

Cardiac macrophages prevent sudden death during heart stress

Junichi Sugita¹, Katsuhito Fujii^{1,2,3✉}, Yukiteru Nakayama¹, Takumi Matsubara¹, Jun Matsuda¹, Tsukasa Oshima¹, Yuxiang Liu¹, Yujin Maru¹, Eriko Hasumi¹, Toshiya Kojima¹, Hiroshi Seno⁴, Keisuke Asano⁴, Ayumu Ishijima⁴, Naoki Tomii⁴, Masatoshi Yamazaki⁴, Fujimi Kudo⁵, Ichiro Sakuma⁴, Ryoza Nagai⁶, Ichiro Manabe^{5✉} & Issei Komuro¹

Cardiac arrhythmias are a primary contributor to sudden cardiac death, a major unmet medical need. Because right ventricular (RV) dysfunction increases the risk for sudden cardiac death, we examined responses to RV stress in mice. Among immune cells accumulated in the RV after pressure overload-induced by pulmonary artery banding, interfering with macrophages caused sudden death from severe arrhythmias. We show that cardiac macrophages crucially maintain cardiac impulse conduction by facilitating myocardial intercellular communication through gap junctions. Amphiregulin (AREG) produced by cardiac macrophages is a key mediator that controls connexin 43 phosphorylation and translocation in cardiomyocytes. Deletion of *Areg* from macrophages led to disorganization of gap junctions and, in turn, lethal arrhythmias during acute stresses, including RV pressure overload and β -adrenergic receptor stimulation. These results suggest that AREG from cardiac resident macrophages is a critical regulator of cardiac impulse conduction and may be a useful therapeutic target for the prevention of sudden death.

¹ Department of Cardiovascular Medicine, the University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan. ² Department of Advanced Cardiology, the University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan. ³ PRESTO, Japan Science and Technology Agency, 4-1-8 Honcho Kawaguchi, Saitama 332-0012, Japan. ⁴ Medical Device Development and Regulation Research Center, Department of Bioengineering/Department of Precision Engineering, School of Engineering, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8656, Japan. ⁵ Department of Disease Biology and Molecular Medicine, Graduate School of Medicine, Chiba University, 1-8-1 Inohana, Chuo-ku, Chiba-shi, Chiba 260-8670, Japan. ⁶ Jichi Medical University, 3311-1 Yakushiji, Shimotsuke-shi, Tochigi-ken, Tochigi 329-0498, Japan. ✉email: fujii-ty@umin.ac.jp; manabe-ty@umin.ac.jp

Hear failure is a growing medical and economic burden worldwide^{1,2}. Research and development efforts have focused largely on left ventricular (LV) dysfunction and left heart failure, but right heart failure has been gaining increasing attention as a serious unmet medical need. Right heart failure associates with adverse outcomes in a variety of heart diseases, including left heart failure, in part by compromising cardiac function and worsening serious arrhythmias. Indeed, right ventricular (RV) dysfunction increases the risk for sudden cardiac death³. However, the molecular mechanisms for right ventricular (RV) dysfunction remain poorly understood.

Recent studies have revealed the various roles played by immune cells in the maintenance of homeostasis and the development of cardiac diseases. For instance, many reports indicate that macrophages promote left ventricular (LV) remodeling⁴ and, conversely, are required for proper adaptive responses to stress and healing after myocardial infarction^{5,6}. Other immune cells such as lymphocytes and mast cells also reportedly play important roles in the LV remodeling^{7,8}; however, very little is known about the actions of immune cells during RV stress responses⁹.

Here, we show that cardiac macrophages play an essential role in survival during cardiac stress by maintaining the cardiac electrical conduction through regulating cardiac gap junction formation. These findings may pave the way for a new therapeutic method for sudden death.

Results

Macrophages are vital for survival after right heart burden.

To assess the functional roles of immune cells during acute RV stress, we imposed pressure overload on the right ventricle by pulmonary artery banding (PAB) (Fig. 1a). Accumulation of various types of immune cells, including macrophages, granulocytes, CD4⁺, and CD8⁺ T cells, and B cells, was observed after PAB (Fig. 1b, Supplementary Fig. 1). To gain insight into the functions of these cell populations during acute RV stress, we performed PAB on mice treated with clodronate liposomes, which depleted or impaired monocytes and macrophages¹⁰, or with anti-Ly6G antibody, which depleted granulocytes¹¹; *Cd4a*^{-/-} and *Cd8a*^{-/-} mice, which lacked CD4⁺ and CD8⁺ T cells, respectively; and *Rag2*^{-/-} mice, which lacked mature T and B cells. While the interventions affecting granulocytes and T and B cells did not impact survival after PAB, more than half (57%) of the clodronate-treated mice died within 4 h after PAB (Fig. 1c). Electrocardiograms (ECGs) showed that, after PAB, 4 of the 7 clodronate-treated mice developed advanced atrioventricular (AV) conduction block, which led to ventricular arrest (Fig. 1d and Supplementary Table 1). To further confirm the involvement of macrophage deficiency in the sudden death phenotype, we used another macrophage depletion model, *Cx3cr1*^{CreER/+;R26iDTR/+} mice, in which tamoxifen induces CX₃CR1⁺ cell-specific expression of the diphtheria toxin receptor¹². Macrophage depletion in this model also led to sudden death after PAB (Supplementary Figures 2 and 3). Given the sudden death phenotype and advanced AV block seen in clodronate-treated mice, these results strongly suggest that monocytes and macrophages are essential for survival during acute RV pressure overload, presumably by suppressing severe arrhythmias.

Macrophage-derived AREG protects against sudden death. We recently demonstrated that cardiac macrophage-derived AREG is essential for the proper cardiac adaptive response to LV pressure overload⁵. Accordingly, we tested whether AREG is similarly important for the response to RV pressure overload. Although we did not observe sudden death in *Areg*^{-/-} mice in the steady-state,

continuous ECG monitoring of freely moving 8- to 12-week-old *Areg*^{-/-} mice showed sinus arrest or sinoatrial (SA) block, intermittent complete AV block, and premature ventricular contractions (PVCs) in all 6 mice tested (Fig. 2a and Supplementary Table 2). By contrast, no arrhythmias were observed in wild-type (WT) mice (*n* = 6). When we imposed RV pressure overload on *Areg*^{-/-} mice with PAB, the mice developed AV block, and >85% of them died within 24 h (Fig. 2b, c and Supplementary Table 3) with no noticeable histological or functional changes in their RVs (Supplementary Figs. 4 and 5). To confirm that macrophage-derived AREG is important for preventing arrhythmias, we transplanted bone marrow (BM) from WT or *Areg*^{-/-} mice into lethally irradiated WT mice. Six weeks after the BM transplantation (BMT), 99.2% of the cardiac-resident macrophages were replaced with BM-derived cells. After PAB, 57% of the chimeric mice receiving *Areg*^{-/-} BM died within several hours, and >80% died within 7 days. By contrast, no mice receiving WT-BMT died after PAB (Fig. 2d). AV block was recorded in the *Areg*^{-/-}-BMT mice after PAB (Fig. 2e and Supplementary Table 3). In a publicly available single-cell RNA-seq dataset from heart stromal cells¹³, most *Areg*-expressing cells were *Cd68*⁺, indicating that macrophages are the major immune cell type expressing *Areg*, although some other cells might express AREG after PAB (Supplementary Fig. 6). Accordingly, the results of *Areg*^{-/-} BMT strongly suggest that AREG produced by macrophages is essential for protection against AV block and sudden death after PAB.

To further investigate whether AREG mediates the anti-arrhythmogenic action of macrophages, we next continuously administered AREG to clodronate-treated mice beginning the day prior of PAB. This AREG supplementation prevented the sudden death otherwise seen after PAB (Fig. 2f and Supplementary Table 3). Collectively, these results demonstrate that *Areg* deficiency confers abnormal cardiac electrical conduction and arrhythmogenicity, even in the steady-state, which leads to severe arrhythmias and sudden death after RV pressure overload.

It has been reported that sympathetic nerve activation triggers and facilitates arrhythmias and is an important mechanism underlying sudden cardiac death¹⁴, and that isoproterenol induces AV block and lethal arrhythmias in mice with conduction disturbances¹⁵⁻¹⁷. This prompted us to test the effects of β -adrenergic receptor stimulation. Isoproterenol administration (5 mg/kg) caused sudden death in ~57% of *Areg*^{-/-} mice within 2 days (Fig. 2g) and induced severe arrhythmias, including advanced AV block and ventricular fibrillation (VF) (Fig. 2h and Supplementary Fig. 7). All ECGs at the time of death showed VF in the isoproterenol-administered *Areg*^{-/-} mice. By contrast, WT mice had only infrequent isolated PVCs and did not die.

We further tested whether AREG deficiency promotes ventricular arrhythmogenic substrates by analyzing electrical activation in Langendorff-perfused whole heart preparations using optical mapping. VT/VF was induced through programmed stimulation or burst pacing. While VT was induced in only 1 of 6 WT hearts, all 6 *Areg*^{-/-} hearts developed VT when subjected to the VT/VF induction protocol (Fig. 2i).

AREG regulates cardiac gap junction formation. Because *Areg*^{-/-} mice exhibited multiple types of arrhythmias, we speculated that they were subjected to global electrical conduction disturbances. Myocardial electrical conduction depends primarily on the intercellular transfer of current through gap junctions composed of connexin protein complexes^{18,19}. We, therefore, performed western blotting to examine the cardiac expression of connexin (Cx) proteins in WT and *Areg*^{-/-} mice. Although

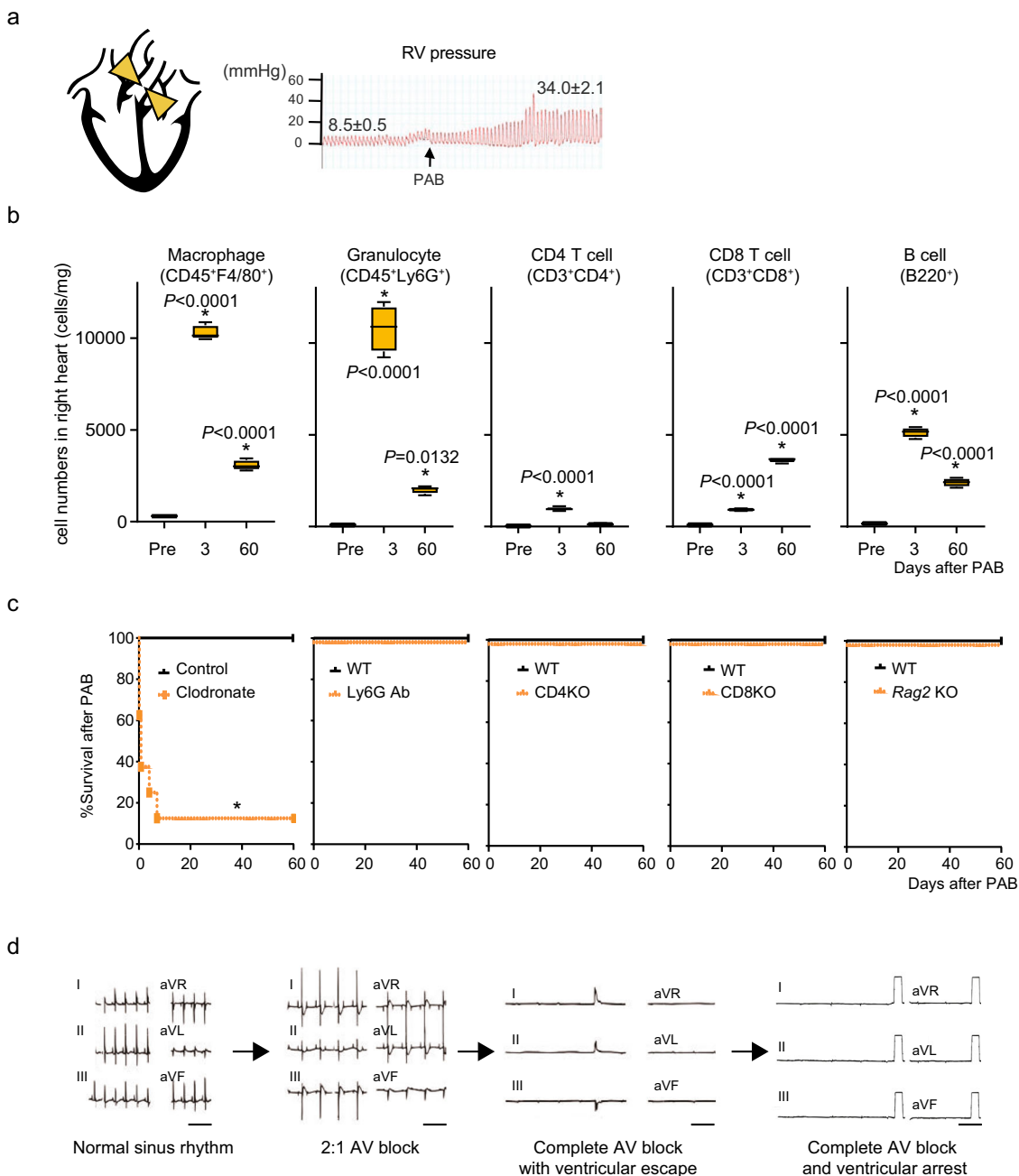


Fig. 1 Macrophage depletion causes sudden death after right heart stress. **a** Mouse model of right heart pressure overload (PAB). The pulmonary artery was ligated to increase the RV pressure to 30–35 mmHg from 5–10 mmHg. **b** Flow cytometric analysis of immune cells in the right atrial (RA) and RV free wall in wild-type (WT) mice after PAB. $n = 4$ in each group. One-way ANOVA followed by Tukey’s post-hoc test. $*P < 0.05$ vs. pre: steady-state. Box plots show center line as median, box limits as upper and lower quartiles, whiskers as minimum to maximum values. **c** Effects of depleting macrophages (clodronate), granulocytes (Ly6G Ab), CD4⁺ (CD4KO) or CD8⁺ (CD8KO) cells, or T and B cells (*Rag2*KO) on survival after PAB. $*P < 0.0001$ (log-rank test). **d** Representative six-channel ECGs from clodronate-treated mice after PAB. Shown is the representative sequence of arrhythmia progression observed in a mouse. Bars, 400 msec.

there were no apparent differences in the levels of Cx40 and Cx45 or their band positions between WT and *Areg*^{-/-} mice, Cx43 bands from WT heart samples exhibited band shifts and smears, which were eliminated by treatment with lambda protein phosphatase before loading (Fig. 3a). This indicates that the band shifts and smearing reflect phosphorylation of Cx43. Levels of Cx43 phosphorylation appeared to be much reduced in *Areg*^{-/-} hearts, and the reduced phosphorylation persisted after PAB. Phosphorylation of Cx43 reportedly affects its localization and function²⁰.

In mice, Cx43 is the primary component of gap junctions in both the atria and ventricles²¹. Phosphorylation of Cx43 is linked to its translocation and modulates gap junction properties, and dephosphorylation of Cx43 is associated with electrical uncoupling and susceptibility to arrhythmia^{20,22}. We immunohistochemically stained Cx43 in mouse hearts (Fig. 3b). In WT mice, Cx43 was primarily localized to the intercalated discs between myocytes, as previously reported²³. By contrast, the distribution of Cx43 was disorganized within cardiomyocytes in *Areg*^{-/-} mice, with Cx43 mislocated to the cells’ lateral surfaces

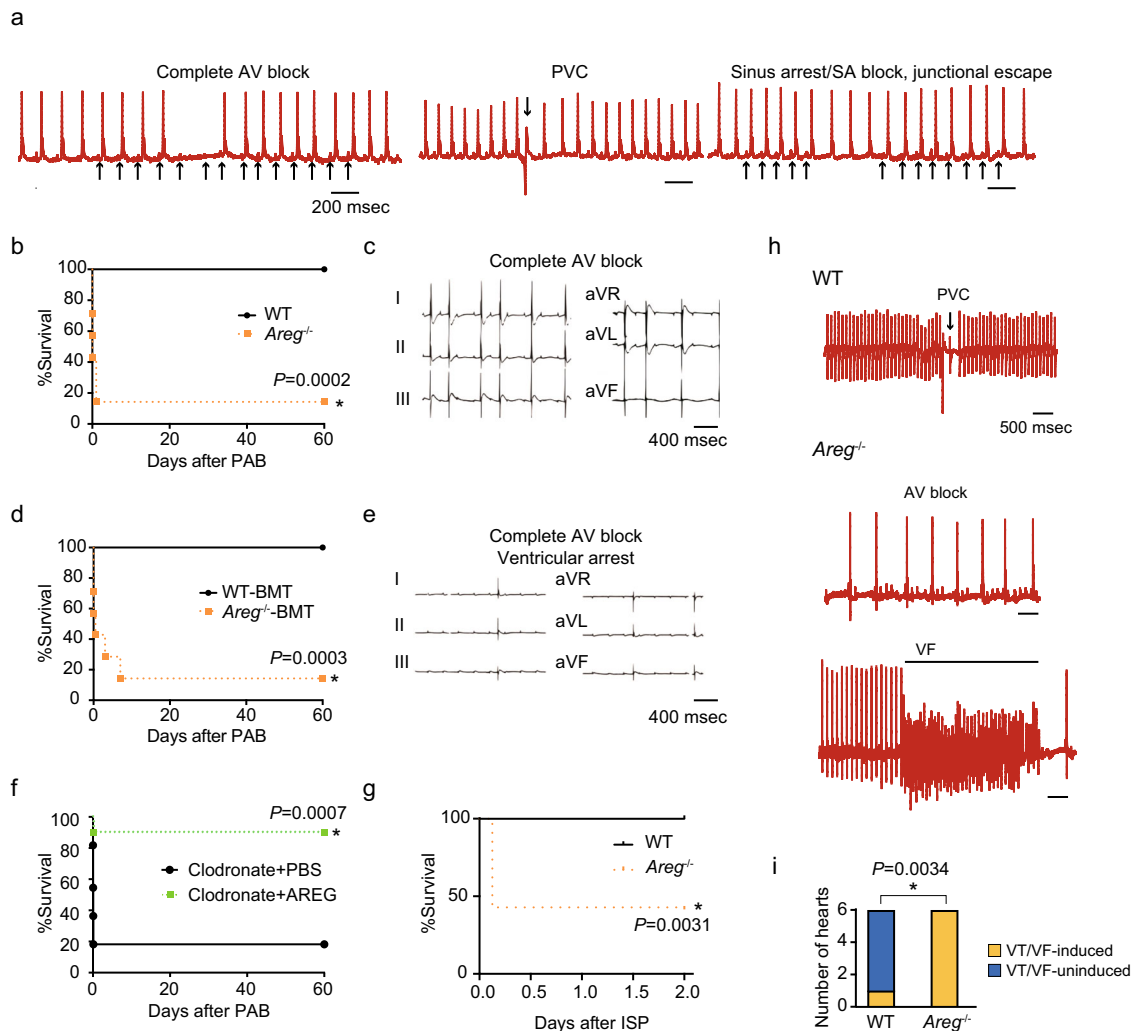


Fig. 2 *Areg* deficiency leads to abnormal cardiac electrical conduction at steady-state and lethal arrhythmias under cardiac stress. **a** ECGs from freely moving *Areg*^{-/-} mice recorded using telemetry. Spontaneous AV block, PVCs, and sinus arrest or SA block were observed in *Areg*^{-/-} mice. Bars indicate 200 msec. **b** Survival rates among WT and *Areg*^{-/-} mice after PAB. * $P < 0.001$, log-rank test. **c** Representative ECG from an *Areg*^{-/-} mouse showing complete AV block and ventricular arrest after PAB. **d**, **e** BM chimeric mice that received WT (WT-BMT) or *Areg*^{-/-} (*Areg*^{-/-}-BMT) BM were subjected to PAB. Survival rates after PAB are in **d**. * $P < 0.001$, log-rank test. In **e**, a representative ECG from an *Areg*^{-/-}-BMT mouse after PAB is shown. **f** Survival curves for clodronate-treated mice with or without continuous administration of AREG (5 μ g/day) from 24 h before PAB. $n = 10$ in each group. * $P < 0.01$, log-rank test. **g**, **h** WT and *Areg*^{-/-} mice were intraperitoneally administered isoproterenol (bolus, 5 mg/kg). Survival rates are shown in **g**. In **h**, representative ECGs from WT and *Areg*^{-/-} mice are shown. * $P < 0.05$, log-rank test. **i** Numbers of Langendorff-perfused hearts that developed VT/VF. * $P < 0.01$ (χ^2 -test).

(Fig. 3b, c and Supplementary Figs. 8 and 9). The disorganization was reversed by intraperitoneal administration of recombinant AREG (Fig. 3b). This mislocation of Cx43, called lateralization, is associated with arrhythmogenicity^{24,25}. One consequence of Cx43 lateralization is the opening of Cx43 hemichannels that are mislocated away from intercalated discs and do not form gap junction channels with adjacent cells. This results in abnormal ion trafficking and electrical conduction^{15,26}. To determine whether Cx43 lateralization is involved in the life-threatening arrhythmias seen in *Areg*^{-/-} mice, we utilized Gap19, which selectively inhibits Cx43 hemichannels without affecting normal gap junctional communication^{17,27}. Intraperitoneal administration of Gap 19 to *Areg*^{-/-} mice clearly reduced the incidence of isoproterenol-induced death (Supplementary Fig. 10). These results suggest that AREG is critically involved in the regulation of Cx43 localization and cardiac gap junction formation, possibly through Cx43 phosphorylation, and that Cx43 lateralization is a

cause of life-threatening arrhythmias triggered by cardiac stress in *Areg*^{-/-} mice.

AREG promotes phosphorylation of Cx43. We next carried out dye transfer assays to further assess the effects of AREG on gap junctional intercellular communication (GJIC) between cultured mouse cardiomyocytes²⁸. In this assay, a line is scraped into a confluent layer of cardiomyocytes, which enables a membrane-impermeant dye, 5(6)-carboxyfluorescein, to enter the affected cells through a transient tear in their plasma membrane. The dye can then migrate to adjacent cells through gap junctions. We quantified the level of dye transfer based on the distance the dye traveled from the scrape line (Fig. 4a). To assess the effects of cardiac macrophages on GJIC, we co-cultured cardiomyocytes with CD45⁺CD11b⁺ Ly6G⁺F4/80⁺ Ly6C^{lo} cardiac macrophages. The level of dye

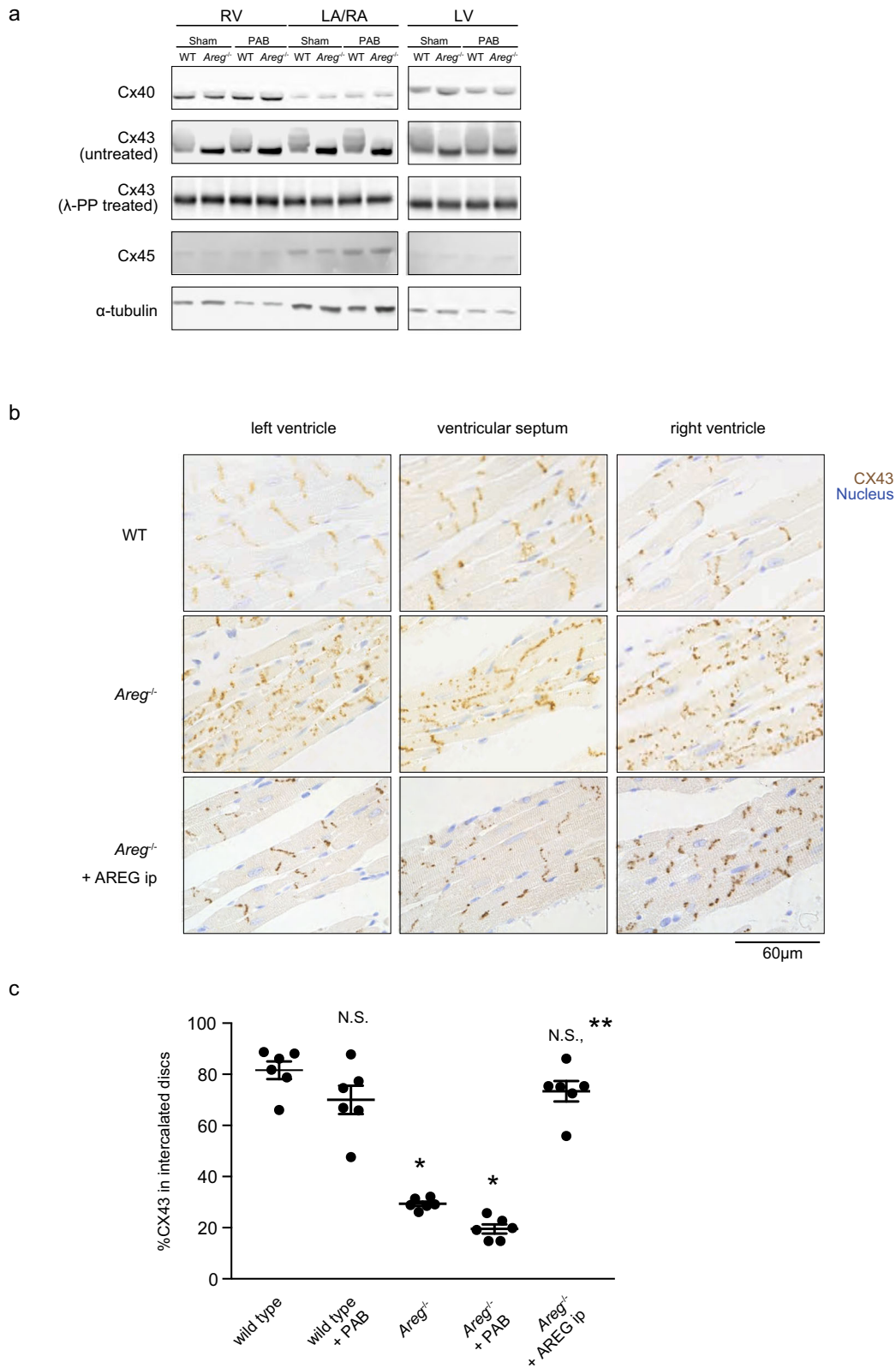
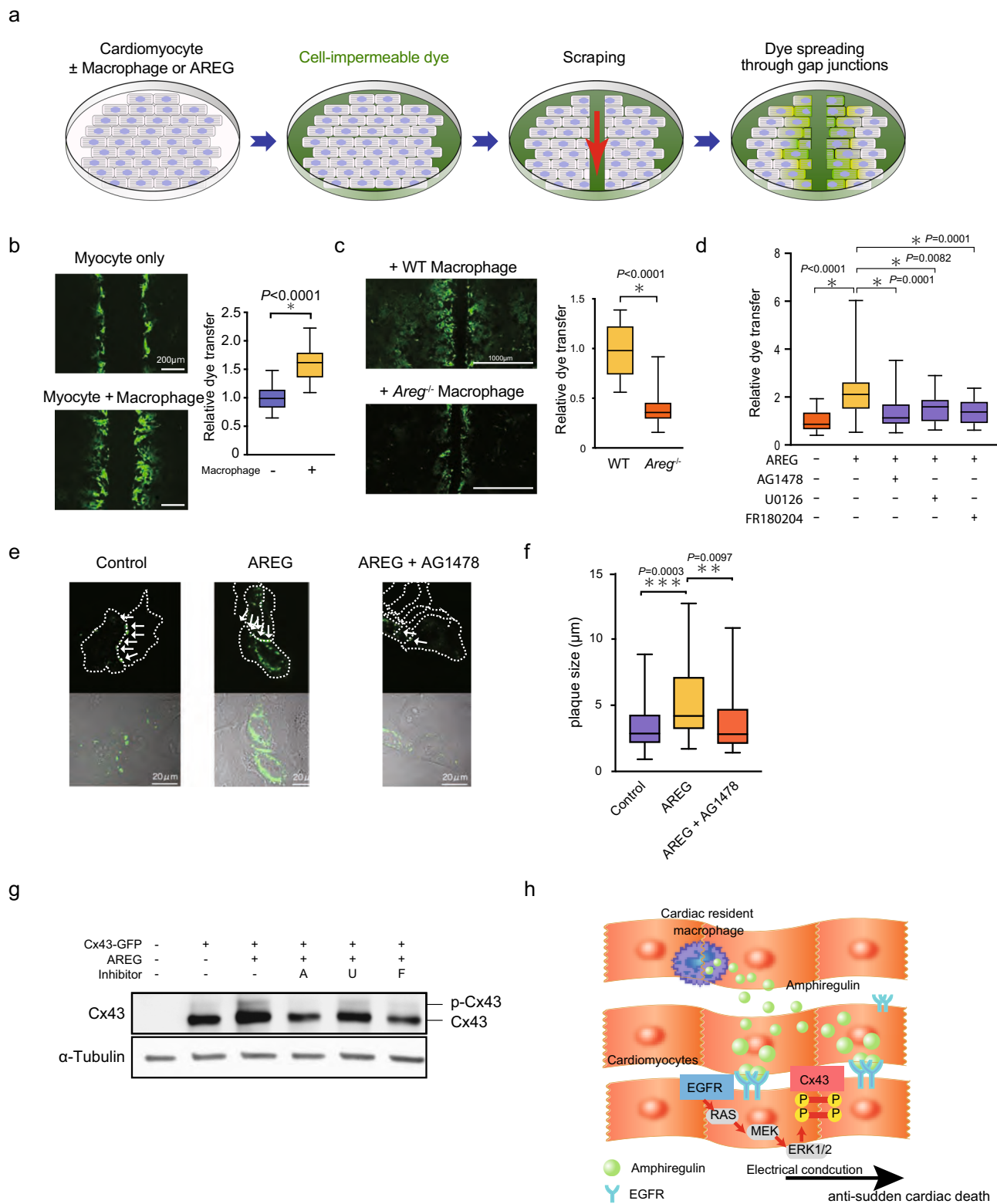


Fig. 3 AREG promotes gap junctional connection between cardiomyocytes through Cx43. **a** Western blot analysis of Cx40, Cx43, and Cx45. Representative immunoblots are shown. α-Tubulin was used as a loading control. Note the mobility shift of Cx43 bands in WT mouse hearts. **b** Immunohistochemical staining of Cx43 (brown) in the myocardium. Nuclei (blue) were counterstained with methylene blue. WT and *Areg*^{-/-} mice were intraperitoneally administered either vehicle (PBS) or recombinant AREG (5 μg), and the hearts were harvested 30 min later. WT and *Areg*^{-/-} mice not receiving AREG were administered vehicle (PBS) 30 min before sacrifice. The bar indicates 60 μm. The images show representative immunohistochemically stained hearts from WT or *Areg*^{-/-} mice at steady-state. **c** Fractions of Cx43 localized to the intercalated discs in cardiomyocytes in RVs. *n* = 6 mice in each group. One-way ANOVA followed by Tukey’s post-hoc test. **P* < 0.0001 vs. WT. ***P* < 0.0001 vs. *Areg*^{-/-}. Data are presented as mean values ± SEM.



transfer was significantly increased in the presence of macrophages (Fig. 4b).

We next tested the effects of AREG produced by macrophages on GJIC. As indicated by the dye transfer, GJIC between cardiomyocytes cocultured with *Areg*^{-/-} macrophages was significantly lower than between cardiomyocytes cocultured with WT macrophages (Fig. 4c). Moreover, recombinant AREG protein similarly increased the level of dye transfer (Fig. 4d).

We then assessed GJIC function by using a cell motion imaging system to analyze the conduction velocity of contraction propagation in cultured cardiomyocytes. Conduction velocities in cardiomyocytes co-cultured with cardiac macrophages were higher than in cardiomyocytes cultured alone (Supplementary Fig. 11). AREG similarly increased conduction velocity in cultured cardiomyocytes. In mouse hearts in vivo, optical mapping showed that electrical propagation was impaired in

Fig. 4 AREG facilitates gap junctional intercellular communication by stabilizing Cx43 gap junctions. **a** Schematic of the dye transfer assay. Mouse neonatal cardiomyocytes were cultured with or without cardiac resident macrophages. The cell-impermeable dye was loaded by scraping that caused tearing of the plasma membrane, and the level of dye transfer between cardiomyocytes was evaluated as the distance of dye-stained cells from the scraped edge after 15 min. **b**, and **c** Mouse neonatal cardiomyocytes were cultured with or without WT or *Areg*^{-/-} cardiac macrophages. Shown are representative images of dye-stained cells spreading from the scrape and the relative extent of dye transfer. Effects of macrophage coculture (**b**) and the lack of AREG in cocultured macrophages (**c**) on dye transfer were analyzed. $n = 24$ and 18 , $n = 22$ and 23 (images) $*P < 0.0001$, two-tailed unpaired Student's *t*-test. **d** Effects of AREG (100 ng/mL) and the indicated signal inhibitors (10 $\mu\text{mol/l}$, each) on dye transfer. $n = 28$, 28 , 24 , 24 , and 30 ; $*P < 0.05$, post-hoc Dunnett's test. **e**, **f** A *Cx43-GFP* plasmid was transfected into HeLa cells, and the cells were treated for 5 h with or without AREG (100 ng/mL) or AG1478 (10 $\mu\text{mol/l}$), as indicated. In **e**, formation of Cx43-GFP gap junction plaques (white arrows) at the cell-cell borders is shown. Cell borders are marked by dashed lines in upper panels. Scale bars, 20 μm . In **f**, plaque size is shown. $n = 47$, 39 , and 36 ; $**p < 0.01$, $***p < 0.001$, post-hoc Tukey's test comparing cell adhesion between two-cardiomyocytes. **g** Western blotting of *Cx43-GFP* transfected HeLa cells treated for 6 h with AREG and 10 $\mu\text{mol/l}$ AG1478 (A), U0126 (U), or FR180204 (F), as indicated. The positions of unphosphorylated and phosphorylated forms of Cx43 are shown. **h** Schematic model of how amphiregulin derived from cardiac macrophage suppresses arrhythmia. Throughout, box-and-whisker plots represent the median, the first and third quartiles, and the minimum and maximum values.

Areg^{-/-} mice (Supplementary Fig. 12). These results indicate that AREG derived from cardiac macrophages facilitates functional coupling between cardiomyocytes and electrical propagation in the heart.

EGFR is reported to be a low-affinity receptor for AREG^{29–31}. To test whether EGFR signaling mediates the effect of AREG on GJIC, we added an EGFR inhibitor (AG1478) or an inhibitor of MEK (U0126) or ERK (FR180204), two effectors downstream of EGFR. All three inhibitors significantly reduced the level of dye transfer mediated by AREG (Fig. 4c, d). This suggests that AREG secreted from cardiac macrophages induces GJIC between cardiomyocytes through EGFR and that AREG facilitates GJIC between cardiomyocytes at least in part via an EGFR/MEK/ERK pathway. To further elucidate how AREG-mediated phosphorylation of Cx43 enhances GJIC, we established HeLa cells expressing GFP-tagged Cx43 (Cx43-GFP) for live-cell imaging of Cx43. While HeLa cells do not endogenously express Cx43, exogenously expressed Cx43 does form functional gap junctions in these cells³². In the absence of AREG, Cx43-GFP was clustered in a few typical gap junction plaques between cells, which is consistent with the aforementioned report (Fig. 4e)³². After 5 h of AREG treatment, however, Cx43 formed much larger gap junctions at cell-cell contacts (Fig. 4e, f and Supplementary Fig. 13). The EGFR inhibitor AG1478 inhibited the formation of larger Cx43 plaques (Fig. 4e, f and Supplementary Fig. 13). Western blotting of the transfected cells showed that AREG induced phosphorylation of Cx43-GFP, which was blocked by inhibition of EGFR, MEK, or ERK. Thus, EGFR/MEK/ERK is a major pathway leading to Cx43 phosphorylation (Fig. 4g). However, Cx43 is known to be phosphorylated at multiple sites by multiple signals^{33,34}. To detect additional AREG-mediated pathways leading to Cx43 phosphorylation, we mutated the target phosphorylation sites on Cx43-GFP that correspond to the Akt, PKC, PKA, CK1, and MAPK signaling pathways. The results showed that the sites corresponding to PKA, CK1, and MAPK are required for the full phosphorylation of Cx43 induced by AREG (Supplementary Fig. 14). Additional mutant analysis of phosphorylation sites targeted by MAPKs revealed the MEK/ERK pathway may be essential for Cx43 phosphorylation (Supplementary Fig. 14).

Among EGFR ligand genes, including *Areg*, *Hbepf*, *Tgfa*, *Egf*, *Ereg*, *Btc*, and *Epg*, cardiac macrophages expressed only *Areg* and *Hbepf* (Supplementary Fig. 15). While AREG promoted phosphorylation of Cx43, HBEGF did not (Supplementary Fig. 14). Similarly, while AREG increased the conduction velocity of cardiomyocyte contraction, HBEGF did not (Supplementary Fig. 11). These results show that the two EGFR ligands produced by cardiac macrophages exhibit differential actions in cardiomyocytes. Taken together, these

findings indicate AREG induces Cx43 phosphorylation via an EGFR/MEK/ERK pathway, which facilitates formation of stable gap junctions, thereby promoting intercellular communication (Fig. 4h).

Discussion

Cardiac arrhythmias are a primary contributor to sudden cardiac death in patients with heart disease³⁵ and preventing sudden cardiac death remains a major unmet medical need. It has been suggested that altered expression of various genes, including genes related to ion channels and gap junctions^{36,37}, contributes to the arrhythmogenicity in heart failure patients. The present study demonstrates that AREG derived from cardiac macrophages maintains electrical impulse conduction by controlling the alignment of Cx43 at gap junctions between cardiomyocytes. Phosphorylation of Cx43 by AREG facilitates intramyocardial conduction by promoting formation of large gap junctions. Previous studies showed that in a normal heart, Cx43 proteins primarily exist in a phosphorylated state and that ischemia induces dephosphorylation²⁰. Our finding that gap junctions are disorganized in *Areg*^{-/-} mice suggests that dysregulation of Cx43 phosphorylation is an important mechanism underlying severe arrhythmia and sudden death induced by RV stress. That said, functions other than Cx43 regulation, such as regulatory roles in other processes in cardiac physiology and stress response⁵, may also be involved in AREG's antiarrhythmic action.

Hulsmans et al. recently reported that macrophages contribute to cardiac conduction in the AV node by directly connecting to cardiomyocytes through Cx43-containing gap junctions³⁸. While *Cx43* deletion from macrophages prolonged AV conduction, it did not cause advanced AV block, suggesting that additional mechanisms, such as AREG-mediated facilitation of intramyocardial electrical coupling, are critically involved in AV nodal conduction¹⁴. More importantly, cardiac macrophages exist throughout the myocardium, and our finding that *Areg* deletion generates substrates for ventricular arrhythmia suggests that macrophages and AREG are required for maintenance of myocardial electrical conduction widely throughout the heart.

In addition to systemic and hematopoietic loss of *Areg*, acute induction of macrophage damage using clodronate liposomes made mice highly susceptible to severe arrhythmia and sudden death. This indicates that macrophages are required not only to establish gap junctional communication, but also to continuously and dynamically regulate gap junctions, particularly during cardiac stresses, such as acute RV pressure overload. AREG supplementation prevents sudden death, which also highlights the dynamic nature of gap junctional regulation and suggests that cardiac macrophages and AREG are potential therapeutic targets for both long-term and acute treatment of life-threatening

arrhythmias. Notably, Son et al. reported that *Areg* expression is reduced in LV myocardial tissues of patients who suffered sudden cardiac death³⁹, suggesting that AREG is also important for preventing severe arrhythmias in humans.

Methods

Animal studies. Male C57BL6/J mice were purchased from CLEA Japan and Jackson Laboratory and maintained on a standard mouse chow diet under sterile barrier conditions, on a 12-h light-dark cycle with 18–23 °C and 40–60% humidity. Male *Areg* homozygous null (*Areg*^{-/-}) mice were backcrossed to C57BL6/J mice over ten generations⁵. *Rag2*^{-/-}, *Cd4*^{-/-}, and *Cd8a*^{-/-} mice were purchased from Taconic (Germantown, NY). The genotypes of all mice were determined using genomic PCR. The isoproterenol challenge entailed bolus administration of isoproterenol (Sigma) 5 mg/kg intraperitoneally under constant ECG recording¹⁷. Clodronate liposomes and control liposomes were purchased from LIPOSOMA B. V. A total of 10 µl of clodronate or control liposome solution per gram of mouse were intravenously administered 24 h before PAB. After the first administration of clodronate or control liposomes, the same volume of liposomes was also administered every 7 days for long term depletion, per the manufacturer's instructions. All protocols for animal experiments were approved by the Animal Care and Use Committee of the University of Tokyo.

Cell culture. Mouse cardiomyocyte cultures were prepared from one- to three-day-old male and female C57BL6/J mouse pups using a neonatal heart dissociation kit (Miltenyi Biotec GmbH, Bergisch Gladbach, Germany). Cardiac macrophages were depleted from primary cultures of cardiomyocytes using anti-CD11b microbeads (Miltenyi Biotec GmbH, Bergisch Gladbach, Germany) with an AutoMACS[®] Pro Separator according to the manufacturer's instructions.

The separated cells were maintained in a 5% CO₂ incubator at 37 °C on collagen-coated 35-mm culture dishes in high glucose Dulbecco's modified Eagle's medium (DMEM; Gibco, 11995-065, USA) supplemented with 10% fetal bovine serum (FBS; HyClone, SH30396.03, USA), 100 IU/ml penicillin and 100 µg/ml streptomycin. The cell cultures were used for scrape loading/dye transfer assays when they reached confluence.

Human epithelioid cervix carcinoma cells (HeLa cells, ATCC, CCL-2) were cultured in DMEM (Gibco, 11995-065, USA) supplemented with 10% FBS, 100 U/ml penicillin, 100 µg/ml streptomycin, and 2 mmol/l glutamine. The cells were split the day before transfection at 70–90% confluency. The cells were transfected using LipofectAMINE 2000 (Life Technologies, 11668-019, USA) according to the manufacturer's instructions. A total of 10 µmol/l AG1478 (Wako, 017-20151, Japan), UO126 (Abcam, ab120241, USA) or FR180204 (Sigma, SML0320, USA) was added 1 h before supplementation with the indicated concentrations of recombinant mouse AREG protein (989-AR/CF, R&D systems, Minneapolis, MN, USA). The transfected cells were visualized using confocal microscopy (LSM 5100 Meta, Carl Zeiss, Germany). For analysis of the effects of EGFR ligands on Cx43 phosphorylation (Supplementary Fig. 3), AREG (100 ng/mL), HBEGF (10 ng/mL, SRP6050, Sigma, USA), or TGF- α (100 ng/mL, 239-A-100, R&D systems, Minneapolis, MN, USA) were added to the cells for 30 min.

Scrape loading/dye transfer assay. GJIC levels were determined using the scrape loading/dye transfer technique as previously reported, with minor modifications²⁸. Briefly, cardiomyocytes were grown to confluence in 35-mm culture dishes. For coculture with cardiac-resident macrophages, sorted cardiac CD11b⁺F4/80⁺Ly6C^{lo}Ly6G⁻ macrophages (15,000 cells/per well) were added to cardiomyocytes and cultured for 24 h. The cells were scraped in phosphate buffered saline (PBS) containing 1 mmol/l 5(6)-carboxyfluorescein, which can pass through gap junctions. Three min after the scraping, the cells were washed with PBS, and fluorescence images were captured using a EVOS FL auto cell imaging system (ThermoFisher Scientific, Waltham MA, USA). The distance the dye spread from the border of the scratch was measured. At least five images per well were used to assess the extent of the dye transfer. For tests of AREG and signal inhibitors, AREG (100 ng/mL) and AG1478, UO126, or FR180204 (10 µmol/l) were added to the cells and incubated for 30 min before scraping.

Plasmid construction. To construct a green fluorescent protein (GFP)-tagged Cx43 expression plasmid, P132 and Cx43 delta dt GFP were purchased from Cecilia Lo (Addgene Plasmid, USA, #17668 and #17671). P132 was digested with KpnI and BpuI, and Cx43 delta dt GFP was digested with KpnI and XbaI. Oligonucleotides were used to link the full-length Cx43 and the *Gfp*-containing backbone. After ligation using T4 DNA ligase (TKR, 6023, Japan), competent DH5a *Escherichia coli* (TKR, 9057, Japan) was transformed with the plasmid, positive colonies were identified and selected, and the insert size was confirmed by gel electrophoresis. Thereafter, the sequence of the cDNA encoding the chimeric protein was verified. The sequence of the oligonucleotides used in these experiments are shown in Supplemental Table 4.

To investigate which amino acids in Cx43 are phosphorylated by AREG, we used a QuikChange Lightning Multi Site-Directed Mutagenesis Kit (Agilent Technologies, California, USA) according to the manufacturer's instructions to

introduce mutations at the consensus phosphorylation sites for the Akt (S373), PKC (S368), cAMP (S365), CK1 (S325/328/330) and MAPK (S255/262/279/282) pathways by substituting the serine residues of Cx43-GFP with alanine. The sequences of the primers used were shown in Supplemental Table 4.

Mouse model preparation. For the BMT experiments, 8-week-old mice were irradiated with a lethal dose (9 Gy). After 24 h, BM cells (1×10^6) isolated from donor mice were injected into the recipient mice through the tail vein. The donor BM was harvested in DMEM, filtered through a 100 µm cell strainer (Greiner Bio-One, 542000, Austria), washed in fresh DMEM with 10% FBS, and resuspended in PBS.

For the PAB experiments, the surgery was performed largely as previously described⁴⁰. We optimized procedures to minimize perioperative death and gain consistent RV pressure overload. Briefly, 8- to 12-week-old mice were anesthetized with intraperitoneal urethane, after which anesthesia was maintained with 2% isoflurane. After orotracheal intubation, the mice were mechanically ventilated. An incision was made in the second left intercostal space using sterile techniques. The pulmonary artery was isolated, and a suture ligature was placed around the vessel. The suture was tied against a 23-gauge needle that was then rapidly removed. After the chest was closed, the mouse was extubated and allowed to recover from the anesthesia. Control mice underwent sham thoracotomies. The RV pressure was around 30 mmHg after PAB when measured in several mice using a pressure catheter (FTH-1211B-0018, Primetech Corporation, Japan).

For repeated AREG administrations to mice, 5 µg recombinant mouse AREG protein (989-AR/CF, R&D systems, Minneapolis, MN, USA) dissolved in 50 µl of PBS was intraperitoneally administered to *Areg*^{-/-} mice 3 times a week, as previously described⁵. For continuous administration of AREG to mice, 5 µg/day of AREG was administered using osmotic minipumps (Alzet, Cupertino, CA, USA).

Immunohistochemistry. Cx43 staining was accomplished using anti-Cx43 antibodies (C6219, Sigma, 1:2000) and detected using a DAB system (R&D Systems, Minneapolis MN, USA). The images were acquired using a FSX1000 (Olympus, Tokyo, Japan). Quantification of Cx43 present at gap junctions was evaluated by measuring the area of Cx43 at gap junctions and total Cx43 using Image J 1.53a software (NIH, USA).

Mice were perfused with PBS and then fixed overnight in a formalin- and methanol-based fixative solution (Ufix, Sakura Fine Tech, Japan). The following day, the fixed tissues were dehydrated and embedded in paraffin. To quantify cardiac fibrosis, we stained heart sections with Picrosirius red.

Paraffin-embedded fixed heart tissues were cut into 5-µm-thick sections, after which the sections were dewaxed, the antigens were retrieved using microwave antigen retrieval (pH 8.0), and the sections were incubated first with mouse anti-N-cadherin antibody (33-3900, ThermoFisher, USA, 1:500) and rabbit anti-CX43 antibody (C6219, Sigma, USA, 1:2000), and then with Alexa-Fluor-488-conjugated anti-mouse-IgG (A-11001, ThermoFisher, USA, 1:2000) and Alexa-Fluor-635-conjugated anti-rabbit-IgG (A-31576, ThermoFisher, USA, 1:2000). The heart sections were visualized using confocal microscopy (LSM 5100 Meta, Carl Zeiss, Germany).

Western blotting. Hearts were removed from the experimental mice and flash-frozen in liquid nitrogen. The tissues were minced using a BioMasher[®] II (Nippi, 300-95441, Tokyo, Japan), after which the cell pellets were lysed with RIPA buffer containing a protease inhibitor cocktail (COMPLETE, EDTA-free, Roche, USA). After protein concentrations were determined using a BCA protein assay kit (Pierce, 23227, Rockford IL, USA), protein samples were separated using SDS-PAGE and transferred to nitrocellulose membranes (BioRad, 1620112, Hercules CA, USA). In the dephosphorylation experiments, the samples were treated for 1 h with lambda protein phosphatase (1 U/µl, sc-200312A, Santa Cruz Biotechnology, USA) before being loaded onto the gel. The following primary antibodies were used for western blotting: rabbit anti-Cx40 polyclonal antibodies (AB1726, Merck Millipore, USA, 1:500), rabbit anti-Cx43 polyclonal antibodies (C6219, Sigma, USA, 1:2000) and rabbit anti-Cx45 polyclonal antibodies (AB1745, Merck Millipore, USA, 1:2000). Mouse anti- α -tubulin monoclonal antibodies (T6199, Sigma, USA, 1:1000) were used to normalize expression. Secondary goat anti-rabbit IgG-HRP antibodies (Cell Signaling Technology, 7074, USA, 1:5000) or goat anti-mouse IgG-HRP antibodies (Cell Signaling Technology, 7076, USA, 1:5000) were used as probes, and peroxidase activity was detected using Clarity[™] Western ECL blotting substrate (Bio-Rad, 170-5061, USA) and an ImageQuant[™] LAS 4000mini (GE Healthcare, Piscataway NJ, USA).

Long-term ECG monitoring. Long-term ECGs were recorded from experimental mice using implanted telemetry devices (Softron, Tokyo, Japan). Under intraperitoneal anesthesia, a radiofrequency transmitter was inserted into a subcutaneous tissue pocket. The leads were tunneled subcutaneously and fixed to the pectoral muscle in the lead II configuration. The experiments were performed 1 week after the surgical instrumentation. The telemetry data were recorded continuously through a receiver placed under the mouse cage. LabChart pro version 8 (AD Instruments, Australia) was used for beat-to-beat analysis to detect arrhythmias manually. VT/VF were diagnosed based on Lambeth Convention guidelines⁴¹.

Optical action potential mapping. We used a high-resolution optical mapping system to examine the electrophysiological properties of the heart. Details of the system and the mapping procedure were described previously⁴². In brief, isolated mouse hearts were continuously perfused on a Langendorff apparatus with modified Krebs-Ringer solution equilibrated with 95% O₂-5% CO₂ (37 °C, pH 7.4). The hearts were then stained with the voltage-sensitive dye 4- β -[2-(di-*n*-butylamino)-6-naphthyl]vinyl]pyridinium (di-4-ANEPPS). We used premature stimulation or burst pacing to induce VT/VF. After rapid stimulation at an S1-S1 interval of 100 ms, a single premature stimulation (S2) was applied, and the sequence was repeated with progressive shortening of the S1-S2 interval. When VT/VF was not induced, burst pacing was applied. In addition, VT/VF induction was also tested in the same protocol under 3 μ M isoproterenol.

Conduction velocity (CV) was measured as previously described⁴³. Briefly, CV was measured during constant stimulation of the center of the LV free wall using a basic cycle length of 150 ms. The longitudinal (L) direction of propagation was determined from the activation map so that it crossed the most widely spaced isochrones. A second line for the transverse (T) direction of propagation was drawn perpendicular to the first line through the most densely spaced isochrone.

Echocardiography. Mice were anesthetized with isoflurane (2–3% for induction; 0.25–2% for maintenance) and set in a supine position. Transthoracic echocardiography was recorded using a Vevo 2000 imaging system (VisualSonics). Pulmonary artery peak velocity, right ventricular fractional shortening, and right ventricular wall-thickness were measured as previously reported⁴⁴.

Flow cytometric analysis and cell sorting. Samples for flow cytometry were prepared as previously described⁵. Briefly, mice were anesthetized, and the heart was exposed and perfused with 10 ml PBS from the left ventricle. Thereafter, the heart tissue was mechanically minced using a scalpel, and the tissue was incubated in Dulbecco's modified Eagle's medium (DMEM) containing 1% elastase (Worthington Biochemical) for 120 min at 37°C. During the incubation, the cells in the suspension were dissociated by sequentially passing the extract through 20-, 21- and 23-gauge needles at 30-min intervals. To further dissociate the cells, they were passed through a 23-gauge needle three times and then filtered through a 40- μ m cell strainer (BD). The cells were then centrifuged at 300 \times g for 5 min, washed with PBS, and resuspended in FACS buffer (PBS supplemented with 1% FBS). After removing erythrocytes in BD PharmLyse solution (BD), the isolated cells were stained with fluorochrome-conjugated antibody and subjected to flow cytometric analysis and cell sorting using a FACS Aria III instrument (BD). Data were acquired on FACSDIVA v8.0.2 (BD) and analyzed using FlowJo (Tree Star). Cells isolated for RNA purification were fixed before staining with antibodies for flow cytometric analysis, as previously reported⁴⁵. FACS-sequential gating strategies were shown in Supplementary Figure 16.

Antibodies. For flow cytometric analysis, anti-CD45.2-V500(562129, BD bioscience, 1:100), anti-CD45.2-APC(109814, BioLegend, 1:100), anti-CD4-PerCP/Cy5.5(100433, BioLegend, 1:100), anti-CD8a-PerCP/Cy5.5(100733, BioLegend, 1:100), anti-CD11b-PerCP/Cy5.5(45-0112-80, eBioscience, 1:100), anti-Ly6g-PerCP/Cy5.5(127615, BioLegend, 1:100), anti-B220-PerCP/Cy5.5(103235, BioLegend, 1:100), anti-CD11b-BV421(101251, BioLegend, 1:100), anti-CD64-APC(139306, BioLegend, 1:100), anti-F4/80-PE(123110, BioLegend, 1:100), anti-Ly6c-PE-Cy7(128018, BioLegend, 1:100) were used. For western blotting, anti-Cx40 (AB1726, Merck Millipore, 1:500), anti-Cx43(C6219, Sigma, 1:2000), anti-Cx45 (AB1745, Merck Millipore, 1:2000), anti- α -tubulin(T6199, Sigma, 1:1000), anti-rabbit IgG-HRP(7074, Cell Signaling Technology, 1:5000), anti-mouse IgG-HRP(7076, Cell Signaling Technology, 1:5000) were used. For immunohistochemistry, anti-F4/80(MCA497G, Serotec, 1:500), anti-Ly6G(127601, BioLegend, 1:500), anti-B220 (103201, BioLegend 1:500), anti-CD3(GTX42110, GeneTex, 1:500), anti-Cx43(C6219, Sigma, 1:2000), anti-N-cadherin(33-3900, ThermoFisher, 1:500) were used. The second antibodies used were Alexa-Fluor-488-conjugated anti-mouse-IgG(A-11001, ThermoFisher, 1:2000) and Alexa-Fluor-635-conjugated anti-rabbit-IgG(A-31576, ThermoFisher, 1:2000).

Video microscopy and propagation analysis of cultured cardiomyocytes.

Video images of cultured cardiomyocytes were recorded as sequential phase-contrast images with a 4x objective at a frame rate of 150 frames/s and a resolution of 2048 \times 2048 pixels using a SI8000 cell motion imaging system (Sony Corporation, Tokyo, Japan). The conduction velocity of the excitation wave was calculated by the SI8000 system. Cardiomyocytes were harvested from neonatal mouse hearts using Neonatal Cardiomyocyte Isolation Kit, mouse (Miltenyi Biotec). All isolated cardiomyocytes from 14 murine hearts of neonates aged from postnatal day 0 were equally divided into all wells of one 48 wells-dish. After the three days-culture with Dulbecco's Modified Eagle Medium (Gibco, Thermo Fisher SCIENTIFIC) containing 10% fetal bovine serum (GE HealthcareHyClone), L-glutamine, Penicillin and Streptomycin, cardiomyocytes reached confluent. AREG was supplemented after 10 h. serum starvation.

Real-time PCR analysis. Total RNA was purified from cells using a RNeasy plus micro kit (Qiagen) according to the manufacturer's instructions. Quantitative real-

time PCR analyses were carried out using a Lightcycler 480 system (Roche), with 18 S rRNA serving as an internal control. The sequences of the primers used were shown in Supplemental Table 4.

Single-cell transcriptome analysis. FASTQ data (E-MTAB-7376)¹³ were downloaded from Array Express and processed using 10xGenomics Cell Ranger count software and Seurat version 3⁴⁶. We filtered out cells that expressed fewer than 200 unique molecular identifiers (UMI), and the percentage of counts mapped to the mitochondrial genome was \geq 5%. To exclude possible doublets, cells with high UMI counts were also excluded. Datasets (sham of total interstitial cells and *Pdgfra*⁺ cells) were aggregated using Seurat's standard integration procedure with 2000 highly variable genes. The expression matrix was dimensionally reduced using principal component analysis of the corrected integrated gene matrix. Clusters were identified using a graph-based approach with the Louvain modularity optimization algorithm. We employed t-SNE for dimensionality reduction and visualization of our datasets. To view gene expression, the MAGIC algorithm was used to impute drop-out values⁴⁶.

Statistics and reproducibility. Data are presented as box-and-whisker plots showing the median, the first and third quartiles, and the minimum and maximum values. The data were analyzed using either an unpaired Student's *t*-test or ANOVA followed by Dunnett's, Tukey's, or Holm-Sidak multiple comparison test, as indicated. Kaplan-Meier analysis followed by a log-rank test with or without post hoc Bonferroni's correction was used to compare survival between mouse groups. The frequencies of VT/VF induction were compared using the χ^2 test. All the statistical analyses were performed using Microsoft Excel 2019 and GraphPad Prism 8 software (GraphPad Software, Inc., San Diego, CA, USA). Each experiment was repeated independently at least 2–3 times with similar results.

Reporting summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

All data supporting the findings of this study are available from the corresponding author upon reasonable request. All source data are provided. Source data are provided with this paper.

Received: 8 August 2019; Accepted: 27 February 2021;

Published online: 26 March 2021

References

- Ziaeian, B. & Fonarow, G. C. Epidemiology and aetiology of heart failure. *Nat. Rev. Cardiol.* **13**, 368 (2016).
- Dunlay, S. M., Roger, V. L. & Redfield, M. M. Epidemiology of heart failure with preserved ejection fraction. *Nat. Rev. Cardiol.* **14**, 591 (2017).
- Naksuk, N. et al. Right ventricular dysfunction and long-term risk of sudden cardiac death in patients with and without severe left ventricular dysfunction. *Circulation: Arrhythmia Electrophysiol.* **11**, e006091 (2018).
- Swirski, F. K. & Nahrendorf, M. Cardioimmunology: the immune system in cardiac homeostasis and disease. *Nat. Rev. Immunol.* **18**, 733–744 (2018).
- Fujiu, K. et al. A heart-brain-kidney network controls adaptation to cardiac stress through tissue macrophage activation. *Nat. Med.* **23**, 611–622 (2017).
- Oishi, Y. & Manabe, I. Macrophages in inflammation, repair and regeneration. *Int. Immunol.* **30**, 511–528 (2018).
- Hofmann, U. & Frantz, S. Role of lymphocytes in myocardial injury, healing, and remodeling after myocardial infarction. *Circ. Res.* **116**, 354–367 (2015).
- Levick, S. P. et al. Cardiac mast cells: the centrepiece in adverse myocardial remodelling. *Cardiovasc Res.* **89**, 12–19 (2011).
- Luitel, H. et al. Pressure overload leads to an increased accumulation and activity of mast cells in the right ventricle. *Physiol. Rep.* **5**, e13146 (2017).
- Weisser, S. B., van Rooijen, N. & Sly, L. M. Depletion and reconstitution of macrophages in mice. *J. Vis. Exp.*, 4105, <https://doi.org/10.3791/4105> (2012).
- Bruhn, K. W., Dekitani, K., Nielsen, T. B., Pantapalankoor, P. & Spellberg, B. Ly6G-mediated depletion of neutrophils is dependent on macrophages. *Results Immunol.* **6**, 5–7 (2015).
- Culemann, S. et al. Locally renewing resident synovial macrophages provide a protective barrier for the joint. *Nature* **572**, 670–675 (2019).
- Farbehi, N. et al. Single-cell expression profiling reveals dynamic flux of cardiac stromal, vascular and immune cells in health and injury. *eLife* **8**, e43882 (2019).
- Kiuchi, M. G. et al. New approaches in the management of sudden cardiac death in patients with heart failure—targeting the sympathetic nervous system. *Int. J. Mol. Sci.* **20**, 2430 (2019).
- Barold, S. S., De Wilde, H. E., Timmers, L., Van Heuverswyn, F. E. & Stroobandt, R. X. Atrioventricular block precipitated by isoproterenol. *Ann. Noninvasive Electrocardiol.* **20**, 397–401 (2015).

16. Brembilla-Perrot, B. et al. Paradoxical effect of isoprenaline infusion. *Europace* **7**, 621–627 (2005).
17. Gonzalez, J. P., Ramachandran, J., Xie, L.-H., Contreras, J. E. & Fraidenraich, D. Selective Connexin43 inhibition prevents isoproterenol-induced arrhythmias and lethality in muscular dystrophy mice. *Sci. Rep.* **5**, 15315–15315 (2015).
18. Beyer, E. C., Paul, D. L. & Goodenough, D. A. Connexin43: a protein from rat heart homologous to a gap junction protein from liver. *J. Cell Biol.* **105**, 2621–2629 (1987).
19. Manjunath, C. K. & Page, E. Cell biology and protein composition of cardiac gap junctions. *Am. J. Physiol.* **248**, H783–H791 (1985).
20. Beardslee, M. A. et al. Dephosphorylation and intracellular redistribution of ventricular connexin43 during electrical uncoupling induced by ischemia. *Circ. Res.* **87**, 656–662 (2000).
21. van Kempen, M. J., Fromaget, C., Gros, D., Moorman, A. F. & Lamers, W. H. Spatial distribution of connexin43, the major cardiac gap junction protein, in the developing and adult rat heart. *Circ. Res.* **68**, 1638–1651 (1991).
22. Remo, B. F. et al. Phosphatase-resistant gap junctions inhibit pathological remodeling and prevent arrhythmias. *Circ. Res.* **108**, 1459–1466 (2011).
23. Hoyt, R. H., Cohen, M. L. & Saffitz, J. E. Distribution and three-dimensional structure of intercellular junctions in canine myocardium. *Circ. Res.* **64**, 563–574 (1989).
24. van Rijen, H. V., van Veen, T. A., Gros, D., Wilders, R. & de Bakker, J. M. Connexins and cardiac arrhythmias. *Adv. Cardiol.* **42**, 150–160 (2006).
25. Duffy, H. S. The molecular mechanisms of gap junction remodeling. *Heart rhythm* **9**, 1331–1334 (2012).
26. Lillo, M. A. et al. S-nitrosylation of connexin43 hemichannels elicits cardiac stress-induced arrhythmias in Duchenne muscular dystrophy mice. *JCI Insight* **4**, <https://doi.org/10.1172/jci.insight.130091> (2019).
27. Abudara, V. et al. The connexin43 mimetic peptide Gap19 inhibits hemichannels without altering gap junctional communication in astrocytes. *Front Cell Neurosci.* **8**, 306–306 (2014).
28. el-Fouly, M. H., Trosko, J. E. & Chang, C. C. Scrape-loading and dye transfer. A rapid and simple technique to study gap junctional intercellular communication. *Exp. Cell Res.* **168**, 422–430 (1987).
29. Shoyab, M., Plowman, G. D., McDonald, V. L., Bradley, J. G. & Todaro, G. J. Structure and function of human amphiregulin: a member of the epidermal growth factor family. *Science* **243**, 1074–1076 (1989).
30. Neelam, B. et al. Structure-function studies of ligand-induced epidermal growth factor receptor dimerization. *Biochemistry* **37**, 4884–4891 (1998).
31. Riese, D. J. et al. The epidermal growth factor receptor couples transforming growth factor- α , heparin-binding epidermal growth factor-like factor, and amphiregulin to Neu, ErbB-3, and ErbB-4. *J. Biol. Chem.* **271**, 20047–20052 (1996).
32. Elfgang, C. et al. Specific permeability and selective formation of gap junction channels in connexin-transfected HeLa cells. *J. Cell Biol.* **129**, 805–817 (1995).
33. Lastwika, K. J., Dunn, C. A., Solan, J. L. & Lampe, P. D. Phosphorylation of connexin 43 at MAPK, PKC or CK1 sites each distinctly alter the kinetics of epidermal wound repair. *J. Cell Sci.* **132**, jcs234633 (2019).
34. Solan, J. L. & Lampe, P. D. Specific Cx43 phosphorylation events regulate gap junction turnover in vivo. *FEBS Lett.* **588**, 1423–1429 (2014).
35. Moss, A. J. et al. Prophylactic implantation of a defibrillator in patients with myocardial infarction and reduced ejection fraction. *N. Engl. J. Med.* **346**, 877–883 (2002).
36. Dupont, E. et al. Altered connexin expression in human congestive heart failure. *J. Mol. Cell. Cardiol.* **33**, 359–371 (2001).
37. Poelzing, S. & Rosenbaum, D. S. Altered connexin43 expression produces arrhythmia substrate in heart failure. *Am. J. Physiol. Heart Circ. Physiol.* **287**, H1762–H1770 (2004).
38. Hulsmans, M. et al. Macrophages facilitate electrical conduction in the heart. *Cell* **169**, 510–522 (2017). e520.
39. Son, G. H. et al. Postmortem mRNA expression patterns in left ventricular myocardial tissues and their implications for forensic diagnosis of sudden cardiac death. *Mol. Cells* **37**, 241–247 (2014).
40. Tarnavski, O. et al. Mouse cardiac surgery: comprehensive techniques for the generation of mouse models of human diseases and their application for genomic studies. *Physiol. Genomics* **16**, 349–360 (2004).
41. Walker, M. J. et al. The Lambeth Conventions: guidelines for the study of arrhythmias in ischaemia infarction, and reperfusion. *Cardiovasc. Res.* **22**, 447–455 (1988).
42. Yamada, C. et al. The renin-angiotensin system promotes arrhythmogenic substrates and lethal arrhythmias in mice with non-ischaemic cardiomyopathy. *Cardiovasc. Res.* **109**, 162–173 (2016).
43. Sato, T. et al. Altered expression of connexin43 contributes to the arrhythmogenic substrate during the development of heart failure in cardiomyopathic hamster. *Am. J. Physiol. Heart Circ. Physiol.* **294**, H1164–H1173 (2008).
44. Cheng, H. W. et al. Assessment of right ventricular structure and function in mouse model of pulmonary artery constriction by transthoracic echocardiography. *J. Vis. Exp.*, e51041, <https://doi.org/10.3791/51041> (2014).
45. Yamada, H. et al. Messenger RNA quantification after fluorescence-activated cell sorting using in situ hybridization. *Cytom. A* **77**, 1032–1037 (2010).
46. van Dijk, D. et al. Recovering gene interactions from single-cell data using data diffusion. *Cell* **174**, 716–729 (2018). e727.

Acknowledgements

We gratefully acknowledge the excellent technical assistance of M. Hayashi, X. Yingda, and N. Yamanaka. This study was supported by the Grant-in-Aid for Scientific Research (20H03672, 19K22615, AMED(19gm6110006h0003) to K.F., 19H03648, 20H04938, 21gm5010002 to I.M. and AMED-CREST(JP20gm0810013), AMED (JP20ek0109440, JP20ek0109487, JP20ek0109406, JP20km0405209, JP20ek021014, JP19bm0804010) to I.K.) and from Takeda Science Foundation and Uehara Memorial Foundation (to I.M.).

Author contributions

J.S., K.F., Y.N., T.M., J.M., T.O., Y.L., Y.M., E.H., and T.K. performed experiments and analyzed the data. J.S., K.F., F.K., I.M., and I.K. wrote the manuscript. K.F. and I.M. designed the study and were involved in the histological studies. J.S., K.F., M.Y., H.S., K.A., A.I., N.T., S.Y., and I.S. were involved in cardiac electrophysiologic studies. K.F. and I.M. directed the project. R.N. and I.K. supervised the overall project.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41467-021-22178-0>.

Correspondence and requests for materials should be addressed to K.F. or I.M.

Peer review information *Nature Communications* thanks Robin Shaw and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021