#### **REVIEW ARTICLE**



# Feline infectious peritonitis (FIP) and coronavirus disease 19 (COVID-19): Are they similar?

Saverio Paltrinieri<sup>1,2</sup> | Alessia Giordano<sup>1,2</sup> | Angelica Stranieri<sup>1,2</sup> | Stefania Lauzi<sup>1,2</sup>

<sup>1</sup>Department of Veterinary Medicine, University of Milan, Lodi, Italy

<sup>2</sup>Veterinary Teaching Hospital, University of Milan, Lodi, Italy

#### Correspondence

Angelica Stranieri, Department of Veterinary Medicine, via dell'Università 6, 26900, University of Milan, Lodi, Italy. Email: angelica.stranieri@unimi.it

#### Abstract

SARS-CoV-2 has radically changed our lives causing hundreds of thousands of victims worldwide and influencing our lifestyle and habits. Feline infectious peritonitis (FIP) is a disease of felids caused by the feline coronaviruses (FCoV). FIP has been considered irremediably deadly until the last few years. Being one of the numerous coronaviruses that are well known in veterinary medicine, information on FCoV could be of interest and might give suggestions on pathogenic aspects of SARS-CoV-2 that are still unclear. The authors of this paper describe the most important aspects of FIP and COVID-19 and the similarities and differences between these important diseases. SARS-CoV-2 and FCoV are taxonomically distant viruses, and recombination events with other coronaviruses have been reported for FCoV and have been suggested for SARS-CoV-2. SARS-CoV-2 and FCoV differ in terms of some pathogenic, clinical and pathological features. However, some of the pathogenic and immunopathogenic events that are well known in cats FIP seem to be present also in people with COVID-19. Moreover, preventive measures currently recommended to prevent SARS-CoV-2 spreading have been shown to allow eradication of FIP in feline households. Finally, one of the most promising therapeutic compounds against FIP, GS-441524, is the active form of Remdesivir, which is being used as one therapeutic option for COVID-19.

#### KEYWORDS

COVID-19, feline coronavirus, feline infectious peritonitis, SARS-CoV-2

# 1 | INTRODUCTION

The current pandemic of coronavirus disease 2019 (COVID-19) is caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Zhou, Zhang, et al., 2020). COVID-19 is now inducing profound changes to our lifestyle and has severe consequences on healthcare systems and finances worldwide. Therefore, a huge research activity on this virus has been developed in the past few months to increase the knowledge on SARS-CoV-2 biology and pathology and to design new strategies for prevention and treatment of COVID-19 (Cyranoski, 2020a). Coronaviral diseases are well known in veterinary medicine, since numerous different species of coronaviruses (CoV) affect wild animals (primarily bats or avian species) as well as domestic species such as bovine, swine, feline and canine (Abdel-Moneim & Abdelwhab, 2020; Decaro & Lorusso, 2020; Su et al., 2016). Since the late 1990s (Paltrinieri et al., 1998), the research activity of our group has been focused on feline coronaviruses and especially on the diagnosis and pathogenesis of feline infectious peritonitis (FIP), a systemic and lethal disease of cats caused by the feline coronavirus (FCoV) (Pedersen, 2009). Therefore, with this review, we want to share our thoughts on the possible similarities and differences between FCoV infection and the preliminary information on SARS-CoV-2 biology and pathology published so far, as an attempt to provide possible points of discussion for future research on human coronaviruses.

# 2 | VIROLOGY

The SARS-CoV-2 belongs to the betacoronavirus genus and to the species Severe acute respiratory syndrome-related coronavirus (Gorbalenya et al., 2020). It is a completely new virus that is thought has originated from recombination events between CoVs of other species, possibly through one or more intermediate hosts. Bats are the most likely candidate reservoir of the SARS-CoV-2 (Andersen et al., 2020; Lau, et al., 2020; Letko et al., 2020; Xiaolu et al., 2020; Zhou, Chen, et al., 2020; Zhou, Yang, et al., 2020). The strong similarities between some pangolin coronaviruses and SARS-CoV-2 in the receptor-binding domain (RBD) suggested that also this species possibly played a role in the spillover to humans (Abdel-Moneim & Abdelwhab, 2020; Chen, et al., 2020; Dong et al., 2020; Lam et al., 2020; Malik et al., 2020; Xiao et al., 2020; Zhang, Wu, et al., 2020). More recently, other authors have reported that the sequence similarity in the spike RBD between SARS-CoV-2 and a sequence from pangolin is probably due to an ancient intergenomic introgression that occurred approximately 40 years ago (Chaw et al., 2020). Moreover, differences have been reported in the key residues of RBD and polybasic cleavage sites between bat and pangolin coronaviruses and SARS-CoV-2 from humans (Malaiyan et al., 2020) suggesting the need of further studies to help in the understanding of SARS-CoV-2 origin and intermediate transmission (Decaro & Lorusso, 2020; Liu, Jiang, et al., 2020; Malaiyan et al., 2020; Mavian, Pond, et al., 2020; Sironi et al., 2020; Tiwari et al., 2020; Wong et al., 2020). Studies are ongoing to identify the possible reservoir and intermediate hosts (Cyranoski, 2020b; Deng et al., 2020).

Recombination is a common behaviour for CoVs that may generate new variants able to infect species other than their natural reservoir (Holmes & Rambaut, 2004). Sequence analysis and phylogenetic studies allowed to identify the recombination events that led to the appearance of new CoV species in many animal species, and to identify bat coronaviruses as the ancestors of several CoVs of swine, ruminants, birds or rodents (Wong et al., 2019). All these animal species may be affected by different CoV species (Decaro & Lorusso, 2020). Conversely, only 2 types of FCoV, that belongs to the alphacoronavirus genus, are currently known: serotype I and serotype II FCoV. The ancestor of type I FCoV is unknown but alphacoronavirus are known to have likely originated from bats (MacLachlan & Dubovi, 2011). Type II FCoV originated by a double recombination between the FCoV type I and the canine coronavirus (CCoV) (Herrewegh et al., 1998). This recombination may have been one of the factors influencing the biology of the virus as type II FCoV, compared to FCoV type I, can easily grow in vitro, and it seems to be associated with a different cellular entry pathway. Indeed, types I and II appear to enter the cytosol through late and early endosomes, respectively (Takano et al., 2019). While the two serotypes show no differences in their pathogenicity, type I FCoV remains the most prevalent strain detected in field cases of FIP (Jaimes & Whittaker, 2018).

Despite recombination events contributed to the generation of new variants of FCoVs, the number of viral variants in cats is

uncary and Emerging

very limited, unlike in other animal species, where several species or strains of species-specific CoVs have been generated, mostly in recent years (Lin et al., 2017; Su et al., 2016). This may depend on the less frequent interaction of FCoVs of pet cats with CoVs of other animal species or on the social/behavioural peculiarities of wild cats that tend to have few interspecific interactions, although wild cats are more exposed to CoVs infecting rodents or birds due to their hunting activity. If this interpretation is correct, the probability of future spillover of CoVs may be reduced by prevention of interspecific interaction of domestic animals and consequently of interspecific exchange of potentially recombining CoVs. Moreover, due to the strict cohabitation between pet cats and people, especially in urban centres, cat infection with the SARS-CoV-2 should be prevented, also in order to avoid possible recombinations. Despite Gao, Bao, et al. (2020) recommended to guarantine pets at home as the better strategy for controlling the spread of SARS-CoV-2 to humans, the prevalence of infected pet cats seems to be very low to represent a threat for owners (Gao, Pan, et al., 2020; Patterson et al., 2020). At the moment, there is evidence that SARS-CoV-2 has infected cats in Wuhan, China, as recently showed by positive serological results in two cats (Chen, Huang, et al., 2020) and in a cohort of 15/102 examined cats using ELISA targeting the receptor-binding domain (RBD) of the virus as reported in a preprint of a research article posted online at bioRxiv (Zhang, Zhang, et al., 2020)). Moreover, SARS-CoV-2 RT-PCR-positive results have been recorded in two cats: one cat from Belgium (positive on stool and vomit) and in one cat from Hong Kong (positive on samples from oral and nasal cavities and rectum). Both cats belong to COVID-19-positive owners. Only the Belgian cat showed mild, respiratory and gastroenteric clinical signs, whereas the other cat was asymptomatic, as reported by the World Organisation for Animal Health (OIE, 2020). More recently, three cats were found positive in the United States, two with mild respiratory symptoms and one with fever, oral lesions and ulcerations on the tongue but only two belonging to a COVID-19positive owner (Newman et al., 2020; OIE, 2020). Two cats showing respiratory signs from owner previously suspected of being infected with COVID-19 in France also tested positive (OIE, 2020; Sailleau et al., 2020). Positive test results were also reported in one cat from Germany, one cat from Russia and two cats from Spain (OIE, 2020). Three cats have tested positive for SARS-CoV-2 in the Netherlands (Hossain et al., 2020). The route of transmission to cats from the Netherland is not known but interestingly these cats lived on a mink farm where minks were reported to be infected with SARS-CoV-2 (Enserink, 2020; Molenaar et al., 2020; Oreshkova et al., 2020). Moreover, lions and tigers have been affected, since cases were recorded in a zoo from New York (OIE, 2020; Wang, Mitchell, et al., 2020).

Nonetheless, a former report about the possible susceptibility of cats to the closely related SARS-CoV was published in the early 2000s, with presence of antibodies in asymptomatic infected cats (Martina et al., 2003).

Despite the low frequency of recombination with other CoVs, the FCoV has a high variability in the feline population. The frequency of

VII FY— Transboundary and Emerging Diseases

mutations, especially in some regions of FCoV RNA, is very high and the high replication rate of FCoV in the intestine of affected cats leads to the generation of 'quasispecies' in each single cat (Battilani et al., 2003). Some of these new variants may bear mutations that, if coupled with a peculiar immune response of infected cats, are likely to play a key role in the pathogenesis of FIP, as specified below (Pedersen, 2014a).

Several variations have been reported also in SARS-CoV-2, some of the being non-synonymous mutations, and reported mutations may affect SARS-CoV-2 virulence, infectivity, and transmissibility (Chaw et al., 2020; Kim et al., 2020; Li, Yang et al., 2020; Tiwari & Mishra, 2020; van Dorp et al., 2020). Based on the high-frequency mutations, SARS-CoV-2 genomes have been classified into different groups (Baay et al., 2020; Wang, Li, et al., 2020).

It is important to keep in mind that information on genetic heterogeneity of SARS-CoV-2 strains is considered for now not conclusive, but only preliminary and hypothesis-generating (Mavian, Marini, et al., 2020). However, the hypothesis of the presence of strains with different virulence (Armengaud et al., 2020; Lau, Wang, et al., 2020) may justify, along with other epidemiological factors, the variability of clinical signs of COVID-19, that span from mild flu-like symptoms to severe and lethal pneumonia. Further investigations on the genetic diversity of SARS-CoV-2 populations are warranted to support this hypothesis (Chen, Zhou, et al., 2020; Guo et al., 2020; Zhang, Yang, et al., 2020).

#### 3 | EPIDEMIOLOGY

Both the SARS-CoV-2 and the FCoV are highly contagious and rapidly spread within susceptible populations (Sanche et al., 2020). This behaviour is typical of many animal CoVs (Cavanagh, 2007; Guy, 2000; Licitra et al., 2014; Song & Park, 2012). When CoVs are introduced into a new population, the infection may show different epidemiological patterns. CoV infection may show an epidemic pattern, as reported with SARS-CoV and currently with SARS-CoV-2 (Petersen et al., 2020). On the other hand, it may become endemic, with a high number of infected asymptomatic individuals and a lower mortality rate, as observed in at least four common human coronaviruses (hCoVs-229E, -NL63, -OC43, and -HKU1) and for almost all the coronaviruses of bats (Corman et al., 2018; Roussel et al., 2020; Vijaykrishna et al., 2007).

An endemic pattern is also observed in cats and FIP. The disease was discovered in the 60s in the United States (Holzworth, 1963; Wolfe & Griesemer, 1966) and afterwards likely spread from the United States to other countries, when single reports on FIP cases diagnosed in different countries appeared in scientific journals starting from the late 60s-early 70s (Lauzi et al., 2020). In the following years, the number of FIP cases and of FCoV-infected cats has been increasing worldwide as such, currently, the rate of FCoV RT-PCR-positive and/or seropositive cats in multi-cat environment often approaches 100%. The infection rate has been reported to increase proportionally with the number of cats per cattery (Paltrinieri et al., 2014; Pedersen, 2009). This high rate of infection, in turn, depends on the only partially protective immunity that cats may mount at the intestinal level. Partial protective immunity allows cats to periodically clear the infection but, once the local immunity decreases, cats can be re-infected if living in FCoV-endemic environments. Therefore, infection develops according to the SIS (susceptible-infected-susceptible) model and most of the cats in FCoV-endemic catteries are recurrent shedders of the virus (Foley et al., 1997), thus contributing to maintain the infection in the environment. Despite this high rate of infection, mortality remains low also due to the peculiar immunopathogenesis of FIP described below (Pedersen, 2014a).

Based on the information on the diffusion of COVID-19 from its origin in the Hubei province (China) to the current worldwide distribution, the spread of the SARS-CoV-2 is now in the epidemic/pandemic phase. The efforts of Public Health Authorities are focused on slowing down the infection (Bruinen de Bruin et al., 2020). It is unlikely that the virus will be eradicated and the infection may enter the endemic phase. In this scenario the risk of additional foci of epidemic infection (as it occurs in cats when FCoVs enter in catteries with low endemicity) may be present, as well as the risk of future mutations of the virus that may modify its virulence.

#### 4 | PATHOGENESIS AND PATHOLOGY

Although virological and epidemiological aspects of FCoV and SARS-CoV-2 infection have some common features, the pathogenesis of the disease seems to be different. This difference starts at the level of cell entry with the SARS-CoV-2 binding the angiotensinconverting enzyme 2 (ACE2) receptor in humans (South et al., 2020). Predicted homology of feline and human ACE2 receptors (Guo et al., 2008) along with the identification of the same ACE-2 receptor in cats and dogs (Luan et al., 2020) confirms that SARS-CoV-2 infection may occur also in these domestic animals. FCoVs binds receptors different than the ACE-2 receptor in cats. Type II FCoVs employ feline aminopeptidase N (fAPN) as a cellular receptor whereas the receptor used by type I FCoVs is still unknown. However, cell membrane lectins (C-type lectin dendritic cell-specific intercellular adhesion molecule-3-grabbing nonintegrin, fDC-SIGN) seem to play a role in the cell entry for both FCoV serotypes (Regan et al., 2010; Van Hamme et al., 2011).

Moreover, the cellular tropism of the two viruses seems to be partly different. FCoV is a systemic disease affecting different organs, whereas SARS-CoV-2 apparently affects mainly the lungs (Wang, Wang, et al., 2020). However, results of RT-PCR on stool samples suggest that oral-faecal route may be an additional mode of infection for SARS-CoV-2 (Cipriano et al., 2020; He et al., 2020). Gastrointestinal involvement was also a characteristic of SARS-CoV that shares the same receptor of SARS-CoV-2 for cellular entry. Indeed, gastrointestinal involvement may be explained by the presence of ACE-2 receptor which are highly expressed in lung epithelial cells as well as in enterocytes from ileum and colon (Zhang, Kang, et al., 2020). More recently, SARS-CoV-2 was demonstrated to also

incary and Emerging Diseases

been able to actively replicate in human enterocytes in vitro (Lamers et al., 2020).

Curiously, enterocytes are the major target of FCoV, although the virulent FCoV strains infect macrophages also. Both type I and type II FCoV, in fact, have 2 distinct pathotypes: one with enteric tropism, formerly known as feline enteric coronavirus (FECV), which may occasionally induce mild gastrointestinal symptoms, and the other, formerly known as feline infectious peritonitis virus (FIPV), which is more able to replicate within macrophages, thus disseminating in the host and inducing FIP.

Feline infectious peritonitis is a systemic disease characterized by granulomatous lesions ('dry' or non-effusive FIP) and/or by vasculitis that induce the development of cavitary effusions ('wet' or effusive FIP). The two pathotypes were for long time considered as distinct viral species, but molecular studies demonstrated that they are two variants of the same virus, with different virulence (Pedersen, 2014a). In turn, the different virulence has been thought to depend on mutations of intestinal strains. However, a single mutation definitely responsible for FIP has not been identified yet, although several candidate genes have been supposed to be involved in this virulence shift (Tekes & Thiel, 2016). Recent studies have identified mutations in the FCoV spike protein as markers of systemic spread of FCoV, regardless of virulence (Barker et al., 2017; Porter et al., 2014).

Differences in the nucleotide and amino acid sequence have been reported in the SARS-CoV-2 viral receptor-binding domain (RBD) compared to SARS-CoV. These mutations seem to be associated with an enhanced affinity of the spike protein of SARS-CoV-2 to the ACE-2 receptor, that may explain the higher morbidity of SARS-CoV-2 (Ou et al., 2020; Singh, 2020). The possible role of mutated variants of the SARS-CoV-2 in the pathogenesis of clinical diseases characterized by more severe clinical signs has not yet been fully exploited or elucidated.

While the mutation to the virulent pathotype is the first key event, the second key event in the pathogenesis of FIP is the activation of the immune response. Cats that seem to be more resistant to the infection (i.e. cats that harbour the FCoV in their intestine for months or the recurrent shedders mentioned above) have a very efficient cell-mediated (Th1) immune response. Conversely, the activation of the humoral (Th2) immune response may exacerbate the course of the disease. Antibodies have been shown to accelerate, in vitro, the uptake of the virus by macrophages, according to the mechanism known as 'antibody-dependent enhancement' (ADE), where anti-FIPV antibodies increase the uptake of FIPV through the macrophage Fc receptor (Olsen et al., 1993). However, the role of ADE was controversial until recently. The first studies on naturally infected cats with repeated exposure to FCoV did not suggest that the presence of antibodies increases the risk of FIP and, at the same time, the high antibody titres of cats that cleared the infection suggested a protective role of antibodies, at least in some cases (Addie et al., 1995). Recently, the role of ADE in disease mechanisms has been confirmed. ADE, in fact, has been documented in experimentally infected cats that were immunized against FIPV and developed FIP, while non-immunized cats did not (Takano et al., 2019).

Interestingly, ADE stimulated by other CoVs is thought to exist for the SARS-CoV-2 and be responsible for the more severe cases recorded in a lower percentage of infected patients, usually among the elderly (Schatzmann Peron & Nakaya, 2020). In particular, this phenomenon could be consequence of the exposure not to SARS-CoV, but to other coronaviruses that cause only mild symptoms and are mistaken for common cold viruses, but allow the host to mount an antibody response (Tetro, 2020). The antibody-dependent infection of macrophages could represent, in fact, a pivotal step towards disease progression from mild to severe symptoms, and may as well explain the dysregulated immune responses in COVID-19, characterized by T-cell lymphopenia and pro-inflammatory cascade with macrophage hyperactivation, both important known phenomena in FIP (Pedersen, 2014a; Tay et al., 2020). Moreover, progression to the critical phase of the disease often coincides with the beginning of humoral immunity antibody response (Ricke & Malone, 2020). However, even though coronaviruses such as MERS-CoV and FCoV have been demonstrated to actively infect monocytes and macrophages, this needs to be more thoroughly investigated for SARS-CoV-2, also at the light of the fact that ACE-2 is expressed on alveolar macrophages and SARS-CoV-2 can replicate in these latter cells (Chu et al., 2020; Kai & Kai, 2020; Pedersen, 2014a; Ricke & Malone, 2020; Schatzmann Peron & Nakaya, 2020). Moreover, the role of CD169<sup>+</sup> macrophages in the systemic spreading of SARS-CoV-2 as well as in the excessive inflammation happening during SARS-CoV-2 infection has been questioned and needs to be further investigate (Merad & Martin, 2020; Park, 2020). Despite interesting results, the real role of ADE in naturally infected cats is still questioned and data regarding ADE in SARS-CoV-2 infection are still too speculative.

Regardless of the ADE mechanism, the role of antibodies in the pathogenesis of FIP has been previously reported by several studies. Indeed, antibodies are known to be involved in the pathogenesis of wet FIP. Intracavitary effusions observed in wet FIP are the consequences of a vasculitis based on the contribution of a type III hypersensitivity reaction, on which immunocomplexes formed by the FCoV and by anti-FCoV antibodies precipitate around the vessel's walls and induce the recruitment of macrophages producing factors. These cytokines damage the tissues and, at the same time, induce the release of neutrophils that intensify the inflammatory process and exacerbate the tissue damage (Acar et al., 2016; Berg et al., 2005; Pedersen, 2009). It is still unclear why only some seropositive cats develop the immune complex disease, while other seropositive cats living in the same cattery do not. Recent studies have observed that serum and tissue patterns of molecules and cells involved in the innate and specific immune response differ in seropositive clinically healthy cats compared with cats with FIP, suggesting that cats may develop or not FIP depending on the type of activation of these responses (Pedersen, 2014a). Some studies suggested that specific sequences of structural proteins of the FCoV may influence the type of immune response mounted by the cat. Therefore, it may be supposed that mutations of the virus may also play a role in the host-virus interactions (Rossi et al., 2011; Satoh et al., 2010; Takano et al., 2014).

The information collected in the few months regarding COVID-19 immune pathogenesis from the first documented cases seems to exclude a direct role of the host's immune response in the development of COVID-19 lesions. Firstly, the median incubation time of the disease (four days in a report of 1,099 cases) is lower than the time required to activate the cells involved in the immune response and the production of antibodies (Guan et al., 2020). Secondly, the administration of neutralizing antibodies containing plasma from patients recovered from COVID-19 seems to be a useful additional therapeutic support for critically ill patients, suggesting that antibodies might have a protective effect against the virus rather than contributing to the development of the disease (Shen et al., 2020). Thirdly, although mononuclear infiltrates composed by lymphocytes, monocytes and macrophages may be found within COVID-19 lesions, the interstitial pneumonia reported in this disease as well as the presence of syncytial multinucleated cells and pneumocytes showing cytopathic lesions (Felsenstein et al., 2019; Liu, Chen, et al., 2020; Xu et al., 2020) seems to be directly induced by the replication of the virus within the cells. In particular, the virus infects primarily ACE-2 expressing cells, which are mainly present in the lungs and were demonstrated to be the primary target of the SARS-CoV, with which SARS-CoV-2 share several pathogenic aspects (He et al., 2006; Sarzi-Puttini et al., 2020).

In contrast, FIP lesions are typically characterized by lymphoplasmocytic infiltrates, admixed with activated macrophages and neutrophils and are centred around vessels that show the fibrinoid necrosis typical of immune complex vasculitis (Kipar et al., 2005; Pedersen, 2009). Conversely, in one report only immunostaining for SARS-CoV-2 in lung lesions showed minimal viral protein expression on vessels (Zhang, Zhou, et al., 2020). However, studies on the possible involvement of the immune system in the development of lesions induced by the SARS-CoV-2 are still scarce. These aspects need to be further exploited through additional studies, also focusing on the possible distribution of viral antigens on tissues other than the lung, or at the light of the recent rising concern regarding atypical COVID-19 manifestations in children. Specifically, multisystem inflammatory syndrome in children (MIS-C) is a rare but devastating consequences of SARS-CoV-2 infection in paediatric patients and it resembles some aspects of Kawasaki disease, with the overexpression of pro-inflammatory cytokines and the production of autoantibodies targeting primarily the heart and blood vessels (Cavouinidis et al., 2020; Mahase, 2020). In fact, if SARS-CoV and SARS-CoV-2 share similar behaviours, the hyperproduction of inflammatory cytokines from infected cells is probably responsible both for the more severe cases and for the multiorgan involvement (Li, Geng, et al., 2020).

Despite the pathogenic differences listed above, the severe acute systemic inflammatory reaction syndrome (SIRS) is common in COVID-19 and in FIP. As above stated, cats may harbour the FCoV without showing clinical signs for years, but when FIP

develops the activation of the innate immune response is rapid. The immune response leads to a pro-inflammatory cytokines overproduction responsible of severe clinical and laboratory changes (Kipar & Meli, 2014). A 'cytokine storm' similar to that reported in FIP cases has been reported also in patients with COVID-19 (Dhama et al., 2020; Kipar et al., 2006; Pedersen & Ho, 2020; Sarzi-Puttini et al., 2020). In turn, this cytokine storm may induce a multiorgan failure that is responsible for the high mortality rate of critically ill patients with COVID-19, This hypothesis is supported by the promising results of immunotherapies with anti-cytokine drugs, especially when using TNF- $\alpha$  blockers (Russell et al., 2020; Sarzi-Puttini et al., 2020). This type of treatment has never been investigated in cats with FIP. mostly due to cost reasons or to the unavailability of drugs registered for the cat. However, the good clinical response of cats to steroidal anti-inflammatory drugs (Pedersen, 2014b) supports the hypothesis that suppression of the hyperinflammatory response may temporarily improve the clinical condition. This is not completely curative in FIP, due to its peculiar immunopathogenic mechanisms, but it may provide COVID-19 patients with precious time for activating the antiviral immune reaction or, hopefully, to enhance the effect of antiviral drugs.

# 5 | CLINICAL PRESENTATION AND DIAGNOSIS

As mentioned above, FCoV may induce a broad spectrum of clinical and laboratory changes, ranging from a mild gastrointestinal disease without peculiar laboratory abnormalities induced by the 'enteric strains' (FECV) to a severe and invariably lethal systemic disease (FIP) induced by the mutation into 'virulent strains' (FIPV) coupled with the peculiar immune response of infected cats mentioned above. FIP may have two main clinical presentations, sometimes overlapped to each other: the wet and the dry form. The wet form is basically characterized by the accumulation of protein-rich fibrinous fluids in body cavities, leading to symptoms consistent with the severe and acute hypovolemia and/or to compression of organs within the affected cavity (e.g. dyspnoea, decreased peristalsis, etc). The clinical presentation of the dry form depends on the main site of formation of granulomatous lesions: these may occur primarily on the kidney, leading to clinical and laboratory changes consistent with renal dysfunction, but extra-renal symptoms may occur if granulomatous lesions are prevalent in other organs such as liver, lung, intestine, but especially in the eye or in the central nervous system (Pedersen, 2009). In both the clinical forms, systemic signs due to the SIRS mentioned above such as fever, depression, asthenia, emaciation as well laboratory changes such as anaemia, lymphopenia, increased serum concentration of globulin fractions and acute phase proteins are commonly found (Pedersen, 2014b; Stranieri et al., 2018).

Conversely, the most frequent clinical presentation of COVID-19 deals with a severe acute respiratory distress associated with interstitial pneumonia, although neurological and hepatic manifestations have been described in SARS-CoV-2-infected patients and, most importantly, there is growing evidence of gastrointestinal tract involvement (Li, Bai, et al., 2020; Tian et al., 2020). In several COVID-19 patients, gastrointestinal symptoms have been reported, often before the beginning of respiratory symptoms (Gu et al., 2020). These clinical changes are associated with fever and fatigue, and with laboratory changes such as lymphopenia and increased serum concentration of acute phase proteins (Borges do Nascimento et al., 2020) likely depending on the 'cytokine storm' mentioned above.

The different biology and the pathogenesis of the virus influences also the diagnostic approach to the disease. In particular, the immunopathogenesis of FIP also affects the possibility to correctly diagnose the disease. No molecular or serological tests are able to differentiate virulent and non-virulent strains of FIP (i.e. the former FIPV and FECV). Therefore, RT-PCR or serology may confirm the infection but not the disease. FIP may be erroneously diagnosed in RT-PCR-positive or seropositive cats with non-specific clinical signs such as fever or weight loss due to diseases other than FIP. Conversely, cats with FIP may become RT-PCR negative or seronegative when immune complexes develop and precipitate in tissues (Meli et al., 2013; Pedersen, 1976). Ultimately, only the detection of the virus in the effusion or within the lesions may confirm the disease, although several clinical or clinico-pathological changes may be highly suggestive of FIP (Felten & Hartmann, 2019; Stranieri et al., 2018; Tasker, 2018). All these obstacles do not seem to occur in patients affected by COVID-19 and, although asymptomatic SARS-CoV-2-positive patients may be frequently found (Shi et al., 2020), the diagnosis may be based on RT-PCR-positive swabs in patients with clinical signs or with chest diagnostic imaging findings consistent with the disease.

### 6 | MANAGEMENT AND PREVENTION

One anti-FIP/anti-FCoV vaccine has been developed and commercialized, but it is available only in few countries (Fehr et al., 1997). It must be administered to kittens of 16 weeks of age or older, which is several weeks after the maternal protective antibodies decrease, leaving a wide-open time window for FIP manifesting (Addie, 2019). Moreover, the risk of stimulating an excessive antibodies production that, based on the aforementioned ADE occurrence, may induce the disease rather than preventing it, is still debated (Bálint et al., 2014) and according to the World Small Animal Veterinary Association the vaccine against FIP is defined as not recommended (Day et al., 2016).

Several research institutions or pharmaceutical companies are currently working in the development of a vaccine for the prevention of SARS-CoV-2 infection and around 200 candidates' vaccines are at different stages of trial. Specifically, as of August 13, 2020, three vaccines are in the last phase of clinical development: two inactivated vaccines developed by the Chinese companies Sinopharm and Sinovac and a sequence-optimized mRNA encoding prefusion-stabilized SARS-CoV-2 S-2P protein (mRNA-1273) by the Kaiser Permanente Washington Health Research Institute (Chugh, 2020; Corbett et al., 2020; Gao, Bao, et al., 2020). Constant updates on boundary and Emerging Diseases

vaccines development can be found at https://www.raps.org/newsand-articles/news-articles/2020/3/covid-19-vaccine-tracker.

Based on what above stated, the genetic diversity of SARS-CoV-2 strains seems to be lower than that of the FCoV and the humoral immune response seems only partially involved in the pathogenesis of COVID-19. Therefore, vaccination could be a promising preventive tool for this disease, even if the question of whether a certain vaccination regimen could induce long-term protection has been addressed for animal and human coronaviruses (Saif, 2020).

In the absence of an effective vaccine, the major preventive measure adopted worldwide to contain and possibly eradicate the SARS-CoV-2 infection is based on quarantine/isolation of poorly symptomatic infected patients, on confinement measures that limit the circulation of people and on social distancing. Although profoundly affecting our habits, our social life and having a great impact on global economy, so far the application of these measured strongly reduced both the rate of infection and mortality in China and in several countries worldwide.

Such an approach has been already recommended in cats. Years ago, prevention strategies based both on the isolation of shedders until repeated RT-PCR tests on their faeces resulted persistently negative and on the early weaning of kittens born from seropositive or PCR-positive queens have been recommended (Addie & Jarrett, 2001). Although these approaches have been proven to be successful on the management of infection in single catteries, their efficiency has been biased by the lack of common rules imposed to all the cat owners by regulatory bodies (e.g. breed associations, veterinary health authorities). Breeders or shelters managers are usually not willing to apply these strategies. Indeed, the participation to the expected activities of breeding catteries (e.g. mating, participation to cat shows,) or shelters (reintroduction of guarantined cats in larger groups of non-tested animals) inevitably exposes cats to the risk of reinfection and ultimately to the risk of FIP, making almost useless the efforts aimed to make their cats FCoV-negative. This experience reinforces the concept that isolation/quarantine strategies must be applied on a large scale, as Public Authorities are doing in many Countries for COVID-19 and that the application of strict measures only to single districts or cities could become ineffective with time.

# 7 | TREATMENT

Although FIP has been historically considered an invariably lethal disease and no successful treatments have been available for decades, effective treatments were recently developed with promising results. Historically, cats with FIP received only supportive or anti-inflammatory treatment, that often ameliorated the quality of life as stated in anecdotal reports, but without clearing from the infection neither stopping the immunopathogenesis of the disease (Addie et al., 2009). This is actually what happens also in COVID-19, where most of the therapeutic efforts to manage sick patients, especially in intensive care units, are based on the

# TABLE 1 Comparison between FCoV/SARS-CoV-2 infection and FIP/COVID diseases

		FCoV/FIP	SARS-CoV-2/COVID-19
Virology	Genus	Alphacoronavirus	Betacoronavirus
	Presence of serotypes/ clades/strains	Yes (serotype I and II)	Yes (research ongoing)
	Origin	Serotype I: unknown (likely bat origin of alphacoronaviruses)	Suggested spillover from other species (bats, pangolins)
		Serotype II: recombination FCoV I/ CCoV	
	Cell Receptor	Serotype I: Unknown (possibly fDC-SIGN)	ACE2 receptor
		Serotype II: fAPN (possibly also fDC-SIGN)	
	Frequency of mutation	High (quasispecies)	Mutations reported, research ongoing (suggested groups with different virulence)
Epidemiology	Transmissibility	High	High
	Epidemiologic pattern	Epidemic -> endemic	Epidemic (up to now)
	Reinfections	Frequent	Rarely reported (up to now)
	Model of infection	SIS (susceptible-infected-susceptible)	Unknown (up to now)
	Lethality	Epidemic phase: high; Endemic phase: low	Epidemic phase: high
Pathogenesis	Route of infection	Faecal-Oral	Respiratory (oral not excluded)
	Cellular tropism	Enterocytes, monocytes/macrophages	Alveolar macrophages, enterocytes
	Role of mutated viral variants	Probable	Postulated
	Main target organs	FECV: intestine, FIPV: multiple organs/ tissues	Lung (less frequently GI tract or other organs)
	Lesions	Granulomatous lesions, vasculitis and effusions, lymphoplasmocytic infiltrates	Cytopathic effect on lung cells, multinucleated syncytial cells, mononuclear infiltrates
Immunopathogenesis	Type III hypersensitivity	Demonstrated	Postulated
	T-cell lymphopenia	Demonstrated	Demonstrated
	ADE	Hypothesized	Hypothesized
	Cytokine storm/SIRS	Demonstrated	Demonstrated
Prevention	Vaccination	Available but not recommended (risk of ADE)	Not available
	Quarantine/isolation	May eradicate the disease from catteries	May reduce the prevalence of infection/disease
Treatment	Symptomatic drugs	Effective as a support therapy	Effective, curative in mild forms
	Anti-inflammatory drugs	Effective as a support therapy	Possibly curative
	Anti-cytokine drugs	Not tested	Effective, curative in mild forms
	Hyper-immune plasma	Not tested	Possibly curative
	Interferon or Th1 modulators	Rarely effective	Studies ongoing
	Antiviral drugs	Effective in a few clinical trials (GS- 441524, GS-5734, Xraphconn)	Possibly effective (GS-5734)

Abbreviations: ACE2, angiotensin-converting enzyme 2; ADE, antibody-dependent enhancement; CCoV, canine coronavirus; fAPN, feline aminopeptidase N; FCoV, feline coronavirus; fDC-SIGN, C-type lectin dendritic cell-specific intercellular adhesion molecule-3-grabbing nonintegrin; SIRS, systemic inflammatory response syndrome.

management of the acute respiratory distress or on the modulation of the inflammatory reactions, as above mentioned (Alhazzani et al., 2020). The stimulation of cell-mediated immunity on FIP cats has been proposed in the past and is still one of the most used therapeutic approach. For this purpose, interferon- $\alpha$  or feline recombinant interferon- $\omega$ , that

possess also antiviral properties, are used, even though the efficacy of these treatments is controversial (Pedersen, 2014b; Hartmann & Ritz, 2008) as it is for polyprenyl immunostimulant (Pedersen, 2014b). The use of interferon for COVID-19 has been questioned mainly for its antiviral activity, and, to the authors' knowledge, trials regarding its use in association with other molecules (i.e. antiviral agent ribavirin) are currently ongoing (Lu et al., 2020).

The most relevant changes on the therapeutic approach to FIP cats occurred over the past few years. The peptidomimetic GC-376 and the nucleoside analogue GS-441524, both able to inhibits FCoV replication in different manners, became available and were tested in vitro and in experimental infections (Kim et al., 2016; Murphy et al., 2018), or in cats naturally affected by FIP (Pedersen et al., 2018, 2019). More specifically, GC-376 caused disease remission in 35% of the treated cats and it appeared more efficient towards certain clinical presentations of FIP (Pedersen et al., 2018). On the other hand, GS-441524 strongly reduced the viral burden in infected cats and induced the remission of clinical signs after one or more cycles of treatment in the large majority (96.1%) of cats. Despite GS-441524 is not registered for use in cats in many countries, this treatment is now widely used by cat owners and breeders that can buy the drug through online distributors. Despite, the use of unlicensed drugs must be abandoned (Letter from Dr. Pedersen, https://sockf ip.org/), several anecdotal reports on successfully treated cats are available on online blogs. Interestingly, the GS-441524 is the biologically active component of the phosphoramidate prodrug GS-5734 (Remdesivir) that has been tested, with some promising results, also in patients with COVID-19 since its efficacy was previously demonstrated against Ebola and Nipah viruses infections (Cao et al., 2020; Ledford, 2020). Recently, Mutian® Xraphconn (Mutian X), an orally administered drug containing inotodiol, an anti-inflammatory sterol of fungal origin, has been shown to completely and rapidly clear the virus from the intestine of FCoV-infected cats likely by reducing viral replication (Addie et al., 2020). Anecdotal reports on therapeutic successes of this drug in cats with FIP are also published on online blogs, despite being an unlicensed drug as the other above mentioned. Clinical studies aimed to evaluate the effectiveness of Remdesivir treatment on COVID-19 patients are currently ongoing (Nili et al., 2020), and it would be interesting to consider how the bioavailability of the drug in the lung of COVID-19 patients may differ from that of the intestine or other tissues of cats with FIP.

# 8 | CONCLUSIVE REMARKS

The FCoV and the SARS-CoV-2 share some common features, such as the rapid spread of the infection within the population, the reduction of the infection rate by isolation of infected patients or the shared success of similar anti-inflammatory or antiviral compounds. However, FCoV and SARS-CoV-2 differ also in terms of biology of the virus, target cells, pathogenesis and clinical features. Differences and similarities of the two infections are summarized in Table 1. Nevertheless, years of studies on FCoV-infected cats demonstrate oundary and Emerging Dise

that increasing the knowledge on virus biology and host-virus interactions may improve the chances to contain and, eventually, combat the infection. Moreover, the information gained so far on the aspects of FCoV infection shared with SARS-CoV-2 may serve as a basis for a rapid development of prevention or therapeutic strategies for COVID-19 as well as for studies on the possible interaction between FCoV and SARS-CoV-2 that may occur due to the strict relationship between people and their pet cats.

#### CONFLICT OF INTEREST

The authors declare that there are no conflicts of interests.

#### ETHICAL STATEMENT

Not applicable.

#### DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analysed in this study.

#### ORCID

Angelica Stranieri ២ https://orcid.org/0000-0002-8529-572X

#### REFERENCES

- Abdel-Moneim, A. S., & Abdelwhab, E. M. (2020). Evidence for SARS-CoV-2 infection of animal hosts. *Pathogens*, 30, 529. https://doi. org/10.3390/pathogens9070529
- Acar, D. D., Olyslaegers, D. A., Dedeurwaerder, A., Roukaerts, I. D., Baetens, W., Van Bockstael, S., De Gryse, G. M., Desmarets, L. M., & Nauwynck, H. J. (2016). Upregulation of endothelial cell adhesion molecules characterizes veins close to granulomatous infiltrates in the renal cortex of cats with feline infectious peritonitis and is indirectly triggered by feline infectious peritonitis virus-infected monocytes in vitro. *Journal of General Virology*, *97*, 2633–2642. https://doi. org/10.1099/jgv.0.000585
- Addie, D. D. (2019). Feline infectious peritonitis: Answers to frequently asked questions concerning FIP and coronavirus. *Veterinary Nursing Journal*, 34, 201–206. https://doi.org/10.1080/17415 349.2019.1629366
- Addie, D., Belák, S., Boucraut-Baralon, C., Egberink, H., Frymus, T., Gruffydd-Jones, T., Hartmann, K., Hosie, M. J., Lloret, A., Lutz, H., Marsilio, F., Pennisi, M. G., Radford, A. D., Thiry, E., Truyen, U., & Horzinek, M. C. (2009). Feline infectious peritonitis. ABCD guidelines on prevention and management. *Journal of Feline Medicine & Surgery*, 11, 594–604. https://doi.org/10.1016/j.jfms.2009.05.008
- Addie, D. D., Curran, S., Bellini, F., Crowe, B., Sheehan, E., Ukrainchuk, L., & Decaro, N. (2020). Oral Mutian®X stopped faecal feline coronavirus shedding by naturally infected cats. *Research in Veterinary Science*, 130, 222–229. https://doi.org/10.1016/j.rvsc.2020.02.012
- Addie, D. D., & Jarrett, O. (2001). Use of a reverse-transcriptase polymerase chain reaction for monitoring the shedding of feline coronavirus by healthy cats. *Veterinary Record*, 148, 649–653. https://doi. org/10.1136/vr.148.21.649
- Addie, D. D., Toth, S., Murray, G. D., & Jarrett, O. (1995). Risk of feline infectious peritonitis in cats naturally infected with feline coronavirus. *American Journal of Veterinary Research*, 56, 429–434.
- Alhazzani, W., Møller, M. H., Arabi, Y. M., Loeb, M., Gong, M. N., Fan, E., Oczkowski, S., Levy, M. M., Derde, L., Dzierba, A., Du, B., Aboodi, M., Wunsch, H., Cecconi, M., Koh, Y., Chertow, D. S., Maitland, K., Alshamsi, F., Belley-Cote, E., ... Rhodes, A. (2020). Surviving sepsis

-WILEY- Transboundary and Emerging Diseases

campaign: Guidelines on the management of critically ill adults with Coronavirus Disease 2019 (COVID-19). *Intensive Care Medicine*, 46(5), 854–887. https://doi.org/10.1007/s00134-020-06022-5

- Andersen, K. G., Rambaut, A., Lipkin, W. I., Holmes, E. C., & Garry, R.
   F. (2020). The proximal origin of SARS-CoV-2. *Nature Medicine*, 26, 450–452. https://doi.org/10.1038/s41591-020-0820-9
- Armengaud, J., Delaunay-Moisan, A., Thuret, J. Y., Van Anken, E., Acosta-Alvear, D., Aragón, T., Arias, C., Blondel, M., Braakman, I., Collet, J. F., & Courcol, R. (2020). The importance of naturally attenuated SARS-CoV-2 in the fight against Covid-19. *Environmental Microbiology*, 22, 1997–2000. https://doi.org/10.1111/1462-2920.15039
- Baay, M., Lina, B., Fontanet, A., Marchant, A., Saville, M., Sabot, P., Duclos, P., Vandeputte, J., & Neels, P. (2020). SARS-CoV-2: Virology, epidemiology, immunology and vaccine development. *Biologicals*, 2, 35–40. https://doi.org/10.1016/j.biologicals.2020.06.005
- Bálint, Á., Farsang, A., Szeredi, L., Zádori, Z., & Belák, S. (2014). Recombinant feline coronaviruses as vaccine candidates confer protection in SPF but not in conventional cats. *Veterinary Microbiology*, 169, 154–162. https://doi.org/10.1016/j.vetmic.2013.10.015
- Barker, E. N., Stranieri, A., Helps, C. R., Porter, E. L., Davidson, A. D., Day, M. J., Knowles, T., Kipar, A., & Tasker, S. (2017). Limitations of using feline coronavirus spike protein gene mutations to diagnose feline infectious peritonitis. *Veterinary Research*, 48, 60–74. https://doi. org/10.1186/s13567-017-0467-9
- Battilani, M., Coradin, T., Scagliarini, A., Ciulli, S., Ostanello, F., Prosperi, S., & Morganti, L. (2003). Quasispecies composition and phylogenetic analysis of feline coronaviruses (FCoVs) in naturally infected cats. FEMS Immunology & Medical Microbiology, 39, 141–147. https:// doi.org/10.1016/S0928-8244(03)00237-2
- Berg, A. L., Ekman, K., Belak, S., & Berg, M. (2005). Cellular composition and interferon-γ expression of the local inflammatory response in feline infectious peritonitis (FIP). *Veterinary Microbiology*, 111, 15–23. https://doi.org/10.1016/j.vetmic.2005.07.017
- Borges do Nascimento I.J, Cacic N, Abdulazeem H. M, von Groote T. C, Jayarajah U, Weerasekara I, Esfahani M. A, Civile V. T, Marusic A, Jeroncic A, Carvas Junior N, Pericic T. P, Zakarija-Grkovic I, Meirelles Guimarães S. M, Luigi Bragazzi N, Bjorklund M, Sofi-Mahmudi A, Altujjar M, Tian M, ... Marcolino M. S (2020). Novel Coronavirus Infection (COVID-19) in Humans: A Scoping Review and Meta-Analysis. Journal of Clinical Medicine, 9, (4), 941. http://dx.doi. org/10.3390/jcm9040941
- Bruinen de Bruin, Y., Lequarre, A. S., McCourt, J., Clevestig, P., Pigazzani, F., Zare Jeddi, M., Colosio, C., & Goulart, M. (2020). Initial impacts of global risk mitigation measures taken during the combatting of the COVID-19 pandemic. *Safety Science*, 128, 104773. https://doi. org/10.1016/j.ssci.2020.104773
- Cao, Y. C., Deng, Q. X., & Dai, S. X. (2020). Remdesivir for severe acute respiratory syndrome coronavirus 2 causing COVID-19: An evaluation of the evidence. *Travel Medicine and Infectious Disease*, 35, 101647. https://doi.org/10.1016/j.tmaid.2020.101647
- Cavanagh, D. (2007). Coronavirus avian infectious bronchitis virus. Veterinary Research, 38, 281–297. https://doi.org/10.1051/vetres:2006055
- Cavounidis, A., Alderson, J., & Quastel, M. (2020). Multisystem inflammatory syndrome in children: Getting to the heart of the matter. *Nature Reviews Immunology*, 20(9), 520. https://doi.org/10.1038/ s41577-020-0409-z
- Chaw, S. M., Tai, J. H., Chen, S. L., Hsieh, C. H., Chang, S. Y., Yeh, S. H., Yang, W. S., Chen, P. J., & Wang, H. Y. (2020). The origin and underlying driving forces of the SARS-CoV-2 outbreak. *Journal of Biomedical Science*, 27, 1–12. https://doi.org/10.1186/s12929-020-00665-8
- Chen, J., Huang, C., Zhang, Y., Zhang, S., & Jin, M. (2020). Severe acute respiratory syndrome coronavirus 2-specific antibodies in pets in Wuhan, China. *Journal of Infection*, 81(3), e68–e69. https://doi. org/10.1016/j.jinf.2020.06.045

- Chen, N., Li, X., Li, S., Xiao, Y., Ye, M., Yan, X., & Li, X. (2020). How related is SARS-CoV-2 to other coronaviruses? *Veterinary Record*, 186, 496. https://doi.org/10.1136/vr.m1452
- Chen, N., Zhou, M., Dong, X., Qu, J., Gong, F., Han, Y., Qiu, Y., Wang, J., Liu, Y., Wei, Y., Xia, J., Yu, T., Zhang, X., & Zhang, L. I. (2020). Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: A descriptive study. *Lancet*, 395, 507–513. https://doi.org/10.1016/S0140-6736(20)30211-7
- Chu, H., Chan, J.-W., Wang, Y., Yuen, T.-T., Chai, Y., Hou, Y., Shuai, H., Yang, D., Hu, B., Huang, X., Zhang, X. I., Cai, J.-P., Zhou, J., Yuan, S., Kok, K.-H., To, K.-W., Chan, I.-Y., Zhang, A. J., Sit, K.-Y., ... Yuen, K.-Y. (2020). Comparative replication and immune activation profiles of SARS-CoV-2 and SARS-CoV in human lungs: An ex vivo study with implications for the pathogenesis of COVID-19. *Clinical Infectious Diseases*, 71(6), 1400–1409. https://doi.org/10.1093/cid/ciaa410
- Chugh, T. (2020). Timelines of COVID-19 vaccines. Current Medicine Research and Practice, 10, 137–138. https://doi.org/10.1016/j. cmrp.2020.07.009
- Cipriano, M., Ruberti, E., & Giacalone, A. (2020). Gastrointestinal infection could be new focus for coronavirus diagnosis. *Cureus*, 12, e7422. https://doi.org/10.7759/cureus.7422
- Corbett, K. S., Flynn, B., Foulds, K. E., Francica, J. R., Boyoglu-Barnum, S., Werner, A. P., Flach, B., O'Connell, S., Bock, K. W., Minai, M., Nagata, B. M., Andersen, H., Martinez, D. R., Noe, A. T., Douek, N., Donaldson, M. M., Nji, N. N., Alvarado, G. S., Edwards, D. K., ... Graham, B. S. (2020). Evaluation of the mRNA-1273 vaccine against SARS-CoV-2 in Nonhuman primates. *New England Journal of Medicine*. https://doi. org/10.1056/NEJMoa2024671
- Corman, V. M., Muth, D., Niemeyer, D., & Drosten, C. (2018). Hosts and sources of endemic human coronaviruses. Advances in Virus Research, 100, 163–188. https://doi.org/10.1016/bs.aivir.2018.01.001
- Cyranoski, D. (2020a). Profile of a killer: The complex biology powering the coronavirus pandemic. *Nature*, 581, 22–26. https://doi. org/10.1038/d41586-020-01315-7
- Cyranoski, D. (2020b). The biggest mystery: What it will take to trace the coronavirus source. *Nature*. https://doi.org/10.1038/d41586-020-01541-z
- Day, M. J., Horzinek, M. C., Schultz, R. D., & Squires, R. A. (2016). WSAVA Guidelines for the vaccination of dogs and cats. *Journal of Small Animal Practice*, 57, E1–E45. https://doi.org/10.1111/jsap.2\_12431
- Decaro, N., & Lorusso, A. (2020). Novel human coronavirus (SARS-CoV-2): A lesson from animal coronaviruses. *Veterinary Microbiology*, 244, 108693. https://doi.org/10.1016/j.vetmic.2020.108693
- Deng, J., Jin, Y., Liu, Y., Sun, J., Hao, L., Bai, J., Huang, T., Lin, D., Jin, Y., & Tian, K. (2020). Serological survey of SARS-CoV-2 for experimental, domestic, companion and wild animals excludes intermediate hosts of 35 different species of animals. *Transboundary and Emerging Diseases*, 67(4), 1745–1749. https://doi.org/10.1111/tbed.13577
- Dhama K, Patel SK, Pathak M, Yatoo MII, Tiwari R, Malik YS, Singh R, Sah R, Rabaan A. A., Bonilla-Aldana D. K., Rodriguez-Morales A. J. (2020). An update on SARS-CoV-2/COVID-19 with particular reference to its clinical pathology, pathogenesis, immunopathology and mitigation strategies. *Travel Medicine and Infectious Disease*, 101755. http://dx.doi.org/10.1016/j.tmaid.2020.101755.
- Dong, R., Pei, S., Yin, C., He, R. L., & Yau, S. S. (2020). Analysis of the hosts and transmission paths of SARS-CoV-2 in the COVID-19 outbreak. *Genes*, 11, 637–653. https://doi.org/10.3390/genes11060637
- Enserink, M. (2020). Coronavirus rips through Dutch mink farms, triggering culls. *Science*, *368*, 1169. https://doi.org/10.1126/science.368. 6496.1169
- Fehr, D., Holznagel, E., Bolla, S., Hauser, B., Herrewegh, A. A., Horzinek, M. C., & Lutz, H. (1997). Placebo-controlled evaluation of a modified life virus vaccine against feline infectious peritonitis: Safety and efficacy under field conditions. *Vaccine*, 15, 1101–1109. https://doi. org/10.1016/S0264-410X(97)00006-6

ransboundary and Emerging Diseases

- Felsenstein, S., Herbert, J. A., McNamara, P. S., & Hedrich, C. M. (2019). COVID-19: Immunology and treatment options. *Clinical Immunology*, 215, 108448. https://doi.org/10.1016/j.clim.2020.108448
- Felten, S., & Hartmann, K. (2019). Diagnosis of feline infectious peritonitis: A review of the current literature. Viruses, 11, 1068. https://doi. org/10.3390/v11111068
- Foley, J. E., Poland, A., Carlson, J., & Pedersen, N. C. (1997). Patterns of feline coronavirus infection and fecal shedding from cats in multiple-cat environments. *Journal of the American Veterinary Medical Association*, 210, 1307–1312.
- Gao, Q., Bao, L., Mao, H., Wang, L., Xu, K., Yang, M., Li, Y., Zhu, L., Wang, N., Lv, Z., Gao, H., Ge, X., Kan, B., Hu, Y., Liu, J., Cai, F., Jiang, D., Yin, Y., Qin, C., ... Qin, C. (2020). Development of an inactivated vaccine candidate for SARS-CoV-2. *Science*, *369*(6499), 77–81. https://doi.org/10.1126/science.abc1932
- Gao, T., Pan, X., & Pan, C. (2020). The fate of house cats during the COVID-19 pandemic. *Microbes and Infection*, 22, 157. https://doi. org/10.1016/j.micinf.2020.04.006
- Gorbalenya, A. E., Baker, S. C., Baric, R. S., de Groot, R. J., Drosten, C., Gulyaeva, A. A., Haagmans, B. L., Lauber, C., Leontovich, A. M., Neuman, B. W., Penzar, D., Perlman, S., Poon, L. L. M., Samborskiy, D., Sidorov, I. A., Sola, I., & Ziebuhr, J. (2020). Coronaviridae Study Group of the International Committee on Taxonomy of Viruses. The species Severe acute respiratory syndrome-related coronavirus: Classifying 2019-nCoV and naming it SARS-CoV-2. Nature Microbiology, 5, 536– 544. https://doi.org/10.1038/s41564-020-0695-z
- Guan, W.-J., Ni, Z.-Y., Hu, Y. U., Liang, W.-H., Ou, C.-Q., He, J.-X., Liu, L., Shan, H., Lei, C.-L., Hui, D. S. C., Du, B., Li, L.-J., Zeng, G., Yuen, K.-Y., Chen, R.-C., Tang, C.-L., Wang, T., Chen, P.-Y., Xiang, J., ... Zhong, N.-S. (2020). Clinical characteristics of coronavirus disease 2019 in China. New England Journal of Medicine, 382, 1708–1720. https://doi. org/10.1056/NEJMoa2002032
- Gu J, Han B, Wang J (2020). COVID-19: Gastrointestinal Manifestations and Potential Fecal-Oral Transmission. *Gastroenterology*, 158, (6), 1518–1519. http://dx.doi.org/10.1053/j.gastro.2020.02.054.
- Guo, H., Guo, A., Wang, C., Yan, B., Lu, H., & Chen, H. (2008). Expression of feline angiotensin converting enzyme 2 and its interaction with SARS-CoV S1 protein. *Research in Veterinary Science*, 84, 494–496. https://doi.org/10.1016/j.rvsc.2007.05.011
- Guo, Y. R., Cao, Q. D., Hong, Z. S., Tan, Y. Y., Chen, S. D., Jin, H. J., Tan, K. S., Wang, D. Y., & Yan, Y. (2020). The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak-an update on the status. *Military Medical Research*, *7*, 1–10. https://doi.org/10.1186/s40779-020-00240-0
- Guy, J. S. (2000). Turkey coronavirus is more closely related to avian infectious bronchitis virus than to mammalian coronaviruses: A review. *Avian Pathology*, 29, 207–212. https://doi.org/10.1080/0307945005 0045459
- He, L., Ding, Y., Zhang, Q., Che, X., He, Y., Shen, H., Wang, H., Li, Z., Zhao, L., Geng, J., & Deng, Y. (2006). Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected ACE2+ cells in SARS patients: Relation to the acute lung injury and pathogenesis of SARS. *Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland*, 210, 288–297. https://doi.org/10.1002/path.2067
- He, Y., Wang, Z., Li, F., & Shi, Y. (2020). Public health might be endangered by possible prolonged discharge of SARS-CoV-2 in stool. *Journal of Infection*, 80, e18–e19. https://doi.org/10.1016/j.jinf.2020.02.031
- Herrewegh, A. A., Smeenk, I., Horzinek, M. C., Rottier, P. J., & de Groot, R. J. (1998). Feline coronavirus type II strains 79–1683 and 79–1146 originate from a double recombination between feline coronavirus type I and canine coronavirus. *Journal of Virology*, 72, 4508–4514. https://doi.org/10.1128/JVI.72.5.4508-4514.1998
- Hartmann K, Ritz S (2008). Treatment of cats with feline infectious peritonitis. Veterinary Immunology and Immunopathology, 123, (1-2), 172– 175. http://dx.doi.org/10.1016/j.vetimm.2008.01.026.

- Holmes, E. C., & Rambaut, A. (2004). Viral evolution and the emergence of SARS coronavirus. *Philosophical Transactions of the Royal Society* of London. Series B: Biological Sciences, 359, 1059–1065. https://doi. org/10.1098/rstb.2004.1478
- Holzworth, J. (1963). Some important disorders of cats. Cornell Veterinarian, 53, 157-160.
- Hossain, M. G., Javed, A., Akter, S., & Saha, S. (2020). SARS-CoV-2 host diversity: An update of natural infections and experimental evidences. *Journal of Microbiology, Immunology and Infection*. in press https://doi.org/10.1016/j.jmii.2020.06.006
- Jaimes, J. A., & Whittaker, G. R. (2018). Feline coronavirus: Insights into viral pathogenesis based on the spike protein structure and function. Virology, 517, 108–121. https://doi.org/10.1016/j. virol.2017.12.027
- Kai, H., & Kai, M. (2020). Interactions of coronaviruses with ACE2, angiotensin II, and RAS inhibitors—lessons from available evidence and insights into COVID-19. *Hypertension Research*, 43(7), 648–654.
- Kim, S. J., Nguyen, V. G., Park, Y. H., Park, B. K., & Chung, H. C. (2020). A novel synonymous mutation of SARS-CoV-2: Is this possible to affect their antigenicity and immunogenicity? *Vaccines*, *8*, 220. https://doi. org/10.3390/vaccines8020220
- Kim, Y., Liu, H., Kankanamalage, A. C. G., Weerasekara, S., Hua, D. H., Groutas, W. C., Chang, K. O., & Pedersen, N. C. (2016). Reversal of the progression of fatal coronavirus infection in cats by a broad-spectrum coronavirus protease inhibitor. *PLoS Path*, 12, e1005531. https://doi.org/10.1371/journal.ppat.1005531
- Kipar, A., May, H., Menger, S., Weber, M., Leukert, W., & Reinacher, M. (2005). Morphologic features and development of granulomatous vasculitis in feline infectious peritonitis. *Veterinary Pathology*, 42, 321–330. https://doi.org/10.1354/vp.42-3-321
- Kipar, A., & Meli, M. L. (2014). Feline infectious peritonitis: Still an enigma? Veterinary Pathology, 51, 505–526. https://doi.org/10.1177/03009 85814522077
- Kipar, A., Meli, M. L., Failing, K., Euler, T., Gomes-Keller, M. A., Schwartz, D., Lutz, H., & Reinacher, M. (2006). Natural feline coronavirus infection: Differences in cytokine patterns in association with the outcome of infection. Veterinary Immunology and Immunopathology, 112, 141–155. https://doi.org/10.1016/j.vetimm.2006.02.004
- Lam, T. T. Y., Shum, M. H. H., Zhu, H. C., Tong, Y. G., Ni, X. B., Liao, Y. S., Wei, W., Cheung, W. Y. M., Li, W. J., Li, L. F., & Leung, G. M. (2020). Identification of 2019-nCoV related coronaviruses in Malayan pangolins in southern China. *Nature*, 583, 282-285. https://doi. org/10.1038/s41586-020-2169-0
- Lamers, M. M., Beumer, J., van der Vaart, J., Knoops, K., Puschhof, J., Breugem, T. I., Ravelli, R. B., van Schayck, J. P., Mykytyn, A. Z., Duimel, H. Q., van Donselaar, E., Riesebosch, S., Kuijpers, H. J. K., Schipper, D., van de Wetering, W. J., de Graaf, M., Koopmans, M., Cuppen, E., Peters, P. J., ... Clevers, H. (2020). SARS-CoV-2 productively infects human gut enterocytes. *Science*, 369(6499), 50–54. https://doi.org/10.1126/science.abc1669
- Lau, S. K. P., Luk, H. K. H., Wong, A. C. P., Li, K. S. M., Zhu, L., He, Z., Fung, J., Chan, T. T. Y., Fung, K. S. C., & Woo, P. C. Y. (2020). Possible bat origin of severe acute respiratory syndrome coronavirus 2. *Emerging Infectious Disease*, 26, 1542–1547. https://doi.org/10.3201/eid26 07.200092
- Lau, S.-Y., Wang, P., Mok, B.-Y., Zhang, A. J., Chu, H., Lee, A.-Y., Deng, S., Chen, P., Chan, K.-H., Song, W., Chen, Z., To, K.-W., Chan, J.-W., Yuen, K.-Y., & Chen, H. (2020). Attenuated SARS-CoV-2 variants with deletions at the S1/S2 junction. *Emerging Microbes & Infections*, *9*, 837–842. https://doi.org/10.1080/22221751.2020.1756700
- Lauzi, S., Stranieri, A., Giordano, A., Luzzago, C., Zehender, G., Paltrinieri, S., & Ebranati, E. (2020). Origin and transmission of Feline coronavirus type I in domestic cats from Northern Italy: A phylogeographic approach. *Veterinary Microbiology*, 244, 108667. https://doi. org/10.1016/j.vetmic.2020.108667

- Ledford, H. (2020). Hopes rise for coronavirus drug Remdesivir. Nature. https://doi.org/10.1038/d41586-020-01295-8
- Letko, M., Seifert, S. N., Olival, K. J., Plowright, R. K., & Munster, V. J. (2020). Bat-borne virus diversity, spillover and emergence. *Nature Reviews Microbiology*, 18, 461–471. https://doi.org/10.1038/s4157 9-020-0394-z
- Li, C., Yang, Y., & Ren, L. (2020). Genetic evolution analysis of 2019 novel coronavirus and coronavirus from other species. *Infection, Genetics and Evolution, 82*, 104285. https://doi.org/10.1016/j. meegid.2020.104285
- Li, X., Geng, M., Peng, Y., Meng, L., & Lu, S. (2020). Molecular immune pathogenesis and diagnosis of COVID-19. *Journal of Pharmaceutical Analysis*, 10(2), 102–108. https://doi.org/10.1016/j. jpha.2020.03.001
- Li, Y. C., Bai, W. Z., & Hashikawa, T. (2020). The neuroinvasive potential of SARS-CoV2 may play a role in the respiratory failure of COVID-19 patients. *Journal of Medical Virology*, 92, 552–555. https://doi. org/10.1002/jmv.25728
- Licitra, B. N., Duhamel, G. E., & Whittaker, G. R. (2014). Canine enteric coronaviruses: Emerging viral pathogens with distinct recombinant spike proteins. *Viruses*, 6, 3363–3376. https://doi.org/10.3390/ v6083363
- Lin, X.-D., Wang, W., Hao, Z.-Y., Wang, Z.-X., Guo, W.-P., Guan, X.-Q., Wang, M.-R., Wang, H.-W., Zhou, R.-H., Li, M.-H., Tang, G.-P., Wu, J., Holmes, E. C., & Zhang, Y.-Z. (2017). Extensive diversity of coronaviruses in bats from China. *Virology*, 507, 1–10. https://doi. org/10.1016/j.virol.2017.03.019
- Liu, K., Chen, Y., Lin, R., & Han, K. (2020). Clinical features of COVID-19 in elderly patients: A comparison with young and middle-aged patients. *Journal of Infection*, 80(6), e14–e18. https://doi.org/10.1016/j. jinf.2020.03.005
- Liu, P., Jiang, J. Z., Wan, X. F., Hua, Y., Li, L., Zhou, J., Wang, X., Hou, F., Chen, J., Zou, J., & Chen, J. (2020). Are pangolins the intermediate host of the 2019 novel coronavirus (SARS-CoV-2)? *PLoS Path*, 14, e1008421. https://doi.org/10.1371/journal.ppat.1008421
- Lu, C.-C., Chen, M.-Y., Lee, W.-S., & Chang, Y.-L. (2020). Potential therapeutic agents against COVID-19: What we know so far. *Journal of the Chinese Medical Association*, 83(6), 534–536. https://doi.org/10.1097/ JCMA.00000000000318
- Luan, J., Lu, Y., Jin, X., & Zhang, L. (2020). Spike protein recognition of mammalian ACE2 predicts the host range and an optimized ACE2 for SARS-CoV-2 infection. *Biochemical and Biophysical Research Communications*, 526, 165–169. https://doi.org/10.1016/j. bbrc.2020.03.047
- MacLachlan, N. J., & Dubovi, E. J. (2011). Coronaviridae. In C. A. San Diego (Ed.), Fenner's veterinary virology, (4th ed., pp. 393–413). Elsevier.
- Mahase, E. (2020). Covid-19: Concerns grow over inflammatory syndrome emerging in children. *The BMJ*, 369, m1710. https://doi. org/10.1136/bmj.m1710
- Malaiyan, J., Arumugam, S., Mohan, K., & Gomathi, R. G. (2020). An update on the origin of SARS-CoV-2: Despite closest identity, bat (RaTG13) and pangolin derived coronaviruses varied in the critical binding site and O-linked glycan residues. *Journal of Medical Virology*. https://doi.org/10.1002/jmv.26261
- Malik, Y. S., Sircar, S., Bhat, S., Sharun, K., Dhama, K., Dadar, M., Tiwari, R., & Chaicumpa, W. (2020). Emerging novel coronavirus (2019-nCoV)-current scenario, evolutionary perspective based on genome analysis and recent developments. *Veterinary Quarterly*, 40, 68–76. https://doi.org/10.1080/01652176.2020.1727993
- Martina, B. E., Haagmans, B. L., Kuiken, T., Fouchier, R. A., Rimmelzwaan, G. F., Van Amerongen, G., Peiris, J. M., Lim, W., & Osterhaus, A. D. (2003). SARS virus infection of cats and ferrets. *Nature*, 425, 915. https://doi.org/10.1038/425915a
- Mavian, C., Marini, S., Manes, C., Capua, I., Prosperi, M., & Salemi, M. (2020). Regaining perspective on

SARS-CoV-2 molecular tracing and its implications. *medRxiv*. https://doi.org/10.1101/2020.03.16.20034470

- Mavian, C., Pond, S. K., Marini, S., Magalis, B. R., Vandamme, A. M., Dellicour, S., Scarpino, S. V., Houldcroft, C., Villabona-Arenas, J., Paisie, T. K., Trovão, N. S., Boucher, C., Zhang, Y., Scheuermann, R. H., Gascuel, O., Lam, T. T., Suchard, M. A., Abecasis, A., Wilkinson, E., ... Salemi, M. (2020). Sampling bias and incorrect rooting make phylogenetic network tracing of SARS-COV-2 infections unreliable. *Proceedings of the National Academy of Science of the United States of America*, 117, 12522–12523. https://doi.org/10.1073/pnas.20072 95117
- Meli, M. L., Burr, P., Decaro, N., Graham, E., Jarrett, O., Lutz, H., McDonald, M., & Addie, D. D. (2013). Samples with high virus load cause a trend toward lower signal in feline coronavirus antibody tests. *Journal of Feline Medicine and Surgery*, 15, 295–299. https://doi. org/10.1177/1098612X12467995
- Merad, M., & Martin, J. C. (2020). Pathological inflammation in patients with COVID-19: A key role for monocytes and macrophages. *Nature Reviews Immunology*, 20, 355–362. https://doi.org/10.1038/s4157 7-020-0331-4
- Molenaar, R. J., Vreman, S., Hakze-van der Honing, R. W., Zwart, R., de Rond, J., Weesendorp, E., Smit, L. A., Koopmans, M., Bouwstra, R., & Stegeman, A.& van der Poel, W. H. (2020). Clinical and pathological findings in SARS-CoV-2 disease outbreaks in farmed mink (*Neovison vison*). Veterinary Pathology, 57, 653–657. https://doi. org/10.1177/0300985820943535
- Murphy, B. G., Perron, M., Murakami, E., Bauer, K., Park, Y., Eckstrand, C., Liepnieks, M., & Pedersen, N. C. (2018). The nucleoside analog GS-441524 strongly inhibits feline infectious peritonitis (FIP) virus in tissue culture and experimental cat infection studies. *Veterinary Microbiology*, 219, 226-233. https://doi.org/10.1016/j. vetmic.2018.04.026
- Newman, A., Smith, D., Ghai, R. R., Wallace, R. M., Torchetti, M. K., Loiacono, C., Murrell, L. S., Carpenter, A., Moroff, S., Rooney, J. A., & Behravesh, C. B. (2020). First reported cases of SARS-CoV-2 infection in companion animals—New York, March–April 2020. Morbidity and Mortality Weekly Report, 69, 710. https://doi.org/10.15585/ mmwr.mm6923e3
- Nili, A., Farbod, A., Neishabouri, A., Mozafarihashjin, M., Tavakolpour, S., & Mahmoudi, H. (2020). Remdesivir: A beacon of hope from Ebola virus disease to COVID-19. *Reviews in Medical Virology*, e2133. https://doi.org/10.1002/rmv.2133
- OIE, World Organisation for Animal Health. Retrieved from https:// www.oie.int/en/scientific-expertise/specific-information-andrecommendations/questions-and-answers-on-2019novel-coron avirus/events-in-animals/
- Olsen, C. W., Corapi, W. V., Jacobson, R. H., Simkins, R. A., Saif, L. J., & Scott, F. W. (1993). Identification of antigenic sites mediating antibody-dependent enhancement of feline infectious peritonitis virus infectivity. *Journal of General Virology*, 74, 745–749. https://doi. org/10.1099/0022-1317-74-4-745
- Oreshkova, N., Molenaar, R. J., Vreman, S., Harders, F., Oude Munnink, B.
  B., Hakze-van der Honing, R. W., Gerhards, N., Tolsma, P., Bouwstra,
  R., Sikkema, R. S., Tacken, M. G. J., de Rooij, M. M. T., Weesendorp,
  E., Engelsma, M. Y., Bruschke, C. J. M., Smit, L. A. M., Koopmans,
  M., van der Poel, W. H. M., & Stegeman, A. (2020). SARS-CoV-2
  infection in farmed minks, the Netherlands, April and May 2020. *Eurosurveillance*, *25*, 2001005. https://doi.org/10.2807/1560-7917.
  ES.2020.25.23.2001005
- Ou, J., Zhou, Z., Zhang, J., Lan, W., Zhao, S., Wu, J., Seto, D., Zhang, G., & Zhang, Q. (2020). RBD mutations from circulating SARS-CoV-2 strains enhance the structural stability and human ACE2 affinity of the spike protein. *bioRxiv*. https://doi.org/10.1101/2020.03.15.991844
- Paltrinieri, S., Cammarata Parodi, M., Cammarata, G., & Mambretti, M. (1998). Type IV hypersensitivity in the pathogenesis of FIPV-induced

lesions. Journal of Veterinary Medicine Series B, 45, 151–159. https:// doi.org/10.1111/j.1439-0450.1998.tb00778.x

- Paltrinieri, S., Rossi, G., & Giordano, A. (2014). Relationship between rate of infection and markers of inflammation/immunity in Holy Birman cats with feline coronavirus. *Research in Veterinary Science*, 97, 263– 270. https://doi.org/10.1016/j.rvsc.2014.08.009
- Park, M. D. (2020). Macrophages: A Trojan horse in COVID-19? Nature Reviews Immunology, 20(6), 351. https://doi.org/10.1038/s4157 7-020-0317-2
- Patterson, E. I., Elia, G., Grassi, A., Giordano, A., Desario, C., Medardo, M., Smith, S. L., Anderson, E. R., Prince, T., Patterson, G. T., Lorusso, E., Lucente, M. S., Lanave, G., Lauzi, S., Bonfanti, U., Stranieri, A., Martella, V., Solari Basano, F., Barrs, V. R., ... Decaro, N. (2020). Evidence of Exposure to SARS-CoV-2 in cats and dogs from households in Italy. *bioRxiv*. https://doi.org/10.1101/2020.07.21.214346
- Pedersen, N. C. (1976). Serologic studies of naturally occurring feline infectious peritonitis. American Journal of Veterinary Research, 37, 1449–1453.
- Pedersen, N. C. (2009). A review of feline infectious peritonitis virus infection: 1963–2008. *Journal of Feline Medicine and Surgery*, 11, 225– 258. https://doi.org/10.1016/j.jfms.2008.09.008
- Pedersen, N. C. (2014a). An update on feline infectious peritonitis: Virology and immunopathogenesis. *Veterinary Journal*, 201, 123–132. https://doi.org/10.1016/j.jfms.2008.09.008
- Pedersen, N. C. (2014b). An update on feline infectious peritonitis: Diagnostics and therapeutics. *Veterinary Journal*, 201, 133–141. https://doi.org/10.1016/j.tvjl.2014.04.016
- Pedersen, N. C., Kim, Y., Liu, H., Galasiti Kankanamalage, A. C., Eckstrand, C., Groutas, W. C., Bannasch, M., Meadows, J. M., & Chang, K. O. (2018). Efficacy of a 3c-like protease inhibitor in treating various forms of acquired feline infectious peritonitis. *Journal of Feline Medicine and Surgery*, 20, 378–392. https://doi.org/10.1177/10986 12X17729626
- Pedersen, N. C., Perron, M., Bannasch, M., Montgomery, E., Murakami, E., Liepnieks, M., & Liu, H. (2019). Efficacy and safety of the nucleoside analog GS-441524 for treatment of cats with naturally occurring feline infectious peritonitis. *Journal of Feline Medicine* and Surgery, 21, 271–281. https://doi.org/10.1177/1098612X19 825701
- Pedersen, S. F., & Ho, Y. C. (2020). SARS-CoV-2: A storm is raging. Journal of Clinical Investigation, 130, 2202–2205. https://doi.org/10.1172/ JCI137647
- Petersen, E., Koopmans, M., Go, U., Hamer, D. H., Petrosillo, N., Castelli, F., Storgaard, M., Al Khalili, S., & Simonsen, L. (2020). Comparing SARS-CoV-2 with SARS-CoV and influenza pandemics. *Lancet Infectious Diseases*, 20(9), e238–e244. https://doi.org/10.1016/ S1473-3099(20)30484-9
- Porter, E., Tasker, S., Day, M. J., Harley, R., Kipar, A., Siddell, S. G., & Helps, C. R. (2014). Amino acid changes in the spike protein of feline coronavirus correlate with systemic spread of virus from the intestine and not with feline infectious peritonitis. *Veterinary Research*, 45, 49. https://doi.org/10.1186/1297-9716-45-49
- Regan, A. D., Ousterout, D. G., & Whittaker, G. R. (2010). Feline lectin activity is critical for the cellular entry of feline infectious peritonitis virus. *Journal of Virology*, 84, 7917–7921. https://doi.org/10.1128/ JVI.00964-10
- Ricke, D., & Malone, R. W. (2020). Medical countermeasures analysis of 2019-nCoV and vaccine risks for antibody-dependent enhancement (ADE). SSRN Electronic Journal. https://doi.org/10.2139/ ssrn.3546070
- Rossi, G., Cornaro, C., Battilani, M., Pocacqua, V., & Paltrinieri, S. (2011). Production of IFN-γ in feline whole blood after incubation with potential T-celle epitopes of the Nucleocapsid protein of feline coronavirus. *Veterinary Microbiology*, 150, 248–256. https://doi. org/10.1016/j.vetmic.2011.02.004

Roussel, Y., Giraud-Gatineau, A., Jimeno, M. T., Rolain, J. M., Zandotti, C., Colson, P., & Raoult, D. (2020). SARS-CoV-2: Fear versus data. *International Journal of Antimicrobial Agents*, 55, 105947. https://doi. org/10.1016/j.ijantimicag.2020.105947

sboundary and Eme

- Russell, B., Moss, C., George, G., Santaolalla, A., Cope, A., Papa, S., & Van Hemelrijck, M. (2020). Associations between immune-suppressive and stimulating drugs and novel COVID-19—a systematic review of current evidence. *Ecancermedicalscience*, 14, 1022. https://doi. org/10.3332/ecancer.2020.1022
- Saif, L. J. (2020). Vaccines for COVID-19: Perspectives, prospects, and challenges based on candidate SARS, MERS, and animal coronavirus vaccines. *European Medical Journal*. https://doi.org/10.33590/ emj/200324
- Sailleau, C., Dumarest, M., Vanhomwegen, J., Delaplace, M., Caro, V., Kwasiborski, A., Hourdel, V., Chevaillier, P., Barbarino, A., Comtet, L., Pourquier, P., Klonjkowski, B., Manuguerra, J.-C., Zientara, S., & Le Poder, S. (2020). First detection and genome sequencing of SARS-CoV-2 in an infected cat in France. *Transboundary and Emerging Diseases*. https://doi.org/10.1111/tbed.13659
- Sanche, S., Lin, Y., Xu, C., Romero-Severson, E., Hengartner, N., & Ke, R. (2020). High contagiousness and rapid spread of severe acute respiratory syndrome coronavirus 2. *Emerging Infectious Diseases*, 26, 1470–1477. https://doi.org/10.3201/eid2607.200282
- Sarzi-Puttini, P., Giorgi, V., Sirotti, S., Marotto, D., Ardizzone, S., Rizzardini, G., Antinori, S., & Galli, M. (2020). COVID-19, cytokines and immunosuppression: What can we learn from severe acute respiratory syndrome? *Clinical and Experimental Rheumatology*, 38, 337–342.
- Satoh, R., Kobayashi, H., Takano, T., Motokawa, K., Kusuhara, H., & Hohdatsu, T. (2010). Characterization of T helper (Th) 1-and Th2-type immune responses caused by baculovirus-expressed protein derived from the S2 domain of feline infectious peritonitis virus, and exploration of the Th1 and Th2 epitopes in a mouse model. *Microbiology and Immunology*, 54, 726-733. https://doi. org/10.1111/j.1348-0421.2010.00275.x
- Schatzmann Peron, J. P., & Nakaya, H. (2020). Susceptibility of the elderly to SARS-CoV-2 infection: ACE-2 overexpression, shedding and antibody-dependent enhancement (ADE). *Preprints*, 75, e1912. https:// doi.org/10.20944/preprints202003.0400.v1
- Shen, C., Wang, Z., Zhao, F., Yang, Y., Li, J., Yuan, J., Wang, F., Li, D., Yang, M., Xing, L. I., Wei, J., Xiao, H., Yang, Y., Qu, J., Qing, L., Chen, L. I., Xu, Z., Peng, L., Li, Y., ... Liu, L. (2020). Treatment of 5 critically III patients with COVID-19 with convalescent plasma. *Journal of the American Medical Association JAMA*, 323, 1582–1589. https://doi.org/10.1001/ jama.2020.4783
- Shi, Y., Wang, Y., Shao, C., Huang, J., Gan, J., Huang, X., Bucci, E., Piacentini, M., Ippolito, G., & Melino, G. (2020). COVID-19 infection: The perspectives on immune responses. COVID-19 infection: The perspectives on immune responses. *Cell Death and Differentiation*, 27, 1451–1454. https://doi.org/10.1038/s4141 8-020-0530-3
- Singh, B. (2020). Understanding the role of key point mutations in receptor binding domain of SARS-CoV-2 spike glycoprotein. *Infectious Disorder Drug Targets*. https://doi.org/10.20944/preprints202003. 0394.v2
- Sironi, M., Hasnain, S. E., Rosenthal, B., Phan, T., Luciani, F., Shaw, M. A., Sallum, M. A., Mirhashemi, M. E., Morand, S., & González-Candelas, F. (2020). SARS-CoV-2 and COVID-19: A genetic, epidemiological, and evolutionary perspective. *Infection, Genetics and Evolution, 29*, 104384. https://doi.org/10.1016/j.meegid.2020.104384
- Song, D., & Park, B. (2012). Porcine epidemic diarrhoea virus: A comprehensive review of molecular epidemiology, diagnosis, and vaccines. Virus Genes, 44, 167–175. https://doi.org/10.1007/s1126 2-012-0713-1
- South, A. M., Diz, D., & Chappell, M. C. (2020). COVID-19, ACE2 and the cardiovascular consequences. *American Journal of*

1798

Physiology, 318(5), H1084-H1090. https://doi.org/10.1152/ajphe art.00217.2020

- Stranieri, A., Giordano, A., Paltrinieri, S., Giudice, C., Cannito, V., & Lauzi, S. (2018). Comparison of the performance of laboratory tests in the diagnosis of feline infectious peritonitis. *Journal of Veterinary Diagnostic Investigation*, 30, 459–463. https://doi.org/10.1177/10406 38718756460
- Su, S., Wong, G., Shi, W., Liu, J., Lai, A. C., Zhou, J., Liu, W., Bi, Y., & Gao, G. F. (2016). Epidemiology, genetic recombination, and pathogenesis of coronaviruses. *Trends in Microbiology*, 24, 490–502. https://doi. org/10.1016/j.tim.2016.03.003
- Takano, T., Morioka, H., Gomi, K., Tomizawa, K., Doki, T., & Hohdatsu, T. (2014). Screening and identification of T helper 1 and linear immunodominant antibody-binding epitopes in spike 1 domain and membrane protein of feline infectious peritonitis virus. *Vaccine*, *32*, 1834–1840. https://doi.org/10.1016/j.vaccine.2014.01.074
- Takano, T., Wakayama, Y., & Doki, T. (2019). Endocytic pathway of feline coronavirus for cell entry: Differences in serotype-dependent viral entry pathway. *Pathogens*, 8, 300. https://doi.org/10.3390/patho gens8040300
- Takano, T., Yamada, S., Doki, T., & Hohdatsu, T. (2019). Pathogenesis of oral type I feline infectious peritonitis virus (FIPV) infection: Antibody-dependent enhancement infection of cats with type I FIPV via the oral route. *Journal of Veterinary Medical Science*, 81, 911–915. https://doi.org/10.1292/jvms.18-0702
- Tang, X., Wu, C., Li, X., Song, Y., Yao, X., Wu, X., Duan, Y., Zhang, H., Wang, Y., Qian, Z., Cui, J., & Lu, J. (2020). On the origin and continuing evolution of SARS-CoV-2. *National Science Review*, 7(6), 1012–1023. https://doi.org/10.1093/nsr/nwaa036
- Tasker, S. (2018). Diagnosis of feline infectious peritonitis: Update on evidence supporting available tests. Journal of Feline Medicine and Surgery, 20, 228–243. https://doi.org/10.1177/1098612X18758592
- Tay, M. Z., Poh, C. M., Rénia, L., MacAry, P. A., & Ng, L. F. (2020). The trinity of COVID-19: Immunity, inflammation and intervention. *Nature Reviews Immunology*, 20(6), 363–374. https://doi.org/10.1038/s4157 7-020-0311-8
- Tekes, G., & Thiel, H. J. (2016). Feline coronaviruses: Pathogenesis of feline infectious peritonitis. Advances in Virus Research, 96, 193–218. https://doi.org/10.1016/bs.aivir.2016.08.002
- Tetro, J. A. (2020). Is COVID-19 receiving ADE from other coronaviruses? *Microbes and Infection*, 22, 72–73. https://doi.org/10.1016/j. micinf.2020.02.006
- Tian, Y., Rong, L., Nian, W., & He, Y. (2020). Review article: Gastrointestinal features in COVID-19 and the possibility of faecal transmission. *Alimentary Pharmacology and Therapeutics*, 51, 843–851. https://doi. org/10.1111/apt.15731
- Tiwari, M., & Mishra, D. (2020). Investigating the genomic landscape of novel coronavirus (2019-nCoV) to identify non-synonymous mutations for use in diagnosis and drug design. *Journal of Clinical Virology*, 128, 104441. https://doi.org/10.1016/j.jcv.2020.104441
- Tiwari, R., Dhama, K., Sharun, K., Iqbal Yatoo, M., Malik, Y. S., Singh, R., Michalak, I., Sah, R., Bonilla-Aldana, D. K., & Rodriguez-Morales, A. J. (2020). COVID-19: Animals, veterinary and zoonotic links. *Veterinary Quarterly*, 40, 169–182. https://doi.org/10.1080/01652 176.2020.1766725
- van Dorp, L., Acman, M., Richard, D., Shaw, L. P., Ford, C. E., Ormond, L., Owen, C. J., Pang, J., Tan, C. C., Boshier, F. A., & Ortiz, A. T. (2020). Emergence of genomic diversity and recurrent mutations in SARS-CoV-2. *Infection, Genetics and Evolution, 83*, 104351. https://doi. org/10.1016/j.meegid.2020.104351
- Van Hamme E, Desmarets L, Dewerchin H. L., Nauwynck H. J. (2011). Intriguing interplay between feline infectious peritonitis virus and its receptors during entry in primary feline monocytes. *Virus Research*, 160, (1-2), 32–39. http://dx.doi.org/10.1016/j.virus res.2011.04.031.

- Vijaykrishna, D., Smith, G. J., Zhang, J. X., Peiris, J. S. M., Chen, H., & Guan, Y. (2007). Evolutionary insights into the ecology of coronaviruses. *Journal of Virology*, 81, 4012–4020. https://doi.org/10.1128/ JVI.02605-06
- Wang, L., Mitchell, P. K., Calle, P. P., Bartlett, S. L., McAloose, D., Killian,
  M. L., Yuan, F., Fang, Y., Goodman, L. B., Fredrickson, R., Elvinger,
  F., Terio, K., Franzen, K., Stuber, T., Diel, D. G., & Torchetti, M. K.
  (2020). Complete genome sequence of SARS-CoV-2 in a tiger from
  a US zoological collection. *Microbiology Resource Announcements*, 28, e00468-e520. https://doi.org/10.1128/MRA.00468-20
- Wang, L. S., Wang, Y. R., Ye, D. W., & Liu, Q. Q. (2020). A review of the 2019 Novel Coronavirus (COVID-19) based on current evidence. *International Journal of Antimicrobial Agents*, 55, 105948. https://doi. org/10.1016/j.ijantimicag.2020.105948
- Wang, M., Li, M., Ren, R., Li, L., Chen, E. Q., Li, W., & Ying, B. (2020). International expansion of a novel SARS-CoV-2 mutant. *Journal* of Virology, 94(12), e00567-20. https://doi.org/10.1128/JVI. 00567-20
- Wolfe, L. G., & Griesemer, R. A. (1966). Feline infectious peritonitis. Pathologia Veterinaria, 3, 255–270. https://doi.org/10.1177/03009 8586600300309
- Wong, A. C., Li, X., Lau, S. K., & Woo, P. C. (2019). Global epidemiology of bat coronaviruses. Viruses, 11, 174. https://doi.org/10.3390/v1102 0174
- Wong, G., Bi, Y. H., Wang, Q. H., Chen, X. W., Zhang, Z. G., & Yao, Y. G. (2020). Zoonotic origins of human coronavirus 2019 (HCoV-19/SARS-CoV-2): Why is this work important? *Zoological Research*, 18, 213–219. https://doi.org/10.24272/j.issn.2095-8137.2020.031
- Xiao, K., Zhai, J., Feng, Y., Zhou, N., Zhang, X., Zou, J. J., Li, N., Guo, Y., Li, X., Shen, X., Zhang, Z., Shu, F., Huang, W., Li, Y., Zhang, Z., Chen, R. A., Wu, Y. J., Peng, S. M., Huang, M., ... Shen, Y. (2020). Isolation of SARS-CoV-2-related coronavirus from Malayan pangolins. *Nature*, *583*, 286–289. https://doi.org/10.1038/s41586-020-2313-x
- Xu, Z., Shi, L., Wang, Y., Zhang, J., Huang, L., Zhang, C., Liu, S., Zhao, P., Liu, H., Zhu, L. I., Tai, Y., Bai, C., Gao, T., Song, J., Xia, P., Dong, J., Zhao, J., & Wang, F.-S. (2020). Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respiratory Medicine*, *8*, 420–422. https://doi.org/10.1016/S2213 -2600(20)30076-X
- Zhang, H., Kang, Z., Gong, H., Xu, D., Wang, J., Li, Z., Cui, X., Xiao, J., Meng, T., Zhou, W., & Liu, J. (2020). The digestive system is a potential route of 2019-nCov infection: A bioinformatics analysis based on single-cell transcriptomes. *BioRxiv.* https://doi. org/10.1101/2020.01.30.927806
- Zhang, H., Zhou, P., Wei, Y., Yue, H., Wang, Y. I., Hu, M., Zhang, S., Cao, T., Yang, C., Li, M., Guo, G., Chen, X., Chen, Y., Lei, M., Liu, H., Zhao, J., Peng, P., Wang, C.-Y., & Du, R. (2020). Histopathologic changes and SARS-CoV-2 immunostaining in the lung of a patient with COVID-19. Annals of Internal Medicine, 172(9), 629–632. https://doi. org/10.7326/M20-0533
- Zhang, L., Yang, J. R., Zhang, Z., & Lin, Z. (2020). Genomic variations of SARS-CoV-2 suggest multiple outbreak sources of transmission. *medRxiv*. https://doi.org/10.1101/2020.02.25.20027953
- Zhang, Q., Zhang, H., Huang, K., Yang, Y., Hui, X., Gao, J., He, X., Li, C., Gong, W., Zhang, Y., & Peng, C. (2020). SARS-CoV-2 neutralizing serum antibodies in cats: A serological investigation. *bioRxiv*. https:// doi.org/10.1101/2020.04.01.021196
- Zhang, T., Wu, Q., & Zhang, Z. (2020). Probable pangolin origin of SARS-CoV-2 associated with the COVID-19 outbreak. *Current Biology*, 30, 1346–1351. https://doi.org/10.1016/j.cub.2020.03.022
- Zhou, H., Chen, X., Hu, T., Li, J., Song, H., Liu, Y., Wang, P., Liu, D., Yang, J., Holmes, E. C., Hughes, A. C., Bi, Y., & Shi, W. (2020). A novel bat coronavirus closely related to SARS-CoV-2 contains

natural insertions at the S1/S2 cleavage site of the Spike protein. *Current Biology*, 30, 2196–2203. https://doi.org/10.1016/j. cub.2020.05.023

- Zhou, M., Zhang, X., & Qu, J. (2020). Coronavirus disease 2019 (COVID-19): A clinical update. Frontiers of Medicine, 14(2), 126–135. https:// doi.org/10.1007/s11684-020-0767-8
- Zhou, P., Yang, X.-L., Wang, X.-G., Hu, B., Zhang, L., Zhang, W., Si, H.-R., Zhu, Y., Li, B., Huang, C.-L., Chen, H.-D., Chen, J., Luo, Y., Guo, H., Jiang, R.-D., Liu, M.-Q., Chen, Y., Shen, X.-R., Wang, X. I., ... Shi, Z.-L. (2020). A pneumonia outbreak associated with a new coronavirus of

probable bat origin. *Nature, 579*, 270–273. https://doi.org/10.1038/ s41586-020-2012-7

ransboundary and Em

How to cite this article: Paltrinieri S, Giordano A, Stranieri A, Lauzi S. Feline infectious peritonitis and coronavirus disease 19 (COVID-19): Are they similar? *Transbound Emerg Dis* 2021;68:1786–1799. https://doi.org/10.1111/tbed.13856