



### **Kv7** Channels in Lung Diseases

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Lung diseases constitute a global health concern causing disability. According to WHO in 2016, respiratory diseases accounted for 24% of world population mortality, the second cause of death after cardiovascular diseases. The Kv7 channels family is a group of voltage-dependent K<sup>+</sup> channels (Kv) encoded by KCNQ genes that are involved in various physiological functions in numerous cell types, especially, cardiac myocytes, smooth muscle cells, neurons, and epithelial cells. Kv7 channel α-subunits are regulated by KCNE1-5 ancillary β-subunits, which modulate several characteristics of Kv7 channels such as biophysical properties, cell-location, channel trafficking, and pharmacological sensitivity. Kv7 channels are mainly expressed in two large groups of lung tissues: pulmonary arteries (PAs) and bronchial tubes. In PA, Kv7 channels are expressed in pulmonary artery smooth muscle cells (PASMCs); while in the airway (trachea, bronchus, and bronchioles), Kv7 channels are expressed in airway smooth muscle cells (ASMCs), airway epithelial cells (AEPs), and vagal airway C-fibers (VACFs). The functional role of Kv7 channels may vary depending on the cell type. Several studies have demonstrated that the impairment of Kv7 channel has a strong impact on pulmonary physiology contributing to the pathophysiology of different respiratory diseases such as cystic fibrosis, asthma, chronic obstructive pulmonary disease, chronic coughing, lung cancer, and pulmonary hypertension. Kv7 channels are now recognized as playing relevant physiological roles in many tissues, which have encouraged the search for Kv7 channel modulators with potential therapeutic use in many diseases including those affecting the lung. Modulation of Kv7 channels has been proposed to provide beneficial effects in a number of lung conditions. Therefore, Kv7 channel openers/enhancers or drugs acting partly through these channels have been proposed as bronchodilators, expectorants, antitussives, chemotherapeutics and pulmonary vasodilators.

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### LUNG DISEASES: ONE WORLD'S HEALTH CONCERN

Lung diseases constitute a global health concern causing severe physical limitations and premature death, and entails enormous costs associated with treatment, hospital-based and primary care (Tobin, 2005). Mortality from overall lung diseases is very striking according to data collected by the WHO in 2016, in which respiratory diseases account for 24% of world population mortality, the second cause of death after cardiovascular diseases (WHO Global Health Estimates, 2020). At the top of mortality rate within lung diseases, the 9% of the population dies of tuberculosis. Second and third positions include chronic obstructive pulmonary disease (COPD) and lower respiratory infections with 5.4 and 5.2%, respectively. These are followed by lung cancer (affecting trachea, bronchus, and lungs) with 3% and, lastly, asthma and other respiratory

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diseases with a 0.7 and 0.6%, respectively (Figure 1). Most major respiratory diseases are avoidable and can be mitigated by reducing exposure to air pollution, toxic smoke of biomass fuel, tobacco smoke, and different allergens (Laumbach and Kipen, 2012). Additional factors, such as aging, genetic predisposition, nutritional status, obesity, physical inactivity, raised blood pressure, human immunodeficiency virus 1 infection (HIV-1), or other infections may influence the risk of developing lung diseases. Lung diseases can be classified into three main groups depending on the lung tissue involved. First, airways diseases including asthma, COPD, emphysema, cystic fibrosis, or bronchitis in which the bronchial tubes are narrowed or blocked and the transport of oxygen and other gases are affected (Athanazio, 2012). Second, lung tissue diseases which affect the structure of alveoli or the interstitium (Corvol et al., 2009), such as pneumonia, tuberculosis, pulmonary edema, acute respiratory distress syndrome, sarcoidosis, and pulmonary fibrosis, where the lungs are unable to expand fully (Dellaripa, 2018). The third group includes diseases affecting the pulmonary vessels, such as pulmonary hypertension characterized by vasoconstriction, vascular remodeling, inflammation, and thrombosis (Cummings and Bhalla, 2015). This last group of diseases may end up affecting heart function.

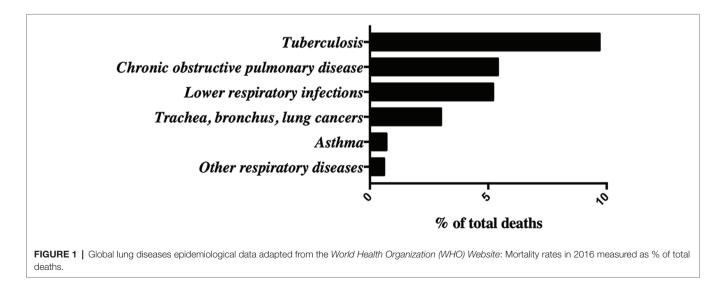
Coronavirus disease 2019 (COVID-19), caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has recently rapidly evolved into a global pandemic. To date, risk factors associated with admission to ICU and mortality include cardiovascular comorbidities while respiratory diseases paradoxically do not appear to increase the prevalence or severity of the viral infection (Halpin et al., 2020; Li et al., 2020).

## Kv7 CHANNEL STRUCTURE AND REGULATION

The Kv7 channel family, is a group of voltage-dependent K<sup>+</sup> channels subfamily Q, which are involved in various physiological activities of numerous cell types, especially, cardiac myocytes,

smooth muscle cells, neurons, and epithelial cells (Stott et al., 2014). The different Kv7 channels isoforms are encoded by KCNQ1-5 genes located at chromosomal loci 11p15, 20q13, 8q24, 1p34, and 6q13, respectively (Barrese et al., 2018). KCNQ genes encode for five Kv7 proteins, Kv7.1-Kv7.5 channels, comprising approximately 650–940 aminoacids. Each pore-forming  $\alpha$ -subunit of Kv7 channels has six helices transmembrane domains (TM). The TM5 and TM6 are part of the pore while the TM4 constitutes the voltage-detection domain. Four  $\alpha$ -subunits form the transmembrane pore assembling into a homo- or heterotetrameric channel (Taylor and Sanders, 2017). Kv7 channels can be regulated by potassium voltage-gated channel subfamily E regulatory subunits (KCNEs) ancillary subunits (KCNE1-5, also called minK-related peptides), which present a single transmembrane domain with 103-177 residues, encoded by the five KCNE1-5 genes located at chromosomal loci 21q11, 21q11, 11q13, 2q36, and Xq23, respectively (Lundquist et al., 2006). These KCNE subunits modulate several features of Kv7 channels such as biophysical properties, cell localization, channel trafficking, and sensitivity to different drugs (Barrese et al., 2018).

Mutations in all five KCNQ and KCNE genes have been shown to underlie excitability hereditary disorders (Jentsch, 2000; Maljevic et al., 2010; Lehman et al., 2017; Barrese et al., 2018). Particularly, mutations affecting KCNQ1 gene cause disorders related to cardiac electrical activity such as long-QT syndrome, short-QT syndrome, and atrial fibrillation leading to severe arrhythmias and sudden death (Li et al., 1998; Chen et al., 2003). KCNQ2 and KCNQ3 gene mutations have been identified in different epilepsy syndromes (Neubauer et al., 2008) and, moreover, KCNQ3 gain-of-function variant have been related to autism and developmental disability (Sands et al., 2019). Kv7.4 channels expressed in the cochlea are essential for normal hearing, and KCNQ4 mutations are linked to hearing loss (Kubisch et al., 1999; Jung et al., 2019). Lastly, mutations in the Kv7.5 channel encoded by KCNQ5 gene cause epilepsy and intellectual disability (Lehman et al., 2017). Different mutations have also been found in all five KCNE genes, mainly related to cardiac arrhythmias (Splawski et al., 2000; Lundby et al., 2008; Ohno et al., 2009; Abbott, 2016).



Different combinations of Kv7 channels and KCNE subunits can be found depending on the cell type. Thus, there is a Kv7 channel isoform-specific expression playing different roles depending on tissue types (see section Kv7 Channel Expression in the Lung). Many KCNE subunits have been shown to modulate Kv7 channels  $\alpha$ -subunits expressed in heterologous systems, providing a valuable experimental platform for elucidating structure-function relationships and mechanistic differences. Thus, all five KCNE proteins have been demonstrated to modulate Kv7.1 channels expressed in heterologous systems with diverse effects (Van Horn et al., 2011; Sun and MacKinnon, 2020). For instance, KCNE1 causes the Kv7.1 channel to activate at more positive voltages, slows its activation and deactivation, increases its conductance, and suppresses its inactivation (Sun and MacKinnon, 2020). While, KCNE2 and KCNE4 have inhibitory effects on Kv7.1 activity (Grunnet et al., 2002; Vanoye et al., 2009; Hu et al., 2019), KCNE3 stabilizes Kv7.1 channels voltage sensors (S4 segment) in an activated state turning the channel voltage-independent (Barro-Soria et al., 2015). Such a mechanism seems to require the participation of the signaling lipid phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>) in non-excitable cells such as lung epithelial cells (Zhou et al., 2019; Sun and MacKinnon, 2020). Sun and MacKinnon have recently identified binding sites of PIP<sub>2</sub> on Kv7.1 channels present in the S0, the S2-S3 loop, and the S4-S5 linker, and proposed a model in which the conformational change following PIP<sub>2</sub> interaction would results in dilation of the pore's gate (Sun and MacKinnon, 2020). In addition, KCNE subunits regulate traffic and cell surface expression of Kv7.1 channels (Roura-Ferrer et al., 2010; Hu et al., 2019). The information about the regulation of Kv7.2 and Kv7.3 channels by KCNE subunits is more limited. In Xenopus oocytes, KCNE1 was found to slow Kv7.2/7.3 heteromer currents and decreased their magnitude (Yang et al., 1998). Likewise, KCNE2 was shown to accelerate the deactivation kinetics of Kv7.2 and Kv7.2/7.3 channels transiently expressed in COS cells (Tinel et al., 2000). Whereas, KCNE1, KCNE2, and especially KCNE4 and KCNE5 increased Kv7.4 currents, co-expression with KCNE3 led to a strong attenuation (Strutz-Seebohm et al., 2006; de Jong and Jepps, 2018). Moreover, KCNE4 was shown to co-localize with Kv7.4 channels, regulating its activity and membrane abundance in mesenteric arterial myocytes (Jepps et al., 2015). Finally, Kv7.5 channels were shown to be affected by KCNE1 and KCNE3, but not by other KCNE subunits in Xenopus oocytes and HEK-293 cells (Roura-Ferrer et al., 2009, 2012). Thus, while KCNE1 was shown to slow activation and increase Kv7.5 currents, KCNE3 drastically inhibited these currents (Roura-Ferrer et al., 2009, 2012).

The activity of Kv7 channels is modulated by G proteincoupled membrane receptors (GPCRs). Thus, activation of different GPCR have been shown to inhibit M-type (Kv7) currents, with a dominant participation of the  $G_q$  and  $G_{11}$  $\alpha$ -subunits (Caulfield et al., 1994; Haley et al., 1998; Delmas and Brown, 2005). For instance, in neurons, Kv7 activity is decreased by  $M_1$  and  $M_3$  muscarinic and  $H_1$  histaminic receptors coupled to  $G_{q/11}$  proteins through different signal transduction pathways, including depletion of PIP<sub>2</sub>, activation of protein kinase C (PKC), or increased intracellular calcium (Delmas and Brown, 2005; Higashida et al., 2005; Taylor and Sanders, 2017; Barrese et al., 2018). In human airway smooth muscle cells, histamine was found to inhibit Kv7.5 channels though a PKC-dependent phosphorylation of Ser441 (Haick et al., 2017). On the other hand, it is well recognized that stimulation of  $\beta$ -adrenergic receptor increases the activity of different Kv7 channels in different tissues such as cardiac (Kv7.1), neuronal (Kv7.2/Kv7.3), and vascular (Kv7.4/Kv7.5 and Kv7.5) channels (Jentsch, 2000; Marx et al., 2002; Chadha et al., 2014; Mani et al., 2016; Barrese et al., 2018). Likewise, in human airway smooth muscle cells, Kv7.5 channels are robustly enhanced by activation of  $\beta$ -adrenoceptors via a mechanism involving cyclic adenosine monophosphate/protein kinase A (cAMP/PKA)-dependent phosphorylation (Brueggemann et al., 2018). This effect might be shared with other GPCRs but does not seem to account for Kv7.4 activation. The mechanism may involve increased membrane expression of Kv7 channels mediated by colchicine-sensitive microtubule disruption (Lindman et al., 2018). Kv7 channels are also regulated by GPCR-independent mediators activating soluble guanylate cyclase (sGC) such as nitric oxide (NO) (Mondejar-Parreño et al., 2019) and natriuretic peptide (Stott et al., 2015). The implications in respiratory diseases and drug therapy are discussed below.

In addition to regulation by KCNE  $\beta$ -subunits and GPCRs, Kv7 channels can be regulated by scaffolding proteins, sodiumcoupled myo-inositol transporter (SMIT), lipids, microRNAs, and/or further signaling mechanisms as extensively reviewed elsewhere (Delmas and Brown, 2005; Zaydman and Cui, 2014; Taylor and Sanders, 2017; Barrese et al., 2018; Byron and Brueggemann, 2018).

# Kv7 CHANNEL EXPRESSION IN THE LUNG

Kv7 channels are expressed mainly in pulmonary arteries (PAs) and in the bronchial tubes (**Figure 2**). In PA, Kv7 channels are expressed in pulmonary artery smooth muscle cells (PASMCs); while in the bronchial tubes (trachea, bronchus and bronchioles), Kv7 channels are expressed in airway smooth muscle cells (ASMCs), airway epithelial cells (AEPs) and vagal airway C-fibers (VACFs). The functional role of Kv7 channels may vary depending on the cell type. While most of the available studies have provided relevant information on the particular expression of α-subunits in these cell types, evidence for the expression of heteromeric Kv7 complexes or KCNE subunits is still very scarce.

#### **Pulmonary Artery Smooth Muscle Cells**

The expression of Kv7 channels in mice and rat PA has been demonstrated in several studies. Specifically, their expression is located in PASMC. In these cells, the expression of Kv7.1, Kv7.4, and Kv7.5 channels predominates; but Kv7.4 channels expression appears to be the most relevant. The expression of Kv7.2 and Kv7.3 channels in PASMCs is still controversial (Joshi et al., 2009; Morecroft et al., 2009; Chadha et al., 2012; Li et al., 2014a; Sedivy et al., 2015; Eid and Gurney, 2018; Mondejar-Parreño et al., 2018). The existence of Kv7.1/7.5 or Kv7.4/7.5 heteromeric

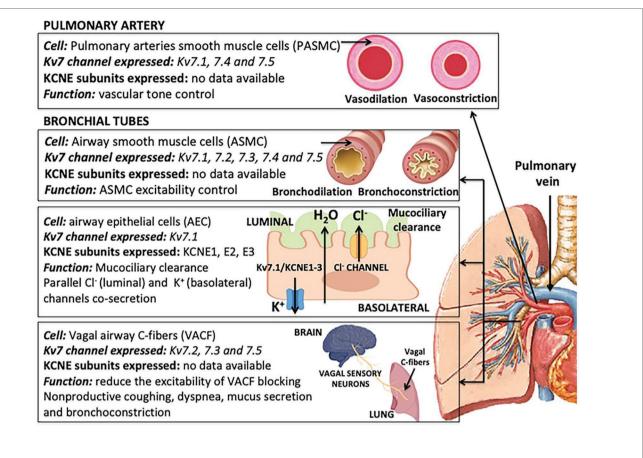


FIGURE 2 | Expression and functions of isoform-specific Kv7 channel and potassium voltage-gated channel subfamily E regulatory subunits (KCNE) ancillary subunit in lung cells.

complexes in vascular smooth muscle has been postulated on several studies (Chadha et al., 2014; Oliveras et al., 2014; Barrese et al., 2018). In particular, Kv7.4/7.5 heteromers have been proposed as the dominant functional vascular Kv7 channels, supported by electrophysiological data showing that Kv7 currents in freshly isolated vascular smooth muscle cells have phenotypes intermediate to the respective homomeric Kv7.4 and Kv7.5 channels (Brueggemann et al., 2014a; Chadha et al., 2014; Fosmo and Skraastad, 2017; Barrese et al., 2018). We have recently confirmed that Kv7.4/7.5 heteromeric complexes are also present in PASMC (authors' unpublished observations). Although, in other types of arteries, the expression of Kv7 channels has been located also in endothelial cells (Goodwill et al., 2016), whether this is also the case for pulmonary artery endothelial cells (PAECs) remains unknown. The main function of these channels is to regulate pulmonary vascular tone, due to their contribution to maintain the membrane potential (Em) of PASMC (Joshi et al., 2009; Sedivy et al., 2015; Mondejar-Parreño et al., 2018, 2019).

#### **Airway Smooth Muscle Cells**

There is some interspecies variability in the expression of Kv7 channel isoforms in the airways. Thus, ASMCs from guinea pigs were shown to express Kv7 channels with a relative abundance of KCNQ2 > KCNQ5 > KCNQ4 > KCNQ3 and, to a lesser

extent, KCNQ1 (Brueggemann et al., 2011). Comparatively, KCNQ mRNA expression in ASMC revealed a high expression of KCNQ4 followed by a fainter expression of KCNQ5 and KCNQ1 transcripts (Brueggemann et al., 2014b). Studies in human tracheal muscle revealed abundant expression of KCNQ1 relative to guinea pig, a modest expression of KCNQ4 and KCNQ5 and no detectable expression of KCNQ2 or KCNQ3 (Brueggemann et al., 2011). A further study showed that cultured human ASMCs express all KCNQ genes with a predominant relative abundance of KCNQ5 mRNA followed by KCNQ3 or KCNQ4, while KCNQ2 and KCNQ1 mRNA are expressed to a lesser extent (Brueggemann et al., 2018). These differences in the expression profile of Kv7 channels in human tissue could be due to phenotypic changes associated with cell culture. Measurements of Kv7 currents and functional studies assessing airway smooth muscle contractility in the bronchioles from different species have pointed to Kv7 channels as important players in maintaining, resting Em, and controlling ASMC excitability (Evseev et al., 2013; Brueggemann et al., 2014b, 2018).

#### **Airway Epithelial Cells**

Airway epithelial cells (AECs) predominates the expression of Kv7.1 channels, also named KvLQT1, along with the expression of KCNE1 and KCNE3 ancillary subunits (Mall et al., 2000;

Boucherot et al., 2001; Trinh et al., 2007; Preston et al., 2010; Frizzell and Hanrahan, 2012; Kroncke et al., 2016; Hynes and Harvey, 2019). Nevertheless, KCNE2 has also been found to be expressed in *Calu-3* cells, a model for human submucosal serous cells of the lung, where it has been suggested to regulate *KCNQ1* (Cowley and Linsdell, 2002a). Chloride (Cl<sup>-</sup>) secretion in AEC is essential to mucociliary clearance and requires the co-activation of Cl<sup>-</sup> channels in luminal membrane and, in parallel, of K<sup>+</sup> channels in the basolateral membrane which involves Kv7.1 channels (Frizzell and Hanrahan, 2012).

#### Vagal Airway C-Fibers

Using single-cell mRNA, Sun et al. have recently demonstrated that *KCNQ2*, *KCNQ3*, and *KCNQ5* are consistently expressed in mouse airway sensory afferent nerves and remarkably, the expression of *KCNQ3* mRNA was the most prevalent (Sun et al., 2019). Most vagal sensory nerves in the respiratory tract are nociceptive C-fibers. Increased VACF activity may contribute to the nonproductive coughing, dyspnea, parasympathetic reflex mucus secretion, and bronchoconstriction. Thus, it has been proposed that activation of Kv7 channels may reduce the vagal C-fibers excitability innervating the airways, and being useful to treat chronic cough.

### Kv7 CHANNELS IN LUNG DISEASES

This section covers the potential pathophysiological role of Kv7 channels in different respiratory diseases such as cystic fibrosis, asthma, COPD, chronic coughing, lung cancer, and pulmonary hypertension.

#### **Cystic Fibrosis**

The airway surface liquid layer represents a key primary defense mechanism of mucociliary clearance, which crucially depends on the transport of chloride into the lumen of the airways, and the subsequent movement of water. Cl- ion transport across the apical membrane is mainly driven by a cAMP-regulated Cl<sup>-</sup> channel named cystic fibrosis transmembrane conductance regulator (CFTR) and by the calcium-activated Cl- channels (CaCC) (Frizzell and Hanrahan, 2012; Bartoszewski et al., 2017). However, a possible overlap between these two pathways has also been proposed (Billet and Hanrahan, 2013). Cystic fibrosis is an autosomal recessive disease due to mutations in the gene encoding CFTR and characterized by alterations in the composition and volume of secreted luminal fluids. The CFTR is a member of the ATP-binding cassette (ABC) transporter superfamily known to function as an ion cannel that conducts chloride ions across epithelial cell membranes and is responsible for salt secretion in response to cAMP/PKA stimulation. Thus, mutations of the CFTR gene lead to defective cAMP-dependent Cl- secretion and enhanced Na+ absorption throughout the apical membrane of airway epithelia.

CFTR has been shown to influence both function and properties of other membrane channels, such as epithelial Na<sup>+</sup> channels, inward rectifying renal outer medullary K<sup>+</sup> channel (ROMK), outwardly rectifying Cl<sup>-</sup> channels, and aquaporin 3 (Schwiebert

et al., 1999). Cl<sup>-</sup> secretion in AEC requires the parallel co-activation of Cl- channels and K+ channels, in the luminal surface and the basolateral membrane, respectively. Whereas Cl- channels are required for Cl<sup>-</sup> efflux to the luminal side of the AEC, K<sup>+</sup> channels are needed for K<sup>+</sup> recycling *via* the basolateral membrane (Boucherot et al., 2001; Bartoszewski et al., 2017). Kv7.1 channels have been shown to contribute to the basolateral cAMP-activated K<sup>+</sup> conductance in human airway epithelium (Mall et al., 1996, 2000; Bardou et al., 2009). This study also showed that the Kv7.1 channel inhibitor chromanol 293B inhibited Cl- secretion suggesting an important role of these K<sup>+</sup> channels in maintaining cAMP-dependent Cl<sup>-</sup> secretion in human airways. Since CFTR is a modulator of other ion channels, several studies have analyzed whether it controls the activity of Kv7.1 channels. Studies analyzing this issue have led to contradictory results. Thus, while initial studies suggested that CFTR could directly modulate Kv7.1 conductance (Cunningham et al., 1992; Loussouarn et al., 1996; Mall et al., 2000), this was not the case in other studies (Boucherot et al., 2001). Interestingly, using CFTR KO mice, Belfodil et al. (2003) found that cAMP-sensitive Kv7.1 currents were regulated by CFTR in renal distal but not in proximal tubule cells, suggesting that the possible modulation of Kv7.1 is unlikely to involve a direct interaction.

As stated above, human airway epithelia also express CaCC that are activated by extracellular nucleotides [adenosine triphosphate (ATP) and/or uracil triphosphate (UTP)] and other Ca2+-increasing agonists. CaCC was shown to be preserved or enhanced in human and murine cystic fibrosis airways as a mechanism to compensate for the lack of CFTR (Clarke et al., 1994; Grubb et al., 1994). Since, CaCC-mediated Clsecretion appears to be preserved in cystic fibrosis its pharmacological stimulation has been suggested as a therapeutic tool for bypassing dysfunctional CFTR. Mall et al. demonstrated that activation of the cAMP pathway increased CaCC-mediated secretion in cystic fibrosis and, conversely, chromanol 293B, an inhibitor of cAMP-activated Kv7.1 channels was able to fully abolish nucleotide-activated Cl- secretion (Mall et al., 2003). Hence, the authors proposed that activation of Kv7.1 channels, together with hSK4 K<sup>+</sup> channels, may improve Cl<sup>-</sup> secretion via CaCC in human cystic fibrosis airway tissues (Mall et al., 2003). Altogether, these studies indicate the Kv7.1 channel activation may help to improve mucociliary clearance in cystic fibrosis airways.

Elevated levels of reactive oxygen species (ROS) lead to oxidative stress and DNA damage in several tissues providing a common pathological mechanism in all inflammatory lung diseases. Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) has been proven to stimulate several components of the Cl<sup>-</sup> secretion in the airway epithelia, including the cAMP-dependent activation of CFTR and the basolateral K<sup>+</sup> channels (Cowley and Linsdell, 2002b; Ivonnet et al., 2015). The basolateral K<sup>+</sup> conductance was abrogated by clofilium, a Kv7.1 channel inhibitor, suggesting a role for these channels in the H<sub>2</sub>O<sub>2</sub>-stimulated response (Cowley and Linsdell, 2002b; Ivonnet et al., 2015). In line with this, H<sub>2</sub>O<sub>2</sub> has been reported to potently enhance Kv7 currents (Gamper et al., 2006; Kim et al., 2016). Activation of transmembrane adenylyl cyclase by H<sub>2</sub>O<sub>2</sub> has been proposed as a triggering mechanism for the cAMP-activated basolateral Kv7.1 current (Ivonnet et al., 2015). The stimulated anion secretion by ROS could act as a compensatory protective mechanism to keep the airways clear from damaging radicals and to hold acceptable airway surface liquid levels.

While the Kv7.1 channel is the member of the KCNQ family with the strongest evidence of its functional expression and physiological role in lung epithelial cells, others (KCNQ3 and KCNQ5) have been detected in the apical or lateral membranes (Moser et al., 2008). In addition, mRNA transcripts for several KCNE accessory subunits (KCNE2 and KCNE3) have been found in human airway Calu-3 cells (Cowley and Linsdell, 2002a). Kv7.1 channels expressed in Xenopus oocytes were shown to be activated by an increase in intracellular cAMP concentration, independently of co-expression with KCNE1 or KCNE3 ancillary subunits (Boucherot et al., 2001). On the other hand, Potet et al. demonstrated that recombinant Kv7.1/KCNE1 complexes expressed in Cos-7 cells were totally insensitive to cAMP (Potet et al., 2001). The co-assembly of Kv7.1 with KCNE3 subunits markedly affects Kv7.1 channel properties (abolishing its voltage dependence) and has been proposed to form the cAMP-activated basolateral K<sup>+</sup> channels contributing to Cl<sup>-</sup> secretion in colonic epithelial cells (Schroeder et al., 2000). Interestingly, Preston et al. evidenced that cAMP-stimulated Cl<sup>-</sup> secretion across tracheal epithelia was severely reduced in KCNE3<sup>(-/-)</sup> mice (Preston et al., 2010). While the involvement of KCNE3 in airway secretion requires confirmation in future studies, the structural basis of the interaction of Kv7.1 with KCNE3 leading to a constitutive activation of Kv7.1 channel activity has been recently established (Kroncke et al., 2016; Sun and MacKinnon, 2020).

#### Asthma

Asthma is characterized by hyper-contractility of bronchial smooth muscle attributed to an increased concentration of different bronchoconstrictor agonists, or enhanced sensitivity to them, leading to aberrant narrowing of the airways (Penn, 2008). Acetylcholine, leukotriene D4, endothelin, and histamine represent the most relevant locally released bronchoconstrictor agonists that activate GPCR on ASMC, leading to increased cytosolic Ca<sup>2+</sup> concentration and contraction (Jude et al., 2008; Penn, 2008). This together with the strong inflammation leads to an acute hypersensitivity to different air stimulants that produce ASMC bronchospasm and that, in the long term, promote smooth muscle proliferation and hypertrophy (Stott et al., 2014). Kv7 channels are considered negative regulators of ASMC contraction due to their major contribution to maintain negative Em negative, preventing L-type-Ca<sup>2+</sup> channel activation and the subsequent bronchoconstriction (Brueggemann et al., 2011). Several members of the Kv7 family are expressed in ASMCs (Brueggemann et al., 2011; Bartoszewski et al., 2017). The relative abundance expression of the Kv7 members appears to differ depending on the location (trachea vs. bronchioles) and depending on the animal species as mentioned above (Brueggemann et al., 2011, 2014b; Evseev et al., 2013). Several studies have evidenced that bronchoconstrictor agonists negatively regulate the activity of Kv7 channels. Thus, methacholine, carbachol, or histamine reduce Kv7 currents in guinea pig, mouse, and rat ASMCs (Brueggemann et al., 2011, 2014; Haick et al., 2017). On the other hand, structurally unrelated Kv7 channel activators cause a concentration-dependent relaxation of small bronchioles (Brueggemann et al., 2014b) and tracheal rings (Evseev et al., 2013). Of note, *in vivo* administration of the Kv7 channel opener retigabine was proven to transiently reduce bronchoconstriction induced by methacholine in mice (Evseev et al., 2013).

Chronic administration of long-acting  $\beta_2$ -adrenergic receptor agonist elicits a time- and concentration-dependent relaxation of rat airways, with remarkable desensitization to short-term or with repeated treatments. Co-administration of retigabine plus the long-acting  $\beta_2$ -adrenergic receptor agonist formoterol was shown to cause a sustained reduction of methacholine-induced bronchoconstriction and reduced the desensitization observed with formoterol alone (Brueggemann et al., 2014b). These findings suggest that using a Kv7 channel enhancer together a  $\beta_2$ -adrenergic receptor agonist may improve bronchodilator therapy.

In addition to exaggerated bronchoconstriction, asthma is characterized by a chronic inflammatory response in the airways that is targeted primarily with inhaled corticosteroids. As stated in the previous section basolateral membrane K<sup>+</sup> channels play a key role in stimulating Cl<sup>-</sup> transport across AEPs (Frizzell and Hanrahan, 2012). Unlike in cystic fibrosis, a decrease in airway mucus secretion ameliorates the symptomatology of asthma. Very recently, Hynes and Harvey have demonstrated that dexamethasone produces an inhibition of transepithelial chloride ion secretion though a rapid non-genomic inhibition of Kv7.1 channels and KCNN4 (K<sub>Ca</sub>3.1) (Hynes and Harvey, 2019). Thus, it is plausible that down-regulation of Kv7.1 channel activity might contribute to the potential benefit of corticosteroids to reverse airway hypersecretion.

## Chronic Obstructive Pulmonary Disease (COPD)

COPD is a progressive life-threatening lung disease characterized by airway obstruction due to inflammation of the small airways. The disease evolves with irreversible limitation of airflow, loss of lung function and severe acute exacerbations which significantly deteriorate the patient's quality of life (Barnes et al., 2003).  $\beta$ -adrenergic receptors ( $\beta$ -AR) agonists promote airway smooth muscle relaxation and are the drugs of choice for rescue from acute bronchoconstriction in asthma and COPD. A recent study has shown that the long-acting  $\beta$ -AR formoterol robustly enhanced Kv7 currents in human ASMCs (Brueggemann et al., 2018). Furthermore, these authors provided evidence that Kv7.5 is the predominant Kv7 α-subunit in human ASMCs and identified a PKA-dependent phosphorylation of S53 on the N-terminus of Kv7.5 channel subunits as a potential mechanism of activation following the stimulation of the  $\beta AR/$ cAMP pathway (Brueggemann et al., 2018).

The exposure to cigarette smoking is the principal cause of COPD (Salvi, 2014). Recently Sevilla-Montero et al. have analyzed the impact of cigarette smoke exposure on the pulmonary vasculature (Sevilla-Montero et al., 2019). Authors showed that cigarette smoke extract exposure had direct effects on human fibroblasts and PASMCs, promoting a senescent phenotype which contributed to the secretion of inflammatory molecules and increase the proliferation of non-exposed cells. Furthermore, cigarette smoke extract exposure affected cell contractility attenuated the pulmonary vascular responses to Kv7 channel modulators and dysregulated the expression and activity of Kv7.4 channels. The results suggest that cigarette smoke may impair pulmonary vascular tone atleast in part through Kv7.4 downregulation (Sevilla-Montero et al., 2019).

A sequence polymorphism of the human *KCNE2* gene has been found to be associated with altered lung function (Soler-Artigas et al., 2011; Sabater-Lleal et al., 2014). The deletion of *KCNE2* reduced pulmonary expression of the Kv7.1  $\alpha$ -subunit suggesting the formation of pulmonary KCNQ1-KCNE2 complexes (Zhou et al., 2019). *KCNE2*<sup>(-/-)</sup> mice had reduced blood O<sub>2</sub>, increased CO<sub>2</sub>, increased pulmonary apoptosis, and increased inflammatory mediators TNF- $\alpha$ , IL-6, and leukocytes in bronchoalveolar lavage fluids (Zhou et al., 2019). These data indicated KCNE2 regulates *KCNQ1* in the lungs and is required for normal lung function.

#### Chronic Cough

Airway sensory afferent nerves are essential for triggering different reflex responses to potential injurious stimuli (Undem and Kollarik, 2005). Most vagal sensory afferent nerves which terminate in the respiratory tract are C-fibers (Undem and Kollarik, 2005). The excessive activity of VACF induces different symptoms associated with inflammatory lung diseases, such as nonproductive coughing, dyspnea, parasympathetic reflex mucus secretion and bronchoconstriction (Mazzone and Undem, 2016). Consequently, ion channels that regulate the excitability of C-fibers in the airways can be an attractive therapeutic target aimed at reducing the symptoms of various inflammatory diseases of the airways. Using single-cell mRNA analysis, Sun and collaborators have very recently shown that KCNQ2, KCNQ3, and KCNQ5 are consistently expressed in mouse lung specific nodose neurons with KCNQ3 being the most predominant (Sun et al., 2019). This study suggested that Kv7.3 channels are the major contributors to the Kv7-M current in the nodal neurons of the mouse lungs. The Kv7 channel activator retigabine increased the amplitude and accelerated the activation rate of the M-current, caused a prominent hyperpolarization, and suppressed the excitability of mouse nodose neurons. These effects were reversed or prevented in the presence of XE991, a Kv7 channel inhibitor. Remarkably, nebulized retigabine was found to attenuate cough evoked by inhalation of irritant gases (SO<sub>2</sub> and NH<sub>3</sub>) in awake mice (Sun et al., 2019). Although future studies are needed to confirm their role in the regulation of C-fiber excitability of the airways in humans, these data indicate that Kv7.3 channels may provide a novel therapeutic target to reduce the conditions associated with pulmonary inflammatory diseases such as chronic coughing.

#### Non-small Cell Lung Cancer

K<sup>+</sup> channels regulate a number of processes of key relevance in cancer biology such as cell volume control, pH regulation, migration, proliferation, differentiation, apoptosis, and drug

resistance (Rao et al., 2015; Prevarskaya et al., 2018; Haworth and Brackenbury, 2019; Serrano-Novillo et al., 2019). Thus, there is increasing evidence that K<sup>+</sup> channels play an important role in cancer; but a clear scenario is still far due to the plethora of K<sup>+</sup> channel isoforms and their diverse repertoire of effects. Notably, K<sup>+</sup> channels regulate cell cycle, but according to the channel isoform, its level of expression, the cell type, and the stage of differentiation the effects are diverse. For instance, Kv channels may exhibit proliferative or anti-proliferative roles, depending on the channel subtype and the phase of the cell cycle (Rao et al., 2015). Moreover changes in the phenotypes, signaling pathways, and pattern of expression of K<sup>+</sup> channels may be observed in cancer, which increases the complexity of K<sup>+</sup> channels regulation on cell viability. In addition to the well-characterized involvement of several Kv channels (most notably Kv1.1, Kv1.3, Kv1.5, Kv10.1, Kv10.2, and Kv11.1) in cancer (Rao et al., 2015), some members of the Kv7 family have also been suggested to participate in cell proliferation and cancer (Serrano-Novillo et al., 2019).

K<sup>+</sup> channels participate in tumor cell invasion and metastasis propagation via regulation of cell migration and growth (O'Grady and Lee, 2005; Schwab et al., 2008; Arcangeli et al., 2009; Prevarskaya et al., 2010). Indeed, inhibiting or silencing K<sup>+</sup> channels reduces the proliferation of cancer cells (Pancrazio et al., 1993; Jang et al., 2011). Kv7.1 channels have been highlighted in cell migration, proliferation, and repair of AECs (Trinh et al., 2007). In 2014, Girault and colleagues showed an increased expression (by 1.5- to 7-fold) of Kv7.1 channels in tumor lung adenocarcinoma samples, compared to non-neoplastic tissues from 17 of 26 patients (Girault et al., 2014). Authors found that the proliferation rates of lung adenocarcinoma cell monolayers were reduced by the Kv7.1 channel blockers clofilium and chromanol, or after silencing Kv7.1 channels using a specific siRNA. These data suggest that Kv7.1 channels could be a potential therapeutic target in lung cancer (Girault et al., 2014).

On the other hand, K<sup>+</sup> channels have also been implicated in both early and late stages of apoptosis. The decrease in cell volume is one of the earliest and necessary morphological changes observed in cells undergoing apoptosis. Active K<sup>+</sup> channels in the plasma membrane leads to K<sup>+</sup> and Cl<sup>-</sup> outflow through K<sup>+</sup> and Cl<sup>-</sup> channels, this generates an osmotic gradient leading to an outward water transport through aquaporins which results in a decrease in cell volume (Maeno et al., 2000). In later stages of apoptosis, the augmented intracellular K<sup>+</sup> concentration ([K<sup>+</sup>]<sub>int</sub>) suppresses caspase and nuclease activity and, conversely, the decreased [K<sup>+</sup>]<sub>int</sub> caused by K<sup>+</sup> channel activation, promotes apoptosis (Burg et al., 2008). There is a wealth of literature supporting that the M- channel activators have a neuroprotective role via Kv7 activation and reduction of hyperactivity and, hence, suppression of glutamate excitotoxicity (Gamper et al., 2006; Bierbower et al., 2015; Vigil et al., 2020). It is likely that flupirtine would work the same way (Panchanathan et al., 2013; Li et al., 2014a). Nevertherless, its ability to inhibit cell viability has also been reported. Thus, Lee et al. showed that flupirtine exerted anti-proliferative effects in canine osteosarcoma cells by enhancing Kv7.5 function and arresting cells in the  $G_0/G_1$  phases (Lee et al., 2014).

Similarly, retigabine was found to exhibit anti-proliferative effects in *C2C12* myoblasts (Iannotti et al., 2010).

Non-small-cell lung cancer (NSCLC) accounts for most cancer deaths. Aberrant activation of the epidermal growth factor receptor (EGFR) is involved in the pathogenesis and progression of several human cancers, including NSCLC (Melosky, 2014). Indeed, approximately 10-38% of NSCLC patients have EGFR gene mutations (Lin et al., 2014). EGFR tyrosine kinase inhibitors, such as Gefitinib have become standard first-line treatment for these patients (Cohen et al., 2003). However, acquired resistance represents a major limitation for drug efficacy. Cancer stem cells have self-renewal properties in various solid tumors, and play a key role in tumor growth and progression (Wang et al., 2013) and in therapy resistance (Steinbichler et al., 2018). A subset of cancer stem cells, termed the "side population" (SP), have elevated clonogenic potential and higher expression levels of ABC-transporters than main-population cells known as non-SP cells (Richard et al., 2013) and are considered to be responsible for anti-cancer drug resistance (Singh et al., 2010; Leon et al., 2016). Choi and colleagues have demonstrated a reduced mRNA expression of KCNQ3 and KCNQ5 in SP cells compared to non-SP cells (Choi et al., 2017). The treatment of flupirtine (a Kv7.2-7.5 channel enhancer) decreased the viability of SP and non-SP cells. In addition, the combination treatment of gefitinib plus flupirtine was more effective, compared to the individual treatment of both drugs, in decreasing the viability of both non-SP cells, and gefitinib-resistant SP cells. According to their results, the combination treatment of gefitinib plus flupirtine was proposed to reduce the viability of gefitinib-resistant cells through inhibition of the EGFR-Ras-Raf-ERK pathway via (Choi et al., 2017). These results suggest that the activation of Kv7.3 and Kv7.5 channels could play an important role in the induction of apoptosis in SP cells, either by participating in the decrease of the cellular volume necessary for apoptosis initiation; or by decreasing [K<sup>+</sup>]<sub>int</sub>, which results in the activation of caspases and nucleases.

#### **Pulmonary Hypertension**

Pulmonary hypertension is defined by an increase in mean pulmonary arterial pressure at rest, as assessed by right heart catheterization. Based on pathophysiological findings, clinical presentation, hemodynamic characteristics, and therapeutic considerations, pulmonary hypertension is categorized into five groups by WHO: pulmonary arterial hypertension (PAH), pulmonary hypertension due to left-sided heart disease, pulmonary hypertension due to lung disease or hypoxia, chronic thromboembolic pulmonary hypertension, and multifactorial pulmonary hypertension (Simonneau et al., 2019). PAH is characterized by excessive pulmonary vasoconstriction and progressive remodeling of the distal PAs, resulting in elevated pulmonary vascular resistance and, eventually, in right ventricular failure (McLaughlin et al., 2015). Reduced expression or function of K<sup>+</sup> channels (notably Kv1.5 and TASK-1 channels) in PASMCs results in a more depolarized Em, and is considered to contribute to the enhanced vasoconstriction and proliferation (Moudgil et al., 2006; Boucherat et al., 2015; Hayabuchi, 2017). In 2009, Joshi et al. demonstrated that Kv7 channel blockers, linopirdine, and XE991, inhibited the  $I_{\mbox{\scriptsize KN}}$  current in PASMCs and raised mean pulmonary arterial pressure (mPAP); while the Kv7 channel openers, retigabine and flupirtine, had the opposite effects (Joshi et al., 2009). Linopirdine can also increase mesenteric vascular resistance and systemic arterial pressure (Mackie et al., 2008). Functional expression of Kv7 channels in the pulmonary circulation was later confirmed by others (Morecroft et al., 2009; Eid and Gurney, 2018; Mondejar-Parreño et al., 2018). As in the systemic circulation, KCNQ1 and especially KCNQ4 and KCNQ5 appear the most predominant KCNQ channels in the pulmonary circulation (Joshi et al., 2009; Morales-Cano et al., 2014; Sedivy et al., 2015; Mondejar-Parreño et al., 2018). The lack of selective drugs for the different Kv7 subunits hinders identification of the functional impact of individual Kv7 members (Barrese et al., 2018). Structurally related Kv7 enhancers such as retigabine and flupirtine (which activate all Kv7 isoforms except Kv7.1 channels) (Barrese et al., 2018) and zinc pyrithione (which activate all Kv7 members but Kv7.3) have been shown to induce pulmonary vasodilation (Joshi et al., 2009; Morecroft et al., 2009; Sedivy et al., 2015; Eid and Gurney, 2018; Mondejar-Parreño et al., 2018). In addition, drugs that block all Kv7 channels (i.e., linopirdine and XE991) are able to contract PAs previously primed (Chadha et al., 2012; Sedivy et al., 2015) or not (Joshi et al., 2009), whereas selective Kv7.1 blockers HMR1556, L768, L673, and JNJ39490282 (Barrese et al., 2018) have no contractile effects in PAs (Chadha et al., 2012). On the other hand, the application of R-L3, a compound purported to activate Kv7.1 channels, relaxed efficiently precontracted intrapulmonary arteries (Chadha et al., 2012; Mondejar-Parreño et al., 2018). These data suggest that even when Kv7.4 and Kv7.5 appear to be the most relevant Kv7 channels in controlling resting Em in PASMC, Kv7.1 channels are also functionally expressed.

The NO/cGMP pathway represents a key physiological signaling controlling tone in PAs, and drugs stimulating this pathway are used to treat PAH (Barnes and Liu, 1995), and drugs activating this pathway are used to treat pulmonary arterial hypertension. Recently, we have found that NO donors and riociguat, a stimulator of sGC used for the treatment of pulmonary arterial hypertension, enhance Kv7 current and induce membrane hyperpolarization in fresh isolated PASMCs contributing to pulmonary vasodilation (Mondejar-Parreño et al., 2019). Further, we identified Kv7.5 channels as a potential member of the Kv7 family targeted by these drugs.

The expression of Kv7 channels has been explored in several experimental models of pulmonary hypertension. While the hypoxic pulmonary vasoconstriction (HPV) reflects a physiological intrinsic property of PASMCs, which allows shifting blood flow from hypoxic to normoxic lung areas, coupling ventilation and perfusion, the chronic exposure to hypoxic air causes the pulmonary circulation to become hypertensive (Sylvester et al., 2012). Sedivy et al. found a significant loss of *KCNQ4* mRNA expression in PA from rats maintained for 3 days in a hypoxic environment, even when a significant reduction of the Kv7.4 protein expression could not be confirmed (Sedivy et al., 2015). On the other hand, the expression of *KCNQ1* and *KCNQ5* was unaffected. However, Li et al. reported that longer incubation on hypoxia (9 days) was associated with a downregulation of

*KCNQ5* in the PAs (Li et al., 2014b). These authors also demonstrated that microRNA-190 (miR-190), whose expression is increased following the hypoxic exposure, directly targeted *KCNQ5*. This led the authors to propose the increased in miR-190 as a mechanism involved in the *KCNQ5* downregulation and enhanced vasoconstriction induced by hypoxia. In addition to the hypoxic pulmonary hypertension models, a slight reduction of *KCNQ5* expression has also been observed in rat lungs from monocrotaline-induced pulmonary hypertension (Morales-Cano et al., 2014). On the other hand, we found the expression of *KCNQ1*, *KCNQ4*, and *KCNQ5* mRNAs unaltered in lungs from Zucker rats, a model of type 2 diabetes, developing increased pulmonary arterial pressure, and right ventricular hypertrophy (Morales-Cano et al., 2019).

The potential impairment of *KCNQ* channels has been also explored in conditions associated with pulmonary hypertension. The HIV-1 infection is an established risk factor for PAH, although the pathological mechanisms underlying the development of HIV-related PAH remain unclear (Butrous, 2015). In a recent study using HIV-transgenic mice expressing seven of the nine HIV viral proteins and wild-type mice we observed that HIV mice had reduced lung expression of *KCNQ1* and *KCNQ5* channels (Mondejar-Parreño et al., 2018). By using vascular reactivity and patch-clamp experimental approaches we found decreased responses to the Kv7.1 channel activator L-364,373 but unaltered responses to retigabine (Mondejar-Parreño et al., 2018). These results suggest that Kv7 channels (especially Kv7.1) are impaired in mice expressing HIV-1 proteins.

Congenital diaphragmatic hernia is a rare congenital anomaly commonly associated with persistent pulmonary hypertension. A recent study showed that the expression of *KCNQ5*, but not that of *KCNQ1* or *KCNQ4*, was significantly downregulated at the mRNA and protein levels in rat lungs from nitrofen-induced congenital diaphragmatic hernia (Zimmer et al., 2017). Likewise, a markedly diminished *KCNQ5* expression was found in the pulmonary vasculature of congenital diaphragmatic hernia fetuses. Since Kv7.5 channels have shown to contribute to NO donorsinduced pulmonary vasodilation (Mondejar-Parreño et al., 2019), its reduced availability in congenital diaphragmatic hernia could partly explain the poor therapeutic response to inhaled NO in these patients (Group (NINOS), 1997; Putnam et al., 2016).

Studies in experimental models have shown evidence that drugs activating Kv7 channels may be of benefit in the treatment of pulmonary hypertension with different etiologies (Morecroft et al., 2009; Sedivy et al., 2015). Morecroft et al. also demonstrated that flupirtine attenuated the development of chronic hypoxiainduced pulmonary hypertension in mice and reversed established pulmonary hypertension in mice that over-express the 5-HT transporter (SERT), apparently *via* Kv7 activation (Morecroft et al., 2009). In this study authors showed that the chronic treatment with flupirtine reduced mean right ventricle pressure (mRVP), pulmonary remodeling, and RV hypertrophy (RVH). More recently, Sedivy et al. confirmed the potential benefit of Kv7 channel openers showing the pulmonary antihypertensive effects of flupirtine in rats exposed to short-term (3–5 days) hypoxia (Sedivy et al., 2015). In line with this, we have found that retigabine-induced vasodilation is increased in PAs from pulmonary hypertensive animals and this is associated with an increased contribution of Kv7 channels to the net K<sup>+</sup> current present in pulmonary arterial myocytes (unpublished data from the author's laboratory).

Altogether, these findings suggest that Kv7 channels play a protective role in the pulmonary circulation limiting pulmonary vasoconstriction, and their activation may represent a promising therapeutic strategy in pulmonary hypertension.

### Kv7 CHANNELS: NEW PHARMACOLOGICAL TARGETS IN RESPIRATORY DISEASES?

In addition to their well-established role in neural and cardiac tissue, studies over the last 20 decades have evidenced the expression and the plethora of cellular functions of Kv7 channels in numerous cell types. Thus, Kv7 channels are now recognized as playing relevant physiological roles in many tissues, which has encouraged the search for the therapeutic potential of Kv7 channel modulators in many diseases including those affected the lung. As stated in the previous section activation of Kv7 channels has been proposed to provide beneficial effects in a number of lung conditions. Therefore, Kv7 channel openers/enhancers or drugs acting partly through these channels could be proposed as bronchodilators, expectorants, antitussives, chemotherapeutics, and pulmonary vasodilators, as summarized below.

Flupirtine and retigabine are structurally related Kv7.2–7.5 channel activators. The discovery of Kv7 channels came later than the extensive use of flupirtine and retigabine as an analgesic and anticonvulsant, respectively, and whose mechanism of action was unknown until then (Schenzer et al., 2005). Retigabine has been shown to increase the open probability and cause a hyperpolarizing shift in the voltage dependence of activation of *KCNQ* channels by binding at the intracellular side of the pore-forming S5 helix around a key tryptophan (Trp) Trp236 in Kv7.2 and analogous residues in Kv7.3, 4, and 5 which is absent in Kv7.1 channels (Schenzer et al., 2005; Lange et al., 2009; Kim et al., 2015). Kim et al. have suggested that the retigabine interaction with this Trp residue depends largely on the formation of a hydrogen bond (Kim et al., 2015).

As the relevant role of Kv7 channels was discovered, the idea that these channels could constitute a new therapeutic strategy became more attractive, leading to investment in the development of new activators. The key residues for retigabinebinding have also been recognized to be also essential for the binding of other compounds such as S1 and BMS-204352; though these last drugs have a greater efficacy enhancing Kv7.4 and Kv7.5 isoforms rather than Kv7.2 or/and Kv7.3 channels (Dupuis et al., 2002; Bentzen et al., 2006). Zinc pyrithione activates the Kv7 channel by interacting with a binding site different to that of retigabine (Xiong et al., 2008). Hence, zinc pyrithione activates all Kv7 channels except for Kv7.3 channels, which is the most retigabine-sensitive. It is worth noting that the active compound in the action of zinc pyrithione on Kv7 channels is zinc, as administration of free zinc also activates these channels by reducing their dependence on PIP<sub>2</sub> (Gao et al., 2017). The activity of ICA-27243, a drug more potent at activating Kv7.2/7.3 heteromeric channels than Kv7.4 homomeric channels or Kv7.3/7.5 heteromeric channels, is determined by a novel site inside the Kv7 channel voltage-detection domain (Wickenden et al., 2008; Blom et al., 2010). R-L3 is a benzodiazepine that activates Kv7.1 homomeric channels, but is considerably weaker when Kv7.1 is coexpressed with KCNE1 subunit, suggesting that R-L3 and KCNE1 subunits compete for the same interaction site on *KCNQ1* subunits (Salata et al., 1998).

Kv7 channels are currently new therapeutic targets in the field of lung diseases; consequently, there is still much to be learned about their physiological role, regulation, and possible impairment by pathological processes. The specific expression of certain Kv7 channel in different types of cells is of great pharmacological interest, since their selective modulation could provide beneficial therapeutic effects while avoiding the adverse effects associated with other isoforms (Stott et al., 2014). A clear example comes from the previous experience with retigabine and flupirtine, which have been used as analgesic and anticonvulsant, respectively, for many years but were withdrawn from the marked due to their adverse effects including the risk of serious liver injury, pigment changes in the retina and urinary retention (Herrmann et al., 1987; Brickel et al., 2012). While, as mentioned, both drugs activate Kv7.2-Kv7.5, they appear to display greater efficacy for Kv7.2 and Kv7.3 channels over Kv7.4 and Kv7.5 channels, which predominate in smooth muscle. Thus, it is expected that compounds selectively activate the Kv7.4 and/or the Kv7.5 subunit may be used in the treatment of vascular smooth muscle disorders with few neurological side effects. For instance, selective activation of Kv7.4 and Kv7.4/Kv7.5 channels (but not Kv7.1, Kv7.2, and Kv7.2/Kv7.3) has been implicated in the vasorelaxant effects of the Rho kinase inhibitor fasudil (Zhang et al., 2016). Thus, it is likely that, in addition to Rho kinase inhibition, activation of these channels may contribute to the beneficial effects of fasudil in cardiovascular conditions such as hypertension, vasospasm, stroke arteriosclerosis, pulmonary hypertension, and heart failure (Shi and Wei, 2013; Shimokawa and Satoh, 2015). Likewise, it is expected that selective Kv7.2 and Kv7.3 channel enhancers may be used to the treatment of neurological disorders with few adverse effects in smooth muscle cells (Stott et al., 2014; Barrese et al., 2018). Remarkably, Manville et al. have recently reported that the neurotransmitter  $\gamma$ -aminobutyric acid (GABA) and various metabolites ( $\beta$ -hydroxybutyric acid and  $\gamma$ -aminoβ-hydroxybutyric acid) directly activate Kv7.3 and Kv7.5 but not Kv7.1, 2, and 4 (Manville et al., 2018). Manville and colleagues found that Trp residues in Kv7.3 (Trp265) and Kv7.5 (Trp270) were required for binding but not sufficient for GABAinduced activation. Thus, while Kv7.2 and Kv7.4 share analogous Trp residues and bind GABA, they are not activated by GABA (Manville and Abbott, 2018). Likewise, the synthetic GABA analog gabapentin, but not pregabalin, was also found to activate Kv7.3 and Kv7.5 (Manville and Abbott, 2018). Very recently, these authors have proposed a model in which the sodiumcoupled myo-inositol transporter 1 (SMIT1) interacts with KCNQ2/3 and tunes the channel response to GABA and related metabolites (Manville and Abbott, 2020).

The search for Kv7 activators with a better safety profile than retigabine and flupirtine as well as the identification of novel potential therapeutic applications, has driven the development of a large number of compounds with Kv7 enhancing properties in the last decade (Jepps et al., 2013; Stott et al., 2014; Haick and Byron, 2016; Barrese et al., 2018).

Given the functional roles and ubiquitous expression of Kv7 channels in many tissues, drugs modulating Kv7 channels are presumed to cause off-target effects including alterations of the QT interval at the heart level, systemic hypotension, urinary retention or neuronal effects. This could be largely avoided by using localized drug delivery to the lungs by oral inhalation. Inhaled administration is the route of choice for many drugs (i.e., corticosteroids; anticholinegics, or  $\beta$ 2-adrenergic receptor agonists) to treat respiratory diseases. Similarly, targeting lung Kv7 channels specifically using inhaled Kv7 activators may be an innovative approach for the treatment of COPD, cystic fibrosis, pulmonary hypertension, asthmatic events, lung cancer, bronchitis, and pulmonary edema.

Progress in the treatment of respiratory diseases is a global challenge because of its health and socio-economic implications. The optimal management and treatment of these diseases should be aimed at achieving adequate control of the disease, improving the quality of life of patients, and reducing hospitalizations and the enormous financial burden on healthcare systems. Targeting Kv7 channels in the treatment of lung disorders could be a promising therapeutic strategy. Here, we summarized the potential usefulness of Kv channel modulators in treating respiratory diseases.

#### **Bronchodilators**

Asthma and COPD are characterized by hyper-contraction of ASMC attributed to abnormally high concentrations of different bronchoconstrictor agonists and/or increased sensitivity to them, leading to pathological narrowing of the airways (Penn, 2008). K<sup>+</sup> channels are key regulators of Em in ASMC and their impairment causes membrane depolarization and bronchoconstriction (Hall, 2000). Among them, Kv7 channels contribute significantly to the negative resting Em in ASMC (Brueggemann et al., 2011). By acting on a different target, Kv7 channel openers offer the potential of being combined with standard bronchodilators such as  $\beta$ 2-adrenergic receptor agonists and antimuscarinics as an adjuvant therapy to increase the efficacy or overcome desensitization (Brueggemann et al., 2014b).

#### **Expectorants**

The function of Kv7.1 together with KCNE1/E2/E3 subunits seems to play a key role in secretions in airway epithelial cells. The co-activation of Cl<sup>-</sup> channels in the luminal surface and K<sup>+</sup> channels in the basolateral membrane in AEC is an essential requirement for Cl<sup>-</sup> secretion (Frizzell and Hanrahan, 2012). The activation of Kv7.1 channels has an expectorant effect provoking or promoting the expulsion of accumulated bronchial secretions and the productive cough helping to airway clearance

(Mall et al., 2000; Al-Hazza et al., 2016; Kroncke et al., 2016; Hynes and Harvey, 2019). Kv7.1 channel openers or drugs that indirectly activate Kv7.1 channels might be beneficial in the treatment of bronchitis, COPD, cystic fibrosis and asthma (Boucherot et al., 2001; Athanazio, 2012; Hynes and Harvey, 2019).

#### Antitussives

Stimulation of airway C-fibers can be activated by inflammatory factors leading to the impulse to unproductive cough, dyspnea, mucus secretion and bronchoconstriction (Mazzone and Undem, 2016). Activation of Kv7.3 channels in VACF by retigabine has been shown to increase the M-current amplitude and to inhibit unproductive coughing (Sun et al., 2019). Thus, Kv7 activators could be useful as chronic coughing relievers in pulmonary inflammatory diseases.

#### Chemotherapy

Although, the role of Kv7 channel modulators as chemotherapy agents has been only barely studied, flupirtine has been shown to attenuate the resistance to gefitinib in a human lung cancer cell line (Choi et al., 2017). The anti-proliferative effect of flupirtine has also been proposed as potentially beneficial in other types of cancer (Lee et al., 2014). Additional studies are required to evaluate the potential of Kv7 channel activators with selectivity for particular isoforms as chemotherapy agents.

#### **Pulmonary Vasodilators**

K<sup>+</sup> channels are also key regulators of cellular Em in PASMCs, controlling the activity of voltage-dependent Ca2+ channels and, subsequently, pulmonary artery smooth muscle contraction. Pulmonary vascular tone is controlled by the interaction of different circulating or locally released vasoactive factors with vasoconstrictor or vasodilating properties. Acute modulation of the activity of K<sup>+</sup> channels has been extensively shown to contribute to the vascular effects of these vasoactive factors. Thus, different vasodilators such as nitric oxide, prostacyclin and calcitonin gene-related activate several K<sup>+</sup> channels and, conversely, vasoconstrictors as serotonin, angiotensin II, thromboxane A2, noradrenaline and endothelin-1 have been shown to inhibit K<sup>+</sup> channels (Cogolludo et al., 2007; Stott et al., 2014). With such a crucial role in modulating and maintaining pulmonary arterial tone, changes in function and expression of K<sup>+</sup> channels during the development and pathogenesis of pulmonary hypertension results in aberrations of normal physiological functions. Consequently, K<sup>+</sup> channels openers are presumed to have a beneficial therapeutic effect, especially if their action can be directed to the pulmonary

#### REFERENCES

- Abbott, G. W. (2016). KCNE4 and KCNE5: K(+) channel regulation and cardiac arrhythmogenesis. *Gene* 593, 249–260. doi: 10.1016/j.gene.2016.07.069
- Al-Hazza, A., Linley, J., Aziz, Q., Hunter, M., and Sandle, G. (2016). Upregulation of basolateral small conductance potassium channels (KCNQ1/KCNE3) in ulcerative colitis. *Biochem. Biophys. Res. Commun.* 470, 473–478. doi: 10.1016/j. bbrc.2015.12.086

circulation (Gurney et al., 2010). Up to the present time, the availability of drugs modulating Kv7 channels has been relatively scarce. Advances in the understanding of the role of Kv7 channels in the pulmonary vasculature and the development of new selective drugs for these channels indicate that they may represent a useful therapeutic target for the treatment of pulmonary arterial hypertension. As detailed above, different studies have provided evidence that treatment with Kv7 channel activators such as flupirtine, retigabine, or fasudil is beneficial in different pulmonary hypertension models (Morecroft et al., 2009; Duong-Quy et al., 2013; Sedivy et al., 2015; Zhang and Wu, 2017). In addition, Kv7 channel activation has been shown to contribute to the pulmonary vasodilating effect induced by drugs approved for the pulmonary hypertension treatment such as nitric oxide or riociguat (Mondejar-Parreño et al., 2019).

#### CONCLUSIONS

In the last decade, a significant progress in the understanding of the role of Kv7 channels in lung physiology has been made. The recognition of the involvement of these channels in cellular functions of crucial relevance in the normal physiology of the lung, such as the control of water and salt transport across the epithelial cell membrane, the excitability of airway afferent nerves, and the regulation of airway and lung vascular smooth muscle tone, makes them attractive therapeutic targets in lung diseases. In this regard, deciphering the involvement and importance of specific Kv7 subunits in these cellular functions will lead to a significant advance that may constitute the basis for the therapeutic potential of selective Kv7 channel modulators in lung diseases.

#### AUTHOR CONTRIBUTIONS

GM-P, FP-V, and AC wrote the manuscript. All authors contributed to the article and approved the submitted version.

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Arcangeli, A., Crociani, O., Lastraioli, E., Masi, A., Pillozzi, S., and Becchetti, A. (2009). Targeting ion channels in cancer: a novel frontier in antineoplastic therapy. *Curr. Med. Chem.* 16, 66–93. doi: 10.2174/092986709787002835

Athanazio, R. (2012). Airway disease: similarities and differences between asthma, COPD and bronchiectasis. *Clinics* 67, 1335–1343. doi: 10.6061/clinics/2012(11)19

Bardou, O., Trinh, N. T. N., and Brochiero, E. (2009). Molecular diversity and function of K<sup>+</sup> channels in airway and alveolar epithelial cells. Am. J. Phys. Lung Cell. Mol. Phys. 296, L145–L155. doi: 10.1152/ajplung.90525.2008

- Barnes, P. J., and Liu, S. F. (1995). Regulation of pulmonary vascular tone. *Pharmacol. Rev.* 47, 87–131.
- Barnes, P. J., Shapiro, S. D., and Pauwels, R. A. (2003). Chronic obstructive pulmonary disease: molecular and cellularmechanisms. *Eur. Respir. J.* 22, 672–688. doi: 10.1183/09031936.03.00040703
- Barrese, V., Stott, J. B., and Greenwood, I. A. (2018). KCNQ-encoded potassium channels as therapeutic targets. *Annu. Rev. Pharmacol. Toxicol.* 58, 625–648. doi: 10.1146/annurev-pharmtox-010617-052912
- Barro-Soria, R., Perez, M. E., and Larsson, H. P. (2015). KCNE3 acts by promoting voltage sensor activation in KCNQ1. Proc. Natl. Acad. Sci. 112, E7286–E7292. doi: 10.1073/pnas.1516238112
- Bartoszewski, R., Matalon, S., and Collawn, J. F. (2017). Ion channels of the lung and their role in disease pathogenesis. Am. J. Phys. Lung Cell. Mol. Phys. 313, L859–L872. doi: 10.1152/ajplung.00285.2017
- Belfodil, R., Barrière, H., Rubera, I., Tauc, M., Poujeol, C., Bidet, M., et al. (2003). CFTR-dependent and -independent swelling-activated K<sup>+</sup> currents in primary cultures of mouse nephron. *Am. J. Physiol. Ren. Physiol.* 284, F812–F828. doi: 10.1152/ajprenal.00238.2002
- Bentzen, B. H., Schmitt, N., Calloe, K., Dalby Brown, W., Grunnet, M., and Olesen, S.-P. (2006). The acrylamide (S)-1 differentially affects Kv7 (KCNQ) potassium channels. *Neuropharmacology* 51, 1068–1077. doi: 10.1016/j. neuropharm.2006.07.001
- Bierbower, S. M., Choveau, F. S., Lechleiter, J. D., and Shapiro, M. S. (2015). Augmentation of M-type (KCNQ) potassium channels as a novel strategy to reduce stroke-induced brain injury. *J. Neurosci.* 35, 2101–2111. doi: 10.1523/ JNEUROSCI.3805-14.2015
- Billet, A., and Hanrahan, J. W. (2013). The secret life of CFTR as a calciumactivated chloride channel. J. Physiol. 591, 5273–5278. doi: 10.1113/ jphysiol.2013.261909
- Blom, S. M., Schmitt, N., and Jensen, H. S. (2010). Differential effects of ICA-27243 on cloned K(V)7 channels. *Pharmacology* 86, 174–181. doi: 10.1159/000317525
- Boucherat, O., Chabot, S., Antigny, F., Perros, F., Provencher, S., and Bonnet, S. (2015). Potassium channels in pulmonary arterial hypertension. *Eur. Respir. J.* 46, 1167–1177. doi: 10.1183/13993003.00798-2015
- Boucherot, A., Schreiber, R., and Kunzelmann, K. (2001). Regulation and properties of KCNQ1 (K(V)LQT1) and impact of the cystic fibrosis transmembrane conductance regulator. J. Membr. Biol. 182, 39–47. doi: 10.1007/s00232-001-0030-4
- Brickel, N., Gandhi, P., VanLandingham, K., Hammond, J., and DeRossett, S. (2012). The urinary safety profile and secondary renal effects of retigabine (ezogabine): a first-in-class antiepileptic drug that targets KCNQ (K(v)7) potassium channels. *Epilepsia* 53, 606–612. doi: 10.1111/j.1528-1167.2012.03441.x
- Brueggemann, L. I., Cribbs, L. L., Schwartz, J., Wang, M., Kouta, A., and Byron, K. L. (2018). Mechanisms of PKA-dependent potentiation of Kv7.5 channel activity in human airway smooth muscle cells. *Int. J. Mol. Sci.* 19:2223. doi: 10.3390/ijms19082223
- Brueggemann, L. I., Haick, J. M., Cribbs, L. L., and Byron, K. L. (2014a). Differential activation of vascular smooth muscle Kv7.4, Kv7.5, and Kv7.4/7.5 channels by ML213 and ICA-069673. *Mol. Pharmacol.* 86, 330–341. doi: 10.1124/mol.114.093799
- Brueggemann, L. I., Haick, J. M., Neuburg, S., Tate, S., Randhawa, D., Cribbs, L. L., et al. (2014b). KCNQ (Kv7) potassium channel activators as bronchodilators: combination with a β2-adrenergic agonist enhances relaxation of rat airways. *Am. J. Phys. Lung Cell. Mol. Phys.* 306, L476–L486. doi: 10.1152/ajplung.00253.2013
- Brueggemann, L. I., Kakad, P. P., Love, R. B., Solway, J., Dowell, M. L., Cribbs, L. L., et al. (2011). Kv7 potassium channels in airway smooth muscle cells: signal transduction intermediates and pharmacological targets for bronchodilator therapy. Am. J. Phys. Lung Cell. Mol. Phys. 302, L120–L132. doi: 10.1152/ajplung.00194.2011
- Burg, E. D., Remillard, C. V., and Yuan, J. X.-J. (2008). Potassium channels in the regulation of pulmonary artery smooth muscle cell proliferation and apoptosis: pharmacotherapeutic implications. *Br. J. Pharmacol.* 153, S99–S111. doi: 10.1038/sj.bjp.0707635
- Butrous, G. (2015). Human immunodeficiency virus-associated pulmonary arterial hypertension: considerations for pulmonary vascular diseases in the developing world. *Circulation* 131, 1361–1370. doi: 10.1161/CIRCULATIONAHA.114.006978
- Byron, K. L., and Brueggemann, L. I. (2018). Kv7 potassium channels as signal transduction intermediates in the control of microvascular tone. *Microcirculation* 25:e12419. doi: 10.1111/micc.12419

- Caulfield, M. P., Jones, S., Vallis, Y., Buckley, N. J., Kim, G. D., Milligan, G., et al. (1994). Muscarinic M-current inhibition via G alpha q/11 and alphaadrenoceptor inhibition of Ca<sup>2+</sup> current via G alpha o in rat sympathetic neurones. J. Physiol. 477, 415–422. doi: 10.1113/jphysiol.1994.sp020203
- Chadha, P. S., Jepps, T. A., Carr, G., Stott, J. B., Zhu, H.-L., Cole, W. C., et al. (2014). Contribution of kv7.4/kv7.5 heteromers to intrinsic and calcitonin gene-related peptide-induced cerebral reactivity. *Arterioscler. Thromb. Vasc. Biol.* 34, 887–893. doi: 10.1161/ATVBAHA.114.303405
- Chadha, P. S., Zunke, F., Davis, A. J., Jepps, T. A., Linders, J. T. M., Schwake, M., et al. (2012). Pharmacological dissection of K(v)7.1 channels in systemic and pulmonary arteries. *Br. J. Pharmacol.* 166, 1377–1387. doi: 10.1111/j. 1476-5381.2012.01863.x
- Chen, Y.-H., Xu, S.-J., Bendahhou, S., Wang, X.-L., Wang, Y., Xu, W.-Y., et al. (2003). KCNQ1 gain-of-function mutation in familial atrial fibrillation. *Science* 299, 251–254. doi: 10.1126/science.1077771
- Choi, S. Y., Kim, H.-R., Ryu, P. D., and Lee, S. Y. (2017). Regulation of voltage-gated potassium channels attenuates resistance of side-population cells to gefitinib in the human lung cancer cell line NCI-H460. BMC Pharmacol. Toxicol. 18:14. doi: 10.1186/s40360-017-0118-9
- Clarke, L. L., Grubb, B. R., Yankaskas, J. R., Cotton, C. U., McKenzie, A., and Boucher, R. C. (1994). Relationship of a non-cystic fibrosis transmembrane conductance regulator-mediated chloride conductance to organ-level disease in Cftr(-/-) mice. *Proc. Natl. Acad. Sci.* 91, 479–483. doi: 10.1073/pnas.91.2.479
- Cogolludo, A., Moreno, L., and Villamor, E. (2007). Mechanisms controlling vascular tone in pulmonary arterial hypertension: implications for vasodilator therapy. *Pharmacology* 79, 65–75. doi: 10.1159/000097754
- Cohen, M. H., Williams, G. A., Sridhara, R., Chen, G., and Pazdur, R. (2003). FDA drug approval summary: gefitinib (ZD1839) (Iressa) tablets. *Oncologist* 8, 303–306. doi: 10.1634/theoncologist.8-4-303
- Corvol, H., Flamein, F., Epaud, R., Clement, A., and Guillot, L. (2009). Lung alveolar epithelium and interstitial lung disease. *Int. J. Biochem. Cell Biol.* 41, 1643–1651. doi: 10.1016/j.biocel.2009.02.009
- Cowley, E. A., and Linsdell, P. (2002a). Characterization of basolateral K<sup>+</sup> channels underlying anion secretion in the human airway cell line Calu-3. *J. Physiol.* 538, 747–757. doi: 10.1113/jphysiol.2001.013300
- Cowley, E. A., and Linsdell, P. (2002b). Oxidant stress stimulates anion secretion from the human airway epithelial cell line Calu-3: implications for cystic fibrosis lung disease. J. Physiol. 543, 201–209. doi: 10.1113/jphysiol.2002.022400
- Cummings, K. W., and Bhalla, S. (2015). Pulmonary vascular diseases. Clin. Chest Med. 36, 235-248. doi: 10.1016/j.ccm.2015.02.007
- Cunningham, S. A., Worrell, R. T., Benos, D. J., and Frizzell, R. A. (1992). CAMP-stimulated ion currents in *Xenopus* oocytes expressing CFTR cRNA. *Am. J. Phys.* 262, C783–C788. doi: 10.1152/ajpcell.1992.262.3.C783
- de Jong, I. E. M., and Jepps, T. A. (2018). Impaired Kv7 channel function in cerebral arteries of a tauopathy mouse model (rTg4510). *Phys. Rep.* 6:e13920. doi: 10.14814/phy2.13920
- Dellaripa, P. F. (2018). Interstitial lung disease in the connective tissue diseases; a paradigm shift in diagnosis and treatment. *Clin. Immunol.* 186, 71–73. doi: 10.1016/j.clim.2017.09.015
- Delmas, P., and Brown, D. A. (2005). Pathways modulating neural KCNQ/M (Kv7) potassium channels. *Nat. Rev. Neurosci.* 6, 850–862. doi: 10.1038/nrn1785
- Duong-Quy, S., Bei, Y., Liu, Z., and Dinh-Xuan, A. T. (2013). Role of rhokinase and its inhibitors in pulmonary hypertension. *Pharmacol. Ther.* 137, 352–364. doi: 10.1016/j.pharmthera.2012.12.003
- Dupuis, D. S., Schrøder, R. L., Jespersen, T., Christensen, J. K., Christophersen, P., Jensen, B. S., et al. (2002). Activation of KCNQ5 channels stably expressed in HEK293 cells by BMS-204352. *Eur. J. Pharmacol.* 437, 129–137. doi: 10.1016/S0014-2999(02)01287-6
- Eid, B. G., and Gurney, A. M. (2018). Zinc pyrithione activates K<sup>+</sup> channels and hyperpolarizes the membrane of rat pulmonary artery smooth muscle cells. *PLoS One* 13:e0192699. doi: 10.1371/journal.pone.0192699
- Evseev, A. I., Semenov, I., Archer, C. R., Medina, J. L., Dube, P. H., Shapiro, M. S., et al. (2013). Functional effects of KCNQ K(+) channels in airway smooth muscle. *Front. Physiol.* 4:277. doi: 10.3389/fphys.2013.00277
- Fosmo, A. L., and Skraastad, Ø. B. (2017). The Kv7 channel and cardiovascular risk factors. Front. Cardiovasc. Med. 4:75. doi: 10.3389/fcvm.2017.00075
- Frizzell, R. A., and Hanrahan, J. W. (2012). Physiology of epithelial chloride and fluid secretion. *Cold Spring Harb. Perspect. Med.* 2:a009563. doi: 10.1101/ cshperspect.a009563

- Gamper, N., Zaika, O., Li, Y., Martin, P., Hernandez, C. C., Perez, M. R., et al. (2006). Oxidative modification of M-type K(+) channels as a mechanism of cytoprotective neuronal silencing. *EMBO J.* 25, 4996–5004. doi: 10.1038/ sj.emboj.7601374
- Gao, H., Boillat, A., Huang, D., Liang, C., Peers, C., and Gamper, N. (2017). Intracellular zinc activates KCNQ channels by reducing their dependence on phosphatidylinositol 4,5-bisphosphate. *Proc. Natl. Acad. Sci. U. S. A.* 114, E6410–E6419. doi: 10.1073/pnas.1620598114
- Girault, A., Privé, A., Trinh, N. T. N., Bardou, O., Ferraro, P., Joubert, P., et al. (2014). Identification of KvLQT1 K<sup>+</sup> channels as new regulators of non-small cell lung cancer cell proliferation and migration. *Int. J. Oncol.* 44, 838–848. doi: 10.3892/ijo.2013.2228
- Goodwill, A. G., Fu, L., Noblet, J. N., Casalini, E. D., Sassoon, D., Berwick, Z. C., et al. (2016). KV7 channels contribute to paracrine, but not metabolic or ischemic, regulation of coronary vascular reactivity in swine. Am. J. Physiol. Heart Circ. Physiol. 310, H693–H704. doi: 10.1152/ajpheart.00688.2015
- Group (NINOS) (1997). Inhaled nitric oxide and hypoxic respiratory failure in infants with congenital diaphragmatic hernia. *Pediatrics* 99, 838–845. doi: 10.1542/peds.99.6.838
- Grubb, B. R., Vick, R. N., and Boucher, R. C. (1994). Hyperabsorption of Na<sup>+</sup> and raised Ca(2+)-mediated Cl<sup>−</sup> secretion in nasal epithelia of CF mice. *Am. J. Phys. Cell Phys.* 266, C1478–C1483. doi: 10.1152/ajpcell.1994.266.5.C1478
- Grunnet, M., Jespersen, T., Rasmussen, H. B., Ljungstrøm, T., Jorgensen, N. K., Olesen, S.-P., et al. (2002). KCNE4 is an inhibitory subunit to the KCNQ1 channel. J. Physiol. 542, 119–130. doi: 10.1113/jphysiol.2002.017301
- Gurney, A. M., Joshi, S., and Manoury, B. (2010). KCNQ potassium channels: new targets for pulmonary vasodilator drugs? Adv. Exp. Med. Biol. 661, 405–417. doi: 10.1007/978-1-60761-500-2\_26
- Haick, J. M., Brueggemann, L. I., Cribbs, L. L., Denning, M. F., Schwartz, J., and Byron, K. L. (2017). PKC-dependent regulation of Kv7.5 channels by the bronchoconstrictor histamine in human airway smooth muscle cells. Am. J. Phys. Lung Cell. Mol. Phys. 312, L822–L834. doi: 10.1152/ajplung.00567.2016
- Haick, J. M., and Byron, K. L. (2016). Novel treatment strategies for smooth muscle disorders: targeting Kv7 potassium channels. *Pharmacol. Ther.* 165, 14–25. doi: 10.1016/j.pharmthera.2016.05.002
- Haley, J. E., Abogadie, F. C., Delmas, P., Dayrell, M., Vallis, Y., Milligan, G., et al. (1998). The alpha subunit of Gq contributes to muscarinic inhibition of the M-type potassium current in sympathetic neurons. *J. Neurosci.* 18, 4521–4531. doi: 10.1523/JNEUROSCI.18-12-04521.1998
- Hall, I. P. (2000). Second messengers, ion channels and pharmacology of airway smooth muscle. *Eur. Respir. J.* 15, 1120–1127. doi: 10.1034/j.1399-3003.2000.01523.x
- Halpin, D. M. G., Faner, R., Sibila, O., Badia, J. R., and Agusti, A. (2020). Do chronic respiratory diseases or their treatment affect the risk of SARS-CoV-2 infection? *Lancet Respir. Med.* 5, 436–438. doi: 10.1016/S2213-2600(20)30167-3
- Haworth, A. S., and Brackenbury, W. J. (2019). Emerging roles for multifunctional ion channel auxiliary subunits in cancer. *Cell Calcium* 80, 125–140. doi: 10.1016/j.ceca.2019.04.005
- Hayabuchi, Y. (2017). The action of smooth muscle cell potassium channels in the pathology of pulmonary arterial hypertension. *Pediatr. Cardiol.* 38, 1–14. doi: 10.1007/s00246-016-1491-7
- Herrmann, W. M., Kern, U., and Aigner, M. (1987). On the adverse reactions and efficacy of long-term treatment with flupirtine: preliminary results of an ongoing twelve-month study with 200 patients suffering from chronic pain states in arthrosis or arthritis. *Postgrad. Med. J.* 63, 87–103.
- Higashida, H., Hoshi, N., Zhang, J.-S., Yokoyama, S., Hashii, M., Jin, D., et al. (2005). Protein kinase C bound with A-kinase anchoring protein is involved in muscarinic receptor-activated modulation of M-type KCNQ potassium channels. *Neurosci. Res.* 51, 231–234. doi: 10.1016/j.neures.2004.11.009
- Hu, B., Zeng, W.-P., Li, X., Al-Sheikh, U., Chen, S.-Y., and Ding, J. (2019). A conserved arginine/lysine-based motif promotes ER export of KCNE1 and KCNE2 to regulate KCNQ1 channel activity. *Channels* 13, 483–497. doi: 10.1080/19336950.2019.1685626
- Hynes, D., and Harvey, B. J. (2019). Dexamethasone reduces airway epithelial Cl<sup>-</sup> secretion by rapid non-genomic inhibition of KCNQ1, KCNN4 and KATP K<sup>+</sup> channels. *Steroids* 151:108459. doi: 10.1016/j.steroids.2019.108459
- Iannotti, F. A., Panza, E., Barrese, V., Viggiano, D., Soldovieri, M. V., and Taglialatela, M. (2010). Expression, localization, and pharmacological role of Kv7 potassium channels in skeletal muscle proliferation, differentiation,

and survival after myotoxic insults. J. Pharmacol. Exp. Ther. 332, 811-820. doi: 10.1124/jpet.109.162800

- Ivonnet, P., Salathe, M., and Conner, G. E. (2015). Hydrogen peroxide stimulation of CFTR reveals an Epac-mediated, soluble AC-dependent cAMP amplification pathway common to GPCR signalling. *Br. J. Pharmacol.* 172, 173–184. doi: 10.1111/bph.12934
- Jang, S. H., Choi, S. Y., Ryu, P. D., and Lee, S. Y. (2011). Anti-proliferative effect of Kv1.3 blockers in A549 human lung adenocarcinoma in vitro and in vivo. *Eur. J. Pharmacol.* 651, 26–32. doi: 10.1016/j.ejphar.2010.10.066
- Jentsch, T. J. (2000). Neuronal KCNQ potassium channels: physiology and role in disease. Nat. Rev. Neurosci. 1, 21-30. doi: 10.1038/35036198
- Jepps, T. A., Carr, G., Lundegaard, P. R., Olesen, S.-P., and Greenwood, I. A. (2015). Fundamental role for the KCNE4 ancillary subunit in Kv7.4 regulation of arterial tone. J. Physiol. 593, 5325–5340. doi: 10.1113/JP271286
- Jepps, T. A., Olesen, S. P., and Greenwood, I. A. (2013). One man's side effect is another man's therapeutic opportunity: targeting Kv7 channels in smooth muscle disorders. Br. J. Pharmacol. 168, 19–27. doi: 10.1111/j.1476-5381.2012.02133.x
- Joshi, S., Sedivy, V., Hodyc, D., Herget, J., and Gurney, A. M. (2009). KCNQ modulators reveal a key role for KCNQ potassium channels in regulating the tone of rat pulmonary artery smooth muscle. *J. Pharmacol. Exp. Ther.* 329, 368–376. doi: 10.1124/jpet.108.147785
- Jude, J. A., Wylam, M. E., Walseth, T. F., and Kannan, M. S. (2008). Calcium signaling in airway smooth muscle. *Proc. Am. Thorac. Soc.* 5, 15–22. doi: 10.1513/pats.200704-047VS
- Jung, J., Lin, H., Koh, Y. I., Ryu, K., Lee, J. S., Rim, J. H., et al. (2019). Rare KCNQ4 variants found in public databases underlie impaired channel activity that may contribute to hearing impairment. *Exp. Mol. Med.* 51, 1–12. doi: 10.1038/s12276-019-0300-9
- Kim, H.-J., Jeong, M.-H., Kim, K.-R., Jung, C.-Y., Lee, S.-Y., Kim, H., et al. (2016). Protein arginine methylation facilitates KCNQ channel-PIP2 interaction leading to seizure suppression. *Elife* 5:e17159. doi: 10.7554/eLife.17159
- Kim, R. Y., Yau, M. C., Galpin, J. D., Seebohm, G., Ahern, C. A., Pless, S. A., et al. (2015). Atomic basis for therapeutic activation of neuronal potassium channels. *Nat. Commun.* 6:8116. doi: 10.1038/ncomms9116
- Kroncke, B. M., Van Horn, W. D., Smith, J., Kang, C., Welch, R. C., Song, Y., et al. (2016). Structural basis for KCNE3 modulation of potassium recycling in epithelia. *Sci. Adv.* 2:e1501228. doi: 10.1126/sciadv.1501228
- Kubisch, C., Schroeder, B. C., Friedrich, T., Lütjohann, B., El-Amraoui, A., Marlin, S., et al. (1999). KCNQ4, a novel potassium channel expressed in sensory outer hair cells, is mutated in dominant deafness. *Cell* 96, 437–446. doi: 10.1016/S0092-8674(00)80556-5
- Lange, W., Geissendörfer, J., Schenzer, A., Grötzinger, J., Seebohm, G., Friedrich, T., et al. (2009). Refinement of the binding site and mode of action of the anticonvulsant retigabine on KCNQ K<sup>+</sup> channels. *Mol. Pharmacol.* 75, 272–280. doi: 10.1124/mol.108.052282
- Laumbach, R. J., and Kipen, H. M. (2012). Respiratory health effects of air pollution: update on biomass smoke and traffic pollution. J. Allergy Clin. Immunol. 129, 3–13. doi: 10.1016/j.jaci.2011.11.021
- Lee, B. H., Ryu, P. D., and Lee, S. Y. (2014). Serum starvation-induced voltagegated potassium channel Kv7.5 expression and its regulation by Sp1 in canine osteosarcoma cells. *Int. J. Mol. Sci.* 15, 977–993. doi: 10.3390/ ijms15010977
- Lehman, A., Thouta, S., Mancini, G. M. S., Naidu, S., van Slegtenhorst, M., and McWalter, K. (2017). Loss-of-function and gain-of-function mutations in KCNQ5 cause intellectual disability or epileptic encephalopathy. Am. J. Hum. Genet. 101, 65–74. doi: 10.1016/j.ajhg.2017.05.016
- Leon, G., MacDonagh, L., Finn, S. P., Cuffe, S., and Barr, M. P. (2016). Cancer stem cells in drug resistant lung cancer: targeting cell surface markers and signaling pathways. *Pharmacol. Ther.* 158, 71–90. doi: 10.1016/j. pharmthera.2015.12.001
- Li, H., Chen, Q., Moss, A. J., Robinson, J., Goytia, V., Perry, J. C., et al. (1998). New mutations in the KVLQT1 potassium channel that cause long-QT syndrome. *Circulation* 97, 1264–1269. doi: 10.1161/01.CIR.97.13.1264
- Li, C., Huang, P., Lu, Q., Zhou, M., Guo, L., and Xu, X. (2014a). KCNQ/Kv7 channel activator flupirtine protects against acute stress-induced impairments of spatial memory retrieval and hippocampal LTP in rats. *Neuroscience* 280, 19–30. doi: 10.1016/j.neuroscience.2014.09.009
- Li, S.-S., Ran, Y.-J., Zhang, D.-D., Li, S.-Z., and Zhu, D. (2014b). MicroRNA-190 regulates hypoxic pulmonary vasoconstriction by targeting a voltage-gated

K<sup>+</sup> channel in arterial smooth muscle cells. *J. Cell. Biochem.* 115, 1196–1205. doi: 10.1002/jcb.24771

- Li, X., Xu, S., Yu, M., Wang, K., Tao, Y., Zhou, Y., et al. (2020). Risk factors for severity and mortality in adult COVID-19 inpatients in Wuhan. J. Allergy Clin. Immunol. S0091-6749(20)30495-4. doi: 10.1016/j.jaci.2020.04.006
- Lin, Y., Wang, X., and Jin, H. (2014). EGFR-TKI resistance in NSCLC patients: mechanisms and strategies. *Am. J. Cancer Res.* 4, 411–435.
- Lindman, J., Khammy, M. M., Lundegaard, P. R., Aalkjær, C., and Jepps, T. A. (2018). Microtubule regulation of Kv7 channels orchestrates cAMP-mediated vasorelaxations in rat arterial smooth muscle. *Hypertension* 71, 336–345. doi: 10.1161/HYPERTENSIONAHA.117.10152
- Loussouarn, G., Demolombe, S., Mohammad-Panah, R., Escande, D., and Baró, I. (1996). Expression of CFTR controls cAMP-dependent activation of epithelial K<sup>+</sup> currents. Am. J. Phys. 271, C1565–C1573. doi: 10.1152/ajpcell.1996. 271.5.C1565
- Lundby, A., Ravn, L. S., Svendsen, J. H., Hauns, S., Olesen, S.-P., and Schmitt, N. (2008). KCNE3 mutation V17M identified in a patient with lone atrial fibrillation. *Cell. Physiol. Biochem.* 21, 47–54. doi: 10.1159/000113746
- Lundquist, A. L., Turner, C. L., Ballester, L. Y., and George, A. L. (2006). Expression and transcriptional control of human KCNE genes. *Genomics* 87, 119–128. doi: 10.1016/j.ygeno.2005.09.004
- Mackie, A. R., Brueggemann, L. I., Henderson, K. K., Shiels, A. J., Cribbs, L. L., Scrogin, K. E., et al. (2008). Vascular KCNQ potassium channels as novel targets for the control of mesenteric artery constriction by vasopressin, based on studies in single cells, pressurized arteries, and in vivo measurements of mesenteric vascular resistance. J. Pharmacol. Exp. Ther. 325, 475–483. doi: 10.1124/jpet.107.135764
- Maeno, E., Ishizaki, Y., Kanaseki, T., Hazama, A., and Okada, Y. (2000). Normotonic cell shrinkage because of disordered volume regulation is an early prerequisite to apoptosis. *Proc. Natl. Acad. Sci. U. S. A.* 97, 9487–9492. doi: 10.1073/pnas.140216197
- Maljevic, S., Wuttke, T. V., Seebohm, G., and Lerche, H. (2010). KV7 channelopathies. Arch. Eur. J. Physiol. 460, 277–288. doi: 10.1007/ s00424-010-0831-3
- Mall, M., Gonska, T., Thomas, J., Schreiber, R., Seydewitz, H. H., Kuehr, J., et al. (2003). Modulation of Ca<sup>2+</sup> activated Cl<sup>-</sup> secretion by basolateral K<sup>+</sup> channels in human normal and cystic fibrosis airway epithelia. *Pediatr. Res.* 53, 608–618. doi: 10.1203/01.PDR.0000057204.51420.DC
- Mall, M., Kunzelmann, K., Hipper, A., Busch, A. E., and Greger, R. (1996). CAMP stimulation of CFTR-expressing *Xenopus* oocytes activates a chromanolinhibitable K<sup>+</sup> conductance. *Arch. Eur. J. Physiol.* 432, 516–522. doi: 10.1007/ s004240050164
- Mall, M., Wissner, A., Schreiber, R., Kuehr, J., Seydewitz, H. H., Brandis, M., et al. (2000). Role of K(V)LQT1 in cyclic adenosine monophosphate-mediated Cl(–) secretion in human airway epithelia. *Am. J. Respir. Cell Mol. Biol.* 23, 283–289. doi: 10.1165/ajrcmb.23.3.4060
- Mani, B. K., Robakowski, C., Brueggemann, L. I., Cribbs, L. L., Tripathi, A., Majetschak, M., et al. (2016). Kv7.5 potassium channel subunits are the primary targets for PKA-dependent enhancement of vascular smooth muscle Kv7 currents. *Mol. Pharmacol.* 89, 323–334. doi: 10.1124/mol. 115.101758
- Manville, R. W., and Abbott, G. W. (2018). Gabapentin is a potent activator of KCNQ3 and KCNQ5 potassium channels. *Mol. Pharmacol.* 94, 1155–1163. doi: 10.1124/mol.118.112953
- Manville, R. W., and Abbott, G. W. (2020). Potassium channels act as chemosensors for solute transporters. *Commun. Biol.* 3:90. doi: 10.1038/s42003-020-0820-9
- Manville, R. W., Papanikolaou, M., and Abbott, G. W. (2018). Direct neurotransmitter activation of voltage-gated potassium channels. *Nat. Commun.* 9:1847. doi: 10.1038/s41467-018-04266-w
- Marx, S. O., Kurokawa, J., Reiken, S., Motoike, H., D'Armiento, J., Marks, A. R., et al. (2002). Requirement of a macromolecular signaling complex for beta adrenergic receptor modulation of the KCNQ1-KCNE1 potassium channel. *Science* 295, 496–499. doi: 10.1126/science.1066843
- Mazzone, S. B., and Undem, B. J. (2016). Vagal afferent innervation of the airways in health and disease. *Physiol. Rev.* 96, 975–1024. doi: 10.1152/ physrev.00039.2015
- McLaughlin, V. V., Shah, S. J., Souza, R., and Humbert, M. (2015). Management of pulmonary arterial hypertension. J. Am. Coll. Cardiol. 65, 1976–1997. doi: 10.1016/j.jacc.2015.03.540

- Melosky, B. (2014). Review of EGFRTKIs in metastatic NSCLC, including ongoing trials. Front. Oncol. 4:244. doi: 10.3389/fonc.2014.00244
- Mondejar-Parreño, G., Morales-Cano, D., Barreira, B., Callejo, M., Ruiz-Cabello, J., Moreno, L., et al. (2018). HIV transgene expression impairs K<sup>+</sup> channel function in the pulmonary vasculature. *Am. J. Phys. Lung Cell. Mol. Phys.* 315, L711–L723. doi: 10.1152/ajplung.00045.2018
- Mondejar-Parreño, G., Moral-Sanz, J., Barreira, B., De la Cruz, A., Gonzalez, T., Callejo, M., et al. (2019). Activation of Kv7 channels as a novel mechanism for NO/cGMP-induced pulmonary vasodilation. *Br. J. Pharmacol.* 176, 2131–2145. doi: 10.1111/bph.14662
- Morales-Cano, D., Callejo, M., Barreira, B., Mondejar-Parreño, G., Esquivel-Ruiz, S., Ramos, S., et al. (2019). Elevated pulmonary arterial pressure in Zucker diabetic fatty rats. *PLoS One* 14:e0211281. doi: 10.1371/journal.pone.0211281
- Morales-Cano, D., Menendez, C., Moreno, E., Moral-Sanz, J., Barreira, B., Galindo, P., et al. (2014). The flavonoid quercetin reverses pulmonary hypertension in rats. *PLoS One* 9:e114492. doi: 10.1371/journal.pone.0114492
- Morecroft, I., Murray, A., Nilsen, M., Gurney, A., and MacLean, M. (2009). Treatment with the Kv7 potassium channel activator flupirtine is beneficial in two independent mouse models of pulmonary hypertension. *Br. J. Pharmacol.* 157, 1241–1249. doi: 10.1111/j.1476-5381.2009.00283.x
- Moser, S. L., Harron, S. A., Crack, J., Fawcett, J. P., and Cowley, E. A. (2008). Multiple KCNQ potassium channel subtypes mediate basal anion secretion from the human airway epithelial cell line Calu-3. *J. Membr. Biol.* 221, 153–163. doi: 10.1007/s00232-008-9093-9
- Moudgil, R., Michelakis, E. D., and Archer, S. L. (2006). The role of K<sup>+</sup> channels in determining pulmonary vascular tone, oxygen sensing, cell proliferation, and apoptosis: implications in hypoxic pulmonary vasoconstriction and pulmonary arterial hypertension. *Microcirculation* 13, 615–632. doi: 10.1080/10739680600930222
- Neubauer, B. A., Waldegger, S., Heinzinger, J., Hahn, A., Kurlemann, G., Fiedler, B., et al. (2008). KCNQ2 and KCNQ3 mutations contribute to different idiopathic epilepsy syndromes. *Neurology* 71, 177–183. doi: 10.1212/01. wnl.0000317090.92185.ec
- O'Grady, S. M., and Lee, S. Y. (2005). Molecular diversity and function of voltage-gated (Kv) potassium channels in epithelial cells. *Int. J. Biochem. Cell Biol.* 37, 1578–1594. doi: 10.1016/j.biocel.2005.04.002
- Ohno, S., Toyoda, F., Zankov, D. P., Yoshida, H., Makiyama, T., Tsuji, K., et al. (2009). Novel KCNE3 mutation reduces repolarizing potassium current and associated with long QT syndrome. *Hum. Mutat.* 30, 557–563. doi: 10.1002/ humu.20834
- Oliveras, A., Roura-Ferrer, M., Solé, L., de la Cruz, A., Prieto, A., Etxebarria, A., et al. (2014). Functional assembly of Kv7.1/Kv7.5 channels with emerging properties on vascular muscle physiology. *Arterioscler. Thromb. Vasc. Biol.* 34, 1522–1530. doi: 10.1161/ATVBAHA.114.303801
- Panchanathan, E., Ramanathan, G., and Lakkakula, B. V. K. S. (2013). Effect of flupirtine on the growth and viability of U373 malignant glioma cells. *Cancer Biol. Med.* 10, 142–147. doi: 10.7497/j.issn.2095-3941. 2013.03.004
- Pancrazio, J. J., Tabbara, I. A., and Kim, Y. I. (1993). Voltage-activated K+ conductance and cell proliferation in small-cell lung cancer. *Anticancer Res.* 13, 1231–1234.
- Penn, R. B. (2008). Embracing emerging paradigms of G protein-coupled receptor agonism and signaling to address airway smooth muscle pathobiology in asthma. *Naunyn Schmiedeberg's Arch. Pharmacol.* 378, 149–169. doi: 10.1007/ s00210-008-0263-1
- Potet, F., Scott, J. D., Mohammad-Panah, R., Escande, D., and Baró, I. (2001). AKAP proteins anchor cAMP-dependent protein kinase to KvLQT1/IsK channel complex. Am. J. Physiol. Heart Circ. Physiol. 280, H2038–H2045. doi: 10.1152/ajpheart.2001.280.5.H2038
- Preston, P., Wartosch, L., Günzel, D., Fromm, M., Kongsuphol, P., Ousingsawat, J., et al. (2010). Disruption of the K<sup>+</sup> channel beta-subunit KCNE3 reveals an important role in intestinal and tracheal Cl<sup>-</sup> transport. *J. Biol. Chem.* 285, 7165–7175. doi: 10.1074/jbc.M109.047829
- Prevarskaya, N., Skryma, R., and Shuba, Y. (2010). Ion channels and the hallmarks of cancer. *Trends Mol. Med.* 16, 107–121. doi: 10.1016/j. molmed.2010.01.005
- Prevarskaya, N., Skryma, R., and Shuba, Y. (2018). Ion channels in cancer: are cancer hallmarks oncochannelopathies? *Physiol. Rev.* 98, 559–621. doi: 10.1152/physrev.00044.2016

- Putnam, L. R., Tsao, K., Morini, F., Lally, P. A., Miller, C. C., and Lally, K. P. (2016). Evaluation of variability in inhaled nitric oxide use and pulmonary hypertension in patients with congenital diaphragmatic hernia. *JAMA Pediatr.* 170, 1188–1194. doi: 10.1001/jamapediatrics.2016.2023
- Rao, V. R., Perez-Neut, M., Kaja, S., and Gentile, S. (2015). Voltage-gated ion channels in cancer cell proliferation. *Cancer* 7, 849–875. doi: 10.3390/ cancers7020813
- Richard, V., Nair, M. G., Santhosh Kumar, T. R., and Pillai, M. R. (2013). Side population cells as prototype of chemoresistant, tumor-initiating cells. *Biomed. Res. Int.* 2013:517237. doi: 10.1155/2013/517237
- Roura-Ferrer, M., Etxebarria, A., Solé, L., Oliveras, A., Comes, N., Villarroel, A., et al. (2009). Functional implications of KCNE subunit expression for the Kv7.5 (KCNQ5) channel. *Cell. Physiol. Biochem.* 24, 325–334. doi: 10.1159/ 000257425
- Roura-Ferrer, M., Solé, L., Oliveras, A., Dahan, R., Bielanska, J., Villarroel, A., et al. (2010). Impact of KCNE subunits on KCNQ1 (Kv7.1) channel membrane surface targeting. *J. Cell. Physiol.* 225, 692–700. doi: 10.1002/jcp.22265
- Roura-Ferrer, M., Solé, L., Oliveras, A., Villarroel, A., Comes, N., and Felipe, A. (2012). Targeting of Kv7.5 (KCNQ5)/KCNE channels to surface microdomains of cell membranes. *Muscle Nerve* 45, 48–54. doi: 10.1002/mus.22231
- Sabater-Lleal, M., Mälarstig, A., Folkersen, L., Soler Artigas, M., Baldassarre, D., Kavousi, M., et al. (2014). Common genetic determinants of lung function, subclinical atherosclerosis and risk of coronary artery disease. *PLoS One* 9:e104082. doi: 10.1371/journal.pone.0104082
- Salata, J. J., Jurkiewicz, N. K., Wang, J., Evans, B. E., Orme, H. T., and Sanguinetti, M. C. (1998). A novel benzodiazepine that activates cardiac slow delayed rectifier K<sup>+</sup> currents. *Mol. Pharmacol.* 54, 220–230. doi: 10.1124/ mol.54.1.220
- Salvi, S. (2014). Tobacco smoking and environmental risk factors for chronic obstructive pulmonary disease. *Clin. Chest Med.* 35, 17–27. doi: 10.1016/j. ccm.2013.09.011
- Sands, T. T., Miceli, F., Lesca, G., Beck, A. E., Sadleir, L. G., Arrington, D. K., et al. (2019). Autism and developmental disability caused by KCNQ3 gainof-function variants. *Ann. Neurol.* 86, 181–192. doi: 10.1002/ana.25522
- Schenzer, A., Friedrich, T., Pusch, M., Saftig, P., Jentsch, T. J., Grötzinger, J., et al. (2005). Molecular determinants of KCNQ (Kv7) K<sup>+</sup> channel sensitivity to the anticonvulsant retigabine. *J. Neurosci.* 25, 5051–5060. doi: 10.1523/ JNEUROSCI.0128-05.2005
- Schroeder, B. C., Waldegger, S., Fehr, S., Bleich, M., Warth, R., Greger, R., et al. (2000). A constitutively open potassium channel formed by KCNQ1 and KCNE3. *Nature* 403, 196–199. doi: 10.1038/35003200
- Schwab, A., Hanley, P., Fabian, A., and Stock, C. (2008). Potassium channels keep mobile cells on the go. *Physiology* 23, 212–220. doi: 10.1152/ physiol.00003.2008
- Schwiebert, E. M., Benos, D. J., Egan, M. E., Stutts, M. J., and Guggino, W. B. (1999). CFTR is a conductance regulator as well as a chloride channel. *Physiol. Rev.* 79, S145–S166. doi: 10.1152/physrev.1999.79.1.S145
- Sedivy, V., Joshi, S., Ghaly, Y., Mizera, R., Zaloudikova, M., Brennan, S., et al. (2015). Role of Kv7 channels in responses of the pulmonary circulation to hypoxia. Am. J. Phys. Lung Cell. Mol. Phys. 308, L48–L57. doi: 10.1152/ ajplung.00362.2013
- Serrano-Novillo, C., Capera, J., Colomer-Molera, M., Condom, E., Ferreres, J. C., and Felipe, A. (2019). Implication of voltage-gated potassium channels in neoplastic cell proliferation. *Cancer* 11:287. doi: 10.3390/cancers11030287
- Sevilla-Montero, J., Labrousse-Arias, D., Fernández-Pérez, C., Barreira, B., Mondejar-Parreño, G., Cogolludo, A., et al. (2019). Direct effects of cigarette smoke in pulmonary arterial cells alter vascular tone through arterial remodeling and Kv7.4 channel dysregulation. *BioRxiv* 555953. [Preprint]. doi: 10.1101/555953
- Shi, J., and Wei, L. (2013). Rho kinases in cardiovascular physiology and pathophysiology: the effect of fasudil. J. Cardiovasc. Pharmacol. 62, 341–354. doi: 10.1097/FJC.0b013e3182a3718f
- Shimokawa, H., and Satoh, K. (2015). 2015 ATVB plenary lecture: translational research on rho-kinase in cardiovascular medicine. *Arterioscler. Thromb. Vasc. Biol.* 35, 1756–1769. doi: 10.1161/ATVBAHA.115.305353
- Simonneau, G., Montani, D., Celermajer, D. S., Denton, C. P., Gatzoulis, M. A., Krowka, M., et al. (2019). Haemodynamic definitions and updated clinical classification of pulmonary hypertension. *Eur. Respir. J.* 53:1801913. doi: 10.1183/13993003.01913-2018

- Singh, A., Wu, H., Zhang, P., Happel, C., Ma, J., and Biswal, S. (2010). Expression of ABCG2 (BCRP) is regulated by Nrf2 in cancer cells that confers side population and chemoresistance phenotype. *Mol. Cancer Ther.* 9, 2365–2376. doi: 10.1158/1535-7163.MCT-10-0108
- Soler Artigas, M., Loth, D. W., Wain, L. V., Gharib, S. A., Obeidat, M., Tang, W., et al. (2011). Genome-wide association and large-scale follow up identifies 16 new loci influencing lung function. *Nat. Genet.* 43, 1082–1090. doi: 10.1038/ng.941
- Splawski, I., Shen, J., Timothy, K. W., Lehmann, M. H., Priori, S., Robinson, J. L., et al. (2000). Spectrum of mutations in long-QT syndrome genes. KVLQT1, HERG, SCN5A, KCNE1, and KCNE2. *Circulation* 102, 1178–1185. doi: 10.1161/01.CIR.102.10.1178
- Steinbichler, T. B., Dudás, J., Skvortsov, S., Ganswindt, U., Riechelmann, H., and Skvortsova, I.-I. (2018). Therapy resistance mediated by cancer stem cells. *Semin. Cancer Biol.* 53, 156–167. doi: 10.1016/j.semcancer.2018.11.006
- Stott, J. B., Barrese, V., Jepps, T. A., Leighton, E. V., and Greenwood, I. A. (2015). Contribution of Kv7 channels to natriuretic peptide mediated vasodilation in normal and hypertensive rats. *Hypertension* 65, 676–682. doi: 10.1161/HYPERTENSIONAHA.114.04373
- Stott, J. B., Jepps, T. A., and Greenwood, I. A. (2014). KV7 potassium channels: a new therapeutic target in smooth muscle disorders. *Drug Discov. Today* 19, 413–424. doi: 10.1016/j.drudis.2013.12.003
- Strutz-Seebohm, N., Seebohm, G., Fedorenko, O., Baltaev, R., Engel, J., Knirsch, M., et al. (2006). Functional coassembly of KCNQ4 with KCNE-beta-subunits in *Xenopus* oocytes. *Cell. Physiol. Biochem.* 18, 57–66. doi: 10.1159/000095158
- Sun, H., Lin, A.-H., Ru, F., Patil, M. J., Meeker, S., Lee, L.-Y., et al. (2019). KCNQ/M-channels regulate mouse vagal bronchopulmonary C-fiber excitability and cough sensitivity. JCI Insight 4:e124467. doi: 10.1172/jci.insight.124467
- Sun, J., and MacKinnon, R. (2020). Structural basis of human KCNQ1 modulation and gating. Cell 180, 340.e9–347.e9. doi: 10.1016/j.cell.2019.12.003
- Sylvester, J. T., Shimoda, L. A., Aaronson, P. I., and Ward, J. P. T. (2012). Hypoxic pulmonary vasoconstriction. *Physiol. Rev.* 92, 367–520. doi: 10.1152/ physrev.00041.2010
- Taylor, K. C., and Sanders, C. R. (2017). Regulation of KCNQ/Kv7 family voltage-gated K<sup>+</sup> channels by lipids. *Biochim. Biophys. Acta Biomembr.* 1859, 586–597. doi: 10.1016/j.bbamem.2016.10.023
- Tinel, N., Diochot, S., Lauritzen, I., Barhanin, J., Lazdunski, M., and Borsotto, M. (2000). M-type KCNQ2-KCNQ3 potassium channels are modulated by the KCNE2 subunit. *FEBS Lett.* 480, 137–141. doi: 10.1016/S0014-5793(00)01918-9
- Tobin, M. J. (2005). Update in pulmonary diseases. Ann. Intern. Med. 142, 283–288. doi: 10.7326/0003-4819-142-4-200502150-00010
- Trinh, N. T. N., Privé, A., Kheir, L., Bourret, J.-C., Hijazi, T., Amraei, M. G., et al. (2007). Involvement of KATP and KvLQT1 K<sup>+</sup> channels in EGFstimulated alveolar epithelial cell repair processes. Am. J. Phys. Lung Cell. Mol. Phys. 293, L870–L882. doi: 10.1152/ajplung.00362.2006
- Undem, B. J., and Kollarik, M. (2005). The role of vagal afferent nerves in chronic obstructive pulmonary disease. *Proc. Am. Thorac. Soc.* 2, 355–360. doi: 10.1513/pats.200504-033SR
- Van Horn, W. D., Vanoye, C. G., and Sanders, C. R. (2011). Working model for the structural basis for KCNE1 modulation of the KCNQ1 potassium channel. *Curr. Opin. Struct. Biol.* 21, 283–291. doi: 10.1016/j.sbi.2011.01.001
- Vanoye, C. G., Welch, R. C., Daniels, M. A., Manderfield, L. J., Tapper, A. R., and Sanders, C. R. (2009). Distinct subdomains of the KCNQ1 S6 segment determine channel modulation by different KCNE subunits. *J. Gen. Physiol.* 134, 207–217. doi: 10.1085/jgp.200910234
- Vigil, F. A., Bozdemir, E., Bugay, V., Chun, S. H., Hobbs, M., Sanchez, I., et al. (2020). Prevention of brain damage after traumatic brain injury by pharmacological enhancement of KCNQ (Kv7, «M-type») K<sup>+</sup> currents in neurons. *J. Cereb. Blood Flow Metab.* 40, 1256–1273. doi: 10.1177/0271678X19857818
- Wang, K., Zeng, J., Luo, L., Yang, J., Chen, J., Li, B., et al. (2013). Identification of a cancer stem cell-like side population in the HeLa human cervical carcinoma cell line. Oncol. Lett. 6, 1673–1680. doi: 10.3892/ol.2013.1607
- WHO Global Health Estimates (2020). WHO; World Health Organization. Available at: http://www.who.int/healthinfo/global\_burden\_disease/en/ (Accessed March 1, 2020).
- Wickenden, A. D., Krajewski, J. L., London, B., Wagoner, P. K., Wilson, W. A., Clark, S., et al. (2008). N-(6-chloro-pyridin-3-yl)-3,4-difluoro-benzamide (ICA-27243): a novel, selective KCNQ2/Q3 potassium channel activator. *Mol. Pharmacol.* 73, 977–986. doi: 10.1124/mol.107.043216

- Xiong, Q., Sun, H., Zhang, Y., Nan, F., and Li, M. (2008). Combinatorial augmentation of voltage-gated KCNQ potassium channels by chemical openers. *Proc. Natl. Acad. Sci. U. S. A.* 105, 3128–3133. doi: 10.1073/pnas.0712256105
- Yang, W. P., Levesque, P. C., Little, W. A., Conder, M. L., Ramakrishnan, P., Neubauer, M. G., et al. (1998). Functional expression of two KvLQT1-related potassium channels responsible for an inherited idiopathic epilepsy. J. Biol. Chem. 273, 19419–19423. doi: 10.1074/jbc.273.31.19419
- Zaydman, M. A., and Cui, J. (2014). PIP2 regulation of KCNQ channels: biophysical and molecular mechanisms for lipid modulation of voltagedependent gating. *Front. Physiol.* 5:195. doi: 10.3389/fphys.2014.00195
- Zhang, X., An, H., Li, J., Zhang, Y., Liu, Y., Jia, Z., et al. (2016). Selective activation of vascular Kv7.4/Kv7.5 K<sup>+</sup> channels by fasudil contributes to its vasorelaxant effect. Br. J. Pharmacol. 173, 3480–3491. doi: 10.1111/bph.13639
- Zhang, Y., and Wu, S. (2017). Effects of fasudil on pulmonary hypertension in clinical practice. *Pulm. Pharmacol. Ther.* 46, 54–63. doi: 10.1016/j. pupt.2017.08.002
- Zhou, L., Köhncke, C., Hu, Z., Roepke, T. K., and Abbott, G. W. (2019). The KCNE2 potassium channel  $\beta$  subunit is required for normal lung function

and resilience to ischemia and reperfusion injury. FASEB J. 33, 9762–9774. doi: 10.1096/fj.201802519R

Zimmer, J., Takahashi, T., Hofmann, A. D., and Puri, P. (2017). Downregulation of KCNQ5 expression in the rat pulmonary vasculature of nitrofen-induced congenital diaphragmatic hernia. *J. Pediatr. Surg.* 52, 702–705. doi: 10.1016/j. jpedsurg.2017.01.016

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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