## **INVITED ARTICLE**

# Approach to the Coagulopathic Patient in the Intensive Care Unit

Manoj Yogendra Singh

#### Key learning objectives:

- · Evaluating patients with coagulopathy in intensive care
- · Managing coagulopathy in intensive care
- · Understand the complications and limitations of various therapies

Keywords: Bleeding and hemorrhage, Coagulation reversal, Coagulation tests, Coagulopathy, Factor concentrate, Fresh frozen plasma, Haemostasis, Recombinant coagulation products

Indian Journal of Critical Care Medicine (2019): 10.5005/jp-journals-10071-23256

#### INTRODUCTION

A critical care physician often has to manage patients who either present with or develop coagulation abnormalities in intensive care unit, and they are a predictor of both, the need for massive transfusion and mortality.<sup>1</sup> These abnormalities can range from something as simple as isolated thrombocytopenia to more complex multisystem coagulation defects. Table 1 shows the common causes of coagulopathy in critically ill patients. Moreover, in critically ill patients, assessing bleeding risk is one of the key management strategies to minimize any procedural or perioperative bleeding. Critically ill patients can be prone to bleeding for a wide variety of reasons including hereditary or acquired bleeding disorders (platelet function abnormalities, factor deficiencies and factor inhibitors), underlying medical conditions such as hepatic or renal disease and concomitant anticoagulation medications. Besides, certain connective tissue disorders can impact on the integrity of blood vessels, which make them more prone to bruising/bleeding.

Department of Intensive Care Medicine, The Canberra Hospital, Garran, ACT, Australia

**Corresponding Author:** Manoj Yogendra Singh, Department of Intensive Care Medicine, The Canberra Hospital, Garran, ACT, Australia, e-mail: Manoj.Singh@act.gov.au

How to cite this article: Singh MY. Approach to the Coagulopathic Patient in the Intensive Care Unit. Indian J Crit Care Med 2019;23(Suppl 3):S215–S220.

Source of support: Nil

Conflict of interest: None

# **UNDERSTANDING NORMAL HEMOSTASIS**

Hemostasis is a complex process and involves multiple steps (Fig. 1). It is subdivided into four phases. The first phase of primary hemostasis involves vasoconstriction and platelet plug formation and is triggered when the subendothelial collagen is

Table 1: Common causes of coagulopathy in critically ill

Secondary disruption of	Derang		
hemostasis	Deranged coagulation	Thrombocytopenia	Hyperfibrinolysis <sup>2</sup>
• Hypothermia (Temp <34°C)	Сог	nsumption	Acquired secondary
<ul> <li>Severe acidosis (pH &lt;7.25)</li> <li>Hypocalcemia (iCa<sup>++</sup> &lt;1 mmol/L)</li> </ul>	<ul> <li>Sepsis</li> <li>DIC</li> <li>Cardiac surgery</li> </ul>	<ul> <li>Sepsis</li> <li>DIC</li> <li>Extracorporeal circuits (CRRT),</li> <li>Enlarged spleen</li> </ul>	<ul> <li>Trauma</li> <li>Thrombolytic therapy</li> <li>Cardiopulmonary bypass</li> <li>Systemic amyloidosis</li> <li>Placental disorders</li> </ul>
	B	Acquired Primary	
	<ul> <li>Multiple trauma and major blood loss</li> </ul>	Multiple trauma and major blood loss	<ul> <li>End-stage liver cirrhosis</li> <li>Acute promyelocytic leukaemia</li> </ul>
	Decreased	Inherited secondary	
	<ul> <li>Vitamin K deficiency</li> <li>Vitamin K antagonists</li> <li>Liver disease and renal failure</li> <li>Hemophilia</li> <li>FXIII deficiency</li> <li>Dysfibrinogenemias</li> <li>Drugs: Heparin, novel oral anticoagulants, direct thrombin inhibitore direct Ya inhibitore</li> </ul>	<ul> <li>Bone marrow suppression</li> <li>Vitamin B<sub>12</sub> and folate deficiency</li> <li>Myelosuppression</li> <li>Drugs: Acetaminophen, carbamazepine, hydrochlorothiazide, cimetidine, ranitidine, quinidine, quinine, bactrim, etc</li> </ul>	<ul> <li>Hemophilia</li> <li>FXIII deficiency</li> <li>Dysfibrinogenemias</li> </ul>

© The Author(s). 2019 Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (https://creativecommons. org/licenses/by-nc/4.0/), which permits unrestricted use, distribution, and non-commercial reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.



Fig. 1: Simplified diagram of Hemostasis

\_ . .

	Screening test	Confirmatory test
Coagulation	aPTT, PT, Thrombin time (TT)	FII, V, VII, VII, IX, X, XI, XII, XII activity Fibrinogen activity
vWF	Platelet function analyzer, vWF antigen	Antigen and platelet activity, genetic testing, propeptide testing
Platelet function	<ul><li> Platelet count</li><li> Bleeding time (BT)</li><li> Platelet function analyzer</li></ul>	Light transmission aggregometry with arachidonic acid, thrombin receptor-activating peptide, collagen, adenosine diphosphate
Fibrinolysis	Euglobulin lysis time	tPA, plasminogen activity inhibitor, alpha-2 antiplasmin
Anticoagulation	aPTT, PT, TT, BT, anti-FXa activity, thrombin inhibition time (TTI), reptilase time	

Table 2: Screening and	l confirmatory	test for hemostasis
------------------------	----------------	---------------------

exposed after vessel wall injury. This leads to platelet adhesion to the subendothelial layer via von Willebrand and glycoprotein Ib and subsequent platelet aggregation. Next comes secondary hemostasis, which involves the activation of coagulation factors and thrombin formation. Phase 3 requires fibrin clot formation and stabilization, and the final step requires inhibition of thrombin generation and fibrinolysis. Traditionally, this process has thought to occur via three pathways. The intrinsic pathway (uses factors VIII, IX, XI, XII), extrinsic pathway (uses factor VII), and common pathway where both converge to activate factor X (uses factors II, V, fibrinogen). In the new proposed coagulation cascade, this complete process does not occur continuously but instead requires three consecutive phases: namely, an initial phase, an amplification phase and the propagation phase.

The liver is responsible for the production of most of the factors, namely; I, II, V, VII, VII, IX, X, XI, XIII and protein C (FXIV) and protein S. Thus, patients with advanced liver disease often have coagulopathy. The coagulation system also has a negative feedback mechanism to prevent overcoagulation and thrombosis. Thrombin also acts by activating plasminogen (to plasmin, which is an active enzyme in fibrinolysis) and stimulating the production of antithrombin (which decreases the production of thrombin and decreases the output of FXa).

#### **EVALUATING COAGULATION**

The most obvious indications that a patient has coagulopathy are an unusual drop in hemoglobin or persistent bleeding. This often manifests as the presence of ecchymosis, petechiae, haematuria, hematomas or prolonged bleeding from puncture sites. Excessive bleeding from surgical drains or incision sites may also occur. If the bleeding is significant and allowed to continue, it may lead to hypovolemic shock, hypoperfusion and organ failure. Apart from above physical signs of bleeding, one should also look for jaundice, splenomegaly, arthropathy, joint and skin laxity (Marfans or Ehlers-Danlos syndrome) as signs of systemic or connective tissue disorder. The other causes of easy bruising like alcohol abuse, purpura simplex, Cushing's disease, vitamin C deficiency should also be screened for. One should also take a detailed medication history to check if the patient is taking medications such as antiplatelet agents, anticoagulants (warfarin or NOVACs) and complementary medications that affect coagulation. Drugs like cephalosporins, ginkgo-biloba, interferon, SSRI, TCA are rare causes that can cause bleeding and bruising.<sup>3</sup>

The screening tests for hemostasis are summarized in Table 2. Despite their limitations, both prothrombin time (PT) and activated partial thromboplastin time (aPTT) remain the most common screening test to evaluate coagulation. PT measures the integrity of extrinsic and common pathway while aPTT measures the integrity of the intrinsic and common pathway. They assess the time it takes for both the pathways to generate cellular plasma and thus only investigate a narrow part of the coagulation system. Figure 2 highlights the factors involved in each pathway. These tests are designed for clinical monitoring of anticoagulation and not coagulopathy and thus only serve as useful starting points of investigation of coagulation. Mixing studies are done to determine whether a prolonged PT or aPTT or both are affected by the presence of a factor deficiency or a factor inhibitor. If the test normalises when plasma is added, it is due to factor deficiency, and if not, it is secondary to the presence of an inhibitor like lupus anticoagulant.



Fig. 2: Factors involved in the extrinsic (yellow), intrinsic (blue) and common pathways (red)

PT	aPTT	Causes				
Normal Normal		von Willebrand's disease FXIII deficiency, Dysfibrinogenemia	Platelet dysfunction $\alpha$ -antiplasmin deficiency			
		Mixing study corrects	Mixing study does not correct			
Prolonged	Normal	<ul> <li>Hereditary</li> <li>Isolated FVII deficiency</li> <li>Acquired</li> <li>Vitamin K deficiency/ antagonists</li> <li>Severe liver impairment</li> </ul>	<ul><li>Inhibitors</li><li>Lupus anticoagulant</li><li>FVII inhibitor (rare)</li></ul>			
Normal	Prolonged	<ul> <li>Bleeding</li> <li>FVIII/vWD deficiency, FIX/XI deficiency</li> <li>No bleeding</li> <li>FXII, HMWK, prekallikrein</li> </ul>	<ul> <li>Bleeding</li> <li>FVIII, FIX, FXI inhibitor, heparin No bleeding</li> <li>FXII inhibitor, lupus anticoagulant HMWK, prekallikrein</li> </ul>			
Prolonged	Prolonged	<ul> <li>FII, FV, FX deficiency</li> <li>FV and FVIII deficiency</li> <li>Severe liver impairment</li> <li>Vitamin K deficiency/antagonist</li> <li>DIC</li> </ul>	<ul><li>FII, FV, FX inhibitor</li><li>Lupus anticoagulant</li></ul>			

Table 3: Initial	assessment of	prolonged PT	and aPTT in a	natient with	bleeding
Table 5. Initial	assessment of	prolongeurr	anuarrinia	patient with	Dieeunig

Table 4: Assessment of coagulation for patients on anticoagulants

	5		-			
Test	Anticoagulant					
	LMWH/ Fondaparinux	Thrombin inhibitors <ul> <li>Argatroban</li> <li>Bivalirudin</li> <li>Dabigatran</li> </ul>	Vitamin K antagonist • Warfarin	FXa inhibitors • Rivaroxaban • Apixaban	range	
PT	Normal	Prolonged	Prolonged	Prolonged	<35 sec	
aPTT	Normal	Prolonged	Normal	Prolonged	<13.5 sec	
TT	Normal	Prolonged	Normal	Normal	<21 sec	
BT	Normal	Normal	Normal	Normal	<21 sec	
Anti-FXa	Detectable	Not detectable	Not detectable	Detectable	<0.1	
TTI	No inhibition	Inhibition	No inhibition	No inhibition		

TT, thrombin time; BT, bleeding time; TTI, tissue thromboplastin inhibition

These tests should accompany a detailed history, including medication and clinical examination. It is also essential to get the peripheral smear examination as it also helps in looking at the morphology of platelets and also identify systemic illness and other hematological disorders. Table 3 highlights the causes of abnormality in these tests and Table 4 highlights the defect associated with the use of anticoagulants.

#### **Evaluation of Preexisting Coagulation Disorder**

The British Committee for Standards in hematology and various Anesthesiology Society guidelines for perioperative assessment recommend evaluation of bleeding risk before surgery or invasive procedures for all patients.<sup>4–6</sup> Bleeding assessment tools (BATs) were developed to offer a simple, structured screening tool to improve the diagnostic accuracy of bleeding disorder with symptom severity, minimise investigations, predict the risk of bleeding and inform about the treatment strategies (Table 5).

It is easy to diagnose major bleeding disorders like hemophilia and vWD but challenging to diagnose or classify some of the mild bleeding disorders. In a prospective Indian study, Kotru et al. reported that of the 164 patients who presented with slight bleeding, epistaxis was the most common presentation with cutaneous bleeding, the next common site. A family history of bleeding was present in only 11 patients. Only 25% of the patients were confirmed to have a bleeding disorder based on the investigation, and the rest labeled as unclassified bleeding disorder.<sup>7</sup>

Tool	Year developed	Scores allotted	Target condition	Time taken	Comments <sup>8,9</sup>
Vincenza bleeding score	2005	0 to +3	Type 1 vWD	40 min	Sensitivity (64%) Specificity (99%)
European molecular and clinical markers for diagnosis and management of type 1 vWD (MCMDM- 1 vWD)	2006	–1 to +4	Type 1 vWD	40 min	Sensitivity (59%) Specificity (96%)
Condensed MCMDM vWD-1	2008	–1 to +4	Type 2B vWD	5–10 min	Sensitivity (76%) Specificity (100%)
			Bleeding disorders		Sensitivity (85%) Specificity (90%)
ISTH-BAT	2010	0 to +4	Type 1 vWD	20 min	Sensitivity (49%) Specificity (47%)
HEMSTOP	2015	0 to 1	Bleeding disorder	5–10 min	Sensitivity (89.5%) Specificity (98.6%)

Table 6: Advantages and disadvantages of blood and pharmacological products

Blood P	roducts	Factor concentrates		
Advantages	Disadvantages	Advantages	Disadvantages	
<ul> <li>FFP contains all factors</li> </ul>	<ul> <li>Low concentration</li> </ul>	<ul> <li>High concentration of</li> </ul>	<ul> <li>Not all factors available</li> </ul>	
including vWF and factor XIII	<ul> <li>Transfusion related risks</li> </ul>	factors	<ul> <li>High costs</li> </ul>	
<ul> <li>Well studied and included</li> </ul>	<ul> <li>Requires cross matching</li> </ul>	<ul> <li>Few transfusion related</li> </ul>	<ul> <li>Point of care testing</li> </ul>	
in standardized massive	<ul> <li>Large fluid load</li> </ul>	complications	required	
transfusion protocols (MTP)	<ul> <li>Shorter storage life</li> </ul>	<ul> <li>No cross match required</li> </ul>	<ul> <li>Need parallel fluid resusci-</li> </ul>	
<ul> <li>Effective volume therapy in</li> </ul>	Thawing takes time and not	<ul> <li>Small fluid volumes and</li> </ul>	tation for volume loss	
shock patients	immediately available	hence no hemodilution		
<ul> <li>Relatively cheap</li> </ul>				

#### **Reversal of Coagulopathy**

When managing a patient with coagulopathy, it is essential to consult a hematologist. It is also crucial to monitor the response to the treatment given by checking targeted tests. One can correct the factor deficiencies using fresh frozen plasma (FFP) when the coagulopathy is undifferentiated or secondary to causes like severe sepsis and DIC. Factor concentrates are used when one knows the coagulopathy is secondary to specific factor deficiency and or anticoagulant (e.g. warfarin). Table 6 highlights the advantages and disadvantages of each.<sup>10</sup>

Plasma transfusion may be useful in a patient who requires volume resuscitation along with multiple factors to correct the coagulopathy (e.g. patients with trauma and or one with massive exsanguination). It is also important to maintain temperature >35°C, ionized calcium levels >1.1 mmol/L and pH >7.25 in these patients. One also needs to activate a systematic transfusion therapy with local massive transfusion protocol to improve the delivery of blood products (packed blood cells, fresh frozen plasma, platelets in fixed ratios along with cryoprecipitate if the fibrinogen is <2.0 g/L). One gram of tranexamic acid followed by an infusion of 1 gram over 8 hours is also used in such scenarios. FFP is effective in correcting high PT as it has a dilution factor secondary to the volume. However, it is uncertain that prophylactically correcting INR decreases the incidence of bleeding. The TOPIC trial, which was an RCT, failed to show that transfusion of FFP (12 mL/kg) to correct the INR (>1.5-3.0) prevented bleeding complications in patients undergoing central venous catheter placement, percutaneous tracheostomy, chest tube or abscess drainage.<sup>11</sup> The amount of FFP needed to increase the desired level of factor concentration also varies. Chowdary et al. found that one may need volume as high as 30 mL/kg to achieve the target.12

 Table 7: Reference for blood component and factor concentrate administration based on factor deficiency

		Factor
Factor deficient	Blood Component	concentrates
Fl (fibrinogen)	Cryoprecipitate (cryo), fresh frozen plasma (FFP)	Fl concentrate
Factor V (labile factor)	FFP	
Factor VII (stable factor/ proaccelerin)	FFP	Factor VII concentrate
Factor VIII (antihemophilic factor)	Cryo, FFP	Factor VIII concentrate
von Willebrand's disease	Cryo, FFP	Factor VIII concentrate
Factor IX (Christmas factor)	FFP	Factor FIX concentrate
Factor X (Stuart–Prower factor)	FFP	Factor FX concentrate
Factor XI (plasma thromboplastin antecedent)	FFP	Factor IX complex (II, VII, IX, X)
Factor XIII (fibrin- stabilizing factor)	FFP	Factor FXIII concentrate

The common risks associated with FFP transfusion are transfusion-related acute lung injury (TRALI), transfusion-related circulatory overload, allergic or anaphylactic reactions. The less frequent complications include risk of transmission of infections, febrile nonhemolytic reactions, hemolytic reactions and red blood cell (RBC) alloimmunization. Factor concentrate therapy (Table 7) guided by viscoelastic testing (TEG or ROTEM) has been tried to

S218

minimize the use of blood products in patients undergoing major surgery and trauma but the benefits of this approach has not been demonstrated in big multicentre randomized control trials (RCT).<sup>13,14</sup>

Prothrombin complex concentrate (PCC) has variable concentrates of 4 factors, namely II, VII, IX, and X and are mainly approved for reversal of coagulopathy secondary to vitamin K antagonist. They are also used in the prevention and treatment of bleeding in patients with hemophilia B. Certain PCC also contain small levels of activated FVII. When compared to FFP, PCCs reverse INR faster and are easier to administer as do not need crossmatching. In scenarios when the INR is not corrected post PCC infusion, one should treat the patient with FFP.

Recombinant coagulation products are proteins which are now more readily available for managing bleeding patients on anticoagulants or with specific coagulation deficiencies. These proteins can be modified and can be used in patients with acquired antibodies and inhibitors to various factors. Recombinant activated factor VIIa is approved for patients with hemophilia with inhibitors but increasingly used in patients with life-threatening massive hemorrhage where conventional blood component therapy is unsuccessful. It only works once hypothermia, hypocalcemia, acidosis are corrected, and PT/aPTT optimized. Its significant side-effects are thromboembolic complications. Inactivated-zhzo recombinant FXa (Andexxa) has recently been approved for reversal

of the anticoagulation effect of direct FXa inhibitors (apixaban and rivaroxaban). The limitation of this product is that the anticoagulant effect lasts while the infusion is ongoing (2 hours) and its high costs.

Factor XIII plays a vital role in the final step of clot formation and stabilization. Many studies have shown a reduction in factor FXIII in patients put on cardiopulmonary bypass and FXIII replacement, along with antifibrinolytic therapy reduces the risk of postoperative bleeding and in patients with major trauma.<sup>15,16</sup> However, the presence alpha-2 antiplasmin protein (which inactivates plasmin) and other clotting factors (VIII, XIII, vWF) in addition to fibrinogen in the cryoprecipitate makes them more effective in terms of their duration and mode of action when compared to factor FXIII concentrates.

In the event when the coagulopathy is secondary to anticoagulants, it is ideal first to use specific anticoagulation reversing agents and it is also necessary to consult a haematologist. Table 8 highlights the specific reversal agents of various anticoagulants.<sup>17</sup>

#### SUMMARY

Coagulopathy is common in intensive care and can be multifactorial. It is crucial to find the underlying cause and understand the limitations of various tests to assess them. Early hematology referral is vital. FFPs remain the broad-spectrum therapy to correct coagulopathy.

Table 8: Anticoagulant and the reversal agents

		Half-life		Reversal agent(s)	Reversal agent
Anticoagulant	Evaluation test	(hours)	Reversible	Consult hematologist	Half-life
		Ar	ntithrombin III ac	tivator (FII and FX inhibitor)	
IV Heparin	aPTT	1.5	Yes	Protamine (1mg/100 units heparin)	7–8 minutes
			Vitan	nin K inhibitor	
Warfarin	PT/INR	40	Yes	Vitamin K 10 mg	2 hours
				Prothrombin complex concentrate	4–12 hours
				20–50 units/kg	
				rFVIIa 90 μg/kg	2.3 hours
				FFP	4–12 hours
			Fact	or X inhibitor	
LMWH Enoxaparin	Anti-FXa	7	Partial	Protamine	7–8 minutes
				If received <2 hours (60–80%)	
				1mg protamine/1mg enoxaparin	
				If received >8 hours before:	
				0.5 mg protamine/1 mg enoxaparin	
				If received >12 hours before: Nil	
				rFVIIa (70–90 mg/kg) in severe bleeds	2.3 hours
LMWH Dalteparin	Anti-FXa	3-5	Partial	Protamine	7–8 minutes
				If received <2 hours: (60%)	
				1 mg protamine/1 mg dalteparin	
				rFVIIa (70–90 mg/kg) in severe bleeds	2.3 hours
			Direct	FXa inhibitors	
Apixaban	Anti-FXa	8–15	No	Andexxa (rXa, inactivated-zhzo)	1 hour
				<8 hours:400 mg–800 mg at 30 mg/minute;	
				>8 hours 400 mg	
				Prothrombin complex concentrate	4–60 hours
				25–50 units/kg	
Rivaroxaban	Anti-FXa	7–11	No	Andexxa (rXa, inactivated-zhzo)	1 hour
				<8 hours:400 mg–800 mg at 30 mg/minute; >8	
				hours 400 mg	
				Prothrombin complex concentrate	4–60 hours
				25–50 units/kg	
				rFVIIa 90 mg/kg	2.3 hours
					Contd

Approach to the Coagulopathic Patient in the Intensive Care Unit

			Direct F	Ila inhibitors	
Dabigatran	Limited value except TT, TEG, anti-Flla	12–17	No	ldarucizumab (if TT is prolonged) 5 mg bolus or infusion over 5–10 minutes Prothrombin complex concentrate	Biphasic: 45 min- utes, 4–8 hours
				50 units/kg	4–12 hours
Argatroban	Limited value except TEG anti-Flla	0.75	No	rFVIIa 90 μg/kg	2.3 hours
Tissue plasminogen activator					
Alteplase	D-dimer	0.5-0.75	Yes	Tranexamic acid 10 mg/kg	2 hours

## References

- 1. Walsh TS, Stanworth SJ, Prescott RJ, Lee RJ, Watson DM, Wyncoll D, et al. Prevalence, management, and outcomes of critically ill patients with prothrombin time prolongation in United Kingdom intensive care units. Crit Care Med. 2010;38(10):1939–1946.
- Kolev K, Longstaff C. Bleeding related to disturbed fibrinolysis. Br J Haematol. 2016;175(1):12–23.
- Tosetto A, Rodeghiero F, Castaman G, Goodeve A, Federici AB, Batlle J, et al. A quantitative analysis of bleeding symptoms in type 1 von Willebrand disease: results from a multicenter European study (MCMDM-1 VWD). J Thromb Haemost. 2006;4(4):766–773.
- Bonhomme F, Ajzenberg N, Schved JF, Molliex S, Samama CM, French A, et al. Pre-interventional haemostatic assessment: Guidelines from the French Society of Anaesthesia and Intensive Care. Eur J Anaesthesiol. 2013;30(4):142–162.
- Committee on S, Practice P, Apfelbaum JL, Connis RT, Nickinovich DG, American Society of Anesthesiologists Task Force on Preanesthesia E, et al. Practice advisory for preanesthesia evaluation: an updated report by the American Society of Anesthesiologists Task Force on Preanesthesia Evaluation. Anesthesiology. 2012;116(3):522–538.
- Chee YL, Crawford JC, Watson HG, Greaves M. Guidelines on the assessment of bleeding risk prior to surgery or invasive procedures. British Committee for Standards in Haematology. Br J Haematol. 2008;140(5):496–504.
- Kotru M, Mutereja D, Purohit A, Tyagi S, Mahapatra M, Saxena R, et al. Mild Bleeders: Diagnosis is Elusive in Large Number of Patients. Mediterr J Hematol Infect Dis. 2016;8(1):e2016049.
- 8. Moenen F, Nelemans PJ, Schols SEM, Schouten HC, Henskens YMC, Beckers EAM. The diagnostic accuracy of bleeding assessment tools

for the identification of patients with mild bleeding disorders: A systematic review. Haemophilia. 2018;24(4):525–535.

- Bonhomme F, Boehlen F, Clergue F, de Moerloose P. Preoperative hemostatic assessment: a new and simple bleeding questionnaire. Can J Anaesth. 2016;63(9):1007–1015.
- Faraoni D, Hardy JF, Van der Linden P. An early, multimodal, goaldirected approach of coagulopathy in the bleeding traumatized patient. Curr Opin Anaesthesiol. 2013;26(2):193–195.
- Muller MC, de Jonge E, Arbous MS, Spoelstra-de Man AM, Karakus A, Vroom MB, et al. Transfusion of fresh frozen plasma in non-bleeding ICU patients--TOPIC trial: study protocol for a randomized controlled trial. Trials. 2011;12:266.
- Chowdary P, Saayman AG, Paulus U, Findlay GP, Collins PW. Efficacy of standard dose and 30 ml/kg fresh frozen plasma in correcting laboratory parameters of haemostasis in critically ill patients. Br J Haematol. 2004;125(1):69–73.
- 13. Shen L, Tabaie S, Ivascu N. Viscoelastic testing inside and beyond the operating room. J Thorac Dis. 2017;9(Suppl 4):S299-S308.
- Mohamed M, Majeske K, Sachwani GR, Kennedy K, Salib M, McCann M. The impact of early thromboelastography directed therapy in trauma resuscitation. Scand J Trauma Resusc Emerg Med. 2017;25(1):99.
- 15. Sniecinski RM, Levy JH. Bleeding and management of coagulopathy. J Thorac Cardiovasc Surg. 2011;142(3):662–667.
- Curry N, Foley C, Wong H, Mora A, Curnow E, Zarankaite A, et al. Early fibrinogen concentrate therapy for major haemorrhage in trauma (E-FIT 1): results from a UK multi-centre, randomised, double blind, placebo-controlled pilot trial. Crit Care. 2018;22(1):164.
- 17. Medow JE, Dierks MR, Williams E, Zacko JC. The emergent reversal of coagulopathies encountered in neurosurgery and neurology: a technical note. Clin Med Res. 2015;13(1):20–31.

