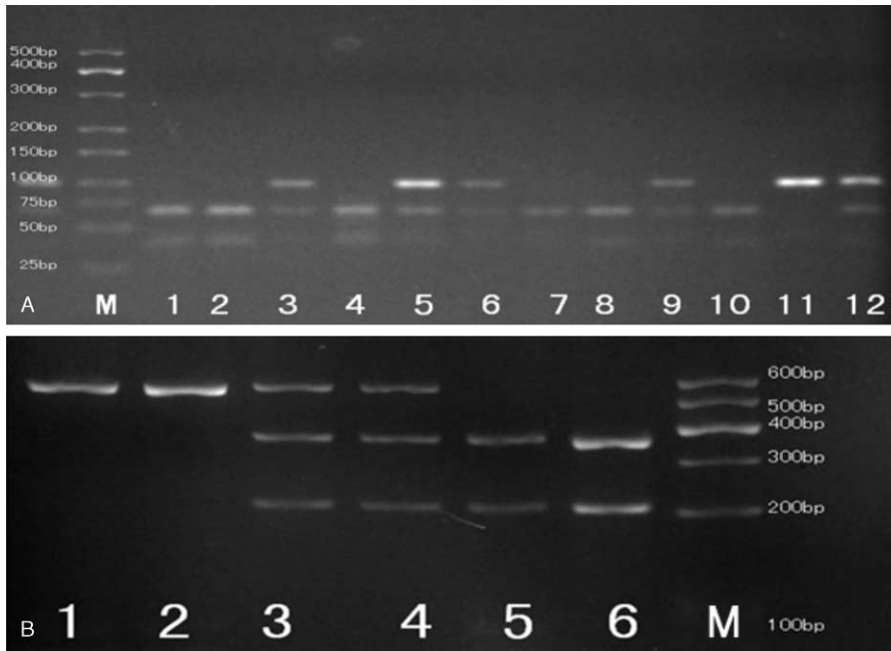


FIGURE 1. PCR-RFLP assay for analyzing the rs662 and rs705382 polymorphisms of the PON1 gene. (A) rs662-lanes M: DNA marker; lanes 1, 2, 4, 7, 8, and 10 show GG genotype; lanes 3, 5, 6, 9, and 12 show AG genotype; lane 11 shows AA genotype. (B) rs705382-lanes M: DNA marker; lanes 1 and 2 show GG genotype; lanes 3 and 4 show CG genotype; lanes 5 and 6 show CC genotype. PCR-RFLP = polymerase chain reaction–restriction fragment length polymorphism, PON1 = paraoxonase 1.

seemed to have a healthier lifestyle in comparison with the HCC patients. Apart from this, most of the healthy controls were from the Zhuang ethnic group, whereas the HBV patients were primarily from the Han ethnic group ($P=0.044$). The mean BMI in the control individuals was similar to that in the HBV patients ($P=0.210$). In healthy controls, the observed genotype frequencies of rs662 and rs705382 were both in agreement with

HWE ($P=0.437$ for rs662 and $P=1.000$ for rs705382) (Table 2).

Association Analysis of Candidate SNPs with CHB, HBV-related LC, and HCC Risk

Table 2 shows the genotype and allele frequencies of the 2 candidate SNPs within the PON1 gene for the patient and

TABLE 1. Clinical and Epidemiological Data for the Patient and Control Groups

Parameters	Controls (n = 221, %)	CHB (n = 99, %)	LC (n = 84, %)	HCC (n = 258, %)	P Value
Age, y, mean ± SD	46.51 ± 6.88	38.58 ± 12.07	49.94 ± 12.40	49.31 ± 11.28	<0.001
BMI, mean ± SD	22.61 ± 3.51	22.34 ± 3.65	23.01 ± 4.47	22.14 ± 3.26	0.210
Sex					
Men	123 (55.7)	76 (76.8)	66 (78.6)	235 (91.1)	<0.001
Women	98 (44.3)	23 (23.2)	18 (21.4)	23 (8.9)	
Tobacco smoking					
Yes	70 (31.7)	48 (48.5)	30 (35.7)	165 (64.0)	<0.001
No	151 (68.3)	51 (51.5)	54 (64.3)	93 (36.0)	
Alcohol drinking					
Yes	60 (27.1)	39 (39.4)	41 (48.8)	165 (64.0)	<0.001
No	161 (72.9)	60 (60.6)	43 (51.2)	93 (36.0)	
Ethnicity					
Han	97 (43.9)	59 (59.6)	50 (59.5)	152 (58.9)	0.044
Zhuang	112 (50.7)	36 (36.4)	31 (36.9)	106 (41.0)	
Others	12 (5.4)	4 (4.0)	3 (3.6)	8 (0.1)	

BMI = body mass index, CHB = chronic hepatitis B, HCC = hepatocellular carcinoma, LC = liver cirrhosis, SD = standard deviation.

TABLE 2. Genotype and Allele Frequencies of Candidate SNPs in Patients and Control Individuals

SNPs	Controls			CHB			LC			HCC		
	n = 221 (%)	n = 99 (%)	P _{OR}	n = 99 (%)	OR (95% CI)*	P _{OR}	n = 84 (%)	OR (95% CI)*	P _{OR}	n = 258 (%)	OR (95% CI)*	P _{OR}
rs662 Genotypes	HWE = 0.437											
AA	33 (14.9%)	14 (14.1%)	1.00 ^{ref}	14 (14.1%)	1.00 ^{ref}	1.00 ^{ref}	12 (14.3%)	1.00 ^{ref}	1.00 ^{ref}	39 (15.1%)	1.00 ^{ref}	0.607
AG	87 (39.4%)	44 (44.5%)	1.16 (0.51–2.61)	44 (44.5%)	1.16 (0.51–2.61)	0.725	33 (39.3%)	1.12 (0.49–2.56)	0.787	108 (41.9%)	1.17 (0.64–2.13)	0.864
GG	101 (45.7%)	41 (41.4%)	1.05 (0.47–2.35)	41 (41.4%)	1.05 (0.47–2.35)	0.906	39 (46.4%)	1.06 (0.47–2.36)	0.892	111 (43.0%)	0.95 (0.53–1.71)	0.874
AG + GG	188 (85.1%)	85 (85.9%)	1.10 (0.52–2.33)	85 (85.9%)	1.10 (0.52–2.33)	0.804	72 (85.7%)	1.09 (0.51–2.32)	0.834	219 (84.9%)	1.05 (0.60–1.81)	
rs662 Alleles												
A	153 (34.6%)	72 (36.4%)	1.00 ^{ref}	72 (36.4%)	1.00 ^{ref}	1.00 ^{ref}	57 (33.9%)	1.00 ^{ref}	1.00 ^{ref}	186 (36.0%)	1.00 ^{ref}	0.629
G	289 (65.4%)	126 (63.6%)	1.00 (0.67–1.50)	126 (63.6%)	1.00 (0.67–1.50)	0.996	111 (66.1%)	1.01 (0.68–1.50)	0.969	330 (64.0%)	0.93 (0.69–1.25)	
rs705382 Genotypes	HWE = 1.000											
CC	54 (24.5%)	15 (15.1%)	1.00 ^{ref}	15 (15.1%)	1.00 ^{ref}	1.00 ^{ref}	21 (25.0%)	1.00 ^{ref}	1.00 ^{ref}	62 (24.0%)	1.00 ^{ref}	0.441
CG	111 (50.2%)	56 (56.6%)	2.45 (1.11–5.43)	56 (56.6%)	2.45 (1.11–5.43)	0.027	36 (42.9%)	0.77 (0.40–1.51)	0.452	137 (53.1%)	1.22 (0.74–2.00)	0.891
GG	56 (25.3%)	28 (28.3%)	2.66 (1.12–6.31)	28 (28.3%)	2.66 (1.12–6.31)	0.027	27 (32.1%)	1.05 (0.51–2.16)	0.901	59 (22.9%)	1.04 (0.59–1.85)	0.548
CG + GG	167 (75.5%)	84 (84.9%)	2.53 (1.19–5.38)	84 (84.9%)	2.53 (1.19–5.38)	0.016	63 (75.0%)	0.87 (0.47–1.61)	0.659	196 (76.0%)	1.15 (0.72–1.84)	
rs705382 Alleles												
C	219 (49.5%)	86 (43.4%)	1.00 ^{ref}	86 (43.4%)	1.00 ^{ref}	1.00 ^{ref}	78 (46.4%)	1.00 ^{ref}	1.00 ^{ref}	261 (50.6%)	1.00 ^{ref}	0.876
G	223 (50.5%)	112 (56.6%)	1.50 (1.01–2.22)	112 (56.6%)	1.50 (1.01–2.22)	0.044	90 (53.6%)	1.03 (0.71–1.51)	0.865	255 (49.4%)	1.02 (0.77–1.36)	

CHB = chronic hepatitis B, CI = confidence interval, HCC = hepatocellular carcinoma, LC = liver cirrhosis, ref = reference, SNPs = single-nucleotide polymorphisms.

For rs662, codominant model: AG vs AA, GG vs AA; dominant model: AG + GG.

For rs705382, codominant model: CG vs CC, GG vs CC; dominant model: CG + GG vs CC.

* Adjusted for age, sex, tobacco smoking, alcohol consumption, and BMI.

control groups. Distributions of the rs662 and rs705382 genotypes and alleles were then compared across the groups, and no statistically significant differences were observed (both $P > 0.05$).

Associations between the PON1 rs662 polymorphism and CHB, HBV-related LC, and HCC risk were assessed using logistic regression analyses adjusted for sex, age, ethnicity, smoking, alcohol consumption, and BMI, and no significant associations were observed in any genetic models (Table 2). However, logistic regression analyses for the PON1 rs705382 polymorphism indicated that individuals carrying the rs705382 G allele were at moderately increased risk of CHB when compared with those carrying the C allele (adjusted OR 1.50, 95% CI 1.01–2.22, $P = 0.044$; Table 2). Similarly, we also found that both the rs705382 GG and CG carriers had more than a 2-fold risk of CHB (GG vs CC genotype: adjusted OR 2.66, 95% CI 1.12–6.31, $P = 0.027$; CG vs CC genotype: adjusted OR 2.45, 95% CI 1.11–5.43, $P = 0.027$) relative to the CC carriers. In the combined analyses, the GG + CG genotype was associated with a significantly increased risk of CHB, with adjusted OR of 2.53 (95% CI 1.19–5.38, $P = 0.016$).

We further examined the associations between the 2 PON1 SNPs and CHB, HBV-related LC, and HCC risk by stratifying each study participant into subgroups according to age and sex. The results suggested that men with rs705382 GG genotype and G allele exhibited an increased risk of CHB (GG vs CC genotype: adjusted OR 3.59, 95% CI 1.23–6.31, $P = 0.020$; G vs C allele: adjusted OR 1.76, 95% CI 1.07–2.88, $P = 0.025$; Table 3). Interestingly, compared with the rs705382 CC

genotype, a marginally decreased risk of LC was associated with the CG and the combined GG + CG genotypes among women (CG vs CC genotype: adjusted OR 0.26, 95% CI 0.07–0.98, $P = 0.047$; dominant model: adjusted OR 0.28, 95% CI 0.08–0.96, $P = 0.042$). On the contrary, neither the allele frequencies nor the genotype distributions of the rs662 SNP showed impacts on men or women in both the case and control groups (Table 3). In addition, among individuals aged 40 years or older, strong evidences of associations between CHB and the rs705382 variant genotypes occur, with adjusted OR of 10.336 for GG versus CC (95% CI 1.27–84.11, $P = 0.029$), adjusted OR of 11.54 for CG versus CC (95% CI 1.47–90.68, $P = 0.020$), and adjusted OR of 11.05 for the dominant model (95% CI 1.44–84.76, $P = 0.021$) (Table 4). Similar to the stratified results for sex, negative results were found for rs662 polymorphism in both older individuals (≥ 40 years old) and younger individuals (< 40 years old).

The genetic background of PON1 polymorphisms may vary between different races and populations. Therefore, the allele and genotype frequencies of the 2 SNPs in the control groups were compared with those previously reported in the Haplotype Map (HapMap) project (<http://www.ncbi.nlm.nih.gov/snp/>) and several human genetic association studies.^{31–33} As is shown in Table 5, the distribution of the 2 SNPs in the present study is significantly different from that in CEU (Utah residents with northern and western European ancestry) and YRI (Yoruba in Ibadan). For the rs662 SNP, the frequencies of the GG genotype and G allele in CEU population are significantly lower, whereas it is higher in the YRI population when compared with our data. In contrast, the GG genotype and

TABLE 3. Stratified Effects of PON1 rs662 and rs705382 Polymorphisms on CHB, LC, and HCC Risk Estimated by Sex

SNPs	Genotypes	CHB		LC		HCC		
		OR (95% CI)*	P _{OR}	OR (95% CI)*	P _{OR}	OR (95% CI)*	P _{OR}	
Men								
rs662	Codominant model	AG vs AA	1.13 (0.40–3.20)	0.813	1.57 (0.60–4.11)	0.361	1.28 (0.66–2.47)	0.459
		GG vs AA	1.46 (0.54–3.90)	0.456	1.32 (0.53–3.32)	0.553	0.97 (0.52–1.83)	0.934
	Dominant model	AG + GG vs AA	1.31 (0.52–3.33)	0.570	1.42 (0.59–3.40)	0.435	1.10 (0.61–1.99)	0.757
		Allele model	G vs A	1.27 (0.76–2.11)	0.358	1.09 (0.69–1.73)	0.720	0.92 (0.67–1.28)
rs705382	Codominant model	CG vs CC	2.71 (1.00–7.33)	0.050	1.06 (0.48–2.38)	0.880	1.26 (0.74–2.00)	0.408
		GG vs CC	3.58 (1.23–10.46)	0.020	1.39 (0.59–3.27)	0.450	1.03 (0.59–1.85)	0.936
	Dominant model	CG + GG vs CC	0.82 (0.51–1.32)	0.418	1.19 (0.57–2.49)	0.644	1.18 (0.72–1.84)	0.534
		Allele model	G vs C	1.76 (1.07–2.88)	0.025	1.20 (0.77–1.87)	0.421	1.02 (0.75–1.40)
Women								
rs662	Codominant model	AG vs AA	0.85 (0.18–4.11)	0.837	0.27 (0.05–1.52)	0.137	1.18 (0.25–5.54)	0.834
		GG vs AA	0.46 (0.08–2.64)	0.383	0.28 (0.05–1.66)	0.161	1.04 (0.22–4.94)	0.959
	Dominant model	AG + GG vs AA	0.68 (0.15–3.12)	0.618	0.27 (0.05–1.40)	0.120	1.11 (0.26–4.75)	0.889
		Allele model	G vs A	0.68 (0.31–1.51)	0.345	0.67 (0.29–1.53)	0.339	0.98 (0.47–2.04)
rs705382	Codominant model	CG vs CC	2.00 (0.44–9.16)	0.372	0.26 (0.07–0.98)	0.047	0.97 (0.28–3.45)	0.967
		GG vs CC	1.53 (0.28–8.50)	0.628	0.33 (0.06–1.69)	0.183	0.68 (0.15–3.06)	0.617
	Dominant model	CG + GG vs CC	1.82 (0.43–7.74)	0.415	0.28 (0.08–0.96)	0.042	0.87 (0.26–2.94)	0.827
		Allele model	G vs C	1.19 (0.54–2.60)	0.672	0.53 (0.23–1.22)	0.137	0.84 (0.42–1.69)

CHB = chronic hepatitis B, CI = confidence interval, HCC = hepatocellular carcinoma, LC = liver cirrhosis, OR = odds ratio, SNPs = single-nucleotide polymorphisms.

* Adjusted for age, sex, tobacco smoking, alcohol consumption, and BMI.

TABLE 4. Stratified Effects of PON1 rs662 and rs705382 Polymorphisms on CHB, LC, and HCC Risk Estimated by Age

SNPs	Genotypes	CHB		LC		HCC	
		OR (95% CI)*	P _{OR}	OR (95% CI)*	P _{OR}	OR (95% CI)*	P _{OR}
Age <40 y old							
rs662							
Codominant model	AG vs AA	1.71 (0.47–6.31)	0.418	2.71 (0.24–30.80)	0.422	2.58 (0.57–11.75)	0.221
	GG vs AA	0.47 (0.12–1.87)	0.283	2.03 (0.19–22.03)	0.559	2.08 (0.458–9.46)	0.342
Dominant model	AG + GG vs AA	1.01 (0.31–3.30)	0.994	2.30 (0.23–23.16)	0.480	2.32 (0.57–9.51)	0.243
Allele model	G vs A	0.60 (0.31–1.17)	0.133	1.14 (0.44–2.95)	0.793	1.29 (0.62–2.68)	0.493
rs705382							
Codominant model	CG vs CC	1.56 (0.51–4.75)	0.437	2.39(0.39–14.13)	0.351	3.20 (0.90–11.38)	0.073
	GG vs CC	1.37 (0.40–4.69)	0.614	3.33(0.58–19.03)	0.177	2.62 (0.62–11.02)	0.189
Dominant model	CG + GG vs CC	1.48 (0.54–4.10)	0.450	2.83(0.57–14.13)	0.204	2.98 (0.91–9.73)	0.071
Allele model	G vs C	1.19 (0.63–2.24)	0.591	1.97(0.79–4.90)	0.147	1.60 (0.80–3.22)	0.187
Age ≥40 y old							
rs662							
Codominant model	AG vs AA	0.85(0.28–2.61)	0.780	0.84 (0.34–2.03)	0.690	1.05 (0.544–2.041)	0.827
	GG vs AA	1.15(0.41–3.25)	0.796	0.87 (0.37–2.04)	0.745	0.86 (0.449–1.636)	0.641
Dominant model	AG + GG vs AA	1.02(0.38–2.79)	0.963	0.85 (0.38–1.91)	0.700	0.94 (0.514–1.731)	0.849
Allele model	G vs A	1.16(0.68–1.98)	0.568	0.95 (0.61–1.47)	0.808	0.89 (0.641–1.226)	0.468
rs705382							
Codominant model	CG vs CC	11.54 (1.47–90.68)	0.020	0.66 (0.27–1.59)	0.354	0.82 (0.43–1.58)	0.551
	GG vs CC	10.34 (1.27–84.11)	0.029	0.94 (0.43–2.07)	0.877	0.82 (0.47–1.43)	0.491
Dominant model	CG + GG vs CC	11.05 (1.44–84.76)	0.021	0.69 (0.33–1.42)	0.309	0.94 (0.55–1.60)	0.805
Allele model	G vs C	12.27 (2.91–51.77)	0.001	0.70 (0.42–1.17)	0.171	0.94 (0.64–1.37)	0.726

CHB = chronic hepatitis B, CI = confidence interval, HCC = hepatocellular carcinoma, LC = liver cirrhosis, OR = odds ratio, SNPs = single-nucleotide polymorphisms.

* Adjusted for age, sex, tobacco smoking, alcohol consumption and BMI.

TABLE 5. Comparison of Genotype and Allele Frequencies in the Healthy Control Participants of Our Study and That From the HapMap Project

Polymorphisms	Samples, N	Genotype Frequency, n (%)			P Value	Alleles Frequency, n (%)		
		AA	AG	GG		A	G	P Value
rs662								
Present study	221	33 (14.9%)	87 (39.4%)	101 (45.7%)		153 (34.6%)	289 (65.4%)	
CEU*	226	98 (43.4%)	106 (46.9%)	22 (9.7%)	<0.001	302 (66.8%)	150 (33.2%)	<0.001
HCB*	86	16 (18.6%)	42 (48.8%)	28 (32.6%)	0.154	74 (43.0%)	98 (57.0%)	0.246
JPT*	170	14 (8.2%)	72 (42.4%)	84 (49.4%)	0.312	100 (29.4%)	240 (70.6%)	0.363
YRI*	226	10 (4.4%)	74 (32.7%)	142 (62.8%)	0.009	94 (20.8%)	358 (79.2%)	0.027
Turkish ³⁰	234	20 (8.6%)	93 (39.7%)	121 (51.7%)	0.392	133 (28.4%)	335 (71.6%)	0.287
Anhui ³¹	136	19 (14.0%)	57 (41.9%)	60 (44.1%)	0.909	95 (34.9%)	177 (65.1%)	1.000
Sichuan ³²	248	39 (15.7%)	101 (40.7%)	108 (43.6%)	0.941	179 (36.1%)	317 (63.9%)	0.883
rs705382								
Present study	221	54 (24.5%)	111 (50.2%)	56 (25.3%)		219 (49.5%)	223 (50.5%)	
CEU*	226	24 (10.6%)	104 (46.0%)	98 (43.4%)	0.006	152 (33.6%)	300 (66.4%)	0.026
HCB*	86	18 (20.9%)	36 (41.9%)	32 (37.2%)	0.186	72 (41.9%)	100 (58.1%)	0.286
JPT*	172	22 (12.8%)	98 (57.0%)	52 (30.2%)	0.095	142 (41.3%)	202 (58.7%)	0.226
YRI*	226	146 (64.6%)	64 (28.3%)	16 (7.1%)	<0.001	356 (78.8%)	96 (21.2%)	<0.001
Sichuan ³²	248	50 (20.2%)	129 (52.0%)	69 (27.8%)	0.682	229 (46.2%)	267 (53.8%)	0.619

CEU = Utah residents with northern and western Europeanancestry; HCB = Han Chinese in Beijing, China; JPT = Japanese in Tokyo, Japan; YRI = Yoruba in Ibadan, Nigeria.

* Data from HapMap Project.

G-allele distributions of the rs705382 SNP are significantly higher and lower in the CEU and YRI populations, respectively.

Haplotype Analysis of Associations Between PON1 Polymorphisms and CHB, HBV-related LC, and HBV-related HCC

We performed further the haplotype analysis to evaluate the linkage strength of nearby SNPs located in the same chromosome regions. A total of 4 two-locus haplotypes (CA, CG, GA, and GG) were analyzed. We identified that GG was the most common haplotype in both patients and controls. CA was the next most common in healthy controls, CHB, and LC patients, whereas CG was in second place for HCC patients (Table 6). The frequency of the haplotype GA was significantly associated with increased HCC risk (OR 1.68, 95% CI 1.08–2.60, $P = 0.020$).

DISCUSSION

In this study, we investigated the association of PON1 gene SNPs (rs662 and rs705382) with risk of CHB, HBV-related LC, and HCC in the Guangxi population. Our results demonstrate that, for the overall population, the candidate rs705382 SNP was statistically significantly related with greater risk of CHB, but not in LC or HCC. When stratified by sex and age, significant associations with increased risk of CHB were observed for the rs705382 SNP in men and older individuals (≥ 40 years old), whereas a marginally decreased risk of HBV-related LC was found for the variant genotype of rs705382 among women. Haplotype analysis of the 2 candidate SNPs revealed that the haplotype GA served as an increase risk factor for HCC.

The pathogenic mechanism linking oxidative stress and hepatocarcinogenesis has previously been studied.^{34,35} PON1, one member of the PON family, plays a crucial role in antioxidant defense and anti-inflammatory.^{11,12} Sun et al³⁶ reported that the expression levels of PON1 protein in patients with LC and HCC were significantly lower than that in the healthy controls. Therefore, it is easily inferred that the significant decline of serum PON1 concentrations or activity can profoundly affect liver disease status. According to previous results, genetic mutations like promoter polymorphisms are the most important factors regarding the variations in PON1 enzyme activity between individuals. Several studies have reported a functional significance of PON1 gene polymorphisms. It has been suggested that the variant alleles of PON1 rs662 and rs705382 seem to induce higher PON1 activity.^{19,20,27} However, there are fewer studies investigating the association of the PON1 rs662 and rs705382 polymorphism with CHB and LC, as well as with HCC.

In 2013, based on 217 confirmed cases and 217 cancer-free controls, Akkiz et al²⁶ firstly reported the association of PON1 rs662 polymorphisms with HCC risk in the Turkish population. In that study, the variant allele frequencies were 0.28 and 0.27 for HCC cases and controls, respectively. Further statistical analysis showed that the rs662 polymorphisms have no direct effect on the risk for developing HCC, which is in agreement with our results. Furthermore, there were 2 studies investigating the associations of the PON1 rs705382 polymorphism with ischemic stroke (IS) and sporadic amyotrophic lateral sclerosis (SALS), respectively. The study on the association between rs705382 SNP and ischemic stroke was conducted by Kim et al²⁷ in 2009, and involved 86 Korean patients with IS. According to the results, the rs705382 G allele was observed with a little lower frequency in cases than in controls. A general

TABLE 6. Haplotype Analysis of PON1 rs662 and rs705382 Polymorphisms With the Risk of CHB, LC, and HCC

Haplotypes	CHB vs Controls			LC vs Controls			HCC vs Controls			
	Controls (n = 442, %)	CHB (n = 198, %)	LC (n = 168, %)	HCC (n = 516, %)	OR (95% CI)	P_{OR}	OR (95% CI)	P_{OR}	OR (95% CI)	P_{OR}
CA	117 (26.4%)	49 (25.0%)	43 (25.8%)	123 (23.9%)	0.93 (0.63–1.36)	0.705	0.97 (0.65–1.46)	0.887	0.87 (0.65–1.17)	0.359
CG	101 (22.9%)	37 (18.5%)	35 (20.6%)	132 (25.6%)	0.76 (0.50–1.16)	0.204	0.87 (0.57–1.35)	0.538	1.16 (0.86–1.56)	0.336
GA	36 (8.2%)	23 (11.4%)	14 (8.1%)	63 (12.2%)	1.44 (0.82–2.50)	0.200	1.06 (0.51–1.89)	0.962	1.68 (1.08–2.60)	0.020
GG	188 (42.5%)	89 (45.1%)	76 (45.5%)	198 (38.3%)	1.12 (0.80–1.57)	0.521	1.13 (0.79–1.62)	0.502	0.84 (0.65–1.09)	0.196

CHB = chronic hepatitis B, CI = confidence interval, HCC = hepatocellular carcinoma, LC = liver cirrhosis, OR = odds ratio.

linear model revealed that the rs705382-1434G was related with higher PON1 activity and acted as a risk factor for IS in the Korean population.²⁷ In contrast, Chen et al³¹ assessed 373 SALS patients and 248 controls from Southwest China, and the rs705382 polymorphism did not seem to contribute to the risk for developing SALS in that study population, as evidenced by the absence of statistical differences in allele frequencies between the control and SALS group, and by the association analysis of rs705382 SNP with SALS risk. Nevertheless, either PON1 rs662 or rs705382 SNP was investigated in these studies. Few studies so far have estimated the influences of both the PON1 rs662 and rs705382 polymorphisms on CHB, LC, and HCC risks. The current study was conducted to investigate the association between 2 SNPs (rs662, rs705382) of the PON1 gene, and CHB, LC, and HCC risks in a substantial number of cases and controls (99 CHB patients, 84 LC, 258 HCC, and 221 healthy controls), which greatly increased statistical power of the study when compared with previous studies. In our study, the rs705382-1434G allele was distributed as follows: 0.57 in CHB, 0.54 in LC, 0.51 in HCC, and 0.51 in healthy controls. Although no significant differences were found in the allele frequencies between patients and controls, the rs705382 SNP was associated with increased risk of CHB, after adjusting for all conventional risk factors. It is possible that these inconsistent findings may be explained by the differences in studied population, sample size, and other potential risk factors that influence the effects of genetic variants on disease risk.

On the basis of the subgroup analysis by sex and age, our observations suggest that there are age and sex-related differences in the correlation between the PON1 rs705382 polymorphism and CHB risk. Individuals with the rs705382 GG genotype had a significantly high risk of CHB among men and people over 40 years old, but no such results were observed among women and people younger than 40 years old. Notably, compared with those of the CC genotype, women carrying the rs705382 CG and the combined CG + GG genotypes were at increased risk of LC. However, this association only bordered on statistical significance and the result should be interpreted cautiously. Previous studies have shown a considerable decrease of PON1 activity with advancing age,^{37,38} and this leads to increased risk of disease among older individuals. On the contrary, Hernandez et al³⁹ reported that women had significant lower levels of PON1 among the Spanish population, suggesting that women may be more susceptible to diseases. Nevertheless, this is not in line with our results. Considering the distinct genetic background and differences in exposure to environmental risk factors between the Chinese and Spanish populations, further similar studies with a larger number of patients should be performed for confirmation.

Analysis of multiple linked SNPs tend to be more powerful for providing robust results than individual SNPs.⁴⁰ Meanwhile, information is lacking regarding PON1 haplotypes and HCC risk. Consequently, haplotype analysis is to be preferred while evaluating the relationship of PON1 polymorphisms with HCC risk by using the SHEsis software. We observed that the haplotype GG (carrying all variants) was the most prevalent one in both the patient and control groups; however, the haplotype GA (carrying 1 variant and 1 wild-type allele) was associated with a 1.68-fold increase in the risk of HCC.

To the best of our knowledge, the present study is the first to assess the potential implications of the PON1 gene rs662 and rs705382 for susceptibility to CHB, HBV-related LC, and HCC.

Despite the strengths and biological plausibility of the associations observed in this study, there are still several limitations need to be addressed. Firstly, the study participants were all recruited from the Guangxi district, and they may not be representative of the entire Chinese population. Data should be applied cautiously to other ethnic groups. Additionally, because of the lack of dietary exposure assessment of aflatoxin, we have not carried out the gene–environment interaction analyses of aflatoxin exposure and PON1 candidate SNPs in risk estimates of HCC. Follow-up studies are needed to examine the detailed molecular interactions between PON1 polymorphisms and aflatoxin exposure.

In conclusion, our results showed that the PON1 rs705382 SNP was associated with the increased risk of CHB in a Guangxi population, especially in men and individuals over 40 years old. Larger prospective studies with detailed information about aflatoxin exposure are warranted to validate our findings.

ACKNOWLEDGMENTS

The authors would like to thank Professor Zhiping Chen (Department of Occupational Health and Environmental Health, School of Public Health at Guangxi Medical University) for the integrity and accuracy of the data analysis.

REFERENCES

1. Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015;136:E359–386.
2. El-Serag HB, Rudolph KL. Hepatocellular carcinoma: epidemiology and molecular carcinogenesis. *Gastroenterology*. 2007;132:2557–2576.
3. Li Q, Du J, Guan P, et al. [Estimation and prediction of incidence, mortality and prevalence on liver cancer, in 2008, China]. *Zhonghua Liu Xing Bing Xue Za Zhi*. 2012;33:554–557.
4. Jemal A, Bray F, Center MM, et al. Global cancer statistics. *CA Cancer J Clin*. 2011;61:69–90.
5. Yu MC, Yuan JM, Govindarajan S, et al. Epidemiology of hepatocellular carcinoma. *Can J Gastroenterol*. 2000;14:703–709.
6. Nguyen VT, Law MG, Dore GJ. Hepatitis B-related hepatocellular carcinoma: epidemiological characteristics and disease burden. *J Viral Hepat*. 2009;16:453–463.
7. Ha NB, Ha NB, Ahmed A, et al. Risk factors for hepatocellular carcinoma in patients with chronic liver disease: a case-control study. *Cancer Causes Control*. 2012;23:455–462.
8. Chuang SC, La Vecchia C, Boffetta P. Liver cancer: descriptive epidemiology and risk factors other than HBV and HCV infection. *Cancer Lett*. 2009;286:9–14.
9. Dragani TA. Risk of HCC: genetic heterogeneity and complex genetics. *J Hepatol*. 2010;52:252–257.
10. Klaunig JE, Kamendulis LM. The role of oxidative stress in carcinogenesis. *Annu Rev Pharmacol Toxicol*. 2004;44:239–267.
11. Mackness MI, Mackness B, Durrington PN. Paraonase and coronary heart disease. *Atheroscler Suppl*. 2002;3:49–55.
12. Mackness B, Durrington PN, Mackness MI. Human serum paraonase. *Gen Pharmacol*. 1998;31:329–336.
13. Mackness B, Durrington PN, Mackness MI. Polymorphisms of paraonase genes and low-density lipoprotein lipid peroxidation. *Lancet*. 1999;353:468–469.
14. Burlina A, Galzigna L. Serum arylesterase isoenzymes in chronic hepatitis. *Clin Biochem*. 1974;7:202–205.

15. Keskin M, Dolar E, Dirican M, et al. Baseline and salt-stimulated paraoxonase and arylesterase activities in patients with chronic liver disease: relation to disease severity. *Intern Med J*. 2009;39:243–248.
16. Mogarekar MR, Talekar SJ. Serum lactonase and arylesterase activities in alcoholic hepatitis and hepatitis B. *Indian J Gastroenterol*. 2013;32:307–310.
17. Brophy VH, Hastings MD, Clendenning JB, et al. Polymorphisms in the human paraoxonase (PON1) promoter. *Pharmacogenetics*. 2001;11:77–84.
18. Brophy VH, Jampsa RL, Clendenning JB, et al. Effects of 5' regulatory-region polymorphisms on paraoxonase-gene (PON1) expression. *Am J Hum Genet*. 2001;68:1428–1436.
19. Mackness MI, Mackness B, Durrington PN, et al. Paraoxonase: biochemistry, genetics and relationship to plasma lipoproteins. *Curr Opin Lipidol*. 1996;7:69–76.
20. Landers JE, Shi L, Cho TJ, et al. A common haplotype within the PON1 promoter region is associated with sporadic ALS. *Amyotroph Lateral Scler*. 2008;9:306–314.
21. Mackness B, Mackness MI, Arrol S, et al. Effect of the human serum paraoxonase 55 and 192 genetic polymorphisms on the protection by high density lipoprotein against low density lipoprotein oxidative modification. *FEBS Lett*. 1998;423:57–60.
22. Lurie G, Wilkens LR, Thompson PJ, et al. Genetic polymorphisms in the Paraoxonase 1 gene and risk of ovarian epithelial carcinoma. *Cancer Epidemiol Biomarkers Prev*. 2008;17:2070–2077.
23. Martinez C, Molina JA, Alonso-Navarro H, et al. Two common nonsynonymous paraoxonase 1 (PON1) gene polymorphisms and brain astrocytoma and meningioma. *BMC Neurol*. 2010;10:71.
24. Conesa-Zamora P, Ruiz-Cosano J, Torres-Moreno D, et al. Polymorphisms in xenobiotic metabolizing genes (EPHX1, NQO1 and PON1) in lymphoma susceptibility: a case control study. *BMC Cancer*. 2013;13:228.
25. Eom SY, Yim DH, Lee CH, et al. Interactions between paraoxonase 1 genetic polymorphisms and smoking and their effects on oxidative stress and lung cancer risk in a Korean population. *PLoS One*. 2015;10:e0119100.
26. Akkiz H, Kuran S, Akgollu E, et al. Effect of PON1 gene polymorphisms in Turkish patients with hepatocellular carcinoma. *Meta Gene*. 2013;1:93–101.
27. Kim NS, Kang BK, Cha MH, et al. Association between PON1 5'-regulatory region polymorphisms, PON1 activity and ischemic stroke. *Clin Biochem*. 2009;42:857–863.
28. Liu Y, Xie L, Zhao J, et al. Association between catalase gene polymorphisms and risk of chronic hepatitis B, hepatitis B virus-related liver cirrhosis and hepatocellular carcinoma in Guangxi population: a case-control study. *Medicine (Baltimore)*. 2015;94:e702.
29. Abdussemet H, Fuat D, Mustafa G, et al. The paraoxonase 1 (PON1) gene polymorphisms in coronary artery disease in the southeastern Turkish population. *Turk J Med Sci*. 2011;6:895–902.
30. Shi YY, He L. SHEsis a powerful software platform for analyses of linkage disequilibrium, haplotype construction, and genetic association at polymorphism loci. *Cell Res*. 2005;15:97–98.
31. Chen Y, Huang R, Chen K, et al. Association analysis of PON polymorphisms in sporadic ALS in a Chinese population. *Neurobiol Aging*. 2012;33:e2941–29432949.
32. Aksoy-Sagirli P, Cakmakoglu B, Isbir T, et al. Paraoxonase-1 192/55 polymorphisms and the risk of lung cancer in a Turkish population. *Anticancer Res*. 2011;31:2225–2229.
33. Y.F Z. The Association between Paraoxonase1Gene Polymorphisms. *Oxidative Stress and Retinopathy in Type2Diabetes Mellitus*. 2013.
34. Fujinaga H, Tsutsumi T, Yotsuyanagi H, et al. Hepatocarcinogenesis in hepatitis C: HCV shrewdly exacerbates oxidative stress by modulating both production and scavenging of reactive oxygen species. *Oncology*. 2011;81(Suppl 1):11–17.
35. Hikita H, Kodama T, Tanaka S, et al. Activation of the mitochondrial apoptotic pathway produces reactive oxygen species and oxidative damage in hepatocytes that contribute to liver tumorigenesis. *Cancer Prev Res (Phila)*. 2015;8:693–701.
36. Sun C, Chen P, Chen Q, et al. Serum paraoxonase 1 heteroplasmon, a fucosylated, and sialylated glycoprotein in distinguishing early hepatocellular carcinoma from liver cirrhosis patients. *Acta Biochim Biophys Sin (Shanghai)*. 2012;44:765–773.
37. Marchegiani F, Marra M, Spazzafumo L, et al. Paraoxonase activity and genotype predispose to successful aging. *J Gerontol A Biol Sci Med Sci*. 2006;61:541–546.
38. Seres I, Paragh G, Deschene E, et al. Study of factors influencing the decreased HDL associated PON1 activity with aging. *Exp Gerontol*. 2004;39:59–66.
39. Hernandez AF, Gil F, Leno E, et al. Interaction between human serum esterases and environmental metal compounds. *Neurotoxicology*. 2009;30:628–635.
40. Gupta N, Singh S, Maturu VN, et al. Paraoxonase 1 (PON1) polymorphisms, haplotypes and activity in predicting cad risk in North-West Indian Punjabis. *PLoS One*. 2011;6:e17805.