Contents lists available at ScienceDirect

Genomics Data

journal homepage: <www.elsevier.com/locate/gdata>

Data in Brief

Draft genome sequence of Kocuria sp. SM24M-10 isolated from coral mucus

Bruna Rafaella Z. Palermo ^{a, 1}, Daniel B.A. Castro ^{a, 1}, Letícia Bianca Pereira ^{a, 1}, Ana Carolina G. Cauz ^b, Beatriz L. Magalhães ^b, Camila Carlos ^a, Fernanda L.P. da Costa ^b, Guilherme P. Scagion ^b, Juliana S. Higa ^c, Ludimila D. Almeida ^b, Meiriele da S. das Neves ^b, Melina Aparecida Cordeiro ^b, Paula F.V. do Prado ^b, Thiago M. da Silva ^a, Thiago Willian A. Balsalobre ^a, Luciana C. Paulino ^d, Renato Vicentini ^a, Lúcio F.C. Ferraz ^e, Laura M.M. Ottoboni ^{a,}*

a Center for Molecular Biology and Genetic Engineering (CBMEG), State University of Campinas - UNICAMP, Campinas, Brazil

^b Department of Genetics, Evolution and Bioagents, State University of Campinas — UNICAMP, Campinas, Brazil

^c Institute Butantan, São Paulo, Brazil

^d Center of Natural and Human Sciences, Federal University of ABC, Santo André, Brazil

^e Department of Molecular Biology and Pharmacology, University of São Francisco, Bragança Paulista, Brazil

article info abstract

Article history: Received 10 December 2015 Accepted 18 December 2015 Available online 19 December 2015

Keywords:

Kocuria sp. SM24M-10 Coral mucus Osmotic stress Oxidative stress

Here, we describe the genomic features of the Actinobacteria Kocuria sp. SM24M-10 isolated from mucus of the Brazilian endemic coral Mussismilia hispida. The sequences are available under accession number LDNX01000000 (http://www.ncbi.nlm.nih.gov/nuccore/LDNX00000000). The genomic analysis revealed interesting information about the adaptation of bacteria to the marine environment (such as genes involved in osmotic and oxidative stress) and to the nutrient-rich environment provided by the coral mucus.

© 2015 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Direct link to deposited data

http://www.ncbi.nlm.nih.gov/nuccore/LDNX00000000

2. Experimental design, materials and methods

The Kocuria genus was established in 1995 by a taxonomic dissection of the Micrococcus genus [\[1\]](#page-1-0). So far, it comprises the species Kocuria rosea, Kocuria varians, Kocuria kristinae [\[1\],](#page-1-0) Kocuria rhizophila, Kocuria palustris [\[2\]](#page-1-0), Kocuria polaris [\[3\]](#page-1-0), Kocuria marina [\[4\]](#page-1-0), Kocuria atrinae [\[5\],](#page-1-0) Kocuria koreensis [\[6\],](#page-1-0) Kocuria gwangalliensis [\[7\],](#page-1-0) Kocuria himachalensis [\[8\]](#page-1-0), Kocuria aegyptia [\[9\],](#page-1-0) Kocuria flava, Kocuria turfanensis [\[10\]](#page-1-0), Kocuria salsicia [\[11\]](#page-1-0), Kocuria halotolerans [\[12\],](#page-1-0) Kocuria carniphila [\[13\],](#page-1-0) Kocuria sediminis [\[14\]](#page-1-0), and Kocuria assamensis [\[15\].](#page-1-0) Members of the Kocuria genus can be found in soil, mammal skin, the rhizosphere, clinical samples, and several other ecological niches, such as the Antarctic cyanobacterial mat [\[3\],](#page-1-0) marine sediment [\[4\],](#page-1-0) fermented food [\[5,6\],](#page-1-0) seawater [\[7\],](#page-1-0) and sulfide ore [\[16\]](#page-2-0). Although isolated from various sources, the size of the Kocuria genus is relatively small among the Actinomycetes, which suggests that each species of Kocuria is highly adapted to its ecological niche [\[8,9\]](#page-1-0). Here, we report a draft genome of Kocuria sp. SM24M-10 isolated from coral mucus, adding new information about this genus and its adaptation to the marine environment.

Kocuria sp. SM24M-10 was grown overnight in 3 mL of liquid GASW (Glycerol Artificial Sea Water) medium [\[17\]](#page-2-0) at 30 °C, followed by centrifugation. The supernatant was discarded and the cells were used for

2213-5960/© 2015 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Corresponding author.

E-mail address: ottoboni@unicamp.br (L.M.M. Ottoboni).

 $^{\rm 1}$ First authors who contributed equally to this work.

<http://dx.doi.org/10.1016/j.gdata.2015.12.016>

DNA extraction with the Wizard® Genomic DNA Purification Kit (Promega), according to the manufacturer's protocol. Sequencing of the genomic DNA was performed at the Life Sciences Core Facility (LaCTAD) at the State University of Campinas (UNICAMP), using a paired-end library of 400 bp in a HiSeq2500 Illumina sequencer, and yielded 65,536,228 paired-end reads of 100 bp. The estimated genome size was 4.56 Mbp, with coverage of $1437\times$, based on k-mer count [\[18,19\].](#page-2-0) All reads were preprocessed using NGS QC Toolkit v.2.2.3 [\[20\],](#page-2-0) with quality and length cutoffs of 20 and 70 bp, respectively. The assem-bly was performed with Edena v.3 [\[21\]](#page-2-0) ($m = 41$) and resulted in 177 contigs with a total length of 4,346,986 bp, N50 of 45,611 bp, and GC content of 72.35%. The identification of RNA genes, the open reading frames (ORFs), and the prediction of protein functions were performed using the Rapid Annotation Subsystem Technology (RAST) online server [\[22\]](#page-2-0), with the gene caller GLIMMER 3.0 [\[23\].](#page-2-0) Kocuria sp. SM24M-10 presented 4013 coding sequences (CDSs), 49 tRNA genes, and 6 rRNA genes (two 5S RNA, two 16S, and two 23S). A total of 1650 (42%) proteincoding genes were classified in 404 subsystems, and a predicted function was assigned to 1574 of them (Table 1).

The genomic analysis of Kocuria sp. SM24M-10 revealed interesting information about its adaptation to the marine environment and to the nutrient-rich environment provided by the coral mucus. The genome presented 22 genes associated with osmotic stress, and 18 of these genes were classified as being involved in betaine uptake and biosynthesis. Betaine is an efficient osmolyte that is promptly accumulated in large amounts in response to osmotic stress [\[24,25\]](#page-2-0). Due to symbiosis with zooxanthellae algae [\[26\]](#page-2-0), coral tissues may contain exceptionally high levels of oxygen, which can generate oxidative stress. For this reason, it is expected that bacteria associated with coral are able to produce enzymes to overcome the toxic effects of reactive oxygen species [\[26\]](#page-2-0). Several genes involved in the oxidative stress response were found in Kocuria sp. SM24M-10. Among these genes, 14 were related to the biosynthesis of glutathione and its analogs (e.g. mycothiol). Glutathione plays a role in preventing cellular damage caused by reactive oxygen species [\[27,28\],](#page-2-0) while mycothiol provides protection against oxidative compounds as well as some classes of antibiotics [\[28\]](#page-2-0). Genes related to the catabolism of sugar alcohols, such as glycerol and inositol, were also observed in the bacteria genome. Both glycerol and inositol are important photosynthetic products that are released by symbiotic Symbiodinium algae and are used by corals [\[29\]](#page-2-0). The presence of genes related to the use of these compounds as energy sources demonstrates the adaptation of Kocuria sp. SM24M-10 to the microenvironment provided by the coral. The genome also presented genes related to mechanisms of resistance against antibiotics and toxic compounds. Considering the fact that Kocuria sp. SM24M-10 was isolated from a marine environment exposed to chemicals derived from high levels of cargo ship traffic, we suggest that some of the resistance genes found in the genome annotation might have contributed to the survival of the bacteria in the coral microenvironment. In addition, genes related to pathways of resistance to heavy metals (cobalt, copper, mercury, and arsenic) were found in the genome.

In conclusion, the first draft genome of Kocuria sp. SM24M-10 revealed a pool of functional categories related to central metabolism, together with characteristic properties associated with the response to osmotic and oxidative stress, carbohydrate metabolism, and resistance

Table 1

Features of the Kocuria sp. SM24M-10 draft genome.

to antibiotics and toxic compounds. These data provide new insights into the adaptation and survival of microorganisms in marine environments, as well as the relationship between microorganisms and coral species.

3. Nucleotide sequence accession number

The draft genome sequence of Kocuria sp. SM24M-10 has been deposited at DDBI/EMBL/GenBank under accession number LDNX01000000.

Conflicts of interest

The authors declare no conflicts of interest concerning the work published in this paper.

Acknowledgments

CC, DBAC, GPS, LBP, LDA, MSN, PFVD, TMS, and TWAB received fellowships from the São Paulo State Research Foundation (FAPESP). BLM and MAC received fellowships from the National Council for Technological and Scientific Development (CNPq). ACGC, BRZP, and JSH received fellowships from Coordination for the Improvement of Higher Education Personnel (CAPES).

References

- [1] [E. Stackebrandt, C. Koch, O. Gvozdiak, P. Schumann, Taxonomic dissection of the](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0005) genus Micrococcus: Kocuria gen. nov., [Nesterenkonia](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0005) gen. nov., Kytococcus gen. nov., Dermacoccus gen. nov., and Micrococcus [Cohn 1872 gen. emend. Int. J. Syst.](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0005) [Bacteriol. 45 \(1995\) 682](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0005)–692.
- [2] G. Kovacs, J. Burghardt, S. Pradella, P. Schumann, E. Stackebrandt, K. Marialigeti, Kocuria palustris sp. nov. and Kocuria rhizophila sp. nov., isolated from the rhizoplane of the narrow-leaved cattail (Typha angustifolia). Int. J. Syst. Bacteriol. 49 (1999) 167–173, http://dx.doi.org[/10.1099/00207713-49-1-167](http://dx.doi.org/10.1099/00207713-49-1-167).
- [3] G.S. Reddy, J.S. Prakash, V. Prabahar, G.I. Matsumoto, E. Stackebrandt, S. Shivaji, Kocuria polaris sp. nov., an orange-pigmented psychrophilic bacterium isolated from an Antarctic cyanobacterial mat sample. Int. J. Syst. Evol. Microbiol. 53 (2003) 183–187, http://dx.doi.org[/10.1099/ijs.0.02336-0](http://dx.doi.org/10.1099/ijs.0.02336-0).
- S.B. Kim, O.I. Nedashkovskaya, V.V. Mikhailov, S.K. Han, K.O. Kim, M.S. Rhee, K.S. Bae, Kocuria marina sp. nov., a novel actinobacterium isolated from marine sediment. Int. J. Syst. Evol. Microbiol. 54 (2004) 1617–1620, http://dx.doi.org[/10.1099/ijs.0.02742-0](http://dx.doi.org/10.1099/ijs.0.02742-0).
- [5] E.J. Park, M.S. Kim, S.W. Roh, M.J. Jung, J.W. Bae, Kocuria atrinae sp. nov., isolated from traditional Korean fermented seafood. Int. J. Syst. Evol. Microbiol. 60 (2010) 914–918, http://dx.doi.org[/10.1099/ijs.0.014506-0](http://dx.doi.org/10.1099/ijs.0.014506-0).
- [6] E.J. Park, S.W. Roh, M.S. Kim, M.J. Jung, K.S. Shin, J.W. Bae, Kocuria koreensis sp. nov., isolated from fermented seafood. Int. J. Syst. Evol. Microbiol. 60 (2010) 140–143, http://dx.doi.org[/10.1099/ijs.0.012310-0.](http://dx.doi.org/10.1099/ijs.0.012310-0)
- [7] Y.B. Seo, D.E. Kim, G.D. Kim, H.W. Kim, S.W. Nam, Y.T. Kim, J.H. Lee, Kocuria gwangalliensis sp. nov., an actinobacterium isolated from seawater. Int. J. Syst. Evol. Microbiol. 59 (2009) 2769–2772, http://dx.doi.org[/10.1099/ijs.0.008482-0.](http://dx.doi.org/10.1099/ijs.0.008482-0)
- [8] S. Mayilraj, R.M. Kroppenstedt, K. Suresh, H.S. Saini, Kocuria himachalensis sp. nov., an actinobacterium isolated from the Indian Himalayas. Int. J. Syst. Evol. Microbiol. 56 (2006) 1971–1975, http://dx.doi.org[/10.1099/ijs.0.63915-0](http://dx.doi.org/10.1099/ijs.0.63915-0).
- [9] W.J. Li, Y.Q. Zhang, P. Schumann, H.H. Chen, W.N. Hozzein, X.P. Tian, L.H. Xu, C.L. Jiang, Kocuria aegyptia sp. nov., a novel actinobacterium isolated from a saline, alkaline desert soil in Egypt. Int. J. Syst. Evol. Microbiol. 56 (2006) 733–737, http://dx. doi.org[/10.1099/ijs.0.63876-0.](http://dx.doi.org/10.1099/ijs.0.63876-0)
- [10] G. Zhou, X. Luo, Y. Tang, L. Zhang, Q. Yang, Y. Qiu, C. Fang, Kocuria flava sp. nov. and Kocuria turfanensis sp. nov., airborne actinobacteria isolated from Xinjiang, China. Int. J. Syst. Evol. Microbiol. 58 (2008) 1304–1307, http://dx.doi.org[/10.1099/ijs.0.](http://dx.doi.org/10.1099/ijs.0.65323-0) [65323-0](http://dx.doi.org/10.1099/ijs.0.65323-0).
- [11] J.H. Yun, S.W. Roh, M.J. Jung, M.S. Kim, E.J. Park, K.S. Shin, Y.D. Nam, J.W. Bae, Kocuria salsicia sp. nov., isolated from salt-fermented seafood. Int. J. Syst. Evol. Microbiol. 61 (2011) 286–289, http://dx.doi.org/[10.1099/ijs.0.021469-0.](http://dx.doi.org/10.1099/ijs.0.021469-0)
- [12] S.K. Tang, Y. Wang, K. Lou, P.H. Mao, L.H. Xu, C.L. Jiang, C.J. Kim, W.J. Li, Kocuria halotolerans sp. nov., an actinobacterium isolated from a saline soil in China. Int. J. Syst. Evol. Microbiol. 59 (2009) 1316–1320, http://dx.doi.org[/10.1099/ijs.0.](http://dx.doi.org/10.1099/ijs.0.006627-0) [006627-0.](http://dx.doi.org/10.1099/ijs.0.006627-0)
- [13] L. Tvrzová, P. Schumann, I. Sedláček, Z. Páčova, C. Spröer, S. Verbarg, R.M. Kroppenstedt, Reclassification of strain CCM 132, previously classified as Kocuria varians, as Kocuria carniphila sp. nov. Int. J. Syst. Evol. Microbiol. 55 (2005) 139–142, http://dx.doi.org/[10.1099/ijs.0.63304-0](http://dx.doi.org/10.1099/ijs.0.63304-0).
- [14] M. Bala, C. Kaur, I. Kaur, F. Khan, S. Mayilraj, Kocuria sediminis sp. nov., isolated from a marine sediment sample. Antonie Van Leeuwenhoek 101 (2012) 469–478, http:// dx.doi.org[/10.1007/s10482-011-9654-2.](http://dx.doi.org/10.1007/s10482-011-9654-2)
- [15] C. Kaur, I. Kaur, R. Raichand, T.C. Bora, S. Mayilraj, Description of a novel actinobacterium Kocuria assamensis sp. nov., isolated from a water sample collected

from the river Brahmaputra, Assam, India. Antonie Van Leeuwenhoek 99 (2011) 721–726, http://dx.doi.org[/10.1007/s10482-010-9547-9.](http://dx.doi.org/10.1007/s10482-010-9547-9)

- [16] D.B.A. Castro, L.B. Pereira, M.V.M. Silva, B.P. Silva, B.R.Z. Palermo, C. Carlos, D.R.B. Belgini, E.E.G. Limache, G.V. Lacerda Júnior, M.B.P. Nery, M.B. Gomes, S.S. Souza, T.M. Silva, V.D. Rodrigues, L.C. Paulino, R. Vicentini, L.F.C. Ferraz, L.M.M. Ottoboni, High-quality draft genome sequence of Kocuria marina SO9-6, an actinobacterium isolated from a copper mine. Genomics Data (2015) 34–35, http://dx.doi.org/[10.](http://dx.doi.org/10.1016/j.gdata.2015.05.006) [1016/j.gdata.2015.05.006.](http://dx.doi.org/10.1016/j.gdata.2015.05.006)
- [17] [G.W. Smith, S.S. Hayasaka, Nitrogenase activity associated with](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0085) Halodule wrightii [roots. Appl. Environ. Microbiol. 43 \(1982\) 1244](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0085)–1248.
- [18] G. Marçais, C. Kingsford, A fast, lock-free approach for efficient parallel counting of occurrences of k-mers. Bioinformatics 27 (2011) 764–770, http://dx.doi.org/[10.](http://dx.doi.org/10.1093/bioinformatics/btr011) [1093/bioinformatics/btr011](http://dx.doi.org/10.1093/bioinformatics/btr011).
- [19] R. Li, W. Fan, G. Tian, H. Zhu, L. He, J. Cai, Q. Huang, Q. Cai, B. Li, Y. Bai, Z. Zhang, Y. Zhang, W. Wang, J. Li, F. Wei, H. Li, M. Jian, J. Li, Z. Zhang, R. Nielsen, D. Li, W. Gu,
Z. Yang, Z. Xuan, O.A. Ryder, F.C.-C. Leung, Y. Zhou, J. Cao, X. Sun, Y. Fu, X. Fang, X. Guo, B. Wang, R. Hou, F. Shen, B. Mu, P. Ni, R. Lin, W. Qian, G. Wang, C. Yu, W. Nie, J. Wang, Z. Wu, H. Liang, J. Min, Q. Wu, S. Cheng, J. Ruan, M. Wang, Z. Shi, M. Wen, B. Liu, X. Ren, H. Zheng, D. Dong, K. Cook, G. Shan, H. Zhang, C. Kosiol, X. Xie, Z. Lu, H. Zheng, Y. Li, C.C. Steiner, T.T.-Y. Lam, S. Lin, Q. Zhang, G. Li, J. Tian, T. Gong, H. Liu, D. Zhang, L. Fang, C. Ye, J. Zhang, W. Hu, A. Xu, Y. Ren, G. Zhang, M.W. Bruford, Q. Li, L. Ma, Y. Guo, N. An, Y. Hu, Y. Zheng, Y. Shi, Z. Li, Q. Liu, Y. Chen, J. Zhao, N. Qu, S. Zhao, F. Tian, X. Wang, H. Wang, L. Xu, X. Liu, T. Vinar, Y. Wang, T.- W. Lam, S.-M. Yiu, S. Liu, H. Zhang, D. Li, Y. Huang, X. Wang, G. Yang, Z. Jiang, J. Wang, N. Qin, L. Li, J. Li, L. Bolund, K. Kristiansen, G.K.-S. Wong, M. Olson, X. Zhang, S. Li, H. Yang, J. Wang, J. Wang, The sequence and de novo assembly of the giant panda genome. Nature 463 (2010) 311–317, http://dx.doi.org[/10.1038/](http://dx.doi.org/10.1038/nature08696) [nature08696](http://dx.doi.org/10.1038/nature08696).
- [20] R.K. Patel, M. Jain, NGS QC Toolkit: a toolkit for quality control of next generation sequencing data. PLoS ONE 7 (2012) e30619, http://dx.doi.org[/10.1371/journal.pone.](http://dx.doi.org/10.1371/journal.pone.0030619) [0030619.](http://dx.doi.org/10.1371/journal.pone.0030619)
- [21] D. Hernandez, P. François, L. Farinelli, M. Osterås, J. Schrenzel, De novo bacterial genome sequencing: millions of very short reads assembled on a desktop computer. Genome Res. 18 (2008) 802–809, http://dx.doi.org[/10.1101/gr.072033.107.](http://dx.doi.org/10.1101/gr.072033.107)
- [22] R.K. Aziz, D. Bartels, A. Best, M. DeJongh, T. Disz, R. Edwards, K. Formsma, S. Gerdes, E. Glass, M. Kubal, F. Meyer, G.J. Olsen, R. Olson, A.L. Osterman, R.A. Overbeek, L.K. McNeil, D. Paarmann, T. Paczian, B. Parrello, G.D. Pusch, C. Reich, R. Stevens, O. Vassieva, V. Vonstein, A. Wilke, O. Zagnitko, The RAST Server: rapid annotations using subsystems technology. BMC Genomics 9 (2008)http://dx.doi.org[/10.1186/](http://dx.doi.org/10.1186/1471-2164-9-75) [1471-2164-9-75.](http://dx.doi.org/10.1186/1471-2164-9-75)
- [23] A.L. Delcher, D. Harmon, S. Kasif, O. White, S.L. Salzberg, Improved microbial gene identification with GLIMMER. Nucleic Acids Res. 27 (1999) 4636–4641, http://dx. doi.org[/10.1093/nar/27.23.4636.](http://dx.doi.org/10.1093/nar/27.23.4636)
- [24] A. Oren, Microbial life at high salt concentrations: phylogenetic and metabolic diversity. Saline Syst. 4 (2008)http://dx.doi.org/[10.1186/1746-1448-4-2.](http://dx.doi.org/10.1186/1746-1448-4-2)
- [25] R.D. Sleator, C. Hill, Bacterial osmoadaptation: the role of osmolytes in bacterial stress and virulence. FEMS Microbiol. Rev. 26 (2002) 49–71, http://dx.doi.org/[10.](http://dx.doi.org/10.1111/j.1574-6976.2002.tb00598.x) [1111/j.1574-6976.2002.tb00598.x](http://dx.doi.org/10.1111/j.1574-6976.2002.tb00598.x).
- [26] C.B. Munn, H.K. Marchant, A.J. Moody, Defences against oxidative stress in vibrios associated with corals. FEMS Microbiol. Lett. 281 (2008) 58–63, http://dx.doi.org/ [10.1111/j.1574-6968.2008.01073.x.](http://dx.doi.org/10.1111/j.1574-6968.2008.01073.x)
- [27] [E. Cabiscol, J. Tamarit, J. Ros, Oxidative stress in bacteria and protein damage by re](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0135)[active oxygen species. Int. Microbiol. 3 \(2010\) 3](http://refhub.elsevier.com/S2213-5960(15)30117-3/rf0135)–8.
- [28] N.A. Buchmeier, G.L. Newton, T. Koledin, R.C. Fahey, Association of mycothiol with protection of Mycobacterium tuberculosis from toxic oxidants and antibiotics. Mol. Microbiol. 47 (2003) 1723–1732, http://dx.doi.org/[10.1046/j.1365-2958.2003.](http://dx.doi.org/10.1046/j.1365-2958.2003.03416.x) [03416.x.](http://dx.doi.org/10.1046/j.1365-2958.2003.03416.x)
- [29] M.S. Burriesci, T.K. Raab, J.R. Pringle, Evidence that glucose is the major transferred metabolite in dinoflagellate–cnidarian symbiosis. J. Exp. Biol. 215 (2012) 3467–3477, http://dx.doi.org[/10.1242/jeb.070946.](http://dx.doi.org/10.1242/jeb.070946)