





Whole-Genome Sequences of *Cronobacter sakazakii* Isolates Obtained from Foods of Plant Origin and Dried-Food Manufacturing Environments

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ABSTRACT Here, we present draft genome sequences of 29 *Cronobacter sakazakii* isolates obtained from foods of plant origin and dried-food manufacturing facilities. Assemblies and annotations resulted in genome sizes ranging from 4.3 to 4.5 Mb and 3,977 to 4,256 gene-coding sequences with G+C contents of \sim 57.0%.

Cronobacter species are Gram-negative opportunistic pathogens associated with life-threatening infections, such as infantile meningitis, septicemia, and necrotizing enterocolitis (1, 2), and urinary tract infections, pneumonia, and wound infections in adults (3, 4). The *Cronobacter* genus consists of seven species, including *C. sakazakii*, *C. malonaticus, C. turicensis, C. muytjensii, C. dublinensis, C. universalis,* and *C. condimenti* (5, 6). The primary pathogen is *C. sakazakii*, and epidemiologically, infections have been linked to consumption of contaminated powdered infant formula. However, surveillance studies have shown that *Cronobacter* spp. are found in a variety of different foods, including dried foods (flour, spices, herbs, and cereal) and fresh ready-to-eat vegetables (7–10). There is a growing body of evidence that plants may serve as a reservoir or ancestral host for *Cronobacter* spp. (11, 12). Although occurrences of *Cronobacter* spp. in foods of plant origin are increasingly being reported, relatively little genomic information on them is available. Here, we report the draft genome sequences of 29 *C. sakazakii* strains isolated from foods of plant origin and dried-food processing environments.

Whole-genome sequencing (WGS) libraries of these *C. sakazakii* strains were constructed using the Nextera XT DNA sample preparation kit (Illumina, San Diego, CA, USA). FASTQ data sets generated on the MiSeq platform (Illumina) were trimmed for *de novo* assembly using CLC Genomics Workbench version 9.0 (CLC bio, Aarhus, Denmark). The genomes were annotated using the Rapid Annotations using Subsystems Technology (RAST) annotation server (13), and 16 sequence types were determined by using the *Cronobacter* multilocus sequence typing (MLST) website (http:// pubmlst.org/cronobacter). The genome sizes and coding sequences (CDSs) of these assemblies ranged from 4.3 to 4.5 Mb and 3,977 to 4,256, respectively, with G+C contents of ~57.0% (Table 1).

All strains harbored a pESA3/pSP291-like virulence plasmid, which was found by comparing their genome assemblies with WGS of *C. sakazakii* BAA-894 (GenBank accession numbers NC_009778 and CP000783) and was confirmed by PCR analysis (14). Four strains also possessed pESA2, and three possessed pCTU3.

Other mobilome-like genes, such as integrase/transposase genes coding for COG0582, Tn7 (TnsA), and TnpA, and 19 to 40 phage-associated proteins, were present in some strains. Interestingly, all strains possessed a fosfomycin resistance (*fosA*) gene. Other genes identified in these strains included genes for multidrug resistance efflux

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TABLE 1 Genomic information and GenBank accession numbers of 29 C. sakazakii isolates obtained from foods of plant origin and dried-
food production environments

Strain	Genome size (kb)	G+C content (%)	No. of CDSs	ST ^a	Source	GenBank accession no.
MOD1_LR631	4,485	56.8	4,134	4, CC4 ^b	Instant oatmeal	PTPF0000000
MOD1_LR632	4,552	56.7	4,225	4, CC4	Environment	PTPE0000000
MOD1_LR634	4,421	56.9	4,118	148, CC16	Dried cut carrots	PTPD0000000
MOD1_LR635	4,428	56.9	4,096	148, CC16	Dried cut carrots	PTPC0000000
MOD1_LR640	4,553	56.7	4,231	1, CC1	Sodium caseinate	PTPB0000000
MOD1_LR654	4,433	57.0	4,061	23, CC23	Adult cereal	PTPA0000000
MOD1_LR707	4,406	57.0	4,055	1, CC1	Org. ^c casein flour	PTOZ0000000
MOD1_LR708	4,441	56.9	4,106	1, CC1	Org. casein flour	PTOY0000000
MOD1_LR715	4,503	56.9	4,179	4, CC4	Org. soy	PTOW0000000
MOD1_LR722	4,489	56.7	4,147	1, CC1	Org. soy	PTOV0000000
MOD1_LR733	4,468	56.9	4,142	1, CC1	Org. casein flour	PTOU0000000
MOD1_LR752	4,363	56.9	4,034	1, CC1	Honey powder	PTOT0000000
MOD1_LR753	4,545	56.7	4,268	31, CC31	Honey powder	PTOS0000000
MOD1_LR757	4,576	56.8	4,256	13, CC13	Org. casein flour	PTOR0000000
MOD1_1-15	4,459	56.9	4,086	136	Nuts	NITJ0000000
MOD1_3-21	4,420	56.9	4,069	1, CC1	Nuts	NITK0000000
MOD1_5-17G	4,422	56.9	4,071	226, CC8	Nuts	NITM0000000
MOD1_5-20G	4,567	56.7	4,215	17, CC17	Nuts	NITL0000000
MOD1_5-21G	4,569	56.8	4,230	17, CC17	Nuts	NIXM0000000
MOD1_KW1	4,539	56.8	4,155	93	Barley	NITI0000000
MOD1_KW4	4,542	56.8	4,233	73, CC73	Dried seaweed	NITG0000000
MOD1_KW11	4,446	56.8	4,085	6 alleles ^d	Black bean	NITE0000000
MOD1_KW18	4,438	57.0	4,095	156, CC21	Mushroom	NITC0000000
MOD1_777122	4,375	57.0	4,057	198, CC52	Hulled sesame seed	PTOQ0000000
MOD1_760029	4,508	56.8	4,197	40, CC40	Whole grain, corn	PTOP0000000
MOD1_16MP002184	4,564	56.6	4,238	40, CC40	Chocolate org. shake	PTOO0000000
MOD1_16MP002185	4,508	56.8	4,168	3, CC3	Chocolate org. shake	PTON0000000
MOD1_Jor109	4,412	57.0	3,997	6 alleles ^e	Grapes	NITQ0000000
MOD1_WNTSBCO4	4,490	56.8	4,137	1, CC1	Walnut	PTOM0000000

^aSequence type (ST) was determined by uploading genome assemblies to https://pubmlst.org/cronobacter.

^bCC, clonal complex. ^cOrg., organic.

^dThe MLST scheme for strain MOD1_KW11 matched only 6 of the 7 alleles, and no closest match was determined.

eThe MLST scheme for strain MOD1_Jor109 matched only 6 of the 7 alleles, and its closest match was determined to be ST40, CC40.

pump-related proteins belonging to the *acrAB* operon, the resistance-nodulationdivision, the major facilitator superfamily, ABC-type drug transport, and bicyclomycin resistance families. Heavy metal resistance genes and gene clusters involved in copper, hydroperoxide, fusaric acid, and tellurite resistance were also found. An albicidin (a phytotoxin that inhibits DNA gyrase in chloroplasts) resistance protein (11, 15) was observed in all the strains. Furthermore, all strains possessed an operon encoding a xylose utilization pathway, supporting the hypothesis that plants may be the ancestral econiche for *Cronobacter* spp., as posited by Chase et al. (11) and Schmid et al. (12). However, the size of this gene cluster varied among the strains.

These results add to the growing number of genomes of *Cronobacter* strains which have plant origins. The availability of genomic information from these strains will provide a better understanding of the genetic features linked to plant association and expands insights into the evolutionary history of this important foodborne pathogen.

Accession number(s). The *C. sakazakii* genome sequences were submitted to NCBI GenBank under BioProject number PRJNA258403 (*Cronobacter* GenomeTrakr Project, FDA-CFSAN), and their accession numbers are listed in Table 1.

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REFERENCES

- Tall BD, Chen Y, Yan QQ, Gopinath GR, Grim CJ, Jarvis KG, Fanning S, Lampel KA. 2014. *Cronobacter*: an emergent pathogen causing meningitis to neonates through their feeds. Sci Prog 97:154–172. https://doi .org/10.3184/003685014X13994743930498.
- Hunter CJ, Petrosyan M, Ford HR, Prasadarao NV. 2008. Enterobacter sakazakii: an emerging pathogen in infants and neonates. Surg Infect 9:533–539. https://doi.org/10.1089/sur.2008.006.
- Holý O, Petrželová J, Hanulík V, Chromá M, Matoušková I, Forsythe SJ. 2014. Epidemiology of *Cronobacter* spp. isolates from patients admitted to the Olomouc University Hospital (Czech Republic). Epidemiol Mikrobiol Imunol 63:69–72.
- Alsonosi A, Hariri S, Kajsík M, Oriešková M, Hanulík V, Röderová M, Petrželová J, Kollárová H, Drahovská H, Forsythe S, Holý O. 2015. The speciation and genotyping of *Cronobacter* isolates from hospitalised patients. Eur J Clin Microbiol Infect Dis 34:1979–1988. https://doi.org/ 10.1007/s10096-015-2440-8.
- 5. Iversen C, Mullane N, McCardell B, Tall BD, Lehner A, Fanning S, Stephan R, Joosten H. 2008. Cronobacter gen. nov., a new genus to accommodate the biogroups of Enterobacter sakazakii, and proposal of Cronobacter sakazakii gen. nov., comb. nov., Cronobacter malonaticus sp. nov., Cronobacter turicensis sp. nov., Cronobacter muytjensii sp. nov., Cronobacter dublinensis sp. nov., Cronobacter genomospecies 1 and of three subspecies, Cronobacter dublinensis subsp. dublinensis subsp. nov., Cronobacter dublinensis subsp. nov. and Cronobacter dublinensis subsp. lactaridi subsp. nov. and Cronobacter dublinensis subsp. lactaridi subsp. nov. Int J Syst Evol Microbiol 58:1442–1447. https://doi.org/10.1099/ijs.0.65577-0.
- Joseph S, Cetinkaya E, Drahovska H, Levican A, Figueras MJ, Forsythe SJ. 2012. Cronobacter condimenti sp. nov., isolated from spiced meat and Cronobacter universalis sp. nov., a species designation for Cronobacter sp. genomospecies 1, recovered from a leg infection, water and food ingredients. Int J Syst Evol Microbiol 62:1277–1283. https://doi.org/10.1099/ ijs.0.032292-0.
- Berthold-Pluta A, Garbowska M, Stefańska I, Pluta A. 2017. Microbiological quality of selected ready-to-eat leaf vegetables, sprouts and nonpasteurized fresh fruit-vegetable juices including the presence of *Cronobacter* spp. Food Microbiol 65:221–230. https://doi.org/10.1016/j.fm.2017.03.005.

- Sani NA, Odeyemi OA. 2015. Occurrence and prevalence of *Cronobacter* spp. in plant and animal derived food sources: a systematic review and meta-analysis. SpringerPlus 4:545. https://doi.org/10.1186/s40064-015-1324-9.
- Lou X, Si G, Yu H, Qi J, Liu T, Fang Z. 2014. Possible reservoir and routes of transmission of *Cronobacter* (*Enterobacter sakazakii*) via wheat flour. Food Control 43:258–262. https://doi.org/10.1016/j.foodcont.2014.03.029.
- Kandhai MC, Heuvelink AE, Reij MW, Beumer RR, Dijk R, van Tilburg JJHC, van Schothorst M, Gorris LGM. 2010. A study into the occurrence of *Cronobacter* spp. in The Netherlands between 2001 and 2005. Food Control 21:1127–1136. https://doi.org/10.1016/j.foodcont.2010.01.007.
- 11. Chase HR, Eberl L, Stephan R, Jeong H, Lee C, Finkelstein S, Negrete F, Gangiredla J, Patel I, Tall BD, Gopinath GR, Lehner A. 2017. Draft genome sequence of *Cronobacter sakazakii* GP1999, sequence type 145, an epiphytic isolate obtained from the tomato's rhizoplane/rhizosphere continuum. Genome Announc 5:e00723-17. https://doi.org/10.1128/genomeA.00723-17.
- Schmid M, Iversen C, Gontia I, Stephan R, Hofmann A, Hartmann A, Jha B, Eberl L, Riedel K, Lehner A. 2009. Evidence for a plant-associated natural habitat for *Cronobacter* spp. Res Microbiol 160:608–614. https:// doi.org/10.1016/j.resmic.2009.08.013.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Franco AA, Hu L, Grim CJ, Gopinath G, Sathyamoorthy V, Jarvis KG, Lee C, Sadowski J, Kim J, Kothary MH, McCardell BA, Tall BD. 2011. Characterization of putative virulence genes on the related RepFIB plasmids harbored by *Cronobacter* spp. Appl Environ Microbiol 77:3255–3267. https://doi.org/10.1128/AEM.03023-10.
- Hashimi SM, Wall MK, Smith AB, Maxwell A, Birch RG. 2007. The phytotoxin albicidin is a novel inhibitor of DNA gyrase. Antimicrob Agents Chemother 51:181–187. https://doi.org/10.1128/AAC.00918-06.