



Review

Mismatch of Glucose Allocation between Different Life Functions in the Transition Period of Dairy Cows

Jonas Habel * and Albert Sundrum

Department of Animal Nutrition and Animal Health, Faculty of Organic Agricultural Sciences, University of Kassel, Nordbahnhofstr. 1a, 37213 Witzenhausen, Germany; sundrum@uni-kassel.de

* Correspondence: jonas.habel@uni-kassel.de; Tel.: +49-5542-981529

Received: 12 May 2020; Accepted: 10 June 2020; Published: 13 June 2020



Simple Summary: The reasons for the development of diseases in the transition period of dairy cows are manifold and highly farm- and cow-specific. Nevertheless, links exist between the degree of negative energy balance (NEB) and disease susceptibility, which suggest a mutual adverse relationship between immune and productive functions. Glucose is the most essential fuel and precursor for both immune cells and mammary epithelial cells (MEC). While the delivery of glucose by the intermediary metabolism is not always able to keep up with whole-body demands, trade-offs between mammary and immune cells emerge. The prioritization of mammary supply during early lactation is a physiologic principle in all mammals. In contrast, tremendous increases in milk yield and the specific demand for glucose in high-yielding dairy cows resulting from decades of selection for milk production override the evolutionary principles of nutrient partitioning. Therefore, high-producing dairy cows face an increased risk of glucose shortages in their immune cells, particularly during early lactation.

Abstract: Immune cell functions such as phagocytosis and synthesis of immunometabolites, as well as immune cell survival, proliferation and differentiation, largely depend on an adequate availability of glucose by immune cells. During inflammation, the glucose demands of the immune system may increase to amounts similar to those required for high milk yields. Similar metabolic pathways are involved in the adaptation to both lactation and inflammation, including changes in the somatotropic axis and glucocorticoid response, as well as adipokine and cytokine release. They affect (i) cell growth, proliferation and activation, which determines the metabolic activity and thus the glucose demand of the respective cells; (ii) the overall availability of glucose through intake, mobilization and gluconeogenesis; and (iii) glucose uptake and utilization by different tissues. Metabolic adaptation to inflammation and milk synthesis is interconnected. An increased demand of one life function has an impact on the supply and utilization of glucose by competing life functions, including glucose receptor expression, blood flow and oxidation characteristics. In cows with high genetic merits for milk production, changes in the somatotropic axis affecting carbohydrate and lipid metabolism as well as immune functions are profound. The ability to cut down milk synthesis during periods when whole-body demand exceeds the supply is limited. Excessive mobilization and allocation of glucose to the mammary gland are likely to contribute considerably to peripartal immune dysfunction.

Keywords: glucose metabolism; immune dysfunction; production diseases; lactational physiology; trade-offs

1. Introduction

Animal welfare concerns, as well as the economic implications of the high prevalence of production diseases [1–3] and increasing rates of involuntary culling in dairy farming [4], emphasize the need

Animals 2020, 10, 1028 2 of 21

to address possible conflicts of aims between these impacts and the level of milk production. Many scientific investigations are based on the assumption that individual genomic and/or metabolomic differences are able to explain why some cows are both high producing and healthy while others fail to cope [5–8]. These approaches try to identify cows that are more efficient in digestion, absorption, synthesis and mammary utilization of nutrients. However, they disregard the limitations in the capacity to deal with deficiencies in a highly heterogeneous and dynamically changing environment that is elusive to deterministic approaches. They disregard that immune functionality is fundamental to ensure health, longevity and productivity of dairy cows, as it is not only essential for pathogen elimination but is part of the coordinated reaction of the organism to all kinds of stressors.

After parturition, high-producing dairy cows generally enter a negative energy balance (NEB), because their level of dry matter intake (DMI) does not meet the demands imposed by the onset of milk production [9]. Consequently, they mobilize body tissue to overcome this shortage. Excessive mobilization can lead to a hypercatabolic response described as metabolic stress, associated with the occurrence of subclinical and clinical diseases [10]. The overall energy budget of an organism includes various energy sources, metabolic pathways and interactions between subsystems of nutrient trafficking that make it difficult to evaluate the consequences of sustained overall NEB on metabolic disorders and health. In fact, plasma concentrations of single metabolites vary substantially between individual cows with similar status of NEB in early lactation [11–14]. Thus, it has been emphasized that we must move "from joules to moles of molecules or groups of molecules" to advance animal nutrition concepts [15].

In dairy cows, the amount of glucose required to fuel milk production outreaches by far energy expenditures of other life functions like reproduction or maintenance [16]. Besides being a precursor for the synthesis of lactose, which is the osmotic regulator of milk volume [17,18], glucose-derived carbon is also found in milk fat and protein [19,20]. Moreover, reduction of nicotinamide adenine dinucleotide phosphate (NADP+) through pentose phosphate pathway as well as the production of ATP, which are required for the synthetic processes depend on the availability of glucose. During peak lactation, mammary epithelial cells (MEC) are able to retrieve up to 2.7 kg of glucose per day from the plasma pool at a milk level of 40 kg [16]. On the other hand, cells of the innate and adaptive immune system rely largely on the uptake of glucose and the storage of glycogen, because glucose supports proliferation, survival and differentiation as well as essential functions like phagocytosis and reactive oxygen species (ROS) production [21]. Recent data about dairy cows exposed to lipopolysaccharide (LPS) challenges showed that a fully activated immune system needs 2.5 to 3.1 kg of glucose per day [22]. In other words, immunoactivities in dairy cows can amount to degrees of glucose demands similar to those required for high milk yields. In contrast to monogastric species, ruminants cover their glucose demand almost exclusively through hepatic gluconeogenesis, which seems limited to about 3 kg of glucose at a milk yield level of ~40 kg/day [23]. These considerations and the quantities at hand clearly indicate a competitive situation between milk production and immune defense and give rise to the question of how allocation of nutrients between different tissues and life functions is regulated when essential nutrients become scarce.

2. Resource Allocation between Maintenance and Productive Life Functions in Early Lactating High-Producing Dairy Cows

2.1. Resource Allocation Theory

According to the resource allocation theory [24], resources including energy and essential nutrients have to be partitioned between all life functions. Common differentiations of life functions that an animal has to fuel include productive processes such as growth, gestation and lactation as well as maintenance functions. However, definitions of maintenance and the requirements of regulatory systems are essentially conceptual, of a qualitative nature and are also imprecise and contradictory throughout the scientific literature [25,26]. However, activities like thermoregulation and immune function are known to impose high demands [27].

Animals **2020**, 10, 1028 3 of 21

As for the relationship between productive and other life functions in dairy cows it was hypothesized that cows with high genetic merit for milk production cover their demand by an increase in feed intake, while cows with low genetic merit—if they do consume more feed—accumulate body reserves [28]. Consequently, maintenance requirements would be unaffected by milk yield, and differences in milk yield could be due to differences in the efficiency of energy and nutrient utilization—a phenomenon that was described as "dilution of maintenance" [28]. Although correlations between yield and intake ranging from 0.46 to 0.65 [29] indicate the strong relationship between these variables, they reveal as well that the increase in feed intake does not keep pace with the increased demand imposed by an increased number of lactocytes in the mammary gland. Accordingly, increases in body weight associated with selection for milk production have been shown to increase maintenance requirements as well [30]. While energy expenditures by visceral organs represent 0.4-fold of the maintenance requirements for nonproductive adults, they increase to 1.2-fold for lactating ruminants [31]. Moreover, increased energy and nutrient demands for milk synthesis may also lead to time constraints since eating and rumination time has to be traded off against all other activities. In fact, there is little sign that high genetic merit cows digest feed more efficiently than low genetic merit cows. It has been shown decades ago that digestive efficiency decreases as feed intake increases [32]. Accordingly, the nonlinear character of the relationship between milk yield and body weight suggest an optimum body weight beyond which the feed-efficiency decreases in specialized breeds [33]. Loncke et al. (2020) recently showed a similar pattern for the efficiency of hepatic glucose synthesis (conversion rate of precursors to glucose), which decreases at high levels of precursor supply [34]. As a result, high-producing dairy cows show increased rates of energy mobilization from body tissues to bridge the gap between the supply of energy from feedstuffs and the energy needed to support milk production along with all other energy-demanding life functions [35].

2.2. Homeorhetic and Allostatic Control of Nutrient Partitioning

Because most organs and tissues lack autonomy to control their nutrient access, the organism needs prioritization rules that coordinate nutrient partitioning between different life functions in all situations where demand exceeds supply. In this regard, different concepts of regulation have been established and adapted to dairy cow physiology. In the concept of homeorhesis, nutrient partitioning is described as a function of biological needs that alternates cyclically between storing and mobilization of energy from body tissues and the associated prioritization of reproductive and productive functions [9]. Following this approach, it is a fundamental biological principle that after parturition, mammalian organisms prioritize mammary tissues to provide an adequate supply to the neonate. Dairy production is based on, and takes advantage of, this principle through a performance-oriented selection of animals.

However, environmental and nutritional stressors associated with varying quality and quantity of feedstuffs, social stress, climate variability and extremes, poor hygiene, technical failure, etc. also affect supply and demand of nutrients independent of the physiological state [27,36]. In this regard, it has been emphasized that systematically reviewing the literature from epidemiological studies is unlikely to support understanding of the effects of metabolic imbalances of each cow in her specific genetic and environmental circumstance due to the fact that intricate biological correlations—besides within- and between-herd confounding effects—exist [37].

The concept of allostasis goes beyond the concepts of homeostasis and homeorhesis by assuming dynamic set points emerging from the integration of both the requirements of different tissues at different physiological states and current nutritional, social and housing conditions [38]. In dairy cows, various genotype x environment interactions have been described, including a reduced ability to adapt to unfavorable conditions (plasticity) in cows with high genetic merit for milk production [39]. This suggests that the process of adaptation to such conditions imposes a certain demand for energy itself, which has been described qualitatively as the "allostatic load" of the animal [40]. If the allostatic load becomes too big (allostatic overload), the capacity to cope with additional stressors is reduced and the animal enters a pre-pathological state [41]. However, by considering maintenance functions as costs,

Animals 2020, 10, 1028 4 of 21

they have to be traded off against productivity goals. Following this approach, resource availability for immune cells provided through a balanced resource allocation is a prerequisite for an organism to be able to cope with internal and external stressors.

3. Glucose Metabolism to Fuel Milk Synthesis and Immune Functions in Dairy Cows

Proinflammatory signaling promotes similar metabolic adaptations, as does the lactational "reprogramming" with the aim of maximizing glucose availability to the respective cell types (Figure 1). These changes are mediated by a network of hormones and affect both mobilization and allocation, including specific regulation of blood flow and receptor expression patterns in peripheral, mammary and immune tissues. In the following chapters, the metabolic processes associated with the onset of lactation (Section 3.1) and those occurring during inflammation (Section 3.2) are discussed separately. Subsequently, trade-offs for glucose between MEC and immune cells in dairy cows are evaluated in Section 4.

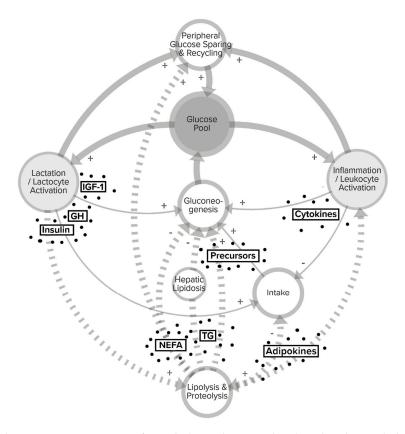


Figure 1. Schematic representation of metabolic pathways related to the glucose balance of dairy cows during lactation and inflammation. Milk synthesis and immune defense rely on an adequate supply with glucose, as it is an important energy source and precursor for lactocytes and leukocytes. To increase overall glucose availability to the respective cell type, inflammatory signals (cytokines) as well as peripartal fluctuation of hormones associated with the somatotropic axis such as growth hormone (GH), insulin and insulin-like growth factor 1 (IGF-1) enhance (+) the rate of gluconeogenesis and affect the level of intake (solid lines, thin), and increase the mobilization of body reserves (dashed arrows). Lipolysis and proteolysis provide endogenous glucose precursors such as alanin and glycerol as well as alternative energy sources like non-esterified fatty acids (NEFA) that help spare glucose in peripheral tissues, where insulin sensitivity is reduced. Because glucose uptake is non-dependent on insulin in both leukocytes and lactocytes, trade-offs for glucose allocation (solid arrows, bold) may arise in situations where inflammation and lactation impose high demands. Limitations may also arise from negative effects (-) of adipokines and cytokines on the hypothalamic regulation of intake and from hepatic accumulation of triglycerides (TG) and NEFA when lipolysis is excessive.

Animals **2020**, 10, 1028 5 of 21

3.1. Adaptation to Lactation

Morphological changes required for mammary growth, morphogenesis, and milk synthesis are created during gestation, with ductal elongation and lobulo-alveolar development being mediated through high plasma concentration of prolactin, growth hormone (GH) and gonadotropic steroids progesterone and estrogen before parturition [42,43]. Subsequently, high concentrations of GH are known to stimulate galactopoiesis, while prolactin and Insulin-like growth factor 1 (IGF-1) are involved in establishing and maintaining milk synthesis through their effect on epithelial cell differentiation and survival [44,45].

To meet the sudden increase in demand during early lactation, further alterations in the endocrine setup support dairy cows to metabolically adapt to lactation. Besides the above-mentioned hormones, insulin, thyroid hormones, glucocorticoids and the gonadotropic axis are main effectors of the new catabolic physiology and plasma concentrations of these hormones typically fluctuate at the transition from a pregnant to a lactating physiology [46]. Especially changes in the somatotropic axis; i.e., decreased pancreatic secretion of insulin and reduced GH-receptor (GHR) expression in the liver [47] are thought to be major levers of the new catabolic physiology [48–50]. Subsequently, hepatic GH resistance and hypoinsulinemia mitigate stimulating effects on hepatic IGF-1 production [51]—a condition that has been described as the "uncoupling of the somatotropic axis" [48].

Insulin signaling is particularly essential for a successful adaptation to lactation by affecting the rate of lipolysis, the rate of uptake and transport of glucose and fatty acids to different tissues, and the expression of key enzymes at the metabolic crossroads of glucose and fatty acid metabolism [52,53]. A main effect of peripartal hypoinsulinemia is the reduction in insulin's antilipolytic properties, which facilitates the mobilization of nutrients from body reserves by increasing the rates of lipolysis and proteolysis [54]. These processes are accompanied by increased rates of gluconeogenesis, reflected by an increased mRNA amount of the important rate-limiting enzymes pyruvate carboxylase (PC) and phosphoenolpyruvatecarboxykinase (PEPCK) postpartum [55]. Precursors for gluconeogenesis include rumen-derived volatile fatty acids, mainly propionate and, to a lesser extent, circulating C3-bodies like glycerol, alanine and lactate from intermediary metabolism [23]. Increased levels of circulating non-esterified fatty acids (NEFA) resulting from adipose tissue remodeling are taken up proportionally to their plasma level by the liver. Together with a simultaneous lack of oxaloacetate, which is highly used for gluconeogenesis, increased NEFA lead to an accumulation of acetyl-CoA in the liver. Subsequently, hepatocytes are forced to switch acetyl-CoA utilization from complete (Krebs cycle) towards incomplete oxidation (ketogenesis) and/or to re-esterification with subsequent storage of triglyceride in the liver [56]. Besides negative effects of hepatic TG accumulation on general hepatic function, increased levels of β -hydroxybutyrate (BHB) were shown to impair gluconeogenic capacity [57,58]. In cultured bovine hepatocytes, increasing levels of NEFA gradually decrease mRNA levels and catalytic activity of PC and PEPCK [59]. Thus, glucose balance is challenged severely when lipolysis becomes excessive. Although the usefulness of plasma glucose as an indicator of a cow's metabolic status is particularly contentious due to the tight regulation of glucose homeostasis [60], hypoglycemia is associated with the onset of ketosis, higher first test-day milk production and milk production at 100 days in milk [61].

Moreover, allocation patterns that regulate the flow of nutrients between different tissues within the organism change according to the new dominant physiological state of lactation. Again, these changes are related to the 'uncoupled' somatotropic axis and in particular, to the phenomenon of reduced insulin sensitivity in peripheral tissues of postpartum dairy cows [62–65]. Because glucose receptors prevailing in the mammary gland are mostly non-dependent on insulin while muscle and adipose cells are highly insulin-responsive cell types [66], reduced peripheral insulin sensitivity favors the glucose supply of lactocytes. Simultaneously, mRNA encoding insulin-independent glucose transporter (GLUT) with the highest affinity to glucose (GLUT1) increases strongly in mammary tissues at the onset of milk synthesis [67]. As lactation advances, the mammary gland becomes more insulin-sensitive and insulin-dependent while glucose uptake via GLUT4 increases [68]. In contrast,

Animals **2020**, 10, 1028 6 of 21

GLUT1 decreases about 6-fold in mRNA and protein levels in adipose tissue of early lactating cows compared with dried off or late lactating cows [67]. However, the extraction of great amounts of glucose from circulation is promoted primarily through a greater blood flow to the mammary gland, which was found to be stimulated by the characteristic endocrine regulation of lactation [69,70]. In particular, thyroxine is thought to enhance mammary nutrient extraction by increased heart rate and subsequent increases in blood flow [71]. This was identified decades ago as a main determinant of quantitative udder metabolism [16,72]. In more recent studies, blood sampling techniques comparing glucose concentrations from the jugular and mammary vein showed a lower jugular/mammary quotient for glucose concentration in dry and low-yielding cows, while revealing significantly higher levels in high yielding cows [73].

In peripheral tissues, modest reductions [74,75] or no significant changes [67] in the expression of insulin-dependent GLUT 4 in peripheral tissues have been reported in early lactating dairy cows. However, these tissues are aligned to save glucose during that period by shifting their glucose metabolism from complete oxidation towards lactate production. Accordingly, irreversible losses of glucose excluding the loss in milk lactose decreases significantly in the first days after parturition [76]. Together with alanine and glycerol derived from muscle resp. adipose tissue, lactate can recirculate to the liver, where it is supposed to have a higher proportional contribution to gluconeogenesis during early lactation [23].

In summary, a complex endocrine network develops to increase glucose availability to the mammary gland. If precursor supply or hepatic synthetic capacity are inadequate, the sudden increase in mammary demand for glucose at the onset of lactation is the main driver of the hypermetabolic reaction that affects a variety of metabolic pathways, tissues and organs within the organism.

3.2. Adaptation to Inflammation

Immune cell activity and inflammation are not only essential for pathogen elimination but are part of the coordinated reaction of the organism to all kinds of stressors, including infective and non-infective, metabolic and environmental stressors. After parturition, dairy cows experience an inflammatory-like status, which is systemically linked to the inherent stress of parturition, social and nutritional changes and the endotoxin-releasing processes of ruminal adaptation and uterine tissue reorganization [77,78]. The response is characterized by a marked increase in plasma concentration of positive acute phase proteins [79]. Their plasma level has been associated with the occurrence of retained placenta, other diseases and impaired reproductive and productive performance during early lactation [80,81]. However, the necessity of some degree of "physiological inflammation" is illustrated by the action of anti-inflammatory drugs that inhibit the synthesis of prostaglandins required to expel the placenta [82]. Following administration of an anti-inflammatory drug after calving, dairy cows have an increased risk of retained placenta (2.5-fold) and metritis (1.5-fold) [83].

Whatever the origin of inflammation, the accumulation of proinflammatory processes implies a supply with energy that adequately meets the requirements of immune response. Qualitatively, the costs of immune activation include (1) a general elevation of metabolic rates due to a rise in body temperature, (2) reduced nutrient availability following anorexic effects of proinflammatory signaling, (3) the precursors and energy needed to fuel the synthesis of acute-phase proteins and immunoglobulins, (4) altered priorities for nutrient utilization in other tissues, (5) the costs associated with the repair of damaged tissues and (6) increased turnover rates of the leukocyte pool [27]. Although an almost infinite number of possible combinations between metabolic and environmental stressors make it impossible to estimate the current degree and duration of inflammation and immunoactivation and to determine the energy demand of immune cells, some quantification has been performed. For instance, it has been shown that the demand for oxygen, glucose and glutamine increases two-to three-fold during lymphocyte activation [84]. By examining the effect of an infection with nematode larvae on the energy requirement of merino sheep, it was estimated that infection increased the requirement for metabolizable energy by 28% [85]. Even more impressively, Kvidera and colleagues

Animals 2020, 10, 1028 7 of 21

combined an intravenous LPS challenge, a euglycemic clamp and measurement of milk yield reduction in cows of parity 2 or 3 that were at 69 ± 7 days in milk to calculate the demand of a fully activated immune system. The authors estimated that dairy cows may require up to 3.1 kg of glucose per day to mount an acute inflammatory response (Figure 2) [22].

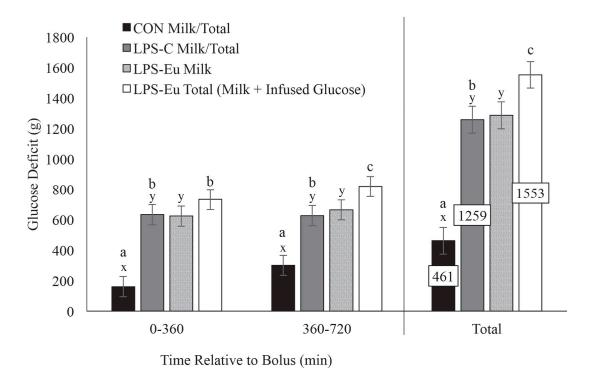


Figure 2. Milk or total glucose deficit from zero to 360, 360 to 720, and accumulated over 720 min in cows administered a bolus of saline (CON), lipopolysaccharide (LPS-C), or lipopolysaccharide accompanied with a euglycemic clamp (LPS-Eu). Different letters (x,y) represent differences between milk glucose deficits ($p \le 0.05$). Different letters (a–c) represent differences between total glucose deficits ($p \le 0.05$); total glucose deficit = milk glucose deficit in CON and LPS-C cows; total deficit = milk glucose deficit + infused glucose in LPS-Eu cows). Results are expressed as least square means \pm standard error of means. Reprinted from Kvidera et al. (2017), Copyright (2017) with permission from Elsevier.

In fact, cells of the innate and adaptive immunity rely largely on the uptake of glucose and the storage of glycogen, because glucose supports proliferation, survival and differentiation as well as essential functions like phagocytosis and ROS production [21]. Moreover, an activation of apoptotic pathways in response to limited glucose uptake in cultured hematopoietic cells was reported [86]. In dairy cows, reduced glycogen concentrations in circulating neutrophils at calving indicate a depletion of glucose depots during this challenging period and are associated with the occurrence of subclinical endometritis and metritis [87]. Although immune cells are able to use alternative energy sources like glutamine and ketone bodies to some extent [88–90], the importance of glucose as their main fuel was corroborated by Noleto et al., who found that supplying increasing amounts of glutamine in the absence of glucose was not sufficient to raise the inflammatory response to LPS in endometrial monocytes and macrophages of dairy cows, whereas supplying more glucose was able to increase inflammation in the absence of glutamine [91].

Not surprisingly, leukocytes trigger a number of metabolic pathways that increase the glucose supply to these cells while reducing consumption of glucose by other tissues. First references describing the link between inflammation and insulins actions date far back [92]. By now it is clear that the interplay between proinflammatory and insulin signaling is common to all the mammals [93]. In dairy cows, the effect of continuous and increasing LPS-infusion on whole-body insulin-resistance has recently been demonstrated [94]. T-cells were shown to shift glucose transporter expression from insulin-dependent

Animals 2020, 10, 1028 8 of 21

GLUT4 towards GLUT1 and GLUT3, which are non-dependent on insulin, to maintain glucose disposal during activation [95–97]. Inflammatory pathways also promote the transcription of gluconeogenic genes via toll-like receptor 4 (TLR-4) [98]. Macrophages and neutrophils undergo a metabolic switch from oxidative phosphorylation towards glycolysis during activation, thereby increasing their demand for glucose as well as their lactate production [99]. Metabolic reactions to the alterations induced by proinflammatory cytokines further encompass increased rates of lipolysis and proteolysis, that could provide energy for leukocyte functions as well as substrates for gluconeogenesis [100,101]. However, the inflammation-mediated metabolic reprogramming appears very similar to the reprogramming mediated by lactation, both aiming at a maximum supply of glucose for the respective cell functions. On a systemic level, this includes increased rates of gluconeogenesis and reduced glucose consumption in peripheral tissues.

4. Trade-Offs for Glucose between Lactocytes and Leukocytes

All mammals favor the supply of nutrients to the mammary gland during early lactation. In contrast, the increases in milk yield and the specific demand for glucose required for high milk yields override evolutionary principles of nutrient partitioning [102]. Because nutritional supply is limited through various factors, e.g., percentage of concentrate in the diet, time to eat, turnover rates in the rumen as well as the synthetic capacity of the liver during this period, most high-producing dairy cows experience a period of glucose shortage. In such situations, trade-offs for glucose between MEC and immune cells are unavoidable as they both rely on this essential substrate. Therefore, a special focus on the allocation dynamics of glucose is necessary during periods when both lactation and inflammation impose high demands.

4.1. Peripartal Immune Dysfunction

The phenomenon of reduced immune cell competence is well established in peripartal dairy cows. It is broadly characterized by a dysfunction of PMN, macrophages and lymphocytes, including an impairment of viability, survival, phagocytosis and respiratory burst capacity [103]. Studying gene expression profiles in the bovine mammary gland during stage I and II of lactogenesis, it was found that most of the genes associated with immune response were downregulated at the end of gestation [104]. This is in line with the interpretation of Goff and Horst, who suggested that neutrophil phagocytosis and lymphocyte proliferation begin to be impaired around three weeks before parturition [105]. Moreover, significant changes in lymphocyte subsets occur. Overall number and proliferation of circulating lymphocytes are reduced, while mammary cell number and proliferation peaks around calving [43,106–108]. Accordingly, altered immune functions during the dry period are associated with the development of metabolic disease during early lactation [106]. Around parturition, elevated levels of glucocorticoids and decreased plasma levels of oestrogens and progesterone also affect immune response through altered MHC-expression, cytokine production, diapedesis capacity and viability of immune cells [109–111]. In summary, the mammary gland prepares for lactation not only by improving functionality but also by suppression of competitive functions, allowing more resources to be used for milk synthesis [104].

4.2. Metabolic Stress and the Immune System

The aforementioned relationships suggest that the substantial but transient suppression of immune functions before parturition is related to the physiological adaptation to lactation. However, not only cell number and proliferation, but also functionality of immune cells is impaired strongest when MEC start the abundant synthetic activity of lactogenesis as was demonstrated by the transient loss of expression of vascular factors and antimicrobial chemokines [104,112–114]. Accordingly, mastectomized cows had a shorter and less marked immune suppression, including less impairment of oxidative burst capacity and faster recovery of myeloperoxidase activity in neutrophils at calving compared to non-mastectomized cows [115,116]. This indicates that some immunosuppressive effects

Animals 2020, 10, 1028 9 of 21

may be independent from the endocrine changes associated with parturition but related directly to the capacity to synthesize milk. In fact, various effects of severe NEB on immunosuppression have been published and many of them are related to the effects of adipose-tissue remodeling on key molecules involved in glucose and lipid metabolism [117]. For instance, high plasma levels of BHB are negatively correlated with DNA replication and repair in leukocytes [118]. Plasma concentration of NEFA correlate with increased hepatic expression of mRNA encoding proinflammatory cytokines and acute-phase proteins [119]. Increased hepatic uptake of NEFA may also result in increased production of reactive oxygen species (ROS) that carry out important tasks of immune defense by facilitating the destruction of pathogens and enhancing the proinflammatory cascade at physiological plasma concentrations [120]. They can affect the integrity of immune cells, which are very susceptible to peroxidation due to high concentrations of polyunsaturated fatty acids in their membranes. Additionally, ROS generated during inflammation have been proposed to play a role in mediating insulin resistance [121]. If NEFA mobilization and ROS production is excessive, host tissues may fail to mitigate the negative effects of ROS by activation of antioxidant pathways, resulting in severe tissue damage [122]. Leukocyte function is also affected by the shift in fatty acid profile resulting from lipomobilization [123]. Altered concentrations of adipokines postpartum mitigate stimulating effects on chemotaxis and phagocytosis of neutrophils, proliferation of native T-cells and the secretion of cytokines as well as anti-inflammatory effects [124,125], presumably via activation of TLR-4 and nuclear factor kappa-B (NFkB) [74,126].

Accordingly, cows with severe NEB have a reduced ability to clear uterine infection postpartum. The active uterine inflammatory response in these cows was associated with impaired local insulin-receptor signaling [127]. In the mammary gland of lactating dairy cows subjected to a dietary-induced NEB, expression of genes related to proinflammatory signaling via NFkB (AKT1, IRAK1, MAPK9 and TRAF6), IL-8 (e.g., CXCR1/R2) and chemokine signaling (e.g., SOCS2) were downregulated [128].

Nevertheless, experimentally induced negative energy balance in advanced lactation was repeatedly shown to be unable to cause alterations of inflammation and immune cell function that are as severe as those occurring during early lactation [129–131]. With regard to the importance of glucose for immune cells, a possible mechanism associated with different reactions to similar NEB could be an increased glucose availability during late lactation, as it was demonstrated that late-lactation induced NEB evoked less severe proportional decreases in plasma glucose concentrations, compared to early-lactation NEB [132,133].

4.3. Competition for Glucose between MEC and Immune Cells

As addressed previously, the usefulness of plasma glucose as an indicator of a cow's metabolic status is unsure due to the tight regulation of glucose homeostasis [60]. However, Graber and colleagues differentiated metabolically robust or vulnerable cows based on the occurrence of various metabolic and (re)productive disorders in previous lactations and identified plasma glucose as the only variable explaining the differences between those groups at both time points investigated (3 weeks before and 4 weeks after parturition) [12]. In another study, plasma concentrations of glucose and insulin during lactation were found to be the single most important predictors related to the development of disease, explaining 36% of the between-cow variability in energy-corrected milk [14].

Generally, dietary energy supply affects glucose oxidation and transport in leukocytes in ruminants [134,135] and provide hints regarding the special competition for this essential metabolite. Inversely, elevated plasma concentrations of the acute phase protein haptoglobin are associated with remarkable decreases in milk yield [81,136]. Anti-inflammatory treatments substantially increase lactational milk yield [137,138]. This demonstrates that inflammation has some kind of regulatory potential of on mammary glucose extraction. Still, the question 'how nutrient partitioning is regulated when resources become scarce' remains. In this regard, it was speculated that decreases in monocyte GLUT1 protein and mRNA expression after calving are due to lactogenesis [135]. In fact, Eger et al. demonstrated a direct negative correlation between lactose yield and overall expression of GLUT1

Animals 2020, 10, 1028 10 of 21

and GLUT3 as well as a decrease in GLUT3/GLUT1 ratio of monocytes with increasing lactose yield (Figure 3A–C) [96]. On the other hand, downregulation of some GLUT isoforms in the mammary gland was observed following LPS-induced mastitis during mid lactation [139]. However, mRNA abundance of mammary GLUT1 transporter, which is the most important one for lactose synthesis [140], does not decrease in cows submitted to a hyperinsulinemic–hypoglycemic clamp, not even when these cows were submitted to an additional intramammary LPS challenge [139]. As described above (Section 3.1), mammary extraction of glucose from the plasma pool is likely to not be limited by GLUT expression of these cells and the plasma concentration of glucose but rather depends on the rate of local blood flow resulting from the metabolic activity of the gland. In contrast to mammary epithelial cells, circulating immune cells rely on the rate of GLUT expression and increases in types of GLUT that are insulin-independent to cover their glucose demand.

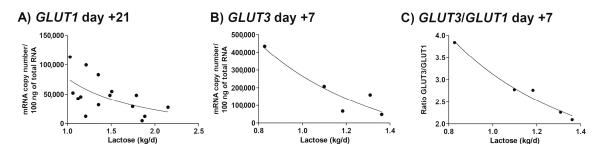


Figure 3. Peripartal monocyte glucose transporter expression is correlated with lactose production. glucose transporter 1 (GLUT1) (**A**) and GLUT3 (**B**) mRNA expression as well as the GLUT3/GLUT1 ratio (**C**) at d + 7 and d + 21 relative to parturition were correlated with milk production data of wk 1 (n = 5) and wk 3 (n = 15) of lactation, respectively, using Pearson (GLUT1 and GLUT3/GLUT1) or Spearman (GLUT3) correlation. Significant correlations with lactose production (at least p < 0.05, r > -0.50) were followed by nonlinear regression analysis (inverse model: Y = B0 + B1/X). (**A**) GLUT1 d + 21 (R² = 0.314, B0 = -30,557, B1 = 107,794), (**B**) GLUT3 d + 7 (R² = 0.871, B0 = -495,507, B1 = 761,265), and (**C**) GLUT3/GLUT1 ratio d + 7 (R² = 0.975, B0 = -0.432, B1 = 3.549). Reprinted from Eger et al. (2016), Copyright (2016) with permission from Elsevier.

Decreases in lactose yield as well as decreases in the mRNA-abundance of the ALA-subunit of lactose synthase were reported in hypoglycemic cows [139,141]. Similarly, reduced lactose content in milk following intramammary infection was reported [142,143] and could be a mechanism to save glucose for immune functions or/and to reduce substrate for bacterial growth during infection. On the other hand, Kreipe et al. showed that fat and protein percentages increased in hypoglycemic cows while energy-corrected milk did not differ significantly between hypoglycemic and control animals [141]. Thus, the extraction of glucose by the mammary gland might be unchanged during hypoglycemia, whereas glucose partitioning within the mammary gland is shifted from lactose synthesis towards glycolysis and pentose phosphate pathway to support protein and fat synthesis as was detected in bovine MEC exposed to various levels of glucose [144].

In fact, high-producing dairy cows were shown to be unable to reduce milk synthesis during early lactation in particular, while being able to reduce milk synthesis during induced energy deficiency at 100 days in milk, even if induced NEB was more severe compared to early-lactation NEB [132]. Accordingly, milk yield reductions following infusion of 100 μ g LPS were found to be more pronounced in late lactation compared to early lactation [145,146]. Milk production of cows challenged with intramammary infusion of 30 cfu [147], 1×10^4 cfu of diluted E. coli per quarter or 1000 μ g LPS [148] decreased to low levels. In contrast, milk yield was unaffected by chronic and exponentially increasing intravenous infusions of LPS (0.017–0.148 μ g/kg of body weight per hour from day 1 to 7) in a recently conducted study [149]. Daily subcutaneous injection of 3 μ g/kg body weight of bovine tumor necrosis factor- α during the first week of lactation decreased milk yield only slightly (33.7 to 28.4 kg at highest

dose) [149]. Osmotic TNF α pumps releasing 14 μ g/kg body weight over 7 days implanted in adipose tissue in late lactation cows had no effect on milk yield [150].

Further evidence for an antagonistic, yet dysbalanced relationship between metabolic pathways involved in adaptation to lactation and adaptation to inflammation is derived from studies examining the effect of the characteristic endocrine alterations required for high milk yields. Compared to low or medium genetic merit cows, high genetic merit cows show lower plasma concentrations of glucose, insulin and IGF-1, as well as higher plasma concentrations of GH [151,152], while insulin resistance is increased [63,153–155]. As described above, hypoinsulinemia favors glucose uptake in both immune cells and MEC because these cells are not dependent on insulin whereas glucose uptake to insulin-dependent cells like adipose and muscle cells is reduced [67]. However, hypoinsulinemia also mitigates stimulating effects of insulin on the rate of glucose utilization and phagocytosis in immune cells [156,157]. Moreover, increased GH-resistance is associated to selection for milk production and might contribute to the dysbalanced allocation of resources between MEC and immune cells in dairy cows. While GH exerts its mammogenic and galactopoietic effects directly in the mammary gland, either through GHR or through mammary IGF-1 production [158,159], many of the immune-stimulating effects attributed to GH are mediated indirectly through induction of hepatic IGF-1 production [160]. However, IGF-1 production in the liver is blunted through hepatic GH-resistance during early lactation [47]. Interestingly, it was shown that different breeds selected for milk production (Holstein-Friesian and Guernsey) showed similar decreases in GHR1A mRNA expression [161], whereas a comparison between Holstein-Friesian and beef cattle revealed decreases in the expression of GHR1A in dairy cows only [162].

5. Management of High-Producing Dairy Cows that Risk Glucose Shortage

The management of high-producing dairy cows should aim for a maximal reduction in metabolic and environmental stress to reduce the energy demand of regulatory systems. Although the specific demand of an activated immune system is difficult to assess, tendencies may be estimated from plasma levels of inflammatory markers, as it has been recently suggested by Trevisi and Minuti [163]. Moreover, the amount of residual glucose left for life functions other than milk synthesis may be estimated by consideration of the glucose demand of quantifiable processes like milk synthesis on one hand and the amount of glucose supply from precursors (derived from feedstuffs and body tissue mobilization) and the hepatic gluconeogenic potential on the other.

Furthermore, dry off feeding and heifer management should be optimized, as it was repeatedly shown that nutrition during these life stages affects availability and allocation of nutrients during early lactation [164]. For instance, overfeeding cows by 50% of predicted requirements decreased postpartum plasma glucose and insulin while increasing glucagon, BHB, and NEFA concentrations after calving compared with cows fed a balanced energy diet during the dry period [165]. During lactation, feeding should be more adapted to meet the need of individual cows in their specific physiological and environmental condition. In particular, supply with glucogenic precursors should be optimized. Although Lucy et al. demonstrated the key role of glucose by showing that infusions of substantial daily doses of glucose (8500 to 1500 g/day) into early postpartum cows were able to completely reverse the hypercatabolic reaction (significant increases in blood concentrations of insulin and IGF-1 along with significant decreases in the concentrations of NEFA and BHB) [166], feeding glucogenic diets is unlikely to significantly reverse lactational energy partitioning, although controversial results can be found in the literature [167–169]. Certainly, nutritional interventions are limited through, e.g., careful use of grain in the diet, time to eat, rumen volume and liver function. There are reasonable doubts whether dairy feeding regimes can further optimize the supply with precursors and the potential of gluconeogenesis and thus increase total glucose availability. For the sake of animal health and welfare and the economic implications of production diseases, dairy farmers should consider a modest but precise reduction on the other side of the equation, i.e., apply management measures that decrease glucose output via milk during periods when dairy cows are challenged simultaneously by both high

Animals 2020, 10, 1028 12 of 21

yields and infectious or non-infectious stressors. Possible management tools include a reduced milking frequency at the onset of lactation [170–172]. In fact, it has been demonstrated that reduced milking frequency reduces both milk yield and inflammation simultaneously [173]. Moreover, instead of implementing general strategies for a very heterogeneous target group, dairy cows should be assessed individually according to their status of NEB. Dairy cows with a high NEB should be allocated to a risk group and dealt with appropriately. In the long term, breeding should be redefined to include increased selection for persistence, lifetime performance and longevity while reducing emphasis on selection for milk yield and early-lactation performance in particular.

6. Conclusions

NEB is commonly thought to identify metabolically instable situations associated with increased risk of disease in dairy cows. However, overall energy balance disregards the reliance of immune cells on glucose as their essential metabolite and synthetic precursor. Although both proinflammatory signaling and lactational reprogramming promote several similar metabolic pathways with the aim of maximizing glucose availability to the respective cell types (lactocytes or leukocytes), adaptation to lactation clearly shifts nutrient partitioning to the favor of the mammary gland. Adaptations are mediated primarily by hormones of the somatotropic axis and affect both mobilization and allocation, including specific regulation of blood flow and receptor expression patterns in peripheral, mammary and immune tissues. Additionally, decades of performance-oriented selection of dairy cows enhanced these patterns substantially by increasing the amount of mammary epithelial cells as well as the metabolic and endocrine setup required to support the demand of these cells. Due to the central role of glucose for milk production and immune cell function, glucose balance is especially submitted to competitive allocation dynamics and is at risk of being overstressed in the early postpartum, high-producing dairy cow, as indicated by reduced responsiveness of lactose synthesis and milk yield to energy or glucose restriction, or other stressors. Therefore, we hypothesize that the uncoupling of the somatotropic axis in cows with high genetic merit for milk production implies, at least in part, an uncoupling of the mammary gland from life function trade-offs. To address possible impacts of glucose shortage on the immune defense, research should focus on the dynamics of glucose supply and demand of immune cells in high producing dairy cows during different periods of lactation.

Author Contributions: Conceptualization, J.H. and A.S.; investigation, J.H.; writing—original draft preparation, J.H.; writing—review and editing, A.S.; visualization, J.H.; All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: The authors are grateful to Regina Vickery-Grund for her careful editing of this manuscript. **Conflicts of Interest:** The authors declare no conflict of interest.

References

- 1. Brunner, N.; Groeger, S.; Canelas Raposo, J.; Bruckmaier, R.M.; Gross, J.J. Prevalence of subclinical ketosis and production diseases in dairy cows in Central and South America, Africa, Asia, Australia, New Zealand, and Eastern Europe1. *Transl. Anim. Sci.* **2019**, *3*, 84–92. [CrossRef]
- 2. Krieger, M.; Sjöström, K.; Blanco-Penedo, I.; Madouasse, A.; Duval, J.E.; Bareille, N.; Fourichon, C.; Sundrum, A.; Emanuelson, U. Prevalence of production disease related indicators in organic dairy herds in four European countries. *Livest. Sci.* **2017**, *198*, 104–108. [CrossRef]
- 3. van Soest, F.J.S.; Mourits, M.C.M.; Blanco-Penedo, I.; Duval, J.; Fall, N.; Krieger, M.; Sjöstrom, K.; Hogeveen, H. Farm-specific failure costs of production disorders in European organic dairy herds. *Prev. Vet. Med.* **2019**, 168, 19–29. [CrossRef]
- Compton, C.W.R.; Heuer, C.; Thomsen, P.T.; Carpenter, T.E.; Phyn, C.V.C.; McDougall, S. Invited review: A systematic literature review and meta-analysis of mortality and culling in dairy cattle. *J. Dairy Sci.* 2017, 100, 1–16. [CrossRef] [PubMed]

5. Fischer, A.; Friggens, N.C.; Berry, D.P.; Faverdin, P. Isolating the cow-specific part of residual energy intake in lactating dairy cows using random regressions. *Animal* **2018**, *12*, 1396–1404. [CrossRef]

- De Koster, J.; Salavati, M.; Grelet, C.; Crowe, M.A.; Matthews, E.; O'Flaherty, R.; Opsomer, G.; Foldager, L.; Hostens, M. Prediction of metabolic clusters in early-lactation dairy cows using models based on milk biomarkers. J. Dairy Sci. 2019, 102, 2631–2644. [CrossRef]
- 7. Weikard, R.; Goldammer, T.; Brunner, R.M.; Kuehn, C. Tissue-specific mRNA expression patterns reveal a coordinated metabolic response associated with genetic selection for milk production in cows. *Physiol. Genom.* **2012**, 44, 728–739. [CrossRef]
- 8. Nayeri, S.; Stothard, P. Tissues, Metabolic Pathways and Genes of Key Importance in Lactating Dairy Cattle. *Springer Sci. Rev.* **2016**, *4*, 49–77. [CrossRef]
- 9. Bauman, D.E.; Currie, W.B. Partitioning of nutrients during pregnancy and lactation: A review of mechanisms involving homeostasis and homeorhesis. *J. Dairy Sci.* **1980**, *63*, 1514–1529. [CrossRef]
- 10. Sordillo, L.M.; Raphael, W. Significance of metabolic stress, lipid mobilization, and inflammation on transition cow disorders. *Vet. Clin. N. Am. Food Anim. Pract.* **2013**, *29*, 267–278. [CrossRef]
- 11. Kessel, S.; Stroehl, M.; Meyer, H.H.D.; Hiss, S.; Sauerwein, H.; Schwarz, F.J.; Bruckmaier, R.M. Individual variability in physiological adaptation to metabolic stress during early lactation in dairy cows kept under equal conditions. *J. Anim. Sci.* **2008**, *86*, 2903–2912. [CrossRef] [PubMed]
- 12. Graber, M.; Kohler, S.; Müller, A.; Burgermeister, K.; Kaufmann, T.; Bruckmaier, R.M.; van Dorland, H.A. Identification of plasma and hepatic parameters related to metabolic robustness in dairy cows. *J. Anim. Physiol. Anim. Nutr.* **2012**, *96*, 75–84. [CrossRef] [PubMed]
- 13. van Dorland, H.A.; Richter, S.; Morel, I.; Doherr, M.G.; Castro, N.; Bruckmaier, R.M. Variation in hepatic regulation of metabolism during the dry period and in early lactation in dairy cows. *J. Dairy Sci.* **2009**, 92, 1924–1940. [CrossRef] [PubMed]
- 14. Ingvartsen, K.L.; Friggens, N.C. To what extent do variabilities in hormones, metabolites and energy intake explain variability in milk yield? *Domest. Anim. Endocrinol.* **2005**, *29*, 294–304. [CrossRef]
- 15. Ortigues-Marty, I.; Agabriel, J.; Vernet, J.; Sepchat, B.; Al-Jammas, M.; Nozière, P. Contribution of nutrient fluxes to the evolution of the net energy systems, example of the INRA feeding system for beef cattle. *Transl. Anim. Sci.* **2019**, *3*, 1048–1063. [CrossRef]
- 16. Bickerstaffe, R.; Annison, E.F.; Linzell, J.L. The metabolism of glucose, acetate, lipids and amino acids in lactating dairy cows. *J. Agric. Sci.* **1974**, *82*, 71. [CrossRef]
- 17. Linzell, J.L. Mechanism of Secretion of the Aqueous Phase of Milk. *J. Dairy Sci.* **1972**, *55*, 1316–1322. [CrossRef]
- 18. Rigout, S.; Lemosquet, S.; van Eys, J.E.; Blum, J.W.; Rulquin, H. Duodenal Glucose Increases Glucose Fluxes and Lactose Synthesis in Grass Silage-Fed Dairy Cows. *J. Dairy Sci.* **2002**, *85*, 595–606. [CrossRef]
- 19. Chaiyabutr, N.; Faulkner, A.; Peaker, M. The utilization of glucose for the synthesis of milk components in the fed and starved lactating goat in vivo. *Biochem. J.* **1980**, *186*, 301–308. [CrossRef]
- 20. Smith, G.H.; Crabtree, B.; Smith, R. Energy metabolism in the mammary gland. In *Biochemistry of Lactation*; Mepham, T.B., Ed.; Elsevier: Amsterdam, The Netherlands, 1983; pp. 121–141. ISBN 0444804897.
- 21. Ingvartsen, K.L.; Moyes, K.M. Factors contributing to immunosuppression in the dairy cow during the periparturient period. *Jpn. J. Vet. Res.* **2015**, *63* (Suppl. 1), S15–S24.
- Kvidera, S.K.; Horst, E.A.; Abuajamieh, M.; Mayorga, E.J.; Fernandez, M.V.S.; Baumgard, L.H. Glucose requirements of an activated immune system in lactating Holstein cows. *J. Dairy Sci.* 2017, 100, 2360–2374. [CrossRef] [PubMed]
- 23. Aschenbach, J.R.; Kristensen, N.B.; Donkin, S.S.; Hammon, H.M.; Penner, G.B. Gluconeogenesis in dairy cows: The secret of making sweet milk from sour dough. *IUBMB Life* **2010**, *62*, 869–877. [CrossRef] [PubMed]
- 24. Rauw, W.M. Introduction. In *Resource Allocation Theory Applied to Farm Animal Production*; Rauw, W.M., Ed.; CABI: Wallingford, UK; Cambridge, MA, USA, 2009; pp. 1–21. ISBN 184593394X.
- 25. Sundrum, A. Nutrition and Health-Management in Dairy Production. In *Livestock Health and Farming*; Abubakar, M., Ed.; IntechOpen: London, UK, 2020.
- 26. Knap, P.W. Allocation of Resources to Maintenance. In *Resource Allocation Theory Applied to Farm Animal Production*; Rauw, W.M., Ed.; CABI: Wallingford, UK; Cambridge, MA, USA, 2009; pp. 110–129. ISBN 184593394X.

27. Colditz, I.G. Allocation of Resources to Immune Responses. In *Resource Allocation Theory Applied to Farm Animal Production*; Rauw, W.M., Ed.; CABI: Wallingford, UK; Cambridge, MA, USA, 2009; pp. 192–209. ISBN 184593394X.

- 28. Baumgard, L.H.; Collier, R.J.; Bauman, D.E. A 100-Year Review: Regulation of nutrient partitioning to support lactation. *J. Dairy Sci.* **2017**, *100*, 10353–10366. [CrossRef] [PubMed]
- 29. Veerkamp, R.F. Selection for economic efficiency of dairy cattle using information on live weight and feed intake: A review. *J. Dairy Sci.* **1998**, *81*, 1109–1119. [CrossRef]
- 30. Huber, K. Invited review: Resource allocation mismatch as pathway to disproportionate growth in farm animals-prerequisite for a disturbed health. *Animal* **2018**, *12*, 528–536. [CrossRef]
- 31. Ortigues-Marty, I.; Cantalapiedra-Hijar, G.; Vernet, J.; Nozière, P. De l'énergie de la ration à l'utilisation des nutriments chez les ruminants: Quel rôle pour les tissus splanchniques? *Cah. Nutr. Diététique* **2017**, *52*, 45–53. [CrossRef]
- 32. Tyrrell, H.F.; Moe, P.W. Effect of Intake on Digestive Efficiency. J. Dairy Sci. 1975, 58, 1151–1163. [CrossRef]
- 33. Ledinek, M.; Gruber, L.; Steininger, F.; Fuerst-Waltl, B.; Zottl, K.; Royer, M.; Krimberger, K.; Mayerhofer, M.; Egger-Danner, C. Analysis of lactating cows on commercial Austrian dairy farms: The influence of genotype and body weight on efficiency parameters. *Arch. Anim. Breed.* **2019**, *62*, 491–500. [CrossRef]
- 34. Loncke, C.; Nozière, P.; Vernet, J.; Lapierre, H.; Bahloul, L.; Al-Jammas, M.; Sauvant, D.; Ortigues-Marty, I. Net hepatic release of glucose from precursor supply in ruminants: A meta-analysis. *Animal* **2020**, *14*, 1–16. [CrossRef]
- 35. Oltenacu, P.A.; Broom, D.M. The impact of genetic selection for increased milk yield on the welfare of dairy cows. *Anim. Welf.* **2010**, *19*, 39–49.
- 36. Domblides, C.; Lartigue, L.; Faustin, B. Metabolic Stress in the Immune Function of T Cells, Macrophages and Dendritic Cells. *Cells* **2018**, 7, 68. [CrossRef] [PubMed]
- 37. Ingvartsen, K.L.; Dewhurst, R.J.; Friggens, N.C. On the relationship between lactational performance and health: Is it yield or metabolic imbalance that cause production diseases in dairy cattle? A position paper. *Livest. Prod. Sci.* **2003**, *83*, 277–308. [CrossRef]
- 38. Sterling, P. Principles of Allostasis: Optimal Design, Predictive Regulation, Pathophysiology, and Rational Therapeutics. In *Allostasis, Homeostasis and the Costs of Physiological Adaptation*; Schulkin, J., Ed.; Cambridge University Press: Cambridge, UK, 2004; pp. 17–64. ISBN 9781316257081.
- 39. Friggens, N.C.; Newbold, J.R. Towards a biological basis for predicting nutrient partitioning: The dairy cow as an example. *Animal* **2007**, *1*, 87–97. [CrossRef] [PubMed]
- 40. McEwen, B.S.; Wingfield, J.C. What's in a name? Integrating homeostasis, allostasis and stress. *Horm. Behav.* **2009**, *57*, 105. [CrossRef] [PubMed]
- 41. Moberg, G.P. Biological response to stress: Implications for animal welfare. In *The Biology of Animal Stress: Basic Principles and Implications for Animal Welfare, Reprinted*; Moberg, G.P., Mench, J.A., Eds.; CABI Publ.: Wallingford, UK, 2001; ISBN 0851993591.
- 42. Tucker, H.A. Physiological Control of Mammary Growth, Lactogenesis, and Lactation. *J. Dairy Sci.* **1981**, *64*, 1403–1421. [CrossRef]
- 43. Capuco, A.V.; Wood, D.L.; Baldwin, R.; Mcleod, K.; Paape, M.J. Mammary Cell Number, Proliferation, and Apoptosis During a Bovine Lactation: Relation to Milk Production and Effect of bST. *J. Dairy Sci.* **2001**, *84*, 2177–2187. [CrossRef]
- 44. Akers, R.M.; Bauman, D.E.; Capuco, A.V.; Goodman, G.T.; Tucker, H.A. Prolactin regulation of milk secretion and biochemical differentiation of mammary epithelial cells in periparturient cows. *Endocrinology* **1981**, *109*, 23–30. [CrossRef]
- 45. Akers, R.M. *Lactation and the Mammary Gland*, 1st ed.; Blackwell Publishing: Ames, Iowa, 2002; pp. 129–198. ISBN 9781119264880.
- 46. Vernon, R.G. Endocrine control of metabolic adaptation during lactation. *Proc. Nutr. Soc.* **1989**, *48*, 23–32. [CrossRef]
- 47. Kobayashi, Y.; Boyd, C.K.; Bracken, C.J.; Lamberson, W.R.; Keisler, D.H.; Lucy, M.C. Reduced growth hormone receptor (GHR) messenger ribonucleic acid in liver of periparturient cattle is caused by a specific down-regulation of GHR 1A that is associated with decreased insulin-like growth factor I. *Endocrinology* **1999**, *140*, 3947–3954. [CrossRef]

Animals **2020**, 10, 1028 15 of 21

48. Lucy, M.C.; Jiang, H.; Kobayashi, Y. Changes in the Somatotrophic Axis Associated with the Initiation of Lactation. *J. Dairy Sci.* **2001**, *84*, E113–E119. [CrossRef]

- 49. Rhoads, R.P.; Kim, J.W.; Leury, B.J.; Baumgard, L.H.; Segoale, N.; Frank, S.J.; Bauman, D.E.; Boisclair, Y.R. Insulin increases the abundance of the growth hormone receptor in liver and adipose tissue of periparturient dairy cows. *J. Nutr.* **2004**, *134*, 1020–1027. [CrossRef] [PubMed]
- 50. Flores-Morales, A.; Greenhalgh, C.J.; Norstedt, G.; Rico-Bautista, E. Negative regulation of growth hormone receptor signaling. *Mol. Endocrinol.* **2006**, *20*, 241–253. [CrossRef] [PubMed]
- 51. Radcliff, R.P.; McCormack, B.L.; Crooker, B.A.; Lucy, M.C. Plasma Hormones and Expression of Growth Hormone Receptor and Insulin-Like Growth Factor-I mRNA in Hepatic Tissue of Periparturient Dairy Cows. *J. Dairy Sci.* 2003, *86*, 3920–3926. [CrossRef]
- 52. Hue, L.; Taegtmeyer, H. The Randle cycle revisited: A new head for an old hat. *Am. J. Physiol. Endocrinol. Metab.* **2009**, 297, E578–E591. [CrossRef] [PubMed]
- 53. Weber, C.; Hametner, C.; Tuchscherer, A.; Losand, B.; Kanitz, E.; Otten, W.; Sauerwein, H.; Bruckmaier, R.M.; Becker, F.; Kanitz, W.; et al. Hepatic gene expression involved in glucose and lipid metabolism in transition cows: Effects of fat mobilization during early lactation in relation to milk performance and metabolic changesade. *J. Dairy Sci.* 2013, *96*, 5670–5681. [CrossRef]
- 54. De Koster, J.D.; Opsomer, G. Insulin resistance in dairy cows. *Vet. Clin. N. Am. Food Anim. Pract.* **2013**, 29, 299–322. [CrossRef]
- 55. Greenfield, R.B.; Cecava, M.J.; Donkin, S.S. Changes in mRNA Expression for Gluconeogenic Enzymes in Liver of Dairy Cattle During the Transition to Lactation1. *J. Dairy Sci.* **2000**, *83*, 1228–1236. [CrossRef]
- 56. Grummer, R.R. Etiology of Lipid-Related Metabolic Disorders in Periparturient Dairy Cows. *J. Dairy Sci.* **1993**, *76*, 3882–3896. [CrossRef]
- 57. Zarrin, M.; Grossen-Rösti, L.; Bruckmaier, R.M.; Gross, J.J. Elevation of blood β-hydroxybutyrate concentration affects glucose metabolism in dairy cows before and after parturition. *J. Dairy Sci.* **2017**, *100*, 2323–2333. [CrossRef]
- 58. Zarrin, M.; De Matteis, L.; Vernay, M.C.M.B.; Wellnitz, O.; van Dorland, H.A.; Bruckmaier, R.M. Long-term elevation of β-hydroxybutyrate in dairy cows through infusion: Effects on feed intake, milk production, and metabolism. *J. Dairy Sci.* **2013**, *96*, 2960–2972. [CrossRef]
- 59. Li, X.; Li, X.; Bai, G.; Chen, H.; Deng, Q.; Liu, Z.; Zhang, L.; Liu, G.; Wang, Z. Effects of non-esterified fatty acids on the gluconeogenesis in bovine hepatocytes. *Mol. Cell. Biochem.* **2012**, 359, 385–388. [CrossRef] [PubMed]
- 60. Herdt, T.H. Variability Characteristics and Test Selection In Herdlevel Nutritional And Metabolic Profile Testing. *Vet. Clin. N. Am. Food Anim. Pract.* **2000**, *16*, 387–403. [CrossRef]
- 61. Ruoff, J.; Borchardt, S.; Heuwieser, W. Short communication: Associations between blood glucose concentration, onset of hyperketonemia, and milk production in early lactation dairy cows. *J. Dairy Sci.* 2017, 100, 5462–5467. [CrossRef] [PubMed]
- 62. Bell, A.W.; Bauman, D.E. Adaptations of Glucose Metabolism During Pregnancy and Lactation. *J. Mammary Gland Biol. Neoplasia* **1997**, 2, 265–278. [CrossRef]
- 63. Davey, A.W.F.; Grainger, C.; MacKenzie, D.D.S.; Flux, D.S.; Wilson, G.F.; Brookes, I.M.; Holmes, C.W. Nutritional and physiological studies of differences between Friesian cows of high and low genetic merit. *Proc. N. Z. Soc. Anim. Prod.* **1983**, 67–70.
- 64. Kerestes, M.; Faigl, V.; Kulcsár, M.; Balogh, O.; Földi, J.; Fébel, H.; Chilliard, Y.; Huszenicza, G. Periparturient insulin secretion and whole-body insulin responsiveness in dairy cows showing various forms of ketone pattern with or without puerperal metritis. *Domest. Anim. Endocrinol.* **2009**, *37*, 250–261. [CrossRef]
- 65. Zachut, M.; Honig, H.; Striem, S.; Zick, Y.; Boura-Halfon, S.; Moallem, U. Periparturient dairy cows do not exhibit hepatic insulin resistance, yet adipose-specific insulin resistance occurs in cows prone to high weight loss. *J. Dairy Sci.* 2013, *96*, 5656–5669. [CrossRef]
- 66. Ostrowska, M.; Jarczak, J. Glucose transporters in cattle—A review. Anim. Sci. Pap. Rep. 2015, 33, 191–212.
- 67. Komatsu, T.; Itoh, F.; Kushibiki, S.; Hodate, K. Changes in gene expression of glucose transporters in lactating and nonlactating cows. *J. Anim. Sci.* **2005**, *83*, 557–564. [CrossRef]
- 68. Mattmiller, S.A.; Corl, C.M.; Gandy, J.C.; Loor, J.J.; Sordillo, L.M. Glucose transporter and hypoxia-associated gene expression in the mammary gland of transition dairy cattle. *J. Dairy Sci.* **2011**, *94*, 2912–2922. [CrossRef]

Animals **2020**, 10, 1028 16 of 21

69. Prosser, C.G.; Davis, S.R.; Farr, V.C.; Lacasse, P. Regulation of Blood Flow in the Mammary Microvasculature1. *J. Dairy Sci.* **1996**, *79*, 1184–1197. [CrossRef]

- 70. Davis, S.R.; Collier, R.J. Mammary Blood Flow and Regulation of Substrate Supply for Milk Synthesis. *J. Dairy Sci.* **1985**, *68*, 1041–1058. [CrossRef]
- 71. Davis, S.R.; Collier, R.J.; McNamara, J.P.; Head, H.H.; Croom, W.J.; Wilcox, C.J. Effects of thyroxine and growth hormone treatment of dairy cows on mammary uptake of glucose, oxygen and other milk fat precursors. *J. Anim. Sci.* **1988**, *66*, 80–89. [CrossRef] [PubMed]
- 72. Kronfeld, D.S.; Raggi, F.; Ramberg, C.F. Mammary blood flow and ketone body metabolism in normal, fasted, and ketotic cows. *Am. J. Physiol.* **1968**, 215, 218–227. [CrossRef] [PubMed]
- 73. Samanc, H.; Kirovski, D.; Lakić, N.; Celeska, I.; Bojković-Kovačević, S.; Sladojević, Z.; Ivanov, I. A comparison of the concentrations of energy-balance-related variables in jugular and mammary vein blood of dairy cows with different milk yield. *Acta Vet. Hung.* **2014**, *62*, 52–63. [CrossRef] [PubMed]
- 74. Sadri, H.; Bruckmaier, R.M.; Rahmani, H.R.; Ghorbani, G.R.; Morel, I.; van Dorland, H.A. Gene expression of tumour necrosis factor and insulin signalling-related factors in subcutaneous adipose tissue during the dry period and in early lactation in dairy cows. *J. Anim. Physiol. Anim. Nutr.* **2010**, 94, e194–e202. [CrossRef]
- 75. Kuhla, B.; Nürnberg, G.; Albrecht, D.; Görs, S.; Hammon, H.M.; Metges, C.C. Involvement of skeletal muscle protein, glycogen, and fat metabolism in the adaptation on early lactation of dairy cows. *J. Proteome Res.* **2011**, *10*, 4252–4262. [CrossRef]
- 76. Larsen, M.; Kristensen, N.B. Precursors for liver gluconeogenesis in periparturient dairy cows. *Animal* **2013**, 7, 1640–1650. [CrossRef]
- 77. Esposito, G.; Irons, P.C.; Webb, E.C.; Chapwanya, A. Interactions between negative energy balance, metabolic diseases, uterine health and immune response in transition dairy cows. *Anim. Reprod. Sci.* **2014**, *144*, 60–71. [CrossRef]
- 78. Minuti, A.; Palladino, A.; Khan, M.J.; Alqarni, S.; Agrawal, A.; Piccioli-Capelli, F.; Hidalgo, F.; Cardoso, F.C.; Trevisi, E.; Loor, J.J. Abundance of ruminal bacteria, epithelial gene expression, and systemic biomarkers of metabolism and inflammation are altered during the peripartal period in dairy cows. *J. Dairy Sci.* **2015**, *98*, 8940–8951. [CrossRef]
- 79. Bradford, B.J.; Yuan, K.; Farney, J.K.; Mamedova, L.K.; Carpenter, A.J. Invited review: Inflammation during the transition to lactation: New adventures with an old flame. *J. Dairy Sci.* **2015**, *98*, 6631–6650. [CrossRef] [PubMed]
- 80. Qu, Y.; Fadden, A.N.; Traber, M.G.; Bobe, G. Potential risk indicators of retained placenta and other diseases in multiparous cows. *J. Dairy Sci.* **2014**, 97, 4151–4165. [CrossRef] [PubMed]
- 81. Huzzey, J.M.; Mann, S.; Nydam, D.V.; Grant, R.J.; Overton, T.R. Associations of peripartum markers of stress and inflammation with milk yield and reproductive performance in Holstein dairy cows. *Prev. Vet. Med.* **2015**, *120*, 291–297. [CrossRef] [PubMed]
- 82. Thorburn, G.D. The placenta, prostaglandins and parturition: A review. *Reprod. Fertil. Dev.* **1991**, *3*, 277–294. [CrossRef] [PubMed]
- 83. Newby, N.C.; Leslie, K.E.; Dingwell, H.D.P.; Kelton, D.F.; Weary, D.M.; Neuder, L.; Millman, S.T.; Duffield, T.F. The effects of periparturient administration of flunixin meglumine on the health and production of dairy cattle. *J. Dairy Sci.* **2017**, *100*, 582–587. [CrossRef] [PubMed]
- 84. Pond, C.M.; Newsholme, E.A. Coping with metabolic stress in wild and domesticated animals. *Bsap Occas. Publ.* **1999**, 24, 9–20. [CrossRef]
- 85. Liu, S.M.; Smith, T.L.; Karlsson, L.J.E.; Palmer, D.G.; Besier, R.B. The costs for protein and energy requirements by nematode infection and resistance in Merino sheep. *Livest. Prod. Sci.* **2005**, *97*, 131–139. [CrossRef]
- 86. Rathmell, J.C.; Fox, C.J.; Plas, D.R.; Hammerman, P.S.; Cinalli, R.M.; Thompson, C.B. Akt-directed glucose metabolism can prevent Bax conformation change and promote growth factor-independent survival. *Mol. Cell. Biol.* **2003**, *23*, 7315–7328. [CrossRef]
- 87. Galvão, K.N.; Flaminio, M.J.B.F.; Brittin, S.B.; Sper, R.; Fraga, M.; Caixeta, L.; Ricci, A.; Guard, C.L.; Butler, W.R.; Gilbert, R.O. Association between uterine disease and indicators of neutrophil and systemic energy status in lactating Holstein cows. *J. Dairy Sci.* **2010**, *93*, 2926–2937. [CrossRef]
- 88. Calder, P.C. Fatty acids, dietary lipids and lymphocyte functions. *Biochem. Soc. Trans.* **1995**, 23, 302–309. [CrossRef]

89. Castell, L.M.; Newsholme, E.A. Glutamine and the effects of exhaustive exercise upon the immune response. *Can. J. Physiol. Pharmacol.* **1998**, *76*, 524–532. [CrossRef]

- 90. Gross, J.J.; Grossen-Rösti, L.; Héritier, R.; Tröscher, A.; Bruckmaier, R.M. Inflammatory and metabolic responses to an intramammary lipopolysaccharide challenge in early lactating cows supplemented with conjugated linoleic acid. *J. Anim. Physiol. Anim. Nutr.* **2018**, *102*, e838–e848. [CrossRef] [PubMed]
- 91. Noleto, P.G.; Saut, J.P.E.; Sheldon, I.M. Short communication: Glutamine modulates inflammatory responses to lipopolysaccharide in ex vivo bovine endometrium. *J. Dairy Sci.* **2017**, *100*, 2207–2212. [CrossRef] [PubMed]
- 92. Williamson, R.T. On the Treatment of Glycosuria and Diabetes Mellitus with Sodium Salicylate. *Br. Med. J.* **1901**, *1*, 760–762. [CrossRef]
- 93. Kyriakis, J.M.; Avruch, J. Mammalian MAPK signal transduction pathways activated by stress and inflammation: A 10-year update. *Physiol. Rev.* **2012**, 92, 689–737. [CrossRef] [PubMed]
- 94. Horst, E.A.; Kvidera, S.K.; Dickson, M.J.; McCarthy, C.S.; Mayorga, E.J.; Al-Qaisi, M.; Ramirez, H.A.; Keating, A.F.; Baumgard, L.H. Effects of continuous and increasing lipopolysaccharide infusion on basal and stimulated metabolism in lactating Holstein cows. *J. Dairy Sci.* **2019**, *102*, 3584–3597. [CrossRef]
- 95. Zhao, F.-Q.; Keating, A.F. Expression and regulation of glucose transporters in the bovine mammary gland. *J. Dairy Sci.* **2007**, *90* (Suppl. 1), E76–E86. [CrossRef]
- 96. Eger, M.; Hussen, J.; Koy, M.; Danicke, S.; Schuberth, H.-J.; Breves, G. Glucose transporter expression differs between bovine monocyte and macrophage subsets and is influenced by milk production. *J. Dairy Sci.* **2016**, 99, 2276–2287. [CrossRef]
- 97. Maratou, E.; Dimitriadis, G.; Kollias, A.; Boutati, E.; Lambadiari, V.; Mitrou, P.; Raptis, S.A. Glucose transporter expression on the plasma membrane of resting and activated white blood cells. *Eur. J. Clin. Investig.* **2007**, 37, 282–290. [CrossRef]
- 98. Mamedova, L.K.; Yuan, K.; Laudick, A.N.; Fleming, S.D.; Mashek, D.G.; Bradford, B.J. Toll-like receptor 4 signaling is required for induction of gluconeogenic gene expression by palmitate in human hepatic carcinoma cells. *J. Nutr. Biochem.* **2013**, 24, 1499–1507. [CrossRef]
- 99. Kominsky, D.J.; Campbell, E.L.; Colgan, S.P. Metabolic shifts in immunity and inflammation. *J. Immunol.* **2010**, *184*, 4062–4068. [CrossRef] [PubMed]
- 100. Wannemacher, R.W.; Beall, F.A.; Canonico, P.G.; Dinterman, R.E.; Hadick, C.L.; Neufeld, H.A. Glucose and alanine metabolism during bacterial infections in rats and rhesus monkeys. *Metabolism* **1980**, *29*, 201–212. [CrossRef]
- 101. Hotamisligil, G.S. Inflammation and metabolic disorders. Nature 2006, 444, 860–867. [CrossRef] [PubMed]
- 102. Gross, J.J.; Bruckmaier, R.M. Invited review: Metabolic challenges and adaptation during different functional stages of the mammary gland in dairy cows: Perspectives for sustainable milk production. *J. Dairy Sci.* **2019**, 102, 2828–2843. [CrossRef] [PubMed]
- 103. Mezzetti, M.; Minuti, A.; Piccioli-Cappelli, F.; Amadori, M.; Bionaz, M.; Trevisi, E. The role of altered immune function during the dry period in promoting the development of subclinical ketosis in early lactation. *J. Dairy Sci.* **2019**. [CrossRef]
- 104. Gao, Y.; Lin, X.; Shi, K.; Yan, Z.; Wang, Z. Bovine mammary gene expression profiling during the onset of lactation. *PLoS ONE* **2013**, *8*, e70393. [CrossRef]
- 105. Goff, J.P.; Horst, R.L. Physiological Changes at Parturition and Their Relationship to Metabolic Disorders. *J. Dairy Sci.* **1997**, *80*, 1260–1268. [CrossRef]
- 106. Kehrli, M.E.; Nonnecke, B.J.; Roth, J.A. Alterations in bovine lymphocyte function during the periparturient period. *Am. J. Vet. Res.* **1989**, *50*, 215–220.
- 107. van Kampen, C.; Mallard, B.A. Effects of peripartum stress and health on circulating bovine lymphocyte subsets. *Vet. Immunol. Immunopathol.* **1997**, *59*, 79–91. [CrossRef]
- 108. Ohtsuka, H.; Koiwa, M.; Fukuda, S.; Satoh, Y.; Hayashi, T.; Hoshi, F.; Yoshino, T.-O.; Kawamura, S.-I. Changes in peripheral leukocyte subsets in dairy cows with inflammatory diseases after calving. *J. Vet. Med. Sci.* **2004**, *66*, 905–909. [CrossRef]
- 109. Lamote, I.; Meyer, E.; Duchateau, L.; Burvenich, C. Influence of 17β-Estradiol, Progesterone, and Dexamethasone on Diapedesis and Viability of Bovine Blood Polymorphonuclear Leukocytes. *J. Dairy Sci.* **2004**, *87*, 3340–3349. [CrossRef]

Animals **2020**, 10, 1028 18 of 21

110. Saama, P.M.; Jacob, J.B.; Kehrli, M.E.; Freeman, A.E.; Kelm, S.C.; Kuck, A.L.; Tempelman, R.J.; Burton, J.L. Genetic Variation in Bovine Mononuclear Leukocyte Responses to Dexamethasone. *J. Dairy Sci.* **2004**, *87*, 3928–3937. [CrossRef]

- 111. Burton, J.L.; Madsen, S.A.; Chang, L.-C.; Weber, P.S.D.; Buckham, K.R.; van Dorp, R.; Hickey, M.-C.; Earley, B. Gene expression signatures in neutrophils exposed to glucocorticoids: A new paradigm to help explain "neutrophil dysfunction" in parturient dairy cows. *Vet. Immunol. Immunopathol.* 2005, 105, 197–219. [CrossRef] [PubMed]
- 112. Jahan, N.; Minuti, A.; Trevisi, E. Assessment of immune response in periparturient dairy cows using ex vivo whole blood stimulation assay with lipopolysaccharides and carrageenan skin test. *Vet. Immunol. Immunopathol.* 2015, 165, 119–126. [CrossRef]
- 113. Lee, E.K.; Kehrli, M.E. Expression of adhesion molecules on neutrophils of periparturient cows and neonatal calves. *Am. J. Vet. Res.* **1998**, *59*, *37*–43.
- 114. Kimura, K.; Goff, J.P.; Kehrli, M.E. Effects of the presence of the mammary gland on expression of neutrophil adhesion molecules and myeloperoxidase activity in periparturient dairy cows. *J. Dairy Sci.* 1999, 82, 2385–2392. [CrossRef]
- 115. Kimura, K.; Goff, J.P.; Kehrli, M.E.; Harp, J.A.; Nonnecke, B.J. Effects of Mastectomy on Composition of Peripheral Blood Mononuclear Cell Populations in Periparturient Dairy Cows1. *J. Dairy Sci.* **2002**, *85*, 1437–1444. [CrossRef]
- 116. Nonnecke, B.J.; Kimura, K.; Goff, J.P.; Kehrli, M.E. Effects of the Mammary Gland on Functional Capacities of Blood Mononuclear Leukocyte Populations from Periparturient Cows1. *J. Dairy Sci.* **2003**, *86*, 2359–2368. [CrossRef]
- 117. Minuti, A.; Bionaz, M.; Lopreiato, V.; Janovick, N.A.; Rodriguez-Zas, S.L.; Drackley, J.K.; Loor, J.J. Prepartum dietary energy intake alters adipose tissue transcriptome profiles during the periparturient period in Holstein dairy cows. *J. Anim. Sci. Biotechnol.* **2020**, *11*, 1–14. [CrossRef]
- 118. Minuti, A.; Jahan, N.; Lopreiato, V.; Piccioli-Cappelli, F.; Bomba, L.; Capomaccio, S.; Loor, J.J.; Ajmone-Marsan, P.; Trevisi, E. Evaluation of circulating leukocyte transcriptome and its relationship with immune function and blood markers in dairy cows during the transition period. *Funct. Integr. Genom.* **2020**, *20*, 293–305. [CrossRef]
- 119. Contreras, G.A.; Strieder-Barboza, C.; De Koster, J. Symposium review: Modulating adipose tissue lipolysis and remodeling to improve immune function during the transition period and early lactation of dairy cows. *J. Dairy Sci.* 2018, 101, 2737–2752. [CrossRef] [PubMed]
- 120. Sordillo, L.M.; Aitken, S.L. Impact of oxidative stress on the health and immune function of dairy cattle. *Vet. Immunol. Immunopathol.* **2009**, *128*, 104–109. [CrossRef] [PubMed]
- 121. Abuelo, A.; Hernández, J.; Benedito, J.L.; Castillo, C. Association of oxidative status and insulin sensitivity in periparturient dairy cattle: An observational study. *J. Anim. Physiol. Anim. Nutr.* **2016**, 100, 279–286. [CrossRef] [PubMed]
- 122. Song, Y.; Li, X.; Li, Y.; Li, N.; Shi, X.; Ding, H.; Zhang, Y.; Li, X.; Liu, G.; Wang, Z. Non-esterified fatty acids activate the ROS-p38-p53/Nrf2 signaling pathway to induce bovine hepatocyte apoptosis in vitro. *Apoptosis* **2014**, *19*, 984–997. [CrossRef]
- 123. Contreras, G.A.; O'Boyle, N.J.; Herdt, T.H.; Sordillo, L.M. Lipomobilization in periparturient dairy cows influences the composition of plasma nonesterified fatty acids and leukocyte phospholipid fatty acids. *J. Dairy Sci.* **2010**, *93*, 2508–2516. [CrossRef] [PubMed]
- 124. Kasimanickam, R.K.; Kasimanickam, V.R.; Olsen, J.R.; Jeffress, E.J.; Moore, D.A.; Kastelic, J.P. Associations among serum pro-and anti-inflammatory cytokines, metabolic mediators, body condition, and uterine disease in postpartum dairy cows. *Reprod. Biol. Endocrinol.* 2013, 11, 103. [CrossRef]
- 125. Mecitoglu, Z.; Senturk, S.; Akgul, G.; Udum, D.; Uzabacı, E.; Kasap, S.; Catik, S. Changes in circulating adiponectin and tumour necrosis factor-α and their relationship with insulin resistance in periparturient dairy cows. *J. Vet. Res.* **2016**, *60*, 163–167. [CrossRef]
- 126. Loor, J.J.; Dann, H.M.; Everts, R.E.; Oliveira, R.; Green, C.A.; Guretzky, N.A.J.; Rodriguez-Zas, S.L.; Lewin, H.A.; Drackley, J.K. Temporal gene expression profiling of liver from periparturient dairy cows reveals complex adaptive mechanisms in hepatic function. *Physiol. Genom.* 2005, 23, 217–226. [CrossRef]

127. Wathes, D.C.; Cheng, Z.; Chowdhury, W.; Fenwick, M.A.; Fitzpatrick, R.; Morris, D.G.; Patton, J.; Murphy, J.J. Negative energy balance alters global gene expression and immune responses in the uterus of postpartum dairy cows. *Physiol. Genom.* **2009**, *39*, 1–13. [CrossRef] [PubMed]

- 128. Moyes, K.M.; Drackley, J.K.; Morin, D.E.; Rodriguez-Zas, S.L.; Everts, R.E.; Lewin, H.A.; Loor, J.J. Mammary gene expression profiles during an intramammary challenge reveal potential mechanisms linking negative energy balance with impaired immune response. *Physiol. Genom.* **2010**, *41*, 161–170. [CrossRef] [PubMed]
- 129. Moyes, K.M.; Drackley, J.K.; Salak-Johnson, J.L.; Morin, D.E.; Hope, J.C.; Loor, J.J. Dietary-induced negative energy balance has minimal effects on innate immunity during a Streptococcus uberis mastitis challenge in dairy cows during midlactation. *J. Dairy Sci.* **2009**, *92*, 4301–4316. [CrossRef] [PubMed]
- 130. Perkins, K.H.; VandeHaar, M.J.; Burton, J.L.; Liesman, J.S.; Erskine, R.J.; Elsasser, T.H. Clinical responses to intramammary endotoxin infusion in dairy cows subjected to feed restriction. *J. Dairy Sci.* **2002**, *85*, 1724–1731. [CrossRef]
- 131. Perkins, K.H.; VandeHaar, M.J.; Tempelman, R.J.; Burton, J.L. Negative energy balance does not decrease expression of leukocyte adhesion or antigen-presenting molecules in cattle. *J. Dairy Sci.* **2001**, *84*, 421–428. [CrossRef]
- 132. Bjerre-Harpøth, V.; Friggens, N.C.; Thorup, V.M.; Larsen, T.; Damgaard, B.M.; Ingvartsen, K.L.; Moyes, K.M. Metabolic and production profiles of dairy cows in response to decreased nutrient density to increase physiological imbalance at different stages of lactation. *J. Dairy Sci.* 2012, 95, 2362–2380. [CrossRef] [PubMed]
- 133. Gross, J.; van Dorland, H.; Bruckmaier, R.M.; Schwarz, F. Performance and metabolic profile of dairy cows during a lactational and deliberately induced negative energy balance with subsequent realimentation. *J. Dairy Sci.* **2011**, *94*, 1820–1830. [CrossRef]
- 134. Eger, M.; Hussen, J.; Drong, C.; Meyer, U.; von Soosten, D.; Frahm, J.; Daenicke, S.; Breves, G.; Schuberth, H.-J. Impacts of parturition and body condition score on glucose uptake capacity of bovine monocyte subsets. *Vet. Immunol. Immunopathol.* **2015**, *166*, 33–42. [CrossRef]
- 135. Bouvier-Muller, J.; Allain, C.; Tabouret, G.; Enjalbert, F.; Portes, D.; Noirot, C.; Rupp, R.; Foucras, G. Whole blood transcriptome analysis reveals potential competition in metabolic pathways between negative energy balance and response to inflammatory challenge. *Sci. Rep.* **2017**, *7*, 2379. [CrossRef]
- 136. Shin, D.H.; Jeong, J.K.; Choi, I.S.; Moon, S.H.; Lee, S.C.; Kang, H.G.; Park, S.B.; Kim, I.H. Associations between serum haptoglobin concentration and peri-and postpartum disorders, milk yield, and reproductive performance in dairy cows. *Livest. Sci.* 2018, 213, 14–18. [CrossRef]
- 137. Winkler, A.; Gessner, D.K.; Koch, C.; Romberg, F.-J.; Dusel, G.; Herzog, E.; Most, E.; Eder, K. Effects of a plant product consisting of green tea and curcuma extract on milk production and the expression of hepatic genes involved in endoplasmic stress response and inflammation in dairy cows. *Arch. Anim. Nutr.* **2015**, *69*, 425–441. [CrossRef]
- 138. Carpenter, A.J.; Ylioja, C.M.; Vargas, C.F.; Mamedova, L.K.; Mendonça, L.G.; Coetzee, J.F.; Hollis, L.C.; Gehring, R.; Bradford, B.J. Hot topic: Early postpartum treatment of commercial dairy cows with nonsteroidal antiinflammatory drugs increases whole-lactation milk yield. *J. Dairy Sci.* 2016, 99, 672–679. [CrossRef]
- 139. Gross, J.; van Dorland, H.; Wellnitz, O.; Bruckmaier, R.M. Glucose transport and milk secretion during manipulated plasma insulin and glucose concentrations and during LPS-induced mastitis in dairy cows. *J. Anim. Physiol. Anim. Nutr.* **2014**, 99. [CrossRef] [PubMed]
- 140. Osorio, J.S.; Lohakare, J.; Bionaz, M. Biosynthesis of milk fat, protein, and lactose: Roles of transcriptional and posttranscriptional regulation. *Physiol. Genom.* **2016**, *48*, 231–256. [CrossRef] [PubMed]
- 141. Kreipe, L.; Vernay, M.C.M.B.; Oppliger, A.; Wellnitz, O.; Bruckmaier, R.M.; van Dorland, H.A. Induced hypoglycemia for 48 hours indicates differential glucose and insulin effects on liver metabolism in dairy cows. *J. Dairy Sci.* 2011, *94*, 5435–5448. [CrossRef] [PubMed]
- 142. Lindmark-Månsson, H.; Bränning, C.; Aldén, G.; Paulsson, M. Relationship between somatic cell count, individual leukocyte populations and milk components in bovine udder quarter milk. *Int. Dairy J.* 2006, 16, 717–727. [CrossRef]
- 143. Moyes, K.M.; Drackley, J.K.; Morin, D.E.; Bionaz, M.; Rodriguez-Zas, S.L.; Everts, R.E.; Lewin, H.A.; Loor, J.J. Gene network and pathway analysis of bovine mammary tissue challenged with Streptococcus uberis reveals induction of cell proliferation and inhibition of PPARgamma signaling as potential mechanism for the negative relationships between immune response and lipid metabolism. *BMC Genom.* **2009**, *10*, 542. [CrossRef]

Animals **2020**, 10, 1028 20 of 21

144. Liu, H.; Zhao, K.; Liu, J. Effects of glucose availability on expression of the key genes involved in synthesis of milk fat, lactose and glucose metabolism in bovine mammary epithelial cells. *PLoS ONE* **2013**, *8*, e66092. [CrossRef]

- 145. Lehtolainen, T.; Suominen, S.; Kutila, T.; Pyörälä, S. Effect of Intramammary Escherichia coli Endotoxin in Early- vs. Late-Lactating Dairy Cows. *J. Dairy Sci.* **2003**, *86*, 2327–2333. [CrossRef]
- 146. Shuster, D.E.; Lee, E.K.; Kehrli, M.E. Bacterial growth, inflammatory cytokine production, and neutrophil recruitment during coliform mastitis in cows within ten days after calving, compared with cows at midlactation. *Am. J. Vet. Res.* **1996**, *57*, 1569–1575.
- 147. Yuan, K.; Farney, J.K.; Mamedova, L.K.; Sordillo, L.M.; Bradford, B.J. TNFα altered inflammatory responses, impaired health and productivity, but did not affect glucose or lipid metabolism in early-lactation dairy cows. *PLoS ONE* **2013**, *8*, e80316. [CrossRef]
- 148. Hoeben, D.; Burvenich, C.; Trevisi, E.; Bertoni, G.; Hamann, J.; Bruckmaier, R.M.; Blum, J.W. Role of endotoxin and TNF-alpha in the pathogenesis of experimentally induced coliform mastitis in periparturient cows. *J. Dairy Res.* 2000, 67, 503–514. [CrossRef]
- 149. Dickson, M.J.; Kvidera, S.K.; Horst, E.A.; Wiley, C.E.; Mayorga, E.J.; Ydstie, J.; Perry, G.A.; Baumgard, L.H.; Keating, A.F. Impacts of chronic and increasing lipopolysaccharide exposure on production and reproductive parameters in lactating Holstein dairy cows. *J. Dairy Sci.* 2019. [CrossRef] [PubMed]
- 150. Martel, C.A.; Mamedova, L.K.; Minton, J.E.; Jones, M.L.; Carroll, J.A.; Bradford, B.J. Continuous low-dose infusion of tumor necrosis factor alpha in adipose tissue elevates adipose tissue interleukin 10 abundance and fails to alter metabolism in lactating dairy cows. *J. Dairy Sci.* **2014**, *97*, 4897–4906. [CrossRef] [PubMed]
- 151. Snijders, S.E.M.; Dillon, P.G.; O'Farrell, K.J.; Diskin, M.; Wylie, A.R.G.; O'Callaghan, D.; Rath, M.; Boland, M.P. Genetic merit for milk production and reproductive success in dairy cows. *Anim. Reprod. Sci.* **2001**, *65*, 17–31. [CrossRef]
- 152. Veerkamp, R.F.; Beerda, B.; van der Lende, T. Effects of genetic selection for milk yield on energy balance, levels of hormones, and metabolites in lactating cattle, and possible links to reduced fertility. *Livest. Prod. Sci.* 2003, 83, 257–275. [CrossRef]
- 153. Chagas, L.M.; Lucy, M.C.; Back, P.J.; Blache, D.; Lee, J.M.; Gore, P.J.S.; Sheahan, A.J.; Roche, J.R. Insulin resistance in divergent strains of Holstein-Friesian dairy cows offered fresh pasture and increasing amounts of concentrate in early lactation. *J. Dairy Sci.* 2009, 92, 216–222. [CrossRef] [PubMed]
- 154. Bossaert, P.; Leroy, J.L.M.R.; De Campeneere, S.; de Vliegher, S.; Opsomer, G. Differences in the glucose-induced insulin response and the peripheral insulin responsiveness between neonatal calves of the Belgian Blue, Holstein-Friesian, and East Flemish breeds. *J. Dairy Sci.* 2009, 92, 4404–4411. [CrossRef] [PubMed]
- 155. Hart, I.; Bines, J.; Morant, S.; Ridley, J. Endocrine control of energy metabolism in the cow: Comparison of the level of hormones (prolactin, growth hormone, insulin and thyroxine) and metabolites in the plasma of high- and low-yielding cattle at various stages of lactation. *J. Endocrinol.* 1978, 77, 333–345. [CrossRef]
- 156. Newsholme, P.; Costa Rosa, L.F.; Newsholme, E.A.; Curi, R. The importance of fuel metabolism to macrophage function. *Cell Biochem. Funct. Cell. Biochem. Its Modul. Act. Agents Dis.* **1996**, *14*, 1–10. [CrossRef]
- 157. Alba-Loureiro, T.C.; Munhoz, C.D.; Martins, J.O.; Cerchiaro, G.A.; Scavone, C.; Curi, R.; Sannomiya, P. Neutrophil function and metabolism in individuals with diabetes mellitus. *Braz. J. Med. Biol. Res.* **2007**, *40*, 1037–1044. [CrossRef]
- 158. Forsyth, I.A. The Insulin-Like Growth Factor and Epidermal Growth Factor Families in Mammary Cell Growth in Ruminants: Action and Interaction with Hormones. *J. Dairy Sci.* **1996**, *79*, 1085–1096. [CrossRef]
- 159. Flint, D.J.; Gardner, M. Evidence that growth hormone stimulates milk synthesis by direct action on the mammary gland and that prolactin exerts effects on milk secretion by maintenance of mammary deoxyribonucleic acid content and tight junction status. *Endocrinology* **1994**, *135*, 1119–1124. [CrossRef]
- 160. Welniak, L.A.; Sun, R.; Murphy, W.J. The role of growth hormone in T-cell development and reconstitution. *J. Leukoc. Biol.* **2002**, *71*, 381–387. [PubMed]
- 161. Okamura, C.S.; Bader, J.F.; Keisler, D.H.; Lucy, M.C. Growth hormone receptor expression in two dairy breeds during the periparturient period. *J. Dairy Sci.* **2009**, 92, 2706–2710. [CrossRef] [PubMed]
- 162. Jiang, H.; Lucy, M.C.; Crooker, B.A.; Beal, W.E. Expression of growth hormone receptor 1A mRNA is decreased in dairy cows but not in beef cows at parturition. *J. Dairy Sci.* **2005**, *88*, 1370–1377. [CrossRef]
- 163. Trevisi, E.; Minuti, A. Assessment of the innate immune response in the periparturient cow. *Res. Vet. Sci.* **2018**, *116*, 47–54. [CrossRef] [PubMed]

Animals 2020, 10, 1028 21 of 21

164. Beever, D.E. The impact of controlled nutrition during the dry period on dairy cow health, fertility and performance. *Anim. Reprod. Sci.* **2006**, *96*, 212–226. [CrossRef]

- 165. Mann, S.; Leal Yepes, F.A.; Duplessis, M.; Wakshlag, J.J.; Overton, T.R.; Cummings, B.P.; Nydam, D.V. Dry period plane of energy: Effects on glucose tolerance in transition dairy cows. *J. Dairy Sci.* **2016**, *99*, 701–717. [CrossRef]
- 166. Lucy, M.C.; Escalante, R.C.; Keisler, D.H.; Lamberson, W.R.; Mathew, D.J. Short communication: Glucose infusion into early postpartum cows defines an upper physiological set point for blood glucose and causes rapid and reversible changes in blood hormones and metabolites. *J. Dairy Sci.* 2013, *96*, 5762–5768. [CrossRef]
- 167. Mesgaran, M.D.; Koolabadi, G.; Banikamalil, A.; Mesgaran, S.D. Effect on animal performance of the source of energy (glucogenic vs. lipogenic) of early lactating dairy cow diets with similar content of metabolizable energy. In Energy and Protein Metabolism and Nutrition. 3rd EAAP International Symposium on Energy and Protein Metabolism and Nutrition, Parma, Italy, 6–10 September 2010; Wageningen Academic Publishers: Wageningen, The Netherlands, 2010; pp. 335–336. ISBN 9789086861538.
- 168. Kokkonen, T.; Taponen, J.; Anttila, T.; Syrjala-Qvist, L.; Delavaud, C.; Chilliard, Y.; Tuori, M.; Tesfa, A.T. Effect of body fatness and glucogenic supplement on lipid and protein mobilization and plasma leptin in dairy cows. *J. Dairy Sci.* 2005, 88, 1127–1141. [CrossRef]
- 169. van Knegsel, A.T.M.; van den Brand, H.; Dijkstra, J.; van Straalen, W.M.; Heetkamp, M.J.W.; Tamminga, S.; Kemp, B. Dietary Energy Source in Dairy Cows in Early Lactation: Energy Partitioning and Milk Composition. *J. Dairy Sci.* 2007, 90, 1467–1476. [CrossRef]
- 170. Bar-Pelled, U.; Maltz, E.; Bruckental, I.; Folman, Y.; Kali, Y.; Gacitua, H.; Lehrer, A.R.; Knight, C.H.; Robinson, B.; Voet, H.; et al. Relationship Between Frequent Milking or Suckling in Early Lactation and Milk Production of High Producing Dairy Cows1. *J. Dairy Sci.* 1995, 78, 2726–2736. [CrossRef]
- 171. Stelwagen, K.; Phyn, C.V.C.; Davis, S.R.; Guinard-Flament, J.; Pomiès, D.; Roche, J.R.; Kay, J.K. Invited review: Reduced milking frequency: Milk production and management implications. *J. Dairy Sci.* **2013**, *96*, 3401–3413. [CrossRef] [PubMed]
- 172. Lacasse, P.; Vanacker, N.; Ollier, S.; Ster, C. Innovative dairy cow management to improve resistance to metabolic and infectious diseases during the transition period. *Res. Vet. Sci.* **2018**, *116*, 40–46. [CrossRef] [PubMed]
- 173. Riley, M.; Garcia, M.; Ylioja, C.; Mamedova, L.K.; Bradford, B.J. Effects of anti-inflammatory treatment and milking frequency on mRNA abundance of adipose tissue from early lactation cows. In *Abstracts of the 2018 American Dairy Science Association Annual Meeting, Proceedings of the American Dairy Science Association Annual Meeting 2018, Knoxville, TN, USA, 24–27 June 2018*; Lucy, M.C., Ed.; Journal of Dairy Science: Champaign, IL, USA, 2018; p. 61820.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).