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REVIEW

The role of Langerhans cells in epidermal homeostasis and pathogenesis of psoriasis

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Abstract

The skin is the main barrier between the human body and the outside world, which not only plays the role of a physical barrier but also functions as the first line of defence of immunology. Langerhans cells (LCs), as dendritic cells (DC) that play an important role in the immune system, are mainly distributed in the epidermis. This review focuses on the role of these epidermal LCs in regulating skin threats (such as microorganisms, ultraviolet radiation and allergens), especially psoriasis. Since human and mouse skin DC subsets share common ontogenetic characteristics, we can further explore the role of LCs in psoriatic inflammation.

KEYWORDS

epidermal homeostasis, inflammatory responses, Langerhans cells, psoriasis

INTRODUCTION OF PSORIASIS 1

Psoriasis is an inflammatory epidermal proliferative dermatosis which affects 2%-3% of the world's population.¹ Clinically, psoriasis is characterized by clear boundaries of skin lesions, erythema and scales on the skin.² Psoriasis was not initially considered as a disease, but a dysfunction of keratinocytes in the epidermis.³ A large number of studies have shown that the innate and adaptive immune responses of cells, especially the T-cell-mediated system, play an important role in the pathogenesis of psoriasis.⁴ In addition, the cytokine network is a key element in psoriasis. The expression levels of interleukin (IL)-1, tumour necrosis factor (TNF), interleukin-12 (IL-12),

interleukin-17 (IL-17), interleukin-22 (IL-22) and interleukin-23 (IL-23) in psoriatic skin are significantly increased. Among them, IL-17A and IL-22 have the most profound effects on keratinocytes.⁵ In psoriatic lesions, keratinocytes are activated and proliferate much faster than normal keratinocytes.⁶ Not only do cytokines have a strong influence on psoriasis but many immune cells also change greatly in psoriasis. Inflammatory cells in psoriatic skin are mainly composed of dendritic cells (DCs), macrophages, dermal T cells and epidermal neutrophils.⁷ Among these immune cells, dendritic cells increase significantly in psoriatic lesions. Although DCs play a central role in the pathogenesis of psoriasis, the specific role of each DC is not clear. At present, it is believed that psoriasis is caused by the imbalance

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of the interaction between innate immunity and adaptive immune components of skin cells, and the interconnection between innate immunity and adaptive immune system is realized by cytokines, such as TNF- α , IFN- γ and IL-1.⁴

T cells are considered to be key effector cells that have complex interactive feedback with antigen-presenting cells, neutrophils, keratinocytes and vascular endothelial cells.⁸ The production of cytokines can be activated by DCs, such as TNF- α , and after TNF- α is activated, it can, in turn, activate some secondary mediators and adhesion molecules. These activated mediators and inflammatory factors play an important role in the pathogenesis of psoriasis. DCs are divided into several types that play different roles in the human immune system and perform their respective duties.

2 | THE ORIGIN OF LANGERHANS CELLS

Dendritic cells play a very important role in the immune system, which are divided into three categories to maintain the balance of the human body: (a) DCs existed in tissue, such as in the interstitial space or the dermis, are called traditional DCs; (b) Plasmacytoid DCs; and (c) Langerhans cells (LCs). LC is a kind of stellate DC located at the base of the epidermis and was discovered by Paul Langerhans in 1868.⁹ One hundred and twenty years later, Nobel laureates Laureate Ralph Steinman and Gerold Schuler found that LCs are derived from dendritic cells that continue to differentiate from lymphoid progenitor cells differentiated from pluripotent haematopoietic stem cells.¹⁰ However, some recent studies have found that LCs are mainly derived from myeloid progenitor cells.¹¹

It is suggested that LCs were originated from primitive yolk sac haematopoietic cells, which are the precursor cells of yolk sac macrophages (Figure 1). Yolk sac macrophages are produced from the primitive red blood cell-myeloid progenitor cell (EMP) after approximately 16-18 days of pregnancy, and some of the LCs migrate to the skin in the form of yolk sac macrophages. Some LCs also migrate to the skin in the form of liver monocytes expressed by c-myb obtained by late EMP.¹² The last portion of the pluripotent haematopoietic stem cells (HSCs) produced by LCs through EMP will appear in the aorta-gonad-mesonephros (AGM) region of the human body since the 32nd day of pregnancy, when it binds to flt3 and begin to migrate to the foetal liver and then to the bone marrow, enter the skin, where they exist for a long time.¹³ C-myb and flt3 are necessary kinases in the process of HSC formation.¹⁴ Therefore, LCs are considered to be a macrophage that retains the function of DCs.¹⁵ It is mainly located in the epithelial cells of many organs and is first present in the epidermis of the skin.¹⁶ LCs



FIGURE 1 Occurrence and development of Langerhans cells (LCs). LCs are mainly differentiated from primordial erythrocytes-myeloid progenitors (EMPs) in the yolk sac. There are three pathways to induce and migrate LCs to the epidermis. The development of LCs in the epidermis depends on CSF-1R, a molecule which could maintain skin homeostasis through combining with IL-34. LCs could be replaced by mononuclear progenitors, which could bind to CSF-1, derived from bone marrow during inflammation

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account for approximately 2%-3% of skin cells.¹⁷ The development of LCs migrate to the epidermis depends on CSF-1R.¹⁸ This is also different from most DC subsets, which depend on another receptor, the tyrosine kinase FLT3.¹⁹ There are two kinds of ligands for CSf-1R. (a) IL-34, derived from keratinocytes, and (b) CSF1, derived from neutrophils, have different ligands in different skin states. CSF1R mainly binds to IL-34 in normal skin.²⁰ During inflammation, it will be replaced by bone marrow-derived mononuclear progenitor cells and bind to CSF-1.²¹ IL-34 is produced in the epidermis of embryonic developmental skin. As a ligand of CSf-1R, it plays an important role in the development of LCs and the maintenance of balance in vivo.^{22,23}

Langerhans cells are epidermal cells characterized by the expression of Langerin (CD207) and CD1a. LCs play a clear role of antigen-presenting cells in skin inflammation.²⁴ Langerin is a type II type c lectin receptor, called Birbeck granule, which is involved in the presentation of non-peptide antigens to T cells²⁵ and is expressed only in LCs in the skin.²⁶ When confronted with pathogens or allergens, LCs act as antigen presentation function triggering a series of immune responses by migrating from the epidermis to the lymph nodes by presenting antigens to T-regulatory cells. The migration of LCs in the epidermis during homeostasis and inflammation comes from different progenitor cells, which perform their respective functions.

3 | MIGRATION OF LANGERHANS CELLS DURING HOMEOSTASIS AND INFLAMMATION

Although LCs in the epidermis are mainly derived from myeloid progenitor cells, they can exist for a long time after migration to the epidermis and can renew themselves in a steady state.²⁷ After the mature LCs migrate to the epidermis, the hydrophilic E-cadherin binds to the surrounding keratinocytes, which can maintain the stability of LCs in the epidermis.²⁸ E-cadherin is a transmembrane glycoprotein that mediates Ca2⁺-dependent intercellular adhesion. It is the main component of adhesion junction and helps to maintain the integrity of the epidermal barrier.²⁹ LCs without E-cadherin showed significant morphological change: more round cell bodies and fewer dendrites. However, the absence of E-cadherin did not affect the turnover, maturation, migration and function of LCs, and E-cadherin was down-regulated during LCs mobilization and migration from the epidermis.²⁸ The non-haematopoietic origin of TGF-β1 is considered to play an important role in promoting LC renewal.³⁰ In vivo, TGFβ1 is secreted by white blood cells and non-haematopoietic cells (including keratinocytes). It has multiple effects in the immune system.³¹ There are three isotypes of TGF- β , among which TGF- β 1 is the main isotype in the immune system. The development of LCs does not require haematopoietic-derived TGF-\beta1. TGF\beta-1 from LCs directly acts on LCs through the autocrine/paracrine loop, which is necessary for LC genesis and survival,³² and it is consistent with the ability of keratinocytes to produce M-CSF and TGF- β 1.³³ The microenvironment of LCs (for example, keratinocyte signal) can induce a large number of resident langerin⁺ LCs to be proliferated²⁰; therefore, it is a crucial element of controlling LC homeostasis.

Exposure to UVB radiation or irritants induces the migration of LCs which originally resident in the epidermis and promotes the migration of bone marrow-derived Gr-1^{hi} mononuclear cells to the epidermis.²⁰ Among them, CCCTC binding factor (CTCF) can promote the outflow of LCs from the epidermis.³⁴ CTCF plays a variety of roles in the haematopoietic lineage, which can regulate the early development of thymocytes and participate in the development and the operation of macrophages.^{35,36} Gr-1 (also known as Ly-6c/G) is a monocyte with high expression of monocyte markers, and it is the precursor of LC in inflammatory epidermis.¹⁸ Macrophage inflammatory protein (MIP)-3 α , as a chemokine produced by epidermis, plays a central role in the summons of LC Precursors to epidermis.³⁷ Gr-1^{hi} monocytes express the inflammatory chemokine CCR2 receptor, which in turn promotes the secretion of proinflammatory CCR2 (monocyte chemoattractant protein 1), CCR7 and other chemokines in the skin.^{20,38}

4 | THE FUNCTION OF LANGERHANS CELLS IN THE INFLAMMATORY RESPONSE

Previous studies have told us that B cells and CD8⁺T cells are the main effectors of the adaptive immune system, while CD4⁺T cells can regulate the function of other types of lymphocytes.³⁹ The CD4⁺T-cell compartment is particularly complex because it includes Th1, Th2, Th17 and T follicle helper (TFH) cells.^{40,41} Among them, TFH cells are very important for the establishment of the germinal centre (GC), and they provide help for the expansion, selection and affinity maturation of antigen-specific B cells.⁴² DCs and LCs, as antigen-presenting cells (APCs), present endogenous peptides under the background of major histocompatibility complex (MHC) I molecules and exogenous peptides under the background of MHC II molecules to CD8⁺ and CD4⁺T cells, respectively,^{43,44} lead to induction of the adaptive immune response and gain the ability to capture and present exogenous antigens through MHC I molecules.⁴⁵ However, the secretion of IL-12 cytokines mediated by DCs and LCs can stimulate the activation of NK and $\gamma\delta T$ cells, thus destroy the target cells of MHC I molecules which lack of self-recognition. Meanwhile, NK and $\gamma\delta T$ can provide positive feedback for the maturation of DCs and LCs and promote the transmission of innate and adaptive immune responses.46

Antigen-presenting cells in the epidermis mainly consist of LCs, and their marker langerin mediates recognition by interacting with sugar conjugates such as mannan, which has a high mannose structure, or β -glucan, and expressed on the surface of pathogens.^{43,47} Langerin mediates ligand internalization for antigen processing and presentation; thus, they can be used to specifically transport antigens that bind to polysaccharides or α -langerin antibodies to LCs (Figure 2).⁴³ The activation of LCs after the binding of langerin to epidermal antigen can directly promote the IL-2-mediated signal conditions to induce the proliferation of Treg cells.⁴⁸ The migration



FIGURE 2 The role of Langerhans cells (LCs) in the occurrence and development of psoriasis. KCs stimulated by stress stimulation (such as cytokines) can stimulate the self-nucleotide and antimicrobial peptide LL-37 complex to activate epidermal LCs, inducing keratinocytes to specifically produce a variety of cytokines to promote the migration of LCs to lymph nodes (DLNs) and T-cell activation. LCs can also activate the LIGHT-LT β R signal axis to activate the expression of CCL21/CCL19 in DLN. At the same time, epidermal LCs can produce IL-2, IL-23 and TLR2. Among them, IL-2 can promote the proliferation of Treg cells; TLR2 signal mediates the activation of the transcription factor NF-κB, promotes LC maturation and migration to lymph nodes and induces Th17 differentiation. The cytokines produced by LCs promote Th17 and Th22 to activate the expression of IL-17, IL-22 and TNF-α and promote the proliferation of keratinocytes and other features of psoriasis. Proliferated KCs once again release cytokines, antimicrobial peptides and chemokines to infiltrate immune cells, further enhancing the role of activated T cells and forming a positive feedback loop between psoriatic epidermal cells and the immune system

of LCs from the epidermis is controlled by IL-1 β , IL-18 and tumour necrosis factor- α (TNF- α), which are specifically produced by keratinocytes.⁴⁹ At the same time, the expression of TNF- α induces the expression of CXCL12 in skin fibroblasts.⁵⁰ CXCR4 has also been shown to play a key role in the maturation and lymph node migration of DC.⁵¹

Therefore, the CXCR4-CXCL12 axis controls the migration of LCs to lymph nodes (DLNs). Other studies have shown that LCs can activate the LIGHT-LT β R signal axis as well as the CCL21/CCL19 expression in lymphatic endothelial cells, thus promote LCs' migration.⁵²

At this time, LCs preferentially induce the proliferation of CD4⁺T cells, which differentiate and secrete Th1 and Th2 cytokines, and the initiation and cross-activation of immature CD8⁺T cells become apparent.^{39,53} LCs are also involved in the differentiation of TFH cells, which is also very important for the induction of TFH cells and the production of germinal centres.⁵⁴ At the same time, LCs derived from CD34⁺ progenitor cells cultured in vitro can effectively promote the proliferation of CD8⁺T cells after internalization of soluble peptides.³⁹ LCs are also equipped with membrane-spanning Toll-like receptors (TLRs) to recognize pathogen-related molecular patterns (PAMPs).⁵⁵ TLR2 signalling is mediated by MyD88-dependent Tollmediated pathways, including phosphorylation of IL-1R-associated kinase (IRAK), which eventually leads to the activation of transcription factors such as NF-KB.⁵⁶ These processes promote the maturation of LCs and increase the expression of costimulatory molecules and MHC II, thus enhancing the ability antigen presentation ability of LCs. The cells also participate in the synergistic release of proinflammatory chemokines, including T-cell-stimulating factor and IL-6, IL-18 and TNF- α , the expression of adhesion molecules and the interaction between immature T cells and cells migrate to lymph nodes, inducing the differentiation of Th17.57,58 At the same time, after the skin is triggered by microbial-derived ligands (such as lipopolysaccharide [LPS] or bacterial cell wall compound peptidoglycan [PGN]) or stimulated by toxins or stimulants (risk-related molecular model, DAMP),⁵⁹ the expression levels of the LCs maturation markers CD40, CD80, HLA-DR and CCR7 and the release of IL-1 β and IL-23 are greatly enhanced. These factors are potential Th17-driven cytokines.⁵⁸ In the epidermis, TLR-2 detects the composition of mycobacteria and Gram-positive bacteria with its related receptors TLR-1 and TLR-6. Exposure to bacterial antigens also leads to the phosphorylation of the MAP kinase ERK, which promotes LCs maturation, which may contribute to the tolerance of LCs.⁵⁵ To further study LCs, scientists have identified LC-like cells that can be cultured in vitro.

5 | DIFFERENCES BETWEEN LCS AND LC-LIKE CELLS IN EPIDERMIS

According to the expression of CD11c and CD1a, the first two species of DCs in human peripheral blood were identified as CD11c⁺DC, composed of the main CD1a⁺/CD11c⁺ component (group 1) and a small amount of a CD1a⁻/CD11c⁺ component (group 2). The CD11c components are monocyte-like and have GM-CSF receptors; the third species is composed of CD1a⁻/CD11c⁻ dendritic cells (group 3), which are similar to plasma cell-like T cells and do not express GM-CSF receptors. Among them, CD1a⁺/CD11c⁺/CD14⁻DC is considered to be the direct precursor of LCs.⁶⁰ CD34 peripheral blood monocytes cultured in GM-CSF, IL-4 and TNF- α differentiate into mature DCs.⁶¹⁻⁶³ If TGF- β 1 is added to the culture medium, LCs may be formed.⁶⁴ As a substitute for human skin, many functional studies of LCs use the above-mentioned LCs derived from CD34⁺ haematopoietic progenitor cells and compare epidermal LCs and LC-like cells.

Thymic interstitial lymphopoietin (TSLP) is an interleukin-(IL)-7-like cytokine. Human LCs treated with TSLP experience phenotypic and functional maturation.⁶⁵ Epidermal LCs treated with TSLP can induce CD4 helper T cells to produce inflammatory TH2 cytokines and thymus and activation-regulated chemokine (TARC)/CCL17.^{66,67} Unlike epidermal LCs, TSLP stimulation does not enhance the survival and maturation of LC-like cells derived from CD34⁺. In addition, TSLP does not increase the immunostimulatory ability of these cells, does not induce them to produce TARC/CCL17 and cannot promote the expression of inflammatory Th2 cytokines in CD4⁺helper T cells.⁶³ Therefore, LC-like cells cannot completely replace epidermal LCs in research. However, there is also a view states that TSLP does not play a major role in controlling the proliferation of LCs in the process of development and inflammation.

6 | THE ROLE OF LANGERHANS CELLS IN PSORIASIS

6.1 | Patients with psoriasis

Since 1970, research data on the number of LCs in psoriasis have been debated, and different reports have detected an increase,^{68,69} decrease^{70,71} or stability^{72,73} of LCs in the epidermis of patients with psoriasis. However, there is a great difference in the number of LCs between normal subjects and patients with psoriasis, and the results of different experiments are different, which may related to the redistribution of LCs and the common surfacers of inflammatory DCs.⁷⁴ The onset of psoriasis has two different age

groups. Early onset occurs before the age of 40, accounting for 75% of cases, while late onset occurs after the age of 40, with a peak between 55 and 60 years old. Although these two types of psoriasis look very similar clinically, their pathogenic genes are different. The HLA-CW6 allele is associated with early-onset psoriasis but has little association with late-onset psoriasis.⁷⁵ The mobilization and migration of LCs in the epidermis of patients with early-onset psoriasis were seriously impaired, and the impaired migration is closely related to bone marrow-derived LCs.⁷⁶ Among them, early-onset chronic plaque psoriasis (CPP) is strongly linked to the inhibition of LCs migration in epidermal. LCs migration damage can be detected in both guttate psoriasis and CPP lesions. LCs migration returned to normal after remission of guttate psoriasis, but LCs will be severely damaged and unable to recover if migrated by guttate psoriasis progressed to CPP.^{77,78} The significant decrease or completely absent of LC migration in early-onset psoriasis is mainly caused by IL-18. TNF- α and contact allergen.⁷⁷ In late-onset psoriasis, keratinocytes do not secrete cytokines that inhibit LC migration.⁷⁹ This may due to the fact that LCs respond to IL-1 β rather than TNF- α . The failure of exogenous TNF- α to mobilize LCs does not necessarily reflect the non-response to TNF- α . Since LC migration needs to receive signals from both TNF- α and IL-1 β , unresponsiveness in this case may be a secondary factor in the production of IL-1^β, biological activation and/ or impaired signal transduction.⁸⁰

It was reported that LCs induce the proliferation of CD4⁺T cells under inflammatory conditions, while CD4⁺T cells, CD8⁺T cells and $\gamma\delta$ T cells produce IL-17, IL-17A, IL-17F, IL-21 and IL-22 as cytokines of Th17 cells.⁸¹ As described in Figure 2, the increased activity of p38 α mitogen-activated protein kinase is related to human susceptibility to psoriasis.⁸² The activity of p38 was also increased in KCs stimulated by stress stimulation (such as cytokines and ultraviolet radiation).⁸³ The p38 α signal in LCs rather than DCs specifically promotes the production of IL-17 in $\gamma\delta$ T and CD4⁺T cells by secreting IL-23 and IL-6, which are essential for the development of the psoriasis.⁸⁴

In addition, studies have shown that the activation of Stat3 in keratinocytes may affect the activation of LCs at least partly through IL-1 α stimulation, and their existence is related to the occurrence or aggravation of psoriasis, called the Koebner phenomenon, which is caused by IL-23.^{85,86} This activation leads to the intraepidermal circuit of KC-LCs and the activation of the IL-23/IL-17 axis, which leads to the occurrence and development of psoriasis. Human β -defensin 3 (HBD3) is a small antimicrobial peptide that has a chemotactic effect on immune cells and plays a small role in promoting the development of psoriasis by inducing the increase of IL-23 produced by epidermal LCs.^{87,88}

Moreover, LCs can induce whole peripheral T cells and immature CD4⁺T cells to produce IL-22, a cytokine that mainly acts on epithelial cells. In the skin, it produces antimicrobial proteins, such as HBD3, indicating that this cytokine is involved in the defence of early hosts against microbial pathogens,⁸⁹ and mediates keratinocyte proliferation and epidermal hyperplasia, which is thought to play an important role in inflammatory diseases with obvious epidermal acanthosis, such as psoriasis.⁸⁹ It has been shown that not only Th17 cells but also T helper cells can produce IL-22, and the type of T helper cells that only produce IL-22 is called Th22.⁹⁰ It is considered that the DCs in the skin, especially the IL-22 cytokines induced by LCs to penetrate into the skin, are produced by Th22 cells.⁹¹ Because there are still many differences between LCs and human epidermal LCs in experiments performed in vitro, more people choose to create mouse models to study LCs.

6.2 | Mouse model of psoriasis

Human skin is different from mouse skin in terms of its anatomy. Human skin is divided into epidermis and dermis, while the epidermis is divided into four layers: a stratum corneum, a transparent layer, a granular layer and a germinal layer, all composed of lamellar flat epithelium. The dermis is divided into two layers: a papillary layer and a reticular layer, and mainly composed of dense connective tissue. The mouse skin has only 2-3 layers, covered under dense hair follicles.⁹² The DCs of mouse skin can be divided into LCs, Langerin⁺DCs and Langerin-DCs. Dermal DCs consist of two main subgroups: Langerin-DC (CD103⁺CD11b⁻) and Langerin-DC (CD103⁻CD11b⁺). The difference between LCs and dermal Langerin-DCs lies in the high expression of CD11b and epithelial cell adhesion molecule (EpCAM) as well as the low expression of CD103.93 At present, the study of LCs mainly uses langerin-diphtheria toxin A (DTA) or diphtheria toxin receptor (DTR) mice with LC gene defects to establish the model of psoriatic dermatitis. In addition, three independent mouse lines have been designed to effectively ablate the endogenous Langerin site introduced by LCs; thus, the primate diphtheria toxin receptor (DTR) and the Langerin-DTR (muLangerin-DTR) mouse strain were established⁹⁴ using human genomic BAC DNA, human Langerin-DTA (huLangerin-DTA) and huLangerin-DTR transgenic mice containing Langerin loci expressing active Diphtheria toxin or DTR.⁹⁵

The mouse psoriatic dermatitis models have been generated by different ways, such as local administration of IL-23, topical application Imiquimod (IMQ) as ligands of TLR7 and TLR8, and the deletion of cre recombinant enzyme in keratin 5 expressing cells (Jun^{f/f} JunB^{f/f} K5creER = DKO * mice) induced by tamoxifen (TX) lead to acanthosis, hypokeratosis and mixed inflammatory infiltration in mice with common psoriasis⁹⁶⁻⁹⁸ (Figure 3). The severity of dermatitis induced by IMQ in LC gene-deficient mice is significantly lower than that in wild-type mice. It is considered that LCs play a very important role in the inflammatory response of psoriatic dermatitis induced by IMQ.⁹⁹ The development of these three common psoriatic dermatitis models is very dependent on IL-23/IL-17/IL-22 axis.^{96,100,101}

In the comparison of the skin transcriptional group of the psoriasis mouse model, the transcriptional group in the IL-23 injection model was most similar to the gene expression pattern found in psoriatic skin.¹⁰² Subcutaneous injection of IL-23, together with IL-12 and TNF produced by LCs and DCs, activates $\alpha\beta T$ and $\gamma\delta T$ cells to produce IL-22 and IL-17A/F, which stimulate keratinocyte proliferation, release S100 protein and β -defensin, and promote the production of growth factors and chemokines in psoriasis.¹⁰⁰

The accumulation of LCs in the epidermis of psoriatic skin lesions induced by IMQ is mainly related to the local proliferation of the LC pool. Local treatment with IMQ for 5-7 days can induce the increase of LC, the production of IL-23, and $\gamma\delta T$ cells which produce IL-17A in mice.¹⁰³ These $\gamma\delta T$ cells can migrate to the epidermis



FIGURE 3 The three kinds of way to establish Psoriasis-like Mouse Model. The first type of psoriasis-like mouse model was intradermal injection of 500 mg of IL-23 every other day for 15 days; the second kind was to smear 62.5 mg of imiquimod on the back every day for 6 days; and the third kind was intraperitoneal injection of tamoxifen for three consecutive days of 1 mg in DKO* mice, which could lead to acanthosis, incomplete keratosis and mixed inflammatory infiltration after 14 days of IMQ-treated skin through CCR6 to further develop psoriasis dermatoses.¹⁰⁴ CCR6 was demonstrated to be expressed on all peripheral T cells expressing IL-17A/F and IL-22.¹⁰⁵ Other studies have shown that Langerin-DTR mice could reduce the inflammation of psoriasis-like dermatitis induced by IMQ and reduce the number of $\gamma\delta T$ cells produced by permeable IL-17A. In the mouse model, it was found that CD1a, a lipid-presenting molecule heavily expressed on LCs, could amplify the inflammatory response mediated by Th17 cells in response to self-lipid antigens.¹⁰⁶ The role of Langerin epidermal LC in inducing the TFH-cell response was confirmed in huLangerin-DTR mice.⁵⁴ The mouse model of psoriasis induced by IMQ mentioned above is only used to study the early stages of the disease.¹⁰⁷ Some studies have adjusted the IMQ treatment time to 14 days to better simulate human conditions and study the late stages of the disease. Langerin-DTR mice can induce IMO-induced psoriatic dermatitis and promote a large number of epidermal neutrophils in the late stage.¹⁰⁸ However, it is still controversial whether this method can really represent the late stage of psoriasis.

Interestingly, the third type of psoriatic model using DKO* mice showed chronic inflammation, remained stable over a long period of time and was thought to mimic the chronic phase of human disease. Studies have shown that in this mouse model, bone marrow-derived LCs decreased significantly, while IL-23 content increased, but did not affect the expression levels of IL-17 and IL-22.¹⁰⁹ The chronic development of this psoriatic dermatitis model may depend on the direct action of IL-23 on keratino-cytes.¹¹⁰ Therefore, the proliferative and activated LCs found in the epidermis In melanoma, the decrease in the number of LC is due to the loss of the antigen presentation function of LC, which promotes the continued survival of tumour cells' immunomodulatory effect in the late stage.

7 | CONCLUSION

As immune cells of the epidermis, LC is essential for sensing danger and triggering congenital and adaptive protective responses in the body. In other skin diseases, such as Atopic dermatitis, LC depletion can help to relief the disease.¹¹¹ In melanoma, the decrease in the number of LC is due to the loss of the antigen presentation function of LC, which promotes the continued survival of tumour cells.¹¹² However, the role and function in psoriasis are controversial; it is possible that there is a certain heterogeneity of LC, there are new functional subsets have not been found, which is also an important content of future research on the role of LC in psoriasis.

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CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

AUTHOR CONTRIBUTION

Bei Yan: Writing – original draft (equal). Nian Liu: Methodology (equal). Jie Li: Writing – review and editing (equal). Jiaoduan Li: Methodology (equal). Wu Zhu: Formal analysis (equal). Yehong Kuang: Formal analysis (equal). Xiang Chen: Conceptualization (equal); project administration (equal). Cong Peng: Writing – review and editing (equal).

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