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Human papillomavirus types in invasive cervical cancer worldwide: a meta-analysis

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This study investigated regional variations in the contribution made by different human papilloma (HPV) types to invasive cervical cancer (ICC). A total of 85 studies using polymerase chain reaction to estimate HPV prevalence in ICC were identified. Data on HPV prevalence were extracted separately for squamous cell carcinoma (SCC) and for adeno- and adenosquamous-carcinoma (ADC). A total of 10 058 cases (8550 SCC, 1508 ADC) were included in pooled analyses. The most common HPV types in ICC were, in order of decreasing prevalence, HPV16, 18, 45, 31, 33, 58, 52, 35, 59, 56, 6, 51, 68, 39, 82, 73, 66 and 70. In SCC, HPV16 was the predominant type (46–63%) followed by HPV18 (10-14%), 45 (2-8%), 31 (2-7%) and 33 (3-5%) in all regions except Asia, where HPV types 58 (6%) and 52 (4%) were more frequently identified. In ADC, HPV prevalence was significantly lower (76.4%) than in SCC (87.3%), and HPV18 was the predominant type in every region (37-41%), followed by 16 (26-36%) and 45 (5-7%). The overall detection of HPV DNA was similar in different regions (83-89%). A majority of ICC was associated with HPV16 or 18 in all regions, but approximately a quarter of all ICC cases were associated with one of 16 other HPV types, their distribution varying by region.

British Journal of Cancer (2003) **88,** 63–73. doi:10.1038/sj.bjc.6600688 www.bjcancer.com © 2003 Cancer Research UK

Keywords: human papillomavirus; cervical carcinoma; squamous cell carcinoma; adenocarcinoma; epidemiology; literature review

Epidemiological studies have clearly established human papillomavirus (HPV) infection as the central cause of invasive cervical cancer (ICC). This is the second most common cancer among women worldwide and the most common female cancer in large areas of the developing world where an estimated 80% of new cases arise (Parkin *et al*, 1999). Studies in 22 countries, coordinated by the International Agency for Research on Cancer (IARC), identified HPV DNA in almost all (99.7%) (of about 1000) cases of cervical cancer (Walboomers *et al*, 1999).

Approximately 40 distinct HPV types are known to infect the genital tract and epidemiological studies to date suggest that at least 14 of these, called oncogenic or high-risk (HR) types, are significantly associated with progression to ICC (Bosch *et al*, 1995). Most of these HR types are phylogenetically related to either HPV16 (31, 33, 35, 52 and 58) or HPV18 (39, 45, 59 and 68) (Chan *et al*, 1995). Limited evidence suggests that their distribution may vary by region (Bosch *et al*, 1995).

HPV vaccines hold great promise to reduce the global burden of ICC any potential vaccine be multivalent since prior infection with one type does not appear to decrease the risk of infection by another HPV type (Koutsky *et al*, 2002; Combita *et al*, 2002; Liaw *et al*, 2001). In this however, to collate all relevant published data to identify the most prevalent HPV types associated with ICC worldwide and within five geographic regions.

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MATERIALS AND METHODS

Study selection

Source material was selected from citations listed in Medline and ISI Current Contents databases and from references cited in the selected papers. Key search terms included: cervical cancer, HPV, human, female, and polymerase chain reaction (PCR). The review was limited to studies that included a minimum of 20 ICC cases; carcinomas in situ were excluded. Studies had to provide a clear description of the use of PCR-based assays to identify HPV DNA. Studies using nonamplified hybridisation methods only were excluded based on the reduced sensitivity of such methods in comparison to PCR (Gravitt et al, 1991; Schiffman et al, 1991; Guerrero et al, 1992). Furthermore, articles were only included if type-specific prevalence of at least one HPV type other than HPV6, 11, 16 or 18 was reported. For articles where study methods suggested that additional type-specific data were available, these data were requested from the authors (Yang et al, 1997; Eluf-Neto et al, 1994; Chaouki et al, 1998; Meyer et al, 1998; Chen et al, 1999; Lin et al, 2001). If data or data subsets had been published in more than one article, only the publication with the largest sample size was included.

Data abstraction

For each study, the following key information was extracted: country of sample; sample size; mean age; study year; distribution of cases by histological type; type of cervical specimen (e.g., fresh/fixed biopsies or exfoliated cells) and PCR primers used to detect HPV positive samples; type-specific and overall prevalence of HPV

Epidemio

infection. Where available, data on HPV-specific prevalence were extracted independently for squamous cell carcinoma (SCC) and for adeno- and adenosquamous carcinoma (henceforth collectively termed ADC). Where histology-specific HPV prevalence was not reported, cases were classified as being of 'unspecified' histology. Each study was classified into one of five geographical regions: Africa, Asia, Europe, North America and Australia, or South and Central America. For studies comparing HPV prevalence across regions (Munoz *et al*, 1992; Bosch *et al*, 1995; Sebbelov *et al*, 2000), data were separated into their regional components.

Studies included

Of studies published up to February 2002 on *Medline* identified by our search criteria, 82 qualified for inclusion (no additional studies were included from *ISI Current Contents*). Three studies were conference abstracts containing the detailed information required for inclusion (Illades-Aguiar *et al*, 2000; Nindl *et al*, 2001; Rabelo-Santos *et al*, 2001). In the course of contacting authors, additional data became available for two studies expanded since the original publication (Burger *et al*, 1996; Andersson *et al*, 2001). Detailed information on the design of each of the 85 included studies is listed in the appendix.

Estimation of type-specific prevalence

HPV prevalence data were expressed as percentages of all cases tested for HPV. Multiple infections (3.7% of all ICC cases) were separated into constituent types, thus type-specific prevalence represents that in either single or multiple infections. Cases with specimens considered to be inadequate for PCR testing were excluded. Type-specific prevalence is presented for the 18 most common HPV types as identified by this review (HPV types 6, 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68, 70, 73 and 82 also known as MM4, W13B or IS39) in order of descending prevalence for each subgroup analysis. Consensus PCR primers My09/11 (Bernard et al, 1994), GP5⁺/6⁺ (Chaouki et al, 1998) and SPF10 (Kleter et al, 1999) were considered to amplify all 18 HPV types, L1C1/L1C2 (Nakagawa et al, 1996) to amplify all types, but HPV73 and 82, GP5/6 to amplify types 6, 11, 16, 18, 31, 33, 35 and 45 only (Roda Husman et al, 1995), and pU1M/2R (Harima et al, 2002) to amplify types 6, 16, 18, 31, 33, 35, 52, 56, 58 and 59 only. For other consensus and type-specific PCR primers, only those HPV types specified in the individual reports were considered amplifiable. For HPV-specific prevalence, only studies testing for a particular HPV type contribute to the analysis for that type, and therefore sample size varies between the type-specific analyses.

Statistical analyses

Sources of variation in overall HPV prevalence were investigated by unconditional multiple logistic regression analysis (Breslow and Day, 1980). The final model included the following sources of variation: geographical region, histological type of ICC, type of specimen for HPV DNA testing, and type of PCR primers used. Mean age and study year were found not to be significantly related to overall HPV prevalence. Adjustment of overall HPV prevalence for these variables was done using the *adjust* command in Stata version 7.0, based on probability estimates from the logistic regression model. Confidence intervals for overall HPV prevalence were calculated assuming the nonindependence of cases within the same study using the *cluster* option in Stata (White, 1980). *P*-values comparing the prevalence of particular HPV types in subsets of ICC cases refer to χ^2 tests.

RESULTS

Meta-analysis of overall HPV prevalence

A total of 10 058 ICC cases from the 85 identified studies were included in this meta-analysis of HPV prevalence (Table 1). A majority of cases came from studies performed in Asia (31%) and Europe (33%), with African studies representing the smallest proportion of cases (6%). HPV prevalence was reported stratified by histological type for 73% of the cases: 5825 SCC cases and 1508 ADC cases. In total, 12 studies included only SCC and seven studies included only ADC.

Adjusted overall HPV prevalence ranged from 79.3% in Asia to 88.1% in North America and Australia, but did not differ significantly between regions (Table 2). HPV DNA was significantly less likely to be detected in ADC (76.5%) than in SCC (87.3%) (P<0.001). DNA detection in ICC of unspecified histology (89.2%) was similar to that in SCC.

Adjusted HPV prevalence was significantly higher from studies testing both cells and biopsies for HPV DNA (92.5%) than from studies testing either cervical exfoliated cells (78.9%) or fixed biopsies (83.3%) only. For PCR primers, highest HPV prevalence

Table I Region- and histology-specific distribution of included studies and ICC cases

				Number (%) of cases with histology-specifi HPV data									
Region	No. of studies	Countries represented	No. of cases	scc	ADC	Unspecified							
Africa	6	Algeria, Benin, Guinea, Mali, Morocco, Senegal, South Africa, Tanzania, Uganda	609	204 (33.5)	21 (3.4)	384 (63.1)							
Asia	28	Mainland China, India, Indonesia, Japan, Korea, Malaysia, Philippines, Taiwan, Thailand	3091	2273 (73.5)	381 (12.3)	437 (14.1)							
Europe	32	Austria, Czech Republic, Denmark, Finland, France, Germany, Greece, Greenland, Holland, Hungary, Ireland, Italy, Norway, Poland, Russia, Sweden, UK	3336	2010 (60.3)	603 (18.1)	723 (21.7)							
North America and Australia	13	Australia, Ćanada, UŚA	1562	914 (58.5)	450 (28.8)	198 (12.7)							
South and Central America	12	Argentina, Bolivia, Brazil, Chile, Colombia, Costa Rica, Cuba, Honduras, Mexico, Panama, Paraguay, Peru	1460	424 (29.0)	53 (3.6)	983 (67.3)							
Total	85		10 058	5825 (57.9)	1508 (15.0)	2725 (27.1)							

HPV=human papillomavirus; ICC=invasive cervical cancer; SCC=squamous cell carcinoma; ADC=adeno/adenosquamous carcinoma.

Table 2 Prevalence of HPV by region, histological type, HPV DNA specimen and PCR primers used

Region Africa 6 609 88.8 86.5 Asia 28 3091 83.1 79.3 Europe 32 3336 85.9 86.7 North America and Australia 13 1562 87.5 88.1 South and Central America 12 1460 89.3 87.7 Histological type Squamous cell carcinoma 47 5825 86.9 87.3 Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2 Cervical exfoliated cells 13 1444 80.1 78.9	5% confidence intervals
Asia 28 3091 83.1 79.3 Europe 32 3336 85.9 86.7 North America and Australia 13 1562 87.5 88.1 South and Central America 12 1460 89.3 87.7 Histological type Squamous cell carcinoma 47 5825 86.9 87.3 Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	
Europe 32 3336 85.9 86.7 North America and Australia 13 1562 87.5 88.1 South and Central America 12 1460 89.3 87.7 Histological type Squamous cell carcinoma 47 5825 86.9 87.3 Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(76.4-92.7)
North America and Australia 13 1562 87.5 88.1 South and Central America 12 1460 89.3 87.7 Histological type Squamous cell carcinoma 47 5825 86.9 87.3 Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(73.7–84.0)
South and Central America 12 1460 89.3 87.7 Histological type Squamous cell carcinoma 47 5825 86.9 87.3 Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(82.5–90.0)
Histological type Squamous cell carcinoma 47 5825 86.9 87.3 Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(83.6–91.5)
Squamous cell carcinoma 47 5825 86.9 87.3 Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(83.1–91.2)
Adeno(squamous) carcinoma 45 1508 76.7 76.5 Unspecified 48 2725 89.0 89.2 HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	
Unspecified 48 2725 89.0 89.2 HPV DNA specimen *** *** *** Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(84.8-89.5)
HPV DNA specimen Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(72.3–80.3)
Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	(85.1–92.3)
Fixed biopsies 34 3324 84.3 83.3 Fresh biopsies 27 3992 88.4 87.8 Unspecified biopsies 8 496 87.1 87.2	
Unspecified biopsies 8 496 87.1 87.2	(78.9-86.9)
	(84.9–90.2)
Can ited as fallowed as la 12 1444 00 L 70 0	(82.3–90.9)
Cervical extollated cells 15 1444 00.1 70.7	(68.5–86.5)
Cells and biopsies 3 802 93.5 92.5	(87.3–95.7)
Primers	
MY09/II 3I 4355 85.9 83.3	(80.1-86.0)
GP5/6 6 506 80.8 77.8	(64.9–87.0)
GP5+/6+ 14 1681 92.2 90.1	(85.0–93.6)
SPF10 3 275 96.7 97.2	(87.9–99.4)
PUIM/2R 6 376 80.9 79.4	(68.8–87.0)
LICI/C2 5 655 91.2 88.0	(77.2–94.1)
Combination 9 1351 88.4 86.4	(77.9–92.0)
Other 4 166 84.3 89.3	(75.2–95.9)
TS-PCR only 7 693 73.6 74.7	(63.8–83.2)

^aAdjusted for histological type, region, HPV DNA specimen and PCR primers.

was obtained in studies using SPF10 primers (97.2%) and the lowest in studies using type-specific PCR (TS-PCR) only (74.7%). Adjusted overall HPV prevalence varied between 77.8 and 90.1% for other primer sets, but these differences were not statistically significant.

Meta-analysis of HPV type-specific prevalence

Owing to their similar overall and type-specific HPV prevalence, ICC of unspecified histology were combined with SCC for comparison of HPV type-specific prevalence by histological type (Figure 1). The most common HPV types identified were, in order of decreasing prevalence, HPV16, 18, 45, 31, 33, 58, 52, 35, 59, 56, 6, 51, 68, 39, 82, 73, 66 and 70. Other HPV types were detected in no more than 0.2% of ICC cases. There was considerable variation in HPV-specific prevalence between SCC and ADC. HPV16 was identified more often in SCC (55.2%) than in ADC (31.3%) (P<0.001). The same was found for the HPV16 phylogenetically related types 31, 33, 52 and 58 (P<0.001), but not 35. Conversely, HPV18 was more prevalent in ADC (37.7%) than in SCC (12.3%) (P<0.001). The HPV18 phylogenetically related type 45 was also more prevalent in ADC (5.8%) than in SCC (3.4%) (P=0.04).

Comparison of HPV-specific prevalence in SCC by region is shown in Figure 2. In SCC, HPV16 was the predominant type in all regions studied, varying from 45.9% in Asia to 62.6% in North America and Australia. HPV18 was found consistently in 10–14% of SCC cases. In most regions, HPV45 (2–8%), 31 (2–7%) and 33 (3–5%) were the most prevalent types in SCC after types 16 and 18. In cases from Africa, the prevalence of HPV45 (8.0%) was more than twice that of either 31 (2.7%) or 33 (3.2%). In cases from Asia, HPV58 (5.8%) and 52 (4.4%) were found more commonly than HPV45, 31 and 33. Other HPV types varied considerably in their

prevalence from region to region, but accounted for no more than 2% of ICC cases from any region.

Sufficient ADC-specific data existed for the comparison of HPV-specific prevalence across Asia, Europe and North America and Australia (Figure 3). HPV18 was the predominant type (37.7%), found consistently in 37–41% of ADC cases in these regions, with HPV16 accounting for a smaller proportion (26–36%). HPV45 was the third most prevalent in each region, present in 5–7% of ADC cases *vs* only in 2–4% of SCC cases from these regions. The HPV16 phylogenetically related types 31, 33, 52 and 58 (but not 35) were all less prevalent in ADC cases than in SCC cases from each region.

DISCUSSION

Two-thirds of ICC cases included in this meta-analysis were associated with HPV16 (51.0%) or 18 (16.2%) infection. However, more than 16 other HPV types were also associated with ICC, of which the most prevalent were types 45, 31, 33, 58 and 52 (collectively accounting for 18.3% of cases). The HPV16 family of viruses were more commonly found in SCC than ADC, whereas the HPV18 family were more common in ADC.

Geographical region

Overall detected HPV prevalence varied little between geographical regions (83–89%), but was low compared to the almost 100% HPV prevalence identified in studies using the most sensitive HPV detection methods (Walboomers *et al*, 1999). This reflects the fact that many studies used HPV DNA detection strategies of suboptimal sensitivity. When comparing prevalence by region



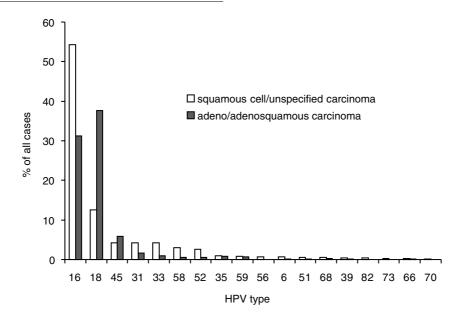


Figure I Type-specific prevalence of HPV in 10058 worldwide cases of invasive cervical cancer by histological type.

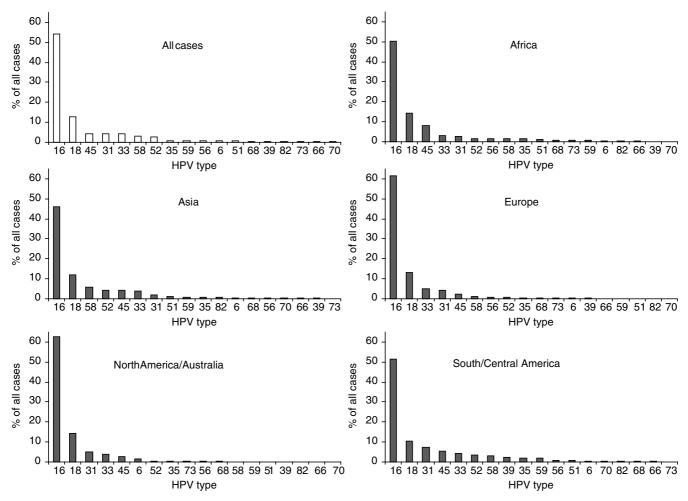


Figure 2 Type-specific prevalence of HPV in 8550 cases of squamous cell and unspecified cervical carcinoma by region.

and histology, we attempted to take account of alternative HPV DNA sources and PCR primers by adjustment. However, it is not known to what extent other unknown sources of variation such as sample storage conditions, specific PCR conditions and quality

of histopathology may affect these comparisons. Residual differences in prevalence between regions could also be because of the yet unknown HPV types not amplified by the existing PCR primers.



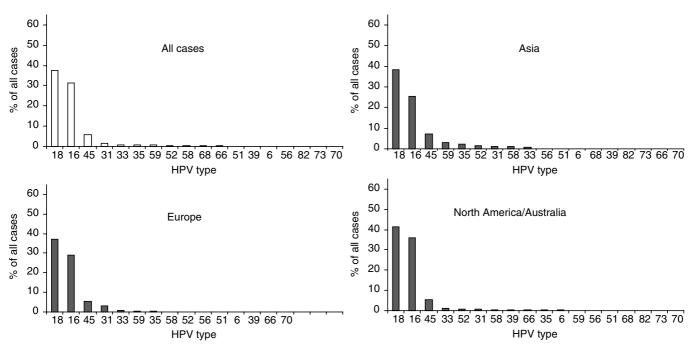


Figure 3 Type-specific prevalence of HPV in 1508 cases of adeno- and adenosquamous cervical carcinoma by region.

There were many similarities in HPV type-specific distribution across the regions studied. In SCC, HPV16 was clearly the predominant type varying from 45.9% in Asia to 62.6% in Europe, with HPV18 being found consistently in 10-14% of the cases. Other rarer types appeared to vary in their distribution. In most regions, HPV45, 31 and 33 were the third, fourth and fifth most common genotypes, although not necessarily in that order. Asia appeared to be different with a larger proportion of cancers associated with HPV58 and 52, as highlighted by a recent study in China of 786 cases in which HPV58 and 52 were the third (10%) and fourth (9%) most common genotypes in ICC (Wong et al, 2000). Other types in SCC were too rare to make inferences on region-specific variations.

Histological type of ICC

This meta-analysis shows that overall HPV prevalence detected in ADC was significantly lower than that detected in SCC. This intriguing finding does not appear to be because of differences with respect to region or HPV detection methods as it persisted even after adjusting for these factors. ADC arises from tissue deeper in the interior of the cervix uteri than SCC, and it has been reported to be more difficult to appropriately sample exfoliated cells of ADC than SCC (Sasieni and Adams, 2001). However, most HPV detected in the present review was based on biopsy specimens (77%). A proportion of cervical ADC could be misclassified ADC arising from the endometrium or other rare histological variants of ADC, for example, clear cell and mesonephric, which have been suggested to be HPV-independent (O'Leary et al, 1998; Pirog et al, 2000).

Whereas HPV16 was the most common type in SCC followed by HPV18, the situation was reversed in ADC where HPV18 was the most common type, followed closely by HPV16. This difference has been described independently by many of the studies in this analysis and by studies outside the scope of this review (IARC, 1995). Compared to HPV16, HPV18 has been shown to be associated with increasing oncogenic potential in cell culture (Barbosa and Schlegel, 1989), as well as a more rapid transition to malignancy (Burger et al, 1996) and a poorer prognosis of cancer patients (Nakagawa et al, 1996; Hildesheim et al, 1999; Schwartz et al, 2001). Given the fact that columnar tissue giving rise to ADC is less accessible, and possibly less susceptible to HPV infections, than the squamous tissue of SCC, the establishment of ADC may require a relatively more aggressive infection. In addition to HPV16 and 18, this large meta-analysis facilitated the identification of differences for some rarer phylogenetically related types: the HPV16-related types 31, 33, 35, 52, and 58 were more prevalent in SCC (15.0% collectively) than in ADC (4.4% collectively); and HPV18-related 45 was more prevalent in ADC (5.8%) than SCC (4.2%). All these differences were seen consistently in all regions where the comparison was possible.

For all regions where histological comparison was possible, the ratio of ADC to SCC was higher than that reported by cancer registries (Parkin et al, 1997). For example, ADC represent 23.1% of histologically verified cases from Europe in this study, but only 15.3% of ICC cases reported to European cancer registries (Parkin et al, 1997). Since all seven studies of only ADC cases were from Europe, Japan or USA, ADC is over-represented in this metaanalysis, particularly in developed countries. No material differences in results were observed when SCC was compared with cancers of unspecified histology.

Study limitations

The different PCR primers employed by the studies covered in this analysis varied in their overall detection of HPV DNA, with the highest prevalence being obtained with SPF10 and GP5+/6+ primers, supporting findings from previous studies (Davies et al, 2001). Such variation is partly because of known differences in the range of HPV DNA types amplifiable by each primer set, and this was taken into consideration when estimating type-specific prevalence. However, there is also evidence that not all primer sets amplify individual HPV types with the same sensitivity (Qu et al, 1997; Kleter et al, 1999), and such differences are a potential source of bias in this analysis.

The type-specific prevalence reported for each individual type includes that in multiple infections, which were reported in a total of 3.7% of our ICC cases. Since many of the included studies tested for only a subset of HPV types, many multiple infections will have been missed. Hence, this meta-analysis was unable to estimate how

often individual HPV types were found in the presence of other types, which limits the conclusions that can be made about individual HPV-type oncogenicity. In particular, a large proportion of cases positive for HPV6, which is not thought to be oncogenic, may be coinfected with an undetected HR HPV type.

The cases included in this meta-analysis are not representative of the worldwide distribution of ICC. The proportion of cases contributed by Africa (6.1%) and Asia (30.7%) in this study underrepresent their proportional burdens of worldwide cervical cancer, which are 14.1 and 49.4%, respectively (Parkin *et al*, 1999). In contrast, the proportion of cases contributed by Europe (33.2%) and North America and Australia (15.5%) over-represent their proportional burdens, which are 15.7 and 4.4%, respectively. Adjustment of type-specific prevalence in all ICC cases (Figure 1) by weighting each region according to their cancer burden, however, did not materially effect the results, highlighting the general similarity of HPV-type distribution across regions (data not shown).

Furthermore, the cases in this meta-analysis were not drawn uniformly from across each region. Large areas have not been included (e.g., the Middle East and Indian subcontinent in Asia), while other specific populations such as Japan in Asia are overrepresented. There is also evidence of inter-regional variation in HPV-type distribution; the high prevalence of HPV52 and 58 in Asia is more apparent in cases from China/Korea/Japan than in those from South East Asia. Hence, for the comparison of alternative regional groupings, HPV-specific prevalence is presented by study in the appendix (Table A1).

Given that HPV is considered a virtually necessary cause of ICC, we further examined results restricted to HPV DNA-positive cases. This increased type-specific prevalence by a factor of ≈ 1.1 consistently for each HPV type, with no impact on the relative distribution of HPV types. However, given that many of the PCR systems used by the included studies amplify only a subset of HPV types, many HPV-'negative' cases may actually be infected with

other, unascertained, HPV types. Thus, we did not consider it appropriate to restrict to HPV-positive cases when comparing type-specific prevalence across studies where PCR methodology differed considerably.

However, in order to make a broad overall estimate if one does assume that, HPV DNA should be detectable in 100% of ICC and that the distribution of undetected types in HPV-negative cases is similar to that in positive cases, this meta-analysis suggests that vaccinating against HPV16 and 18 should prevent over 70% of worldwide ICC. However, a worldwide vaccine against only HPV16/18 may prevent a larger proportion of ICC in Europe, North America and Australia (\approx 75%), than in Africa, Central and South America and Asia (59–64%), where a larger proportion of ICC cases were associated with other HPV types. Although this study identifies at least 18 HPV types associated with ICC from around the world, the most important type after HPV16 and 18 appears to be HPV45, followed by types 31, 33, 58 and 52, the relative importances of which vary by region.

ACKNOWLEDGEMENTS

The work reported in this paper was undertaken by Dr Gary Clifford during the tenure of an IARC Postdoctoral Fellowship from the International Agency for Research on Cancer. We thank the Vaccine Development team (VAD) of the Department of Vaccines and Biologicals of the World Health Organization, Geneva and Swiss Bridge (Award 2001) who supported and contributed to the funding of this work. We are grateful to Dr Teresa Aguado, Coordinator, VAD and Dr Sonia Pagliusi, Scientist, VAD for their critical comments during the preparation of the manuscript, as well as to Dr Rolando Herrero. We also thank those authors who made additional data available from their published studies.

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APPENDIX A

A Comparison of alternative regional groupings, HPV prevalence is presented by the study in Table A1

Table AI Study methods and prevalence of human papillomavirus by study and by region

		Country		PCR primers used to	No. cases	SCC (incl. unspec)/ s ADC	HPV prevalence (% of all cases tested)																	
First author	Reference		HPV DNA source	identify all HPV +ve			Any	16	18	45	31	33	58	52	35	59	56	51	6	68	39	82	73	66 7
Africa																								
Bosch FX	JNCI (1995)	Algeria, Benin, Guinea, Mali, Uganda,Tanzania	Fresh biopsies	My09/11	186	186/0	89.8	42.5	17.7	12.4	2.7	2.7	2.7	2.2	2.2	0.0	3.2	1.1	0.0	2.2		0.5		0.0
Bayo S	Int J Epidemiol (2002)	Mali	Fresh biopsies	GP5+/6+	65	65/0	96.9	47.7	12.3	10.8	0.0	1.5	3.1	0.0	1.5	0.0	0.0	6.2	0.0	0.0	0.0		3.1	0.0
Chaouki N	Int J Cancer (1998)	Morocco	Exfol. cells	GP5+/6+	186	173/13	94.6	67.7	12.4	4.8	3.8	2.7	0.0	1.1	0.5	1.6	0.5	0.0	0.5	0.0	0.0	0.0	0.0	0.5 0
Lin P	Cancer Epid Biomark Prev (2001)	Senegal	Exfol. cells	MY09/11 +HMB01	51	51/0	64.7	37.3	7.8	9.8	0.0	5.9	2.0	3.9	2.0	0.0	2.0	0.0	2.0	0.0	0.0		0.0	0.0
Williamson AL	I Med Virol (1994)	South Africa	Fresh biopsies	MY09/11	68	60/8	80.9	45.6	1.5	1.5	5.9	5.9							0.0					
ter Meulen J	Int Cancer (1992)	Tanzania	Exfol. cells	GP 5/6	53	53/0	88.7	37.7	32.1	5.7	0.0	1.9												
Region subtotal	• ()				609	588/21	88.8	50.2	14.1	7.9	2.6	3.1	1.6	1.6	1.4	0.6	1.6	1.2	0.4	8.0	0.0	0.3	0.7	0.2 0
Asia																								
Huang S	Int J Cancer (1997)	China	Fresh biopsies	MY09/11	40	35/5			30.0	0.0	0.0		27.5		0.0		0.0	0.0	0.0	0.0	0.0	0.0		0.0 0
Lin QQ	Int J Cancer (1998)	China	Fresh/fixed biopsies	MY09/11, GP5+/6+	77	77/0	93.5	48.1	5.2	1.3	2.6	3.9	18.2	5.2	0.0	2.6	0.0	0.0	10.4	0.0	0.0	0.0	0.0	0.0 1
Lo KWK	Gynecol Obstet Invest (2001)	China	Fresh biopsies	MY09/11	121	107/14	78.5	48.8	11.6	0.0	0.8	5.0	6.6	0.8	0.0	0.0	0.8	0.0	0.0	0.0	0.0		0.0	0.0 0
Peng H	Int J Cancer (1991)	China	Exfol. cells	TS-PCR only	101	92/9	34.7	31.7				3.0												
Stephen AL	Int J Cancer (2000)	China	Fixed biopsies	GP5+/6+ of TS-PCR neg samples only	34	24/10	88.2	61.8	8.8	2.9	0.0	2.9	2.9	0.0	0.0	2.9	0.0	0.0	0.0	0.0	0.0		0.0	0.0
Munirajan AK	Gynecol Oncol (1998)	India	Fresh biopsies	pU-1M/pU-2R	43	43/0	69.8	53.5	9.3		0.0	2.3	2.3	0.0	2.3	0.0	0.0		0.0					
Bosch FX	JNCI (1995)	Indonesia, Philippines, Thailand	Fresh biopsies	My09/11	98	98/0	96.9		31.6	8.2	1.0	2.0	2.0	2.0	1.0		3.1	0.0	0.0	1.0		4.1		0.0
Fujinaga Y	Gen Virol (1991)	Japan	Fresh biopsies	pU-IM/pU-2R	39	39/0	84.6	48.7	12.8		5.1	5.1	7.7	2.6					0.0					
Harima Y	Int J Radiat Oncol Biol Phys (2002)	Japan	Fresh biopsies	pU-1M/pU-2R	84	79/5	76.2	26.2	4.8		2.4	2.4	7.1	2.4	0.0	0.0	0.0		1.2					
Kashiwabara K	Acta Pathol Japan (1992)	Japan	Fixed biopsies	LICI/C2	93	68/25	58.1	48.4	6.5		0.0	1.1	0.0	3.2					0.0					
Maki H	Jpn J Cancer Res (1991)	Japan	Biopsies	LI PCR	29	29/0	82.8		20.7			6.9	6.9	6.9					0.0					
Nagai Y	Am J Clin Oncol (2001)	Japan	Exfol. cells	LICI/C2	293	239/54	85.3		3.1		3.8	4.1	3.1		1.7									
Nakagawa S	Cancer (1996)	Japan	Fresh biopsies	LICI/C2 +C2M	146	116/30	88.4		18.5	0.7	2.1	6.2	8.2	10.3	2.1	0.0	0.0	0.7	0.0	2.1	0.0			0.0 0
Nawa A	Cancer (1995)	Japan	Fresh/fixed biopsies	E6C1/C2	23	23/0	87.0		13.0			0.0												
Saito J	Gynecol Obstet Invest (2000)	Japan	Fixed biopsies	pU-1M/pU-2R	66	66/0	75.8	34.8	12.1			10.6	1.5	6.1										
Sasagawa T	Cancer Epid Biomark Prev (2001)	Japan	Exfol. cells	LCR-E7	84	72/12	89.3	42.9	14.3	1.2	6.0	1.2	3.6	10.7	1.2	0.0	0.0	4.8	1.2	0.0	0.0		0.0	0.0 0
Yamakawa Y	Gynecol Oncol (1994)	Japan	Fixed biopsies	TS-PCR only	64	0/64	67.2	32.8	39.1		1.6	0.0			7.8				0.0					
Hwang T	J Korean Med Sci (1999)	Korea	Exfol. cells	pU-1M/pU-2R	41	39/2	92.7	36.6	9.8		7.3	9.8	17.1	7.3	2.4									
Kim KH	Yonsei Med J (1995)	Korea	Fixed biopsies	WD72/76 + WD66/67/154	30	30/0	70.0	53.3	16.7		0.0	0.0												
Yadav M	Med Malaysia (1995)	Malaysia	Fresh biopsies	MY09/11	23	23/0	95.7	73.9	65.2		13.0	4.3												
Ngelangel C	JNCI (1998)	Philippines	Fresh biopsies +exfol. cells	GP5+/6+	356	323/33	93.5		25.6	12.9	0.6	0.0	2.5	2.8	0.0	2.0	0.6	2.5	0.0	0.6	0.3	0.3	0.3	1.1 0
Chen SL	Cancer (1993)	Taiwan	Fresh biopsies	L1C1/C2, E6C1/C2/C3	43	40/3	72.1	46.5	4.7		0.0	2.3	2.3	2.3					0.0					

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		Country	HPV DNA source	PCR primers used to		SCC (incl.	HPV į	V prevalence (% of all cases tested)																	
First author	Reference			identify all HPV +ve	No. cases	unspec)/ ADC	Any	16	18	45	31	33	58	52	35	59	56	51	6	68	39	82	73	66	70
Chen TM	Int J Cancer (1994)	Taiwan	Fresh biopsies	MY09/11, pU-1M/pU-2R	433	382/51	79.0	46.2	12.2			6.5							0.0						
Lai HC Yang YC	Int J Cancer (1999) Gynecol Oncol (1997)	Taiwan Taiwan	Fresh biopsies Fixed biopsies	MY09/II MY09/II of TS-PCR neg samples only	94 136	87/7 120/16	86.2 83.1	47.9 63.2		0.0	2.1 0.7	4.3 5.1	18.1	2.1	0.0	0.0	0.0	0.0	0.0	1.1	0.0	0.0	0.0	0.0	0.0
Bhattarakosol P Chichareon S	J Med Assoc Thai (1996) JNCI (1998)	Thailand Thailand	Fixed biopsies Fresh biopsies +exfol. cells	MY09/11 GP5+/6+	100 377	100/0 338/39	82.0 94.7		17.0 20.7	1.6	1.9	3.0 1.3	2.7	2.4	0.5	1.6	0.0	0.0	0.0 0.5	0.0	0.5	0.0	0.0	0.0	0.5
Siritantikom S	Southeast Asian J Trop Med Public Health (1997)	Thailand	Lavage	My09/11	23	21/2	60.9	56.5	4.3		4.3	4.3			0.0				0.0						
Region subtotal	,				3091	2710/381	83.1	43.4	15.3	4.5	2.0	3.4	5.4	4.2	1.0	1.1	0.4	1.0	0.5	0.5	0.2	0.4	0.1	0.3	0.3
Europe	M		E 11:	CD F . W .	0.4	0710	00.4		0.1	7.0	0.1	41.0	2.2	0.0		0.0	0.0	0.0	0.0	0.0	0.0	0.0		0.0	0.0
Birner P Baay MFD	Mod Pathol (2001) J Clin Microbiol (2001)	Austria Belgium	Fixed biopsies Fixed biopsies	GP 5+/6+ GP5+/6+ of TS-PCR neg	86 115	86/0 95/20	88.4 87.8			7.0 1.7	8.1 2.6	1.7	2.3 0.0	0.0	0.0		0.0	0.0	0.0		0.0	0.0	0.0		0.0
Tachezy R	Med Virol (1999)	Czech Republic	Fresh biopsies	samples only MY09/11	49	49/0	73.5	59.2	10.2	2.0	4.1	0.0	2.0	0.0	0.0	0.0	0.0	0.0		0.0	4.1	0.0	0.0	0.0	0.0
Hording U	APMIS (1997)	Denmark	Fixed biopsies	TS-PCR only	50	0/50	70.0		52.0			0.0													
Sebbelov AM	Microbes Infect (2000)	Denmark	Fixed biopsies	GP5/6 of TS-PCR neg samples only	34	34/0	85.3	70.6	0.0	0.0	0.0	5.9			0.0										
lwasawa A	Cancer (1996)	Finland	Fixed biopsies	MY09/11	460	352/108	88.0		25.0			2.6												0.0	
Lombard I	J Clin Oncol (1998)	France	Fresh biopsies	TS-PCR of SBH neg samples only	297	269/28	82.8	50.5	10.1	0.3	1.0	2.0	1.0	0.3	0.3				0.0		0.0				
Riou G	Lancet (1990)	France	Biopsies	TS-PCR of SBH neg samples only	106	89/17	84.0	54.7	16.0			5.7							0.0						
Milde-Langosch K	Int J Cancer (1995)	Germany	Biopsies	MY09/11	51	25/26	80.4	51.0	25.5		3.9	0.0			0.0				0.0						
Nindl I	International Papillomavirus Conference Proceedings (2001)	Germany	Fixed biopsies	My09/11	77	77/0	89.6	42.0	16.0		18.0	6.0	0.0	1.0	1.0			0.0	8.0		1.0			1.0	
Bosch FX	JNCI (1995)	Germany, Poland, Spain	Fresh biopsies	MY09/11	86	86/0	95.3	65.I	8.1	2.3	5.8	1.2	1.2	3.5	1.2	0.0	2.3	0.0	0.0	3.5	0.0	0.0		1.0	
Dokianakis DN	Oncol Rep (1999)	Greece	Pap smears	GP 5/6	77	75/2	58.4		36.4			3.9													
Koffa M	Int J Oncol (1994)	Greece	Fixed biopsies	GP 5/6	39	32/7	76.9	35.9		0.0		10.3	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0		0.0	
Labropoulou V Sebbelov AM	Sex Transmis Dis (1997) Microbes Infect (2000)	Greece Greenland	Fresh biopsies Fixed biopsies	MY09/11 GP5/6 of TS-PCR neg	35 32	35/0 32/0	97.1 84.4	54.3 81.3			5.7 3.1	0.0 3.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0		0.0	
Konya J	J Med Virol (1995)	Hungary	Fresh biopsies	samples only L1C1/C2 +C2M	47	41/6	97.9	55.3	40.4		2.1	0.0	0.0	2.1					0.0						
O'Leary JJ	J Clin Pathol (1998)	Ireland	Fixed biopsies	GP5/6, GP1/2	20	20/0	90.0		10.0			0.0							0.0						
Sjyldberg BM Garzetti GG	Mod Pathol (1999) Cancer (1998)	Ireland, Sweden Italy	Fixed biopsies	GP 5+/6+ GP 5/6	38 32	0/38 32/0	60.5 68.8		26.3 15.6		0.0 6.3	0.0			0.0				0.0						
Voglino G	Pathologica (2000)	Italy Italy	Fresh biopsies Fixed biopsies	MY09/11,	145	120/25	98.6				16.6	2.1			0.6				0.0						
3	J ()	,		pU1M/2R																					
Karlsen F	J Clin Microbiol (1996)	Norway	Fresh biopsies	My09/11, GP5+6+, Oli1b/2l, Cpl/ll	361	361/0	98.3	68.4	14.1		1.1	8.3			0.6										

 Table AI (continued)

				PCR primers used to identify all HPV +ve	No.	SCC (incl. . unspec)/ es ADC	HPV prevalence (% of all cases tested)																		
First author	Reference	Country	HPV DNA source				Any	16	18	45	31	33	58	52	35	59	56	51	6	68	39	82	73	66	70
Kleter B	J Clin Microbiol (1999)	Russia	Fixed biopsies	SPF10	180	129/51	100.0	64.4	9.4	7.8	3.9	1.1	1.7	1.1	1.7	0.0	2.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Munoz N	Int J Cancer (1992)	Spain	Exfol. cells	MY09/11	142	142/0	69.0				3.5	3.5			1.4				0.0		0.0			0.0	
Rodriguez JA	Diag Mol Pathol (1998)	Spain	Fixed biopsies	GP 5+/6+	54	54/0	85.2					1.9	1.9	0.0	0.0		0.0		0.0	0.0	0.0	0.0	0.0	0.0	0.0
Andersson S	Eur J Cancer (2001)	Sweden	Fixed biopsies	My09/11	173	0/173	68.2	23.9		5.2		0.0	0.0	0.0	0.0	0.6	0.0	0.0	0.0		0.0				
Hagmar B	Med Oncol Tumor Pharmacother (1992)	Sweden	Fixed biopsies	MY09/11	71	71/0	74.6				16.9	8.5							0.0						
Wallin KL	NEJM (1999)	Sweden	Fixed biopsies	MY09/11, GP 5+/6+	104	85/19	76.9	47.1				5.8	0.0	0.0	0.0	0.0	0.0			0.0	0.0	0.0	1.0	0.0	0.0
Zehbe I	J Pathol (1997)	Sweden	Fixed biopsies	GP 5+/6+	45	38/7	95.6		20.0		0.0		2.2	0.0	0.0		0.0		0.0						
Baay MFD	Eur J Gynaec Oncol (1996)	The Netherlands	Fixed biopsies	MY09/11, GP5/6, Cpl/II	162	162/0	87.7		14.2	0.6		4.3	0.6	0.0	0.0	0.0	0.0	0.0	0.0		0.0	0.0		0.0	0.0
van den Brule AJC	Int J Cancer (1991)	The Netherlands	Fresh biopsies	GP5/6 + GP1/2	50	50/0	100.0		26.0		4.0	2.0							0.0						
Arends MJ	Hum Pathol (1993)	UK	Fixed biopsies	TS-PCR only	47	26/21			29.8			0.0							0.0						
Crook T	Lancet (1992)	UK	Fresh biopsies	TS-PCR of SBH neg samples only	28	23/5	89.3	/1.4	17.9		0.0	0.0							0.0						
Giannudis A	Int Cancer (1999)	UK	Fixed biopsies	GP 5+/6+	43	43/0	100.0	81.4	9.3	2.3	0.0	4.7	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.3			0.0	
Region subtotal	me y cuncer (1777)	OK .	Tixed biopsies	GI 3 170 1	3336	2733/603			17.5					0.5								0.0	0.3	0.1	0.0
North America and Australia																									
Chen S	Int Gynecol Obstet (1999)	Australia	Fresh biopsies	My09/11	186	153/33	91.9	53.8	17.2	4.8															
Thompson CH	Gynecol Oncol (1994)	Australia	Fresh/fixed biopsies	pÚ-1M/pU-2R	103	103/0	86.4		18.4		2.9	0.0	0.0												
Duggan MA	Human Pathol (1995)	Canada	Fixed biopsies	LICI/C2 of DBH neg	76	0/76	69.7	35.6	39.5		0.0	2.6			0.0				0.0						0.0
D 1 D/	(N.C. (100E)	1164 6 1	E 111	samples only		F7/0	02.0	F70	150	140		0.0	0.0	0.0	0.0	0.0	2.5	0.0	0.0			0.0		0.0	
Bosch FX	JNCI (1995)	USA, Canada	Fresh biopsies	My09/11	57	57/0	93.0					0.0	0.0	0.0	0.0		3.5		0.0		0.0	0.0	0.2	0.0	0.0
Burger RA Burnett AF	JNCI (1996) Gynecol Oncol (1992)	USA USA	Fresh biopsies Fixed biopsies	My09/11 My09/11	40 I 2 I	297/104 18/3	84.8 100.0	51.4	28.6	2.2	2.5 42.9	1.0 4.8	0.5	1.0	0.0	0.0	0.0	0.0	0.5		0.0	0.0	0.2	0.0	0.0
Ferguson AW	Mod Pathol (1998)	USA	Fixed biopsies	MY09/11	27	0/27	59.3		25.9	7.4	72.7	T.0			17.0				0.0						
Paquette RL	Cancer (1993)	USA	Fresh biopsies	MY09/11	45	28/17	93.3	48.9		7.1	4.4								0.0						
Schwartz SM	Clin Oncol (2001)	USA	Fixed biopsies	MY09/11	465	354/111	88.2		23.2	1.3	2.4	3.4	0.0	0.0	0.2	0.0	0.0	0.0			0.2	0.0	0.2	0.2	0.0
Sebbelov AM	Microbes Infect (2000)	USA	Fixed biopsies	GP5/6 of TS-PCR neg	53	53/0	98.1		3.8				0.0	0.0	0.0	0.0	0.0	0.0		0.0	0.2	0.0	0.2	0.2	0.0
				samples only																					
Wistuba II	Cancer Res (1997)	USA	Fixed biopsies	SPF10	20	20/0	90.0		20.0		5.0	5.0													
Resnick RM	JNCI (1990)	USA (+Holland)	Fixed biopsies	My09/11, WD72/76	33	29/4			15.2			3.0							0.0						
Pirog EC Region subtotal	Am J Pathol (2000)	USA (+Poland)	Fixed biopsies	SPF10	76 1562	0/75 2/450	89.5 87.5		38.2 22.1		1.3 3.9	1.3 3.2	0.0 0.2	2.7 0.6	0.0 0.4	0.0 0.0	0.0 0.2	0.0 0.0	0.0 I.I		0.0 0.1	0.0	0.2	0.0 0.1	0.0
South and Central Amen																									
Bosch FX	JNCI (1995)	Argentina,Bolivia, Brazil,Chile, Colombia,Cuba, Panama,Paraguay	Fresh biopsies	MY09/11	505	505/0	92.9	50.5	9.5	7.3	6.9	3.6	2.2	3.2	2.0	2.8	0.6	1.0	0.2	0.4		0.2			
Alonio LV	MEDICINA (2000)	Argentina	Biopsies	GP 5+/6+	30	30/0	93.3	467	20.0		3.3	3.3							6.7						
Eluf-Neto J	Br Cancer (1994)	Brazil	Exfol. cells	GP 5/6	186	186/0	84.4				2.2	3.2							0.0						
Lorenzato F	Int J Gynecol Cancer (2000)	Brazil	Exfol. cells	MY09/11	59	59/0			38.5	0.0	11.9		3.4	1.7	3.4				0.0						

SCC

(incl.

unspec)/

No.

HPV prevalence (% of all cases tested)

PCR primers

used to

identify

HPV DNA