

# A Systems Biology Workflow for Drug and Vaccine Repurposing: Identifying Small-Molecule BCG Mimics to Reduce or Prevent COVID-19 Mortality

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#### **ABSTRACT**

**Purpose** Coronavirus disease 2019 (COVID-19) is expected to continue to cause worldwide fatalities until the World population develops 'herd immunity', or until a vaccine is developed and used as a prevention. Meanwhile, there is an urgent need to identify alternative means of antiviral defense. Bacillus Calmette–Guérin (BCG) vaccine that has been recognized for its off-target beneficial effects on the immune system can be exploited to boast immunity and protect from emerging novel viruses.

**Methods** We developed and employed a systems biology workflow capable of identifying small-molecule antiviral drugs and vaccines that can boast immunity and affect a wide variety of viral disease pathways to protect from the fatal consequences of emerging viruses.

**Results** Our analysis demonstrates that BCG vaccine affects the production and maturation of naïve T cells resulting in enhanced, long-lasting trained innate immune responses that can provide protection against novel viruses. We have identified small-molecule BCG mimics, including antiviral drugs such as raltegravir and lopinavir as high confidence hits. Strikingly, our top hits emetine and lopinavir were independently validated by recent experimental findings that these compounds inhibit the growth of SARS-CoV-2 in vitro.

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**Conclusions** Our results provide systems biology support for using BCG and small-molecule BCG mimics as putative vaccine and drug candidates against emergent viruses including SARS-CoV-2.

**KEY WORDS** BCG vaccine · COVID-19 · innate immunity · SARS-CoV-2 · systems biology

# **ABBREVIATIONS**

BCG	Bacillus Calmette-Guérin
BIND	Biomolecular interaction network database
BioGRID	Biological general repository for interaction
	datasets
CMap	Connectivity map

COVID-19 Coronavirus disease 2019
DIP Database of interacting proteins
FDA The Food and Drug Administration

GEO Gene expression omnibus

GO Gene ontology

KEGG Kyoto encyclopedia of genes and genomes

Mtb Mycobacterium tuberculosis
RCTs Randomized clinical trials

STRING Search tool for the retrieval of interacting

genes/proteins

WHO World Health Organization

# INTRODUCTION

Few months after the declaration of COVID-19 pandemic by the World Health Organization (WHO), the disease-causing virus is still sweeping the globe, causing more fatalities, failing health care systems, and resulting in severe economic losses. Currently there are no approved drugs to treat COVID-19,



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and new vaccine development is expected to take at least 12–18 months (1,2), with growing fears of possible failure associated with changes in viral antigenic determinants (3) or short-lived immunity (4). Additionally, the highly specific virus-neutralizing antibodies in recovered patients may be short lived and ineffective in preventing the disease caused by the emerging variable strains of the virus (4). With these uncertainties regarding an eminent specific SARS-CoV-2 vaccine, there is a need to search for current alternatives, such as agents that can stimulate or emulate the unique capabilities of our innate immune system.

Recent immuno-oncology success stories indicate that the best cancer-fighting strategies results from unleashing the patients' immune power (5–8). There is an increased awareness that harnessing innate immune responses, opens up new possibilities for long-term, multifaceted tumor control (9,10) and infectious disease prevention (11–13). Therefore, next generation antiviral vaccines should be capable of boosting innate immune responses to tackle a wide range of novel pathogens very early after exposure, as single treatments or adjuvants to traditional vaccines targeting the adaptive immune system.

Accumulating evidence from the biomedical literature indicates that SARS-CoV-mediated pathology, a very similar pathology to SARS-CoV-2, was mainly caused by ineffective innate immune responses, associated with a severe reduction in the number of T cells in the blood (14). Recent evidence indicated that SARS-CoV-2 and Mycobacterium tuberculosis (Mtb) share unique similarities in terms of the host protein interaction partners, and both pathogens infect lung tissues (15). On the other hand, old 'polypharmacological' vaccines, such as the BCG vaccine for tuberculosis (TB), has shown promising therapeutic effects for a wide range of infectious and non-infectious diseases including bladder cancer (16–18). Studies showed that BCG's polypharmacological effects were not limited to memory T cell immunity, but promoted strong, beneficial, and long-lasting effects on innate immunity. The WHO also recognized these beneficial 'offtarget' effects of BCG, calling for a further investigation to repurpose this vaccine for other orphan life-threatening diseases (19). Indeed, there are multiple clinical trials testing BCG for 216 conditions other than TB including 19 studies for COVID-19 as reported on clinicaltrials.gov (20). Additionally, few recent peer-reviewed reports have pointed to an epidemiological relationship between BCG and COVID-19 without providing substantial evidence (21–24). One should expect that the results of the randomized clinical trials (RCTs) will help establish the value of the BCG vaccine as a treatment or prophylactic against the disease.

Herein, we describe a unique drug and vaccine repurposing workflow, and list high confidence proteins and pharmacological classes of compounds, that work as BCG mimics at the system level by inducing beneficial long lasting trained immune response. We also propose that BCG mimics can be used as alternatives to BCG in protecting from COVID-19 and other emergent infectious diseases.

# **MATERIALS AND METHODS**

#### **Integrative Systems Biology Workflow**

We have developed and applied a systems biology workflow to study the BCG network pharmacology and prioritize small-molecule BCG mimics and antivirals. This workflow is based on our original chemocentric informatics workflow described thoroughly in a previous report (25). Our current workflow (Fig. 1) incorporates three major components: (1) a module for mining and prioritizing gene signatures representative of a condition or a biological state; (2) a network-mining module to identify genetic perturbations that induce gene expression profiles that are highly enriched with the genes constituting the condition gene signature; and (3) a pathway enrichment module to understand the biological processes involved in the mechanism of action of BCG and highly correlated genetic perturbagens.

# **BCG** Consensus Gene Signature

A consensus gene signature for BCG vaccine was derived from gene expression profiles in peripheral blood mononuclear cells (PBMCs) in response to a BCG challenge test reported by Matsumiya et al (76), GSE58636 dataset on NCBI Gene Expression Omnibus (GEO) (27). We used the data collected from whole blood samples taken from healthy human subjects enrolled in phase 1 trial (clinical trials registration: NCT01194180). For the purposes of this study we used the gene expression profiles generated from two human subject groups included in the above trial: group 1 (BCG naive), and group 2 (BCG vaccinated; median time since vaccination, 10 years). To study network pharmacology and query the connectivity map, we developed a consensus gene signature using genes that showed significant differential gene expression in response to a BCG challenge test (stimulated) in comparison with controls (unstimulated) on days 0 and 14 in both groups 1 and 2.

#### **Network Building**

A systematic search, for nearest neighbor (NN) genes/proteins of the upregulated and downregulated genes in BCG's gene signature, was conducted in Cytoscape (77) version 3.8.0 using the STRING (78) protein query application. All retrieved protein-protein interactions (PPIs), including both physical and functional interactions were retrieved from widely used and reliable databases such as MINT (79), HPRD (80), BIND



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(81), DIP (82), BioGRID (83), KEGG (84), Reactome (60), EcoCyc (85), NCI-Nature Pathway Interaction Database (86), and Gene Ontology (GO) (87) protein complexes. Network building tools in Cytoscape version 3.7.2 were used to generate PPI networks for BCG-CGS.

# **Enrichment Analysis**

Enrichment analysis was conducted in Cytoscape (77) and MetaCore to identify pathways and biological processes associated with BCG-CGS and CMap genetic connections. The significance of the enrichment was determined by the hypergeometric test (88). All terms from the ontology were ranked based on their calculated p values. Ontology terms with p values less than the p value threshold 0.05 are defined as statistically significant and therefore relevant to the studied list of genes. All terms from the ontology were ranked according to their calculated p values.

# The Connectivity Map (CMap)

The CMap (27,89) is a chemogenomics database that catalogs 1.3 Million profiles of transcriptional responses of human cells to chemical and genetic perturbations. Currently, there are 27,927 perturbagens (19,911 small molecules, and 7494 genetic perturbagens) producing 476,251 expression signatures in 9 human cell lines: PC3, VCAP, A375, A459, HA1E, HCC515, HT29, MCF7, HEPG2. This database of cellular signatures has been produced using the L1000 platform (27); a high-throughput gene expression assay that measures the mRNA transcript abundance of 978 "landmark" genes from human cells.

# **Causal Reasoning**

Causal reasoning (90) analysis identifies genes and proteins of a 'topological significance' in order to make decisions whether these genes/proteins are eligible for targeting in the studied phenotype. In this study, we applied causal reasoning to identify molecular regulators that most likely directly cause the observed expression changes in transcriptional profiles in response to BCG. In this approach, changes in gene expression in both directions as well as the effect of edges in the network are taken into account. For each node (i.e., gene) in the causal reasoning network, observed changes in expression are matched with the expected changes inferred from the network structure given the hypothesis that the observed gene expression is decreased or increased due to its activity. Each node has an outgoing activation or inhibition effects on other objects in the knowledge database, and a key hub with a predicted increase in activity shows increased expression for those genes that the hub is known to activate, and it shows decreased expression for genes it is known to inhibit. Each predicted key hub has a prediction P value which is produced as a result of a binomial test used to assess the probability of making a given number of supportive data out of all defined differentially expressed genes (DEGs) in the examined data. It is noteworthy that causal reasoning examines both direct neighbors of differentially expressed genes, and remote (several steps away) regulators. All causal reasoning predictions were performed in Key Pathway Advisor from Clarivate Analytics, using the Pollard method (91).

#### **R-Package Gplots**

Gplots (92) v3.0.1.2 was used for plotting enhanced heatmaps for transcriptional data (e.g., heatmap representing BCG-CGS in Fig. 2). Heat maps were generated using the heatmap.2 function included in this package.

# **RESULTS**

# **Drug and Vaccine Repurposing Workflow**

To study the BCG polypharmacology and potential beneficial effects of this vaccine in preventing the fatal consequences of COVID-19, we have devised and implemented a 'network biology' workflow (Fig. 1) to interrogate the hypothesis that BCG vaccination may protect from COVID-19 fatalities. This workflow is based on our drug repurposing chemocentric informatics workflow which has been validated previously for small-molecule drug repurposing (25). The current workflow is tweaked towards vaccine repurposing by employing novel bioinformatic approaches to computationally model and connect molecular networks in an effort to understand the underlying 'network' biology of vaccines, and pinpoint the regulatory genes and proteins responsible for causing the observed beneficial multitherapeutic effects. Although we are not the first group to use network biology approaches to study the transcriptional changes of vaccines, to our knowledge, this is the first study that uses these approaches both to support vaccine repurposing, specifically for COVID-19, as well as identify putative small molecule drugs that can mimic the vaccine effects.

# **BCG Consensus Gene Signature**

Our workflow starts with the prioritization of a gene signature to study the BCG network pharmacology. First, we derived a consensus gene signature (CGS) for BCG based on GEO's dataset GSE58636 (26). Details on BCG-CGS signature are found in Table S1 (Supporting Information). Twenty-two differentially-expressed genes across all 4 experiments (2 Groups × 2 time points discussed in Methods) formed



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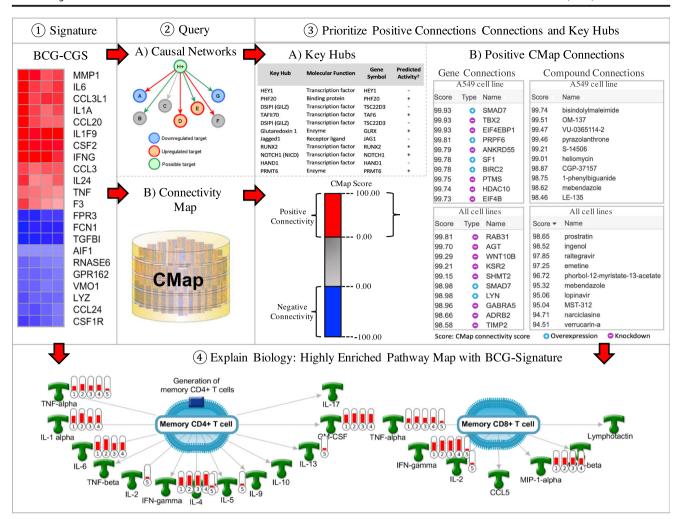


Fig. 1 Workflow for drug and vaccine repurposing. ① A gene signature is identified/derived and a consensus gene signature is prioritized, all genes are nomenclated according to HUGO Gene Nomenclature Committee (HGNC). ② The consensus gene signature is used to query the CMap to identify positive connections capable of producing gene signatures similar to BCG-CGS. ③ Prioritize genes and compounds that induce transcriptional changes similar to those induced by BCG: A) key hubs predicted by causal reasoning; B) Positive genes and compound connections from the connectivity map. ④ Prioritize top enriched pathway map explaining the biological effects of BCG (gene annotations on the pathway map are assigned by Clarivate Analytics, IFN-gamma is an alias for INFG, GM-CSF is an alias for CSF2, MIP-I-alpha is an alias for CCL3).

BCG's consensus gene signature (BCG-CGS) shown in Fig. 2a.

# **BCG Protein-Protein Interactions (PPIs)**

All 22 genes in BCG-CGS were used as seed nodes to build a protein-protein interaction network for signature genes (Fig. 2b). Interactions were extracted from STRING database, and high confidence interactions included physical interactions (e.g., binding), functional interactions (e.g., activation, inhibition, catalysis), or gene co-expression. Two types of networks were generated: 1) high-confidence 'core' network restricted to BCG signature genes as network nodes and high confidence (≥0.70) interactions as network edges, and 2) medium-confidence interaction network obtained from expanding the core network by 20 additional nodes (Fig. 3).

Enrichment analysis results performed in Cytoscape, using STRING's protein-protein interactions, indicated that BCG-CGS is enriched in inflammatory cytokines and immune response modulators (Fig. 2b). Some signature genes are also involved in the negative control of important viral processes (e.g., (FCN1, TNF and CCL3), and others are involved in the response to viral infections (e.g., IFNG, RNASE6, IL6 and TNF). The complete lists of enriched pathways are included in Tables S2 and S3 (Supporting Information).

#### **Identification of Key Hubs**

We identified 291 key hubs using the causal reasoning method, which seeks to identify molecular regulators that can directly cause the observed transcriptional changes in response to BCG vaccination. Key regulators can be transcriptional factors and proteins with potentially altered activity that



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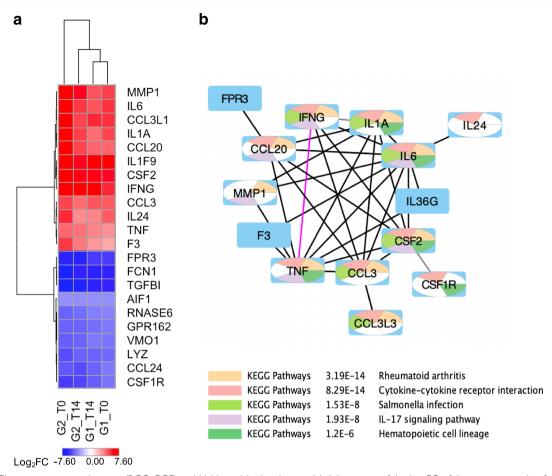


Fig. 2 BCG's consensus gene signature (BCG-CGS) and highly enriched pathways. (a) A heat map of the log<sub>2</sub>FC of the gene expression for differentially expressed genes representing BCG's consensus gene signature. Upregulated genes have positive log<sub>2</sub>FC denoted in red color, and down regulated genes have negative values for log<sub>2</sub>FC denoted in blue color. (b) Core network for BCG-CGS showing highly connected genes in BCG-CGS, deleting all singleton genes. Nodes are color-coded using a split pie chart coloring scheme indicating pathway/gene set contribution to each node from the top 5 most enriched pathways/gene lists. All details about pathway/gene set ID are found in Table S2 (Supporting Information).

explains the transcriptional changes. Top five statistically significant inhibited key hubs included HEY1, DSIPI (GILZ), Jagged1, HAND1 and miR-129-1-3p, whereas top five statistically significant activated key hubs were PHF20, TAFII70, Glutaredoxin, RUNX2 and NOTCH1 (NICD). Top 30 causal key hubs are shown in Table I and all identified 291 key hubs are included in Table S4 (Supporting Information).

# **Identifying BCG 'Mimics'**

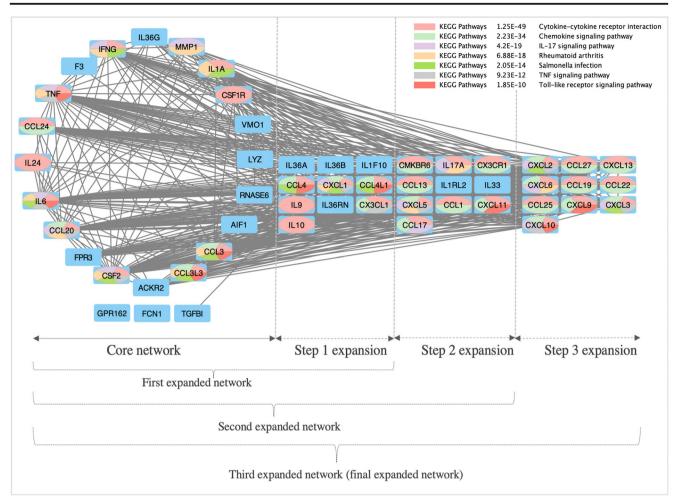
In order to identify experimentally validated upstream regulators that cause transcriptional changes similar to those induced by BCG, we queried the Connectivity Map (CMap) (27) database of the Broad Institute with BCG-CGS and identified proteins and small-molecule drugs that have strong connectivity scores with BCG (Fig. 1). The CMap approach enabled us to compare BCG-CGS with 'experimentally' predefined signatures of therapeutic compounds and genetic perturbations (i.e., over expression or knockdown) included in the CMap and ranked according

to a connectivity scores (ranging from +100 to -100), representing relative similarity to BCG-CGS. The connectivity score itself is derived using a nonparametric, rankbased, pattern-matching strategy based on the Kolmogorov-Smirnov statistics (28). All instances in the database are then ranked according to their connectivity scores with BCG-CGS; those at the top (+) are most strongly correlated to the query signature and looked at as BCG mimics, and those at the bottom (-) are most strongly anticorrelated and can reverse BCG's gene signature.

Our analysis identified three highly enriched classes of genetic knockdown (KD) perturbagens and one pharmacological class of drugs that have positive connectivity scores in alveolar A549 cells (i.e., caused similar transcriptional changes to those induced by BCG in alveolar A549 cells). These hits can be considered as BCG mimics capable of inducing transcriptional changes similar to those caused by the BCG vaccine. Therefore, we suggest that BCG mimics can be used as alternatives to BCG vaccination to promote long-lasting



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**Fig. 3** High-confidence expanded network for BCG-CGS. Nodes are color-coded using a split pie chart coloring scheme indicating pathway/gene set contribution to each node from the top 5 most enriched pathways/gene lists. Core network is composed of genes in the BCG-CGS that are not singletons. Step I expansion, added 10 additional nodes (i.e., genes) to the core network. Step 2 expansion, added another 10 nodes for the first expansion. Step 3 expansion, added another 10 nodes to the second expansion. Expansions were performed to see which pathways remained most statistically significant, and therefore are considered high confidence pathways.

beneficial effects on immune cells. The three enriched protein classes are: protein phosphatases (with best positive connection for PPP4C KD), histone deacetylases (with best positive connection for HDAC10 KD followed by HDAC11 KD), and mediator complex proteins (with best positive connection for MED6 KD followed by MED7 KD). Additionally, protein kinase C (PKC) activators were enriched as a drug class; and top three PKC activators with highest CMap connectivity scores to BCG-CGS were prostratin, phorbol-12-myristate-13-acetate, and ingenol. It is evident that all of the above four classes of proteins share one common feature: they participate in the transcriptional and metabolic regulation of immune cells in response to environmental cues including responses to pathogens (29–32). All top-scoring PKC activators from the CMap, are also known to have antiviral effects or affect T cell activation (33-37).

Remarkably, analyzing top ten CMap positive connections with BCG-CGS obtained from nine cell lines

indicated that two compounds are approved antiviral drugs: raltegravir (top 3rd positive connection, an HIV integrase inhibitor) and lopinavir (top 6th positive connection, an HIV protease inhibitor). More interestingly, emetine (top 4th positive connection) and lopinavir were recently validated to inhibit SARS-CoV-2 replication *in vitro* (38). We also found evidence in the biomedical literature indicating that MST-312 (39), narciclasine (40) and verrucarin-a (41) possess antiviral activities. All CMap hits are provided in Tables S5 and S8 (Supporting Information).

Initial reports from clinical studies evaluating the use of lopinavir in COVID-19 patients showed that the unbound lopinavir concentrations in the lungs were calculated to be subtherapeutic against SARS-CoV-2 (42,43). Another study found that the unbound drug concentrations of lopinavir are far from reaching the EC50 of SARS-CoV-2 (16.4  $\mu$ g/mL), although they clearly suffice to inhibit HIV-1 (44). The authors mentioned



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**Table I** Top Thirty Key Hubs Predicted by Causal Reasoning

TAFII70         Transcription factor         TAF6         +         14/14         6.10E-05         3           DSIPI (GILZ)         Transcription factor         TSC22D3         -         13/13         1.22E-04         2           Glutaredoxin I         Enzyme         GLRX         +         13/13         1.22E-04         3	Key Hub	Molecular function	Gene symbol	Predicted activity†	Correct/Total network predictions‡	Activity prediction <i>P</i> value*	Calculation distance§
DSIPI (GILZ)         Transcription factor         TSC22D3         -         14/14         6.10E-05         3           TARITO         Transcription factor         TAF6         +         14/14         6.10E-05         3           DSIPI (GILZ)         Transcription factor         TSC22D3         -         13/13         1.22E-04         2           Glutaredoxin I         Enzyme         GLRX         +         13/13         1.22E-04         3           Jagged I         Receptor ligand         JAG1         -         13/13         1.22E-04         3           RUNX2         Transcription factor         RUNX2         +         13/13         1.22E-04         2           NOTCHI (NICD)         Transcription factor         NOTCHI         +         16/17         1.37E-04         3           HAND1         Transcription factor         HAND1         -         12/12         2.44E-04         3           PRMT6         Enzyme         PRMT6         +         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           MSK1         Protein kinase         RPS6KAS         +         12/12 <t< td=""><td>HEYI</td><td>Transcription factor</td><td>HEYI</td><td>_</td><td>15/15</td><td>3.05E-05</td><td>3</td></t<>	HEYI	Transcription factor	HEYI	_	15/15	3.05E-05	3
TAFI170         Transcription factor         TAF6         +         14/14         6.10E-05         3           DSIPI (GILZ)         Transcription factor         TSC22D3         -         13/13         1.22E-04         2           Glutaredoxin I         Enzyme         GLRX         +         13/13         1.22E-04         3           Jagged I         Receptor ligand         JAG I         -         13/13         1.22E-04         3           RUNX2         Transcription factor         RUNX2         +         13/13         1.22E-04         3           NOTCHI (NICD)         Transcription factor         NOTCHI         +         16/17         1.37E-04         3           HANDI         Transcription factor         NOTCHI         +         16/17         1.37E-04         3           HANDI         Transcription factor         PRMT6         +         12/12         2.44E-04         3           PRMT6         Enzyme         PRMT6         +         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           MSKI         Protease         USP28         +         15/16         2.59E-04 <td>PHF20</td> <td>Binding protein</td> <td>PHF20</td> <td>+</td> <td>15/15</td> <td>3.05E-05</td> <td>3</td>	PHF20	Binding protein	PHF20	+	15/15	3.05E-05	3
DSIPI (GILZ)         Transcription factor         TSC22D3         -         13/13         1.22E-04         2           Glutaredoxin I         Enzyme         GLRX         +         13/13         1.22E-04         3           Jagged I         Receptor ligand         JAG I         -         13/13         1.22E-04         3           RUNX2         Transcription factor         RUNX2         +         13/13         1.22E-04         3           NOTCH I (NICD)         Transcription factor         NOTCHI         +         16/17         1.37E-04         3           HANDI         Transcription factor         HANDI         -         12/12         2.44E-04         3           PRMT6         Enzyme         PRMT6         +         12/12         2.44E-04         2           miR-129-1-3p         RNA         MIR129-1         -         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           MSKI         Protein kinase         RPS6KAS         +         12/12         2.44E-04         3           C-Fos         Transcription factor         VSP28         +         15/16         2.59E-04 <td>DSIPI (GILZ)</td> <td>Transcription factor</td> <td>TSC22D3</td> <td>_</td> <td>14/14</td> <td>6.10E-05</td> <td>3</td>	DSIPI (GILZ)	Transcription factor	TSC22D3	_	14/14	6.10E-05	3
Glutaredoxin I         Enzyme         GLRX         +         13/13         1.22E-04         3           Jagged I         Receptor ligand         JAG I         -         13/13         1.22E-04         3           RUNX2         Transcription factor         RUNX2         +         13/13         1.22E-04         2           NOTCHI (NICD)         Transcription factor         NOTCHI         +         16/17         1.37E-04         3           HANDI         Transcription factor         NOTCHI         +         16/17         1.37E-04         3           PRMT6         Enzyme         PRMT6         +         12/12         2.44E-04         3           PRMT6         Finzyme         PRMT6         +         12/12         2.44E-04         2           miR-129-1-3p         RNA         MIR129-1         -         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           MSK1         Protein kinase         RPS6KA5         +         12/12         2.44E-04         3           USP28         Protease         USP28         +         15/16         2.59E-04         3	TAFII70	Transcription factor	TAF6	+	14/14	6.10E-05	3
Jagged I         Receptor ligand         JAG I         -         13/13         1.22E-04         3           RUNX2         Transcription factor         RUNX2         +         13/13         1.22E-04         2           NOTCH I (NICD)         Transcription factor         NOTCH I         +         16/17         1.37E-04         3           HAND I         Transcription factor         HAND I         -         12/12         2.44E-04         3           PRNT6         Enzyme         PRNT6         +         12/12         2.44E-04         2           miR-129-1-3p         RNA         MIR129-I         -         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           HAND2         Transcription factor         HAND2         -         12/12         2.44E-04         3           MSKI         Protein kinase         RPS6KA5         +         12/12         2.44E-04         2           USP28         Proteise         LSP36         +         15/16         2.59E-04         3           USP4         Transcription factor         USTF         +         11/11         4.88E-04	DSIPI (GILZ)	Transcription factor	TSC22D3	_	13/13	1.22E-04	2
RUNX2         Transcription factor         RUNX2         +         13/13         1,22E-04         2           NOTCH1 (NICD)         Transcription factor         NOTCH1         +         16/17         1,37E-04         3           HAND1         Transcription factor         HAND1         -         12/12         2,44E-04         3           PRNT6         Enzyme         PRNT6         +         12/12         2,44E-04         2           miR-129-1-3p         RNA         MIR129-1         -         12/12         2,44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2,44E-04         3           HAND2         Transcription factor         HAND2         -         12/12         2,44E-04         3           MSK1         Protein kinase         RPS6KA5         +         12/12         2,44E-04         2           USP28         Proteise         LPS16         2,59E-04         3           c-Fos         Transcription factor         FOS         +         15/16         2,59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4,88E-04         3           UBF	Glutaredoxin I	Enzyme	GLRX	+	13/13	1.22E-04	3
NOTCHI (NICD) Transcription factor NOTCHI + 16/17 1.37E-04 3 HANDI Transcription factor HANDI - 12/12 2.44E-04 3 PRMT6 Enzyme PRMT6 + 12/12 2.44E-04 2 miR-129-1-3p RNA MIR129-1 - 12/12 2.44E-04 3 SOX10 Transcription factor SOX10 + 12/12 2.44E-04 3 HAND2 Transcription factor HAND2 - 12/12 2.44E-04 3 MSKI Protein kinase RPS6KA5 + 12/12 2.44E-04 3 MSKI Protein kinase RPS6KA5 + 12/12 2.44E-04 2 USP28 Protease USP28 + 15/16 2.59E-04 3 C-Fos Transcription factor FOS + 15/16 2.59E-04 3 UBF Transcription factor UBTF + 11/11 4.88E-04 3 MiR-520e-3p RNA MIRS20E - 11/11 4.88E-04 2 TMEMI19 Protein TMEMI19 + 11/11 4.88E-04 2 LRP16 Binding protein MACRODI + 11/11 4.88E-04 3 CaMK II gamma Protein kinase CAMK2G + 14/15 4.88E-04 3 MIR-4500 RNA MIR4500 - 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3 NOTCHI (NICD) Transcription factor NOTCHI + 14/15 4.88E-04 3	Jagged I	Receptor ligand	JAG I	_	13/13	1.22E-04	3
HAND I         Transcription factor         HAND I         -         12/12         2.44E-04         3           PRMT6         Enzyme         PRMT6         +         12/12         2.44E-04         2           miR-129-1-3p         RNA         MIR129-1         -         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           HAND2         Transcription factor         HAND2         -         12/12         2.44E-04         3           MSK1         Protein kinase         RPS6KA5         +         12/12         2.44E-04         2           USP28         Protease         USP28         +         15/16         2.59E-04         3           c-Fos         Transcription factor         FOS         +         15/16         2.59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         2           TMEM119         Protein         MES20E         -         11/11         4.88E-04         3	RUNX2	Transcription factor	RUNX2	+	13/13	1.22E-04	2
PRMT16         Enzyme         PRMT6         +         12/12         2.44E-04         2           miR-129-1-3p         RNA         MIR129-1         -         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           HAND2         Transcription factor         HAND2         -         12/12         2.44E-04         3           MSKI         Protein kinase         RPS6KA5         +         12/12         2.44E-04         2           USP28         Protease         USP28         +         15/16         2.59E-04         3           c-Fos         Transcription factor         FOS         +         15/16         2.59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           MIR-520e-3p         RNA         MIRS20E         -         11/11         4.88E-04         2           MEMI119         Protein         TMEMI19         +         11/11         4.88E-04         2           LRP16         Binding protein         MACRODI         +         14/15         4.88E-04         2	NOTCHI (NICD)	Transcription factor	NOTCHI	+	16/17	1.37E-04	3
miR-129-1-3p         RNA         MIR129-I         -         12/12         2.44E-04         3           SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           HAND2         Transcription factor         HAND2         -         12/12         2.44E-04         3           MSKI         Protein kinase         RPS6KAS         +         12/12         2.44E-04         2           USP28         Protease         USP28         +         15/16         2.59E-04         3           c-Fos         Transcription factor         FOS         +         15/16         2.59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           MiR-520e-3p         RNA         MIRS20E         -         11/11         4.88E-04         2           TMEM119         Protein         TMEM119         +         11/11         4.88E-04         3           LRP16         Binding protein         MACRODI         +         11/11         4.88E-04         2           LRP16         Binding protein         MACRODI         +         14/15         4.88E-04         3 <tr< td=""><td>HANDI</td><td>Transcription factor</td><td>HANDI</td><td>_</td><td>12/12</td><td>2.44E-04</td><td>3</td></tr<>	HANDI	Transcription factor	HANDI	_	12/12	2.44E-04	3
SOX10         Transcription factor         SOX10         +         12/12         2.44E-04         3           HAND2         Transcription factor         HAND2         -         12/12         2.44E-04         3           MSKI         Protein kinase         RPS6KAS         +         12/12         2.44E-04         2           USP28         Protease         USP28         +         15/16         2.59E-04         3           c-Fos         Transcription factor         FOS         +         15/16         2.59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         2           TMEM119         Protein         TMEM119         +         11/11         4.88E-04         3           LRP16         Binding protein         MACRODI         +         11/11         4.88E-04         3           CaMK II gamma         Protein kinase         CAMK2G         +         11/11         4.88E-04         3	PRMT6	Enzyme	PRMT6	+	12/12	2.44E-04	2
HAND2         Transcription factor         HAND2         -         12/12         2.44E-04         3           MSKI         Protein kinase         RPS6KA5         +         12/12         2.44E-04         2           USP28         Protease         USP28         +         15/16         2.59E-04         3           c-Fos         Transcription factor         FOS         +         15/16         2.59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           MiR-520e-3p         RNA         MIR520E         -         11/11         4.88E-04         2           TMEM119         Protein         TMEM119         +         11/11         4.88E-04         3           LRP16         Binding protein         MACRODI         +         11/11         4.88E-04         2           LRP16         Binding protein         MACRODI         +         14/15         4.88E-04         3           CaMK II gamma         Protein kinase         CAMK2G         +         14/15         4.88E-04         3	miR-129-1-3p	RNA	MIR129-1	_	12/12	2.44E-04	3
MSK I         Protein kinase         RPS6KAS         +         12/12         2.44E-04         2           USP28         Protein kinase         USP28         +         15/16         2.59E-04         3           c-Fos         Transcription factor         FOS         +         15/16         2.59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           miR-520e-3p         RNA         MIR520E         -         11/11         4.88E-04         2           TMEM119         Protein         TMEM119         +         11/11         4.88E-04         3           LRP16         Binding protein         MACRODI         +         11/11         4.88E-04         2           LRP16         Binding protein         MACRODI         +         14/15         4.88E-04         3           CaMK II gamma         Protein kinase         CAMK2G         +         11/11         4.88E-04         3           CaMK II gamma         Protein kinase         CAMK2G         +         14/15         4.88E-04         3           NOTCHI (NICD)         Transcription factor         NOTCHI         +         14/15         4.88E-04	SOX10	Transcription factor	SOX10	+	12/12	2.44E-04	3
USP28 Protease USP28 + 15/16 2.59E-04 3 c-Fos Transcription factor FOS + 15/16 2.59E-04 3 UBF Transcription factor UBTF + 11/11 4.88E-04 3 miR-520e-3p RNA MIR520E - 11/11 4.88E-04 2 TMEM119 Protein TMEM119 + 11/11 4.88E-04 3 LRP16 Binding protein MACRODI + 11/11 4.88E-04 2 LRP16 Binding protein MACRODI + 14/15 4.88E-04 3 CaMK II gamma Protein kinase CAMK2G + 11/11 4.88E-04 2 CaMK II gamma Protein kinase CAMK2G + 14/15 4.88E-04 3 miR-4500 RNA MIR4500 - 14/15 4.88E-04 3 NOTCH1 (NICD) Transcription factor NOTCH1 + 14/15 4.88E-04 2 miR-4516 RNA MIR4516 - 11/11 4.88E-04 3 NDPK B Protein kinase NME2 - 11/11 4.88E-04 3 KLF11 (TIEG2) Transcription factor KLF11 - 11/11 4.88E-04 2	HAND2	Transcription factor	HAND2	_	12/12	2.44E-04	3
c-Fos         Transcription factor         FOS         +         15/16         2.59E-04         3           UBF         Transcription factor         UBTF         +         11/11         4.88E-04         3           miR-520e-3p         RNA         MIR520E         -         11/11         4.88E-04         2           TMEM119         Protein         TMEM119         +         11/11         4.88E-04         3           LRP16         Binding protein         MACROD I         +         11/11         4.88E-04         2           LRP16         Binding protein         MACROD I         +         14/15         4.88E-04         3           CaMK II gamma         Protein kinase         CAMK2G         +         11/11         4.88E-04         2           CaMK II gamma         Protein kinase         CAMK2G         +         14/15         4.88E-04         3           NOTCHI (NICD)         Transcription factor         NOTCHI         +         14/15         4.88E-04         3           NOPK B         Protein kinase         NME2         -         11/11         4.88E-04         3           KLFII (TIEG2)         Transcription factor         KLFII         -         11/11         4.88E-0	MSKI	Protein kinase	RPS6KA5	+	12/12	2.44E-04	2
UBF         Transcription factor         UBTF         +         I I/I I         4.88E-04         3           miR-520e-3p         RNA         MIR520E         -         I I/I I         4.88E-04         2           TMEM119         Protein         TMEM119         +         I I/I I         4.88E-04         3           LRP16         Binding protein         MACROD I         +         I I/I I         4.88E-04         2           LRP16         Binding protein         MACROD I         +         I I/I I         4.88E-04         2           LRP16         Binding protein         MACROD I         +         I I/I I         4.88E-04         2           LRP16         Binding protein         MACROD I         +         I I/I I         4.88E-04         3           CaMK II gamma         Protein kinase         CAMK2G         +         I I/I I         4.88E-04         3           MOTCHI (NICD)         Transcription factor         NOTCHI         +         I I/I I         4.88E-04         3           NDPK B         Protein kinase         NME2         -         I I/I I         4.88E-04         3           KLF11 (TIEG2)         Transcription factor         KLF1 I         -         I I/I I	USP28	Protease	USP28	+	15/16	2.59E-04	3
miR-520e-3p       RNA       MIR520E       —       I I/I I       4.88E-04       2         TMEM119       Protein       TMEM119       +       I I/I I       4.88E-04       3         LRP16       Binding protein       MACROD I       +       I I/I I       4.88E-04       2         LRP16       Binding protein       MACROD I       +       I I/I I       4.88E-04       3         CaMK II gamma       Protein kinase       CAMK2G       +       I I/I I       4.88E-04       2         CaMK II gamma       Protein kinase       CAMK2G       +       I I/I I       4.88E-04       3         MiR-4500       RNA       MIR4500       —       I 4/I 5       4.88E-04       3         NOTCH I (NICD)       Transcription factor       NOTCH I       +       I 4/I 5       4.88E-04       2         miR-45 I6       RNA       MIR45 I6       —       I I/I I       4.88E-04       3         NDPK B       Protein kinase       NME2       —       I I/I I       4.88E-04       2         KLF I I (TIEG2)       Transcription factor       KLF I I       —       I I/I I       4.88E-04       2	c-Fos	Transcription factor	FOS	+	15/16	2.59E-04	3
TMEM119         Protein         TMEM119         +         I I/I I         4.88E-04         3           LRP16         Binding protein         MACROD I         +         I I/I I         4.88E-04         2           LRP16         Binding protein         MACROD I         +         I 4/I 5         4.88E-04         3           CaMK II gamma         Protein kinase         CAMK2G         +         I 1/I I         4.88E-04         2           CaMK II gamma         Protein kinase         CAMK2G         +         I 4/I 5         4.88E-04         3           miR-4500         RNA         MIR4500         -         I 4/I 5         4.88E-04         3           NOTCH1 (NICD)         Transcription factor         NOTCH1         +         I 4/I 5         4.88E-04         2           miR-4516         RNA         MIR4516         -         I I/I I         4.88E-04         3           NDPK B         Protein kinase         NME2         -         I I/I I         4.88E-04         2           KLF11 (TIEG2)         Transcription factor         KLF1I         -         I I/I I         4.88E-04         2	UBF	Transcription factor	UBTF	+	11/11	4.88E-04	3
LRP16       Binding protein       MACROD I       +       I I/I I       4.88E-04       2         LRP16       Binding protein       MACROD I       +       I 4/I 5       4.88E-04       3         CaMK II gamma       Protein kinase       CAMK2G       +       I I/I I       4.88E-04       2         CaMK II gamma       Protein kinase       CAMK2G       +       I 4/I 5       4.88E-04       3         miR-4500       RNA       MIR4500       -       I 4/I 5       4.88E-04       3         NOTCHI (NICD)       Transcription factor       NOTCHI       +       I 4/I 5       4.88E-04       2         miR-4516       RNA       MIR4516       -       I I/I I       4.88E-04       3         NDPK B       Protein kinase       NME2       -       I I/I I       4.88E-04       3         KLF11 (TIEG2)       Transcription factor       KLF1I       -       I I/I I       4.88E-04       2	miR-520e-3p	RNA	MIR520E	_	11/11	4.88E-04	2
LRP16       Binding protein       MACROD I       +       14/15       4.88E-04       3         CaMK II gamma       Protein kinase       CAMK2G       +       11/11       4.88E-04       2         CaMK II gamma       Protein kinase       CAMK2G       +       14/15       4.88E-04       3         miR-4500       RNA       MIR4500       -       14/15       4.88E-04       3         NOTCHI (NICD)       Transcription factor       NOTCHI       +       14/15       4.88E-04       2         miR-4516       RNA       MIR4516       -       11/11       4.88E-04       3         NDPK B       Protein kinase       NME2       -       11/11       4.88E-04       3         KLF11 (TIEG2)       Transcription factor       KLF11       -       11/11       4.88E-04       2	TMEM119	Protein	TMEM119	+	11/11	4.88E-04	3
CaMK II gamma       Protein kinase       CAMK2G       +       I I/I I       4.88E-04       2         CaMK II gamma       Protein kinase       CAMK2G       +       I 4/I 5       4.88E-04       3         miR-4500       RNA       MIR4500       -       I 4/I 5       4.88E-04       3         NOTCHI (NICD)       Transcription factor       NOTCHI       +       I 4/I 5       4.88E-04       2         miR-4516       RNA       MIR4516       -       I I/I I       4.88E-04       3         NDPK B       Protein kinase       NME2       -       I I/I I       4.88E-04       3         KLF1 I (TIEG2)       Transcription factor       KLF1 I       -       I I/I I       4.88E-04       2	LRP16	Binding protein	MACRODI	+	11/11	4.88E-04	2
CaMK II gamma       Protein kinase       CAMK2G       +       14/15       4.88E-04       3         miR-4500       RNA       MIR4500       -       14/15       4.88E-04       3         NOTCHI (NICD)       Transcription factor       NOTCHI       +       14/15       4.88E-04       2         miR-4516       RNA       MIR4516       -       11/11       4.88E-04       3         NDPK B       Protein kinase       NME2       -       11/11       4.88E-04       3         KLF11 (TIEG2)       Transcription factor       KLF11       -       11/11       4.88E-04       2	LRP16	Binding protein	MACRODI	+	14/15	4.88E-04	3
miR-4500       RNA       MIR4500       -       14/15       4.88E-04       3         NOTCH1 (NICD)       Transcription factor       NOTCH1       +       14/15       4.88E-04       2         miR-4516       RNA       MIR4516       -       11/11       4.88E-04       3         NDPK B       Protein kinase       NME2       -       11/11       4.88E-04       3         KLF11 (TIEG2)       Transcription factor       KLF11       -       11/11       4.88E-04       2	CaMK II gamma	Protein kinase	CAMK2G	+	11/11	4.88E-04	2
NOTCHI (NICD)       Transcription factor       NOTCHI       +       14/15       4.88E-04       2         miR-4516       RNA       MIR4516       -       11/11       4.88E-04       3         NDPK B       Protein kinase       NME2       -       11/11       4.88E-04       3         KLF11 (TIEG2)       Transcription factor       KLF11       -       11/11       4.88E-04       2	CaMK II gamma	Protein kinase	CAMK2G	+	14/15	4.88E-04	3
miR-4516       RNA       MIR4516       -       I 1/I 1       4.88E-04       3         NDPK B       Protein kinase       NME2       -       I 1/I 1       4.88E-04       3         KLF11 (TIEG2)       Transcription factor       KLF11       -       I 1/I 1       4.88E-04       2	miR-4500	RNA	MIR4500	_	14/15	4.88E-04	3
NDPK B         Protein kinase         NME2         -         I I/I I         4.88E-04         3           KLFI I (TIEG2)         Transcription factor         KLFI I         -         I I/I I         4.88E-04         2	NOTCHI (NICD)	Transcription factor	NOTCHI	+	14/15	4.88E-04	2
KLFII (TIEG2) Transcription factor KLFII – II/II 4.88E-04 2	miR-4516	RNA	MIR4516	_	11/11	4.88E-04	3
	NDPK B	Protein kinase	NME2	_	11/11	4.88E-04	3
miR-320d RNA MIR320D1 - 14/15 4.88E-04 3	KLFII (TIEG2)	Transcription factor	KLFII	_	11/11	4.88E-04	2
	miR-320d	RNA	MIR320D1	_	14/15	4.88E-04	3

<sup>†</sup> Predicted activity of the key hub by causal reasoning is denoted by – if the hub is inhibited, and denoted by + if the hub is activated

that approximately 60- to 120-fold higher concentrations than those found in COVID-19 patients treated with lopinavir-ritonavir, are required to reach the assumed EC50 at trough levels, making effective treatment of COVID-19 with lopinavir and ritonavir at the currently used doses unlikely (44).

In order to prioritize high confidence BCG genetic mimics, we integrated hypotheses derived independently from the CMap with those predicted by causal reasoning, and accepted common hits only (i.e., CMap positive connections with BCG-CGS that are also predicted as beneficial drug targets by causal reasoning). This analysis resulted in 30 high confidence common hits reported in Table S9 (Supporting Information).

# Can We Validate Functional Connections Between BCG and SARS-CoV-2?

We tested whether BCG-CGS, CMap positive connections, or predicted key hubs will have any impact on COVID-19 by identifying overlaps with SARS-CoV-2 interactome, i.e., human proteins that were experimentally validated to interact with SARS-CoV-2 and extracted from two recent reports (45,46). This analysis (Fig. 4a) validated 3 protein hits to have physical links to SARS-CoV-2. The three proteins are transcribed by BRD4, PRKACA and SIRT5; they all were positive connections from the CMap, predicted as statistically



<sup>‡</sup> Correct/total network predictions: correct for the genes in the dataset predicted correctly; total for the total number of genes in the causal reasoning network § Calculation distance: Using causal reasoning one-step key hubs are defined as statistically significant transcriptional factors that are associated with experimental differential expressed genes regulation. Two-step and three-step key hubs are distant key hubs that regulate one-step transcriptional factors

<sup>\*</sup>P-value calcualted for the polynomial test

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significant key hubs, and were also validated as SARS-CoV-2 interacting proteins (15).

Additionally, 14 high-confidence CMap positive connections, were validated to make physical interactions with SARS-CoV-2 proteins. These proteins are: PSEN2, PABPC1, HMOX1, CIT, PLAT, IGF2R, RIPK1, NDUFS3, NDUFA5, GGH, NEU1, SCARB1, CSNK2B, F2RL1. And two positive connections, MARK2 and MARK3, were reported to have interactions with corona viruses (45). Predicted causal key hubs, SIGMAR1 and GNB1, were also validated to have physical links to SARS-CoV-2 (15), and a third key hub PPIA was known as a human protein interacting with proteins from corona viruses (45).

Additionally, we mined the biomedical literature to identify evidence for linking BCG small molecule mimics with SARS-CoV-2, corona viruses or viral infections in general. We found that two out of ten top positive compound connections (emetine and lopinavir), were recently validated to inhibit SARS-CoV-2 replication *in vitro* (38). Other compounds we found to inhibit the growth of corona viruses, or had general antiviral activities (Table II).

# **DISCUSSION**

Previous peer-reviewed reports indicated that BCG's nonspecific effects on the immune system, can reduce all-cause child mortality (47), protect individuals from numerous viral infections (48-52), and it can even enhance the efficiency of some viral vaccines (53-55). Recently, several peer-reviewed studies have pointed to a striking correlation between universal BCG vaccination policies and reduced COVID-19 mortality (23). However, most epidemiological studies identified this correlation without acknowledging other important study confounders like social, economic, and demographic differences between countries. Lately, Escobar et al. mitigated multiple confounding factors for the first time and still observed several significant associations between BCG vaccination and reduced COVID-19 deaths (21). The authors of this study highlighted the need for mechanistic studies behind the effect of BCG vaccination on COVID-19, and for clinical evaluation of the effectiveness of BCG vaccination to protect from severe COVID-19. Earlier studies suggested that the documented beneficial off-target effects of BCG in protecting from non-TB infections, including perhaps COVID-19, involve a potentiation of innate immune responses through epigenetic mechanisms (56-58).

To our knowledge, we report here on the first study providing a mechanistic insight to explain the relationships between BCG and COVID-19 at the molecular and systems biology levels as well as extend this insight toward proposing several BCG mimetics among known drugs as candidates for repurposing against the disease. Our results indicate that

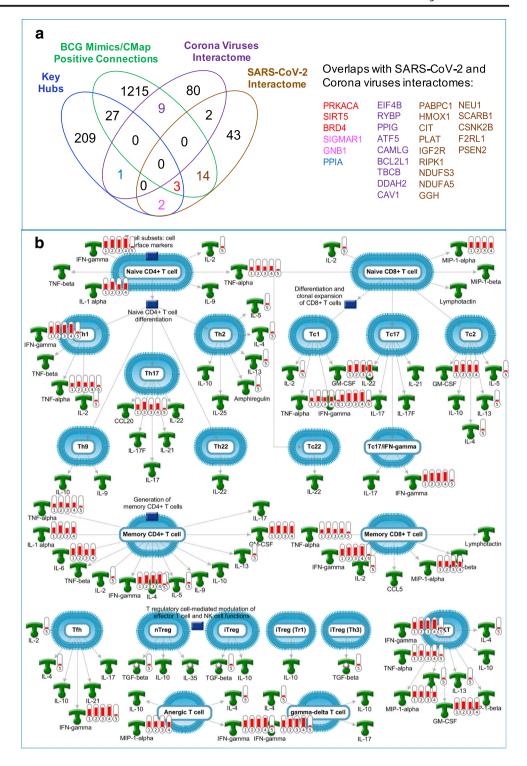
BCG-CGS, key regulatory hubs and BCG-mimics identified from the CMap enrich common biological pathways important for key viral processes such as RNA synthesis and processing, virus-host interactions, positive regulation of viral genome replication, and they are also important for the immune response mounted against the virus. Supporting evidence from the biomedical literature confirms that BCG has many beneficial 'off-target' effects that can protect humans from emerging novel pathogens by boasting their innate immune responses (59). Our studies suggest that BCG promotes a wide-range of transcriptional and metabolic changes, including beneficial gene commensalism, that have been shown to reduce mortality and morbidity from non-TB infectious diseases (51,52). We show that BCG can produce these protective 'off target' effects mainly by increasing the production of thymus-generated short-lived undifferentiated CD4+ cells known as naive T cells (Th<sub>0</sub>), and triggering their differentiation into the long-lived mature naive T cells (MNTs), such as CD4+ and CD8+ T cells (60). Interestingly, a very recent study published in Science (61) showed that many unexposed patients (20–50%) carry selective and cross-reactive SARS-CoV-2 T cell epitopes protecting patients against severe infection. Another recent study in Cell (62) reported similar observation of strong SARS-CoV-2 selective memory T cell immunity (reminiscent of the functional patterns observed after successful vaccine immunizations) in patients with asymptomatic or mild infections. Although these studies make no connection to any previous BCG vaccination as a source of selective epitopes, we observe that these observations are consistent with our mechanistic hypothesis concerning the protective effect of BCG against COVID-19. These conclusions are supported by the enrichment results produced using the 'Compare Experiment' algorithm in MetaCore from Clarivate Analytics, which looks for significant coordinated gene expression effects across all experiments to test whether the pathway is being up- or down-regulated in a manner that is unlikely to be accounted for by random chance. The top enriched pathway map, with upregulated genes in response to BCG, is 'Immune response T cell subsets: secreted signals' (Fig. 4b). A recent study showing that SARS-CoV-2 reshapes central cellular pathways, such as translation, splicing, carbon metabolism and nucleic acid metabolism (53), provides further support for this observation.

Naturally, bioinformatics techniques relying on gene expression, pathway over-representation and network biology have some limitations and biases: 1) results are impacted by the user-selected cut-off thresholds used to determine significant genes, which could make the results user-dependent (63); 2) all components in the pathway are given equal weights without paying attention to the nature of the interactions between the different components (64); 3) there are underlying assumptions that pathways are independent of each other, contrary to the fact that pathways



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Fig. 4 (a) A venn diagram showing overlaps between BCG genetic mimics and key hubs with SARS-CoV-2 and Corona viruses interactomes. (b) Top "pathway map" with the highest level of enrichment by genes in BCG-CGS. This map is generated using MetaCore from Clarivate Analytics. Red thermometers indicate genes overexpressed in response to BCG treatment, and the hight of the red bars is representative of the differential gene expression level (i.e., log<sub>2</sub> values of the fold change). The numbers under the thermometers I-5 refer to the experiment number: 1) gene expression on day 0 in response to BCG vaccination to a BCG-naïve population on day 1; 2) gene expression on day 0 in response to BCG re-vaccination to a previously vaccinated population; 3) gene expression on day 14 in response to BCG vaccination to a BCG-naïve population; 4) gene expression on day 14 in response to BCG revaccination to a previously vaccinated population, and 5) positive connections from the connectivity map, and the red bar in the thermometer number 5 represents presence of the gene only.



cross-talk and overlap (65); 4) assuming independence between genes may result in false positive predictions of highly enriched genes or gene sets (66); 5) these methods are incapable of modeling an organism's biology as a dynamic system, and cannot predict changes in the system due genetic mutations or environmental changes (67); 6) most pathway knowledge databases are built by curating

experiments performed in different cell types at different time points under different conditions, so they are missing condition- and cell-specific information (64).

In order to mitigate some of the aforementioned limitations, we used a consensus gene signature since it is more stable than other gene expression signatures, we paired overrepresentation pathway analysis with causal reasoning to



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**Table II** Small-Molecule BCG Mimics with Potential Antiviral Effects

Compound	Score†	Description	Validation‡
prostratin	98.65	PKC activator	Antiviral (34)
ingenol	98.52	PKC activator	Antiviral (33)
raltegravir	97.85	HIV integrase inhibitor	Antiviral (93)
emetine	97.25	Protein synthesis inhibitor	SARS-CoV-2 (38)
phorbol-12-myristate-13-acetate	96.72	PKC activator	Antiviral (36,37)
mebendazole	95.32	Tubulin inhibitor	Antiviral (94)
lopinavir	95.06	HIV protease inhibitor	SARS-CoV-2 (38,39)
MST-312	95.04	Telomerase inhibitor	Antiviral (36,37,39)
narciclasine	94.71	Coflilin signaling pathway activator	Antiviral (40)
verrucarin-a	94.51	Protein synthesis inhibitor	Antiviral (95)
anisomycin	94.40	DNA synthesis inhibitor	Corona viruses (96)
azacitidine	94.29	DNA methyltransferase inhibitor	Antiviral (97)
cytochalasin-b	93.90	Microtubule inhibitor	Antiviral (98)
cephaeline	93.88	Protein synthesis inhibitor	Antiviral (99)
homoharringtonine	93.42	Protein synthesis inhibitor	Antiviral (100)
ruxolitinib	92.81	JAK inhibitor	COVID-19 CT§
HU-211	92.64	Glutamate receptor antagonist	Unknown
vinblastine	92.36	Microtubule inhibitor	Unknown
RO-28-1675	92.12	Glucokinase activator	Unknown
vincristine	91.61	Tubulin inhibitor	Unknown

†Score refer to the CMap score. It represents the level of similarity between transcriptional effects induced by BCG and each of the compounds

‡ Validation refers to the presence of any supporting evidence from the biomedical literature that the predicted BCG mimics have any antiviral activities. Antiviral means there is evidence that the compound is used as or has antiviral activity; SARS-CoV-2 means that the compound should antiviral activity against SARS-CoV-2; Corona viruses means that the compound showed antiviral activity against corona viruses other than SARS-CoV-2

§ COVID-19 CT: there is evidence that the compound is being tested in clinical trials for COVID-19. There are 12 Studies found for Ruxolitinib in COVID-19 on clinicaltrials.gov.

predict protein activities based on the nature of interactions between upregulated or downregulated genes, and we also integrated results from several bioinformatics methods such as causal reasoning and CMap predictions to prioritize common hypotheses.

A recent publication (68) in Lancet has questioned whether BCG's effects can last for a long time. Our top enriched pathway map (Fig. 4b) indicates that BCG's effects can be longlasting if the effects were exerted on thymus-generated Th<sub>0</sub> cells, which can occur to a greater extent very early in life before reaching thymic involution by puberty (69). This pathway map indicates that BCG is capable of affecting both the numbers and the types of produced innate immune cells, as well as their maturation to long-lived memory T cells (i.e., what is known as trained immunity). This is very significant in the context of BCG's protective effects from SARS-CoV-2 and other emergent novel viruses where the individual's ability to eradicate such viruses is dictated by the number and diversity of naive T cell reservoir (70,71). Our analysis suggest that BCG may protect individuals from novel pathogens by priming their trained immunity to fight such pathogens, including SARS-CoV-2.

Supporting evidence for this hypothesis is found in the literature (72) indicating that the protective effects of the BCG against TB, can last from 15 to 60 years after vaccination (72,73), with longer lasting effects when the vaccine is administered during the first year of life (74,60). A recent study indicated that "school-aged BCG vaccination offered moderate protection against tuberculosis for at least 20 years, which is much longer than previously thought" (60,72). Another 60year follow-up study, showed that BCG vaccine efficacy persisted for 50 to 60 years after a single dose of BCG (60). Of special interest is a recent study that showed that mucosal vaccination resulted in an increased frequency of antigenspecific lung tissue-resident CD4+ T cells that provide longterm immunity (75). These studies serve as additional evidence from the literature supporting our claim that a single dose of an 'effective' BCG vaccination to infants can have a very long duration of protection against pathogens including SARS-CoV-2.

Our findings provided systems biology support for using BCG to protect from the severe consequences of COVID-19. BCG is currently on WHO's List of Essential Medicines; it is considered one of the safest and most effective medicines



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needed in a health system. There is also evidence indicating that BCG can improve the response to vaccines directed against viral infections (48,53–55), which may prove useful when SARS-CoV-2-specific vaccines become available. Therefore, we suggest that administering the BCG vaccine to all newborns may protect them from the infection by SARS-CoV-2 and other emerging pathogens.

Since this is an approved vaccine for TB, it can directly enter Phase III testing for the protection from COVID-19 caused fatalities. However, we caution that running these experiments during an active COVID-19 outbreak, might expose participants to aggravated immune responses if they contract COVID-19 during the study. We also advise that clinical study design takes into account several factors that are known to affect the performance of BCG vaccine, such as: the age of the participants, geographies, ethnicities, route of administration and the mycobacterium strain used in the vaccine. It is equally important to run experimental validation studies, to evaluate the effects of BCG mimics, in preventing COVID-19 or for treating urological cancers.

# **CONCLUSION**

Our results provide systems biology support for using BCG and small-molecule BCG mimics as putative vaccine and drug candidates against emergent viruses including SARS-CoV-2. Of course, any practical actions to repurpose this vaccine as a means of protection against SARS-CoV-2, or other novel viruses, should be preceded by the successful *in vitro* and animal experimentation. We also caution that previous studies showed that the protective effects of BCG were found to be weaker when the vaccine was given after the first year of life and particularly after puberty (68).

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# **AUTHOR CONTRIBUTIONS**

R. Hajjo generated the idea, designed the workflow, generated content, performed data analysis and wrote the manuscript. A. Tropsha was engaged in the critical discussion of the study design, recommended some studies, and edited the manuscript.

**Data Availability** BCG dataset, GSE58636 is available on the Gene Expression Omnibus GEO: https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE58636

#### **COMPLIANCE WITH ETHICAL STANDARDS**

Conflict of Interest The authors declare no competing interests.

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