



Original Article

Enteroparasitism and Risk Factors Associated with Clinical Manifestations in Children and Adults of Jalisco State in Western Mexico

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ABSTRACT

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Objectives: To determine the prevalence and risk factors associated with intestinal parasites in the population of San Juan Cosala, Jalisco, Mexico.

Methods: A total of 277 samples from 104 participants were analysed using direct smear, flotation, formaldehyde/ethyl acetate, and modified Kinyoun's acid-fast stain methods. The Graham method was applied only for samples from children under 12 years of age for the diagnosis of *Enterobius vermicularis*.

Results: The prevalence of parasite infections in the study population was 77.9% including: *Entamoeba histolytica*/*E. dispar*/*E. moshkovskii*/*E. bangladeshi* (37.5%), *Giardia intestinalis* (11.5%); commensals: *Endolimax nana* (44.2%), *Entamoeba coli* (27.9%), *Chilomastix mesnili* (6.7%) and *Iodamoeba bütschlii*, (2.9%); emerging intestinal protozoans: *Blastocystis* spp. (49%), *Cryptosporidium* spp. (7.7%) and *Cyclospora cayetanensis* (2.9%); and helminths: *Enterobius vermicularis* (18.3%) and *Ascaris lumbricoides* (5.8%). The results also showed that 58.64% of the studied population presented polyparasitism. A significant association was found between protozoan infections and housewives, and houses that were not built with concrete ceilings, brick walls and cement floors ($p < 0.05$).

Conclusion: Polyparasitism was observed in over half the study population. The most prevalent parasite was *Blastocystis* spp, whilst the prevalence of helminths was less than that of protozoans. The risk factors for infection to intestinal parasites were being a housewife and not having solid brick, cement and concrete materials for house construction.

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Introduction

Intestinal parasite infections are a public health problem worldwide, especially in developing countries, and in children under 5 years old [1]. Parasitic infections are amongst the top 20 causes of illness in Mexico with approximately 53% of the general population diagnosed with enteroparasites [2]. A study conducted in 2 states of Mexico (Sonora and Oaxaca) showed a prevalence of 47.2% in children [3]. The lack of knowledge to avoid transmission, poor hygiene, homes with lack of sanitation

facilities, low family income, lower levels of education, overcrowding, living with animals and lower consumption of haem iron are factors that generate greater risk of acquisition of intestinal parasites [2,4,5].

In the state of Jalisco, Mexico, there have been few studies on the prevalence of intestinal parasites. A study was conducted in the highlands area in the town of Arandas, showing similar prevalence as reported in Sonora and Oaxaca [3,4]. In the Cienega area in the town of San Pedro Itzicán, the prevalence of intestinal parasites was 60%. Egg counts were determined

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to establish treatment regimens and consequently its effectiveness (at a follow up appointment). San Juan Cosala, as well as San Pedro Itzicán, has a population located on the shores of Lake Chapala where there is a lack of sanitation facilities, thus increasing the risk of infection with intestinal parasites [6]. Information regarding prevalence, clinical manifestations and risk factors are of great importance in determining the necessary healthcare response. This is the first study to perform research on intestinal parasites in this location.

Materials and Methods

1. Study area and population

This study was performed in San Juan Cosala, a small town located on the lakeshore of Chapala, Jalisco state, Mexico, Global Positioning System (GPS) coordinates: longitude 103°19'56.90 W, latitude 20°17'04.50" N. For this region, the average height above sea level is 1,530 metres, the town has a moderate and cool climate, and the rainfall and temperature ranges are 192 to 760 mm, and 19.5 to 30°C, respectively (Figure 1).

The town has 6,973 inhabitants, 48.76% of which are male and 51.24% are female. In this study, a total of 104 people with ages ranging from 1 to 83 years old were eligible to participate

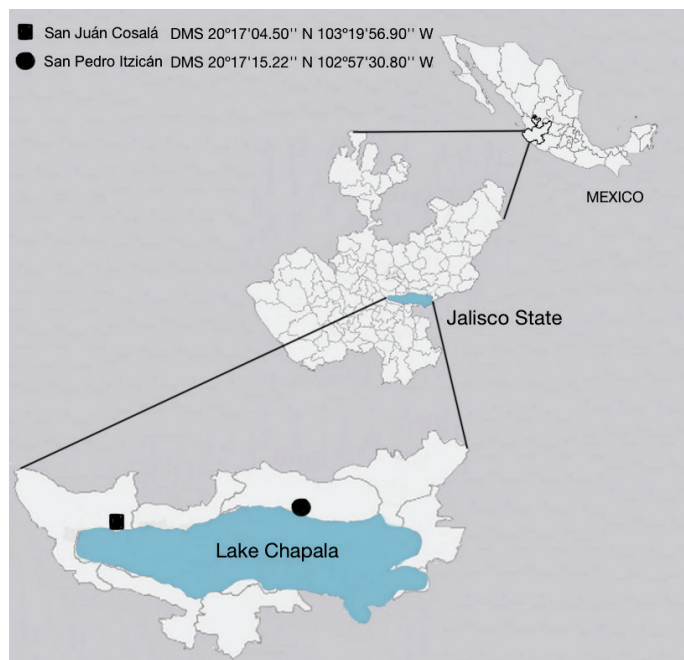


Figure 1. Map of location of small-town San Juan Cosala, situated on the banks of Lake Chapala, Jalisco state, Mexico, Global Positioning System (GPS) coordinates longitude: 103° 19'56.90 W, latitude 20°17'04.50" N.

in this study. The population lives in an overcrowded environment with poor hygiene and lack of sanitation. There are 1,325 houses; 1.3% of these have no sanitary facilities, 0.8% do not have electric lights, 5.5% have no potable water, 5.9% have dirty floors, 29% have one bedroom and 13% do not have a refrigerator. All of the study participants were beneficiaries of the Feed Operation Project, which is a civil association that helps poor people with food pantries, health information, some schooling and cultural programmes for the community of San Juan Cosala, Jalisco.

2. Questionnaire

A structured questionnaire with open ended questions was used to collect information from the adults and parents of children regarding housing, personal hygiene and clinical manifestations. These data included information on age, gender, education, residence and number of household occupants, occupation, community residence site, and housing conditions (number of rooms and bathrooms). Characteristics of housing construction were analysed as a risk factor for infection with intestinal parasites. The aspects of the house included: ceilings, floors, walls and public services (sewer system, electric lighting, and plumbing). Health conditions that were analysed covered a history of symptoms including nausea, vomiting, diminution of appetite, abdominal and epigastric pain, watery, mucous or bloody diarrhea, flatulence, constipation, tenesmus, anal pruritus, and distended abdomen. These conditions were studied in the population and reported along with frequency (daily, occasionally or never) for analysis with parasite tests (+/-). The questionnaire was performed, applied and reviewed by researchers and medical doctors from the Health Centre of that population.

3. Samples and laboratory methods

From 104 participants, samples were obtained on 3 consecutive days; 11 only delivered 1 sample, 22 delivered 2 samples and 74 delivered 3 samples, for a total of 277 samples.

3.1. Laboratory methods

Each stool sample met the acceptance criteria of the pre-analytical phase i.e. free of urine, soil, water and any contamination. The samples were transported and stored at 4°C. Then, samples were examined by direct fecal smear and iodine solution emulsification, concentration by flotation (zinc sulphate), and concentration by sedimentation (formaldehyde/ethyl acetate) [7].

3.1.1. Direct smear

One drop of 5% iodine was placed on a slide. Approximately 2 mg of stool sample was smeared and then thoroughly im-

mersed in the iodine solution. A coverslip was placed on top of the suspension.

3.1.2. Flotation (zinc sulphate)

Four grams of fresh stool was transferred into 10 mL of 10% formalin and was mixed and fixed for 30 minutes. The mixture was filtered through a gauze. Then, 15 mL 0.85% NaCl was added to each tube and centrifuged for 10 minutes at 500 x g. The supernatant was decanted, and the sediment was suspended in 15 mL of zinc sulphate. This was centrifuged for 1 minute at 500 x g, 1 drop of the surface film and 1 drop of sediment were placed onto a slide, and a drop of iodine solution was added and a coverslip was placed on top of each preparation.

4. Formaldehyde/ethyl acetate

Four grams of fresh stool was transferred into 10 mL 10% formalin, and the suspension mixed and fixed for 30 minutes. The mixture was filtered through a gauze and transferred to a 15 mL conical centrifuge tube. A 0.85% NaCl solution was added to make the volume up to 15 mL and centrifuged for 10 minutes at 500 x g. The supernatant fluid was decanted, and the sediment was suspended in 10% formalin. Then, 4 to 5 mL of ethyl acetate was added, and the mixture was shaken vigorously and centrifuged for 10 minutes at 500 x g. The plug of debris was decanted with an applicator stick; the top layers of supernatant were decanted and the sediment was observed.

In the methods described above, the iodine solution was used to detect and identify the parasites with optical microscopy at 10× and 40× magnification.

5. Modified Kinyoun's acid-fast stain

One drop of specimen was smeared onto the slide and allowed to air dry. The samples were then fixed with absolute methanol for 1 minute and stained with Kinyoun's carbol fuchsin for 5 minutes. The samples were rinsed with 50% ethanol and water. Then, the slides were decolourized using 1% sulphuric acid for 2 minutes and rinsed with water and then drained. The samples were stained with methylene blue for 2 minutes and rinsed with water and air dried, and a coverslip was added. The preparation was covered with oil immersion and examined with a 100× objective. Additionally, mothers were instructed to collect 3 perianal scrapings from children under 12 years old to perform the diagnosis of *E. vermicularis* by Graham's method ($n = 52$). These samples were taken in the morning before defecation (as long as no ointments, talc, creams or oils had been applied in the perianal area) [7]. These samples were transported at room temperature in a plastic bag to the Municipal Health Centre of San Juan Cosala.

6. Graham method

The sample was obtained from the perianal area, holding the tape and the slide against the tongue depressor and pressing against the right and left perianal folds. The tape on the slide was placed (with the adhesive side now on the glass) and pressed firmly into position, and the sample was observed. These samples were observed with light microscopy at 10× and 40× magnification.

7. Statistical analysis

The quantitative variables were expressed as mean, standard deviation, and standard error. The qualitative variables were expressed through frequency and percentage and were entered into 2 x 2 contingency tables indicating the positive and negative parasite test results for individuals for each risk factor. Chi-square (X^2) and Fisher's exact tests were used to establish statistical significance. Multiple logistic regression analysis was used when the risk factor was calculated by means of the odds ratio (OR) with a confidence interval of 95% ($_{95}$ CI). The statistical analysis was performed using SPSS software version 20.0 (IBM Corp., Armonk, NY, USA) and Epi-Info. A $p < 0.05$ was considered to be significant.

8. Ethics approval

The project was approved by the Institutional Research, Ethics, and Biosecurity Committee recorded under registration number CI 053-2014, University Centre of Health Sciences, University of Guadalajara, Sierra Mojada # 950 Colonia Independencia, Guadalajara, Jalisco, Mexico. The study was performed in accordance with Official Mexican Norm NOM-087-ECOL-SSA1-2002. All patients who tested positive for enteroparasites were referred to the Municipal Health Centre for their respective treatment.

Results

The prevalence of individuals infected with intestinal parasites in the study population was 77.9% (81/104). The most prevalent intestinal parasites were the protozoans, *Blastocystis* spp. 49% (51/104) and *Endolimax nana* 44.2% (46/104). *Cyclospora cayetanensis* and *Iodamoeba bütschlii* were the least prevalent intestinal parasite, with an incidence of 2.9% (3/104), (Figure 2).

The rate of monoparasitism was 19.23% (20/104), with 58.64% (61/104) of the population having polyparasitism. In relation to the number of parasites, a higher prevalence of protozoans than helminths was observed in children (Figure 3). When the study population was divided into 2 age groups, children (< 18 years old) and adults (≥ 18 years old), there

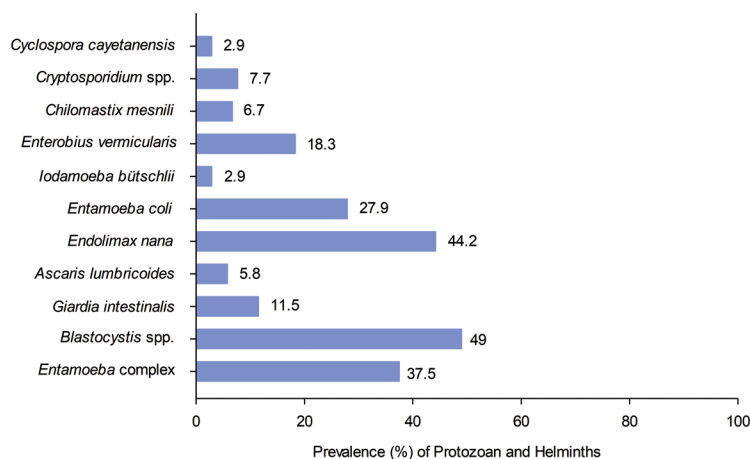


Figure 2. It shows the prevalence by parasitic species, the most prevalent protozoan was *Blastocysts* spp.

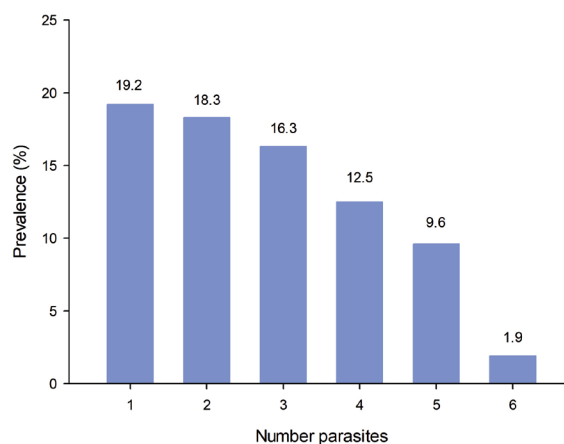


Figure 3. The polyparasitism rate was 58.64% in the population of San Juan Cosala, analysed by the number of parasites; up to 6 parasites were found simultaneously.

Table 1. Socio-demographic characteristics and association with intestinal parasites in the population of San Juan Cosala.

Variable	Number	Positives	%	X ²	p	OR	95% CI
Age (y)							
< 18	56	46	82.1	1.27	0.259	1.70	0.67-4.35
≥ 18 (Adult)	48	35	72.9				
Occupation							
Housewife	30	25	83.3	4.6	0.039	4.37	1.08-17.68
Other	31	21	53.3				
Student	43	35	81.3	1.8	0.083	0.48	0.15-1.4
Gender							
Female	62	47	75.8	0.385	0.535	0.73	0.28-1.94
Male	42	34	81.0				

was no statistically significant difference in the prevalence of parasites ($X^2 = 1.277$). The distribution of parasitism by age group is shown in (Table 1).

There was also no statistically significant difference in the presence of parasites according to gender ($X^2 = 0.385$, $p = 0.535$; Table 1).

There was a significantly higher incidence of housewives with intestinal parasites (83.3%), compared with the incidence in the group consisting of other occupations (53.3%, $X^2 = 4.602$, $p = 0.039$). The relative risk of infection was 4.375 times higher for housewives than for other occupations to acquire intestinal parasites (Table 1).

Personal hygiene (washing hands before eating and after going to the bathroom; habit vs no habit) and overcrowding

were analysed between participants. The people who lived in an overcrowded environment had a higher prevalence of infection (79%) than those who did not live in an overcrowded environment (19/25), 76 %, nevertheless, this difference was not significant (Table 2).

The presence of rodents and insects (such as rats, flies, and cockroaches) inside the house was analysed with a positive intestinal parasite test. No difference was observed between those who had rats, flies and/or cockroaches compared with those households that did not (Table 2).

The highest prevalence of intestinal parasites was in individuals whose house did not have a concrete ceiling or cement floor, which was statistically significant ($p < 0.01$). In addition, the relative risk of intestinal parasites was 6.1 times

Table 2. Personal hygiene, presence of rodents, insects inside the house and association with enteroparasites.

Variable	Number	Positives	%	X ²	p	OR	95% CI
Washing hands before eating and after going to the bathroom							
No	21	17	81.0	0.144	0.959	1.262	0.38-4.20
Yes	83	64	77.1				
Overcrowding							
Yes	78	62	79.5	0.137	0.711	1.224	0.42-3.57
No	25	19	76.0				
Rodents and insects inside the house							
Yes, rats	55	43	78.2	0.006	0.938	1.037	0.41-2.62
No, rats	49	38	77.6				
Yes, flies	66	51	77.3	0.039	0.843	0.907	0.34-2.39
No, flies	38	30	78.9				
Yes, cockroaches	88	68	77.3	0.124	1.000	0.785	0.20-3.03
No, cockroaches	16	13	81.3				

Table 3. Construction characteristics and public housing services and association with enteroparasites.

Characteristics	Number	Positive	%	X ²	p	OR	95% CI
Concrete ceiling							
No	34	29	85.3	1.61	0.205	2.008	0.68-5.97
Yes	70	52	74.3				
Cement floor							
No	67	47	91.9	6.54	0.011	0.207	0.06-0.75
Yes	37	34	70.1				
Brick walls or decks							
No	32	30	93.8	6.75	0.009	6.176	1.35-28.21
Yes	72	51	70.8				
Public services							
Sewer System							
No	13	13	100	2.66	0.1029	4.87	0.61-39.02
Yes	91	68	74.5				
Electric lighting							
No	4	4	100	0.15	1.000	1.538	0.61-39.09
Yes	100	77	77				
Plumbing							
No	5	5	100	0.331	0.975	1.87	0.21-16.31
Yes	99	76	76.8				

Table 4. Clinical manifestations and association with enteroparasites in the San Juan Cosala population.

Clinical manifestation	Number	Positive	%	X ²	p	OR	95% CI
Nausea							
No	84	66	81.4	1.11	0.72	0.81	0.26-2.8
Yes	20	15	18.51				
Vomiting							
No	89	70	78.7	0.21	0.64	0.74	0.21-2.9
Yes	15	11	13.5				
Reduced appetite							
No	55	42	76.4	0.15	0.69	1.2	0.46-3.4
Yes	49	39	79.54				
Abdominal pain							
No	45	38	84.4	1.98	0.15	0.49	0.17-1.3
Yes	59	43	72.8				
Epigastric pain							
No	63	53	84.1	3.6	0.51	0.41	0.15-1.06
Yes	41	28	68.29				
Watery diarrhea							
No	66	54	81.8	0.16	0.20	0.54	0.21-1.4
Yes	38	24	63.15				
Mucousy diarrhea							
No	86	68	79.1	0.26	0.52	0.69	0.21-2.4
Yes	18	76	72.22				
Bloody diarrhea							
No	93	74	79.6	1.4	0.22	0.45	0.11-1.9
Ye	11	7	64.6				
Flatulence							
No	66	54	81.8	1.62	0.20	0.54	0.21-1.4
Yes	38	27	71.0				
Constipation							
No	59	48	81.4	0.95	0.32	0.63	0.24-1.6
Yes	45	33	73.3				
Tenesmus							
No	54	45	83.3	1.93	0.16	0.51	0.19-1.3
Yes	50	36	72.0				
Anal itching							
No	61	49	80.3	0.51	0.47	0.71	0.27-1.8
Yes	43	32	74.41				
Abdominal distension							
No	42	32	76.2	0.11	0.73	1.17	0.44-3.0
Yes	62	49	79.0				

higher for people who did not have brick walls and decks compared to those that did (Table 3). Furthermore, a high prevalence of infection was observed in people who did not have access to public services such as a sewer system, electric lighting, and plumbing, nevertheless, there were no statistically significant differences observed (Table 3).

The highest prevalence of intestinal parasites was observed among those people who were asymptomatic. When the presence of 13 symptoms were compared between people who tested positive or negative for parasites, there were no statistically significant differences observed (Table 4).

Discussion

The prevalence of parasitism in San Juan Cosala, Jalisco, Mexico in this study population was 77.9% which was similar to the prevalence reported in Argentina and Venezuela [8,9] and other states of Mexico, including Colima (38%) [10] and San Luis Potosí (30%) [11].

However, one study reported that in the state of Jalisco, adults of San Pedro Itzicán, municipality of Poncitlán (both native and mestizo), showed a 60% prevalence of infection [6] which was lower than the present study. Nevertheless, the incidence of infections for both study populations in Jalisco was high. This may be due to their location on the lakeshore of Chapala where a shared water source that supplies the towns may be contaminated, as well as similar conditions of public services and hygienic habits. The highest prevalence of infection was in San Pedro Itzicán and San Juan Cosalá, compared with Arandas (47.2%). This suggests that environmental factors such as weather and latitude may influence the prevalence of parasites, as Arandas is located at a higher elevation above sea level, and is less warm, therefore, providing geographical reasons for a lower prevalence rate of intestinal parasite infection [4].

Age did not influence the predominance of parasite infection in this study, although young children (4 to 5 years old) were shown to have a high prevalence rate (86.7%) [12], despite the deficient hygienic conditions to which all age groups were exposed. The results obtained in this age group showed a higher prevalence than in those reported in the municipality of Pantepec, Chiapas (62.8%) [13].

The percentage of *Blastocystis* spp. infections in San Juan Cosala (49%) may be influenced by the presence of this microorganism in the Chapala lake region. This population is located on the lakeshore of Chapala, and it is assumed that the local population consumes contaminated water from Chapala lake. This hypothesis would need to be tested with the water from the lake.

The prevalence of *Blastocystis* spp. infection has been

reported globally with 26.9% in Iran [14], 51.3% in Venezuela [15], 57% in Sydney, Australia [16]. In Mexico, for Guerrero and Veracruz states, the prevalence of *Blastocystis* spp. infection was reported as 23.02% and 80%, respectively [17,18], whilst in San Pedro Itzicán, a higher prevalence (60%) was found for *Blastocystis* spp. [6]. The prevalence of *Blastocystis* spp. infection has been shown to be influenced by host susceptibility as well as the type of population, socio-demographic characteristics, risk factors in housing, deficiencies in alimentary hygiene, water contamination, and contact with harmful fauna [19], providing similar results to those in this current study.

Blastocystis spp., is associated with the causes of irritable colon syndrome [20], problems with dialysis patients [21], recipients of renal transplantation [22] and patients with primary and acquired immunodeficiency [23]. However, the pathogenesis is associated with subtypes. In Sonora and Morelia, Mexico, the ST3 and ST1 subtypes were found in children [24].

The differentiation between *Entamoeba histolytica*/*E. dispar*/*E. moshkovskii*/*E. bangladeshi* was not performed because a molecular diagnosis was not used in this study protocol to differentiate between these 4 protozoans. For the patients who reported diarrhea, (77 with mucousy diarrhea and 7 with bloody diarrhea), the most advisable course of action was to determine the species of *Entamoeba* and provide specific treatment as detailed in other studies [25,26]. With respect to the commensal protozoans, *E. nana* (44.2%), *E. coli* (27.9%), *C. mesnili* (6.7%) and *I. bütschlii* (2.9%) were detected. The identification of these types of protozoans is important because they are transmitted through the ingestion of mature cysts via the fecal-oral route, similar to intestinal pathogen protozoans [27]. Another reason is that these protozoans can be indicative of fecal contamination [28].

In this study, the prevalence rate of *Cryptosporidium* spp. infection was 7.7%, which was higher than the percentage reported in Argentina (0.2%) and Vietnam (5%); however, it was less than that reported in Sudan (13.3%) [29-31]. Similarly, in a study performed in San Pedro Itzicán, located on the border of Chapala lake, the prevalence of *Cryptosporidium* spp. was 7% [6]. It is important to consider the high transmissibility of *Cryptosporidium* spp. since the oocysts that are eliminated in feces are immediately infectious for humans and animals. Its zoonotic component should also be considered, because oocysts are highly resistant to chlorine and could be transmitted by inhalation. Autoinfection occurs through the thin layer of the oocyst, which loses its layer in the intestinal epithelial layer to initiate the cycle again, which could explain a persistent chronic infection [8].

In the present study, the prevalence of *C. cayetanensis* infection was 2.9%; however, there are studies where the prevalence is lower [32]. In contrast, other studies showed higher rates of

infection [33-34]. This coccidian parasite can be found in immunocompetent and immunocompromised patients [35,36] and is responsible for traveller's diarrhea [37].

Contaminated food and water consumption are risk factors for *Cryptosporidium* spp. and *C. cayetanensis* infection [38]. In Mexico, the consumption of mixed vegetable salads from Taylor Farms in Guanajuato has been attributed to infection with *Cryptosporidium* spp., and some cases were specifically related to fresh coriander originating from Puebla. Interestingly, most cases and outbreaks of cyclosporiasis detected in the United States of America have occurred in the Spring and Summer [39].

With respect to the helminths, the prevalence of *A. lumbricoides* infection in this study was 5.8% (6/104), which is less than observations reported in the study of the general population of San Pedro Itzicán, which was 3 times higher (17.5%) than in San Juan Cosala [6]. The low prevalence of helminths found in San Juan Cosala could be, at least in part, the result of the massive deworming campaigns that have been implemented in Mexico [40]. Other studies have reported a decrease in the prevalence of this type of enteric parasite [41]. The soil environment favours the development of geohelminth eggs such as *A. lumbricoides*, *T. trichiura* and hookworms [42]. In this study, the patients who inhabited homes with a cement floor had less parasite infections than those who did not have this type of floor, although this result could be attributed to low numbers (only 4 patients lived in these conditions as opposed to those who had a solid floor). However, when the type of floor in the house was analysed to determine if it influenced the prevalence of parasite infections, a significant difference was found. There were 91.9% of parasitized people who inhabited houses without a tile or cement floor while 35.7% of parasitized people did have a tile or cement floor. This result is possible because the eggs and larvae of geohelminths eliminated in the feces need moist, warm, shaded soil to become infective. Other studies have reported the influence of the type of floor at home and the prevalence of parasitism [41]. In the present study in addition to the house floor, the presence of walls and the material of which they were built were also evaluated. A statistically significant higher prevalence of parasite infection was observed in the people who lived in a house without brick walls or with a roof, and the risk was 6.1 times higher compared with people who lived in a house with brick walls or with a roof. These factors may be influential because there are no physical barriers that prevent polluted air entering the house, and dust is a mode of transmission for parasites such as *Cryptosporidium* spp., and *E. vermicularis*, among others. Additionally, studies that analysed these variables found significant differences [43].

In all the participants that lacked a sewer system in their house, 100% tested positive for parasite infections, although 74.5% participants who lived in houses with a sewer system

were also infected, showing no statistically significant difference, suggesting other associated risk factors were involved in those individuals. However, it has been reported that the lack of a sewer system is an important factor in the development of intestinal parasites [44].

In this study, the Graham method of parasite investigation was applied to children under 12 years old, and *E. vermicularis* was observed in (19/52) 36.5%. In this same group, (4/52) only 7.69% were identified as being infected by using the direct method and concentration method. When these last 2 methods were performed to diagnose adults, only 1 case (1.92%) was detected. These findings reflect the importance of applying the Graham method for the specific diagnosis of *E. vermicularis*. This method has a greater sensitivity than the direct and concentration methods because gravid females migrate nocturnally outside the anus and oviposit while crawling on the skin of the perianal region. The eggs are contained on the adhesive tape [45]. However, the Graham method is not used for adults because of the way the sample is collected [7]. A perianal swab and anal brushing method, has been proposed for the diagnosis of this parasite in children over 12 years and adults [46,47]. Other causes of prevalence of infection can be associated with the spread of eggs in beds, and clothing in the bedroom, in addition to overcrowding suggesting that transmission is intra-domiciliary, affecting children and adults [45].

Of the 104 patients analysed, 58.64% presented polyparasitism. There are reports of multiparasitism in Colima, Jalisco, and Chiapas states of Mexico [6,10], and in other countries such as Malaysia [48]. These results demonstrate the diverse socio-demographic and hygienic factors that provide the optimal conditions required for infection with more than 1 parasite. The majority of the participants who were positive for parasites were asymptomatic, possibly due to coinfection with commensal protozoans that do not induce symptoms. Other possible reasons for the absence of clinical symptoms include low parasitic load, and the immune response of the host. This finding is important, because parasites are being disseminated unknowingly as the infecting parasitic forms are eliminated [44,46].

The strengths of this study were the varied analysis of samples. These included several coproparasitoscopic methods, such as direct smear and iodine solution, concentration by flotation (zinc sulphate), and concentration by sedimentation (formaldehyde/ethyl acetate), the Graham method and the Kinyoun's modified stain method. The use of the flotation method (zinc sulphate) permitted the diagnosis of protozoan cysts and helminth eggs on the surface film, and infertile eggs of *A. lumbricoides* in the sediment were observed.

This study included the general population; other studies consider immunosuppressed populations or children [36]. It has been reported that 3 fecal samples have 94% sensitivity in intestinal parasitic diagnostics [49]. In this study not all pa-

tients provided the 3 samples, which may bias towards an underestimated prevalence of infection for certain parasites such as *G. lamblia* that have an irregular excretion of cysts.

This is the first study reporting the findings of enteroparasitism in San Juan Cosala, Jalisco, Mexico which can support public health, and decision-making strategies to reduce intestinal parasitic infections.

This study observed a high prevalence of *Blastocystis* spp., as well as commensal protozoans. This is important because pathogenic and commensal enteroparasites share the same transmission route, and indicate the deficient hygienic conditions that people are exposed to. A significant association and higher risk of pathogenic and commensal enteroparasite infections were found between the detection of enteroparasites, and housewives, cement floors and brick walls or decks. The clinical symptoms referred to by the participants were not statistically significantly associated with these pathogens.

Sanitary education in the population needs to be implemented. Anti-parasitic albendazole treatment programmes are not specific to protozoans, and since these parasites were found in people who have been treated annually, this treatment programme alone does not address the whole problem.

Conflicts of Interest

The authors declare that they have no competing interests.

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References

- [1] Gebretsadik D, Metaferia Y, Seid A, et al. Prevalence of intestinal parasitic infection among children under 5 years of age at Dessie Referral Hospital: cross sectional study. *BMC Res Notes* 2018;11:771.
- [2] Aquino MJM, Vargas SGB, López MB, et al. Comparación de dos nuevas técnicas de sedimentación y métodos convencionales para la recuperación de parásitos intestinales. *Rev Latinoam Patol Clin* 2012;59(4):233-42. [in Spanish].
- [3] Quihui L, Valencia ME, Crompton DW, et al. Role of the employment status and education of mothers in the prevalence of intestinal parasitic infections in Mexican rural schoolchildren. *BMC Public Health* 2006;6:225.
- [4] Vásquez-Garibay EM, Romero-Velarde E, Napoles-Rodríguez F, et al. [Prevalence of iron and iodine deficiency, and parasitosis among children from Arandas, Jalisco, Mexico]. *Salud Publ Mex* 2002;44(3):195-200. [in Spanish].
- [5] Vásquez-Garibay EM, Campos Barrera LR, Romero Velarde E, et al. Risk factors associated with iron depletion and parasites in preschool and school children of Arandas, Jalisco, Mexico. *Nutr Hosp* 2014;31(1):244-50.
- [6] Galván-Ramírez ML, Madriz-Elisondo AL, Bernal-Redondo R. Biodiversidad parasitaria entre indígenas y mestizos adultos de San Pedro Itzcán, Jalisco, Mexico. *Salud Publica Mex* 2007;49(5):321-2. [in Spanish].
- [7] Clinical and Laboratories Standards Institute, 2005. Procedures for the recovery and identification of parasites from the intestinal tract; approved guideline, 2nd ed. CLSI M28-A2.
- [8] Gamboa MI, Giambelluca LA, Navone GT. Distribución espacial de las parasitosis intestinales en la ciudad de La Plata, Argentina. *Medicina Buenos Aires* 2014;74(5):363-70. [in Spanish].
- [9] Cazorla D, Acosta ME, Acosta ME, et al. [Clinical and epidemiological study of intestinal coccidiosis in a rural population of a semiarid region from Falcon state, Venezuela]. *Invest Clin* 2012;53(3):273-88. [in Spanish].
- [10] Galvan-Ramirez ML, Rivera N, Loeza M, et al. Nitazoxanide in the treatment of *Ascaris lumbricoides* in a rural zone of Colima, Mexico. *J Helminthol* 2007;81(3):255-9.
- [11] Sánchez de la Barquera-Ramos MA, Miramontes-Zapata M. Parasitosis intestinales en 14 comunidades rurales del altiplano de Mexico. *Rev Mex Patol Clin* 2011;58(1):16-25. [in Spanish].
- [12] Guerrero Hernández MT, Hernández Molinar Y, Rada Espinosa ME, et al. Parasitosis intestinal y alternativas de disposición de excreta en municipios de alta marginalidad. *Rev Cub Salud Publica* 2008;34(2):1-4. [in Spanish].
- [13] Gutierrez-Jimenez J, Torres-Sanchez MG, Fajardo-Martinez LP, et al. Malnutrition and the presence of intestinal parasites in children from the poorest municipalities of Mexico. *J Infec Dev Ctries* 2013;7(10):741-7.
- [14] Kiani H, Haghghi A, Rostami A, et al. Prevalence, risk factors and symptoms associated to intestinal parasite infections among patients with gastrointestinal disorders in Nahav and, Western Iran. *Rev Inst Med Trop Sao Paulo* 2016;58:42.
- [15] Devera R, Blanco Y, Amaya I, et al. Parásitos intestinales en habitantes de la comunidad rural "La Canoa", estado Anzoátegui, Venezuela. *Rev Venz Salud Publica* 2014;2(1):15-22. [in Spanish].
- [16] Fletcher S, Caprarelli G, Merif J, et al. Epidemiology and geographical distribution of enteric protozoan infections in Sydney, Australia. *J Public Health Res* 2014;3(2):298.
- [17] Rodríguez E, Mateos B, González JC, et al. Transición parasitaria a *Blastocystis hominis* niños de la zona centro del estado de Guerrero, Mexico. *Parasitol Latinoam* 2008;63(1):20-8. [in Spanish].
- [18] Martínez-Barbabosa I, Gutiérrez-Quiroz M, Ruiz-González L, et al. *Blastocystis hominis* y su relación con el estado nutricional de escolares en una comunidad de la sierra de Huayacocotla, Veracruz, Mexico. *Rev Biomed* 2010;21(2):77-84. [in Spanish].
- [19] Tan KS. New insights on classification, identification, and clinical relevance of *Blastocystis* spp. *Clin Microbiol Rev* 2008;21(4):639-65.
- [20] Dogruman-Al F, Simsek Z, Boorum K, et al. Comparison of methods for detection of *Blastocystis* infection in routinely submitted stool samples, and also in IBS/IBD Patients in Ankara, Turkey. *PloS one* 2010;5:e15484.
- [21] Karadag G, Tamer GS, Dervisoglu E. Investigation of intestinal parasites in dialysis patients. *Saudi Med J* 2013;34(7):714-8.
- [22] Azami M, Sharifi M, Hejazi SH, et al. Intestinal parasitic infections in renal transplant recipients. *Braz J Infect Dis* 2010;14(1):15-8.
- [23] Bednarska M, Jankowska I, Pawelas A, et al. Prevalence of *Cryptosporidium*, *Blastocystis*, and other opportunistic infections in patients with primary and acquired immunodeficiency. *Parasitol Res* 2018;117(9):2869-79.
- [24] Villegas-Gómez I, Martínez-Hernández F, Urrea-Quezada A, et al. Comparison of the genetic variability of *Blastocystis* subtypes between human carriers from two contrasting climatic regions of Mexico. *Infect Genet Evol* 2016;44:334-40.
- [25] Hung CC, Chang SY, Ji DD. *Entamoeba histolytica* infection in men who have sex with men. *Lancet Infect Dis* 2012;12(9):729-36.
- [26] Santos RV, Nunes Jda S, Camargo JA. High occurrence of *Entamoeba histolytica* in the municipalities of Ariquemes and Monte Negro, State of Rondônia, Western Amazonia, Brazil. *Rev Inst Med Tropical Sao Paulo* 2013;55(3):193-6.

- [27] Sard BG, Navarro RT, Sanchis JGE. Amebas intestinales no patógenas: una visión clinicoanalítica. *Enferm Infecc Microbiol Clin* 2011;29:20-8. [in Spanish].
- [28] Calchi La Corte M, Rivero de Rodríguez Z, Bracho Mora A, et al. Prevalencia de *Blastocystis* sp. y otros protozoarios comensales en individuos de Santa Rosa de Agua, Maracaibo, estado Zulia. *Rev Soc Venezolana Microbiol* 2013;33(1):66-71. [in Spanish].
- [29] Pezzani BC, Minvielle MC, Ciarmela ML, et al. [Community participation in the control of intestinal parasitoses at a rural site in Argentina]. *Rev Panam Salud Publica* 2009;26(6):471-7. [in Spanish].
- [30] Pham-Duc P, Nguyen-Viet H, Hattendorf J, et al. *Ascaris lumbricoides* and *Trichuris trichiura* infections associated with wastewater and human excreta use in agriculture in Vietnam. *Parasitol Int* 2013;62(2):172-80.
- [31] Sim S, Yu JR, Lee YH, et al. Prevalence of *Cryptosporidium* Infection among Inhabitants of 2 Rural Areas in White Nile State, Sudan. *Korean J Parasitol* 2015;53(6):745-7.
- [32] Orozco-Mosqueda GE, Martínez-Loya OA, Ortega YR. *Cyclospora cayetanensis* in a pediatric hospital in Morelia, Mexico. *Am J Trop Med Hyg* 2014;91(3):537-40.
- [33] Devera R, Blanco Y, Cabello E. Elevada prevalencia de *Cyclospora cayetanensis* en indígenas del estado Bolívar, Venezuela. *Cad. Saude Publica* 2005;21(6):1778-84. [in Spanish].
- [34] Dhanabal J, Selvadoss PP, Muthuswamy K. Comparative study of the prevalence of intestinal parasites in low socioeconomic areas from South Chennai, India. *J Parasitology Res* 2014;2014:630968.
- [35] Ghoshal U, Dev A, Ranjan P, et al. Identification of opportunistic enteric parasites among immunocompetent patients with diarrhea from Northern India and genetic characterisation of *Cryptosporidium* and *Microsporidia*. *Indian J Med Microbiol* 2016;34(1):60-6.
- [36] Nsagha DS, Njunda AL, Assob NJC, et al. Intestinal parasitic infections in relation to CD4 (+) T cell counts and diarrhea in HIV/AIDS patients with or without antiretroviral therapy in Cameroon. *BMC Infect Dis* 2016;16:9.
- [37] Yu JR, Sohn WM. A case of human cyclosporiasis causing traveler's diarrhea after visiting Indonesia. *J Korean Med Sci* 2003;18(5):738-41.
- [38] Dowd SE, John D, Eliopolus J, et al. Confirmed detection of *Cyclospora cayetanensis*, *Encephalitozoon intestinalis* and *Cryptosporidium parvum* in water used for drinking. *J Water Health* 2003;1(3):117-23.
- [39] Abanyie F, Harvey RR, Harris JR, et al. 2013 multistate outbreaks of *Cyclospora cayetanensis* infections associated with fresh produce: focus on the Texas investigations. *Epidemiol Infect* 2015;143(16):3451-8.
- [40] Tapia-Romero R, Martínez-Méndez LG, Dávila-Solís BL, et al. Transición parasitaria: experiencia en un hospital pediátrico de tercer nivel (1990-2010). *Bol Med Hosp Infant Mex* 2015;72(3):174-80. [in Spanish].
- [41] Chammartin F, Utzinger J, Guimarães LH, et al. Spatio-temporal distribution of soil-transmitted helminth infections in Brazil. *Parasit Vectors* 2014;7:440.
- [42] Tchakounté BN, Nkouayep VR, Poné JW. Soil Contamination Rate, Prevalence, Intensity of Infection of Geohelminths and Associated Risk Factors among Residents in Bazou (West Cameroon). *Ethiop J Health Sci* 2018;28(1):63-72.
- [43] Nyantekyi L, Legesse M, Medhin G, et al. Community awareness of intestinal parasites and the prevalence of infection among community members of rural Abaye Deneba area, Ethiopia. *Asian Pac J Trop Biomed* 2014;4(suppl 1):S152-7.
- [44] Farghly AM, Mohamed SM, Abdel-Rahman SA, Mohammed FE, El-Bahaie ES, El-Shafey MA. The relation between the prevalence of soil transmitted parasites in the soil and among school children in Zagazig district, Sharkyia Governorate, Egypt. *J Parasit Dis* 2016;40(3):1021-9.
- [45] Afrakhteh N, Marhaba Z, Mahdavi SA, et al. Prevalence of *Enterobius vermicularis* amongst kindergartens and preschool children in Mazandaran Province, North of Iran. *J Parasit Dis* 2016;40(4):1332-6.
- [46] Pezzani BC, Minvielle MC, de Luca MM, Cordoba MA, Apezteguia MC, Basualdo JA. *Enterobius vermicularis* infection among population of General Mansilla, Argentina. *World J Gastroenterol* 2004;10(17):2535-9.
- [47] Jeric-Lara MI, Oyarce-Fierro A [Internet]. Recomendaciones para la búsqueda de huevos de *Enterobius vermicularis*. Instituto de Salud Pública de Chile. Available from: http://www.ispch.cl/sites/default/files/Recomendacion_Busqueda_Enterobius_vermicularis.pdf.
- [48] Al-Delaimy AK, Al-Mekhlafi HM, Nasr NA, et al. Epidemiology of Intestinal Polyparasitism among Orang Asli School Children in Rural Malaysia. *PLoS Negl Trop Dis* 2014;8(8):e3074.
- [49] Hanson KL, Cartwright CP. Use of an enzyme immunoassay does not eliminate the need to analyze multiple stool specimens for sensitive detection of *Giardia lamblia*. *J Clin Microbiol* 2001;39(2):474-7.