

Pluripotent stem cells: A therapeutic source for age-related macular degeneration

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Age-related macular degeneration (AMD) leads to progressive loss of central vision in the elderly. At a cellular level, there is aging of the retinal pigment epithelial (RPE) cells, and accumulation of lipofuscin that interferes with the proper functioning of RPE which eventually leads to apoptosis. Treatment depends on the stage of the disease. Wet AMD which has neovascularization is managed by local therapies such as laser photocoagulation and photodynamic therapy and is managed with injections of antivascular endothelial growth factor-based therapy. Unlike the wet AMD, an effective therapy does not exist for dry AMD and geographic atrophy. Cell replacement therapy has shown promise. This review discusses the opportunities in the various types of cell-based therapy, their limitations, and what is possible for India.

Key words: Age-related macular degeneration, embryonic stem cells, retinal pigment epithelial, stem cells

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Age-related Macular Degeneration

Age-related macular degeneration (AMD) leads to progressive loss of central vision. AMD was believed to be a leading cause of blindness in elderly population only in the Western countries. However, a recent study suggested that the prevalence of AMD is similar in India.^[1] AMD is classified into dry (nonexudative) and wet (exudative) AMDs. The dry AMD is more common (85%) than the wet AMD, with 10% of the dry forms progressing to wet AMD. AMD is believed to be caused by the primary failure of retinal pigment epithelial (RPE) cell functions.^[2] RPE plays an important role in the survival of photoreceptor. However, during the aging process, the RPE undergoes senescence and its dysfunction leads to accumulation of lipofuscin. *N*-retinylidene-*N*-retinylethanolamine (A2E) is a major component of lipofuscin that interferes with the proper functioning of RPE and its apoptosis. The loss of RPE subsequently leads to the development of geographic atrophy (GA).^[3] Amorphous deposits containing lipids and proteins called drusen accumulate between RPE and Bruch's membrane (BM). The presence of drusen has been attributed to increased risk of AMD development. In addition, the drusens lead to the detachment of RPE from BM, which is considered to induce choroidal neovascularization (CNV) observed in wet AMD.^[4]

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Strategies for Management of Age-related Macular Degeneration

Preventive strategies

Since AMD is a multifactorial disease, which has both genetic and environmental components, the patients are stratified based on the known risk factors. Those individuals who have risk alleles in genes such as complement factor H, ARMS2, and vascular endothelial growth factor A (VEGFA) have been predicted to develop AMD compared to those who do not carry the risk alleles. In addition, smoking and diet low in antioxidants have been attributed to the progression of AMD in individuals with known risk alleles. Hence, cessation of smoking and diet rich in antioxidants have been advocated as preventive measures in high-risk individuals.^[5,6] The results of the studies on the efficacy of antioxidant therapy have been controversial. A recent review suggested that there is no role for antioxidants in AMD progression.^[7] However, the age-related eye disease study suggested an increased reduction in risk of AMD progression in individuals consuming antioxidant and mineral supplements.

Treatment strategies for age-related macular degeneration

Several treatment strategies are available for treating wet AMD. The strategies that are widely employed as therapy for wet AMD include laser photocoagulation,^[8-11] photodynamic

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therapy,^[12,13] and anti-VEGF therapy.^[14-21] However, there is no effective therapy that exists with regard to dry AMD and GA. Several strategies are being attempted for treating dry AMD and GA such as drugs involved in reducing or blocking drusen formation,^[22-24] reducing and eliminating inflammation.^[25-27]

Cell Replacement Therapy as a Suitable Alternative

The initial surgical procedure for replacing diseased macula involved 360° retinal rotation to translocate the macula.^[28] This surgery was always complemented with strabismus surgery to counterrotate the globe. The surgery resulted in reasonable visual outcome. However, complication of proliferative vitreoretinopathy was noted in 8%–18% of the cases. The recurrence of the disease in RPE was noted in new foveal regions owing to compromised photoreceptor function in the region in most cases of GA.^[29] Recently, transplantation of RPE cells has emerged as a suitable alternative to conventional therapies.

Points to be Considered during Retinal Pigment Epithelial Transplantation

The success of RPE transplantation depends on the following conditions:

- i. A renewable source of RPE cells
- ii. Surgical method for transplanting RPE cells in subretinal region
- iii. Survival and functioning of RPE in the transplanted region
- iv. Restoration of retinal architecture
- v. Improvement of visual acuity.

Hence, the most important criteria for successful RPE transplantation are the source of RPE cells.

Source of Retinal Pigment Epithelial Cells for Transplantation

The RPE cells in the adult retina are postmitotic and hence cannot be utilized for transplantation purposes. The ideal characteristics of cells for transplantation are as follows: (i) Available in sufficient numbers, preferably renewable, (ii) autologous or less immunogenic, and (iii) ethical and less complex. Several types of stem cells have been studied for their ability to replace or rejuvenate the degenerating retinal cells which have one or more of the ideal characteristics mentioned above. These cells have been used in one of the two therapeutic strategies: (i) Source of neurotrophic support and (ii) source of functional cells during cell replacement therapies.

Source of Neurotrophic Support

Neural stem cells

Neural stem cells (NSCs) are multipotent cells isolated from both adult and fetal brain tissues that can give rise to neurons, glia, and oligodendrocytes. Transplantation of the NSCs isolated from aborted fetuses of 16–20 weeks of gestation into the subretinal space of postnatal day 21 Royal College of Surgeons (RCS) rats revealed that the transplanted cells could survive for at least 7 months in the retina without any evidence of tumor formation.^[30] In addition, the treatment revealed sustained visual acuity and luminance sensitivity

improvement. Further studies in this line confirmed that the transplanted cells rescue the degenerating cells predominantly by providing trophic support and do not transdifferentiate into retinal phenotypes. In addition, NSC-directed phagocytosis of photoreceptor outer segments has also been contemplated as the reason for the increased visual acuity.^[31] Recently, human central nervous system stem cell (HuCNS-SC), a Current Good Manufacturing Practices (cGMP) compliant adult NSC, has been developed by Stem Cells Inc. (Newark, CA, USA) along with HuCNS-SC which has been developed by the Retina Foundation of the Southwest (Dallas, TX, USA). Recently, a clinical trial with HuCNS-SC was conducted on patients with GA secondary to AMD and the results are awaited.

Adult mesenchymal stem cells

Mesenchymal stem cells (MSCs) are multipotent stem cells initially derived from bone marrow. Currently, several adult as well as fetal tissues have been shown to harbor these stem cells.^[32-45] Of the available sources, bone marrow-derived and human umbilical tissue-derived MSCs have been used in treating outer retinal degenerations in animal models. Studies involving subretinal as well as intravitreal transplantation of bone marrow-derived MSCs showed increased survival of RPE and photoreceptors with sustained retinal function in treated versus the sham-treated control in RCS rats and mouse model of retinitis pigmentosa (RP).^[46,47] A study by Wang *et al.* suggested that bone marrow-derived MSCs injected through intravenous route have the ability to home into the sites of retinal injury.^[48] A study on mouse model of laser-induced CNV suggested that MSCs that are genetically modified to secrete pigment epithelium-derived factor were shown to be a source of long-term antiangiogenic factor, thus providing an alternative for repeated intravitreal injections of anti-VEGF in wet AMD-associated CNV.^[49] Another source of MSC that has been proven to be effective in the RCS rats is from human umbilical tissue. Subretinal injection of the umbilical tissue-derived MSCs revealed better visual outcomes in the treated eye compared to the control.^[50] It is hypothesized that MSCs, like in other neural degenerations, have paracrine effect which rescue the degenerating cells. These findings subsequently led to a Phase I/II clinical trial by Janssen Research and Development (Philadelphia, PA, USA) utilizing cGMP compliant cell line of human umbilical tissue-derived stem cells (CNT0 2476). In this clinical trial, the cells were delivered to the subretinal space using a catheter-based delivery system. The results are yet to be published.

Bone marrow-derived hematopoietic stem cells or bone marrow stem cells

Li *et al.* showed that sodium iodate-induced RPE damage in C57Bl6 mice led to an active migration of bone marrow stem cell (BMSC) to the subretinal space and it was hypothesized that these BMSCs can orchestrate retinal repair.^[51] Atmac-Sonmez *et al.* showed that green fluorescent protein (GFP)-labeled allogeneic BMSC delivered intravenously into the mouse model of sodium iodate-induced RPE damage, migrated and survived in subretinal spaces with loss of RPE sparing the normal subretinal sites. It was suggested that these GFP-positive cells also expressed RPE65 an RPE-specific marker but did not show morphological or functional attributes of RPE.^[52] There are at least four clinical trials that are ongoing utilizing autologous BMSCs to treat AMD [Table 1]. The results of these studies are awaited.

Table 1: Stem-cell-based clinical trials for age-related macular degeneration registered under the Food and Drug Administration/Clinical Trials Registry India/Japanese Ministry of Health

Identifier	Funding source	Location	Source cell	Route of administration	Status	Published
NCT01632527	Stem Cells Inc.	USA	HuCNS-SC	Subretinal	Completed	No
NCT01226628	Janssen Research and Development	USA	UTSC	Subretinal	Ongoing	No
CTRI/2010/091/000639	AIIMS	India	BMSC (autologous)	Intravitreal	Ongoing	No
NCT02016508	Al-Azhar University	Egypt	BMSC (autologous)	Intravitreal	Ongoing	No
NCT01518127	University of Sao Paulo	Brazil	BMSC (autologous)	Intravitreal	Ongoing	No
NCT01344993	Advanced cell Technologies*	USA	hESC-RPE	Subretinal	Ongoing	[53,54]
NCT02463344						
NCT01674829	CHA Biotech	Korea	hESC-RPE	Subretinal	Ongoing	[55]
NCT02749734	Southwest Hospital	China	hESC-RPE	Subretinal	Ongoing	No
NCT02755428	Chinese Academy of Sciences	China	hESC-RPE	Subretinal	Ongoing	No
NCT02286089	Cell Cure Neurosciences Ltd.	Israel	hESC-RPE	Subretinal	Ongoing	No
NCT01691261	Pfizer, UCL	UK	hESC-RPE	Subretinal	Ongoing	No
UMIN000011929	Riken	Japan	iPSC-RPE	Subretinal	Ongoing	[56]

*Aliases Ocata Therapeutics; Astellas Institute of Regenerative Medicine. CTRI: Clinical trials registry India, NCT: National Clinical Trial, AIIMS: All India Institute of Medical Sciences, UCL: University College of London, UMIN: University Medical Information Network, HuCNS-SC: Human central nervous system-stem cell, UTSC: Umbilical tissue-derived stem cell, BMSC: Bone marrow-derived stem cell, hESC-RPE: Human embryonic stem cell-derived retinal pigment epithelium, iPSC-RPE: Induced pluripotent stem cell-derived retinal pigment epithelium

Source of Functional Cells during Cell Replacement Therapies

Human fetal retinal pigment epithelial

The first RPE transplant in AMD patients involved human fetal RPE cultured *in vitro*. The *in vitro* cultured human fetal RPE cells were transplanted as a patch into the foveal region after membrane excision.^[57,58] In another trial, fetal RPE cell suspension was subretinally injected for rescuing dry AMD.^[59] In both scenarios, the transplantation led to rejection of graft and no significant visual improvement.^[58] In the pursuit of autologous cells for transplantation, iris pigment epithelial (IPE) cells obtained by peripheral iridectomy surgery was expanded *in vitro* in culture followed by subretinal transplantation of the cells. The results showed visual acuity improvement in approximately 80% of the patients with minimal complications.^[60] However, the procedure of obtaining IPE cells itself was considered to be complicated, and the IPE cells *in vitro*, although capable of phagocytosis of rod photoreceptor outer segment, is considered to lack enzymes involved in retinoid visual cycle.^[61] Both fetal RPE and IPE do not have the ideal characteristics of cells for RPE replacement strategy. Recently, pluripotent stem cells' source such as human embryonic stem cell (hESC) has been shown to be a renewable source or functional RPE cells.

Human embryonic stem cells

Unlike adult stem cells which are either unipotent or multipotent, ES cells are pluripotent and can differentiate into almost all the cells in the body except for the placental tissues. Recently, several studies have shown the capacity of hESCs to differentiate toward RPE cells.^[62-69] Currently, there are at least seven protocols available for RPE differentiation from hESCs. The protocols include spontaneous differentiation methods, induction by stromal cell-derived factors, serum-free floating culture of embryoid body-like aggregates, retinal determination, sorting of spherical neural masses,

small-molecule-based induction, and three-dimensional culture.^[70] The hESC-derived RPE cells expressed RPE-specific transcripts involved in melanin production and visual retinoid cycle. Global gene expression revealed significant similarity to human fetal RPE. In addition, the studies on hESC-derived RPE confirmed the potential of these cells to phagocytose rod photoreceptor outer segments.^[65,69,71] Recently, clinical trials utilizing hESC-derived RPE cells for the treatment of AMD is in progress worldwide.

Induced pluripotent stem cells

hESCs, although renewable and has the potential to differentiate into RPE, suffer from limitations such as immunogenicity and related ethical issues. In 2006, the autologous and ethical source of pluripotent stem cells was discovered by Takahashi and Yamanaka. In this study, it was established that introduction of the pluripotency factors, namely, Oct4, Sox2, Klf4, and cMyc, is sufficient to induce pluripotency in somatic cells. The cells that are reprogrammed through the pluripotency factors are referred to as induced pluripotent stem cells (iPSCs).^[72] These reprogrammed cells are shown to be similar to ESCs with respect to their morphological, immunocytochemical, and differentiation properties. The global genetic profiles of these cells are mostly similar to hESCs. However, they do not have the limitations that are associated with the hESCs, such as the ethical issues and immune rejection. Various sources of cells including peripheral blood monocytes, NSCs, and primordial germ cells have been used for reprogramming. Both viral-based and nonviral strategies have been widely employed. The nonintegrative strategies by means of Sendai viruses and episomal vector transfection are currently employed to generate most iPSC lines.^[73-78] With respect to the protocols for deriving RPE from the iPSC lines, several studies have successfully employed the protocols already established in hESCs on most iPSC lines.^[67,79-85] Almost all of these studies provide evidence that iPSCs have potential similar to hESCs in terms of RPE differentiation.

Clinical Trials Using Pluripotent Stem Cell-Derived Retinal Pigment Epithelial

The clinical trials and their outcomes are shown in Table 1. The first clinical trial using hESC-derived RPE cells was performed by Advanced Cell Technology (Santa Monica, California, USA) in 2011. This Phase I/II clinical trial was carried out to understand the safety and efficiency of hESC-derived RPE transplantation on advanced dry AMD and Stargardt's disease (clinical trial registration number: NCT01345006 and NCT01344993).^[53] The preliminary results of the clinical trial established that the subretinal injection of 5×10^4 hESC-derived RPE cells in two patients, one with dry AMD and the other with Stargardt's disease, did not lead to teratoma or immune rejection. The clinical trial suggested that hESC-derived RPE cells are safe and lead to marginal improvement in visual acuity. Recently, the medium to long-term safety and efficiency of the clinical trial outcomes was published. Dose escalation study revealed that 1.5×10^5 RPE cells were well tolerated in most patients with increased visual acuity from 16 to 25 letters over a period of 3–12 months posttransplantation.^[54] Although the clinical trial did not lead to any adverse events, the use of immunosuppressive drugs has been looked into as one of the disadvantages of the procedure. A commentary by Zhang *et al.* suggested that future clinical trials should include optimization of dose and duration of immune suppressive regimen necessary during the subretinal procedure and inclusion of a larger cohort to decipher both safety and efficacy of the procedure.^[86] In addition, Sunness, 2015, suggested that the patients who are recruited in the clinical trial should undergo microperimetry analysis and low-vision training before the subretinal procedure to authenticate the absolute visual improvement of the procedure and to remove the bias that can exist among the individuals.^[87] The authors recently published a reply indicating that the microperimetry analysis was in fact carried out on some patients and suggested that it did not lead to any significant difference in the reported results.^[88] However, they noted merits in the comments provided by Zhang *et al.* and agreed that the incorporation of the suggestions in future clinical trials will provide an unbiased assessment of the treatment.^[86,88] The findings of this clinical trial were reproduced and reported recently by an independent trial conducted in Korea.^[55]

The first clinical trial on utilizing iPSCs for AMD was initiated by Masayo Takahashi, RIKEN, Japan, in September 2014. However, the study was suspended in March 2015.^[56] The study reported mutations in the second patient's iPSCs that were not detectable in the patient's original fibroblasts. The mutations included three single-nucleotide variations and three copy-number variants. It is not definitely known whether the reprogramming process induced the iPSC abnormalities, although iPSCs often acquire mutations and epigenetic and chromosomal changes in culture.^[56] The current report on the trial revealed that the trial protocol was modified to utilize allogeneic iPSC cells instead of autologous iPSC lines. Toward this, an iPSC cell line bank is being created at the center for iPSC cell research and application at Kyoto University, Japan. These iPSC lines are being created from human leukocyte antigen (HLA) typed peripheral blood and cord blood samples. It is expected that the RPE differentiated from the HLA-matched allogeneic iPSCs could be relatively less expensive and could

be used without immunosuppressive therapy, thereby being advantageous than the hESC-RPE.

Current Scenario on Cell-based Therapy for Retinal Degeneration in India

With respect to the clinical trials conducted for retinal degenerations in India, the first long-term safety study of transplanting human fetal neuroretinal cells was carried out by Dr. Taraprasad Das, from L. V. Prasad Eye Institute, India. In this Phase I clinical trial, fetal neuroretinal cells isolated from 14–18-week gestation were transplanted into subretinal space into 14 patients with advanced RP. A constant dose of cells (0.6 million cells in 150 μ l) in suspension was injected into the subretinal space. The study did not observe any detrimental effect at least for 40-month posttransplant except for a retinal detachment in one patient.^[89] However, the status of the Phase II study is not known. Currently, a clinical trial is being carried out by the All India Institute of Medical Sciences, where patients with AMD and RP are treated with intravitreal injections of autologous bone marrow-derived stem cells under the Clinical Trial Registry of India CTRI/2010/091/000639. The results of this trial are yet to be published. With regard to the pluripotent stem cells, there are a few reports available on the transplantation of hESCs in several degenerative conditions and emphysematous chronic obstructive pulmonary disease (COPD) hESCs in the treatment of emphysematous COPD: A case report.^[90] These reports are rudimentary and inconclusive on the actual safety and efficacy of these pluripotent stem cells. Recently, Mariappan *et al.* established the feasibility of generating RPE cells from Indian hESC line BJNh20.^[91] With respect to the use of iPSCs in ophthalmology, there has been a single study utilizing mouse iPSCs to generate retinal progenitors and RPE.^[82] However, there are no reports available on the differentiation of human iPSC lines to retinal cells from India.

Conclusion: Toward Pluripotent Stem Cell-based Clinical Trials in India

Fig. 1 shows the step that needs to be taken toward clinical trials for AMD in India. Collective efforts from reputed ophthalmic institutes are required for feasibility of clinical

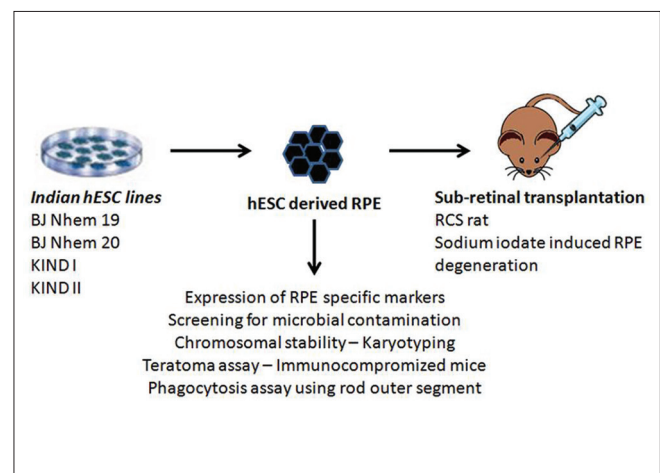


Figure 1: Schematic showing the steps toward clinical trials for age-related macular degeneration in India

trials for AMD in India. Efforts are required to characterize hESC lines that are generated in India (BJNhem19 and 20 and KINDI and KINDII) for their potential to differentiate into RPE. In addition, preclinical studies on animal models of AMD are required to confirm the feasibility of the approach before the clinical trials on the patients become a reality [Fig. 1]. The alternative approach would be to join hands with Steven Schwartz of Advance Cell Technology and initiate the clinical studies in India through proper channel. This approach will hasten the clinical work in India as the earlier approach has some bottlenecks. The vital step of formulating the guidelines for conducting research and clinical trials utilizing pluripotent stem cells in India has already been clearly laid down by the Department of Biotechnology and the Indian Council of Medical Research which ensures ethical and unbiased evaluation of these procedures. With a clear regulatory guideline in place and progress made in the pluripotent stem cell-based research, we can expect the clinical trials to initiate in the near future.

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Conflicts of interest

There are no conflicts of interest.

References

- Krishnan T, Ravindran RD, Murthy GV, Vashist P, Fitzpatrick KE, Thulasiraj RD, *et al.* Prevalence of early and late age-related macular degeneration in India: The INDEYE study. *Invest Ophthalmol Vis Sci* 2010;51:701-7.
- Rakoczy PE, Zhang D, Robertson T, Barnett NL, Papadimitriou J, Constable IJ, *et al.* Progressive age-related changes similar to age-related macular degeneration in a transgenic mouse model. *Am J Pathol* 2002;161:1515-24.
- Khan M, Agarwal K, Loutfi M, Kamal A. Present and possible therapies for age-related macular degeneration. *ISRN Ophthalmol* 2014;2014:608390.
- Iriyama A, Fujiki R, Inoue Y, Takahashi H, Tamaki Y, Takezawa S, *et al.* A2E, a pigment of the lipofuscin of retinal pigment epithelial cells, is an endogenous ligand for retinoic acid receptor. *J Biol Chem* 2008;283:11947-53.
- Chakravarthy U, Augood C, Bentham GC, de Jong PT, Rahu M, Seland J, *et al.* Cigarette smoking and age-related macular degeneration in the EUREYE Study. *Ophthalmology* 2007;114:1157-63.
- Finkel T, Holbrook NJ. Oxidants, oxidative stress and the biology of ageing. *Nature* 2000;408:239-47.
- Evans JR, Lawrenson JG. Antioxidant vitamin and mineral supplements for slowing the progression of age-related macular degeneration. *Cochrane Database Syst Rev* 2012;11:CD000254.
- Virgili G, Bini A. Laser photocoagulation for neovascular age-related macular degeneration. *Cochrane Database Syst Rev* 2007;3: CD004763.
- Argon laser photocoagulation for neovascular maculopathy. Five-year results from randomized clinical trials. Macular Photocoagulation Study Group. *Arch Ophthalmol* 1991;109:1109-14.
- Laser photocoagulation of subfoveal recurrent neovascular lesions in age-related macular degeneration. Results of a randomized clinical trial. Macular Photocoagulation Study Group. *Arch Ophthalmol* 1991;109:1232-41.
- Persistent and recurrent neovascularization after laser photocoagulation for subfoveal choroidal neovascularization of age-related macular degeneration. Macular Photocoagulation Study Group. *Arch Ophthalmol* 1994;112:489-99.
- Verteporfin Roundtable and Participants; Treatment of Age-Related Macular Degeneration with Photodynamic Therapy (TAP) Study Group Principal Investigators; Verteporfin in Photodynamic Therapy (VIP) Study Group Principal Investigators. Guidelines for using verteporfin (visudyne) in photodynamic therapy to treat choroidal neovascularization due to age-related macular degeneration and other causes. *Retina* 2002;22:6-18.
- Kaiser PK; Visudyne In Occult CNV (VIO) Study Group. Verteporfin PDT for subfoveal occult CNV in AMD: Two-year results of a randomized trial. *Curr Med Res Opin* 2009;25:1853-60.
- Gragoudas ES, Adamis AP, Cunningham ET Jr., Feinsod M, Guyer DR; VEGF Inhibition Study in Ocular Neovascularization Clinical Trial Group. Pegaptanib for neovascular age-related macular degeneration. *N Engl J Med* 2004;351:2805-16.
- Ruckman J, Green LS, Beeson J, Waugh S, Gillette WL, Henninger DD, *et al.* 2'-Fluoropyrimidine RNA-based aptamers to the 165-amino acid form of vascular endothelial growth factor (VEGF165). Inhibition of receptor binding and VEGF-induced vascular permeability through interactions requiring the exon 7-encoded domain. *J Biol Chem* 1998;273:20556-67.
- Brown DM, Michels M, Kaiser PK, Heier JS, Sy JP, Ianchulev T; ANCHOR Study Group. Ranibizumab versus verteporfin photodynamic therapy for neovascular age-related macular degeneration: Two-year results of the ANCHOR study. *Ophthalmology* 2009;116:57-65.e5.
- Rosenfeld PJ, Brown DM, Heier JS, Boyer DS, Kaiser PK, Chung CY, *et al.* Ranibizumab for neovascular age-related macular degeneration. *N Engl J Med* 2006;355:1419-31.
- Boyer DS, Heier JS, Brown DM, Francom SF, Ianchulev T, Rubio RG. A Phase IIIb study to evaluate the safety of ranibizumab in subjects with neovascular age-related macular degeneration. *Ophthalmology* 2009;116:1731-9.
- CATT Research Group, Martin DF, Maguire MG, Ying GS, Grunwald JE, Fine SL, *et al.* Ranibizumab and bevacizumab for neovascular age-related macular degeneration. *N Engl J Med* 2011;364:1897-908.
- Brown DM, Heier JS, Ciulla T, Benz M, Abraham P, Yancopoulos G, *et al.* Primary endpoint results of a phase II study of vascular endothelial growth factor trap-eye in wet age-related macular degeneration. *Ophthalmology* 2011;118:1089-97.
- Heier JS, Brown DM, Chong V, Korobelnik JF, Kaiser PK, Nguyen QD, *et al.* Intravitreal aflibercept (VEGF trap-eye) in wet age-related macular degeneration. *Ophthalmology* 2012;119:2537-48.
- Butovsky O, Koronyo-Hamaoui M, Kunis G, Ophir E, Landa G, Cohen H, *et al.* Glatiramer acetate fights against Alzheimer's disease by inducing dendritic-like microglia expressing insulin-like growth factor 1. *Proc Natl Acad Sci U S A* 2006;103:11784-9.
- Landa G, Butovsky O, Shoshani J, Schwartz M, Pollack A. Weekly vaccination with Copaxone (glatiramer acetate) as a potential therapy for dry age-related macular degeneration. *Curr Eye Res* 2008;33:1011-3.
- Landa G, Rosen RB, Patel A, Lima VC, Tai KW, Perez VR, *et al.* Qualitative spectral OCT/SLO analysis of drusen change in dry age-related macular degeneration patients treated with Copaxone. *J Ocul Pharmacol Ther* 2011;27:77-82.
- Wong WT, Dresner S, Forooghian F, Glaser T, Doss L, Zhou M, *et al.* Treatment of geographic atrophy with subconjunctival sirolimus: Results of a phase I/II clinical trial. *Invest Ophthalmol Vis Sci* 2013;54:2941-50.
- Gehlbach P, Li T, Hatfield E. Statins for age-related macular degeneration. *Cochrane Database Syst Rev* 2012;3: CD006927.

27. Buschini E, Fea AM, Lavia CA, Nassisi M, Pignata G, Zola M, *et al.* Recent developments in the management of dry age-related macular degeneration. *Clin Ophthalmol* 2015;9:563-74.
28. Machemer R, Steinhorst UH. Retinal separation, retinotomy, and macular relocation: II. A surgical approach for age-related macular degeneration? *Graefes Arch Clin Exp Ophthalmol* 1993;231:635-41.
29. Cahill MT, Mruthyunjaya P, Bowes Rickman C, Toth CA. Recurrence of retinal pigment epithelial changes after macular translocation with 360 degrees peripheral retinectomy for geographic atrophy. *Arch Ophthalmol* 2005;123:935-8.
30. McGill TJ, Cottam B, Lu B, Wang S, Girman S, Tian C, *et al.* Transplantation of human central nervous system stem cells - neuroprotection in retinal degeneration. *Eur J Neurosci* 2012;35:468-77.
31. Cuenca N, Fernández-Sánchez L, McGill TJ, Lu B, Wang S, Lund R, *et al.* Phagocytosis of photoreceptor outer segments by transplanted human neural stem cells as a neuroprotective mechanism in retinal degeneration. *Invest Ophthalmol Vis Sci* 2013;54:6745-56.
32. Friedenstein AJ, Chailakhjan RK, Lalykina KS. The development of fibroblast colonies in monolayer cultures of guinea-pig bone marrow and spleen cells. *Cell Tissue Kinet* 1970;3:393-403.
33. Stewart MC, Stewart AA. Mesenchymal stem cells: Characteristics, sources, and mechanisms of action. *Vet Clin North Am Equine Pract* 2011;27:243-61.
34. Huang GT, Gronthos S, Shi S. Mesenchymal stem cells derived from dental tissues vs. those from other sources: Their biology and role in regenerative medicine. *J Dent Res* 2009;88:792-806.
35. Marquez-Curtis LA, Janowska-Wieczorek A, McGann LE, Elliott JA. Mesenchymal stromal cells derived from various tissues: Biological, clinical and cryopreservation aspects. *Cryobiology* 2015;71:181-97.
36. Hoogduijn MJ, Betjes MG, Baan CC. Mesenchymal stromal cells for organ transplantation: Different sources and unique characteristics? *Curr Opin Organ Transplant* 2014;19:41-6.
37. Mosna F, Sensebé L, Krampera M. Human bone marrow and adipose tissue mesenchymal stem cells: A user's guide. *Stem Cells Dev* 2010;19:1449-70.
38. Majore I, Moretti P, Stahl F, Hass R, Kasper C. Growth and differentiation properties of mesenchymal stromal cell populations derived from whole human umbilical cord. *Stem Cell Rev* 2011;7:17-31.
39. Bieback K, Netsch P. Isolation, culture, and characterization of human umbilical cord blood-derived mesenchymal stromal cells. *Methods Mol Biol* 2016;1416:245-58.
40. Steigman SA, Fauza DO. Isolation of mesenchymal stem cells from amniotic fluid and placenta. *Curr Protoc Stem Cell Biol* 2007;35:1E.2.1-2.14.
41. Portmann-Lanz CB, Schoeberlein A, Huber A, Sager R, Malek A, Holzgreve W, *et al.* Placental mesenchymal stem cells as potential autologous graft for pre- and perinatal neuroregeneration. *Am J Obstet Gynecol* 2006;194:664-73.
42. Tamagawa T, Oi S, Ishiwata I, Ishikawa H, Nakamura Y. Differentiation of mesenchymal cells derived from human amniotic membranes into hepatocyte-like cells *in vitro*. *Hum Cell* 2007;20:77-84.
43. Macias MI, Grande J, Moreno A, Domínguez I, Bornstein R, Flores AI. Isolation and characterization of true mesenchymal stem cells derived from human term decidua capable of multilineage differentiation into all 3 embryonic layers. *Am J Obstet Gynecol* 2010;203:495.e9-495.e23.
44. Kim DW, Staples M, Shinozuka K, Pantcheva P, Kang SD, Borlongan CV. Wharton's jelly-derived mesenchymal stem cells: Phenotypic characterization and optimizing their therapeutic potential for clinical applications. *Int J Mol Sci* 2013;14:11692-712.
45. Hass R, Kasper C, Böhm S, Jacobs R. Different populations and sources of human mesenchymal stem cells (MSC): A comparison of adult and neonatal tissue-derived MSC. *Cell Commun Signal* 2011;9:12.
46. Inoue Y, Iriyama A, Ueno S, Takahashi H, Kondo M, Tamaki Y, *et al.* Subretinal transplantation of bone marrow mesenchymal stem cells delays retinal degeneration in the RCS rat model of retinal degeneration. *Exp Eye Res* 2007;85:234-41.
47. Arnhold S, Absenger Y, Klein H, Addicks K, Schraermeyer U. Transplantation of bone marrow-derived mesenchymal stem cells rescue photoreceptor cells in the dystrophic retina of the rhodopsin knockout mouse. *Graefes Arch Clin Exp Ophthalmol* 2007;245:414-22.
48. Wang S, Lu B, Girman S, Duan J, McFarland T, Zhang QS, *et al.* Non-invasive stem cell therapy in a rat model for retinal degeneration and vascular pathology. *PLoS One* 2010;5:e9200.
49. Arnhold S, Heiduschka P, Klein H, Absenger Y, Basnaoglu S, Kreppel F, *et al.* Adenovirally transduced bone marrow stromal cells differentiate into pigment epithelial cells and induce rescue effects in RCS rats. *Invest Ophthalmol Vis Sci* 2006;47:4121-9.
50. Lund RD, Wang S, Lu B, Girman S, Holmes T, Sauvé Y, *et al.* Cells isolated from umbilical cord tissue rescue photoreceptors and visual functions in a rodent model of retinal disease. *Stem Cells* 2007;25:602-11.
51. Li Y, Reza RG, Atmaca-Sonmez P, Ratajczak MZ, Ildstad ST, Kaplan HJ, *et al.* Retinal pigment epithelium damage enhances expression of chemoattractants and migration of bone marrow-derived stem cells. *Invest Ophthalmol Vis Sci* 2006;47:1646-52.
52. Atmaca-Sonmez P, Li Y, Yamauchi Y, Schanie CL, Ildstad ST, Kaplan HJ, *et al.* Systemically transferred hematopoietic stem cells home to the subretinal space and express RPE-65 in a mouse model of retinal pigment epithelium damage. *Exp Eye Res* 2006;83:1295-302.
53. Schwartz SD, Hubschman JP, Heilwell G, Franco-Cardenas V, Pan CK, Ostrick RM, *et al.* Embryonic stem cell trials for macular degeneration: A preliminary report. *Lancet* 2012;379:713-20.
54. Schwartz SD, Regillo CD, Lam BL, Elliott D, Rosenfeld PJ, Gregori NZ, *et al.* Human embryonic stem cell-derived retinal pigment epithelium in patients with age-related macular degeneration and Stargardt's macular dystrophy: Follow-up of two open-label phase 1/2 studies. *Lancet* 2015;385:509-16.
55. Song WK, Park KM, Kim HJ, Lee JH, Choi J, Chong SY, *et al.* Treatment of macular degeneration using embryonic stem cell-derived retinal pigment epithelium: Preliminary results in Asian patients. *Stem Cell Reports* 2015;4:860-72.
56. Garber K. RIKEN suspends first clinical trial involving induced pluripotent stem cells. *Nat Biotechnol* 2015;33:890-1.
57. Algrever PV, Berglin L, Gouras P, Sheng Y. Transplantation of fetal retinal pigment epithelium in age-related macular degeneration with subfoveal neovascularization. *Graefes Arch Clin Exp Ophthalmol* 1994;232:707-16.
58. Binder S, Stanzel BV, Krebs I, Glittenberg C. Transplantation of the RPE in AMD. *Prog Retin Eye Res* 2007;26:516-54.
59. Algrever PV, Gouras P, Dalfard Kopp E. Long-term outcome of RPE allografts in non-immunosuppressed patients with AMD. *Eur J Ophthalmol* 1999;9:217-30.
60. Thumann G, Aisenbrey S, Schraermeyer U, Lafaut B, Esser P, Walter P, *et al.* Transplantation of autologous iris pigment epithelium after removal of choroidal neovascular membranes. *Arch Ophthalmol* 2000;118:1350-5.
61. Cai H, Shin MC, Tezel TH, Kaplan HJ, Del Priore LV. Use of iris pigment epithelium to replace retinal pigment epithelium in age-related macular degeneration: A gene expression analysis. *Arch Ophthalmol* 2006;124:1276-85.

62. Haruta M, Sasai Y, Kawasaki H, Amemiya K, Ooto S, Kitada M, *et al.* *In vitro* and *in vivo* characterization of pigment epithelial cells differentiated from primate embryonic stem cells. *Invest Ophthalmol Vis Sci* 2004;45:1020-5.
63. Klimanskaya I, Hipp J, Rezai KA, West M, Atala A, Lanza R. Derivation and comparative assessment of retinal pigment epithelium from human embryonic stem cells using transcriptomics. *Cloning Stem Cells* 2004;6:217-45.
64. Lund RD, Wang S, Klimanskaya I, Holmes T, Ramos-Kelsey R, Lu B, *et al.* Human embryonic stem cell-derived cells rescue visual function in dystrophic RCS rats. *Cloning Stem Cells* 2006;8:189-99.
65. Idelson M, Alper R, Obolensky A, Ben-Shushan E, Hemo I, Yachimovich-Cohen N, *et al.* Directed differentiation of human embryonic stem cells into functional retinal pigment epithelium cells. *Cell Stem Cell* 2009;5:396-408.
66. Yue F, Johkura K, Shirasawa S, Yokoyama T, Inoue Y, Tomotsune D, *et al.* Differentiation of primate ES cells into retinal cells induced by ES cell-derived pigmented cells. *Biochem Biophys Res Commun* 2010;394:877-83.
67. Maruotti J, Sripathi SR, Bharti K, Fuller J, Wahlin KJ, Ranganathan V, *et al.* Small-molecule-directed, efficient generation of retinal pigment epithelium from human pluripotent stem cells. *Proc Natl Acad Sci U S A* 2015;112:10950-5.
68. Carr AJ, Vugler A, Lawrence J, Chen LL, Ahmado A, Chen FK, *et al.* Molecular characterization and functional analysis of phagocytosis by human embryonic stem cell-derived RPE cells using a novel human retinal assay. *Mol Vis* 2009;15:283-95.
69. Buchholz DE, Pennington BO, Croze RH, Hinman CR, Coffey PJ, Clegg DO. Rapid and efficient directed differentiation of human pluripotent stem cells into retinal pigmented epithelium. *Stem Cells Transl Med* 2013;2:384-93.
70. Mu Y, Zhao M, Su G. Stem cell-based therapies for age-related macular degeneration: Current status and prospects. *Int J Clin Exp Med* 2014;7:3843-52.
71. Lu B, Malcuit C, Wang S, Girman S, Francis P, Lemieux L, *et al.* Long-term safety and function of RPE from human embryonic stem cells in preclinical models of macular degeneration. *Stem Cells* 2009;27:2126-35.
72. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 2006;126:663-76.
73. Bazley FA, Liu CF, Yuan X, Hao H, All AH, De Los Angeles A, *et al.* Direct reprogramming of human primordial germ cells into induced pluripotent stem cells: Efficient generation of genetically engineered germ cells. *Stem Cells Dev* 2015;24:2634-48.
74. Brown ME, Rondon E, Rajesh D, Mack A, Lewis R, Feng X, *et al.* Derivation of induced pluripotent stem cells from human peripheral blood T lymphocytes. *PLoS One* 2010;5:e11373.
75. Mack AA, Kroboth S, Rajesh D, Wang WB. Generation of induced pluripotent stem cells from CD34+ cells across blood drawn from multiple donors with non-integrating episomal vectors. *PLoS One* 2011;6:e27956.
76. Fusaki N, Ban H, Nishiyama A, Saeki K, Hasegawa M. Efficient induction of transgene-free human pluripotent stem cells using a vector based on Sendai virus, an RNA virus that does not integrate into the host genome. *Proc Jpn Acad Ser B Phys Biol Sci* 2009;85:348-62.
77. Seki T, Yuasa S, Fukuda K. Generation of induced pluripotent stem cells from a small amount of human peripheral blood using a combination of activated T cells and Sendai virus. *Nat Protoc* 2012;7:718-28.
78. Malik N, Rao MS. A review of the methods for human iPSC derivation. *Methods Mol Biol* 2013;997:23-33.
79. Okamoto S, Takahashi M. Induction of retinal pigment epithelial cells from monkey iPSCs. *Invest Ophthalmol Vis Sci* 2011;52:8785-90.
80. Rowland TJ, Blaschke AJ, Buchholz DE, Hikita ST, Johnson LV, Clegg DO. Differentiation of human pluripotent stem cells to retinal pigmented epithelium in defined conditions using purified extracellular matrix proteins. *J Tissue Eng Regen Med* 2013;7:642-53.
81. Maeda T, Lee MJ, Palczewska G, Marsili S, Tesar PJ, Palczewski K, *et al.* Retinal pigmented epithelial cells obtained from human induced pluripotent stem cells possess functional visual cycle enzymes *in vitro* and *in vivo*. *J Biol Chem* 2013;288:34484-93.
82. Mekala SR, Vauhini V, Nagarajan U, Maddileti S, Gaddipati S, Mariappan I. Derivation, characterization and retinal differentiation of induced pluripotent stem cells. *J Biosci* 2013;38:123-34.
83. Croze RH, Buchholz DE, Radeke MJ, Thi WJ, Hu Q, Coffey PJ, *et al.* ROCK inhibition extends passage of pluripotent stem cell-derived retinal pigmented epithelium. *Stem Cells Transl Med* 2014;3:1066-78.
84. Gong J, Fields MA, Moreira EF, Bowrey HE, Gooz M, Ablonczy Z, *et al.* Differentiation of human protein-induced pluripotent stem cells toward a retinal pigment epithelial cell fate. *PLoS One* 2015;10:e0143272.
85. Leach LL, Croze RH, Hu Q, Nadar VP, Clevenger TN, Pennington BO, *et al.* Induced pluripotent stem cell-derived retinal pigmented epithelium: A comparative study between cell lines and differentiation methods. *J Ocul Pharmacol Ther* 2016;32:317-30.
86. Zhang GY, Liao T, Fu XB, Li QF. Stem cells in age-related macular degeneration and Stargardt's macular dystrophy. *Lancet* 2015;386:29-30.
87. Sunness JS. Stem cells in age-related macular degeneration and Stargardt's macular dystrophy. *Lancet* 2015;386:29.
88. Schwartz SD, Anglade E, Lanza R; Ocata Macular Disease Investigator Group. Stem cells in age-related macular degeneration and Stargardt's macular dystrophy – Authors' reply. *Lancet* 2015;386:30.
89. Das T, del Cerro M, Jalali S, Rao VS, Gullapalli VK, Little C, *et al.* The transplantation of human fetal neuroretinal cells in advanced retinitis pigmentosa patients: Results of a long-term safety study. *Exp Neurol* 1999;157:58-68.
90. Shroff G. Human embryonic stem cells (hESCs) in the treatment of emphysematous COPD: A case report. *Clin Case Rep* 2015;3:632-4.
91. Mariappan I, Maddileti S, Joseph P, Siamwala JH, Vauhini V. Enriched cultures of retinal cells from BJNhem20 human embryonic stem cell line of Indian origin. *Invest Ophthalmol Vis Sci* 2015;56:6714-23.