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The complete chloroplast genome of *Dryas octopetala* var. *asiatica* (Dryadoideae, Rosaceae) and phylogenetic analysis

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ABSTRACT

Dryas octopetala L. var. *asiatica* (Nakai) Nakai 1918 is a dwarf shrub that mainly grow in alpine and arctic zones of the Northern Hemisphere, representing an endemic variety in Asia. In the present study, the complete chloroplast (cp) genome of *D. octopetala* var. *asiatica* was first characterized and used for its phylogenetic analysis. The cp genome span 158,271 bp with an overall GC content of 36.5%. A total of 129 genes were identified, including 84 protein-coding genes (PCGs), 37 tRNA genes, and 8 rRNA genes. In addition, repetitive sequences and microsatellites were detected within this species. Phylogenetic analysis involving 39 cp genomes from Rosaceae family indicated that *D. octopetala* var. *asiatica* was sister to the clade of Amygdaloideae. This study contributes fundamental insights into the cp genome of *Dryas octopetala* var. *asiatica*, which will have expanded its use in photosynthesis and evolutionary study.

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Introduction

Dryas octopetala L. var. asiatica (Nakai) Nakai 1918 is a dwarf shrub belonging to Dryadoideae, Rosaceae. Typically found in alpine and arctic zones of the Northern Hemisphere, this plant faces various environmental challenges such as low temperatures and limited nutrient availability that impact its growth and reproduction. Known for forming ectomycorrhiza (ECM) and engaging in nitrogen-fixing root nodule symbiosis (Gardes and Dahlberg 1996; Bjorbækmo et al. 2010; Li et al. 2016; Billault-Penneteau et al. 2019), *Dryas* species play a significant role in mycology community ecology. Moreover, *D. octopetala* is of particular importance in research on climate



Figure 1. Reference image of *Dryas octopetala* var. *asiatica* taken by Limin Yang at Changbai Mountain of Jilin province, China. Note: *Dryas octopetala* var. *asiatica* is characterized by its prostrate growth habit, leathery leaves, showy white flowers, and its ability to thrive in harsh alpine conditions.

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change (McGraw et al. 2014; Panchen and Gorelick 2017). *Dryas octopetala* var. *asiatica* as the Asian endemic variety is sparsely distributed in alpine regions of northeastern Asia. A recent study has highlighted the sensitivity of photosynthesis to climate warming and its adaptability to temperature increase in *D. octopetala* var. *asiatica* (Zhou et al. 2019). Chloroplast (cp) is the site of photosynthesis, in which light

 Table 1. Information of the chloroplast genome in Dryas octopetala var. asiatica.

| Regions | Length(bp) | GC(%) | Number | | |
|---------|------------|-------|----------------------|-------|-------|
| | | | protein-coding genes | tRNAs | rRNAs |
| Genome | 158,271 | 36.5 | 84 | 37 | 8 |
| LSC | 86,980 | 34.1 | 62 | 22 | 0 |
| SSC | 18,461 | 30.2 | 12 | 1 | 0 |
| IRA | 26,415 | 42.5 | 5 | 7 | 4 |
| IRB | 26,415 | 42.5 | 5 | 7 | 4 |

energy is converted to chemical energy and the oxygen and energy-rich organic compounds are produced. However, little is known about the whole cp genome for this variety. In this study, the cp genome of *D. octopetala* var. *asiatica* was first sequenced using next-generation sequencing. We mainly focused on the analyses of cp genome features, repeat sequences and phylogenetic reconstruction. This study not only offers the valuable cp genome data but also contributes to understanding the phylogenetic relationships within Rosaceae.

Materials and methods

Dryas octopetala var. *asiatica* was collected from Changbai Mountain in Jilin province, China (Figure 1; GPS coordinates: 42°03'17"N 128°04'00"E). A voucher specimen (voucher



Figure 2. Chloroplast genome map of Dryas octopetala var. asiatica.

Note: Genes are depicted as differently sized and colored boxes on the outermost circle, with inner and outer boxes representing genes transcribed in clockwise and counter-clockwise directions. The middle circle illustrates changes in GC content at different positions, while the inner circle highlights the regions and lengths indicated by the tetrad structure (LSC, SSC, IRa, and IRb) in different colors.



Figure 3. The phylogenetic tree based on the complete chloroplast genomes of 40 species of Rosaceae. Numbers at nodes correspond to maximum likelihood bootstrap percentage and the posterior probability of Bayesian inference. The sequences employed for the construction of the phylogenetic tree are detailed as follows: *Adenostoma fasciculatum* (KY387915, Zhang et al. 2017), *Agrimonia coreana* (MF135594, direct submission), *Alchemilla argyrophylla* (MT382661, direct submission), *Argentina phanerophlebia* (MT114192, Aogan 2020a), *Aruncus aethusifolius* (MZ882398, direct submission), *Bencomia exstipulata* (M682353, direct submission), *Chamaebatiaria millefolium* (KY420017, Zhang et al. 2017), *Comarum salesovianum* (MT017928, Aogan 2020b), *Dasiphora fruticosa* (MF683841, Zhao et al. 2018), *Dryas octopetala* var. *asiatica* (OQ420424, this study), *Fragaria chiloensis* (MW537844, direct submission), *Geum elatum* (MT982432, direct submission), *Hagenia abyssinica* (KX008604, direct submission), *Holdoiscus argenteus* (KY420013, Zhang et al. 2017), *Kelseya uniflora* (KY419988, Zhang et al. 2017), *Kerria japonica* (MN418902, Huo et al. 2019), *Lindleya mespiloides* (MN068248, direct submission), *Luetkea pectinata* (KY419971, Zhang et al. 2017), *Kelseya uniflora* (KY419988, Zhang et al. 2017), *Kerria japonica* (MN418902, Huo et al. 2017), *Malus prattii* (MH929090, direct submission), *Morus alba* (MW465954, direct submission), *Neillia incisa* (MT683856, direct submission), *Pentactina rupicola* (JQ041763, Kim and Kim 2016), *Petrophytum caespitosum* (KY419970, Zhang et al. 2017), *Phipspiomeles mexicana* (MN062003, Liu et al. 2019), *Potaninia mongolica* (MN691039, Corg and Jiang 2020), *Potantilla anserina* (OW176989, direct submission), *Prinsepia uniflora* (MZ270554, direct submission), *Prunus cerasus* (MW477422, direct submission), *Sprusos* (HG737342, Korotkova et al. 2014), *Rhamnus globosa* (MT360052, Xie et al. 2020), *Rosa acicularis* (MK741016, Chen et al. 2019), *Rubus amabilis* (MN652918, di

number: ZhouHC1243) was deposited in the Herbarium of Kunming Institute of Botany, CAS (KUN, http://www.kun.ac. cn/, Tao Deng, dengtao@mail.kib.ac.cn). Genomic DNA was extracted from the silica gel-dried leave using a modified CTAB method (Yang et al. 2015) and the constructed libraries were sequenced using Illumina NovaSeq PE150 platform. High-quality reads were filtered and *de novo* assembled with SPAdes software (Prjibelski et al. 2020). The cp genome of *D. octopetala* var. *asiatica* was annotated using PGA software (Qu et al. 2019) with manual adjustments and visualized with the online CPGView program (Liu et al. 2023). The annotated cp genome of *D. octopetala* var. *asiatica* was deposited in GenBank under accession number OQ420424. Phylogenetic reconstruction was performed with

one representative species from 39 genera of Rosaceae downloaded from NCBI GenBank database with *D. octopetala* var. *asiatica. Morus alba* from Moraceae and *Rhamnus globose* from Rhamnaceae were used for the outgroups. Multiple sequence alignments of the complete cp genomes were carried out using MAFFT v.6.833 (Katoh et al. 2005) and used to construct phylogenetic trees with maximum likelihood (ML) and Bayesian inference (BI) methods as previously described (Zhang et al. 2017).

Results and discussion

The complete cp genome of *D. octopetala* var. *asiatica* was 158,271 bp long with an average sequencing depth of

 $2696 \times$ and the minimum sequencing depth of $11 \times$ (supplemental Figure S1). The overall GC content of this cp genome was 36.5% (Table 1). The cp genome exhibited a typical quadripartite structure, consisting of two copies of inverted repeats (IRs) and one large single- copy (LSC) and one small single-copy (SSC) region (Figure 2). The length of the four segments in this cp genome varied, with the LSC region being the largest at 86,980 bp, followed by the two IR regions at 26,415 bp each (Table 1). The GC contents of the four segments differed, with the IR regions having the highest GC content at 42.5% and the SSC region having the lowest at 30.2%. The IR regions were flanked by the LSC and SSC region, forming a circular structure (Figure 2). These results indicated the cp genome structure of D. octopetala var. asiatica is typical with most other land plants (Han et al. 2019; Huang et al. 2021; Zhang and Ling 2022).

The cp genome of *D. octopetala* var. *asiatica* was found to contain a total of 129 genes, including 84 protein-coding genes (PCGs), 37 tRNA, and 8 rRNA genes (Figure 2). The major functions of these genes were associated with photosynthesis and metabolic processes (Figure 2), with 20 genes specifically encoding proteins related to photosynthesis (Figure 2). The majority of PCGs were located in the LSC region, while 16 genes, including 7 tRNA genes, all 4 rRNA genes and 5 PCGs were duplicated in the IR region (Table 1 and Figure 2). Furthermore, 11 genes contained one intron and 2 genes had two introns (supplemental Figure S2), potentially undergoing cis or trans-splicing during the transcription process (supplemental Figure S2 and S3).

Repeats are often associated with genome rearrangement through gene inversion that reverses the order and orientation of multiple genes (Cavalier-Smith 2002). In this study, the dispersed repeats and long tandem repeats were identified and shown in the first two tracks (from the center going outward) (Figure 2). Of them, the majority were located in the LSC region, with a few repeats in the SSC region (Figure 2). Short tandem repeats or microsatellite sequences were shown in the third track and exhibited the similar distribution pattern. These repeat types were also found in other Rosaceae genera like Sorbus (Zhang and Ling 2022). Previous studies have indicated that repeat sequences have promoted the generation of substitutions and indels during chloroplast genome evolution (McDonald et al. 2011; Ahmed et al. 2012). Therefore, the repeats within D. octopetala var. asiatica cp genomes could potentially induce mutations, influencing mutation rates and the evolution of this species.

The phylogenetic position of *D. octopetala* var. *asiatica* as the representative species of Dryadoideae was investigated using 39 species from different genera of Rosaceae. Both BI and ML methods were employed, resulting in the identical topology (Figure 3). Rosaceae was resolved into three main clades representing the subfamilies: Amygdaloideae, Dryadoideae and Rosoideae. Amygdaloideae and Dryadoideae (the respective species, *D. octopetala* var. *asiatica*) had the closer relationship and formed two sister clades with strong supports (Figure 3), consistent with the previous studies (Potter et al. 2007; Zhang et al. 2017). In fact, Amygdaloideae and Dryadoideae have the similar chemical compounds, like

cyanogenic glycosides and sorbitol (Zhang et al. 2017). However, some analyses based on the partial sequences of cp genome (Zhang et al. 2017) suggested a sister relationship between Dryadoideae and Rosoideae. Given the presence of hybridization, apomixis and polyploidy present in Rosaceae (Vamosi and Dickinson 2006; Burgess et al. 2015), further research is needed to definitively resolve the phylogenetic relationships among these subfamilies.

Conclusions

In this study, we first sequenced and characterized the complete cp genome of *D. octopetala* var. *asiatica*. Our results revealed that the cp genome of *D. octopetala* var. *asiatica* follows a typical quadripartite and circular DNA structure. Various important features related to structure and sequence divergence such as genome size, gene/intron content, repeats and microsatellite, were analyzed in this species. Additionally, the phylogenetic analysis positioned *D. octopetala* var. *asiatica* as a representative species of Dryadoideae sister to the clade of Amygdaloideae. These basic knowledge of the cp genome of *D. octopetala* var. *asiatica* will enhance its utility in comparative studies and contribute valuable data for research on photosynthesis.

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Ethical approval

Dryas octopetala var. *asiatica* is not an endangered or protected species and collected without requirement of the specific permission from authorities. This plant used in the study is complied with national and international standards and local laws and regulations.

Author contributions

L.Z. L and S.D. Z designed the experiment and obtained the funding. L.Z. L and Y.G. Z performed laboratory work. L.Z. L, L.M. Y and S.D. Z performed bioinformatics analyses, wrote and revised the manuscript. All authors have read and agreed to the published version of the manuscript.

Disclosure statement

No potential conflict of interest was reported by the author(s).

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Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at (https://www.ncbi.nlm.nih.gov/)

under the accession no. OQ420424. The associated BioProject, SRA, and Bio-Sample numbers are PRJNA724898, SRR28753508, and SAMN41028577 respectively.

References

- Ahmed I, Biggs PJ, Matthews PJ, Collins LJ, Hendy MD, Lockhart PJ. 2012. Mutational dynamics of aroid chloroplast genomes. Genome Biol Evol. 4(12):1316–1323. doi:10.1093/gbe/evs110.
- Aogan K, Li QQ. 2020a. The complete chloroplast genome of *Argentina phanerophlebia* (Rosaceae: Potentilleae). Mitochondrial DNA B Resour. 5(2):1763–1764. doi:10.1080/23802359.2020.1748549.
- Aogan K, Li QQ. 2020b. The complete chloroplast genome of Farinopsis salesoviana (Rosaceae: Potentilleae). Mitochondrial DNA B Resour. 5(2): 1363–1364. doi:10.1080/23802359.2020.1735275.
- Billault-Penneteau B, Sandré A, Folgmann J, Parniske M, Pawlowski K. 2019. Dryas as a model for studying the root symbioses of the Rosaceae. Front Plant Sci. 10:661. doi:10.3389/fpls.2019.00661.
- Bjorbækmo MF, Carlsen T, Brysting A, Vrålstad T, Høiland K, Ugland KI, Geml J, Schumacher T, Kauserud H. 2010. High diversity of root associated fungi in both alpine and arctic *Dryas octopetala*. BMC Plant Biol. 10(1):244–255. doi:10.1186/1471-2229-10-244.
- Burgess MB, Cushman KR, Doucette ET, Frye CT, Campbell CS. 2015. Understanding diploid diversity: a first step in unraveling polyploid, apomictic complexity in *Amelanchier*. Am J Bot. 102(12):2041–2057. doi:10. 3732/ajb.1500330.
- Cavalier-Smith T. 2002. Chloroplast evolution: secondary symbiogenesis and multiple losses. Curr Biol. 12(2):R62–R64. doi:10.1016/S0960-9822(01) 00675-3.
- Chen X, Liu Y, Sun J, Wang L, Zhou S. 2019. The complete chloroplast genome sequence of *Rosa acicularis* in Rosaceae. Mitochondrial DNA B Resour. 4(1):1743–1744. doi:10.1080/23802359.2019.1610100.
- Cong L, Jiang H. 2020. The complete chloroplast genome of *Potaninia* mongolica (Rosaceae) from China. Mitochondrial DNA B Resour. 5(1): 574–575. doi:10.1080/23802359.2019.1710295.
- Gardes M, Dahlberg A. 1996. Mycorrhizal diversity in arctic and alpine tundra: an open question. New Phytol. 133(1):147–157. doi:10.1111/j. 1469-8137.1996.tb04350.x.
- Han T, Li M, Li J, Lv H, Ren B, Chen J, Li W. 2019. Comparison of chloroplast genomes of *Gynura* species: sequence variation, genome rearrangement and divergence studies. BMC Genomics. 20(1):791. doi: 10.1186/s12864-019-6196-x.
- Huang R, Xie X, Chen A, Li F, Tian E, Chao Z. 2021. The chloroplast genomes of four *Bupleurum* (Apiaceae) species endemic to Southwestern China, a diversity center of the genus, as well as their evolutionary implications and phylogenetic inferences. BMC Genomics. 22(1):714. doi:10.1186/s12864-021-08008-z.
- Huo Y, Yan M, Zhao X, Zhu Z, Yuan Z. 2019. The complete chloroplast genome sequence of *Kerria japonica* (L.) DC. 'pleniflora' (Rosaceae). Mitochondrial DNA B Resour. 4(2):3723–3724. doi:10.1080/23802359. 2019.1678433.
- Katoh K, Kuma K, Toh H, Miyata T. 2005. MAFFT version 5: improvement in accuracy of multiple sequence alignment. Nucleic Acids Res. 33(2): 511–518. doi:10.1093/nar/gki198.
- Kim HW, Kim KJ. 2016. The complete plastome sequence of *Pentactina rupicola* Nakai (Rosaceae), a genus endemic to Korea. Mitochondrial DNA B Resour. 1(1):698–700. doi:10.1080/23802359.2016.1225523.
- Korotkova N, Nauheimer L, Ter-Voskanyan H, Allgaier M, Borsch T. 2014. Variability among the most rapidly evolving plastid genomic regions is lineage-specific: implications of pairwise genome comparisons in *Pyrus* (Rosaceae) and other angiosperms for marker choice. PLoS One. 9(11):E112998. doi:10.1371/journal.pone.0112998.
- Li HL, Wang W, Li RQ, Zhang JB, Sun M, Naeem R, Su JX, Xiang XG, Mortimer PE, Li DZ, et al. 2016. Global versus Chinese perspectives on the phylogeny of the N-fixing clade. J Syst Evol. 54(4):392–399. doi:10. 1111/jse.12201.
- Liu BB, Hong DY, Zhou SL, Xu C, Dong WP, Johnson G, Wen J. 2019. Phylogenomic analyses of the *Photinia* complex support the recognition

of a new genus *Phippsiomeles* and the resurrection of a redefined *Stranvaesia* in Maleae (Rosaceae). J Syst Evol. 57(6):678–694. doi:10.1111/jse.12542.

- Liu S, Ni Y, Li J, Zhang X, Yang H, Chen H, Liu C. 2023. CPGView: a package for visualizing detailed chloroplast genome structures. Mol Ecol Resour. 23(3):694–704. doi:10.1111/1755-0998.13729.
- McDonald MJ, Wang WC, Huang HD, Leu JY. 2011. Clusters of nucleotide substitutions and insertion/deletion mutations are associated with repeat sequences. PLoS Biol. 9(6):e1000622. doi:10.1371/journal.pbio. 1000622.
- McGraw JB, Turner JB, Chandler JL, Vavrek MC. 2014. Disturbances as hot spots of ecotypic variation: a case study with *Dryas octopetala*. Arct Antarct Alp Res. 46(3):542–547. doi:10.1657/1938-4246-46.3.542.
- Meng XX, Xian YF, Xiang L, Zhang D, Shi YH, Wu ML, Dong GQ, Ip SP, Lin ZX, Wu L, et al. 2018. Complete chloroplast genomes from *Sanguisorba*: identity and variation among four species. Molecules. 23(9):2137. doi:10.3390/molecules23092137.
- Panchen ZA, Gorelick R. 2017. Prediction of Arctic plant phenological sensitivity to climate change from historical records. Ecol Evol. 7(5): 1325–1338. doi:10.1002/ece3.2702.
- Potter D, Eriksson T, Evans RC, Oh S, Smedmark JEE, Morgan DR, Kerr M, Robertson KR, Arsenault M, Dickinson TA, et al. 2007. Phylogeny and classification of Rosaceae. Plant Syst Evol. 266(1–2):5–43. doi:10.1007/ s00606-007-0539-9.
- Prjibelski A, Antipov D, Meleshko D, Lapidus A, Korobeynikov A. 2020. Using SPAdes De Novo Assembler. Curr Protoc Bioinformatics. 70(1): e102. doi:10.1002/cpbi.102.
- Qu XJ, Moore MJ, Li DZ, Yi TS. 2019. PGA: a software package for rapid, accurate, and flexible batch annotation of plastomes. Plant Methods. 15(1):50. doi:10.1186/s13007-019-0435-7.
- Sun W, Chen Z, Lv Z, Li Z. 2020. Sequencing and characterization of the complete chloroplast genome of *Sorbaria arborea* (Rosaceae). Mitochondrial DNA B Resour. 5(1):1090–1091. doi:10.1080/23802359.2020.1721359.
- Tian, Wei-Jie, Li, Qin-Qin, Khasbagan, 2020. The complete chloroplast genome of *Sibbaldianthe adpressa* (Rosaceae: Potentilleae). Mitochondrial DNA B Resour. 5(2):1563–1564. doi:10.1080/23802359.2020.1742601.
- Vamosi JC, Dickinson TA. 2006. Polyploidy and diversification: a phylogenetic investigation in Rosaceae. Int J Plant Sci. 167(2):349–358. doi: 10.1086/499251.
- Xie Y, Wang Z, Jiang X, Zhang X. 2020. The complete chloroplast genome of *Rhamnus globosa* (Rhamnaceae). Mitochondrial DNA B Resour. 5(3):2830–2831. doi:10.1080/23802359.2020.1791010.
- Yang JB, Li DZ, Li HT. 2015. Highly effective sequencing whole chloroplast genomes of angiosperms by nine novel universal primer pairs. Mol Ecol Resour. 14(5):1024–1031. doi:10.1111/1755-0998.12251.
- Zhang SD, Jin JJ, Chen SY, Chase MW, Soltis DE, Li HT, Yang JB, Li DZ, Yi TS. 2017. Diversification of Rosaceae since the Late Cretaceous based on plastid phylogenomics. New Phytol. 214(3):1355–1367. doi:10.1111/ nph.14461.
- Zhang SD, Ling LZ. 2022. Molecular structure and phylogenetic analyses of the plastomes of eight *Sorbus sensu stricto* species. Biomolecules. 12(11):1648. doi:10.3390/biom12111648.
- Zhang SD, Yan K, Ling LZ. 2023. Characterization and phylogenetic analyses of ten complete plastomes of *Spiraea* species. BMC Genomics. 24(1):137–148. doi:10.1186/s12864-023-09242-3.
- Zhang, Xiao-Hui, Li, Qin-Qin, Khasbagan. 2020. The complete chloroplast genome of *Sibbaldia aphanopetala* (Rosaceae: Potentilleae). Mitochondrial DNA B Resour. 5(3):2026–2027. doi:10.1080/23802359.2020.1756945.
- Zhao Y, Lu D, Han R, Wang L, Qin P. 2018. The complete chloroplast genome sequence of the shrubby cinquefoil *Dasiphora fruticosa* (Rosales: Rosaceae). Conservation Genet Resour. 10(4):675–678. doi:10.1007/ s12686-017-0899-6.
- Zhou Y, Deng J, Tai Z, Jiang L, Han J, Meng G, Li MH. 2019. Leaf anatomy, morphology and photosynthesis of three tundra shrubs after 7year experimental warming on Changbai Mountain. Plants (Basel). 8(8):271–284. doi:10.3390/plants8080271.