



Soluble CD40L activates soluble and cell-surface integrin $\alpha\beta3$, $\alpha5\beta1$, and $\alpha4\beta1$ by binding to the allosteric ligand-binding site (site 2)

Received for publication, November 14, 2020, and in revised form, January 27, 2021. Published, Papers in Press, February 9, 2021,

<https://doi.org/10.1016/j.jbc.2021.100399>

Yoko K. Takada^{1,2}, Michiko Shimoda¹, Emanuel Maverakis¹, Brunie H. Felding³ , R. Holland Cheng⁴, and Yoshikazu Takada^{1,2,*}

From the Departments of ¹Dermatology, and ²Biochemistry and Molecular Medicine, UC Davis School of Medicine, Sacramento, California, USA; ³The Scripps Research Institute, Department of Molecular and Experimental Medicine, La Jolla, California, USA; and ⁴Department of Molecular and Cellular Biology, UC Davis, Davis, California, USA

Edited by Peter Cresswell

CD40L is a member of the TNF superfamily that participates in immune cell activation. It binds to and signals through several integrins, including $\alpha\beta3$ and $\alpha5\beta1$, which bind to the trimeric interface of CD40L. We previously showed that several integrin ligands can bind to the allosteric site (site 2), which is distinct from the classical ligand-binding site (site 1), raising the question of if CD40L activates integrins. In our explorations of this question, we determined that integrin $\alpha4\beta1$, which is prevalently expressed on the same CD4⁺ T cells as CD40L, is another receptor for CD40L. Soluble (s)CD40L activated soluble integrins $\alpha\beta3$, $\alpha5\beta1$, and $\alpha4\beta1$ in cell-free conditions, indicating that this activation does not require inside-out signaling. Moreover, sCD40L activated cell-surface integrins in CHO cells that do not express CD40. To learn more about the mechanism of binding, we determined that sCD40L bound to a cyclic peptide from site 2. Docking simulations predicted that the residues of CD40L that bind to site 2 are located outside of the CD40L trimer interface, at a site where four HIGM1 (hyper-IgM syndrome type 1) mutations are clustered. We tested the effect of these mutations, finding that the K143T and G144E mutants were the most defective in integrin activation, providing support that this region interacts with site 2. We propose that allosteric integrin activation by CD40L also plays a role in CD40L signaling, and defective site 2 binding may be related to the impaired CD40L signaling functions of these HIGM1 mutants.

CD40L is a type II protein ligand member of the tumor necrosis factor (TNF) superfamily expressed by activated T cells. CD40L is a costimulatory molecule critical in a variety of T cell–antigen presenting cell (APC) interactions, including activating APCs to provide help to cytotoxic T cells (1). It also provides help to B cell to promote class switching (2, 3). In addition to its transmembrane form, CD40L is also released as a soluble ligand (sCD40L) by proteolytic cleavage, allowing it to interact at more distant sites. CD40L appears to modulate other cell types as well (4). CD40L can initiate inflammatory and procoagulatory responses in vascular endothelial cells (5–7). Findings such as these have led to

the belief that CD40–CD40L interactions play a more general role in immune regulation. Given its diverse functions, it is not surprising that CD40L is critical in a variety of chronic autoimmune and inflammatory diseases, including systemic lupus erythematosus (SLE), diabetes, chronic kidney disease (8, 9), among others.

CD40L functions through its interactions with cell surface proteins CD40, and $\alpha5\beta1$ and $\alpha\text{IIb}\beta3$ integrins. CD40 belongs to the tumor necrosis factor receptor (TNF-R) family and was first identified and functionally characterized on B lymphocytes (10). CD40–CD40L interactions play a more general role in immune regulation and stabilize arterial thrombi through binding to integrin $\alpha\text{IIb}\beta3$ (11). $\alpha\text{IIb}\beta3$ recognizes the KGD motif at the N terminus of CD40L (residues 115–117 of CD40L). It has also been reported that CD40L binds to integrin $\alpha5\beta1$ and transduces signals through this integrin in a CD40 and $\alpha\text{IIb}\beta3$ -independent manner. Finally, data suggests that CD40 and integrin $\alpha5\beta1$ can bind to CD40L simultaneously (12).

We recently identified vascular integrin $\alpha\beta3$ as a new receptor for CD40L (13). We localized the $\alpha5\beta1$ and $\alpha\beta3$ binding to the trimeric interface of CD40L. CD40L mutants defective in integrin binding in the predicted integrin-binding site were defective in CD40L/CD40 signaling and acted as antagonists of CD40L/CD40 signaling (13). Furthermore, we demonstrated that CD40L binding to $\alpha\beta3$ activates $\alpha\beta3$ in an allosteric manner. Of relevance to our discovery is the finding that eight X-Linked Hyper IgM Syndrome (HIGM1)-causative variants have alterations in the CD40L integrin binding, and they are defective in integrin binding and signaling, suggesting that the loss of integrin binding is related to the defect in CD40L signaling in HIGM1. Also, our previous studies found that several proinflammatory integrin ligands (e.g., CX3CL1, CXCL12, and secreted phospholipase A2 type IIA (sPLA-IIA)) activated integrins by binding to a second ligand-binding site (site 2) in an allosteric manner in addition to binding to a primary site (site 1) (14–16). The recent discovery on the binding of 25-hydroxycholesterol to integrin site 2 upregulates inflammatory cytokines, TNF, and IL-6, production (17) indicates that site-2-mediated integrin activation is involved in proinflammatory signaling.

* For correspondence: Yoshikazu Takada, ytakada@ucdavis.edu.

Allosteric integrin activation by CD40L

Herein, we now present data that integrin $\alpha 4\beta 1$ is a new receptor for CD40L. Notably, we showed that CD40L is an allosteric activator of integrins $\alpha \beta 3$, $\alpha 5\beta 1$, and $\alpha 4\beta 1$. Specifically, we demonstrate that CD40L can bind to an allosteric ligand-binding site (Site 2) (14–16), which is distinct from $\alpha \beta 3$'s classical ligand-binding site (site 1). Also, four HIGM1 mutants are clustered in the site-2-binding site of CD40L, indicating that site-2-binding is potentially involved in CD40L/CD40 signaling. We propose that CD40L acts as a ligand and an allosteric activator of several integrins and mediates proinflammatory signaling independent of CD40. We further propose that this may be a common mechanism for a group of proinflammatory proteins and that CD40L-induced integrin activation is required for CD40L signaling.

Results

sCD40L activates soluble integrin $\alpha \beta 3$ in cell-free conditions

Previous studies showed that several proinflammatory integrin ligands activated integrins in an allosteric manner (see Introduction). We hypothesized that another proinflammatory cytokine CD40L activates integrins. Previous studies showed that soluble integrin $\alpha \beta 3$ bound to many ligands including the fibrinogen γ -chain C-terminal domain with truncation at the C terminus (γ C399tr) (See Introduction). γ C399tr specifically binds to site 1 of $\alpha \beta 3$, but not to site 2 (14). We studied if sCD40L enhances the binding of soluble $\alpha \beta 3$ to γ C399tr. γ C399tr was immobilized and incubated with soluble $\alpha \beta 3$ in the presence of 1 mM Ca^{2+} to keep $\alpha \beta 3$ inactive (Fig. 1A). sCD40L enhanced binding of soluble $\alpha \beta 3$ to immobilized γ C399tr in a dose-dependent manner. Activation is defined by the increase in binding of soluble integrin $\alpha \beta 3$ to immobilized ligand (γ C399tr) by soluble activators (sCD40L). These findings suggest that CD40L activates integrin $\alpha \beta 3$ in an allosteric manner.

sCD40L activates cell-surface integrin $\alpha \beta 3$

We next studied if sCD40L activates integrin $\alpha \beta 3$ on the cell surface using CHO cells (CD40-negative) that express recombinant $\alpha \beta 3$ ($\beta 3$ -CHO cells). $\beta 3$ -CHO cells were incubated with sCD40L and FITC-labeled γ C399tr in the assays medium with 1 mM Ca^{2+} to keep $\alpha \beta 3$ inactive and bound FITC-labeled γ C399tr was measured in flow cytometry. sCD40L markedly enhanced the binding of γ C399tr, indicating that sCD40L activated $\alpha \beta 3$ on the cell surface (Fig. 1B). The activation of $\alpha \beta 3$ by sCD40L was dose-dependent (up to 50 $\mu\text{g}/\text{ml}$) and can be detected at 6 $\mu\text{g}/\text{ml}$. The binding of FITC-labeled γ C399tr

shown by mean fluorescent intensity (MFI) correlated well with the sCD40L concentrations (Fig. 1D). Although high concentrations of sCD40L were required to activate soluble integrins, it is likely that integrin activation by sCD40L is biologically relevant, since CD40L is a membrane-bound protein and is highly concentrated on the cell surface.

CD40L binds to another ligand-binding site (site 2) in integrin $\alpha \beta 3$

We previously showed that several integrin ligands (e.g., CX3CL1, CXCL12, and sPLA2-IIA) bound to an allosteric site (site 2) and activated several integrins (14–16). We performed docking simulation of interaction between $\alpha \beta 3$ (1JV2.pdb, with closed headpiece) and CD40L monomer (1ALY.pdb) using Autodock3. The simulation predicts that monomeric CD40L binds to site 2 well (docking energy -20.5 kcal/mol) (Fig. 1D).

The site-2-binding interface of sCD40L predicted to be located in the outside of trimer

We previously reported that the peptide from site 2 of $\beta 3$ (QPNDGQSHVGSNDNHYSASTTM, residues 267–287 of $\beta 3$, Cys-273 is changed to S, fused to GST) directly bound to CX3CL1, sPLA2-IIA, and CXCL12 and the peptide suppressed integrin activation by these activators, suggesting that they directly bind to site 2 and mediate integrin activation (14–16). To prove if CD40L binds to site 2, we tested if CD40L binds to peptides derived from site 2. Linear site 2 peptides did not show good binding to CD40L (not shown). We thus designed disulfide-linked cyclic 28-mer peptides by introducing two Cys residues at both ends to enhance affinity and stability of the peptides. To predict the positions of the two Cys residues that do not affect the peptide conformation of the peptides we used the Disulfide by Design-2 (DbD2) software (18). We found that cyclic site 2 peptide from $\beta 3$ (C260-RLAGIVQPNDGQSHVGSNDNHYSASTTMC288) and the corresponding $\beta 1$ peptide bound to sCD40L, suggesting that sCD40L binds to site 2 (Fig. 2A). The position of cyclic site 2 peptide is shown in Figure 2B. These findings suggest that sCD40L activates integrins by binding to site 2.

Localization of amino acid residues of CD40L involved integrin activation

The docking simulation predicts that four HIGM1 mutations (S128R/E129G, K143T, G144E, and L155P) are clustered

Table 1
Amino acid residues involved in the predicted CD40L- $\alpha \beta 3$ (closed form) interaction

CD40L	αV	$\beta 3$
Glu129 , Ala130, Ser131, Lys133, Thr134, Thr135, Glu142, Lys143 , Gly144 , Tyr145, Tyr146, Leu155 , Lys159, Phe177, Cys178, Ser179, Asn180, Arg181, Ala183, Ser184, Pro217, Cys218, Gln220, Pro244, Ser245, Gln246, Val247, Ser248, His249, Gly250, Thr251,	Pro14, Glu15, Asn44, Thr45, Thr46, Gln47, Pro48, Gly49, Ile50, Val51, Glu52, Gln68, Gly76, Asn77, Arg78, Asp79, Ala81, Lys82, Asp83, Asp84, Pro85, Glu87, Phe88, Lys89, Ser90, His91, Arg122,	Met165, Ser168, Glu171, Glu174, Asn175, Pro186, <u>Asp278</u> , <u>His280</u> , <u>Tyr281</u> , Ser282, Ala283, Ser284, <u>Thr285</u> , <u>Thr286</u>

The amino acid residues in CD40L that are mutated in HIGM1 are in bold.
The amino acid residues in $\beta 3$ that are involved in site 2 peptide are underlined.

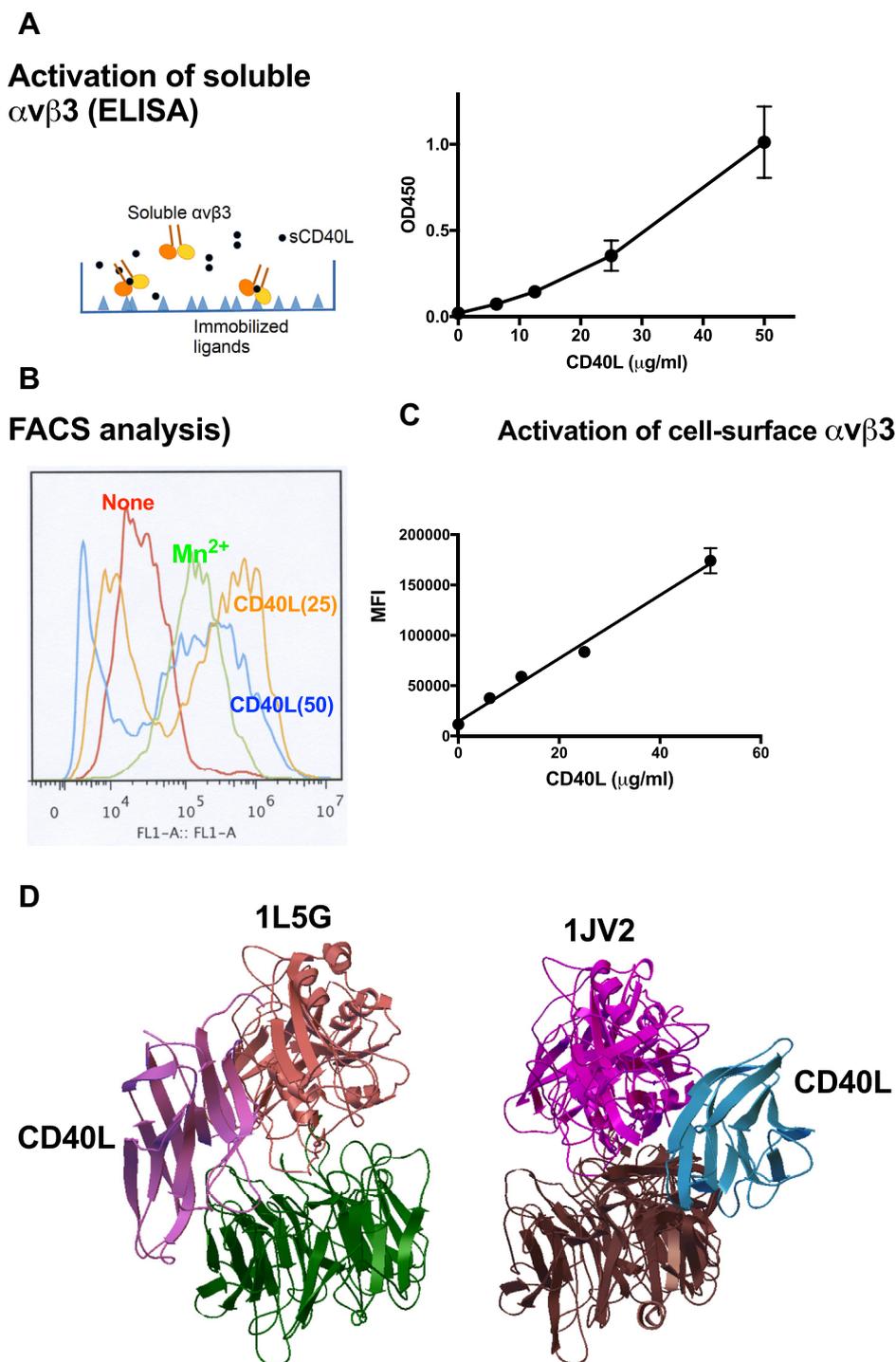


Figure 1. CD40L activates soluble and cell-surface integrin $\alpha\beta 3$. A, WT soluble CD40L activates soluble integrin $\alpha\beta 3$. Wells of 96-well microtiter plate were coated with γC399tr (50 $\mu\text{g/ml}$ in PBS) for 2 h at room temperature and incubated with soluble $\alpha\beta 3$ (5 $\mu\text{g/ml}$) and sCD40L for 1 h in HEPES-Tyrodes buffer with 1 mM Ca^{2+} (which keeps $\alpha\beta 3$ in an inactive form). Bound $\alpha\beta 3$ was quantified using anti- $\beta 3$ antibody (AV10). B, WT soluble CD40L activates cell-surface $\alpha\beta 3$. $\beta 3$ -CHO cells were incubated with sCD40L (25 $\mu\text{g/ml}$) for 30 min on ice in HEPES-Tyrodes buffer with 0.02% BSA and 1 mM Ca^{2+} and then with FITC-labeled γC399tr for 1 h at room temperature. The binding of γC399tr was measured in flow cytometry. A typical flow cytometric profile is shown. C, dose dependency of activation of cell surface $\alpha\beta 3$ (MFI, mean fluorescent intensity). D, Docking models. We performed docking simulation between CD40L (1AYP.pdb) and inactive $\alpha\beta 3$ using open (1L5G.pdb) or closed headpiece (1JV2.pdb) as a target. Docking model of interaction between $\alpha\beta 3$ (open headpiece, 1L5G.pdb)-CD40L (1AYP.pdb) was taken from (13).

in the predicted site-2-binding interface of CD40L (Fig. 2B). Amino acid residues involved in the predicted CD40L- $\alpha\beta 3$ (closed form) interaction are shown in Table 1. We studied if these HIGM1 mutations affect CD40L-induced $\alpha\beta 3$

activation. Notably, the K143T and G144E mutants were defective in activating soluble $\alpha\beta 3$ (Fig. 2C) and were defective in activating cell surface $\alpha\beta 3$ (Fig. 2D). These findings are consistent with the docking model that these mutations

Allosteric integrin activation by CD40L

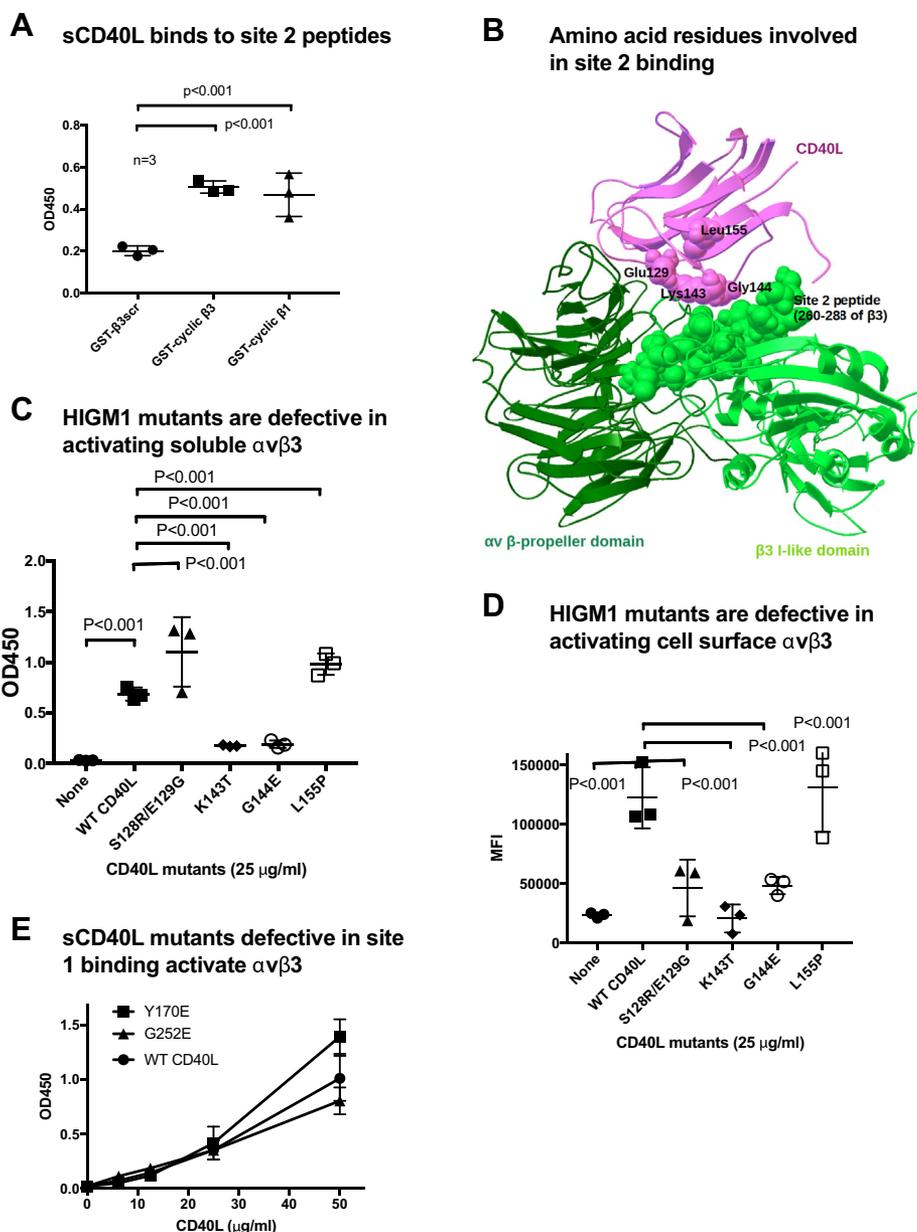


Figure 2. Localization of CD40L-binding site in integrin $\alpha v \beta 3$ and the integrin-binding site in CD40L. **A**, CD40L binds to cyclic site 2 peptide. CD40L was immobilized and incubated with cyclic site 2 peptides of integrin $\beta 1$ or $\beta 3$ subunit fused to GST. Bound GST was measured using HRP-conjugated anti-GST antibodies. Scrambled linear $\beta 3$ site 2 peptide fused to GST was used as a control. **B**, positions of four HIGM1 mutants in CD40L in the predicted site 2 binding site in CD40L. Docking simulation of CD40L binding to site 2 between CD40L (1ALY.pdb) and the $\alpha v \beta 3$ headpiece (1JV2.pdb, closed-headpiece) was performed. Position of site-2-derived peptide in integrin $\beta 3$ was shown. **C**, activation of soluble integrin $\alpha v \beta 3$ by sCD40L HIGM1 mutants in ELISA-type assays. These assays were performed as described in Figure 1 legend. Data are shown as means \pm SD ($n = 3$). **D**, activation of cell-surface integrin $\alpha v \beta 3$ by sCD40L HIGM1 mutants by cell binding assays and flow cytometry. These assays were performed as described in Figure 1 legend. Data are shown as means \pm SD ($n = 3$). **E**, CD40L mutants defective in binding to site 1 allosterically activate integrin. The CD40L Y170E and G252E mutants (13), which do not bind to the classical ligand-binding site (site 1), activate soluble integrin $\alpha v \beta 3$.

(K143T and G144E) are located in the predicted site-2-binding site of CD40L (Fig. 2B).

Previous studies identified amino acid residues in the trimeric interface that are involved in binding to the classical ligand-binding site (site 1) of activated integrins (13). Consistently, the two CD40L mutations Y170E and G252E in the trimeric interface are defective in binding to site 1 (13), but activated soluble $\alpha v \beta 3$ (Fig. 2E). This finding is consistent with

the model that site-1-binding and site-2-binding sites are distinct.

sCD40L activates soluble and cell-surface integrin $\alpha 5 \beta 1$

Previous studies showed that integrin $\alpha 5 \beta 1$ binds to CD40L and induces signals independent of CD40 (12). We previously identified the binding site for $\alpha 5 \beta 1$ in the trimeric interface of CD40L, indicating that $\alpha 5 \beta 1$ and $\alpha v \beta 3$ -binding sites overlap in

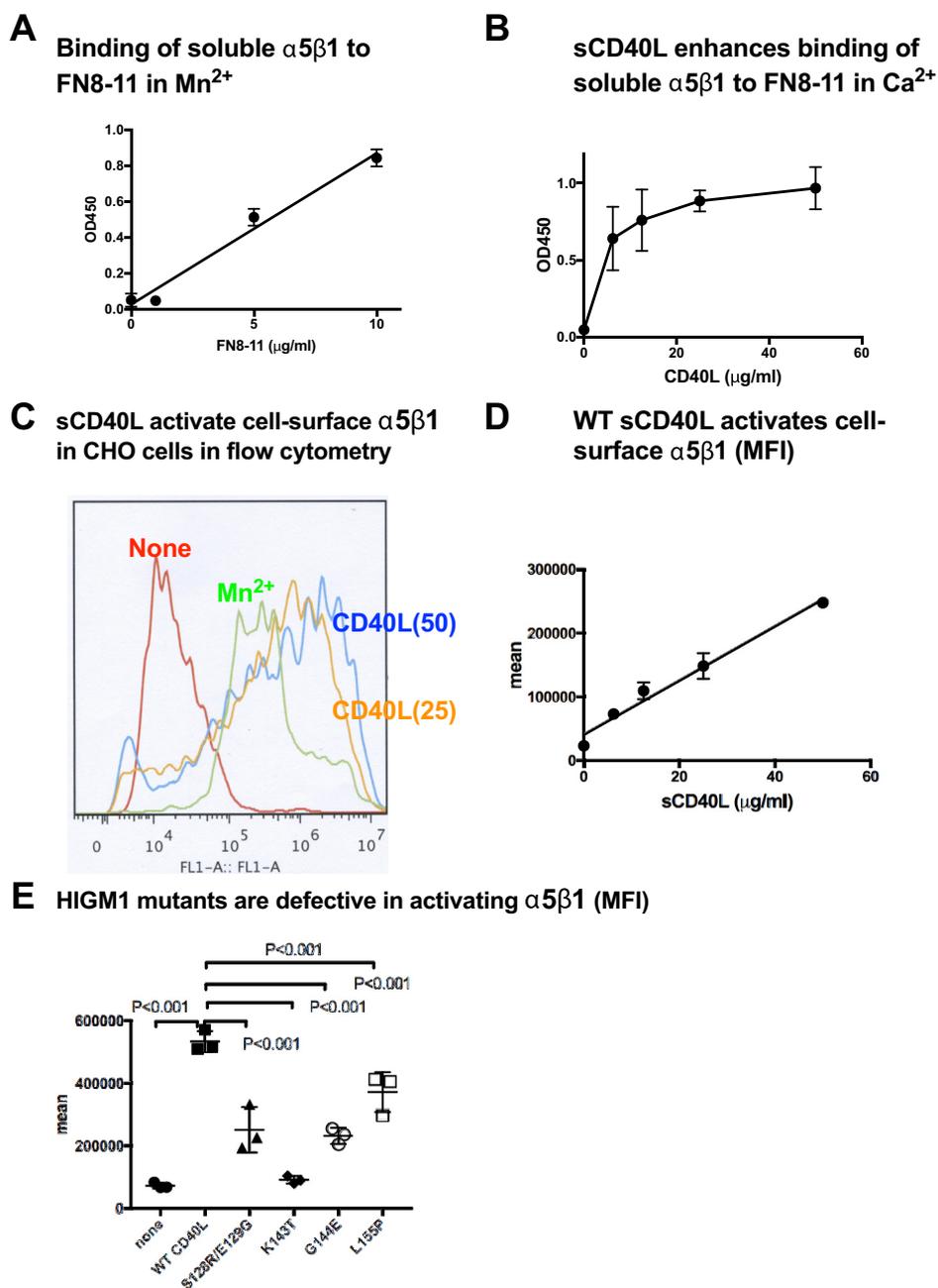


Figure 3. sCD40L activates soluble and cell-surface $\alpha 5\beta 1$. A, biotinylated soluble $\alpha 5\beta 1$ binds to the fibronectin cell-binding fragment (FN8-11). The fibronectin domains 8-11 conjugated to GST was coated to wells of 96-well microtiter plate. Remaining protein-binding sites were blocked with BSA. Soluble biotinylated soluble $\alpha 5\beta 1$ (1 $\mu g/ml$) in 50 μl Tyrode-HEPES buffer with 1 mM Mn^{2+} was added and incubated for 1 h at 37 $^{\circ}C$. Bound $\alpha 5\beta 1$ was quantified using HRP-conjugated streptavidin. Data are shown as means \pm SD (n = 3). B, WT sCD40L activates soluble integrin $\alpha 5\beta 1$. Wells of 96-well microtiter plate were coated with FN8-11 (50 $\mu g/ml$ in PBS) for 2 h at room temperature and incubated with biotinylated soluble $\alpha 5\beta 1$ (1 $\mu g/ml$) and sCD40L for 1 h in HEPES-Tyrodes buffer with 1 mM Ca^{2+} (which keeps $\alpha 5\beta 1$ in an inactive form). Bound $\alpha 5\beta 1$ was quantified using HRP-conjugated streptavidin. C, activation of cell-surface $\alpha 5\beta 1$ in CHO cells. This assay was performed as described in Figure 1 legend, except that CHO cells and FITC-FN8-11 were used. The data were analyzed using FlowJo. A typical flow cytometric profile is shown. D, dose dependency of activation of cell surface $\alpha 5\beta 1$ (mean fluorescent intensity). This assay was performed as described in Figure 1 legend. Data are shown as means \pm SD (n = 3). E, activation of cell-surface integrin $\alpha 5\beta 1$ by sCD40L HIGM1 mutants by cell binding assays and flow cytometry. These assays were performed as described in Figure 1 legend. Data are shown as means \pm SD (n = 3).

the trimeric interface of CD40L (13). We hypothesized that CD40L activates $\alpha 5\beta 1$ as well in an allosteric manner.

We confirmed that biotinylated soluble $\alpha 5\beta 1$ binds to the fibronectin fragment (FN8-11), a specific ligand to $\alpha 5\beta 1$, in ELISA-type activation assays in the presence of 1 mM Mn^{2+} (Fig. 3A). FN8-11 binds to site 1, but does not bind to site 2

(14). We studied if sCD40L activates soluble $\alpha 5\beta 1$. Activation is defined by the increase in integrin binding to immobilized FN8-11 by soluble sCD40L. FN8-11 was immobilized and incubated with biotinylated soluble $\alpha 5\beta 1$ in the presence of 1 mM Ca^{2+} . sCD40L enhanced the binding of soluble $\alpha 5\beta 1$ to immobilized FN8-11 in a dose-dependent manner (Fig. 3B).

Allosteric integrin activation by CD40L

These findings indicate that sCD40L activates soluble integrin $\alpha 5\beta 1$ in cell-free conditions.

We showed that sCD40L activated cell-surface $\alpha 5\beta 1$ in CHO cells ($\alpha 5\beta 1+$, CD40 $-$) using FITC-labeled FN8-11 in a dose-dependent manner (Fig. 3B). The levels of $\alpha 5\beta 1$ activation by sCD40L in MFI and sCD40L concentrations correlated well (Fig. 3, C and D). We studied if four HIGM1 mutants can activate cell-surface $\alpha 5\beta 1$ in CHO cells. They were defective in activating $\alpha 5\beta 1$ and the K143T was the most defective (Fig. 3E). These findings suggest that sCD40L binds to site 2 of $\alpha 5\beta 1$ in a manner similar to that of $\alpha \nu\beta 3$ and activates $\alpha 5\beta 1$.

sCD40L activates soluble and cell-surface integrin $\alpha 4\beta 1$

CD40L is primarily expressed in activated CD4 $+$ T cells, but $\alpha \nu\beta 3$ and $\alpha 5\beta 1$ are not major integrins in T cells. sCD40L binds to site 1 of activated $\alpha \nu\beta 3$ and $\alpha 5\beta 1$ (13). It is unclear if integrin $\alpha 4\beta 1$, which is expressed primarily in immune-competent cells, interacts with CD40L. We used biotinylated soluble $\alpha 4\beta 1$ to study if $\alpha 4\beta 1$ binds to CD40L. We found that soluble $\alpha 4\beta 1$ activated by 1 mM Mn^{2+} bound to immobilized WT sCD40L in a dose-dependent manner (Fig. 4A), indicating that CD40L is a ligand for $\alpha 4\beta 1$ that binds to site 1.

We then studied if sCD40L activates $\alpha 4\beta 1$ by binding to site 2. A fibronectin fragment H120, a ligand specific to $\alpha 4\beta 1$, binds to site 1, but does not bind to site 2 (14). Activation is defined by the increase in the binding of soluble integrin $\alpha 4\beta 1$ to immobilized H120 by sCD40L. We found that sCD40L enhanced the binding of soluble $\alpha 4\beta 1$ to H120 in a dose-dependent manner in the presence of 1 mM Ca^{2+} (Fig. 4B).

Also, sCD40L enhanced the binding of FITC-labeled H120 to CHO cells that express recombinant $\alpha 4\beta 1$ ($\alpha 4$ -CHO cells) in the presence of 1 mM Ca^{2+} (Fig. 4C). The $\alpha 4\beta 1$ activation (MFI) correlated well with the sCD40L concentrations, indicating that sCD40L activated cell-surface $\alpha 4\beta 1$ (Fig. 4D). We studied if the four HIGM1 mutants that are clustered in the predicted site-2-binding site of CD40L can activate $\alpha 4\beta 1$ on $\alpha 4$ -CHO cells. The HIGM1 mutants were defective in activating cell surface $\alpha 4\beta 1$. The K143T and G144E mutants were the most defective in activating $\alpha 4\beta 1$ among four HIGM1 mutants (Fig. 4E), indicating that site-2-binding interface in $\alpha 4\beta 1$ is similar to those of $\alpha \nu\beta 3$ and $\alpha 5\beta 1$. These findings suggest that $\alpha 4\beta 1$ is a new receptor for CD40L. CD40L binds to site 2 and activates $\alpha 4\beta 1$ in a manner similar to those of $\alpha \nu\beta 3$ and $\alpha 5\beta 1$.

Characteristics of four HIGM1 mutants

We further characterized the four HIGM1 mutants to determine the role of site 2 in CD40L signaling. They all bound to CD40 in ELISA (Fig. 5A). We tested their ability to bind to activated soluble $\alpha \nu\beta 3$ in ELISA in the presence of 1 mM Mn^{2+} , which reflects their ability to bind to site 1. S128R/E129G and L155P were defective and K143T and G144E were partially defective in binding to activated integrin $\alpha \nu\beta 3$, suggesting that K143T and G144E are still able to bind to site 1 (Fig. 5B). S128R/E129G and L155P did not bind to activated $\alpha \nu\beta 3$ (to site 1) although these mutations are not in the site-1-binding site in

the trimeric interface. It is possible that these mutations induced conformational changes in the trimeric interface.

We determined the ability of HIGM1 mutants to induce NF- κ B activation in HEK293 reporter cells. The four mutants were all defective in inducing NF- κ B activation in reporter cells, except that K143T is slightly active (Fig. 5C). K143T at high concentration (5 μ g/ml) induced substantial NF- κ B activation (Fig. 5D). We tested if excess (20-fold) mutants can suppress NF- κ B activation by WT sCD40L. We found that all mutants tested suppressed NF- κ B activation by WT sCD40L (Fig. 5D), indicating that they act as antagonists. We previously showed that eight other HIGM1 mutants, which are clustered in the trimeric interface, were defective in integrin binding (to site 1). Their defect in site 1 binding is likely related to their defect in CD40L signaling (13). The two mutants, K143T and G144E, still bind to site 1 but defective in integrin activation by binding to site 2. We thus propose that the binding of CD40L to site 2 is required for NF- κ B activation.

Discussion

Potential biological role of allosteric integrin activation by CD40L

In the present study, we showed that sCD40L activated soluble and cell-surface integrins $\alpha \nu\beta 3$, $\alpha 5\beta 1$, and $\alpha 4\beta 1$ in a dose-dependent manner. These findings indicate that this activation does not require inside-out signaling. Since cyclic site 2 peptide bound to sCD40L, it is suggested that sCD40L bound to site 2. Docking simulation using inactive/close headpiece $\alpha \nu\beta 3$ as a target predicts that CD40L binds to site 2 and four HIGM1 mutants are clustered in the predicted site-2-binding site in CD40L (Fig. 6A). We showed that the HIGM1 mutants (particularly K143T and G144E) are defective in activating integrins, consistent with the prediction. The present study showed that sCD40L is a new allosteric activator of integrins by binding to site 2. The four HIGM1 mutants, including K143T and G144E, were defective in activating integrins $\alpha \nu\beta 3$, $\alpha 5\beta 1$, and $\alpha 4\beta 1$ because they are defective in site 2 binding. This suggests that the defective binding of CD40L to site 2 is related to defective CD40L/CD40 signaling. Since integrins on normal blood cells (e.g., B cells) are not activated, it is possible that CD40L-mediated integrin activation by binding to site 2 is required for CD40L/CD40 signaling, in addition to site 1 binding. Previous study showed that eight HIGM1 mutants clustered in the trimeric interface are defective in integrin binding and in signaling because they are defective in site 1 binding (13), indicating that the defect in integrin binding to site 1 is related to defective CD40L/CD40 signaling. We thus propose that CD40L-mediated integrin activation by binding to site 2 is also required for CD40L/CD40 signaling. Since integrins are not activated in normal leukocytes, integrin activation by this mechanism will facilitate signaling by CD40L.

It is well known that high concentrations of sCD40L (>1000-fold) are required to induce CD40L/CD40 signaling compared with membrane-bound CD40L. In our previous

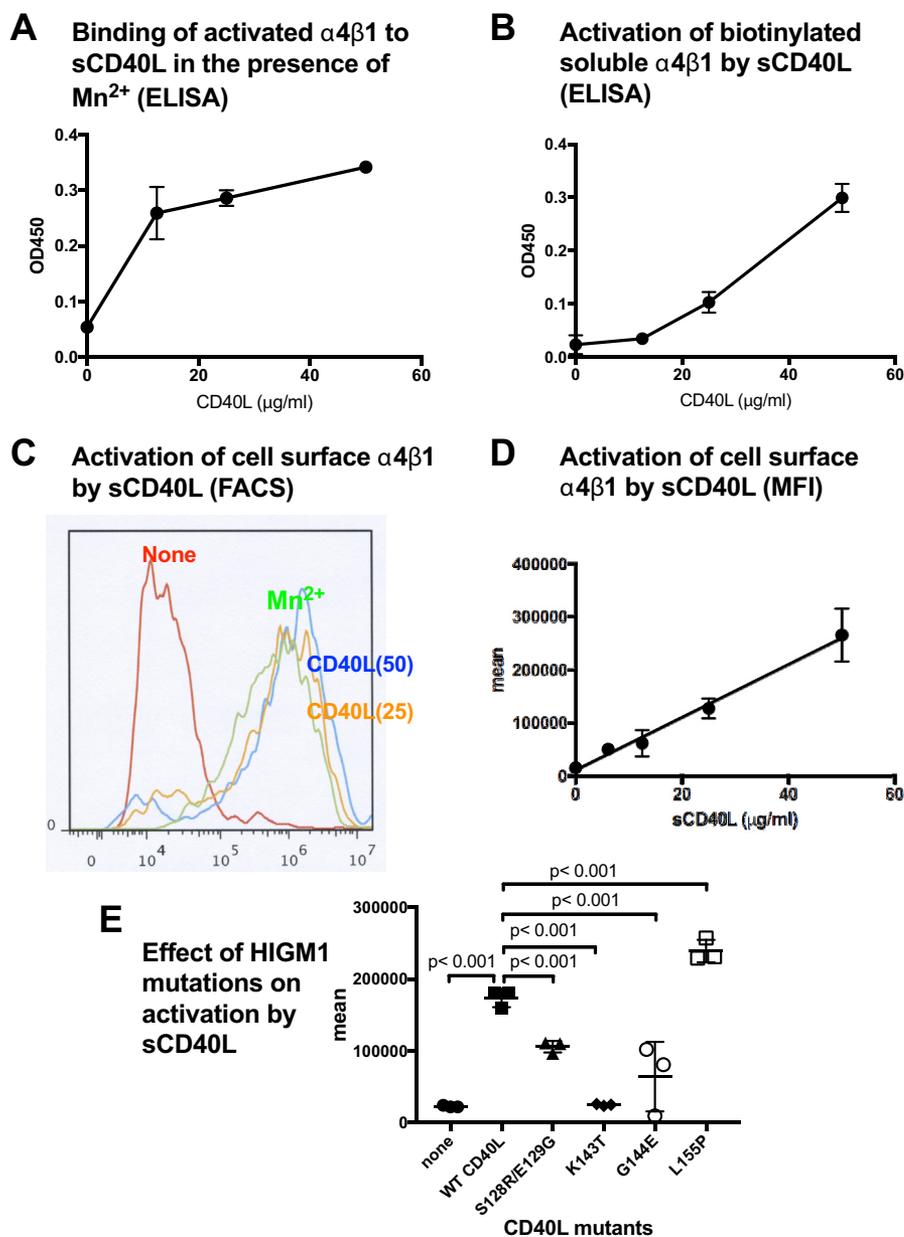


Figure 4. sCD40L activates soluble and cell-surface $\alpha 4\beta 1$. A, WT sCD40L activates soluble integrin $\alpha 4\beta 1$. Wells of 96-well microtiter plate were coated with H120 (50 $\mu\text{g/ml}$ in PBS) for 2 h at room temperature and incubated with biotinylated soluble $\alpha 4\beta 1$ (1 $\mu\text{g/ml}$) and sCD40L for 1 h in HEPES-Tyrodes buffer with 1 mM Ca^{2+} (which keeps $\alpha 4\beta 1$ in an inactive form). Bound $\alpha 4\beta 1$ was quantified using HRP-conjugated streptavidin. Data are shown as means \pm SD ($n = 3$). B, WT sCD40L activates soluble integrin $\alpha 4\beta 1$. Wells of 96-well microtiter plate were coated with FN8-11 (50 $\mu\text{g/ml}$ in PBS) for 2 h at room temperature and incubated with biotinylated soluble $\alpha 5\beta 1$ (1 $\mu\text{g/ml}$) and sCD40L for 1 h in HEPES-Tyrodes buffer with 1 mM Ca^{2+} (which keeps $\alpha 5\beta 1$ in an inactive form). Bound $\alpha 5\beta 1$ was quantified using HRP-conjugated streptavidin. C, WT sCD40L activates cell-surface $\alpha 4\beta 1$. $\alpha 4$ -CHO cells were incubated with sCD40L (25 $\mu\text{g/ml}$) for 30 min on ice in HEPES-Tyrodes buffer with 0.02% BSA and 1 mM Ca^{2+} and then with FITC-labeled H120 for 1 h at room temperature. The binding of H120 was measured in flow cytometry. A typical flow cytometric profile is shown. D, dose dependency of activation of cell surface $\alpha 4\beta 1$ (mean fluorescent intensity). Data are shown as means \pm SD ($n = 3$). E, activation of cell-surface integrin $\alpha 4\beta 1$ by sCD40L HIGM1 mutants by cell binding assays and flow cytometry. These assays were performed as described in Figure 1 legend. Data are shown as means \pm SD ($n = 3$).

study, sCD40L at $>1 \mu\text{g/ml}$ was needed to achieve maximal NF- κB activation in HEK293 reporter cells (13). We proposed that CD40L/CD40 signaling requires direct integrin binding to CD40L (and subsequent integrin-CD40L-CD40 ternary complex formation). Integrins are not high-affinity receptors compared with CD40. This partly explains that high concentrations of sCD40L are required for CD40L/CD40 signaling. In

the present study, very high concentrations of sCD40L were required to detect integrin activation. It is unlikely that integrin activation can be induced by sCD40L under physiological conditions even in autoimmune disease patients (e.g., SLE). We thus propose that integrin activation is primarily induced by membrane-bound CD40L in activated T cells or platelets, in which CD40L is brought to the cell surface upon

Allosteric integrin activation by CD40L

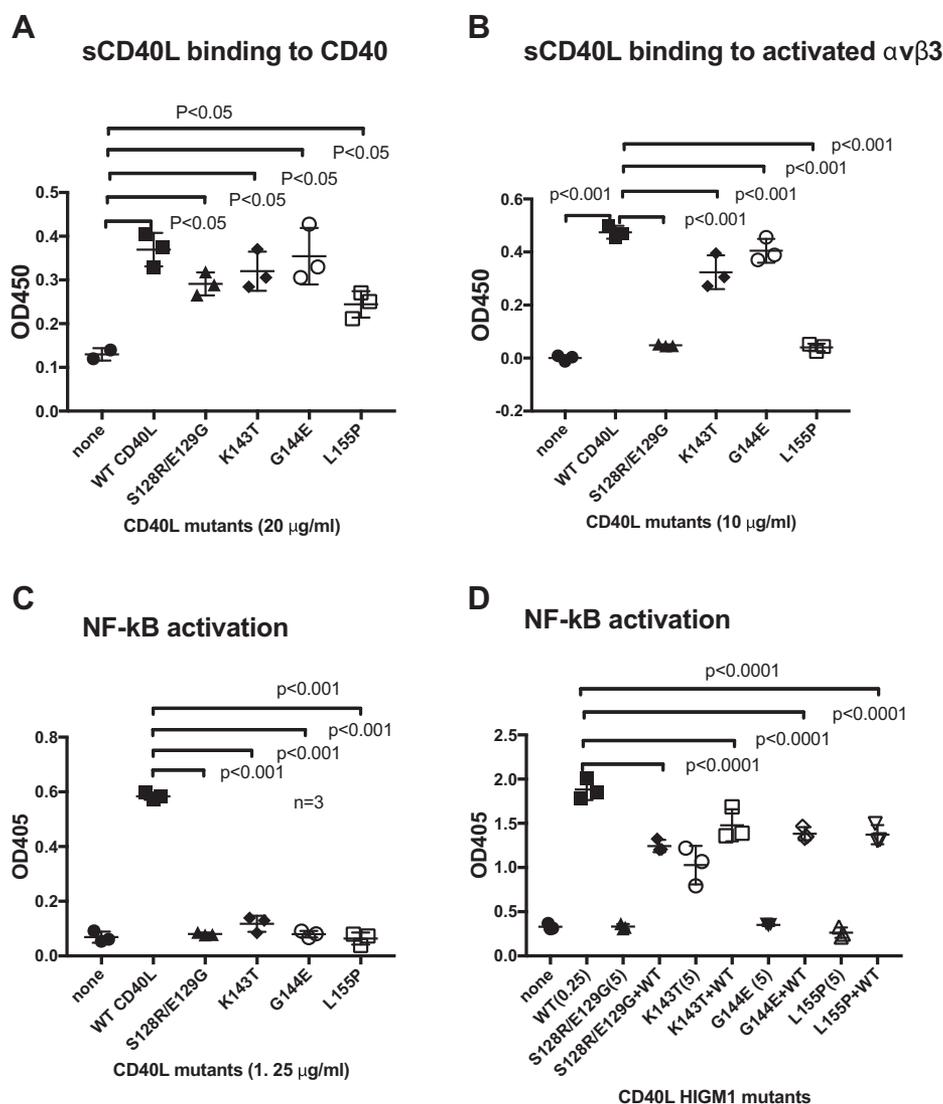


Figure 5. Four HIGM1 mutants clustered in the predicted site-2-binding region of CD40L are defective in signaling and act as antagonists. *A*, binding of the CD40 fragment fused to GST to CD40L (WT and HIGM1 mutants). The CD40 fragment (residues 21–144) fused to GST (100 µg/ml in PBS) was immobilized to wells of a 96-well microtiter plate and incubated with sCD40L mutants. Bound sCD40L mutants were measured using anti-histidine Abs. Data are shown as means ± SD ($n = 3$). *B*, binding of sCD40L to site 1 of soluble integrin $\alpha v \beta 3$ in the presence of 1 mM Mn^{2+} . sCD40L (residues 118–261) was immobilized to wells of a 96-well microtiter plate and incubated with soluble $\alpha v \beta 3$ (5 µg/ml) in HEPES-Tyroses buffer (1 mM Mn^{2+}). Bound $\alpha v \beta 3$ was measured using anti- $\beta 3$ (AV10). Data are shown as means ± SD ($n = 3$). *C*, NF- κB activation by sCD40L (WT and HIGM1 mutants). HEK293 cells that express CD40 and NF- κB reporter gene (SEAP) were incubated with sCD40L in serum-free Dulbecco's modified Eagle's medium for 4 h, and SEAP in the medium was determined. Data are shown as means ± SD ($n = 3$). *D*, suppression of NF- κB activation by HIGM1 mutants. NF- κB activation was measured in reporter cells as described in (*C*). WT CD40L (0.25 µg/ml) and mutants (5 µg/ml) were used. Data are shown as means ± SD ($n = 3$).

activation of T cells or platelets, and CD40L is highly concentrated on the cell surface. It is, however, possible that sCD40L can be concentrated by binding to cell surface CD40. If so, sCD40L at levels in autoimmune disease patients may sufficiently trigger integrin activation and subsequent CD40/CD40L signaling for inflammatory responses. The present results provide the first evidence that CD40L can induce integrin activation, regardless of whether it is soluble or membrane-bound form.

The binding of CD40L to site 2 of integrins may lead to proinflammatory outside-in signaling, as shown by the binding of 25-hydroxycholesterol to site 2 (17). It is possible that integrin activation by binding of CD40L to site 2 will facilitate other growth factor/cytokine signaling, since direct integrin binding of growth

factors/cytokines/proinflammatory proteins to the classical ligand-binding site (site 1) (Fig. 6B) is required for their signaling functions. This includes FGF-1 (19), IGF-1 (20), and neuregulin-1 (21), CX3CL1 (22), IL-1 β (23), CD40L (13), and sPLA2-IIA (24). It is imperative to study the role of allosteric integrin activation in growth factor/cytokine signaling in future studies.

Integrin activation by inside-out signaling versus integrin activation by site 2 binding

It has been generally accepted that integrins are activated by inside-out signaling. Integrin activation is accompanied by global conformational changes (25, 26). It has been, however, reported that inside-out signaling enhanced clustering of

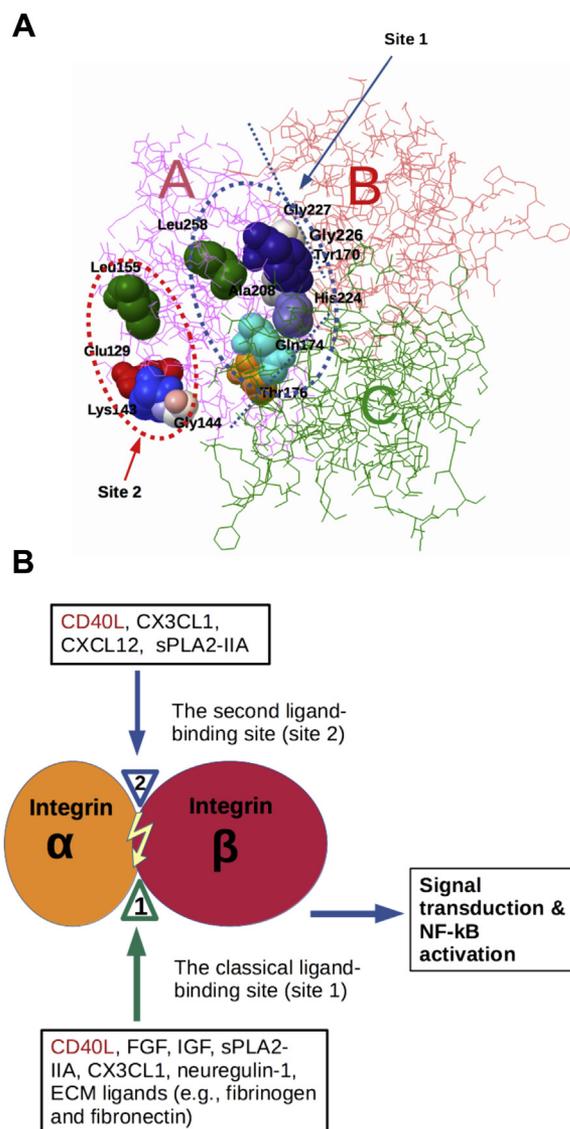


Figure 6. CD40L integrin interaction and its potential roles. *A*, clustering of HIGM1 mutants in site 1 and site-2-binding sites in CD40L. Previous study showed that eight HIGM1 mutants are clustered in the trimeric interface of CD40L. The present study shows that four HIGM1 mutants are clustered in the site-2-binding site of CD40L in the outside of the CD40L trimer. *B*, integrin ligands that bind to site 1 and site 2. We identified the second ligand-binding site (site 2) by docking simulation of ligand binding using inactive integrin as a target. CD40L binding to site 2 enhances ligand binding to site 1 of integrins. The ligand specificity to site 2 overlaps with that of site 1, but ECM ligands do not appear to bind to site 2.

integrin α IIB β 3 and global conformational changes in thrombin-activated platelets but did not enhance ligand-binding affinity to monovalent ligand (27). Also, inside-out signaling induced by cross-linking of T cell receptor did not enhance ligand-binding affinity of leukocyte integrins to monovalent ligand (28). It is possible that inside-out signaling does not enhance ligand affinity (the integrin headpiece is still in close conformation). The present study showed that sCD40L enhanced ligand-binding affinity of integrins to monomeric ligands by binding of CD40L to site 2 in the extracellular milieu. Previous studies showed that CX3CL1 (14), CXCL-12 (16), and sPLA2-IIA (15) activated integrins by binding to site 2 in the absence of inside-out signaling. We

propose that site-2-mediated integrin activation and inside-out signaling occur independently and complement to each other.

Experimental procedure

Materials

Recombinant soluble $\alpha\beta$ 3 was synthesized in Chinese hamster ovary (CHO) K1 cells using the soluble α and β expression constructs and purified by Ni-NTA affinity chromatography, as described (29). Biotinylated soluble α 5 β 1 and α 4 β 1 were obtained from Acro Biosystems. β 3-CHO and α 4-CHO cells have been described (24). Fibrinogen γ -chain C-terminal domain that lacks residues 400–411 (γ C399tr) was synthesized as described (30). GST-fusion proteins of fibronectin type III domains 8–11 (FN8-11) and fibronectin H120 fragment (FN-H120) were described (22). Anti-human β 3 mAb AV10 was provided by B. Felding (The Scripps Research Institute, La Jolla, CA). HRP-conjugated anti-His tag antibody was purchased from Qiagen (Valencia, CA).

Synthesis of recombinant sCD40L with no KGD motif

We synthesized recombinant sCD40L (residues 118–261, QNPQIAAHVISEASSKTTSTVLQWAEKGYTMSNNLVTLENGKQLTVKRQGLYYIYAQVTFCSNREASSQAPFIASLCLKSPGRFERILLRAANTHSSAKPCGQQSIHLGGVFELQPGASVFNVTDPSSQVSHGTGFTSFGLLKL) with no N-terminal KGD motif as described (13). sCD40L mutants were generated by site-directed mutagenesis of wild-type (WT) sCD40L (residues 118–261) and synthesized as described for WT sCD49L.

Synthesis of CD40 fused to GST

The cDNA fragment encoding the CD40 fragment (residues 21–144) was amplified by PCR and subcloned into the BamHI/EcoRI site of PGEX2T. We synthesized the proteins in BL21 cells and purified using glutathione-Sepharose affinity chromatography.

Binding of the CD40 fragment fused to GST to CD40L

The CD40 fragment fused to GST was coated to wells of a 96-well microtiter plate (100 μ g/ml in PBS) for 1 h, and the remaining protein-binding sites were blocked by BSA (0.1%). We then incubated the wells with sCD40L and incubated for 1 h, and bound sCD40L was measured using HRP-conjugated anti-His.

Synthesis of cyclic site 2 peptides

We introduced 6His tag to the BamHI site of pGEX-2T (resulting vector is designated pGEX-2T6His). We synthesized GST fusion protein of site 2 peptide (QPNDGQSHVGSNDNHYSASTTM, residues 267–287 of β 3, C273 is changed to S) and a scrambled site 2 peptide (VHDSHYSGQGAMSDNTNSPQT) by subcloning oligonucleotides that encodes these sequences into the BamHI/EcoRI site of pGEX-2T6His as described (14, 15). We introduced a disulfide linkage that connects both ends of the site 2 peptide

Allosteric integrin activation by CD40L

without affecting its conformation using Disulfide by Design-2 (DbD2) software (<http://cptweb.cpt.wayne.edu/DbD2/>) (18). It predicted that mutating Gly260 and Asp288 to Cys disulfide-linked cyclic site 2 peptide of $\beta 3$ does not affect the conformation of the peptide. We generated C260-RLAGI VQPNDGQSHVGSNDHYSASTTMC288, 29-mer cyclic $\beta 3$ peptide. We designed the corresponding cyclic $\beta 1$ peptide (C268-KLGGIVLPNDGQSHLENNMYTMSHYCY295, 28-mer cyclic $\beta 1$ peptide) in which C281 is converted to S. We synthesized the proteins in BL21 cells and purified using glutathione-Sepharose affinity chromatography.

Binding of soluble integrins to ligands

ELISA-type binding assays were performed as described previously (22). Briefly, wells of 96-well Immulon 2 microtiter plates (Dynatech Laboratories, Chantilly, VA) were coated with 100 μ l PBS containing γ C399tr for 2 h at 37 °C. Remaining protein-binding sites were blocked by incubating with PBS/0.1% BSA for 30 min at room temperature. After washing with PBS, soluble recombinant $\alpha\beta 3$ (5 μ g/ml) in the presence or absence of CD40L (WT or mutant) was added to the wells and incubated in HEPES-Tyroses buffer (10 mM HEPES, 150 mM NaCl, 12 mM NaHCO₃, 0.4 mM NaH₂PO₄, 2.5 mM KCl, 0.1% glucose, 0.1% BSA) with 1 mM CaCl₂ for 1 h at room temperature. After unbound $\alpha\beta 3$ was removed by rinsing the wells with binding buffer, bound $\alpha\beta 3$ was measured using anti-integrin $\beta 3$ mAb (AV-10) followed by HRP-conjugated goat anti-mouse IgG and peroxidase substrates. For activation assays with $\alpha 5\beta 1$, biotinylated soluble $\alpha 5\beta 1$ and $\alpha 4\beta 1$ (AcroBio) and their specific ligands (FN8-11 and H120, respectively) were used, and bound biotinylated soluble integrins were measured by using streptavidin conjugated to HRP.

Flow cytometry

CHO cells were cultured in Dulbecco's modified Eagle's medium (DMEM)/10% fetal calf serum. The cells were resuspended with HEPES-Tyroses buffer/0.02% BSA (heat-treated at 80 °C for 20 min to remove contaminating cell adhesion molecules). The $\beta 3$ -CHO or CHO cells were then incubated with WT or mutant CD40L for 30 min on ice and then incubated with FITC-labeled integrin ligands (γ C399tr, FN8-11 and H120) for 30 min at room temperature. The cells were washed with PBS/0.02% BSA and analyzed by BD Accuri flow cytometer (Becton Dickinson, Mountain View, CA). The data were analyzed using FlowJo 7.6.5.

CD40L reporter assays

We used HEK293 cells that express human CD40L-soluble embryonic alkaline phosphatase (SEAP) reporter (Invivogen). Cells were maintained in DMEM/10% fetal calf serum. For reporter assays, cells were plated in wells of a 24-well culture plate (2.5 $\times 10^5$ cells in 500 μ l DMEM without fetal calf serum) and stimulated with CD40L for 4 h. We measured the levels of alkaline phosphatase activity in the medium, as described (31).

Docking simulation

Docking simulation of interaction between CD40L (Protein Data Bank code 1ALY), which does not contain the N-terminal KGD motif, and integrin $\alpha\beta 3$ was performed using AutoDock3, as described (24). In the current study, we used the headpiece (residues 1–438 of α and residues 55–432 of $\beta 3$) of $\alpha\beta 3$ (closed-headpiece form, Protein Data Bank code 1JV2). Cations were not present in $\alpha\beta 3$ during docking simulation (15, 16).

Statistical analysis

Treatment differences were tested using ANOVA using Prism 7 (GraphPad Software).

Data availability

Coordinate of the docking model presented in this paper is available upon request. All remaining data are contained within the article.

Acknowledgments—We are grateful to Dr Sam Hwang for allowing us to use flow cytometry.

Author contributions—Y. K. T. performed the experiments. Y. T. conceived the project, designed the experiments, analyzed the data, and wrote the article. M. S., E. M., R. H. C., and B. H. F. analyzed the data, discussed, and edited the article.

Funding and additional information—This work was supported by UC Davis STAIR grant (Y. T., M. S.), NCI R01 CA208669-01A1 (B. H. F.), 3R01CA232845-02S1 (M. S.), and UCDCCC Pilot Funding (Y. T., M. S.) and NIH UG3TR002866 and NIFA CADMCB-7399-H (R. H. C.). This work is partly supported by the UC Davis Comprehensive Cancer Center Support Grant (CCSG) awarded by the National Cancer Institute (NCI P30CA093373).

Conflict of interest—The authors declare that they have no conflicts of interest with the contents of this article.

Abbreviations—The abbreviations used are: APC, antigen presenting cell; DMEM, Dulbecco's modified Eagle's medium; HIGM1, high IgM syndrome type 1; SLE, systemic lupus erythematosus; TNF, tumor necrosis factor.

References

1. Schoenberger, S. P., Toes, R. E., van der Voort, E. I., Offringa, R., and Melief, C. J. (1998) T-cell help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. *Nature* **393**, 480–483
2. Korthauer, U., Graf, D., Mages, H. W., Briere, F., Padayachee, M., Malcolm, S., Ugazio, A. G., Notarangelo, L. D., Levinsky, R. J., and Kroccek, R. A. (1993) Defective expression of T-cell CD40 ligand causes X-linked immunodeficiency with hyper-IgM. *Nature* **361**, 539–541
3. Schonbeck, U., and Libby, P. (2001) The CD40/CD154 receptor/ligand dyad. *Cell Mol. Life Sci.* **58**, 4–43
4. Chatzigeorgiou, A., Lyberis, M., Chatzilymperis, G., Nezos, A., and Kamper, E. (2009) CD40/CD40L signaling and its implication in health and disease. *Biofactors* **35**, 474–483
5. Cha, J. K., Jeong, M. H., Bae, H. R., Han, J. Y., Jeong, S. J., Jin, H. J., Lim, Y. J., Kim, S. H., and Kim, J. W. (2000) Activated platelets induce secretion of interleukin-1 β , monocyte chemoattractant protein-1, and macrophage

- inflammatory protein-1 α and surface expression of intercellular adhesion molecule-1 on cultured endothelial cells. *J. Korean Med. Sci.* **15**, 273–278
6. Daub, K., Langer, H., Seizer, P., Stellos, K., May, A. E., Goyal, P., Bigalke, B., Schonberger, T., Geisler, T., Siegel-Axel, D., Oostendorp, R. A., Lindemann, S., and Gawaz, M. (2006) Platelets induce differentiation of human CD34⁺ progenitor cells into foam cells and endothelial cells. *FASEB J.* **20**, 2559–2561
 7. Henn, V., Slupsky, J. R., Grafe, M., Anagnostopoulos, I., Forster, R., Muller-Berghaus, G., and Kroczek, R. A. (1998) CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. *Nature* **391**, 591–594
 8. Crow, M. K., and Kirou, K. A. (2001) Regulation of CD40 ligand expression in systemic lupus erythematosus. *Curr. Opin. Rheumatol.* **13**, 361–369
 9. Delmas, Y., Viallard, J. F., Solanilla, A., Villeneuve, J., Pasquet, J. M., Belloc, F., Dubus, I., Dechanet-Merville, J., Merville, P., Blanco, P., Pellegrin, J. L., Nurden, A. T., Combe, C., and Ripoche, J. (2005) Activation of mesangial cells by platelets in systemic lupus erythematosus via a CD154-dependent induction of CD40. *Kidney Int.* **68**, 2068–2078
 10. Grewal, I. S., and Flavell, R. A. (1998) CD40 and CD154 in cell-mediated immunity. *Annu. Rev. Immunol.* **16**, 111–135
 11. Andre, P., Prasad, K. S., Denis, C. V., He, M., Papalia, J. M., Hynes, R. O., Phillips, D. R., and Wagner, D. D. (2002) CD40L stabilizes arterial thrombi by a β 3 integrin-dependent mechanism. *Nat. Med.* **8**, 247–252
 12. El Fakhry, Y., Alturaihi, H., Yacoub, D., Liu, L., Guo, W., Leveille, C., Jung, D., Khzam, L. B., Merhi, Y., Wilkins, J. A., Li, H., and Mourad, W. (2012) Functional interaction of CD154 protein with α 5 β 1 integrin is totally independent from its binding to α IIb β 3 integrin and CD40 molecules. *J. Biol. Chem.* **287**, 18055–18066
 13. Takada, Y. K., Yu, J., Shimoda, M., and Takada, Y. (2019) Integrin binding to the trimeric interface of CD40L plays a critical role in CD40/CD40L signaling. *J. Immunol.* **203**, 1383–1391
 14. Fujita, M., Takada, Y. K., and Takada, Y. (2014) The chemokine fractalkine can activate integrins without CX3CR1 through direct binding to a ligand-binding site distinct from the classical RGD-binding site. *PLoS One* **9**, e96372
 15. Fujita, M., Zhu, K., Fujita, C. K., Zhao, M., Lam, K. S., Kurth, M. J., Takada, Y. K., and Takada, Y. (2015) Proinflammatory secreted phospholipase A2 type IIA (sPLA-IIA) induces integrin activation through direct binding to a newly identified binding site (site 2) in integrins α v β 3, α 4 β 1, and α 5 β 1. *J. Biol. Chem.* **290**, 259–271
 16. Fujita, M., Davari, P., Takada, Y. K., and Takada, Y. (2018) Stromal cell-derived factor-1 (CXCL12) activates integrins by direct binding to an allosteric ligand-binding site (site 2) of integrins without CXCR4. *Biochem. J.* **475**, 723–732
 17. Pokharel, S. M., Shil, N. K., Gc, J. B., Colburn, Z. T., Tsai, S. Y., Segovia, J. A., Chang, T. H., Bandyopadhyay, S., Natesan, S., Jones, J. C. R., and Bose, S. (2019) Integrin activation by the lipid molecule 25-hydroxycholesterol induces a proinflammatory response. *Nat. Commun.* **10**, 1482
 18. Craig, D. B., and Dombkowski, A. A. (2013) Disulfide by Design 2.0: A web-based tool for disulfide engineering in proteins. *BMC Bioinformatics* **14**, 346
 19. Mori, S., Wu, C. Y., Yamaji, S., Saegusa, J., Shi, B., Ma, Z., Kuwabara, Y., Lam, K. S., Isseroff, R. R., Takada, Y. K., and Takada, Y. (2008) Direct binding of integrin α v β 3 to FGF1 plays a role in FGF1 signaling. *J. Biol. Chem.* **283**, 18066–18075
 20. Saegusa, J., Yamaji, S., Ieguchi, K., Wu, C. Y., Lam, K. S., Liu, F. T., Takada, Y. K., and Takada, Y. (2009) The direct binding of insulin-like growth factor-1 (IGF-1) to integrin α v β 3 is involved in IGF-1 signaling. *J. Biol. Chem.* **284**, 24106–24114
 21. Ieguchi, K., Fujita, M., Ma, Z., Davari, P., Taniguchi, Y., Sekiguchi, K., Wang, B., Takada, Y. K., and Takada, Y. (2010) Direct binding of the EGF-like domain of neuregulin-1 to integrins (α v β 3 and α 6 β 4) is involved in neuregulin-1/ErbB signaling. *J. Biol. Chem.* **285**, 31388–31398
 22. Fujita, M., Takada, Y. K., and Takada, Y. (2012) Integrins α v β 3 and α 4 β 1 act as coreceptors for fractalkine, and the integrin-binding defective mutant of fractalkine is an antagonist of CX3CR1. *J. Immunol.* **189**, 5809–5819
 23. Takada, Y. K., Yu, J., Fujita, M., Saegusa, J., Wu, C. Y., and Takada, Y. (2017) Direct binding to integrins and loss of disulfide linkage in interleukin-1 β (IL-1 β) are involved in the agonistic action of IL-1 β . *J. Biol. Chem.* **292**, 20067–20075
 24. Saegusa, J., Akakura, N., Wu, C. Y., Hoogland, C., Ma, Z., Lam, K. S., Liu, F. T., Takada, Y. K., and Takada, Y. (2008) Pro-inflammatory secretory phospholipase A2 type IIA binds to integrins α v β 3 and α 4 β 1 and induces proliferation of monocytic cells in an integrin-dependent manner. *J. Biol. Chem.* **283**, 26107–26115
 25. Shattil, S. J., Kim, C., and Ginsberg, M. H. (2010) The final steps of integrin activation: The end game. *Nat. Rev. Mol. Cell Biol.* **11**, 288–300
 26. Calderwood, D. A., Campbell, I. D., and Critchley, D. R. (2013) Talins and kindlins: Partners in integrin-mediated adhesion. *Nat. Rev. Mol. Cell Biol.* **14**, 503–517
 27. Bunch, T. A. (2010) Integrin α IIb β 3 activation in Chinese hamster ovary cells and platelets increases clustering rather than affinity. *J. Biol. Chem.* **285**, 1841–1849
 28. Schurpf, T., and Springer, T. A. (2011) Regulation of integrin affinity on cell surfaces. *EMBO J.* **30**, 4712–4727
 29. Takagi, J., Erickson, H. P., and Springer, T. A. (2001) C-terminal opening mimics 'inside-out' activation of integrin α 5 β 1. *Nat. Struct. Biol.* **8**, 412–416
 30. Yokoyama, K., Zhang, X. P., Medved, L., and Takada, Y. (1999) Specific binding of integrin α v β 3 to the fibrinogen γ and α E chain C-terminal domains. *Biochemistry* **38**, 5872–5877
 31. Berger, J., Hauber, J., Hauber, R., Geiger, R., and Cullen, B. R. (1988) Secreted placental alkaline phosphatase: A powerful new quantitative indicator of gene expression in eukaryotic cells. *Gene* **66**, 1–10