SUPPLEMENTARY INFORMATION

Novel Fidaxomicin Antibiotics through Site-Selective Catalysis

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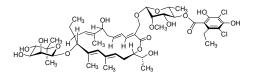
Supplementary Methods

General Methods and Materials

Unless otherwise stated, all chemicals were of reagent grade and purchased from Sigma-Aldrich, Merck, Fluorochem or Honeywell. Fidaxomicin was either obtained by fermentation of Actinoplanes deccanensis (ATCC 21983) or purchased from commercial suppliers (BOC Sciences). Reactions were carried out under protecting gas (N2 or Ar) and, unless otherweise stated, monitored for completion by UHPLC-MS (ESI). Solvents for reactions were of p.a. grade. Evaporation of solvents in vacuo was carried out on a rotary evaporator at 40 °C bath temperature and appropriate pressure. Ultra high performance liquid chromatography coupled to mass spectrometry (UHPLC-MS): Ultimate 3000 LC instrument (Thermo Fisher Scientific) coupled to a triple guadrupole Quantum Ultra EMR MS (Thermo Fisher Scientific) using a reversed-phase column (Kinetex® EVO C18; 1.7 µm; 100 Å, 50 x 2.1 mm; Phenomenex). The LC was equipped with an HPG-3400RS pump, a WPS-3000TRS autosampler, a TCC-3000RS column oven and a Vanquish DAD detector (all Thermo Fisher Scientific). The following solvents were applied: H₂O + 0.1% HCOOH (A), MeCN + 0.1% HCOOH (B). Samples were prepared using HPLC grade solvents (MeCN, MeOH, H2O) and filtered over a 4 mm syringe filter, PTFE (hydrophilic), pore size: 0.22 µm obtained from BGB Analytik AG. The MS was equipped with an H-ESI II ion source. The source temperature was 250 °C, the capillary temperature 270 °C and capillary voltage 3500 V, and datasets were acquired at resolution 0.7 on Q3 in centroid mode. High performance liquid chromatography (HPLC): All samples were pre-purified with a Discovery® DSC-18 SPE cartridge. Prominence modular HPLC instrument (Shimadzu) coupled to an SPD-20A UV/Vis detector (Shimadzu) using a reversed-phase column (Gemini NX C18, 3 µm, 10 Å, 150 mm x 4.6 mm) for analytical HPLC, and a reversed-phase column (Gemini NX C18, 5 μm, 110 Å, 250 mm × 21.2 mm; for preparative HPLC. The LC was equipped with a CBM-20A system controller, LC-20A solvent delivery unit, a DGU-20A degassing unit, FRC-10A fraction collector (all Shimadzu). The following solvents were used: H₂O + 0.1% HCOOH (A), MeCN + 0.1% HCOOH (B). Specific optical rotation $[\alpha]_D^T$: Jasco P-2000 Polarimeter; measured at the indicated temperature T. All given values for $[\alpha]_D^T$ have the dimension ° mL dm⁻¹ g⁻¹. Infrared spectra (IR): SpectrumTwo FT-IR Spectrometer (Perkin-Elmer) equipped with a Specac Golden Gate™ ATR (attenuated total reflection) accessory; applied as neat samples or as films; 1/λ in cm⁻¹. Nuclear magnetic resonance spectra (NMR): ¹H NMR spectra were recorded in CDCl₃ or acetone- d_6 on the instruments AV-500 (500 MHz) or AV-400 (400 MHz); chemical shift δ in ppm relative to solvent signals ($\delta = 7.26$ ppm for CDCl₃, 2.05 ppm for acetone- d_6 , 3.31 ppm for CD₃OD), ¹ coupling constant J is given in Hz. ¹³C NMR spectra were recorded in CDCl₃ or acetone- d_6 on the instruments Bruker AV-500 (125 MHz) or AV-400 (100 MHz); chemical shift in ppm relative to solvent signals (δ = 77.16 ppm for CDCl₃, 29.84 ppm for acetone- d_6 , 49.00 ppm for CD₃OD). High-resolution electrospray ionization mass spectra (HRMS): On flow injection: High-resolution mass spectra were acquired on a Qexactive instrument (ThermoFisher Scientific, Bremen, Germany) equipped with a heated electrospray (ESI) ionization source and connected to a Dionex Ultimate 3000 UHPLC system (ThermoFischer Scientifics, Germering, Germany). The samples were dissolved in MeOH, MeOH/CH₂Cl₂ 3:1, MeOH/H₂O 1:1, DMSO/H₂O 1:10 or H₂O at a concentration of ca. 50 μg mL⁻¹ thereof

1 μL was injected on-flow with a XRS auto-sampler (CTC, Zwingen, Switzerland). The mobile phase (120 µL mL⁻¹ flow rate) consisting of MeOH + 0.1% HCOOH or MeCN/H₂O 2:8 + 0.1% HCOOH was chosen according to the solubility. Ion source parameters were set as follow: spray voltage 3.0 kV; capillary temperature 280 °C; sheath gas 30 L min⁻¹; aux gas 8; 30 L min⁻¹; s-lens RF level 55.0; and aux gas temperature 250 °C. Full scan MS were acquired in the alternating (+)/(–)-ESI mode and over the ranges m/z = 80-1'200, 133-2'000, or 200-3'000 at 70'000 resolution (full width half-maximum) and with automatic gain control (AGC) target of 3.00E+06. The maximum allowed ion transfer time (IT) was 30 ms. Masses were calibrated below 2 ppm accuracy between m/z = 130.06619 and 1621.96509 in the positive and between 265.14790 and 1779.96528 in the negative ESI mode using the Pierce® ESI calibration solutions (ThermoFisher Scientific, Rockford, USA). Additionally, contaminations of erucamide (m/z = 338.34174, (+)-ESI) and palmitic acid (m/z = 255.23295, (–)-ESI) were used as lock masses in (+)-and (-)-ESI, respectively. LC-MS: Samples (1 µL injection) were analyzed with a Dionex Ultimate 3000UHPLC system (ThermoFischer Scientifics, Germering, Germany) connected to an Acquity ex detector and a Qexactive high-resolution mass spectrometer (ThermoFisher Scientific, Bremen, Germany) equipped with a heated electrospray (ESI) ionization source. Separation was performed with an AcquityBEH C18 HPLC column (1.7 µm particle size, 2x100 mm, Waters) kept at 30 °C. The mobile phase was consisting of A: H₂O + 0.1% HCOOH and B: CH₃CN + 0.1% HCOOH. A linear gradient was run from 5 to 98% B within 5 min followed by flushing with 98% B for 1 min at 400 μL min⁻¹ flow rate. UV spectra were recorded between 200 and 600 nm at 1.2 nm resolution and 20 points s⁻¹. MS ion source parameters were set as follows: spray voltage, 3.5 kV; capillary temperature, 260 °C; sheath gas 45 L min⁻¹; aux gas 15 L min⁻¹; sweep gas 2 L min⁻¹; s-lens RF level 450.0, and aux gas temperature 250 °C. Full scan MS were acquired in the (+)-ESI mode and over the ranges m/z = 80-1'200, 133-2'000, or 200-3'000 at 70'000 resolution (full width half-maximum) and with automatic gain control (AGC) target of 3.00E+06. The maximum allowed ion transfer time (IT) was 30 ms. Masses were calibrated below 2 ppm accuracy between m/z = 130.06619 and 1621.96509 in the positive and between 265.14790 and 1779.96528 in the negative ESI mode using the Pierce® ESI calibration solutions (*ThermoFisher Scientific*, Rockford, USA). Additionally, contaminations of erucamide (m/z =338.34174, (+)-ESI) and palmitic acid (m/z = 255.23295, (-)-ESI) were used as lock masses in (+)-and (–)-ESI, respectively.

Experimental Procedures C3"-Functionalizations OP-1118 (4)



Fidaxomicin (1) (1.50 g, 1.42 mmol, 1.0 eq.) was dissolved in MeOH (15 mL) and K_2CO_3 (393 mg, 2.84 mmol, 2.0 eq.) was added. The reaction mixture was stirred at room temperature for 2 h. The reaction mixture was quenched with aq. sat.

NH₄Cl. The aqueous phase was extracted with EtOAc (3x). The combined organic layers were washed with aq. sat. NaCl, dried over MgSO₄ and the solvent was evaporated under reduced pressure. The crude was purified by RP-HPLC [Gemini NX C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH; 20 mL/min; LC time program (min –% B): 0.0 min – 36%, 15.0 min – 36%, 100 min – 41%, 110 min – 100%] to afford, after lyophilization, **OP-1118** (4) (t_R = 31.0 min, 995 mg, 1.01 mmol, 71%) as a colorless solid.

Specific Rotation [α] $_{D}^{25\,^{\circ}C}$ = -23.0 (c = 0.81, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3430, 2975, 2936, 1697, 1591, 1378, 1311, 1242, 1068, 1024 cm $^{-1}$; ¹**H NMR** (500 MHz, acetone- d_{6}) δ 7.15 (d, J = 11.4 Hz, 1H), 6.59 – 6.47 (m, 1H), 5.87 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.73 (s, 1H), 5.53 (t, J = 7.4 Hz, 1H), 5.12 (dt, J = 10.6, 1.6 Hz, 1H), 5.01 (t, J = 9.7 Hz, 1H), 4.64 (dt, J = 6.7, 4.9 Hz, 1H), 4.61 (d, J = 1.3 Hz, 1H), 4.59 (s, 1H), 4.51 (d, J = 11.4 Hz, 1H), 4.33 (d, J = 11.5 Hz, 1H), 4.17 (m, 1H), 3.94 (quint, J = 6.3 Hz, 1H), 3.79 (dd, J = 2.9, 1.3 Hz, 1H), 3.71 (dd, J = 9.9, 3.4 Hz, 1H), 3.62 (d, J = 9.7 Hz, 1H), 3.56 – 3.47 (m, 2H), 3.42 (s, 3H), 3.41 – 3.38 (m, 2H), 2.91 (qd, J = 7.4, 2.3 Hz, 2H), 2.70 – 2.48 (m, 3H), 2.45 – 2.28 (m, 2H), 1.88 – 1.78 (m, 1H), 1.71 (d, J = 1.3 Hz, 3H), 1.64 (d, J = 1.4 Hz, 3H), 1.56 (s, 3H), 1.21 (d, J = 6.1 Hz, 3H), 1.19 – 1.15 (m, 1H), 1.14 – 1.07 (m, 9H), 0.98 (s, 3H), 0.73 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_{6}) δ 169.5, 167.8, 155.9, 153.8, 145.5, 143.5, 142.7, 136.8, 136.1, 136.1, 133.7, 128.1, 126.2, 125.3, 124.1, 114.5, 110.6, 108.2, 101.8, 96.8, 92.9, 81.6, 78.2, 77.6, 75.0, 74.8, 72.9, 72.7, 72.3, 72.2, 70.6, 67.7, 63.4, 61.7, 42.1, 37.3, 29.0, 28.4, 26.5, 26.2, 20.7, 18.2, 17.6, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH), calculated for C₄₈H₇₂Cl₂O₁₇N [M+NH₄]*: 1004.41718, found: 1004.41728.

BisallyI-OP-1118 (5)

A flame-dired two-necked flask under an inert atmosphere was charged with OP-1118 (4) (978 mg, 0.989 mmol, 1.0 eq.) and K_2CO_3 (410 mg, 2.97 mmol, 3.0 eq.). The solids were dissolved in DMF (20 mL) and allyl bromide (215 μ L, 2.47 mmol, 2.5 eq.) was added at room temperature. The

reaction mixture was stirred at 50 °C until full conversion was observed (UHPLC-MS). If required, additional K_2CO_3 and allyl bromide were added to reach full conversion. The reaction mixture was diluted with EtOAc and quenched with aq. sat. NH₄Cl. The aqueous phase was extracted with EtOAc (3x). The combined organic layers were washed with water (5x), dried over MgSO₄ and the solvent was evaporated under reduced pressure to afford **bisallyl-OP-1118** (5) (1.06 g, 0.989 mmol, quant.) as a colorless solid. The crude was used without further purification in the next step.

Specific Rotation [α]_D²⁵ °C = -43.64 (c = 0.71, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3454, 3083, 2975, 2935, 2877, 1735, 1700, 1642, 1568, 1455, 1402, 1379, 1352, 1314, 1277, 1248, 1198, 1176, 1161, 1132, 1068, 1024 cm⁻¹; ¹**H NMR** (400 MHz, acetone- d_6) δ 7.22 (d, J = 11.4 Hz, 1H), 6.67 – 6.57 (m, 1H), 6.23 – 6.02 (m, 2H), 5.95 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.82 (s, 1H), 5.62 (t, J = 8.2 Hz, 1H), 5.49 – 5.38 (m, 2H), 5.31 – 5.23 (m, 2H), 5.20 (d, J = 10.7 Hz, 1H), 5.02 (t, J = 9.7 Hz, 1H), 4.73 (q, J = 5.4 Hz, 1H), 4.69 (s, 1H), 4.65 – 4.49 (m, 6H), 4.40 (d, J = 11.5 Hz, 1H), 4.28 – 4.23 (m, 1H), 4.08 – 3.99 (m, 2H), 3.92 – 3.84 (m, 2H), 3.75 – 3.66 (m, 4H), 3.56 – 3.47 (m, 8H), 3.41 (d, J = 3.8 Hz, 1H), 3.33 (d, J = 6.7 Hz, 1H), 2.87 – 2.79 (m, 2H), 2.76 – 2.57 (m, 3H), 2.54 – 2.37 (m, 2H), 1.97 – 1.87 (m, 1H), 1.80 (s, 3H), 1.73 (s, 3H), 1.65 (s, 3H), 1.33 (d, J = 6.1 Hz, 3H), 1.29 – 1.23 (m, 9H), 1.22 – 1.13 (m, 11H), 1.07 (s, 3H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 167.9, 166.5, 153.9, 152.0, 145.3, 143.3, 140.0, 136.8, 136.1, 136.1, 134.1, 134.1, 133.7, 128.7, 128.2, 126.3, 126.0, 125.4, 124.1, 122.3, 118.8, 118.7, 101.9, 96.8, 92.9, 81.9, 78.3, 77.5, 76.4, 75.0, 74.8, 72.9, 72.7, 72.4, 72.3, 70.7, 67.8, 63.4, 61.7, 42.1, 37.3, 29.0, 28.4, 26.5, 25.6, 20.7, 18.3, 17.6, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+) (MeOH), calculated for C₅₄H₇₆Cl₂O₁₇Na [M+Na]*: 1089.43518, found: 1089.43487.

7a

Benzoyl chloride (9.51 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of bisallyl-OP-1118 (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid^[1] (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in

1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 10 min – 45%, 40 min – 65%, 50 min – 100%] to afford, after lyophilization, 7a (t_R = 32.5 min, 17.0 mg, 15.6 μ mol, 67%) as a slightly yellow solid.

Specific Rotation $[α]_D^{25\,°C} = -20.86$ (c = 0.33, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3441, 2975, 2932, 1699, 1587, 1451, 1376, 1314, 1279, 1246, 1178, 1114, 1066, 1024, 902, 809, 757, 720 cm⁻¹; ¹**H NMR** (500 MHz, acetone- $d_θ$) δ 8.11 – 8.07 (m, 2H), 7.66 – 7.61 (m, 1H), 7.54 – 7.49 (m, 2H), 7.25 (d, J = 11.4 Hz, 1H), 6.64 (dd, J = 15.3, 12.0 Hz, 1H), 6.01 – 5.94 (m, 1H), 5.91 (s, 1H), 5.68 (t, J = 8.1 Hz, 1H), 5.23 (dt, J = 10.5, 1.6 Hz, 1H), 5.11 (t, J = 9.7 Hz, 1H), 5.04 (dd, J = 10.2, 3.1 Hz, 1H), 4.88 (d, J = 1.2 Hz, 1H), 4.80 – 4.72 (m, 1H), 4.69 (d, J = 0.9 Hz, 1H), 4.61 (d, J = 11.5 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.18 (dd, J = 3.2, 1.2 Hz, 1H), 4.10 – 4.02 (m, 1H), 4.00 (d, J = 10.3 Hz, 1H), 3.81 (dt, J = 9.9, 1.8 Hz, 1H), 3.67 – 3.58 (m, 2H), 3.52 (s, 3H), 3.01 (qd, J = 7.4, 1.5 Hz, 2H), 2.82 – 2.60 (m, 3H), 2.54 – 2.43 (m, 2H), 1.97 – 1.88 (m, 1H), 1.84 (d, J = 1.3 Hz, 3H), 1.76 (d, J = 1.4 Hz, 3H), 1.66 (dd, J = 1.4, 0.7 Hz, 3H), 1.33 – 1.18 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- $d_θ$) δ 168.6, 167.0, 165.8, 155.0, 153.0, 144.6, 143.5, 142.6, 141.8, 136.0, 135.3, 135.0, 133.1, 129.7, 128.9, 128.0, 127.3, 125.5, 124.4, 123.1, 113.7, 109.7, 107.3, 100.9, 95.4, 92.2, 80.8, 77.3, 76.7, 74.9, 74.4, 72.0, 71.4, 70.6, 69.9, 69.7, 66.9, 62.5, 60.8, 41.1, 36.4, 28.1, 27.5, 25.6, 25.3, 20.7, 19.8, 17.3, 16.9, 16.7, 14.3, 13.5, 12.9 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₅H₇₂O₁₈Cl₂Na [M+Na]*: 1113.39934, found: 1113.39856.

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^[1] The boronic acid catalyst was obtained according to the procedure of Shimada and coworkers.8

7b

4-Methylbenzoyl chloride (10.8 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L,

93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 40 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, **7b** (t_R = 34.3 min, 17.5 mg, 15.8 μ mol, 68%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°} c = -16.0 (c = 0.25, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3466, 2975, 2934, 2173, 2031, 1983, 1697, 1612, 1590, 1408, 1378, 1311, 1278, 1243, 1179, 1111, 1066, 1021, 900, 755, 664, 585, 541, 518 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.97 (d, J = 8.2 Hz, 2H), 7.31 (d, J = 8.0 Hz, 2H), 7.25 (d, J = 11.4 Hz, 1H), 6.69 – 6.59 (m, 1H), 5.98 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.91 (s, 1H), 5.68 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.11 (t, J = 9.7 Hz, 1H), 5.02 (dd, J = 10.2, 3.2 Hz, 1H), 4.86 (s, 1H), 4.79 – 4.72 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.17 (s, 1H), 4.10 – 4.00 (m, 1H), 3.99 (d, J = 10.2 Hz, 1H), 3.86 – 3.76 (m, 2H), 3.66 – 3.52 (m, 2H), 3.52 (s, 3H), 3.05 – 2.95 (m, 2H), 2.83 – 2.60 (m, 3H), 2.55 – 2.42 (m, 2H), 2.41 (s, 3H), 1.98 – 1.88 (m, 1H), 1.83 (s, 3H), 1.76 (s, 3H), 1.66 (s, 3H), 1.33 – 1.18 (m, 16H), 0.82 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 166.7, 155.9, 153.9, 145.4, 144.4, 143.4, 142.7, 136.9, 136.1, 135.9, 133.9, 130.6, 129.8, 128.9, 128.2, 126.4, 125.3, 124.0, 114.6, 110.6, 108.2, 101.8, 96.3, 93.1, 81.6, 78.2, 77.6, 75.8, 75.3, 72.9, 72.3, 71.5, 70.8, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.2, 21.6, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₆H₇₄O₁₈Cl₂Na [M+Na]⁺: 1127.41499, found: 1127.41495.

7c

2-Fluorobenzoyl chloride (9.69 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient

atmosphere. The reaction mixture was stirred at 30 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, **7c** (t_R = 27.6 min, 11.3 mg, 10.2 μ mol, 44%) as a slightly yellow solid.]

Specific Rotation $[\alpha]_0^{25}$ ° $^c = -25.47$ (c = 0.29, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3457, 2976, 2934, 2108, 1701, 1613, 1588, 1489, 1456, 1377, 1307, 1244, 1160, 1125, 1066, 1022, 901, 801, 757, 662, 525 cm⁻¹; ¹H **NMR** (500 MHz, acetone- d_6) δ 8.03 (td, J = 7.6, 1.8 Hz, 1H), 7.70 – 7.62 (m, 1H), 7.31 (t, J = 7.6 Hz, 1H), 7.30 - 7.22 (m, 2H), 6.64 (dd, J = 15.0, 11.5 Hz, 1H), 5.98 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.91 (s, 1H), 5.68 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 10.6 Hz, 1H), 5.11 (t, J = 9.7 Hz, 1H), 5.05 (dd, J = 10.3, 3.2 Hz, 1H), 4.88 (s, 1H), 4.79 - 4.72 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.19 (s, 1H), 4.09 - 4.00 (m, 1H), 3.97 (d, J = 10.2 Hz, 1H), 3.84 - 3.77 (m, 2H), 3.67 -3.58 (m, 2H), 3.52 (s, 3H), 3.05 - 2.96 (m, 2H), 2.83 - 2.60 (m, 3H), 2.56 - 2.42 (m, 2H), 1.98 - 1.88(m, 1H), 1.83 (s, 3H), 1.76 (s, 3H), 1.66 (s, 3H), 1.34 – 1.10 (m, 16H), 0.82 (t, J = 7.5 Hz, 3H) ppm; ¹³C **NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 163.98 (d, J = 3.3 Hz), 162.72 (d, J = 258.9 Hz), 156.0, 154.2, 145.4, 143.5, 142.6, 136.9, 136.1, 135.9, 135.55 (d, J = 8.7 Hz), 134.0, 133.0, 128.2, 126.4, 125.3, 124.96 (d, J = 4.1 Hz), 124.0, 120.18 (d, J = 9.9 Hz), 117.65 (d, J = 22.0 Hz), 114.7, 110.3, 108.2, 101.8, 96.2, 93.1, 81.7, 78.2, 77.6, 76.3, 75.3, 72.9, 72.3, 71.4, 70.7, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.3, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; ¹⁹F NMR (376.5 MHz, acetone- d_6) δ -111.4 ppm; **HRMS** ESI(+), (MeOH) calculated for $C_{55}H_{71}O_{18}CI_2FNa$ [M+Na]⁺: 1131.38992, found: 1131.38906.

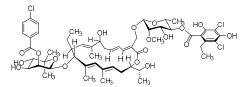
7d

4-Chlorobenzoyl chloride (10.5 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol,

4.0 eq.) in 1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, 7d (t_R = 39.6 min, 10.0 mg, 8.87 μ mol, 38%) as a slightly yellow solid and 7d-C2 (t_R = 36.4 min, 6.80 mg, 6.03 μ mol, 26%) as a slightly yellow solid.

Specific Rotation $[a]_D^{25\text{ °C}} = -24.38$ (c = 0.28, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3454, 2975, 2934, 1702, 1593, 1402, 1377, 1311, 1274, 1242, 1175, 1116, 1090, 1066, 1018, 900, 853, 759, 514 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 8.09 (d, J = 8.4 Hz, 2H), 7.56 (d, J = 8.4 Hz, 2H), 7.25 (d, J = 11.5 Hz, 1H), 6.68 – 6.59 (m, 1H), 5.97 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.91 (s, 1H), 5.67 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 5.04 (dd, J = 10.2, 3.1 Hz, 1H), 4.87 (s, 1H), 4.79 – 4.72 (m, 4H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.18 (s, 1H), 4.09 – 4.00 (m, 1H), 3.99 (d, J = 10.2 Hz, 1H), 3.84 – 3.78 (m, 2H), 3.67 – 3.57 (m, 2H), 3.52 (s, 3H), 3.05 – 2.96 (m, 2H), 2.83 – 2.63 (m, 3H), 2.58 – 2.42 (m, 2H), 1.98 – 1.87 (m, 1H), 1.83 (s, 3H), 1.76 (s, 3H), 1.66 (s, 3H), 1.36 – 1.18 (m, 16H), 0.81 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 165.8, 156.0, 154.2, 145.4, 143.4, 142.6, 139.6, 136.90, 136.1, 135.9, 134.0, 132.2, 130.4, 129.5, 129.5, 128.2, 126.4, 125.3, 124.0, 114.7, 108.2, 101.8, 96.2, 93.1, 81.6, 78.2, 77.6, 76.4, 75.3, 72.9, 72.3, 71.4, 70.7, 70.6, 67.7, 63.4, 61.7, 42.0, 37.3, 28.92, 28.4, 26.5, 26.3, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₅H₇₁O₁₈Cl₃Na [M+Na]⁺: 1147.36037, found: 1147.35900.

7d-C2



Specific Rotation [α]_D²⁵° c = -18.68 (c = 0.26, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3431, 2975, 2931, 2108, 1707, 1593, 1402, 1371, 1272, 1243, 1174, 1068, 1019, 900, 850, 756, 666, 527 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 8.08 (d, J = 8.6 Hz, 2H), 7.58

(d, J = 8.6 Hz, 2H), 7.22 (d, J = 11.4 Hz, 1H), 6.61 (dd, J = 15.0, 11.5 Hz, 1H), 5.93 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.86 (s, 1H), 5.65 (t, J = 8.3 Hz, 1H), 5.61 (dd, J = 3.3, 1.2 Hz, 1H), 5.18 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.95 (d, J = 1.3 Hz, 1H), 4.78 – 4.72 (m, 1H), 4.67 (s, 1H), 4.59 (d, J = 11.5 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.23 (s, 1H), 4.08 – 4.01 (m, 1H), 3.89 (dd, J = 10.0, 3.4 Hz, 1H), 3.80 (dd, J = 9.9, 3.3 Hz, 1H), 3.73 (d, J = 9.8 Hz, 1H), 3.66 (d, J = 9.9 Hz, 1H), 3.63 – 3.58 (m, 2H), 3.51 (s, 3H), 3.00 (q, J = 7.4 Hz, 2H), 2.80 – 2.74 (m, 1H), 2.71 – 2.63 (m, 1H), 2.52 – 2.41 (m, 3H), 1.81 – 1.76 (m, 1H), 1.75 (s, 3H), 1.73 (s, 3H), 1.56 (s, 3H), 1.32 (s, 3H), 1.30 (d, J = 6.2 Hz, 3H), 1.24 – 1.14 (m, 10H), 0.72 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone-d₆) δ 169.5, 167.8, 165.6, 156.0, 154.2, 145.4, 143.5, 142.6, 139.3, 136.8, 136.1, 135.8, 134.2, 132.2, 130.9, 129.5, 128.1, 126.5, 125.3, 124.0, 114.7, 110.3, 108.2, 101.7, 94.8, 92.2, 81.6, 78.2, 77.6, 75.42, 75.2, 74.6, 72.9, 72.3, 70.6, 70.5, 67.8, 63.4, 61.7, 41.9, 37.3, 29.2, 28.39, 26.3, 26.1, 20.7, 18.2, 17.7, 17.5, 15.1, 14.4, 13.7, 11.02 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₅H₇₁O₁₈Cl₃Na [M+Na]⁺: 1147.36037, found: 1147.36005.

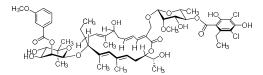
7e

3-Anisoyl chloride (11.5 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L,

93.6 µmol, 4.0 eq.) in 1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: $H_2O + 0.1\%$ HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, **7e** ($t_R = 30.8$ min, 10.2 mg, 9.09 µmol, 39%) as a slightly yellow solid and **7e-C2** ($t_R = 26.8$ min, 6.8 mg, 6.06 µmol, 26%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°C} = -27.89 (c = 0.38, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3450, 2974, 2934, 1697, 1588, 1454, 1375, 1280, 1237, 1142, 1110, 1065, 1022, 1004, 898, 800, 755, 683, 666 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.67 (d, J = 7.7 Hz, 1H), 7.63 – 7.58 (m, 1H), 7.42 (t, J = 8.0 Hz, 1H), 7.25 (d, J = 11.4 Hz, 1H), 7.23 – 7.16 (m, 1H), 6.68 – 6.59 (m, 1H), 5.98 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.91 (s, 1H), 5.68 (t, J = 8.1 Hz, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.11 (t, J = 9.7 Hz, 1H), 5.02 (dd, J = 10.2, 3.1 Hz, 1H), 4.87 (s, 1H), 4.79 – 4.72 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.19 (s, 1H), 4.10 – 4.00 (m, 1H), 4.00 (d, J = 10.2 Hz, 1H), 3.86 (s, 3H), 3.83 – 3.79 (m, 2H), 3.67 – 3.58 (m, 2H), 3.52 (s, 3H), 3.01 (qd, J = 7.4, 2.0 Hz, 2H), 2.84 – 2.75 (m, 1H), 2.74 – 2.61 (m, 2H), 2.56 – 2.43 (m, 2H), 1.98 – 1.89 (m, 1H), 1.84 (d, J = 1.3 Hz, 3H), 1.76 (s, 3H), 1.66 (s, 3H), 1.34 – 1.17 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.6, 167.8, 166.5, 160.6, 156.1, 154.4, 145.5, 143.5, 142.6, 136.9, 136.1, 135.9, 133.9, 133.0, 130.3, 128.2, 126.4, 125.3, 124.0, 122.7, 119.5, 115.6, 114.8, 110.0, 108.2, 101.8, 96.2, 93.1, 81.7, 78.2, 77.6, 76.2, 75.3, 72.9, 72.3, 71.4, 70.8, 70.6, 67.8, 63.4, 61.7, 55.8, 42.0, 37.3, 28.9, 28.4, 26.5, 26.3, 20.7, 18.2, 17.8, 17.6, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₆H₇₄O₁₉Cl₂Na [M+Na]*: 1143.40991, found: 1143.40905.

7e-C2



Specific Rotation $[\alpha]_D^{25\ °c} = -56.30$ (c = 0.24, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3436, 2974, 2932, 1704, 1588, 1454, 1372, 1278, 1239, 1069, 1024, 900, 756 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.67 (d, J = 7.8 Hz, 1H), 7.60 (d, J = 3.2 Hz,

1H), 7.44 (t, J = 8.0 Hz, 1H), 7.22 (d, J = 11.5 Hz, 1H), 7.19 (dd, J = 8.0, 3.2 Hz, 1H), 6.67 – 6.57 (m, 1H), 5.93 (ddd, J = 14.5, 9.5, 4.5 Hz, 1H), 5.86 (s, 1H), 5.65 (t, J = 8.4 Hz, 1H), 5.60 (d, J = 3.3 Hz, 1H), 5.19 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.95 (s, 1H), 4.74 (q, J = 5.3 Hz, 1H), 4.68 (s, 1H), 4.59 (d, J = 11.5 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.23 (s, 1H), 4.06 – 4.03 (m, 1H), 3.91 – 3.85 (m, 1H), 3.87 (s, 3H), 3.80 (dd, J = 9.7, 3.5 Hz, 1H), 3.76 – 3.71 (m, 1H), 3.67 (d, J = 9.9 Hz, 1H), 3.64 – 3.58 (m, 2H), 3.51 (s, 3H), 3.00 (q, J = 7.3 Hz, 2H), 2.80 – 2.74 (m, 1H), 2.72 – 2.65 (m, 1H), 2.54 – 2.40 (m, 3H), 1.86 – 1.78 (m, 1H), 1.75 (d, J = 1.3 Hz, 3H), 1.74 (s, 3H), 1.56 (s, 3H), 1.33 (s, 3H), 1.30 (d, J = 6.2 Hz, 3H), 1.24 – 1.15 (m, 10H), 0.73 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 166.3, 160.6, 155.9, 153.8, 145.5, 143.5, 142.7, 136.7, 136.1, 135.8, 134.1, 133.6, 130.3, 128.1, 126.4, 125.3, 124.1, 122.7, 119.2, 115.6, 114.5, 110.6, 108.2, 101.7, 95.0, 92.3, 81.6, 78.2, 77.6, 75.4, 75.3, 74.3, 72.9, 72.3, 70.6, 70.6, 67.8, 63.4, 61.7, 55.7, 41.9, 37.3, 29.2, 28.4, 26.2, 26.1, 20.7, 18.2, 17.7, 17.5, 15.1, 14.4, 13.7, 11.0 ppm; **HRMS** ESI(+), (MeOH) calculated for $C_{56}H_{74}O_{19}Cl_2Na$ [M+Na]*: 1143.40991, found: 1143.40961.

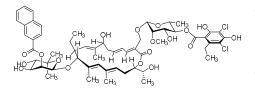
7f

2-Naphthoyl chloride (15.6 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L,

93.6 µmol, 4.0 eq.) in 1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 70%, 70 min – 100%] to afford, after lyophilization, **7f** (t_R = 37.9 min, 10.9 mg, 9.54 µmol, 41%) as a slightly yellow solid and **7f-C2** (t_R = 34.1 min, 7.90 mg, 6.92 µmol, 30%) as a slightly yellow solid.

Specific Rotation $[a]_D^{25\text{ °C}} = -24.18$ (c = 0.24, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3446, 2975, 2933, 1696, 1590, 1374, 1285, 1231, 1197, 1131, 1064, 1021, 903, 756, 666 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 8.73 (s, 1H), 8.13 (dd, J = 8.6, 1.5 Hz, 1H), 8.11 (d, J = 8.1 Hz, 1H), 8.02 (dd, J = 8.5, 2.8 Hz, 2H), 7.72-7.60 (m, 2H), 7.28 (d, J = 11.4 Hz, 1H), 6.70 – 6.61 (m, 1H), 6.00 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.94 (s, 1H), 5.71 (t, J = 8.1 Hz, 1H), 5.26 (d, J = 10.5 Hz, 1H), 5.13 (ddd, J = 9.8, 6.2, 3.1 Hz, 1H), 4.92 (s, 1H), 4.82-4.74 (m, 1H), 4.71 (s, 1H), 4.63 (d, J = 11.4 Hz, 1H), 4.45 (d, J = 11.4 Hz, 1H), 4.29 (s, 1H), 4.28 – 4.24 (m, 1H), 4.11 – 4.04 (m, 2H), 3.87 – 3.80 (m, 2H), 3.68 – 3.60 (m, 2H), 3.54 (s, 3H), 3.02 (q, J = 7.0 Hz, 3H), 2.85 – 2.78 (m, 1H), 2.76 – 2.62 (m, 2H), 2.56 – 2.40 (m, 2H), 2.00 – 1.91 (m, 1H), 1.87 (s, 3H), 1.78 (s, 3H), 1.68 (s, 3H), 1.35 – 1.19 (m, 16H), 0.84 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 166.8, 156.0, 154.2, 145.4, 143.4, 142.6, 136.9, 136.5, 136.2, 135.9, 134.0, 133.4, 131.8, 130.1, 129.2, 128.9, 128.9, 128.7, 128.2, 127.7, 126.4, 126.2, 125.3, 124.0, 114.7, 110.2, 108.2, 101.7, 96.3, 93.1, 81.7, 78.2, 77.6, 76.2, 75.3, 72.9, 72.3, 71.5, 70.9, 70.6, 67.7, 63.4, 61.7, 42.0, 37.3, 29.0, 28.4, 26.5, 26.3, 20.7, 18.2, 17.9, 17.6, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+), (MeOH) calculated for $C_{59}H_{74}O_{18}Cl_2Na$ [M+Na]*: 1163.41499, found: 1163.41462.

7f-C2



Specific Rotation [α]_D²⁵°^C = -58.61 (c = 0.24, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3435, 2975, 2932, 1704, 1590, 1371, 1284, 1232, 1197, 1069, 1023, 902, 778, 757, 666 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 8.69 (d, J = 1.5 Hz, 1H), 8.13 (dd, J = 8.6, 1.7 Hz, 1H), 8.10 (d, J = 8.1 Hz, 1H), 8.06 – 7.97 (m, 2H), 7.68 – 7.60

(m, 2H), 7.22 (d, J = 11.4 Hz, 1H), 6.67 – 6.50 (m, 1H), 5.92 (ddd, J = 14.7, 9.7, 4.7 Hz, 1H), 5.87 (s, 1H), 5.72 – 5.62 (m, 2H), 5.19 (d, J = 10.4 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.99 (d, J = 1.3 Hz, 1H), 4.75 (q, J = 5.4 Hz, 1H), 4.67 (s, 1H), 4.59 (d, J = 11.5 Hz, 1H), 4.41 (d, J = 11.5 Hz, 1H), 4.21 (s, 1H), 4.08 – 4.00 (m, 1H), 3.92 (dd, J = 10.0, 3.4 Hz, 1H), 3.83 – 3.74 (m, 3H), 3.65 – 3.57 (m, 2H), 3.51 (s, 3H), 3.00 (q, J = 7.4 Hz, 2H), 2.83 – 2.75 (m, 1H), 2.69 – 2.63 (m, 1H), 2.51 – 2.42 (m, 3H), 1.84 – 1.79 (m, 1H), 1.75 (d, J = 1.3 Hz, 3H), 1.73 (d, J = 1.4 Hz, 3H), 1.53 (d, J = 1.2 Hz, 3H), 1.38 (s, 3H), 1.30 (d, J = 6.1 Hz, 3H), 1.25 – 1.16 (m, 10H), 0.72 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone-d₆) δ 169.4, 167.8, 166.6, 155.9, 153.9, 145.5, 143.5, 142.7, 136.7, 136.4, 136.1, 135.8, 134.2, 133.5, 131.7, 130.1, 129.6, 129.1, 128.9, 128.7, 128.1, 127.6, 126.4, 126.4, 125.2, 124.1, 114.6, 110.5, 108.1, 101.7, 95.0, 92.2, 81.6, 78.2, 77.6, 75.4, 75.2, 74.4, 72.8, 72.3, 70.6, 70.5, 67.7, 63.4, 61.7, 41.9, 37.2, 28.4, 28.1, 26.3, 26.1, 20.6, 18.2, 17.7, 17.5, 15.1, 14.4, 13.7, 11.0 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₉H₇₄O₁₈Cl₂Na [M+Na]⁺: 1163.41499, found: 1163.41473.

7j

3-Furoyl chloride (15.5 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in

1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 40 °C for 2 h (*in this case, prolonged reaction time led to decomposition*) and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, **7j** (t_R = 20 min, 22.5 mg, 20.8 µmol, 89%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°C} = -23.68 (c = 0.27, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3468, 2975, 2935, 2317, 2223, 2207, 2174, 2105, 1981, 1963, 1703 1582, 1507, 1378, 1312, 1243, 1161, 1141, 1066, 1021, 899, 874, 800, 766, 601, 541, 525 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 8.24 – 8.20 (m, 1H), 7.66 (t, J = 1.8 Hz, 1H), 7.25 (d, J = 11.4 Hz, 1H), 6.80 (d, J = 1.8 Hz, 1H), 6.69 – 6.57 (m, 1H), 5.97 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 8.1 Hz, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.94 (dd, J = 10.3, 3.1 Hz, 1H), 4.84 (s, 1H), 4.77 – 4.72 (m, 1H), 4.68 (s, 1H), 4.61 (d, J = 11.5 Hz, 1H), 4.42 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.15 (d, J = 3.1 Hz, 1H), 4.08 – 3.98 (m, 1H), 3.91 (d, J = 10.2 Hz, 1H), 3.83 – 3.73 (m, 2H), 3.67 – 3.56 (m, 2H), 3.52 (s, 3H), 3.00 (qd, J = 7.4, 1.5 Hz, 2H), 2.82 – 2.74 (m, 1H), 2.73 – 2.60 (m, 2H), 2.52 – 2.43 (m, 2H), 1.98 – 1.86 (m, 1H), 1.83 (d, J = 1.3 Hz, 3H), 1.75 (d, J = 1.3 Hz, 3H), 1.65 (s, 3H), 1.43 – 1.14 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 163.3, 155.9, 154.0, 149.1, 145.4, 145.1, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 120.6, 114.6, 110.7, 110.5, 108.2, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.6, 75.2, 72.9, 72.3, 71.4, 70.8, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.2, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₃H₇₀O₁₉Cl₂Na [M+Na]*: 1103.37861, found: 1103.37844.

7k

3-Thiophenecarbonyl chloride (12.0 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L,

93.6 µmol, 4.0 eq.) in 1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 40 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, **7k** (t_R = 23.8 min, 22.7 mg, 19.3 µmol, 82%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°} = -36.14 (c = 0.28, CHCl₃); **FT-IR** $\tilde{\nu}$ (film 3440, 2973, 2931, 1695, 1589, 1522, 1408, 1379, 1311, 1245, 1197, 1162, 1142, 1109, 1064, 1021, 1003, 986, 899, 873, 800, 752, 666, 626 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 8.32 – 8.27 (m, 1H), 7.57 – 7.52 (m, 2H), 7.25 (d, J = 11.5 Hz, 1H), 6.69 – 6.59 (m, 1H), 5.98 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.91 (s, 1H), 5.72 – 5.64 (m, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.11 (t, J = 9.7 Hz, 1H), 4.97 (dd, J = 10.3, 3.1 Hz, 1H), 4.86 (s, 1H), 4.78 – 4.72 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.5 Hz, 1H), 4.28 – 4.25 (m, 1H), 4.18 – 4.16 (m, 1H), 4.08 – 4.02 (m, 1H), 3.95 (d, J = 10.2 Hz, 1H), 3.84 – 3.78 (m, 2H), 3.65 – 3.58 (m, 2H), 3.52 (s, 3H), 3.01 (qd, J = 7.4, 1.8 Hz, 2H), 2.80 – 2.61 (m, 3H), 2.55 – 2.42 (m, 2H), 1.98 – 1.89 (m, 1H), 1.83 (s, 3H), 1.75 (s, 3H), 1.66 (s, 3H), 1.37 – 1.14 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 162.9, 156.0, 154.1, 145.4, 143.4, 142.7, 136.9, 136.14, 135.9, 135.0, 133.9, 133.9, 128.8, 128.2, 127.2, 126.4, 125.3, 124.0, 114.6, 110.4, 108.2, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.8, 75.3, 72.9, 72.3, 71.4, 70.8, 70.6, 67.7, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.3, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₆₃H₇₀O₁₈Cl₂NaS [M+Na]⁺: 1119.35576, found: 1119.35543.

71

2-Furoyl chloride (8.10 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in

1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min to 60 min – 50%, 70 min – 100%] to afford, after lyophilization, **7I** (t_R = 20 min, 17.3 mg, 16.0 μ mol, 68%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°C} = -2.64 (c = 0.34, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3458, 2932, 1702, 1379, 1305, 1242,1119, 1066, 1021, 761 cm⁻¹; ¹**H NMR** (400 MHz, acetone- d_6) δ 7.83 – 7.78 (m, 1H), 7.30 – 7.21 (m, 2H), 6.69 – 6.58 (m, 2H), 5.97 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 7.9 Hz, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.99 (dd, J = 10.2, 3.0 Hz, 1H), 4.86 (s, 1H), 4.75 (q, J = 5.0 Hz, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.5 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.15 (s, 1H), 4.09 – 4.00 (m, 1H), 3.93 (d, J = 10.1 Hz, 1H), 3.81 (m, 2H), 3.67 – 3.57 (m, 2H), 3.52 (s, 3H), 3.01 (q, J = 7.3 Hz, 2H), 2.81 – 2.60 (m, 3H), 2.54 – 2.42 (m, 2H), 1.97 – 1.87 (m, 1H), 1.84 – 1.80 (m, 3H), 1.75 (s, 3H), 1.66 (s, 3H), 1.35 – 1.14 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 158.8, 155.9, 153.9, 147.7, 145.9, 145.4, 143.4, 142.7, 136.9, 136.1, 135.9, 134.0, 128.2, 126.4, 125.3, 124.0, 118.8, 114.6, 112.7, 110.5, 108.2, 101.8, 96.2, 93.1, 81.6, 78.2, 77.6, 75.9, 75.3, 72.9, 72.3, 71.4, 70.8, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.2, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₃H₇₀O₁₉Cl₂Na [M+Na]⁺: 1103.37861, found: 1103.37839.

7m

2-Thiophenecarbonyl chloride (8.76 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L,

93.6 µmol, 4.0 eq.) in 1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 40 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min LC time program (min –% B): 12 min – 50%, 60 min – 70%, 70 min – 100%] to afford, after lyophilization, **7m** (t_R = 24.4 min, 15.0 mg, 12.7 µmol, 54%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25\,°c}$ = +10.87 (c = 0.25, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3460, 2975, 2934, 2034, 1981, 1694, 1589, 1524, 1416, 1372, 1246, 1142, 1093, 1067, 1022, 900, 755, 601, 524 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.87 – 7.80 (m, 2H), 7.25 (d, J = 11.5 Hz, 1H), 7.020 (dd, J = 5.0, 3.7 Hz, 1H), 6.69 – 6.57 (m, 1H), 5.97 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 10.5, 1H), 5.11 (t, J = 9.7 Hz, 1H), 4.99 (dd, J = 10.2, 3.1 Hz, 1H), 4.86 (d, J = 1.1 Hz, 1H), 4.75 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.5 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (m, 1H), 4.16 (m, 1H), 4.04 (m, 1H), 3.95 (m, 1H), 3.81 (m, 2H), 3.61 (m, 2H), 3.52 (s, 3H), 3.13 – 2.92 (m, 2H), 2.74 – 2.60 (m, 3H), 2.48 (m, 2H), 1.98 – 1.88 (m, 1H), 1.83 (d, J = 1.3 Hz, 3H), 1.75 (d, J = 1.3 Hz, 3H), 1.68 – 1.63 (m, 3H), 1.37 – 1.16 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 162.5, 156.0, 154.1, 145.4, 143.5, 142.6, 136.9, 136.1, 135.9, 135.1, 134.4, 134.0, 133.8, 128.7, 128.2, 126.4, 125.3, 124.0, 114.7, 110.3, 108.2, 101.8, 96.2, 93.1, 81.7, 78.2, 77.6, 76.23, 75.3, 72.9, 72.3, 71.4, 70.8, 70.7, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.3, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₃H₇₀O₁₈Cl₂NaS [M+Na]⁺: 1119.35576, found: 1119.35551.

7n

1-Methylpyrrole-2-carbonyl chloride (11.8 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient

atmosphere. The reaction mixture was stirred at 30 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, **7n** (t_R = 24.2 min, 17.5 mg, 14.9 μ mol, 64%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25\text{ °C}} = -25.00$ (c = 0.26, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3464, 2975, 2935, 2067, 2013, 1983, 1964, 1689, 1589, 1532, 1411, 1383, 1323, 1245, 1141, 1112, 1065, 1020, 898, 801, 755, 665, 601, 584, 540, 510 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.25 (d, J = 11.5 Hz, 1H), 7.00 – 6.92 (m, 2H), 6.63 (dd, J = 15.0, 11.5 Hz, 1H), 6.07 (dd, J = 4.0, 2.5 Hz, 1H), 5.97 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 8.1 Hz, 1H), 5.23 (d, J = 10.6 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.96 (dd, J = 10.3, 3.1 Hz, 1H), 4.84 (s, 1H), 4.79 – 4.72 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.5 Hz, 1H), 4.27 (s, 1H), 4.12 (s, 1H), 4.08 – 4.01 (m, 1H), 3.95 – 3.89 (m, 4H), 3.84 – 3.77 (m, 2H), 3.66 – 3.58 (m, 2H), 3.52 (s, 3H), 3.00 (q, J = 7.3 Hz, 2H), 2.75 – 2.60 (m, 3H), 2.57 – 2.41 (m, 2H), 1.95 – 1.90 (m, 1H), 1.83 (s, 3H), 1.75 (s, 3H), 1.66 (s, 3H), 1.38 – 1.15 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 161.5, 155.9, 153.8, 145.4, 143.4, 142.7, 136.9, 136.1, 136.0, 133.9, 130.5, 128.2, 126.4, 125.3, 124.0, 123.6, 118.9, 114.5, 110.6, 108.3, 108.2, 101.8, 96.3, 93.0, 81.7, 78.2, 77.6, 75.3, 74.7, 72.9, 72.3, 71.5, 71.0, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 37.1, 28.9, 28.4, 26.5, 26.2, 20.7, 18.23, 17.8, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₄Hr₃O₁₈Cl₂NNa [M+Na]⁺: 1116.41024, found: 1116.40971.

7p

2,5-Dimethylfuran-3-carbonyl chloride (10.9 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane

(125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 2 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, 7p (t_R = 30.4 min, 19.4 mg, 17.5 μ mol, 75%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25 \, \circ c}$ = -42.65 (c = 0.49, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3463, 2975, 1701, 1589, 1367, 1235, 1068, 1022 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.25 (d, J = 11.4 Hz, 1H), 6.63 (dd, J = 15.0, 11.5 Hz, 1H), 6.28 (s, 1H), 5.97 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 10.4 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.95 (dd, J = 10.3, 3.1 Hz, 1H), 4.83 (s, 1H), 4.79 – 4.71 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.10 (d, J = 3.0 Hz, 1H), 4.09 – 4.00 (m, 1H), 3.90 (d, J = 10.3 Hz, 1H), 3.85 – 3.76 (m, 2H), 3.66 – 3.57 (m, 2H), 3.52 (s, 3H), 3.06 – 2.95 (m, 2H), 2.81 – 2.74 (m, 1H), 2.75 – 2.60 (m, 2H), 2.52 (s, 3H), 2.52 – 2.41 (m, 2H), 2.23 (s, 3H), 1.98 – 1.86 (m, 1H), 1.83 (s, 3H), 1.75 (s, 3H), 1.66 (s, 3H), 1.34 – 1.16 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 164.2, 158.3, 156.0, 154.1, 150.8, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 115.2, 114.6, 110.4, 108.2, 107.3, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.3, 74.9, 72.9, 72.2, 71.5, 70.9, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.2, 20.7, 18.2, 17.80, 17.5, 15.2, 14.4, 13.8, 13.8, 13.7, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₅H₇₄Cl₂O₁₉Na [M+Na]*: 1131.40936, found: 1131.40922.

7q

5-tert-Butyl-2-methylfuran-3-carbonylchloride (16.4 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%)

and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 2 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, **7q** (t_R = 37.7 min, 18.5 mg, 16.1 μ mol, 69%) as a slightly yellow solid.

Specific Rotation $[a]_D^{25\text{ °C}} = -31.87$ (c = 0.46, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3428, 2971, 2934, 2874, 1704, 1611, 1584, 1402, 1365, 1312, 1233, 1208, 1177, 1143, 1118, 1067, 1022, 1004, 987 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.25 (d, J = 11.5 Hz, 1H), 6.63 (dd, J = 15.0, 11.5 Hz, 1H), 6.28 (s, 1H), 5.97 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.95 (dd, J = 10.3, 3.1 Hz, 1H), 4.83 (s, 1H), 4.79 – 4.71 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.42 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.11 (s, 1H), 4.11 – 4.03 (m, 1H), 3.86 – 3.75 (m, 3H), 3.65 – 3.58 (m, 2H), 3.52 (s, 3H), 3.00 (q, J = 7.3 Hz, 2H), 2.79 – 2.60 (m, 3H), 2.54 (s, 3H), 2.52 – 2.42 (m, 2H), 1.97 – 1.89 (m, 1H), 1.83 (d, J = 1.4 Hz, 3H), 1.75 (d, J = 1.3 Hz, 3H), 1.65 (s, 3H), 1.35 – 1.15 (m, 25H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.8, 164.4, 162.7, 158.2, 155.8, 153.8, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.7, 114.5, 110.7, 108.2, 103.9, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.3, 74.9, 72.9, 72.3, 71.5, 70.9, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 33.0, 29.1, 28.9, 28.4, 26.5, 26.2, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.9, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₈H₈₀Cl₂O₁₉Na [M+Na]⁺: 1173.45631, found: 1173.45632.

7r

2-Methyl-5-phenylfuran-3-carbonyl chloride (18.1 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane

(125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 2 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, 7r (t_R = 53.4 min, 21.5 mg, 18.3 μ mol, 78%) as a slightly yellow solid.

Specific Rotation $[\alpha]_D^{25}{}^{\circ}C = -25.00$ (c = 0.38, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3416, 2975, 2934, 2875, 1704, 1644, 1613, 1589, 1560, 1532, 1450, 1404, 1366, 1312, 1294, 1233, 1199, 1176, 1163, 1144, 1093 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.73 (d, J = 7.4 Hz, 2H), 7.44 (t, J = 7.8 Hz, 2H), 7.32 (t, J = 7.4 Hz, 1H), 7.25 (d, J = 11.4 Hz, 1H), 7.08 (s, 1H), 6.80 – 6.59 (m, 1H), 5.97 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.91 (s, 1H), 5.67 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 11.0 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 5.00 (dd, J = 10.3, 3.1 Hz, 1H), 4.86 (s, 1H), 4.81 – 4.72 (m, 1H), 4.69 (s, 1H), 4.61 (d, J = 11.4 Hz, 1H), 4.43 (d, J = 11.4 Hz, 1H), 4.27 (s, 1H), 4.15 (d, J = 2.7 Hz, 1H), 4.11 – 4.00 (m, 1H), 3.95 (d, J = 10.3 Hz, 1H), 3.86 – 3.78 (m, 2H), 3.69 – 3.57 (m, 2H), 3.52 (s, 3H), 3.04 – 2.96 (m, 2H), 2.83 – 2.62 (m, 3H), 2.67 (s, 3H), 2.56 – 2.42 (m, 2H), 1.96 – 1.90 (m, 1H), 1.83 (d, J = 1.3 Hz, 3H), 1.76 (s, 3H), 1.66 (s, 3H), 1.34 – 1.17 (m, 16H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 169.5, 167.9, 164.0, 159.6, 155.8, 153.9, 152.5, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 134.0, 130.9, 129.7, 128.6, 128.2, 126.4, 125.3, 124.3, 124.0, 116.6, 114.5, 110.7, 108.2, 106.8, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.3, 75.2, 72.9, 72.3, 71.5, 70.8, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.2, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 14.0, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₆₀H₇₆Cl₂O₁₉Na [M+Na]*: 1193.42501, found: 1193.42554.

7s

Acetyl chloride (5.84 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in

1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 10 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 45%, 60 min – 55%, 70 min – 100%] to afford, after lyophilization, **7s** (t_R = 18.1 min, 15.0 mg, 14.6 μ mol, 62%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°C} = -27.33 (c = 0.29, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3477, 2977, 2015, 1979, 1704, 1377, 1243, 1067, 772, 524 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.24 (dd, J = 11.4, 0.9 Hz, 1H), 6.70 – 6.59 (m, 1H), 5.96 (ddd, J = 14.5, 9.5, 4.6 Hz, 1H), 5.88 (s, 1H), 5.76 – 5.58 (m, 1H), 5.22 (d, J = 10.6 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.78 (d, J = 1.2 Hz, 1H), 4.77 – 4.73 (m, 2H), 4.68 (d, J = 0.9 Hz, 1H), 4.60 (d, J = 11.4 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.26 (s, 1H), 4.08 – 3.99 (m, 2H), 3.85 – 3.75 (m, 3H), 3.68 – 3.58 (m, 2H), 3.52 (s, 3H), 3.08 – 2.94 (m, 2H), 2.81 – 2.74 (m, 1H), 2.73 – 2.68 (m, 1H), 2.67 – 2.58 (m, 1H), 2.54 – 2.41 (m, 2H), 2.02 (s, 3H), 1.97 – 1.88 (m, 1H), 1.81 (d, J = 1.3 Hz, 3H), 1.74 (d, J = 1.5 Hz, 3H), 1.65 (d, J = 0.7 Hz, 3H), 1.31 (d, J = 6.2 Hz, 3H), 1.28 – 1.17 (m, 10H), 1.14 (s, 3H), 0.82 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6)) δ 171.0, 169.5, 167.8, 156.0, 154.2, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.7, 110.2, 108.3, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.2 (2C), 72.9, 72.3, 71.3, 70.7, 70.6, 67.8, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.2, 21.1, 20.7, 18.2, 17.7, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₀H₇₀O₁₈Cl₂Na [M+Na]⁺: 1051.38314, found: 1051.38344.

7t

Isovaleryl chloride (10.0 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient

atmosphere. The reaction mixture was stirred at 10 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 60 min – 50%, 70 min – 100%] to afford, after lyophilization, **7t** (t_R = 28.7 min, 20.0 mg, 17.4 μ mol, 74%) as a slightly yellow solid.

Specific Rotation $[a]_D^{25 \text{ °C}} = -26.42$ (c = 0.44, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3456, 2968, 2934, 2874, 1698, 1590, 1374, 1310, 1244, 1198, 1163, 1143, 1066, 1021, 899, 800, 758, 665, 600 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.26 (d, J = 11.5 Hz, 1H), 6.65 (dd, J = 15.0, 11.5 Hz, 1H), 5.98 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 8.3 Hz, 1H), 5.23 (d, J = 10.7 Hz, 1H), 5.12 (t, J = 9.7 Hz, 1H), 4.84 – 4.79 (m, 2H), 4.78 – 4.74 (m, 1H), 4.70 (s, 1H), 4.62 (d, J = 11.5 Hz, 1H), 4.44 (d, J = 11.4 Hz, 1H), 4.28 (s, 1H), 4.09 – 4.02 (m, 2H), 3.86 – 3.78 (m, 3H), 3.69 – 3.59 (m, 2H), 3.54 (s, 3H), 3.02 (q, J = 7.3 Hz, 2H), 2.82 – 2.76 (m, 1H), 2.75 – 2.69 (m, 1H), 2.68 – 2.62 (m, 1H), 2.56 – 2.43 (m, 2H), 2.23 (dd, J = 7.1, 2.1 Hz, 2H), 2.15 – 2.08 (m, 1H), 1.99 – 1.88 (m, 1H), 1.83 (s, 3H), 1.76 (s, 3H), 1.67 (s, 3H), 1.33 (d, J = 6.2 Hz, 3H), 1.27 – 1.19 (m, 10H), 1.16 (s, 3H), 0.97 (d, J = 6.6 Hz, 6H), 0.83 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 173.0, 169.5, 167.8, 155.9, 153.9, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.6, 110.5, 108.2, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.2, 74.8, 72.9, 72.3, 71.4, 70.8, 70.6, 67.7, 63.4, 61.7, 43.9, 42.1, 37.3, 28.9, 28.4, 26.5, 26.3, 26.2, 22.7 (2C), 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+), (MeOH) calculated for $C_{53}H_{76}O_{18}Cl_2Na$ [M+Na]*: 1093.43064, found: 1093.42969.

7u

Isobutyryl chloride (8.56 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (2.71 mg, 11.7 μ mol, 50 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-

dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 40%, 60 min – 60%, 61 min – 100%] to afford, after lyophilization, 7u (t_R = 35.4 min, 15.1 mg, 14.3 μ mol, 61%) as a slightly yellow solid.

Specific Rotation [a] $_D^{25\text{ °C}} = -35.55$ (c = 0.25, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3466, 2967, 2932, 1692, 1590, 1382, 1311, 1259, 1199, 1143, 1066, 1021, 898, 799, 758; cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.26 (d, J = 11.4 Hz, 1H), 6.65 (dd, J = 15.0, 11.5 Hz, 1H), 5.98 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.89 (s, 1H), 5.67 (t, J = 8.2 Hz, 1H), 5.23 (d, J = 10.5 Hz, 1H), 5.12 (t, J = 9.7 Hz, 1H), 4.85 – 4.73 (m, 3H), 4.70 (s, 1H), 4.62 (d, J = 11.4 Hz, 1H), 4.44 (d, J = 11.5 Hz, 1H), 4.28 (s, 1H), 4.10 – 3.97 (m, 2H), 3.91 – 3.77 (m, 3H), 3.71 – 3.60 (m, 2H), 3.54 (s, 3H), 3.09 – 2.96 (m, 2H), 2.83 – 2.76 (m, 1H), 2.75 – 2.69 (m, 1H), 2.68 – 2.64 (m, 1H), 2.63 – 2.57 (m, 1H), 2.55 – 2.44 (m, 2H), 1.97 – 1.89 (m, 1H), 1.83 (s, 3H), 1.76 (s, 3H), 1.67 (s, 3H), 1.33 (d, J = 6.2 Hz, 3H), 1.28 – 1.07 (m, 19H), 0.83 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 177.1, 169.5, 167.8, 155.9, 154.0, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.6, 110.5, 108.2, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.2, 74.8, 72.9, 72.3, 71.4, 70.7, 70.6, 67.7, 63.4, 61.7, 42.0, 37.3, 34.7, 28.9, 28.4, 26.5, 26.2, 20.7, 19.4, 19.2, 18.2, 17.7, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(-), (MeOH) calculated for C₅₂H₇₃Cl₂O₁₈ [M-H]⁻: 1055.41588, found: 1055.41794

7w

2-Ethylbutyryl chloride (11.2 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (2.71 mg, 11.7 μ mol, 50 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient atmosphere. The reaction

mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 45%, 60 min – 65%, 60.5 min – 100%] to afford, after lyophilization, **7w** (t_R = 36.0 min, 15.1 mg, 13.9 µmol, 59%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25\,°C}$ = -29.56 (c = 0.36, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3455, 2967, 2933, 1692, 1589, 1459, 1381, 1311, 1242, 1197, 1142, 1066, 1022, 898, 799, 755 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.12 (d, J = 11.5 Hz, 1H), 6.51 (dd, J = 15.0, 11.5 Hz, 1H), 5.84 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.76 (s, 1H), 5.53 (t, J = 8.3 Hz, 1H), 5.09 (d, J = 10.3 Hz, 1H), 4.98 (t, J = 9.7 Hz, 1H), 4.69 (dd, J = 10.4, 3.1 Hz, 1H), 4.67 (s, 1H), 4.67 – 4.59 (m, 1H), 4.56 (s, 1H), 4.48 (d, J = 11.5 Hz, 1H), 4.30 (d, J = 11.4 Hz, 1H), 4.14 (s, 1H), 3.94 – 3.89 (m, 2H), 3.73 – 3.61 (m, 3H), 3.56 – 3.45 (m, 2H), 3.40 (s, 3H), 2.88 (q, J = 7.2 Hz, 2H), 2.65 (m, 1H), 2.61 – 2.56 (m, 1H), 2.55 – 2.47 (m, 1H), 2.44 – 2.27 (m, 2H), 2.15 – 2.08 (m, 1H), 1.84 – 1.76 (m, 1H), 1.69 (s, 3H), 1.62 (s, 3H), 1.57 – 1.47 (m, 5H), 1.45 – 1.35 (m, 2H), 1.19 (d, J = 6.1 Hz, 3H), 1.13 – 1.05 (m, 10H), 1.03 (s, 3H), 0.83 – 0.74 (m, 6H), 0.69 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 176.1, 169.5, 167.8, 155.9, 154.0, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.6, 110.5, 108.2, 101.8, 96.2, 93.1, 81.6, 78.2, 77.6, 75.3, 74.6, 72.9, 72.3, 71.4, 70.9, 70.6, 67.8, 63.4, 61.7, 49.7, 42.1, 37.3, 28.9, 28.4, 26.5, 26.2, 25.8, 25.6, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 12.1, 12.1, 11.2 ppm; **HRMS** ESI(-), (MeOH) calculated for C₅₄H₇₇Cl₂O₁₈ [M-H]⁻: 1083.44924, found: 1083.44979.

7x

Cyclopropanecarbonyl chloride (7.45 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (1.36 mg, 5.85 μ mol, 25 mol%) and DIPEA (15.8 μ L, 93.6 μ mol,

4.0 eq.) in 1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: $H_2O + 0.1\%$ HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 40%, 60 min – 60%, 70 min – 100%] to afford, after lyophilization, $\mathbf{7x}$ ($t_R = 36.8$ min, 10.0 mg, 9.46 µmol, 41%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°C} = -36.59 (c = 0.32, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3462, 2976, 2934, 1981, 1701, 1590, 1381, 1312, 1243, 1204, 1067, 1022, 952, 900, 764, 520 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.24 (d, J = 11.4 Hz, 1H), 6.70 – 6.58 (m, 1H), 5.96 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.87 (s, 1H), 5.65 (t, J = 8.3 Hz, 1H), 5.22 (d, J = 10.6 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.81 – 4.72 (m, 3H), 4.68 (s, 1H), 4.60 (d, J = 11.4 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.26 (s, 1H), 4.07 – 3.97 (m, 2H), 3.86 – 3.74 (m, 3H), 3.65 – 3.56 (m, 2H), 3.52 (s, 3H), 3.07 – 2.93 (m, 2H), 2.80 – 2.61 (m, 3H), 2.56 – 2.42 (m, 2H), 1.95 – 1.87 (m, 1H), 1.81 (s, 3H), 1.74 (s, 3H), 1.65 (s, 3H), 1.67 – 1.59 (m, 1H), 1.31 (d, J = 6.1 Hz, 3H), 1.25 – 1.17 (m, 10H), 1.14 (s, 3H), 0.94 – 0.90 (m, 2H), 0.90 – 0.84 (m, 2H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 174.9, 169.5, 167.8, 155.9, 154.1, 145.4, 143.4, 142.7, 136.9, 136.1, 135.9, 133.9, 128.1, 126.3, 125.3, 124.0, 114.7, 110.3, 108.2, 101.8, 96.2, 93.0, 81.6, 78.2, 77.5, 75.2, 75.1, 72.8, 72.3, 71.3, 70.6 (2C), 67.7, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.3, 20.7, 18.2, 17.7, 17.5, 15.2, 14.4, 13.8, 13.5, 11.1, 8.5, 8.5 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₂H₇₂O₁₈Cl₂Na [M+Na]*: 1077.39934, found: 1077.39903.

7у

1-Methylcyclopropane-1-carbonyl chloride (9.71 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (2.71 mg, 11.7 μ mol, 50 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient

atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 45%, 60 min – 65%, 61 min – 100%] to afford, after lyophilization, **7y** (t_R = 29.7 min, 7.3 mg, 6.82 μ mol, 29%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25\text{ °C}} = -24.40$ (c = 0.25, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3355, 2964, 2925, 1635, 1456, 1381, 1312, 1247, 1176, 1066, 1026, 895, 798 cm⁻¹; ¹**H NMR** (400 MHz, acetone- d_6) δ 7.24 (d, J = 11.5 Hz, 1H), 6.77 – 6.52 (m, 1H), 5.96 (ddd, J = 14.6, 9.5, 4.6 Hz, 1H), 5.88 (s, 1H), 5.65 (t, J = 8.4 Hz, 1H), 5.22 (d, J = 10.6 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.76 (s, 1H), 4.75 – 4.71 (m, 2H), 4.68 (s, 1H), 4.60 (d, J = 11.4 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.27 (s, 1H), 4.07 – 4.02 (m, 1H), 3.99 (s, 1H), 3.84 – 3.73 (m, 3H), 3.69 – 3.58 (m, 2H), 3.52 (s, 3H), 3.01 (q, J = 7.4 Hz, 2H), 2.82 – 2.78 (m, 1H), 2.77 – 2.67 (m, 1H), 2.65 – 2.57 (d, J = 9.5 Hz, 1H), 2.53 – 2.40 (m, 2H), 1.97 – 1.87 (m, 1H), 1.81 (s, 3H), 1.74 (s, 3H), 1.65 (s, 3H), 1.32 – 1.17 (m, 18H), 1.13 (s, 3H), 0.82 (t, J = 7.4 Hz, 3H), 0.74 – 0.63 (m, 2H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 175.8, 169.5, 167.8, 156.0, 154.2, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.3, 125.3, 124.0, 114.7, 110.2, 108.2, 101.8, 96.1, 93.0, 81.7, 78.2, 77.6, 75.2, 75.1, 72.9, 72.3, 71.3, 70.7, 70.6, 67.7, 63.4, 61.7, 42.0, 37.3, 28.9, 28.4, 26.5, 26.3, 20.7, 19.6, 19.2, 18.2, 17.7, 17.5, 17.0, 16.7, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(-), (MeOH) calculated for $C_{53}H_{73}Cl_2O_{18}$ [M-H]: 1067.41794, found: 1067.41552.

7z

Cyclobutanecarbonyl chloride (9.34 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (1.36 mg, 5.85 μ mol, 25 mol%) and DIPEA (15.8 μ L, 93.6 μ mol,

4.0 eq.) in 1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 45%, 60 min – 65%, 60.5 min – 100%] to afford, after lyophilization, **7z** (t_R = 27.8 min, 15.8 mg, 14.8 μ mol, 63%) as a slightly yellow solid.

Specific Rotation [α] $_{D}^{25 \circ c} = -25.15$ (c = 0.34, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3439, 2975, 2936, 2874, 1698, 1643, 1589, 1404, 1379, 1312, 1248, 1198, 1162, 1144, 1111, 1066, 1023 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_{6}) δ 7.26 (d, J = 11.4 Hz, 1H), 6.65 (dd, J = 15.0, 11.5 Hz, 1H), 5.98 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.90 (s, 1H), 5.67 (t, J = 8.2 Hz, 1H), 5.24 (d, J = 10.5 Hz, 1H), 5.12 (t, J = 9.7 Hz, 1H), 4.87 – 4.74 (m, 3H), 4.70 (s, 1H), 4.62 (d, J = 11.4 Hz, 1H), 4.44 (d, J = 11.5 Hz, 1H), 4.28 (s, 1H), 4.15 – 3.98 (m, 2H), 3.86 – 3.78 (m, 3H), 3.68 – 3.60 (m, 2H), 3.54 (s, 3H), 3.20 (quint, J = 8.4 Hz, 1H), 3.02 (qd, J = 7.4, 2.1 Hz, 2H), 2.83 – 2.76 (m, 1H), 2.76 – 2.70 (m, 1H), 2.70 – 2.62 (m, 1H), 2.53 – 2.44 (m, 2H), 2.38 – 2.26 (m, 2H), 2.25 – 2.15 (m, 2H), 2.05 – 1.86 (m, 3H), 1.86 – 1.82 (m, 3H), 1.78 – 1.71 (m, 3H), 1.67 (s, 3H), 1.33 (d, J = 6.2 Hz, 3H), 1.29 – 1.19 (m, 10H), 1.16 (s, 3H), 0.83 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_{6}) δ 175.4, 169.6, 167.8, 156.1, 154.4, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.8, 110.0, 108.2, 101.8, 96.2, 93.0, 81.7, 78.2, 77.5, 75.2, 74.9, 72.9, 72.3, 71.4, 70.7, 70.6, 67.7, 63.4, 61.7, 42.0, 39.0, 37.3, 28.9, 28.4, 26.5, 26.3, 25.9, 25.7, 20.7, 18.9, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(-), (MeOH) calculated for C₅₃H₇₃Cl₂O₁₈ [M-H]: 1067.41554, found: 1067.41496.

7aa

Cyclopentanecarbonyl chloride (9.96 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118 (5)** (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (1.36 mg, 5.85 μ mol, 25 mol%) and DIPEA (15.8 μ L, 93.6 μ mol,

4.0 eq.) in 1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 15 min – 45%, 80 min – 65%, 80.5 min – 100%] to afford, after lyophilization, **7aa** (t_R = 43.8 min, 9.50 mg, 8.20 μ mol, 35%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25\text{ °C}} = -37.34$ (c = 0.23, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3439, 2975, 2936, 2874, 1698, 1643, 1589, 1404, 1379, 1312, 1248, 1198, 1162, 1144, 1111, 1066, 1023 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.24 (d, J = 11.4 Hz, 1H), 6.80 – 6.57 (m, 1H), 5.96 (ddd, J = 14.6, 9.6, 4.6 Hz, 1H), 5.88 (s, 1H), 5.69 – 5.61 (m, 1H), 5.22 (d, J = 10.6 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.82 – 4.71 (m, 3H), 4.68 (s, 1H), 4.60 (d, J = 11.5 Hz, 1H), 4.42 (d, J = 11.4 Hz, 1H), 4.26 (s, 1H), 4.10 – 3.99 (m, 2H), 3.86 – 3.77 (m, 2H), 3.77 (d, J = 9.7 Hz, 1H), 3.67 – 3.56 (m, 2H), 3.52 (s, 3H), 3.00 (qd, J = 7.4, 1.8 Hz, 2H), 2.86 – 2.72 (m, 2H), 2.75 – 2.66 (m, 1H), 2.70 – 2.58 (m, 1H), 2.55 – 2.40 (m, 2H), 1.94 – 1.89 (m, 1H), 1.88 – 1.82 (m, 4H), 1.81 (d, J = 1.3 Hz, 3H), 1.74 (d, J = 1.4 Hz, 3H), 1.68 – 1.61 (m, 5H), 1.60 – 1.53 (m, 2H), 1.31 (d, J = 6.1 Hz, 3H), 1.26 – 1.16 (m, 10H), 1.14 (s, 3H), 0.82 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 176.7, 169.6, 167.8, 156.0, 154.3, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.7, 110.2, 108.2, 101.8, 96.2, 93.0, 81.7, 78.2, 77.6, 75.2, 74.8, 72.9, 72.3, 71.4, 70.7, 70.6, 67.8, 63.4, 61.7, 44.5, 42.1, 37.3, 30.7, 30.4, 28.9, 28.4, 26.5, 26.4, 26.4, 26.3, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₄H₇₆Cl₂O₁₈Na [M+Na]⁺:1105.43009, found: 1105.43035.

7ab

Cyclohexanecarbonyl chloride (10.9 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (2.71 mg, 11.7 μ mol, 50 mol%) and DIPEA (15.8 μ L,

93.6 µmol, 4.0 eq.) in 1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 50 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 50%, 60 min – 70%, 61 min – 100%] to afford, after lyophilization, **7ab** (t_R = 28.1 min, 17.6 mg, 16.0 µmol, 69%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25\text{ °}C}$ = -31.25 (c = 0.30, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3449, 2930, 2856, 1692, 1590, 1452, 1405, 1380, 1312, 1246, 1198, 1176, 1162, 1134, 1066, 1023, 949, 897, 800, 757 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.24 (d, J = 11.4 Hz, 1H), 6.63 (dd, J = 15.0, 11.5 Hz, 1H), 5.96 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.88 (s, 1H), 5.65 (t, J = 8.3 Hz, 1H), 5.22 (d, J = 10.6 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.77 (s, 1H), 4.76 – 4.71 (m, 2H), 4.68 (s, 1H), 4.60 (d, J = 11.4 Hz, 1H), 4.42 (d, J = 11.4 Hz, 1H), 4.26 (s, 1H), 4.10 – 3.99 (m, 2H), 3.85 – 3.79 (m, 2H), 3.77 (d, J = 9.7 Hz, 1H), 3.65 – 3.59 (m, 2H), 3.52 (s, 3H), 3.00 (q, J = 7.4 Hz, 2H), 2.84 – 2.74 (m, 1H), 2.73 – 2.67 (m, 1H), 2.67 – 2.59 (m, 1H), 2.55 – 2.41 (m, 2H), 2.34 (tt, J = 11.0, 3.7 Hz, 1H), 1.95 – 1.87 (m, 3H), 1.81 (s, 3H), 1.74 (s, 3H), 1.73 – 1.70 (m, 2H), 1.65 (s, 3H), 1.64 – 1.60 (m, 1H), 1.50 – 1.39 (m, 2H), 1.35 – 1.17 (m, 16H), 1.14 (s, 3H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 175.9, 169.5, 167.8, 155.9, 153.9, 145.4, 143.4, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.6, 110.6, 108.2, 101.8, 96.2, 93.0, 81.6, 78.2, 77.6, 75.2, 74.7, 72.9, 72.3, 71.4, 70.7, 70.6, 67.8, 63.4, 61.7, 43.8, 42.0, 37.3, 28.9, 28.4, 26.6, 26.5, 26.2, 26.1, 26.0, 20.7, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(-), (MeOH) calculated for $C_{65}H_{77}Cl_2O_{18}$ [M-H]: 1095.44924, found: 1095.44910.

7ad

Methoxyacetyl chloride (7.47 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol,

4.0 eq.) in 1,4-dioxane (125 μ L) under ambient atmosphere. The reaction mixture was stirred at 30 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 μ mol, 10 mol%) followed by morpholine (8.15 μ L, 93.6 μ mol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 12 min – 45%, 60 min – 50%, 70 min – 100%] to afford, after lyophilization, **7ad** (t_R = 17.6 min, 16.0 mg, 15.1 μ mol, 65%) as a slightly yellow solid.

Specific Rotation $[a]_D^{25\text{ °C}} = -34.52$ (c = 0.27, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3452, 2975, 2934, 2170, 1741, 1693, 1642, 1587, 1452, 1377, 1310, 1245, 1197, 1120, 1066, 1022, 899, 758, 663, 592, 516 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.24 (d, J = 11.4 Hz, 1H), 6.69 – 6.51 (m, 1H), 5.96 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.88 (s, 1H), 5.65 (t, J = 8.1 Hz, 1H), 5.22 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.86 (dd, J = 10.3, 3.1 Hz, 1H), 4.81 (s, 1H), 4.77 – 4.72 (m, 1H), 4.68 (s, 1H), 4.60 (d, J = 11.5 Hz, 1H), 4.26 (s, 1H), 4.08 – 4.02 (m, 4H), 3.85 – 3.75 (m, 3H), 3.67 – 3.57 (m, 2H), 3.52 (s, 3H), 3.38 (s, 3H), 3.00 (q, J = 7.4, 1.4Hz, 2H), 2.80 – 2.59 (m, 3H), 2.55 – 2.40 (m, 2H), 1.95 – 1.88 (m, 1H), 1.81 (d, J = 1.2 Hz, 3H), 1.74 (d, J = 1.3 Hz, 3H), 1.65 (s, 3H), 1.31 (d, J = 6.1 Hz, 3H), 1.25 – 1.16 (m, 10H), 1.15 (s, 3H), 0.82 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 170.8, 169.4, 167.8, 155.8, 153.8, 145.4, 143.4, 142.7, 136.9, 136.1, 135.9, 133.9, 128.1, 126.4, 125.3, 124.0, 144.5, 110.6, 108.1, 101.8, 96.1, 93.0, 81.6, 78.2, 77.6, 75.4, 75.2, 72.8, 72.3, 71.2, 70.6 (2C), 70.0, 67.6, 63.4, 61.7, 59.1, 42.0, 37.2, 28.9, 28.4, 26.5, 26.2, 20.6, 18.2, 17.7, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for $C_{51}H_{72}O_{19}Cl_2Na$ [M+Na]+: 1081.39426, found: 1081.39396.

7ag

4-Chlorobutyryl chloride (9.35 μ L, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (5) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol,

4.0 eq.) in 1,4-dioxane (125 µL) under ambient atmosphere. The reaction mixture was stirred at 40 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min LC time program (min –% B): 12 min – 50%, 60 min – 55%, 70 min – 100%] to afford, after lyophilization, **7ag** ($t_R = 23.5$ min, 16.8 mg, 15.4 µmol, 66%) as a slightly yellow solid.

Specific Rotation [α]_D^{25°C} = -15.73 (c = 0.39, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 2976, 2034, 1693, 1242, 1066, 771 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.24 (d, J = 11.5 Hz, 1H), 6.68 – 6.46 (m, 1H), 5.96 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.88 (s, 1H), 5.65 (t, J = 8.2 Hz, 1H), 5.20 (d, J = 1.6 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.84 – 4.79 (m, 1H), 4.79 (s, 1H), 4.78 – 4.71 (m, 1H), 4.68 (s, 1H), 4.60 (d, J = 11.5 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.26 (s, 1H), 4.06 – 4.00 (m, 2H), 3.84 – 3.75 (m, 3H), 3.69 (t, J = 6.6 Hz, 2H), 3.64 – 3.59 (m, 2H), 3.52 (s, 3H), 3.00 (q, J = 7.3 Hz, 2H), 2.81 – 2.74 (m, 1H), 2.73 – 2.60 (m, 2H), 2.54 (t, J = 7.2 Hz, 2H), 2.52 – 2.42 (m, 2H), 2.12 – 2.06 (m, 2H), 1.95 – 1.88 (m, 1H), 1.81 (s, 3H), 1.74 (d, J = 1.3 Hz, 3H), 1.65 (d, J = 1.2 Hz, 3H), 1.31 (d, J = 6.1 Hz, 3H), 1.28 – 1.17 (m, 10H), 1.14 (s, 3H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 172.9, 169.4, 167.8, 155.8, 153.8, 145.4, 143.4, 142.7, 136.9, 136.1, 135.9, 133.9, 128.1, 126.4, 125.3, 124.0, 114.5, 110.6, 108.2, 101.8, 96.1, 93.0, 81.6, 78.2, 77.6, 75.3, 75.2, 72.8, 72.3, 71.3, 70.6 (2C), 67.7, 63.4, 61.7, 45.0, 42.0, 37.3, 32.0, 28.9, 28.8, 28.4, 26.5, 26.2, 20.7, 18.2, 17.7, 17.5, 15.2, 14.4, 13.8, 11.1 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₂H₇₃O₁₈Cl₃Na [M+Na]⁺: 1113.37602, found: 1113.37442.

7ah

Hex-5-ynoyl chloride (10.7 mg, 81.9 μ mol, 3.5 eq.) was added to a stirred mixture of **bisallyl-OP-1118** (**5**) (35.6 mg, 23.4 μ mol, 1.0 eq.), boronic acid (679 μ g, 2.93 μ mol, 12.5 mol%) and DIPEA (15.8 μ L, 93.6 μ mol, 4.0 eq.) in 1,4-dioxane (125 μ L) under ambient

atmosphere. The reaction mixture was stirred at 30 °C for 16 h and the reaction was allowed to reach room temperature. Pd(PPh₃)₄ (2.70 mg, 2.34 µmol, 10 mol%) followed by morpholine (8.15 µL, 93.6 µmol, 4.0 eq.) were added to the crude mixture under ambient atmosphere and the reaction was stirred at room temperature for 45 min. The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 µ, 110 Å, 250 mm × 21.2 mm, solvent A: $H_2O + 0.1\%$ HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min LC time program (min -% B): 12 min -50%, 60 min -60%, 70 min -100%] to afford, after lyophilization, **7ah** ($t_R = 21.8$ min, 17.5 mg, 16.2 µmol, 69%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25 \circ c}$ = -38.70 (c = 0.24, CHCl₃); **FT-IR** $\tilde{\nu}$ (film) 3453, 2975, 2934, 1692, 1589, 1379, 1312, 1243, 1144, 1066, 1021, 899, 800, 758, 638, 514 cm⁻¹; ¹**H NMR** (400 MHz, acetone- d_6) δ 7.24 (d, J = 11.4 Hz, 1H), 6.63 (dd, J = 15.0, 11.5 Hz, 1H), 5.97 (ddd, J = 14.6, 9.5, 4.6 Hz, 1H), 5.88 (s, 1H), 5.65 (t, J = 8.3 Hz, 1H), 5.22 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.83 – 4.77 (m, 2H), 4.77 – 4.71 (m, 1H), 4.68 (s, 1H), 4.60 (d, J = 11.4 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.26 (s, 1H), 4.12 – 3.96 (m, 2H), 3.84 – 3.74 (m, 3H), 3.66 – 3.57 (m, 2H), 3.52 (s, 3H), 3.06 – 2.96 (m, 2H), 2.84 – 2.59 (m, 3H), 2.56 – 2.41 (m, 4H), 2.37 (t, J = 2.7 Hz, 1H), 2.27 (td, J = 7.1, 2.6 Hz, 2H), 1.95 – 1.88 (m, 1H), 1.87 – 1.78 (m, 5H), 1.74 (s, 3H), 1.65 (s, 3H), 1.31 (d, J = 6.2 Hz, 3H), 1.26 – 1.17 (m, 10H), 1.14 (s, 3H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 173.2, 169.6, 167.8, 156.1, 154.5, 145.4, 143.3, 142.6, 136.9, 136.1, 135.9, 133.9, 128.2, 126.4, 125.3, 124.0, 114.8, 109.9, 108.2, 101.8, 96.2, 93.0, 84.3, 81.7, 78.2, 77.5, 75.2, 75.2, 72.8, 72.3, 71.3, 70.6 (2C), 70.4, 67.7, 63.4, 61.7, 42.0, 37.3, 33.6, 28.9, 28.4, 26.5, 26.3, 24.8, 20.7, 18.2, 18.2, 17.7, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+), (MeOH) calculated for C₅₄H₇₄O₁₈Cl₂Na [M+Na]⁺: 1103.41444, found: 1103.41499.

7ai

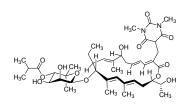
A vial under an inert atmosphere was charged with **7ah** (10 mg, 9.25 μ mol, 1.0 eq.) and **Dinuclear Copper Catalyst** (0.65 mg, 0.92 μ mol, 0.1 eq.). The solids were dissolved in DCM (100 μ L) and a solution of azidoacetic acid (1.38 μ L, 18.5 μ mol, 2 eq.) in THF (100 μ L) was added at room temperature. The

reaction mixture was stirred at room temperature until full conversion was observed (UHPLC-MS). The solvent was removed under reduced pressure and the crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min LC time program (min –% B): 12 min – 50%, 60 min – 60%, 61 min – 100%] to afford, after lyophilization, **7ai** (t_R = 10.3 min, 10.0 mg, 8.45 μ mol, 91%) as a slightly yellow solid.

Specific Rotation [α] $_D^{25}$ ° $_C$ = -8.39 (c = 0.37, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3425, 2974, 2934, 1710, 1379, 1312, 1242, 1142, 1066, 1022, 900, 800, 772, 600 cm $^{-1}$; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.81 (s, 1H), 7.24 (d, J = 11.5 Hz, 1H), 6.63 (dd, J = 15.0, 11.5 Hz, 1H), 5.97 (ddd, J = 14.7, 9.6, 4.6 Hz, 1H), 5.88 (s, 1H), 5.65 (t, J = 8.3 Hz, 1H), 5.28 (s, 2H), 5.22 (d, J = 10.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.84 – 4.79 (m, 1H), 4.79 (s, 1H), 4.74 (q, J = 5.3 Hz, 1H), 4.68 (s, 1H), 4.60 (d, J = 11.4 Hz, 1H), 4.42 (d, J = 11.5 Hz, 1H), 4.26 (s, 1H), 4.06 – 4.00 (m, 2H), 3.86 – 3.79 (m, 2H), 3.77 (d, J = 9.8 Hz, 1H), 3.67 – 3.58 (m, 2H), 3.52 (s, 3H), 3.05 – 2.96 (m, 2H), 2.83 – 2.73 (m, 3H), 2.73 – 2.68 (m, 1H), 2.67 – 2.60 (m, 1H), 2.54 – 2.42 (m, 2H), 2.39 (td, J = 7.3, 2.1 Hz, 2H), 2.02 – 1.95 (m, 2H), 1.91 (m, 1H), 1.81 (s, 3H), 1.74 (s, 3H), 1.65 (s, 3H), 1.31 (d, J = 6.1 Hz, 3H), 1.26 – 1.17 (m, 10H), 1.15 (s, 3H), 0.81 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 173.4, 169.4, 168.7, 167.8, 155.7, 153.7, 147.5, 145.5, 143.5, 142.7, 136.8, 136.1, 136.0, 133.9, 128.1, 126.3, 125.3, 124.0, 123.9, 114.5, 110.6, 108.1, 101.7, 96.2, 93.0, 81.6, 78.2, 77.6, 75.2, 75.2, 72.8, 72.2, 71.2, 70.6 (2C), 67.6, 63.4, 61.7, 51.1, 42.0, 37.2, 33.9, 28.9, 28.4, 26.5, 26.2, 25.7, 25.2, 20.6, 18.2, 17.8, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+), (MeOH) calculated for $C_{56}H_{77}O_{20}Cl_2N_3Na$ [M+Na]*: 1204.43697, found: 1204.43724.

Experimental Procedures Tsuji-Trost Functionalizations

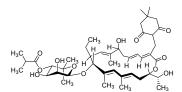
9



In a flame-dried microwave tube, a mixture of fidaxomicin (**1**, 30.0 mg, 28.4 μ mol, 1.0 eq.), 1,3-dimethylbarbituric acid (17.7 mg, 0.114 mmol, 4.0 eq.), Pd(OAc)₂ (3.19 mg, 14.2 μ mol, 50 mol%) and Xantphos (16.4 mg, 28.4 μ mol, 1.0 eq.) was dissolved in dry toluene (500 μ L, degassed by freeze-pump-thaw (3x)). The microwave tube was

evacuated, flushed with argon several times and sealed. The mixture was heated to 90 °C for 5 h and then diluted with EtOAc (2 mL) and washed with H₂O (3 x 2 mL). The organic phase was dried over MgSO₄ and the solvent was evaporated under reduced pressure. The crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 10 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 5%, 10 min – 5%, 12 min – 40%, 55 min – 60%, 56 min – 100%] to afford, after lyophilization, barbituric acid **9** (t_R = 30.1 min, 9.5 mg, 12 μ mol, 42%) as a slightly yellow solid.

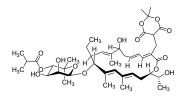
TLC (pentane:acetone, 2:3 v/v) = 0.13; **Specific Rotation** [α] $_D^{26}$ ° c = +32.42 (c = 0.29, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3386, 2972, 2970, 2874, 2857, 1677, 1585, 1446, 1424, 1381, 1292, 1253, 1198, 1147, 1074, 1031, 951, 899, 842, 797, 756, 717, 674, 584, 505, 472 cm $^{-1}$; ¹**H NMR** (500 MHz, CD₃OD) δ 7.14 (d, J = 11.4 Hz, 1H), 6.40 (dd, J = 14.6, 12.2 Hz, 1H), 5.89 (ddd, J = 14.6, 9.4, 4.7 Hz, 1H), 5.81 (s, 1H), 5.54 (t, J = 8.3 Hz, 1H), 5.13 (dt, J = 10.5, 1.6 Hz, 1H), 5.01 (d, J = 10.2 Hz, 1H), 4.71 (s, 1H), 4.63 (dt, J = 6.9, 5.0 Hz, 1H), 4.22 (m, 1H), 3.95 (quint, J = 6.4 Hz, 1H), 3.92 (d, J = 3.1 Hz, 1H), 3.75 – 3.68 (m, 3H), 3.18 (s, 3H), 3.17 (s, 3H), 3.12 (d, J = 14.3 Hz, 1H), 3.05 (d, J = 14.2 Hz, 1H), 2.74 – 2.63 (m, 3H), 2.59 (sept, J = 7.0 Hz, 1H), 2.48 (ddd, J = 16.2, 9.4, 4.4 Hz, 1H), 2.39 (ddd, J = 13.9, 9.0, 4.5 Hz, 1H), 2.05 – 1.96 (m, 1H), 1.79 (d, J = 1.3 Hz, 3H), 1.70 (d, J = 1.3 Hz, 3H), 1.65 (s, 3H), 1.33 – 1.22 (m, 1H), 1.19 – 1.10 (m, 15H), 0.88 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CD₃OD) δ 178.4, 170.4, 170.2, 169.3, 153.4, 144.9, 143.0, 136.94, 136.92, 136.3, 134.6, 128.5, 126.5, 124.6, 124.4, 97.2, 94.3, 78.7, 75.9, 74.6, 73.5, 73.2, 70.5, 68.2, 49.9, 42.5, 37.2, 35.4, 29.1, 28.8, 28.73, 28.70, 28.4, 26.9, 20.4, 19.5, 19.1, 18.7, 17.5, 15.4, 13.9, 11.3 ppm; **HRMS** ESI(+), (MeOH) calculated for C₄₂H₆₂N₂O₁₃Na [M+Na]*: 825.41441, found: 825.41465.



In a flame-dried microwave tube, a mixture of fidaxomicin (1, 20.3 mg, 19.2 µmol, 1.0 eq.), dimedone (10.6 mg, 75.6 µmol, 4.0 eq.) and Pd(PPh₃)₄ (5.0 mg, 4.3 µmol, 25 mol%) was dissolved in dry toluene (100 µL, degassed by freeze-pump-thaw (3x)). The microwave tube was

evacuated and flushed with argon several times and the tube was sealed. The mixture was heated to 90 °C for 5 h and then diluted with EtOAc (2 mL) and washed with H₂O (3 x 2 mL). The organic phase was dried over MgSO₄ and the solvent was evaporated under reduced pressure. The crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 5%, 10 min – 5%, 12 min – 40%, 55 min –60%, 56 min – 100%] to afford, after lyophilization, dimedone **10** (t_R = 38.2 min, 4.10 mg, 5.2 μ mol, 27%) as a slightly yellow solid.

TLC (pentane:acetone, 1:1 v/v) = 0.82; **Specific Rotation** [α]_D²⁴ °C = -12.86 (c = 0.22, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3421, 2971, 2933, 2874, 1732, 1698, 1656, 1637, 1607, 1470, 1385, 1370, 1355, 1292, 1257, 1208, 1150, 1078, 1034, 897, 797, 712 cm⁻¹; ¹**H NMR** (500 MHz, CD₃OD) δ 6.86 (d, J = 11.3 Hz, 1H), 6.65 – 6.55 (m, 1H), 5.83 (s, 1H), 5.71 (ddd, J = 14.7, 9.4, 4.9 Hz, 1H), 5.53 (t, J = 8.2 Hz, 1H), 5.09 (dt, J = 10.4, 1.6 Hz, 1H), 5.01 (d, J = 10.3 Hz, 1H), 4.70 (s, 1H), 4.66 (q, J = 6.2 Hz, 1H), 4.17 (s, 1H), 4.04 (quint, J = 6.3 Hz, 1H), 3.91 (d, J = 2.8 Hz, 1H), 3.72 (dd, J = 10.2, 3.2 Hz, 1H), 3.69 (d, J = 9.8 Hz, 1H), 3.32 (d, J = 14.5 Hz, 1H), 3.25 (d, J = 14.6 Hz, 1H), 2.77 – 2.54 (m, 4H), 2.48 – 2.33 (m, 2H), 2.23 (s, 4H), 2.01 (m, 1H), 1.78 (d, J = 1.3 Hz, 3H), 1.76 (d, J = 1.4 Hz, 3H), 1.63 (d, J = 1.3 Hz, 3H), 1.34 – 1.22 (m, 1H), 1.20 – 1.09 (m, 15H), 1.00 (s, 6H), 0.87 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CD₃OD) δ 208.4 (HMBC), 178.4, 172.6, 142.0, 140.0, 137.1, 136.9, 136.2, 134.7, 130.1, 129.6, 127.1, 124.6, 114.0, 97.1, 94.3, 79.1, 76.0, 74.5, 73.7, 73.2, 70.6, 68.3, 48.3 (HMBC), 42.6, 37.37, 35.4, 32.8, 28.7, 28.5, 28.3, 28.2, 26.9, 20.2, 19.5, 19.1, 18.7, 17.4, 15.5, 13.9, 11.3 ppm; **HRMS** ESI(+), (MeOH) calculated for C₄₄H₆₆O₁₂Na [M+Na]⁺: 809.44465, found: 809.44499.



A flame-dried microwave tube was charged with fidaxomicin (1, 100 mg, 94.5 μ mol, 1.0 eq.), Meldrum's acid (54.5 mg, 0.378 mmol, 4.0 eq.) and Pd(PPh₃)₄ (27.3 mg, 23.6 μ mol, 25 mol%). The tube was sealed and evacuated and flushed with argon several times. Then, the solids were dissolved in dry toluene (1.9 mL, degassed by freeze-pump-thaw (3x))

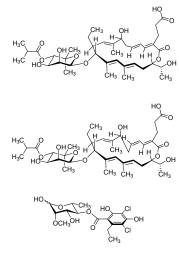
and the mixture was stirred for 3 h at 70 °C. After evaporation of the solvent the crude mixture was purified by RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 5%, 10 min – 5%, 11 min – 55%, 55 min – 60%, 56 min – 100%] to afford, after lyophilization, Meldrum's acid **11** (t_R = 19.7 min, 19.5 mg, 24.6 μ mol, 26%) as a colorless solid.

TLC (pentane:acetone, 1:1 v/v) = 0.30; **Specific Rotation** [α]_D²⁴°^C = +36.52 (c = 0.69, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3452, 2976, 2934, 2876, 1782, 1744, 1690, 1641, 1573, 1438, 1385, 1299, 1254, 1205, 1151, 1075, 1031, 948, 898, 798, 718 cm⁻¹; ¹**H NMR** (500 MHz, CD₃OD) δ 7.10 (d, J = 11.4 Hz, 1H), 6.54 (dd, J = 14.9, 11.9 Hz, 1H), 5.87 (ddd, J = 14.6, 9.6, 4.9 Hz, 1H), 5.83 (s, 1H), 5.54 (t, J = 8.1 Hz, 1H), 5.13 (dt, J = 10.5 Hz, 1.6 Hz, 1H), 5.02 (d, J = 10.3 Hz, 1H), 4.71 (d, J = 1.2 Hz, 1H), 4.66 (td, J = 6.1, 4.3 Hz, 1H), 4.21 (m, 1H), 3.99 (quint, J = 6.3 Hz, 1H), 3.92 (dd, J = 3.3, 1.1 Hz, 1H), 3.76 – 3.68 (m, 2H), 3.12 (d, J = 15.3 Hz, 1H), 3.03 (d, J = 15.4 Hz, 1H), 2.76 – 2.64 (m, 3H), 2.59 (sept, J = 7.0 Hz, 1H), 2.49 (ddd, J = 14.9, 9.5, 4.5 Hz, 1H), 2.40 (ddd, J = 13.8, 8.6, 4.4 Hz, 1H), 2.07 – 1.95 (m, 1H), 1.81 (s, 3H), 1.80 (s, 3H), 1.76 (s, 3H), 1.74 (s, 3H), 1.66 (s, 3H), 1.33 – 1.24 (m, 1H), 1.19 – 1.10 (m, 15H), 0.88 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CD₃OD) δ 178.4, 170.0, 167.3, 167.2, 143.6, 141.5, 137.1, 137.0, 136.2, 134.7, 129.0, 126.9, 125.3, 124.6, 106.3, 97.1, 94.3, 78.6, 76.0, 74.5, 73.6, 73.2, 70.55, 68.3, 45.8, 42.6, 37.3, 35.4, 28.8, 28.7, 28.4, 26.9, 26.5, 24.6, 20.2, 19.5, 19.1, 18.7, 17.5, 15.4, 14.0, 11.3 ppm; **HRMS** ESI(+), (MeOH) calculated for C₄₂H₆₂O₁₄Na [M+Na]⁺: 813.40375, found: 813.40318.

Baylis-Hillman type mechanism - Control experiment

A flame-dried microwave tube was charged with fidaxomicin (1, 5 mg, 4.72 μ mol, 1.0 eq.), Meldrum's acid (2.72 mg, 0.018 mmol, 4.0 eq.) and PPh₃ (0.62 mg, 2.36 μ mol, 50 mol% or 2.5 mg, 9.44 μ mol, 200 mol%). The tube was sealed and evacuated and flushed with argon several times. Then, the solids were dissolved in dry toluene (100 \Box L, degassed by freeze-pump-thaw (3x)) and the mixture was stirred for 3 h at 70 °C. After evaporation of the solvent, the crude mixture was analyzed by UPLC/MS analysis and 1 H NMR. In both cases, mainly starting material was recovered with slight degradation.

(E)-13, (Z)-13 and 12



A flame-dried microwave tube was charged with fidaxomicin (1, 500 mg, 0.473 mmol, 1.0 eq.), Meldrum's acid (136 mg, 0.946 mmol, 2.0 eq.) and Pd(PPh₃)₄ (137 mg, 0.118 mmol, 25 mol%). The tube was sealed and evacuated and flushed with argon several times. Then, the solids were dissolved in dry toluene (4.5 mL, degassed by freeze-pump-thaw (3x)) and the mixture was stirred for 3 h at 70 °C. The solvent was evaporated and the residue was dissolved in DMF/H₂O (9:1, 4.5 mL) and heated to 100 °C for 5 h. The reaction was filtered over Celite^â and the solvent was evaporated. The crude mixture was purified by RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –%

B): 0.0 min - 35%, 15 min - 35%, 37.5 min - 40%, 38 min - 100%] to afford, after lyophilization, sugar **12** (α/β = 4:1, t_R = 9 min, 137 mg, 0.333 mmol, 70%) carboxylic acid (*E*)-**13** (t_R = 22.5 min, 122 mg, 0.173 mmol, 37%) and carboxylic acid (*Z*)-**13** (t_R = 26 min, 42.9 mg, 60.6 μ mol, 13%)

(*E*)-13

TLC (pentane:acetone, 1:1 v/v) = 0.20; **Specific Rotation** [α]_D^{22°C} = +33.75 (c = 0.40, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3417, 2975, 2934, 2875, 1700, 1641, 1602, 1447, 1386, 1370, 1353, 1295, 1250, 1207, 1152, 1082, 1033, 976, 951, 898, 798, 764 cm⁻¹; ¹**H NMR** (400 MHz, CD₃OD) δ 7.06 (d, J = 11.4 Hz, 1H), 6.46 (dd, J = 14.6, 11.4 Hz, 1H), 5.85 (ddd, J = 14.6, 9.6, 4.4, 1H), 5.82 (s, 1H), 5.55 (t, J = 8.2 Hz, 1H), 5.12 (dt, J = 10.5, 1.6 Hz, 1H), 5.02 (d, J = 10.2 Hz, 1H), 4.71 (d, J = 1.1 Hz, 1H), 4.67 (td, J = 6.2, 4.5 Hz, 1H), 4.22 (m, 1H), 4.02 (quint, J = 6.2 Hz, 1H), 3.92 (d, J = 3.1 Hz, 1H), 3.76 – 3.66 (m, 2H), 2.75 – 2.54 (m, 6H),2.49 (ddd, J = 15.0, 9.6, 4.4 Hz, 1H), 2.45 – 2.33 (m, 3H), 2.04 – 1.95 (m, 1H), 1.81 (d, J = 1.3 Hz, 3H), 1.74 (d, J = 1.3 Hz, 3H), 1.66 (d, J = 1.2 Hz, 3H), 1.34 – 1.23 (m, 1H), 1.19 – 1.10 (m, 15H), 0.88 (t, J = 7.4 Hz, 3H); ¹³**C NMR** (126 MHz, CD₃OD) δ 178.4, 177.0, 169.8, 142.7, 141.1, 137.1, 136.9, 136.2, 134.7, 128.84, 128.80, 126.9, 124.5, 97.1, 94.3, 78.5, 76.0, 74.5, 73.6, 73.2, 70.6, 68.2, 42.5, 37.3, 35.4, 34.7, 28.7, 28.2, 26.9, 23.8, 20.2, 19.5, 19.1, 18.7, 17.5, 15.4, 13.9, 11.3 ppm; **HRMS** ESI(+), (MeOH) calculated for C₃₈H₅₈O₁₂Na [M+Na]*: 729.38150, found: 729.38205.

(*Z*)-13

TLC (pentane:acetone, 1:1) = 0.20; **Specific Rotation** [α]_D²⁵°^C = +57.57 (c = 1.00, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3453, 2976, 2934, 2875, 1704, 1633, 1593, 1385, 1260, 1203, 1149, 1078, 1034, 1035, 878, 796 cm⁻¹; **¹H NMR** (500 MHz, CD₃OD) δ 7.38 (d, J = 11.8 Hz, 1H), 6.48 (t, J = 11.3, 1H), 6.01 (dt, J = 9.4, 5.6 Hz, 1H), 5.74 (s, 1H), 5.55 (t, J = 7.7 Hz, 1H), 5.14 (dt, J = 10.9, 1.4 Hz, 1H), 5.02 (d, J = 10.2 Hz, 1H), 4.84 – 4.78 (m, 1H), 4.70 (d, J = 1.2 Hz, 1H), 4.04 – 3.95 (m, 2H), 3.93 (d, J = 3.0 Hz, 1H), 3.73 (dd, J = 10.2, 3.2 Hz, 1H), 3.69 (d, J = 9.4 Hz, 1H), 2.74 – 2.64 (m, 3H), 2.63 – 2.54 (m, 2H), 2.49 (m, 1H), 2.44 – 2.33 (m, 3H), 2.25 – 2.14 (m, 1H), 2.07 – 1.97 (m, 1H), 1.62 (s, 3H), 1.61 (s, 3H), 1.59 (s, 3H), 1.29 – 1.22 (m, 1H), 1.19 – 1.10 (m, 15H), 0.89 (t, J = 7.4 Hz, 3H); 13C **NMR** (126 MHz, CD₃OD) δ 178.4, 177.4, 168.9, 140.0, 139.9, 136.9, 136.5, 135.4, 133.5, 131.4, 125.5, 125.3, 123.9, 97.4, 94.5, 78.6, 75.9, 74.9, 74.6, 73.0, 70.5, 67.9, 42.8, 37.2, 35.4, 35.0, 28.8, 28.7, 26.8, 23.7, 20.4, 19.5, 19.1, 18.8, 18.0, 16.0, 14.1, 11.8 ppm; **HRMS** ESI(+), (MeOH) calculated for C₃₈H₅₈O₁₂Na [M+Na]⁺: 729.38150, found: 729.38250.

12

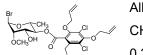
TLC (acetone:pentane, 3:2 v/v) = 0.43; **FT-IR** (film) $\tilde{\nu}$ =3401, 2986, 2939, 1655, 1560, 1456, 1411, 1390, 1374, 1314, 1290, 1242, 1195, 1145, 1128, 1100, 1073, 1039, 1018, 996, 910, 834, 799, 759, 732, 710, 688, 648, 596, 519 cm⁻¹; α: ¹**H NMR** (500 MHz, CDCl₃) δ 5.38 (d, J = 1.5 Hz, 1H), 5.20 (t, J = 9.9 Hz, 1H), 4.16–4.03 (m, 2H), 3.61 (dd, J = 3.7, 1.5 Hz, 1H); 3.53 (s, 3H), 3.17–2.97 (m, 2H), 1.27–1.18 (m, 6H) ppm; ¹³**C NMR** (126 MHz, CDCl₃) δ 170.26, 157.7, 152.6, 143.3, 113.8, 107.4, 107.1, 90.8, 80.5, 76.7, 69.1, 65.9, 59.0, 26.2, 17.8, 14.0 ppm; β: ¹**H NMR** (500 MHz, CDCl₃) δ 5.15 (t, J = 9.8 Hz, 1H), 4.85 (d, J = 1.3 Hz, 1H), 3.80 (dd, J = 9.9, 3.5 Hz, 1H), 3.74 (s, 1H), 3.67 (dd, J = 3.6, 1.3 Hz, 1H); 3.58–3.55 (m, 1H), 3.17–2.97 (m, 2H), 1.31 (d, J = 6.2 Hz, 1H), 1.27–1.18 (m, 3H) ppm; ¹³**C NMR** (126 MHz, CDCl₃) δ 170.34, 157.6, 152.7, 143.1, 113.9, 107.3, 107.2, 94.7, 81.4, 76.2, 72.8, 69.9, 63.2, 26.2, 17.8, 14.0 ppm; **HRMS** ESI(–) (MeOH) calculated for C₁₆H₁₉Cl₂O₈ [M–H]⁻: 409.04625, found: 409.04625.

S1

The Rhamnosyl-resorcylate moiety (**12**, 100 mg, 0.243 mmol, 1.0 eq.) and K_2CO_3 (134 mg, 0.972 mmol, 4.0 eq.) were dissolved in dry DMF (4.8 mL) and allyl bromide (63 μ L, 0.73 mmol, 3.0 eq.) was added dropwise. The mixture was

warmed to 45 °C and it was stirred for 3 h. After complete conversion as indicated by TLC (EtOAc/pentane 1:1), the reaction mixture was diluted with EtOAc (10 mL) and washed with aq. sat. NH₄Cl (3 x 10 mL). The combined organic layers were dried over MgSO₄ and the solvent was evaporated under reduced pressure. The crude was purified by silica-gel column chromatography (EtOAc/penante 1:2) to afford the desired allyl-protected rhamnosyl-resorcylate **S1** (a/b = 6:1, 112 mg, 0.229 mmol, 94%) as a colorless oil.

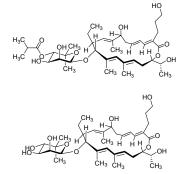
TLC (EtOAc:pentane, 1:1 v/v) = 0.60; **FT-IR** (film) \tilde{v} =3452, 2939, 1734, 1651, 1568, 1456, 1403, 1351, 1315, 1252, 1193, 1103, 1065, 1042, 1019, 933, 799, 749, 601 cm⁻¹; α: ¹**H NMR** (400 MHz, CDCl₃) δ 6.19–5.95 (m, 2H), 5.46–5.18 (m, 4H), 5.33 (s, 1H), 5.09 (t, J = 9.8 Hz, 1H), 4.62–4.44 (m, 4H), 4.08–3.92 (m, 2H), 3.59–3.57 (m, 1H), 3.53 (s, 3H), 3.1 (d, J = 3.5 Hz, 1H), 2.89–2.70 (m, 2H), 2.54 (d, J = 10.7 Hz, 1H), 1.30 (d, J = 6.3 Hz, 3H), 1.19 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CDCl₃) δ 166.6, 153.1, 151.0, 139.2, 132.9, 132.8, 127.3, 125.6, 121.6, 119.1, 119.0, 91.0, 80.7, 76.6, 75.9, 74.4, 69.2, 66.0, 59.1, 25.3, 17.7, 14.1 ppm; β: ¹**H NMR** (400 MHz, CDCl₃) 6.19–5.95 (m, 2H), 5.46–5.18 (m, 4H), 5.05 (t, J = 9.8 Hz, 1H), 4.76 (d, J = 1.4 Hz, 1H), 4.62–4.44 (m, 4H), 3.78–3.75 (m, 1H), 3.74 (s, 3H), 3.63 (dd, J = 3.4, 1.3 Hz, 1H), 3.51–3.44 (m, 1H), 2.89–2.70 (m, 2H), 1.35 (d, J = 6.2 Hz, 3H), 1.19 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CDCl₃) δ166.7, 153.3, 150.9, 138.9, 132.8, 132.5, 127.0, 125.7, 121.6, 119.5, 119.2, 94.4, 81.3, 76.3, 76.0, 73.2, 70.0, 65.9, 62.8, 25.3, 17.7, 14.1 ppm; **HRMS** ESI(–) (MeOH) calculated for C₂₃H₂₉Cl₂O₁₀ [M+HCOO]⁻: 535.11433, found: 535.11416.



Allyl-protected sugar **S2** (50.0 mg, 0.102 mmol, 1.0 eq.) was dissolved in dry CH_2Cl_2 (500 μ L) and cooled to 0 °C. Then, HBr (33 wt% in AcOH, 51 μ L, 0.31 mmol, 3.0 eq.) was added dropwise and the mixture was allowed to warm to

room temperature over 1 h and stirred for an additional 1 h at room temperature. The reaction was monitored by TLC (EtOAc/pentane 1:2). Upon completion, the reaction was poured onto a water-ice mixture (5 mL) and stirred for 15 min. Next, CH₂Cl₂ (5 mL) was added and the layers were separated. The organic layer was washed with aq. Sat. NaHCO₃ (5 mL) and brine (5 mL). The organic layer was dried over MgSO₄, filtered and the solvent was evaporated. The bromo-sugar **14** (46.3 mg, 83.5 mmol, 82%) was obtained as slightly yellow oil and was used without further purification.

TLC (EtOAc:pentane, 1:2 v/v) = 0.79; **FT-IR** (film) \tilde{v} = 3531, 2981, 2938, 1738, 1568, 1458, 1403, 1371, 1314, 1277, 1247, 1126, 1116, 1076, 1046, 1001, 974, 932, 842, 792, 767, 748, 680 cm⁻¹; ¹**H NMR** (500 MHz, CDCl₃) δ 6.56 (m, 2H), 6.15 (ddt, J = 16.5, 10.3, 5.9 Hz, 1H), 6.04 (ddt, J = 16.5, 10.3, 5.9 Hz, 1H), 5.45 (dd, J = 17.1, 1.3 Hz, 1H), 5.37 (dd, J = 17.1, 1.3 Hz, 1H), 5.31 (dd, J = 10.2, 1.3 Hz, 1H), 5.26 (dd, J = 10.2, 1.3 Hz, 1H), 5.16 (t, J = 9.9 Hz, 1H), 4.62 – 4.45 (m, 4H), 4.31 (dd, J = 9.9 Hz, 3.6 Hz, 1H), 4.01 (dq, J = 9.9, 6.2 Hz, 1H), 3.81 (dd, J = 3.6, 1.4 Hz, 1H), 3.55 (s, 3H), 2.81 (m, 2H), 1.35 (d, J = 6.2 Hz, 3H), 1.21 (t, J = 7.5 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CDCl₃) δ 166.4, 153.4, 151.1, 139.2, 132.90, 132.85, 127.0, 125.7, 121.7, 119.1, 119.0, 85.9, 83.7, 75.9, 75.6, 74.5, 70.9, 68.5, 59.1, 25.3, 17.2, 14.2 ppm; **HRMS** ESI(+) (MeOH) calculated for C₂₂H₂₇BrCl₂O₇Na [M+Na]⁺: 575.02094, found: 575.02045.



In a flame dried flask under an atmosphere of argon, carboxylic acid (*E*)-13 (117 mg, 0.166 mmol, 1.0 eq.) was dissolved in dry THF (500 μ L) and *N*-methylmorpholine (36 μ L, 0.33 mmol, 2.0 eq.) was added. The mixture was cooled to 0 °C and isobutyl chloroformate (43 μ L, 0.33 mmol, 2.0 eq.) was added. The reaction mixture was to stirred for 1 h at 0 °C before it was filtered over a plug of Celite^â into a solution of NaBH₄ (7.5 mg, 0.198 mmol, 1.2 eq.) in H₂O (400 μ L) at 0 °C. It was stirred for 1 h at 0 °C and was then allowed warm to room temperature over 1 h.

 H_2O (2 mL) was added and the reaction was extracted with EtOAc (3 x 2 mL). The crude product was purified by silica-gel column chromatography (MeOH/CH₂Cl₂ 1:19 to 1:9) to obtain alcohol **15** (68.3 mg, 98.6 µmol, 60%) as a colorless solid. Hydrolyzed **S2** (7.0 mg, 11 µmol, 20%) was obtained as a byproduct.

<u>15</u>

TLC (MeOH:CH₂Cl₂, 1:9 v/v) = 0.44; **Specific Rotation** [α]_D²⁸°^C = +68.17 (c = 0.35, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3420, 2974, 2932, 2875, 1732, 1696, 1641, 1447, 1385, 1250, 1198, 1151, 1072, 1033, 898, 796, 511 cm⁻¹; ¹**H NMR** (500 MHz, CD₃OD) δ 7.06 (d, J = 11.3 Hz, 1H), 6.49 – 6.40 (m, 1H), 5.87 – 5.77 (m, 2H), 5.56 (t, J = 8.3 Hz, 1H), 5.13 (dt, J = 10.5, 1.6 Hz, 1H), 5.02 (d, J = 10.2 Hz, 1H), 4.71 (d, J = 1.2 Hz, 1H), 4.67 (q, J = 5.5 Hz, 1H), 4.21 (m, 1H), 4.00 (quint, J = 6.4 Hz, 1H), 3.92 (d, J = 2.9 Hz, 1H), 3.78 – 3.66 (m, 2H), 3.53 (t, J = 6.4 Hz, 2H), 2.75 – 2.63 (m, 3H), 2.59 (sept, J = 6.9 Hz, 1H), 2.50–2.43 (m, 4H), 2.06 – 1.97 (m, 1H), 1.81 (d, J = 1.3 Hz, 3H), 1.73 (d, J = 1.3 Hz, 3H), 1.67 – 1.65 (m, 3H), 1.65 – 1.58 (m, 2H), 1.33 – 1.24 (m, 1H), 1.20 – 1.09 (m, 15H), 0.88 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CD₃OD) δ 178.4, 170.1, 142.4, 140.8, 137.00, 136.95, 136.2, 134.7, 129.7, 128.8, 126.7, 124.5, 97.2, 94.3, 78.5, 75.9, 74.5, 73.6, 73.2, 70.5, 68.2, 62.3, 42.5, 37.2, 35.4, 33.2, 28.7, 28.3, 26.9, 24.0, 20.3, 19.5, 19.1, 18.7, 17.5, 15.4, 13.9, 11.3 ppm; **HRMS** ESI(+) (MeOH) calculated for C₃₈H₆₀O₁₁Na [M+Na]*: 715.40250, found: 715.40278.

<u>S2</u>

TLC (MeOH:CH₂Cl₂, 1:9 v/v) = 0.15; **Specific Rotation** [α]_D^{28°C} = +71.2 (c = 0.49, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3382, 2972, 2931, 2874, 1683, 1640, 1381, 1249, 1209, 1160, 1070, 1029, 977, 895, 787 cm⁻¹; ¹**H NMR** (500 MHz, CD₃OD) δ 7.07 (d, J = 11.4 Hz, 1H), 6.48 – 6.40 (m, 1H), 5.86 – 5.78 (m, 2H), 5.56 (t, J = 8.2 Hz, 1H), 5.13 (dd, J = 10.4, 1.8 Hz, 1H), 4.68 (q, J = 5.6 Hz, 1H), 4.65 (s, 1H), 4.23 – 4.19 (m, 1H), 4.01 (quint, J = 6.4 Hz, 1H), 3.88 (d, J = 3.2 Hz, 1H), 3.69 (d, J = 9.7 Hz, 1H), 3.56 – 3.50 (m, 3H), 3.47 (d, J = 10.0 Hz, 1H), 2.75 – 2.62 (m, 4H), 2.50 – 2.34 (m, 4H), 2.06 – 1.95 (m, 1H), 1.81 (s, 3H), 1.74 (s, 3H), 1.65 (s, 3H), 1.64 – 1.58 (m, 2H), 1.32 – 1.26 (m, 1H), 1.24 (s, 3H), 1.17 (d, J = 6.3 Hz, 3H), 1.07 (s, 3H), 0.88 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, CD₃OD) δ 170.1, 142.4, 140.7, 137.0, 136.9, 136.1, 134.7, 129.7, 128.9, 126.7, 124.6, 97.0, 94.0, 78.5, 75.9, 74.8, 73.6, 73.1, 72.2, 68.3, 62.3, 42.6, 37.2, 33.2, 30.7, 28.8, 28.4, 26.9, 23.4, 20.2, 17.8, 17.5, 15.4, 13.9, 11.3 ppm; **HRMS** ESI(+) (MeOH) calculated for C₃₄H₅₄O₁₀Na [M+Na]*: 645.36092, found: 645.36072.

In a flame dried flask under an atmosphere of argon, to a solution alcohol **15** (63.7 mg, 91.9 μ mol, 1.0 eq.) and bromide **14** (56.5 mg, 0.102 mmol, 1.1 eq.) in dry CH₂Cl₂ (8.0 mL), molecular sieves (4Å, 800 mg) was added and the suspension was stirred for 30 min at

room temperature. Then, Ag_2CO_3 (253 mg, 0.919 mmol, 10 eq.) was added and the reaction mixture was stirred for 24 h at room temperature. It was filtered over Celite^a and the solvent was evaporated. The crude product was purified by RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: $H_2O + 0.1\%$ HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 70%, 15 min – 70%, 45 min – 90%, 45.20 min –100%] to afford, after lyophilization, allyl-protected C2-elongated fidaxomicin **S3** ($t_R = 25.5$ min, 15.0 mg, 12.8 μ mol, 14%) as a colorless solid.

TLC (MeOH:CH₂Cl₂, 1:19 v/v) = 0.79; **Specific Rotation** $[\alpha]_D^{24}$ °C = +6.16 (c = 0.75, MeOH); **FT-IR** (film) $\tilde{\nu}$ = 3464, 2975, 2931, 2876, 1736, 1699, 1641, 1568, 1455, 1402, 1384, 1369, 1352, 1316, 1249, 1199, 1135, 1070, 1025, 995, 975, 932, 901, 791, 775, 750, 713 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.10 (d, J = 11.4 Hz, 1H), 6.51-6.44 (m, 1H), 6.17 (ddt, J = 17.2, 10.4, 5.8 Hz, 1H), 6.08 (ddt, J = 17.1, 10.4, 10.4)5.8 Hz, 1H), 5.90-5.82 (m, 2H), 5.61 (t, J = 8.4 Hz, 1H), 5.49-5.37 (m, 2H), 5.30-5.23 (m, 2H), 5.21 (dt, J = 8.4 Hz, 1 Hz), 5.49-5.37 (m, 2H), 5.30-5.23 (m, 2H), 5.21 (dt, J = 8.4 Hz, 1 Hz), 5.49-5.37 (m, 2H), 5.30-5.23 (m, 2H), 5.21 (dt, J = 8.4 Hz, 1 Hz), 5.49-5.37 (m, 2H), 5.30-5.23 (m, 2H), 5.21 (dt, J = 8.4 Hz, 1 Hz), 5.49-5.37 (m, 2H), 5.30-5.23 (m, 2H), 5.21 (dt, J = 8.4 Hz, 1 Hz)J = 10.5, 1.5 Hz, 1H), 5.03 (t, J = 9.7 Hz, 1H), 4.99 (d, J = 10.1 Hz, 1H), 4.77 (d, J = 1.3 Hz, 1H), 4.71-10.00 (d)4.66 (m, 1H), 4.64–4.57 (m, 4H), 4.64–4.57 (m, 1H), 4.29–4.21 (m, 1H), 4.07–4.00 (m, 2H), 3.97–3.94 (m, 1H), 3.90-3.85 (m, 2H), 3.81 (d, J = 3.6 Hz, 1H), 3.79-3.69 (m, 3H), 3.68 (d, J = 4.1 Hz, 1H),3.65-3.63 (m, 1H), 3.64 (s, 3H), 3.52-3.46 (m, 1H), 3.46-3.40 (m, 1H), 2.92-2.71 (m, 2H), 2.71-2.60 (m, 3H), 2.56 (sept, J = 7.0 Hz, 1H), 2.51-2.35 (m, 4H), 1.97-1.91 (m, 1H), 1.81 (d, J = 1.3 Hz, 3H),1.79 - 1.67 (m, 2H), 1.73 (d, J = 1.4 Hz, 3H), 1.65 (d, J = 1.3 Hz, 3H), 1.29 (d, J = 6.1 Hz, 3H), 1.26 - 1.18(m, 1H), 1.18–1.13 (m, 15H), 1.13 (s, 3H), 0.81 (t, J = 7.4 Hz, 3H) ppm; ¹³C NMR (126 MHz acetone- d_6) δ 176.8, 168.7, 166.5, 153.9, 151.9, 141.6, 140.6, 140.0, 136.8, 136.2, 136.0, 134.13, 134.07, 133.9, 129.3, 128.7, 128.5, 126.6, 126.0, 124.0, 122.3, 118.8, 118.7, 102.6, 96.7, 93.3, 82.0, 78.2, 77.6, 76.4, 75.7, 75.0, 73.7, 73.0, 72.8, 72.3, 70.6, 70.2, 69.4, 67.7, 61.9, 42.1, 37.3, 34.8, 30.1, 28.7, 28.2, 26.5, 25.6, 24.1, 20.6, 19.4, 19.2, 18.6, 18.3, 17.4, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for C₆₀H₈₆Cl₂O₁₈Na [M+Na]⁺: 1187.50834, found: 1187.50826

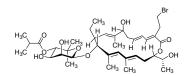
In a flame-dried flask under argon atmosphere, to a solution of allyl-protected C2-elongated fidaxomicin (**S3**, 15.0 mg, 12.9 μ mol, 1.0 eq.) in dry THF (1.2 mL) was added morpholine (1 μ L, 13 μ mol, 1.0 eq.). After cooling

the mixture to 0 °C, Pd(PPh₃)₄ (1.5 mg, 1.3 μ mol, 10 mol%) was added and it was stirred for 10 min. Then, the reaction mixture was diluted with EtOAc (2 mL) and washed with sat. aq. NH₄Cl (2 mL). The phases were separated and the aqueous layer was extracted with EtOAc (3 x 3 mL). The combined organic layers were dried over MgSO₄, filtered and the solvent was evaporated under reduced pressure. The crude residue was further purified by RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 60%, 5.0 min – 60%, 30.0 min – 70%, 30.5 min –100%] to afford, after lyophilization, C2-elongated fidaxomicin **17** (t_R = 13 min, 9.1 mg, 8.4 μ mol, 65%) as a colorless solid.

TLC (MeOH:CH₂Cl₂, 1:19 v/v) = 0.16; **Specific Rotation** [α]_{D^{4}} $^{\circ}C$ = +8.24 (c = 0.46, MeOH); **FT-IR** (film) $\tilde{\nu}$ = 3447, 2976, 2935, 2876, 1734, 1697, 1590, 1406, 1384, 1369, 1312, 1247, 1198, 1146, 1111, 1070, 1024, 975, 901, 873, 799, 760, 738, 720, 695, 582, 504, 477 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.10 (d, J = 11.4 Hz, 1H), 6.54–6.41 (m, 1H), 5.91–5.82 (m, 2H), 5.61 (t, J = 8.3 Hz, 1H), 5.21 (dt, J = 10.6, 1.5 Hz, 1H), 5.10 (t, J = 9.7 Hz, 1H), 4.99 (d, J = 10.1 Hz, 1H), 4.77 (d, J = 1.2 Hz, 1H), 4.71–4.66 (m, 1H), 4.65 (d, J = 0.8 Hz, 1H), 4.28–4.23 (m, 1H), 4.07–3.99 (m, 1H), 3.95 (d, J = 3.3 Hz, 1H), 3.91–3.79 (m, 3H), 3.76–3.65 (m, 3H), 3.63 (s, 3H), 3.62–3.56 (m, 1H), 3.50–3.43 (m, 1H), 3.04–2.96 (m, 2H), 2.79–2.61 (m, 3H), 2.56 (sept, J = 6.7 Hz, 1H), 2.51–2.36 (m, 4H), 1.99–1.90 (m, 1H), 1.81 (d, J = 1.3 Hz, 3H), 1.79–1.67 (m, 2H), 1.73 (d, J = 1.4 Hz, 3H), 1.65 (s, 3H), 1.27 (d, J = 6.1 Hz, 3H), 1.26–1.22 (m, 1H), 1.21 (t, J = 7.3 Hz, 3H), 1.17–1.12 (m, 12H), 1.09 (s, 3H), 0.83 (t, J = 7.4 Hz, 3H) ppm; 13C NMR (126 MHz, acetone- d_6) δ 176.8, 169.4, 168.7, 155.7, 153.8, 142.6, 141.6, 140.7, 136.8, 136.2, 136.0, 133.9, 129.3, 128.4, 126.5, 124.0, 114.5, 110.8, 108.2, 102.6, 96.7, 93.3, 81.8, 78.2, 77.7, 75.7, 73.7, 72.9, 72.8, 72.2, 70.5, 70.1, 69.4, 67.6, 61.8, 42.1, 37.3, 34.8, 30.1, 28.7, 28.2, 26.5, 26.2, 24.0, 20.6, 19.4, 19.2, 18.6, 18.1, 17.4, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for C₆₀H₈₆Cl₂O₁₈Na [M+Na]*: 1107.44574, found: 1107.44501.



Supplementary Figure 1. Observed NOESY correlations in glycosyl donor 14 and C2-elongated 17.

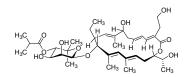


In a flame-dried flask under argon atmosphere, carboxylic acid (*E*)-13 (122 mg, 0.173 mmol, 1.0 eq.) and HOTT (95.8 mg, 0.258 mmol, 1.5 eq.) were dissolved in dry CH_2Cl_2 (1.8 mL). Next, DIPEA (71 μ L, 0.43 mmol, 2.5 eq.) was added and flask was covered in aluminum foil. The reaction

mixture was stirred at room temperature for 1 h, the aluminum foil was removed and CCl₃Br (1.2 mL) was added. The reaction was allowed to stir for 15 h at room temperature, the solvent was evaporated and the residue was purified by silica-gel column chromatography (EtOAc/pentane 3:2). The bromide **16** (79.1 mg, 0.107 mmol, 62%) was obtained as a slightly yellow solid.

TLC (EtOAc:pentane, 1:1 v/v) = 0.13; **Specific Rotation** [α]_D²⁶ $^{\circ}$ C = +17.7 (c = 0.38, MeOH); **FT-IR** (film) $\tilde{\nu}$ = 3441, 2974, 2933, 2873, 1735, 1698, 1640, 1446, 1385, 1297, 1249, 1198, 1153, 1072, 1033, 898, 844, 798, 504 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.18 (d, J = 11.4 Hz, 1H), 6.56-6.38 (m, 1H), 5.92 (ddd, J = 14.6, 9.7, 4.5 Hz, 1H), 5.82 (s, 1H), 5.61 (t, J = 8.2 Hz, 1H), 5.21 (dt, J = 10.7, 1.6 Hz, 1H), 4.99 (d, J = 10.1 Hz, 1H), 4.78 (d, J = 1.2 Hz, 1H), 4.73-4.65 (m, 1H), 4.33-4.21 (m, 1H), 4.09-3.99 (m, 2H), 3.98-3.94 (m, 1H), 3.85 (d, J = 3.6 Hz, 1H), 3.76-3.70 (m, 2H), 3.68 (d, J = 4.2 Hz, 1H), 3.54-3.44 (m, 2H), 3.32 (d, J = 9.2 Hz, 1H), 2.99-2.80 (m, 2H), 2.80-2.60 (m, 3H), 2.56 (sept, J = 7.0 Hz, 1H), 2.49 (ddd, J = 14.8, 9.7, 4.2 Hz, 1H), 2.42 (ddd, J = 14.0, 9.1, 4.4 Hz, 1H), 1.99-1.89 (m, 1H), 1.80 (d, J = 1.3 Hz, 3H), 1.71 (d, J = 1.4 Hz, 3H), 1.65 (s, 3H), 1.29-1.22 (m, 1H), 1.19-1.11 (m, 12H), 1.09 (s, 3H), 0.83 (t, J = 7.4 Hz, 3H) ppm; ¹³C NMR (126 MHz, acetone- d_6) δ 176.8, 168.2, 143.7, 142.4, 136.8, 136.2, 136.0, 133.9, 128.1, 126.31, 126.25, 123.9, 96.7, 93.3, 78.3, 75.7, 73.8, 72.89, 72.83, 70.1, 67.6, 42.0, 37.2, 34.7, 32.1, 31.2, 28.7, 28.2, 26.5, 20.7, 19.4, 19.2, 18.6, 17.4, 15.2, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for C₃₇H₅₇BrO₁₀Na [M+Na]⁺: 763.30273, found: 763.30285.

16.1



Bromide **16** (52.9 mg, 71.3 μ mol, 1.0 eq.) was dissolved in MeCN/H₂O (1:1, 1.4 mL) and AgF (18.1 mg, 0.143 mmol, 2.0 eq.) was added. The reaction mixture was stirred at 50 °C for 14 h before it was filtered over Celite^â and the solvent was removed under reduced pressure. The

residue was redissolved in MeCN (2 mL) and filtered again. The solvent was evaporated and the residue was purified by RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min – % B): 0.0 min – 35%, 10.0 min – 35%, 45.0 min – 45%, 45.2 min – 100%] to afford, after lyophilization, alcohol **16.1** (t_R = 18 min, 10.7 mg, 15.7 μ mol, 44%) as a colorless solid.

TLC (MeOH:CH₂Cl₂, 1:19 v/v) = 0.12; **Specific Rotation** [α]_D²⁶°^c = +36.0 (c = 0.36, MeOH); **FT-IR** (film) $\tilde{\nu}$ = 3423, 2975, 2933, 2876, 1734, 1695, 1641, 1443, 1386, 1370, 1296, 1252, 1202, 1151, 1085, 1035, 975, 951, 899, 797 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) 7.11 (d, J = 11.4 Hz, 1H), 6.46 (dd, J = 15.0, 11.4 Hz, 1H), 5.88–5.80 (m, 2H), 5.59 (t, J = 8.3 Hz, 1H), 5.19 (d, J = 10.5 Hz, 1H), 4.99 (d, J = 10.1 Hz, 1H), 4.77 (s, 1H), 4.67 (q, J = 5.5 Hz, 1H), 4.25–4.22 (m, 1H), 4.02 (quint, J = 6.3 Hz, 1H), 3.95 (d, J = 3.2 Hz, 1H), 3.74–3.69 (m, 2H), 3.54 (t, J = 6.8 Hz, 2H), 2.78–2.43 (m, 7H), 2.39 (ddd, J = 13.8, 9.0, 4.4 Hz, 1H), 1.98–1.91 (m, 1H), 1.80 (s, 3H), 1.72 (s, 3H), 1.65 (s, 3H), 1.31–1.20 (m, 1H), 1.16–1.11 (m, 12H), 1.08 (s, 3H), 0.82 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 176.8, 169.1, 142.6, 140.7, 136.9, 136.2, 135.9, 133.9, 128.6, 126.7, 126.6, 123.9, 96.7, 93.3, 78.2, 75.7, 73.7, 72.9, 72.8, 70.2, 67.6, 61.9, 42.1, 37.2, 34.8, 31.4, 28.7, 28.2, 26.5, 20.6, 19.4, 19.1, 18.6, 17.4, 15.2, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeCN) calculated for C₃₇H₅₈O₁₁Na [M+Na]*: 701.38713, found: 701.38334.

In a flame-dried flask under argon atmosphere, molecular sieves (4Å, 200 mg) were added to a solution of alcohol **16.1** (15.8 mg, 23.3 μ mol, 1.0 eq.) and bromide **14** (33.0 mg, 59.5 μ mol, 2.5 eq.) in dry CH₂Cl₂ (2.0 mL), and the suspension was stirred for 30 min at

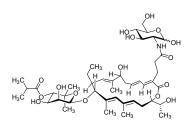
room temperature. Next, Ag₂CO₃ (64.2 mg, 0.233 mmol, 10 eq.) was added and the reaction mixture was stirred for 3 h at 40 °C. Subsequently, it was diluted with CH₂Cl₂ (2 mL) and filtered over Celite^å. The crude product was purified by RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 68%, 15.0 min – 68%, 45.0 min – 73%, 45.2 min –100%] to afford, after lyophilization, allyl-protected C1-elongated fidaxomicin **S4** (t_R = 25.8 min, 19.7 mg, 4.6 μ mol, 20%) as a colorless solid.

TLC (MeOH:CH₂Cl₂, 1:19) = 0.28; **Specific Rotation** $[\alpha]_D^{25}$ ° c = -7.02 (c = 0.26, MeOH); **FT-IR** (film) $\tilde{\nu}$ = 3462, 2976, 2934, 2877, 1735, 1698, 1642, 1568, 1403, 1385, 1369, 1353, 1318, 1250, 1199, 1135, 1073, 1028, 975, 900 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.17 (d, J = 11.4 Hz, 1H), 6.50 (dd, J = 11.4 Hz, 1H 14.9, 11.8 Hz, 1H), 6.17 (ddt, J = 17.3, 10.4, 5.8 Hz, 1H), 6.08 (ddt, J = 17.1, 10.6, 5.8 Hz, 1H), 5.88 (ddd, J = 14.7, 9.5, 4.7 Hz, 1H), 5.82 (s, 1H), 5.62 (t, J = 8.3 Hz, 1H), 5.49 - 5.36 (m, 2H), 5.31 - 5.23(m, 2H), 5.21 (dt, J = 10.7, 1.8 Hz, 1H), 5.01 (t, J = 9.7 Hz, 1H), 4.98 (d, J = 10.1 Hz, 1H), 4.77 (d, J = 10.1 Hz, 1H), 4.78 (d, J = 10.1 Hz, 1H), 4.77 (d, J = 10.1 Hz, 1H), 4.78 (d, J = 10.1 Hz, 1H), 4.77 (d, J = 10.1 Hz, 1H), 4.78 (d, J = 10.1 Hz, 1H), 4.781.3 Hz, 1H), 4.69 (q, J = 5.2 Hz, 1H), 4.63 - 4.58 (m, 4H), 4.54 - 4.49 (m, 1H), 4.28 - 4.23 (m, 1H), 4.07-4.00 (m, 2H), 3.97 - 3.94 (m, 1H), 3.89 - 3.82 (m, 3H), 3.76 - 3.67 (m, 4H), 3.61 - 3.55 (m, 2H), 3.54(s, 3H), 3.53 – 3.45 (m, 1H), 3.29 (d, J = 9.4 Hz, 1H), 2.90 – 2.77 (m, 2H), 2.76 – 2.60 (m, 5H), 2.56 (sept, J = 7.0 Hz, 1H), 2.48 (ddd, J = 14.8, 9.5, 4.1 Hz, 1H), 2.41 (ddd, J = 13.9, 9.2, 4.4 Hz, 1H), 1.98 – 1.89 (m, 1H), 1.80 (d, J = 1.3 Hz, 3H), 1.72 (s, 3H), 1.65 (s, 3H), 1.29 (d, J = 6.1 Hz, 3H), 1.27 - 1.21 (m, 1.20 Hz, 1.201H), 1.17 – 1.12 (m, 15H), 1.09 (s, 3H), 0.83 (t, J = 7.4 Hz, 3H) ppm; ; ¹³C NMR (126 MHz, acetone- d_6) δ 176.8, 168.7, 166.5, 153.9, 151.9, 143.0, 141.2, 140.0, 136.9, 136.2, 136.0, 134.13, 134.05, 133.9, 128.7, 128.6, 126.4, 126.1, 126.0, 123.9, 122.3, 118.8, 118.7, 102.5, 96.8, 93.3, 81.8, 78.2, 77.5, 76.4, 75.7, 75.0, 73.8, 72.9, 72.8, 72.3, 70.6, 70.2, 69.1, 67.6, 61.8, 42.0. 37.2, 34.8, 28.7, 28.2, 28.1, 26.5, 25.6, 20.7, 19.4, 19.2, 18.6, 18.3, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(-) (MeCN) calculated for $C_{60}H_{85}Cl_2O_{20}$ [M+HCOO]⁻: 1195.50167, found: 1195.50218.

In a flame-dried flask under argon atmosphere, morpholine (0.4 μ L, 4.5 μ mol, 1.0 eq.) was added to a solution of allyl-protected C1-elongated fidaxomicin **S4** (5.2 mg, 4.5 μ mol, 1.0 eq.) in dry THF (450 μ L). This

mixture was cooled to 0 °C and Pd(PPh₃)₄ (0.5 mg, 0.4 μ mol, 10 mol%) was added and the reaction was stirred for 20 min. Next, the reaction mixture was diluted with EtOAc (2 mL) and washed with sat. aq. NH₄Cl (2 mL). The phases were separated and the aqueous layer was extracted with EtOAc (3 x 2 mL). The combined organic layers were dried over MgSO₄, filtered and the solvent was evaporated under reduced pressure. The crude residue was further purified by RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 60%, 5.0 min – 60%, 30.0 min – 70%, 30.5 min – 100%] to afford, after lyophilization, C1-elongated fidaxomicin **18** (t_R = 10.5 min, 3.6 mg, 3.4 μ mol, 75%) as a colorless solid.

Specific Rotation [α]_D^{24°C} = -6.94 (c = 0.72, MeOH); **FT-IR** (film) $\tilde{\nu}$ = 3432, 2976, 2933, 2876, 1733, 1697, 1589, 1385, 1370, 1313, 1250, 1200, 1073, 1028, 899, 799, 761 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 7.17 (d, J = 11.4 Hz, 1H), 6.51 (dd, J = 15.0, 11.4 Hz, 1H), 5.89 (ddd, J = 14.6, 9.4, 4.7 Hz, 1H), 5.82 (s, 1H), 5.62 (t, J = 8.3 Hz, 1H), 5.22 (dt, J = 10.6, 1.7 Hz, 1H), 5.09 (t, J = 9.7 Hz, 1H), 4.99 (d, J = 10.1 Hz, 1H), 4.77 (d, J = 1.2 Hz, 1H), 4.70 (q, J = 5.2 Hz, 1H), 4.67 (s, 1H), 4.29 – 4.23 (m, 1H), 4.07 – 3.98 (m, 2H), 3.95 (d, J = 3.5 Hz, 1H), 3.90 – 3.84 (m, 1H), 3.83 – 3.78 (m, 2H), 3.74 – 3.68 (m, 3H), 3.63 – 3.56 (m, 3H), 3.54 (s, 3H), 3.00 (q, J = 7.3 Hz, 2H), 2.76 – 2.60 (m, 5H), 2.56 (sept, J = 7.0 Hz, 1H), 2.52 – 2.38 (m, 2H), 1.98 – 1.89 (m, 1H), 1.80 (s, 3H), 1.72 (s, 3H), 1.65 (s, 3H), 1.27 (d, J = 6.2 Hz, 3H), 1.26 – 1.23 (m, 1H), 1.21 (t, J = 7.4 Hz, 3H), 1.18 – 1.11 (m, 12H), 1.09 (s, 3H), 0.83 (t, J = 7.4 Hz, 3H) ppm; ¹³**C NMR** (126 MHz, acetone- d_6) δ 176.77, 169.41, 168.63, 155.80, 153.81, 143.09, 142.60, 141.28, 136.86, 136.14, 136.04, 133.84, 128.54, 126.30, 126.08, 123.93, 114.51, 110.66, 108.19, 102.53, 96.76, 93.35, 81.61, 78.16, 77.63, 75.70, 73.76, 72.92, 72.82, 72.24, 70.52, 70.15, 69.08, 67.62, 61.77, 42.00, 37.16, 34.78, 28.72, 28.24, 28.09, 26.52, 26.20, 20.74, 19.41, 19.17, 18.59, 18.17, 17.48, 15.19, 14.37, 13.75, 11.17 ppm; **HRMS** ESI(-) (MeCN) calculated for C₅₃H₇₅Cl₂O₁₈ [M–H]⁻: 1069.43359, found: 1069.43427.



In a flame-dried flask under argon atmosphere, carboxylic acid (*E*)-13 (18.3 mg, 25.9 μ mol, 1.0 eq.), D-glucosamine hydrochlorid (8.5 mg, 39 μ mol, 1.5 eq.) and HATU (14.8 mg, 38.8 μ mol, 1.5 eq.) were dissolved in dry DMF (0.2 mL). Next, freshly distilled DIPEA (11 μ L, 63 μ mol, 2.4 eq.) was added and the resulting mixture was stirred at room temperature for 3 h. It was then quenched with water (2 mL) and

lyophilized. The crude residue was purified by preparative RP-HPLC [Gemini NX, C18, 5 μ , 110 Å, 250 mm × 21.2 mm, solvent A: H₂O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH, 20 mL/min; LC time program (min –% B): 0.0 min – 30%, 15.0 min – 30%, 45.0 min – 35%, 45.2 min –100%] to afford, after lyophilization, FDX-glucosamide **19** (a/b=3:1, t_R = 16.8 and 18.5 min, 15.8 mg, 18.2 μ mol, 70%) as a colorless solid.

TLC (MeOH:CH₂Cl₂. 1:4 v/v) = 0.38; **Specific Rotation** $[\alpha]_D^{25}$ °C = +23.70 (c = 0.75, MeOH); **FT-IR** $\tilde{\nu}$ (film) 363, 3404, 2974, 2939, 1680, 1643, 1539, 1386, 1307, 1250, 1212, 1147, 1077, 1037, 849, 559 cm⁻¹; α : ¹H NMR (500 MHz, CD₃OD) δ 7.08 (d, J = 11.4 Hz, 1H), 6.52 – 6.44 (m, 1H), 5.87 – 5.80 (m, 2H), 5.55 (t, J = 8.3 Hz, 1H), 5.12 (d, J = 11.5 Hz, 1H), 5.10 (d, J = 3.5 Hz, 1H), 5.02 (d, J = 10.2 Hz, 1H), 4.73 - 4.70 (m, 1H), 4.67 (d, J = 5.0 Hz, 1H), 4.22 (s, 1H), 4.04 (quint, J = 6.3 Hz, 1H), 3.92 (dd, J = 3.3, 1.2 Hz, 1H, 3.82 - 3.78 (m, 2H), 3.74 - 3.66 (m, 4H), 3.35 (t, J = 9.2 Hz, 1H), 2.76 - 2.55 (m, 2H)6H), 2.51 – 2.43 (m, 1H), 2.43 – 2.31 (m, 3H), 2.05 – 1.98 (m, 1H), 1.82 – 1.79 (m, 3H), 1.75 (s, 3H), 1.66 (s, 3H), 1.33 - 1.24 (m, 1H), 1.20 - 1.11 (m, 15H), 0.88 (t, J = 7.4 Hz, 3H) ppm; ¹³C NMR (126 MHz, 126 MHz)CD₃OD) δ 178.4, 175.5, 169.9, 143.0, 141.3, 137.16, 137.0, 136.1, 134.7, 128.9, 128.64, 126.9, 124.6, 97.2, 94.3, 92.5, 78.6, 76.0, 74.5, 73.6, 73.2, 73.1, 72.6, 72.5, 70.6, 68.2, 62.9, 56.0, 42.5, 37.3, 36.5, 35.4, 28.7, 28.1, 26.9, 24.2, 20.1, 19.5, 19.1, 18.7, 17.5, 15.4, 13.9, 11.3 ppm; β: ¹H NMR (500 MHz, CD₃OD) δ 7.08 (d, J = 11.4 Hz, 1H), 6.52 - 6.44 (m, 1H), 5.87 - 5.80 (m, 2H), 5.55 (t, J = 8.3 Hz, 1H), 5.12 (d, J = 11.5 Hz, 1H), 5.02 (d, J = 10.2 Hz, 1H), 4.73 – 4.70 (m, 1H), 4.67 (d, J = 5.0 Hz, 1H), 4.58 $(d, J = 8.3 \text{ Hz}, 1\text{H}), 4.22 \text{ (s, 1H)}, 4.04 \text{ (quint, } J = 6.3 \text{ Hz}, 1\text{H}), 3.92 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{H}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{H}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{H}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{H}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{H}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{H}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{H}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{Hz}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{Hz}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz}, 1\text{Hz}), 3.86 \text{ (dd, } J = 3.3, 1.2 \text{ Hz$ J = 12.0, 2.1 Hz, 1H, 3.66 (d, J = 5.5 Hz, 1H), 3.62 - 3.54 (m, 1H), 3.46 - 3.41 (m, 1H), 3.30 - 3.27 (m, 1H)1H), 2.76 - 2.55 (m, 6H), 2.51 - 2.43 (m, 1H), 2.43 - 2.31 (m, 3H), 2.05 - 1.98 (m, 1H), 1.82 - 1.79 (m, 3H), 1.75 (s, 3H), 1.66 (s, 3H), 1.33 – 1.24 (m, 1H), 1.20 – 1.11 (m, 15H), 0.88 (t, J = 7.4 Hz, 3H) ppm; ¹³C NMR (126 MHz, CD₃OD) δ 178.4, 175.5, 169.9, 142.9, 141.3, 137.16, 137.0, 136.1, 134.7, 128.9, 128.58, 126.8, 124.6, 97.2, 97.1, 94.3, 78.6, 76.10, 76.0, 74.5, 73.6, 73.2, 72.2, 70.6, 68.2, 62.9, 58.9, 42.5, 37.3, 36.8, 35.4, 28.7, 28.1, 26.9, 24.2, 20.1, 19.5, 19.1, 18.7, 17.5, 15.4, 13.9, 11.3 ppm; **HRMS** ESI(+), (MeOH) calculated for C₄₄H₆₉NO₁₆Na [M+Na]⁺: 890.45086, found: 890.45082.

Alcohol **15** (20.0 mg, 28.9 μ mol, 1.0 eq.) and Bis(2,4-dichlorophenyl) chlorophosphate (35.2 mg, 86.7 μ mol, 3.0 eq.) was dissolved in CH₂Cl₂ (200 μ L). Then DIPEA (19 μ L, 116 μ mol. 4.0 eq.) was added and the reaction mixture was

stirred for 1 h at room temperature. Upon completion of the reaction as monitored by TLC (MeOH/CH₂Cl₂ 1:19), the reaction was quenched with water and the layers were separated. The organic layer was extracted with water (3x). The combined organic layers were dried over MgSO₄, filtered and the solvent was evaporated under a nitrogen flow. The resulting crude phosphate ester was dissolved in DMF (200 μ L), NaN₃ (5.6 mg, 87 μ mol, 3.0 eq.) was added and the reaction mixture was heated to 80 °C for 1.5 h. Upon completion of the reaction as indicated by UHPLC-MS, the reaction mixture was diluted with CH₂Cl₂ and quenched with water. The layers were separated and the organic layer was washed with brine (2x). The combined organic layers were dried over MgSO₄, filtered and the solvent was evaporated under reduced pressure. The resulting azide was dissolved in CH₂Cl₂ (500 μ L) and the reaction mixture was degassed by freeze-pump-thaw (3x). Then, Cu(I)-cat. **21**² (3.9 mg, 5.5 μ mol, 20 mol%) and 1-Ethynyl-4-methylbenzene (7 μ L, 55 μ mol, 2.0 eq.) were added and the reaction was stirred for 4 h at room temperature. The reaction was diluted with CH₂Cl₂ and washed with brine (3x). The organic layer was dried over MgSO₄, filtered and the solvent was evaporated. The crude product was purified by silica-gel column chromatography (MeOH/CH₂Cl₂ 1:39) to give triazole **20** (4.9 mg, 5.9 μ mol, 21%) as a white solid.

or

Alcohol **15** (10.0 mg, 14.4 µmol, 1.0 eq.) was dissolved in toluene (500 µL) and a solution of DPPA-NO $_2^{[2]}$ (52.6 mg, 144 mol, 10 eq.) and DBU (25.8 µL, 173 µmol, 12 eq.) in toluene (1.5 mL) was added dropwise at room temperature. The reaction mixture was stirred at 50°C until completion of the reaction was observed by UHPLC-MS (2h30). The reaction mixture was concentrated under reduced pressure and the resulting crude mixture was dissolved in CH $_2$ Cl $_2$ (250 µL) followed by freeze-pump-thaw (3x). Then, Cu(I)-cat. **21** 2 (5.1 mg, 7.2 µmol, 50 mol%) and 1-Ethynyl-4-methylbenzene (3.65 µL, 29 µmol, 2.0 eq.) were added and the reaction was stirred for 4 h at room temperature. The reaction was diluted with CH $_2$ Cl $_2$ and washed with brine (3x). The layers were separated and the organic layer was washed with brine (2x). The combined organic layers were dried over MgSO $_4$, filtered and the solvent was evaporated under reduced pressure. The crude product was purified by silica-gel column chromatography (MeOH/CH $_2$ Cl $_2$ 1:39) to give triazole **20** (6.3 mg, 8 µmol, 53%) as a slightly yellow solid.

Specific Rotation [α] $_D^{23}$ ° $_C$ = 170.0 (c = 1.15, MeOH); **FT-IR** $\tilde{\nu}$ (film) cm⁻¹; 3452, 2973, 2930, 2874, 1735, 1697, 1641, 1500, 1384, 1368, 1317, 1249, 1202, 1150, 1079, 1034, 900, 820, 797 cm⁻¹; ¹**H NMR** (500 MHz, acetone- d_6) δ 8.28 (s, 1H), 7.78 (d, J = 8.1 Hz, 2H), 7.25 (d, J = 7.9 Hz, 2H), 7.03 (d, J = 11.4 Hz, 1H), 6.39 – 6.29 (m, 1H), 5.87 – 5.77 (m, 2H), 5.56 (t, J = 8.3 Hz, 1H), 5.18 (d, J = 10.6 Hz, 1H), 4.98 (d, J = 10.1 Hz, 1H), 4.75 (s, 1H), 4.69 – 4.64 (m, 1H), 4.45 (t, J = 7.0 Hz, 2H), 4.25 – 4.21

-

^[2] DPPA-NO₂ was prepared according to the procedure of Shioiri and Yamada.⁹

(m, 1H), 4.12 (d, J = 5.2 Hz, 1H), 4.06 (quint, J = 6.0 Hz, 1H), 3.96 - 3.92 (m, 1H), 3.81 (d, J = 3.6 Hz, 1H), 3.75 - 3.66 (m, 3H), 3.62 - 3.59 (m, 1H), 3.25 (d, J = 9.5 Hz, 1H), 2.78 - 2.73 (m, 2H), 2.67 - 2.36 (m, 6H), 2.35 (s, 3H), 2.19 - 2.08 (m, 2H), 1.99 - 1.90 (m, 1H), 1.78 (d, J = 1.3 Hz, 3H), 1.70 (d, J = 1.3 Hz, 3H), 1.64 (s, 3H), 1.26 - 1.21 (m, 0H), 1.14 (dd, J = 8.5, 6.2 Hz, 12H), 1.07 (s, 3H), 0.82 (t, J = 7.5 Hz, 3H) ppm. ¹³C NMR (126 MHz, acetone- d_6) δ 176.77, 168.63, 147.86, 141.87, 141.11, 138.17, 136.90, 136.30, 135.86, 133.96, 130.19, 129.63, 128.31, 128.24, 126.74, 126.20, 123.99, 121.10, 96.68, 93.27, 78.23, 75.72, 73.72, 72.99, 72.81, 70.18, 67.62, 50.39, 42.11, 37.29, 34.78, ~30 (under solvent signal), 28.70, 27.92, 26.47, 24.55, 21.22, 20.43, 19.41, 19.17, 18.61, 17.38, 15.24, 13.83, 11.18 ppm; HRMS ESI(+), (MeOH) calculated for $C_{47}H_{68}N_3O_{10}Na$ [M+H]*: 834.48992, found: 834.49000.

Determination of the Minimum Inhibitory Concentration

General procedure for the determination of MIC values of M. tuberculosis

MIC determination was carried out by Daniel Schäfle in the research group of Prof. Dr. Peter Sander (University of Zurich). MIC determination was essentially conducted as described recently.³ Briefly, the Green-Fluorescent Protein (GFP) expressing recombinant Mycobacterium tuberculosis H37Rv rpsL⁴ transformed with pOLYG-Pr-GFP5 was grown in Middlebrook 7H9-OADC with 0.05% Tween 80 until mid-log phase (optical density at 600 nm $OD_{600} = 0.3 - 1.0$), diluted to an OD_{600} of 0.04 and 20 μ l of the suspension were added to an equal volume of 12-point 2-fold serial dilutions of the compounds in 7H9-OADC-Tween in 384-well plates in triplicates. Compound concentrations were in the range of 62.5 to 0.031 µM. Fluorescence was measured immediately after inoculation (background) and after 10 days of incubation at 37 °C. Dose response curves were fitted with a 4-parameter log-normal model. P_{MIN} [-,-] and PMAX [-, 120] are the minimum and the maximum, respectively, PHIII [0,-] indicates the steepness, and EC₅₀ [-,-] the log-back transformed Minimal Effective Concentration 50. The computational and statistical analysis was conducted with R (3.0.1 - 3.1.1; https://www.r-project.org/). Dose response curves were fitted with the 'drc' package. The inhibitory potency I was calculated with the equation I = 100-[100●(S-P)/N-P)]. S is the sample's fluorescence while P and N derive from growth inhibition with the control drug (Kanamycin A) and solvent growth control measurements (DMSO 1.25% vol./vol.), respectively. A fluorescence reduction of 90% as compared to the no-drug control was reported as Minimal Inhibitory Concentration (MIC₉₀).

General procedure for the determination of MIC values of C. difficile

MIC determination was carried out by Micromyx, LLC, 4717 Campus Drive, Kalamazoo, MI, USA 49008. Approximately 5 mg of each of 20 test compounds were provided. These were stored at -20°C until testing. On the day of the assay, the test articles were dissolved in 100% DMSO (dimethyl sulfoxide, Sigma; St. Louis, MO, Cat. No. 472301-500ML, Lot No. SHBH5551V) to a stock concentration of 3232 μ g/mL. The concentration range tested for these test agents was 16 – 0.015 μ g/mL. The comparator agents, metronidazole and clindamycin were supplied by Micromyx, as shown in the table below:

Comparator Drug	Supplier	Catalog No.	Lot No.	Solvent/Diluent	Testing Range (µg/mL)
Metronidazole	Sigma	M3761-100G	095K0693	DMSO/dH2O	64 – 0.06
Clindamycin	Sigma	C5269-100MG	021M1533	dH2O/dH2O	32 – 0.03

Test Organisms

Test organisms consisted of reference strains from the American Type Culture Collection (ATCC; Manassas, VA) and clinical isolates from the Micromyx repository (MMX; Kalamazoo, MI). Organisms were initially received at Micromyx and were streaked for isolation. Colonies were picked by sterile swab from the medium and suspended in the appropriate broth containing cryoprotectant. The suspensions were aliquoted into cryogenic vials and maintained at -80° C.

Prior to testing, all isolates were streaked onto Brucella Agar supplemented with hemin, Vitamin K and 5% sheep blood (Becton Dickinson [BD]; Sparks, MD, Cat. No. 297716, Lot No. 8256909) and incubated anaerobically at 35 – 37°C for 44 – 48 h.

Additionally, *B. fragilis* ATCC 25285 and *C. difficile* ATCC 700057 were tested for purposes of quality control.

Test Medium

The medium employed for anaerobic testing in the broth microdilution MIC assay was Brucella Broth (BD, Cat. No. 211088, Lot No. 7128995), supplemented with hemin (Sigma, Lot No. SLBP5720V), Vitamin K (Sigma, Lot No. MKCG2075) and 5% laked horse blood (LHB, Cleveland Scientific; Bath, OH, Lot No. 474990).

Broth Microdilution Assay

The MIC assay method followed the procedure described by the CLSI^{6,7} and employed automated liquid handlers (Multidrop 384, Labsystems, Helsinki, Finland; Biomek 2000 and Biomek FX, Beckman Coulter, Fullerton CA) to conduct serial dilutions and liquid transfers. The wells in columns 2 through 12 in a standard 96-well microdilution plate (Costar) were filled with 150 μ L of the appropriate diluent (DMSO for the test agents; dH2O for metronidazole and clindamycin). The drugs (300 μ L at 101X the desired top concentration in the test plates) were dispensed into the appropriate well in column 1 of the mother plates. The Biomek 2000 was used to make serial 2-fold dilutions through column 11 in the "mother plate". The wells of column 12 contained no drug and were the organism growth control wells.

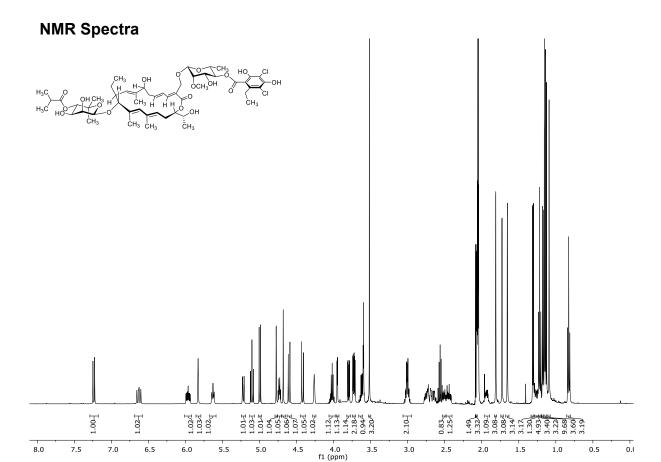
The daughter plates for testing of all isolates were loaded with 190 μ L per well of supplemented Brucella broth with 5% LHB using the Multidrop 384. The daughter plates were prepared on the Biomek FX instrument which transferred 2 μ L of 101X drug solution from each well of a mother plate to the corresponding well of each daughter plate in a single step. The wells of the daughter plates ultimately contained 190 μ L of medium, 2 μ L of drug solution, and 10 μ L of bacterial inoculum prepared in broth.

A standardized inoculum of each organism was prepared per CLSI methods.^{6,7} For all bacteria, suspensions were prepared in supplemented Brucella broth supplemented with hemin and Vitamin K to equal the turbidity of a 0.5 McFarland standard. These suspensions were further diluted 1:10 in supplemented Brucella broth with 5% LHB. The inoculum was dispensed into sterile reservoirs (Beckman Coulter) and transferred by hand in the Bactron Anaerobe chamber so that inoculation took place from low to high drug concentration. A 10 μ L aliquot of inoculum was delivered into each well. Inoculated daughter plates were stacked and placed in an anaerobic box with GasPak sachets (BD; Lot No. 6309689), covered with a lid on the top plate, and incubated at 35 – 37°C.

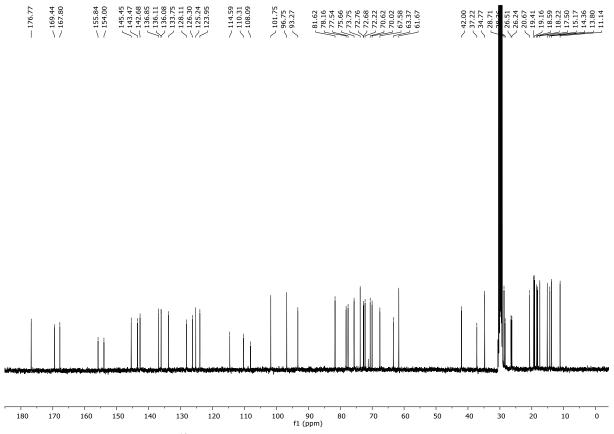
The microplates were viewed from the bottom using a plate viewer after 46 h. For each mother plate, an un-inoculated solubility control plate was observed for evidence of drug precipitation. The MIC was read and recorded as the lowest concentration of drug that inhibited visible growth of the organism.

Supplementary References

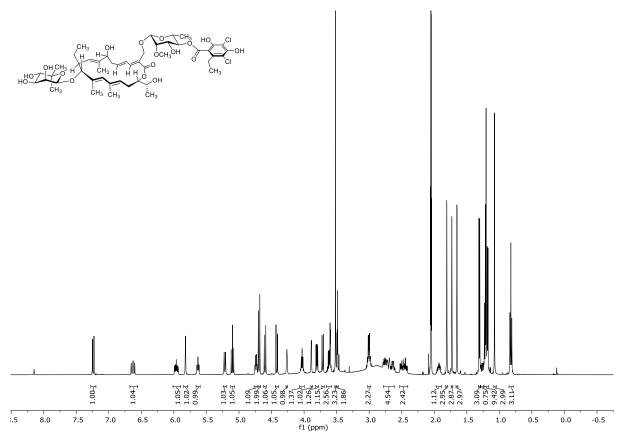
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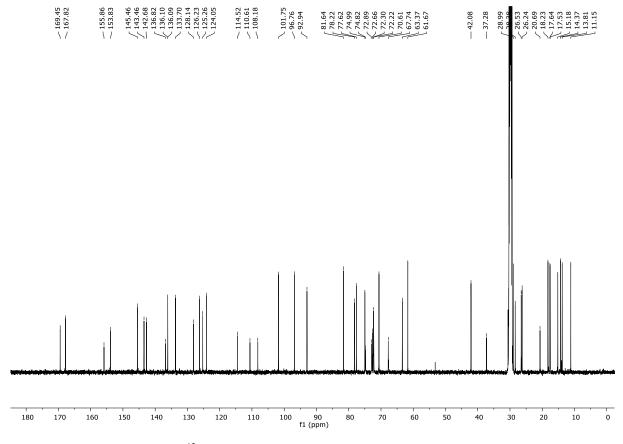
Supplementary Figure 2. ¹H NMR (500 MHz, acetone-*d*₆) of Fidaxomicin (1).



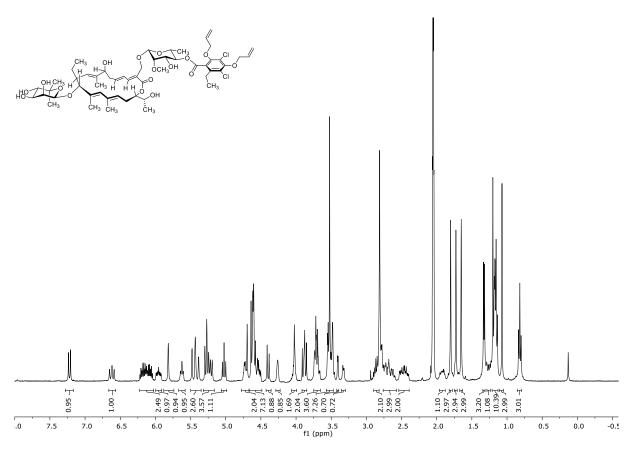
Supplementary Figure 3. ¹³C NMR (101 MHz, acetone-*d*₆) of Fidaxomicin (1).



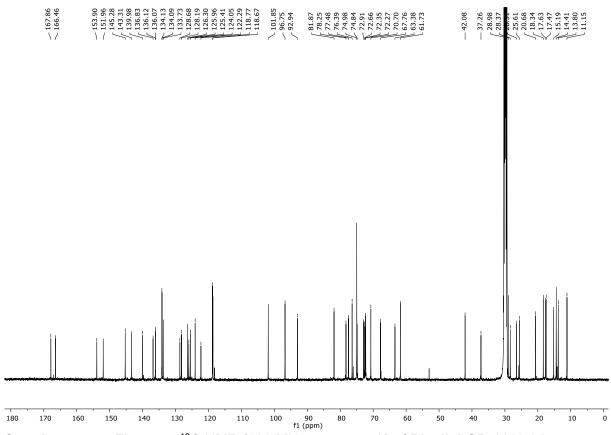
Supplementary Figure 4. 1 H NMR (500 MHz, acetone- d_{6}) of OP-1118 (4).



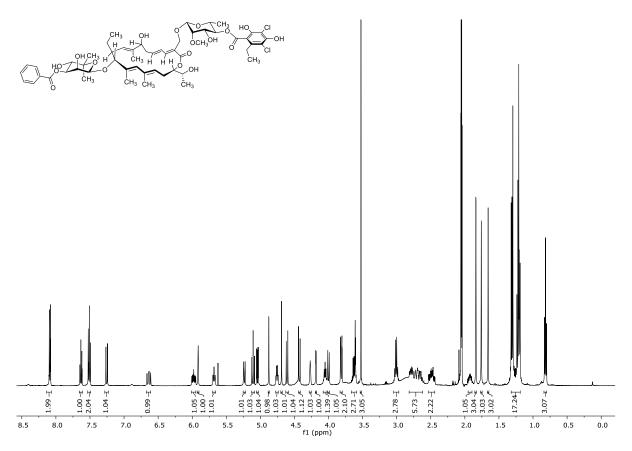
Supplementary Figure 5. 13 C NMR (126 MHz, acetone- d_6) of **OP-1118 (4)**.



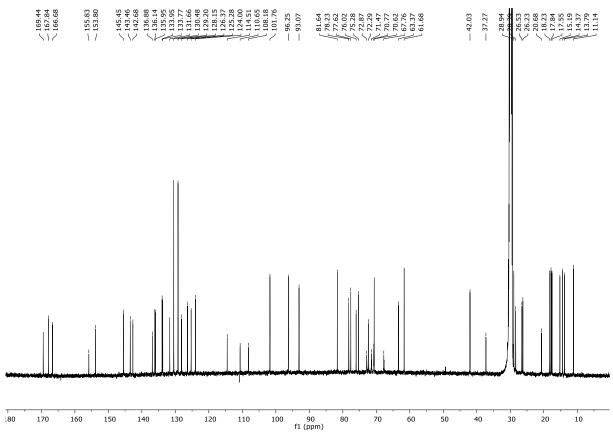
Supplementary Figure 6. ¹H NMR (400 MHz, acetone-*d*₆) of Bisallyl-OP-1118 (5).



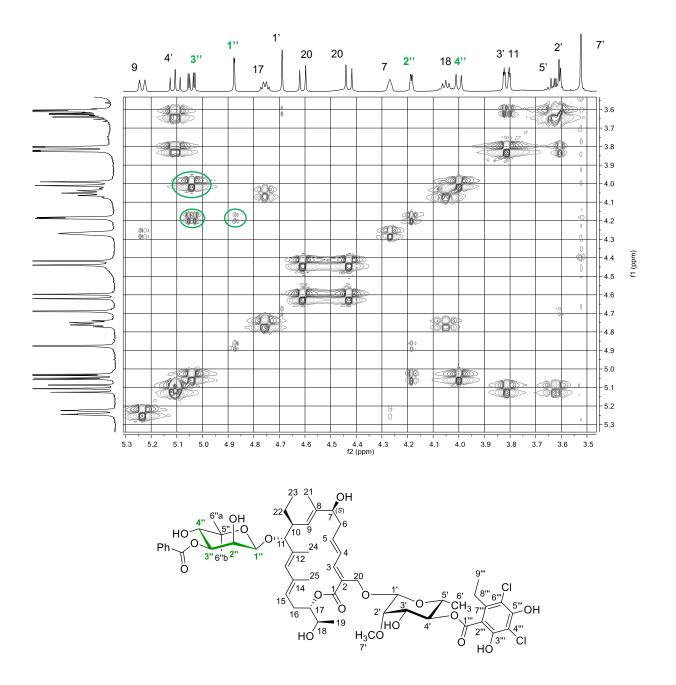
Supplementary Figure 7. ¹³C NMR (126 MHz, acetone-*d*₆) of Bisallyl-OP-1118 (4).



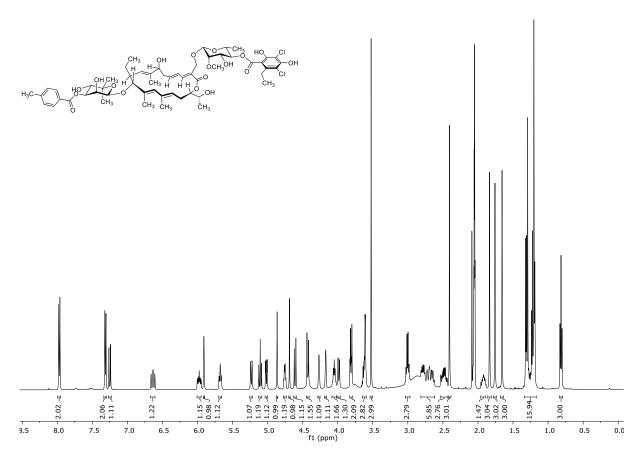
Supplementary Figure 8. ¹H NMR (500 MHz, acetone-d₆) of 7a.



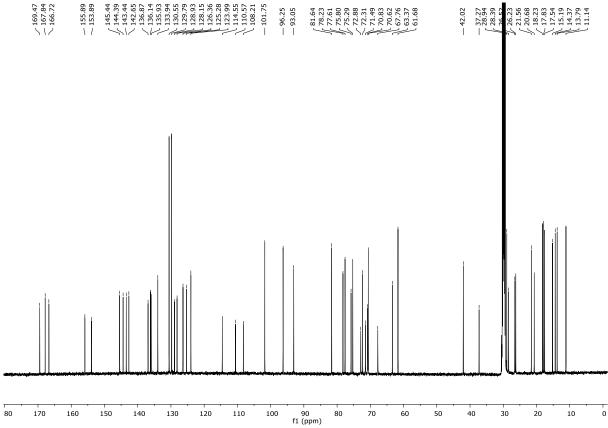
Supplementary Figure 9. 13 C NMR (126 MHz, acetone- d_6) of 7a.



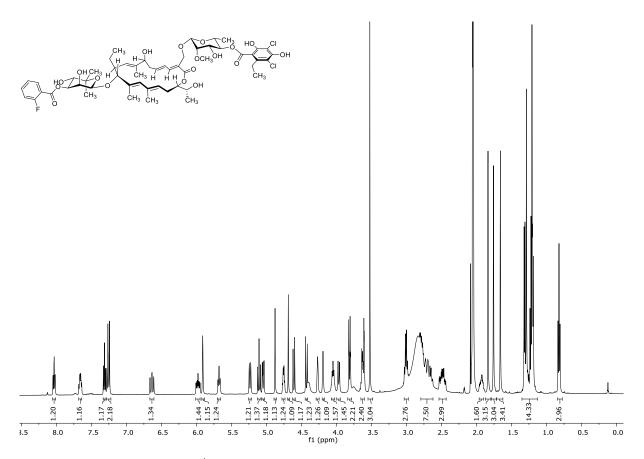
Supplementary Figure 10. COSY NMR (500 MHz, acetone- d_6) of 7a.



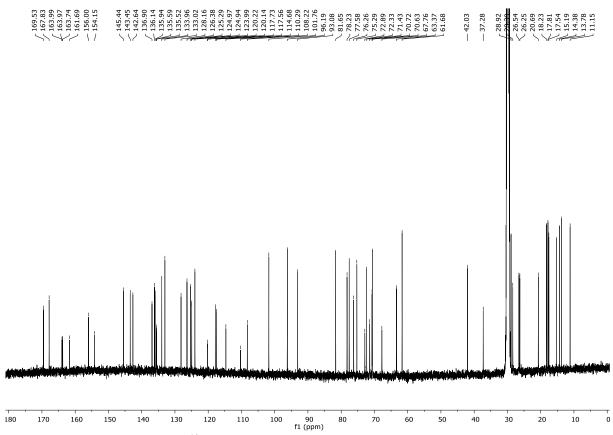
Supplementary Figure 11. ¹H NMR (500 MHz, acetone- d_6) of 7b.



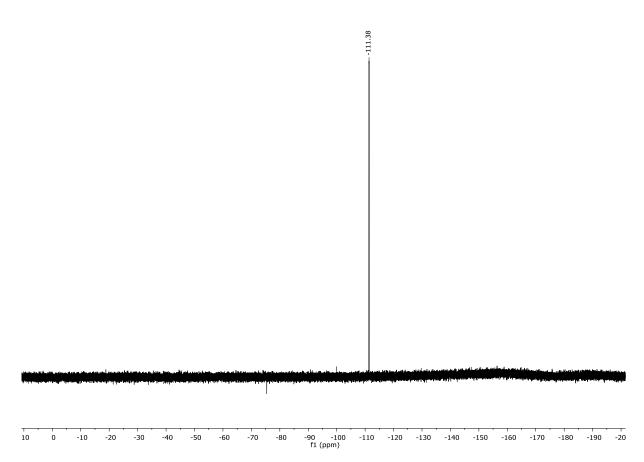
Supplementary Figure 12. 13 C NMR (126 MHz, acetone- d_6) of 7b.



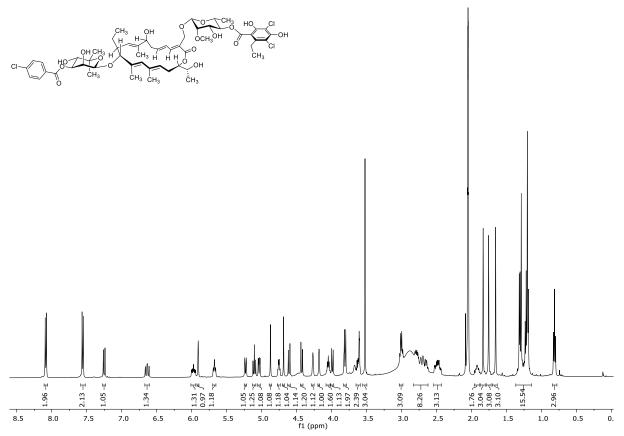
Supplementary Figure 13. ¹H NMR (500 MHz, acetone- d_6) of 7c.



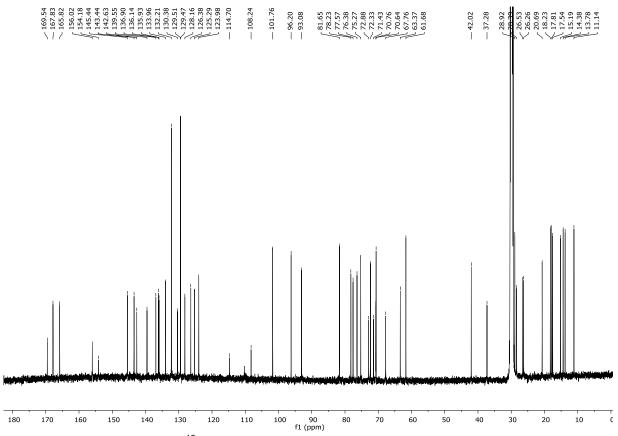
Supplementary Figure 14. 13 C NMR (126 MHz, acetone- d_6) of **7c**.



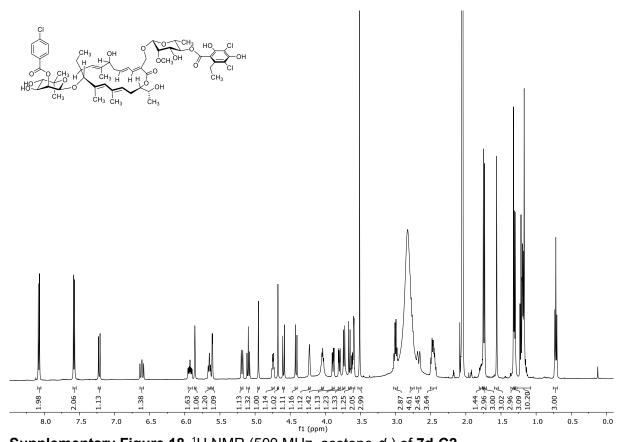
Supplementary Figure 15. 19 F NMR (376.5 MHz, acetone- d_6) of 7c.



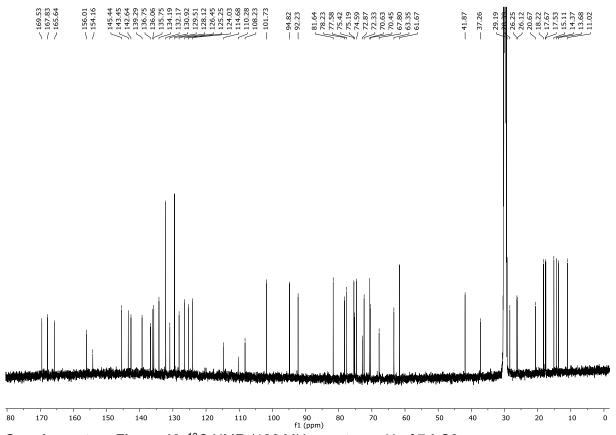
Supplementary Figure 16. 1 H NMR (500 MHz, acetone- d_{6}) of 7d.



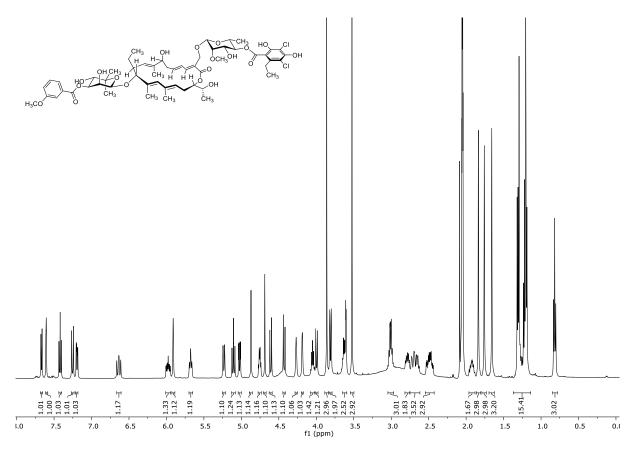
Supplementary Figure 17. 13 C NMR (126 MHz, acetone- d_6) of 7d.



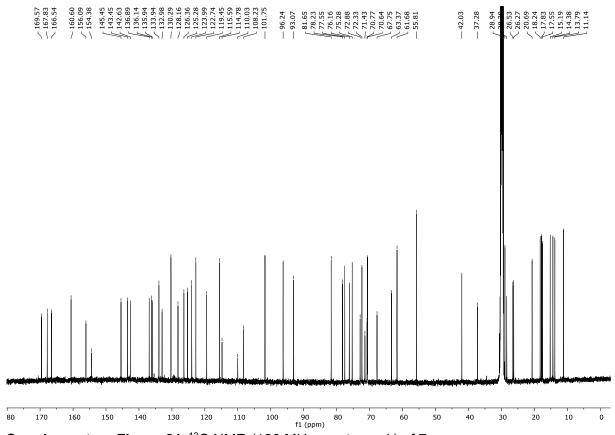
Supplementary Figure 18. ¹H NMR (500 MHz, acetone-d₆) of 7d-C2.



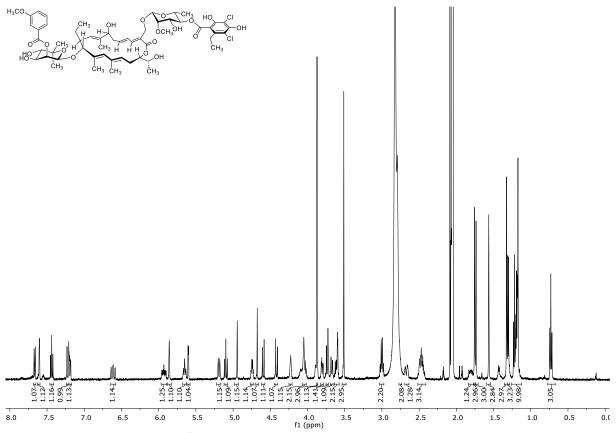
Supplementary Figure 19. ¹³C NMR (126 MHz, acetone-*d*₆) of **7d-C2**.



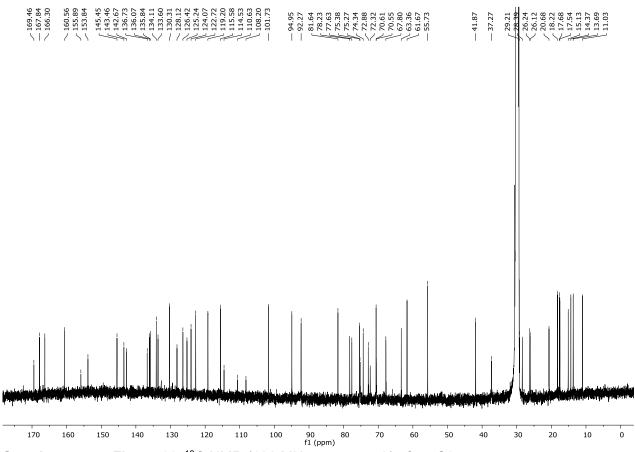
Supplementary Figure 20. ¹H NMR (500 MHz, acetone- d_6) of 7e.



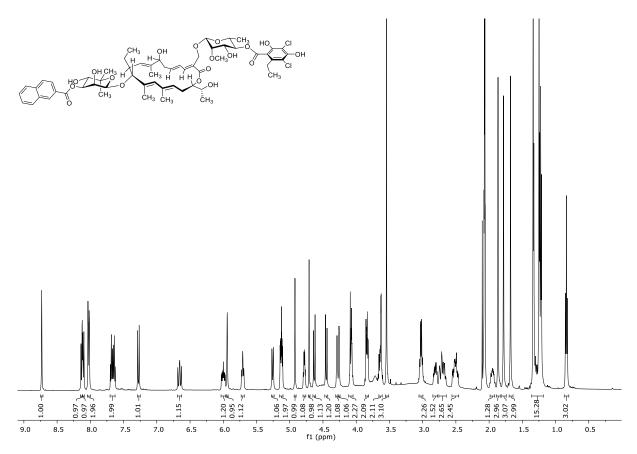
Supplementary Figure 21. 13 C NMR (126 MHz, acetone- d_6) of 7e.



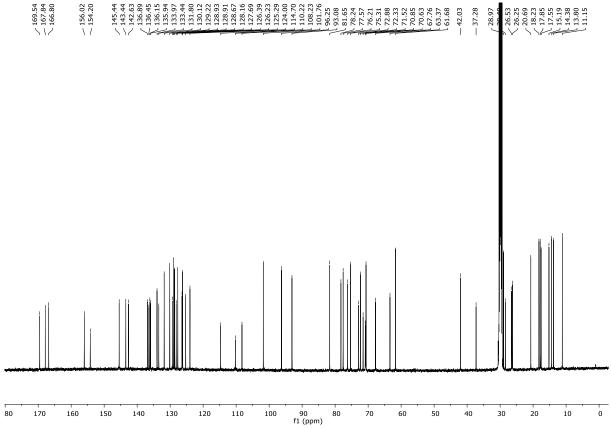
Supplementary Figure 22. ¹H NMR (500 MHz, acetone-*d*₆) of **7e-C2**.



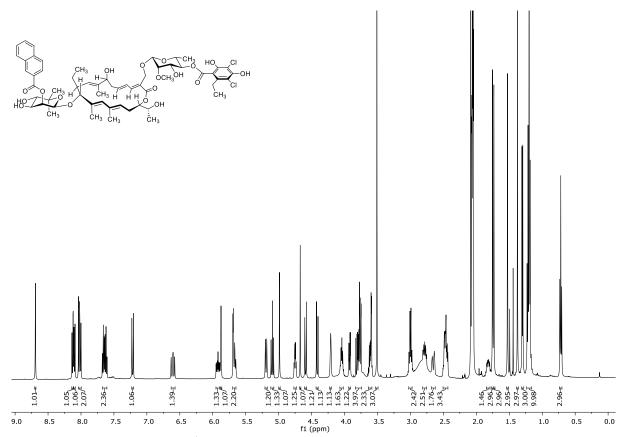
Supplementary Figure 23. 13 C NMR (126 MHz, acetone- d_6) of 7e-C2.



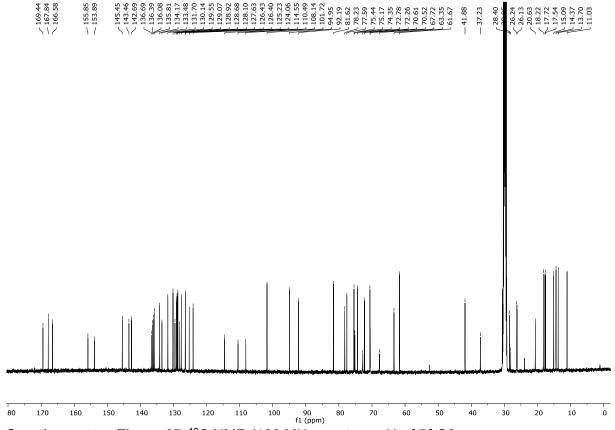
Supplementary Figure 24. ¹H NMR (500 MHz, acetone-d₆) of 7f.



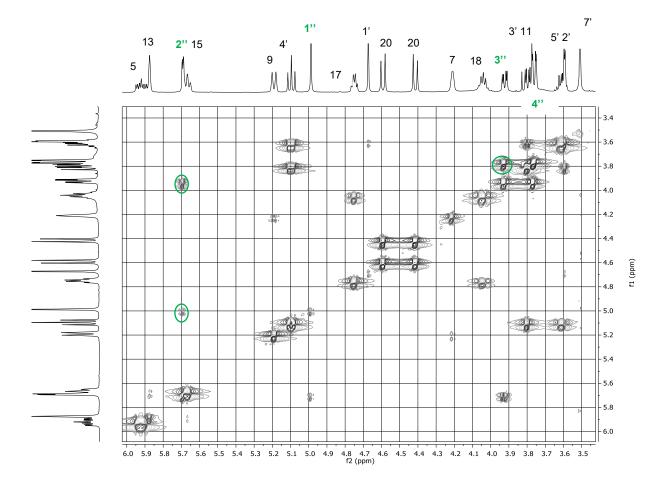
Supplementary Figure 25. ¹³C NMR (126 MHz, acetone-d₆) of 7f.



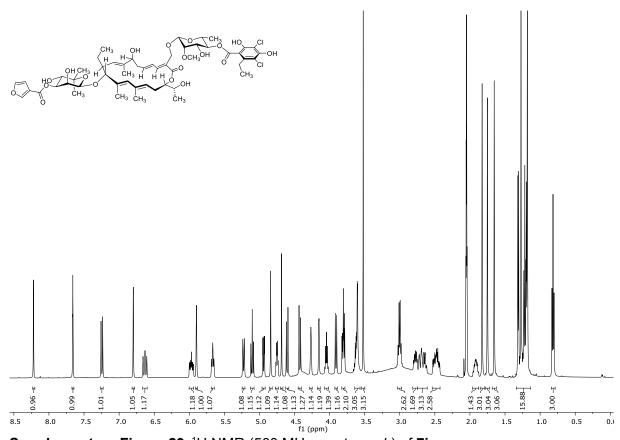




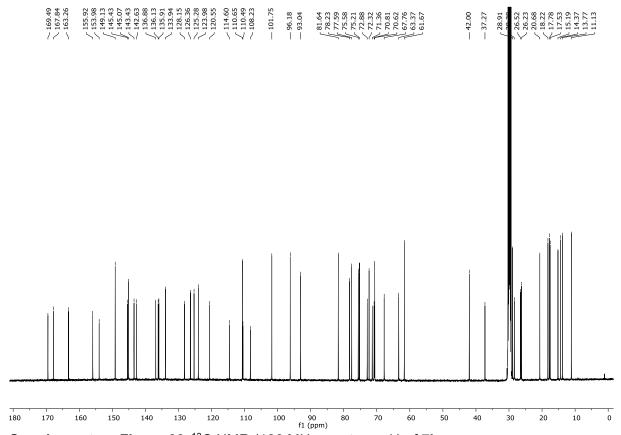
Supplementary Figure 27. ¹³C NMR (126 MHz, acetone-*d*₆) of 7f-C2.



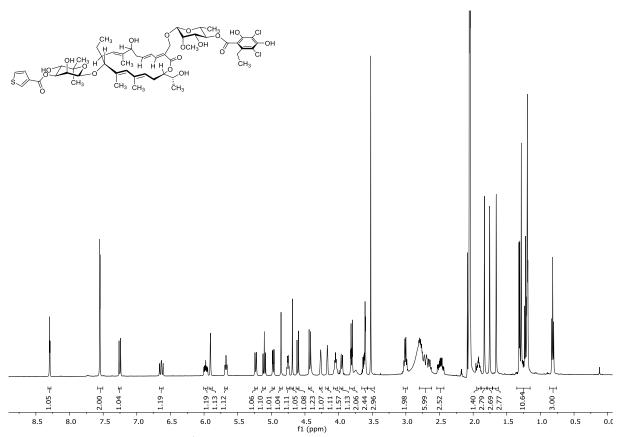
Supplementary Figure 28. COSY NMR (126 MHz, acetone- d_6) of 7f-C2.



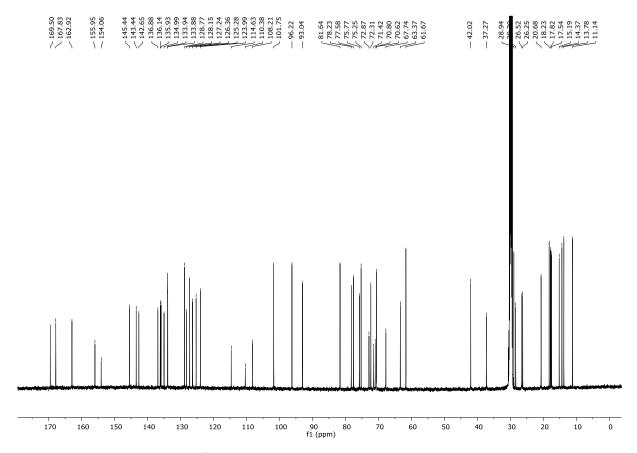
Supplementary Figure 29. ¹H NMR (500 MHz, acetone-d₆) of 7j.



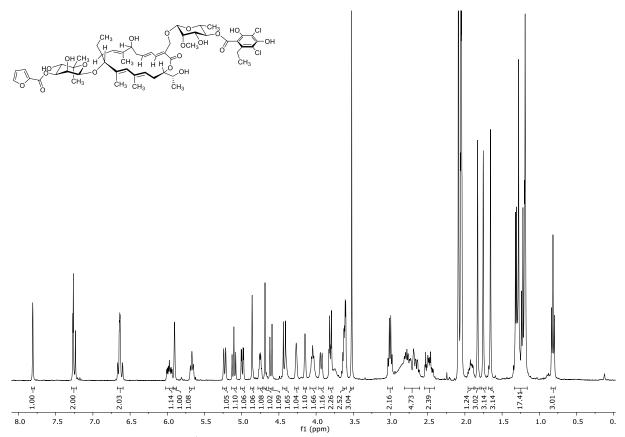
Supplementary Figure 30. 13 C NMR (126 MHz, acetone- d_6) of 7j.



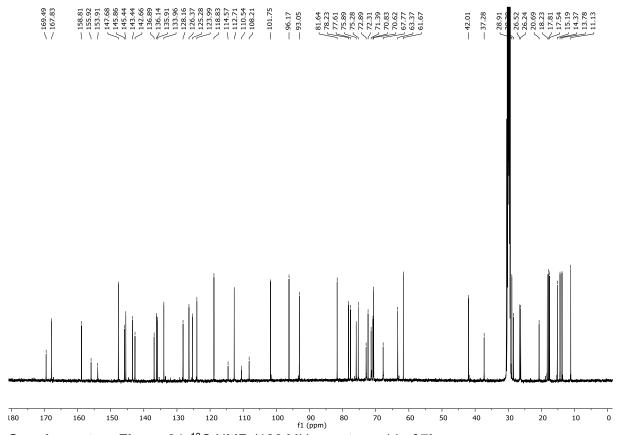
Supplementary Figure 31. ¹H NMR (500 MHz, acetone-d₆) of 7k.



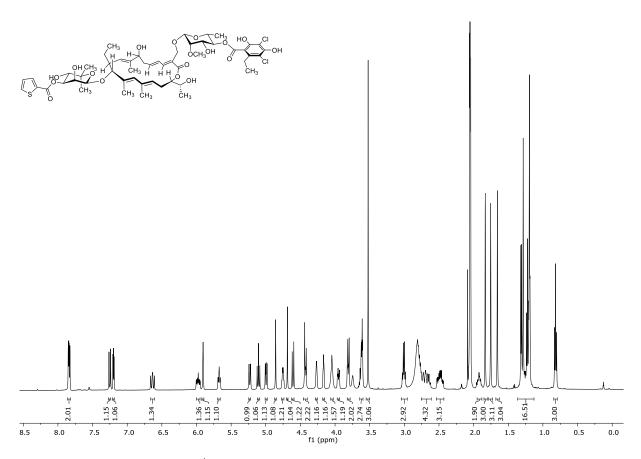
Supplementary Figure 32. ^{13}C NMR (126 MHz, acetone-d₆) of 7k.



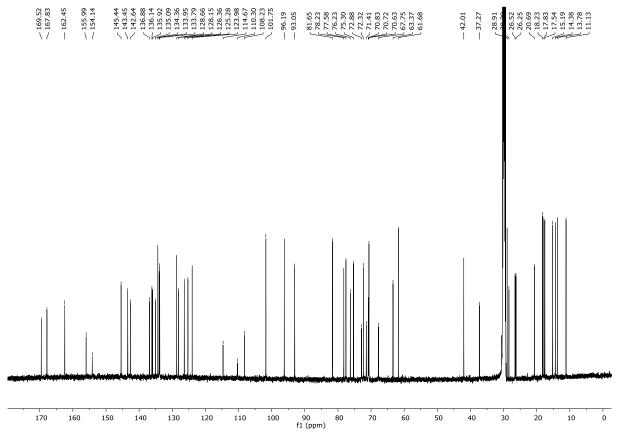
Supplementary Figure 33. ¹H NMR (400 MHz, acetone-d₆) of 71.



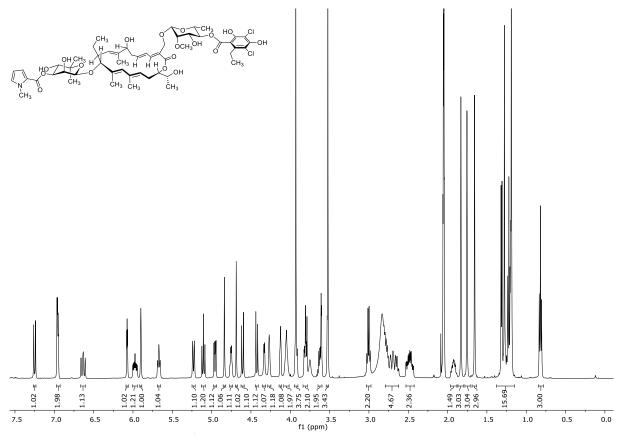
Supplementary Figure 34. ¹³C NMR (126 MHz, acetone-d₆) of 7I.



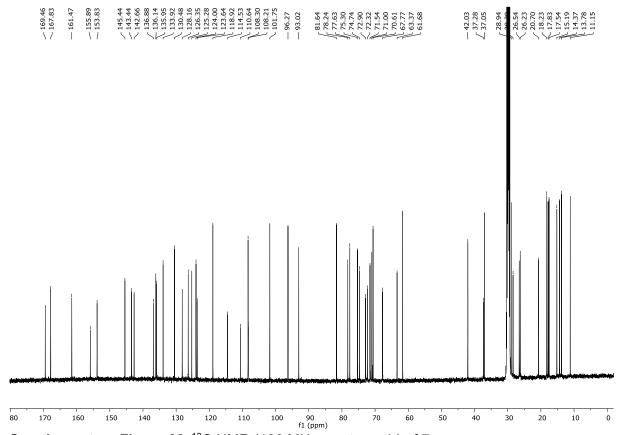
Supplementary Figure 35. ¹H NMR (500 MHz, acetone-*d*₆) of **7m**.



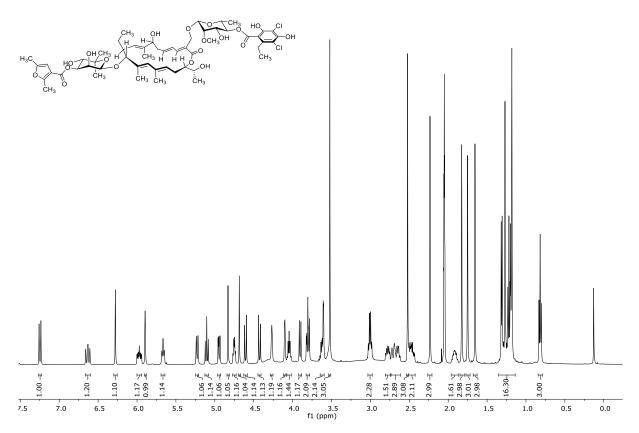
Supplementary Figure 36. ¹³C NMR (126 MHz, acetone-d₆) of 7m.



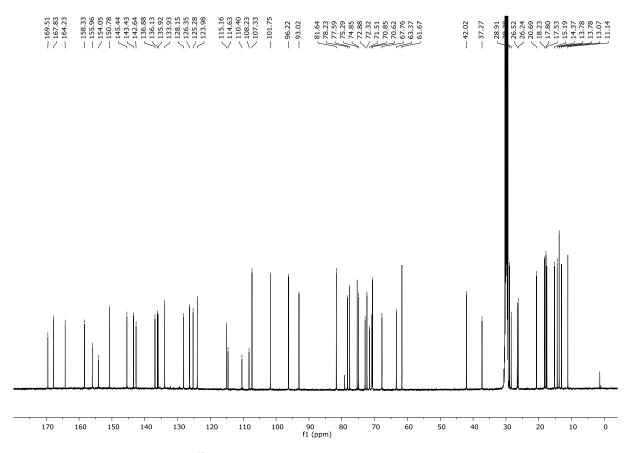
Supplementary Figure 37. ¹H NMR (500 MHz, acetone- d_6) of 7n.



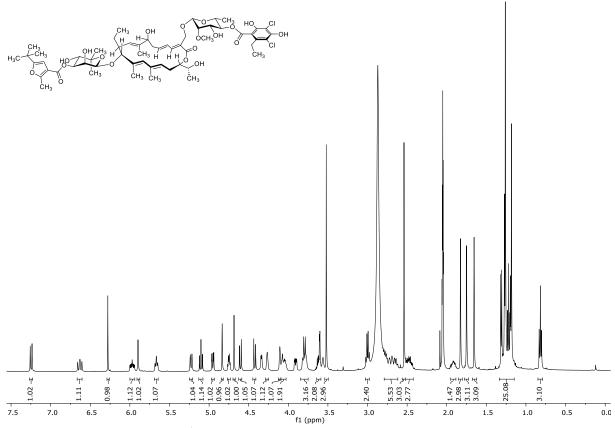
Supplementary Figure 38. ¹³C NMR (126 MHz, acetone-d₆) of 7n.



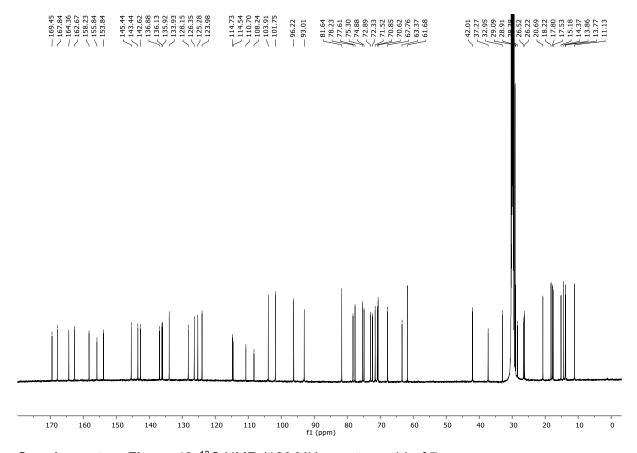
Supplementary Figure 39. 1 H NMR (500 MHz, acetone- d_{6}) of 7p.



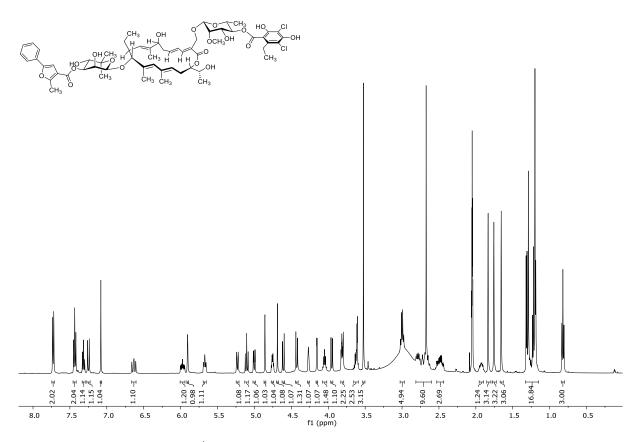
Supplementary Figure 40. ^{13}C NMR (126 MHz, acetone-d₆) of **7p**.



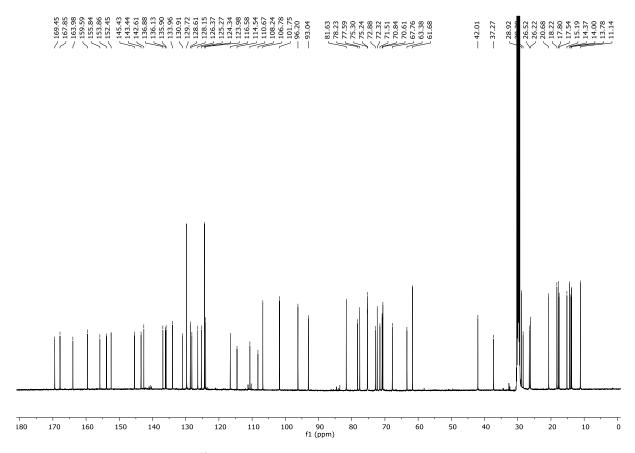
Supplementary Figure 41. ¹H NMR (500 MHz, acetone-d₆) of 7q.



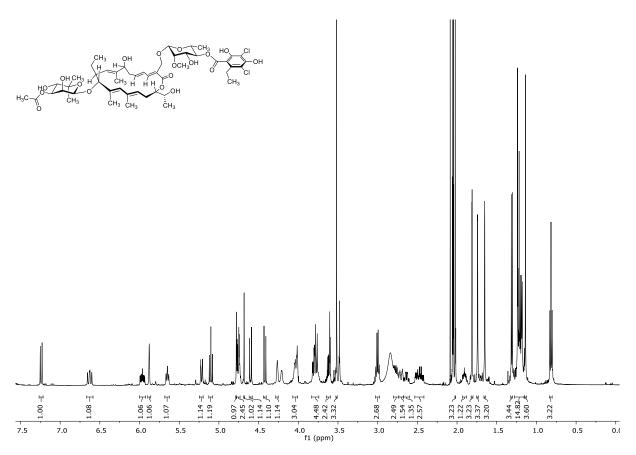
Supplementary Figure 42. ¹³C NMR (126 MHz, acetone-d₆) of 7q.



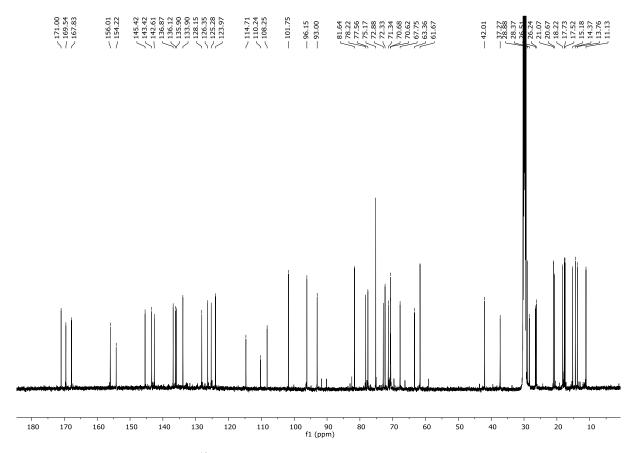
Supplementary Figure 43. ¹H NMR (500 MHz, acetone-d₆) of 7r.



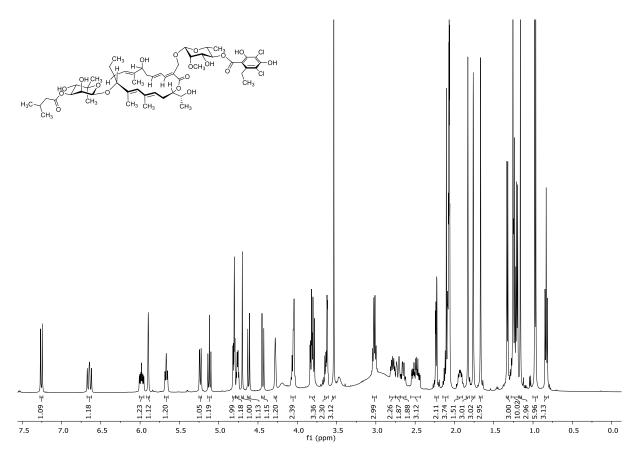
Supplementary Figure 44. ^{13}C NMR (126 MHz, acetone-d₆) of 7r.



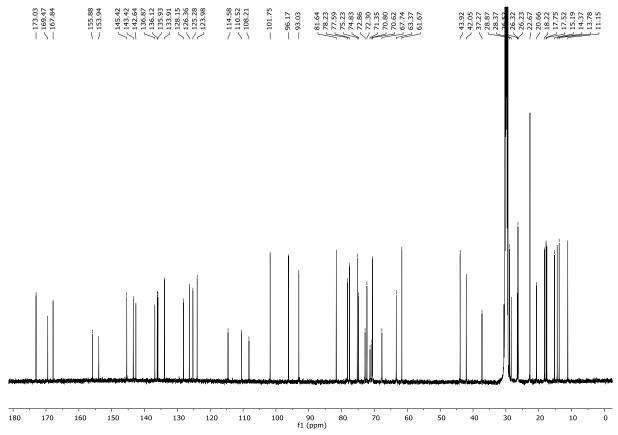
Supplementary Figure 45. ¹H NMR (500 MHz, acetone- d_6) of 7s.



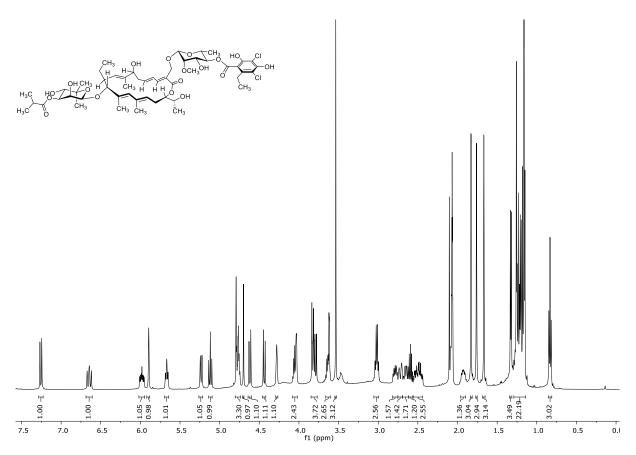
Supplementary Figure 46. ^{13}C NMR (126 MHz, acetone-d₆) of 7s.



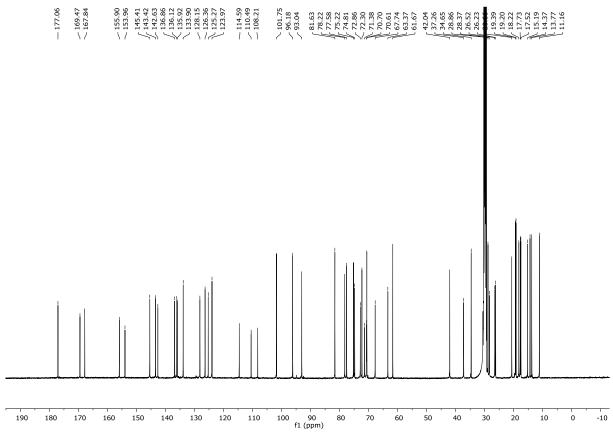
Supplementary Figure 47. ¹H NMR (500 MHz, acetone-*d*₆) of 7t.



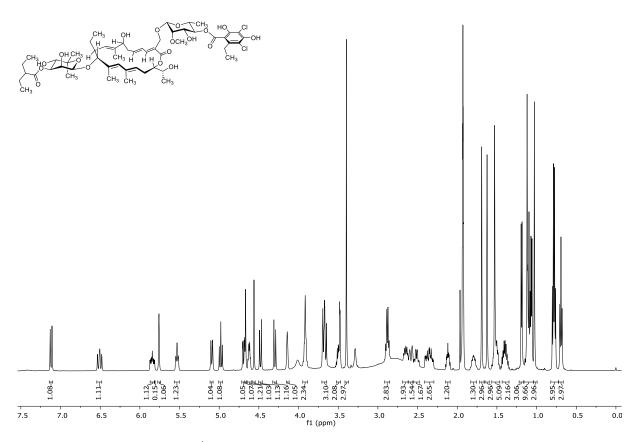
Supplementary Figure 48. ¹³C NMR (126 MHz, acetone-d₆) of 7t.



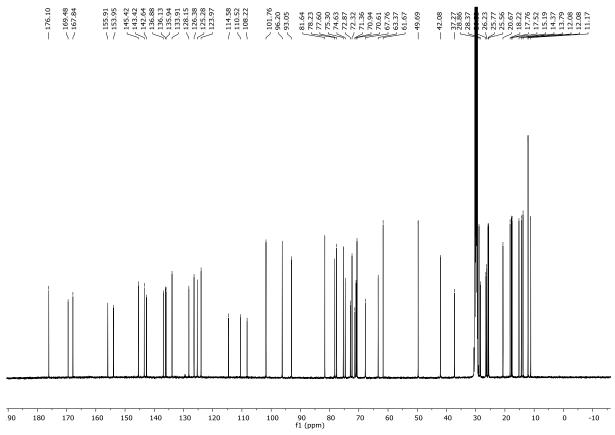
Supplementary Figure 49. 1 H NMR (500 MHz, acetone- d_{6}) of 7u.



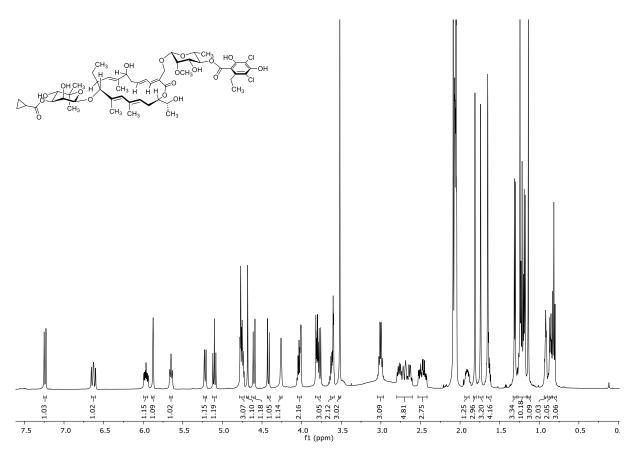
Supplementary Figure 50. ¹³C NMR (126 MHz, acetone-d₆) of 7u.



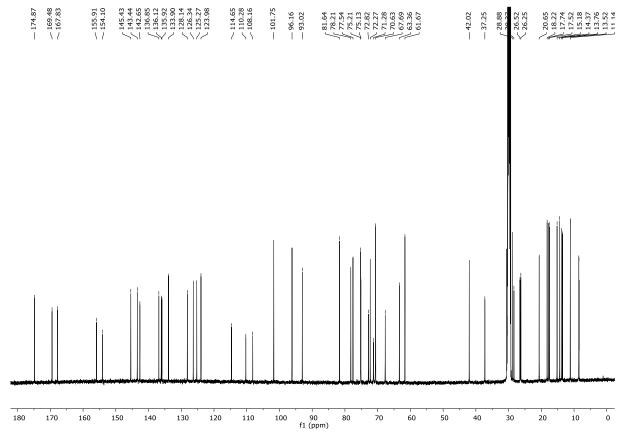
Supplementary Figure 51. ¹H NMR (500 MHz, acetone-*d*₆) of **7w**.



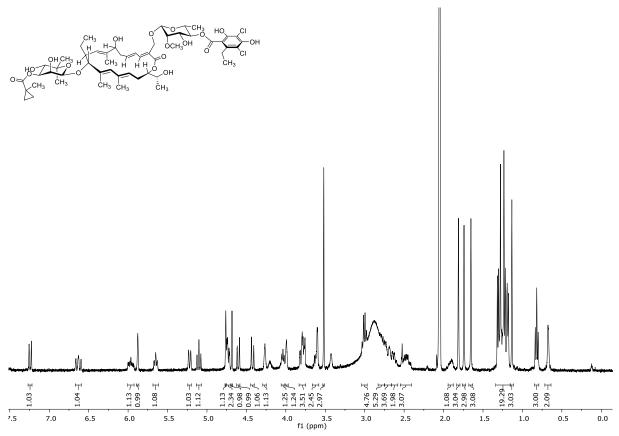
Supplementary Figure 52. ¹³C NMR (126 MHz, acetone-d₆) of 7w.



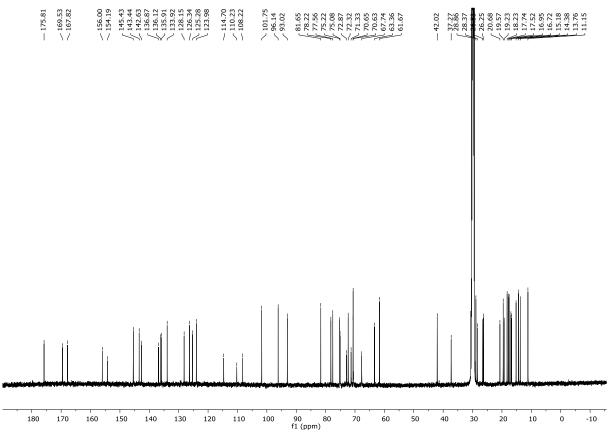
Supplementary Figure 53. ¹H NMR (500 MHz, acetone- d_6) of 7x.



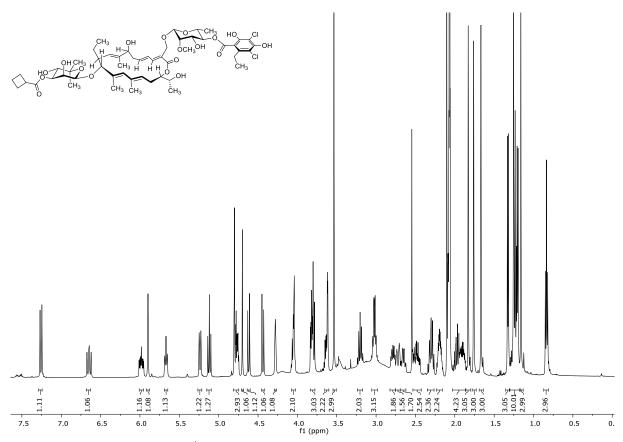
Supplementary Figure 54. ¹³C NMR (126 MHz, acetone-d₆) of 7x.



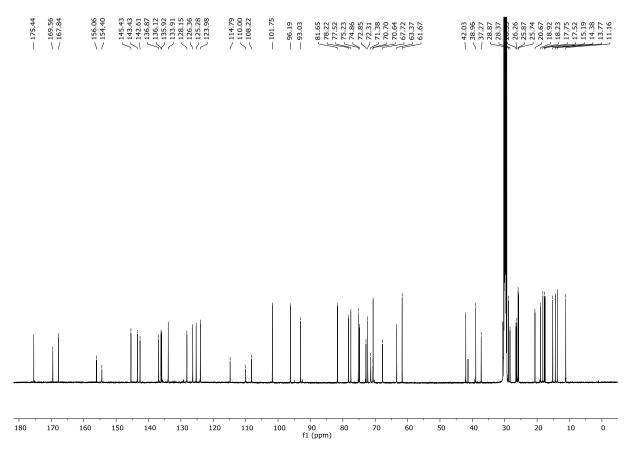
Supplementary Figure 55. ¹H NMR (500 MHz, acetone-*d*₆) of 7y.



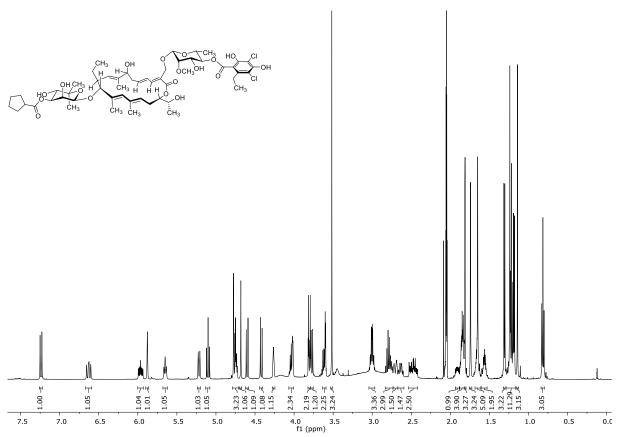
Supplementary Figure 56. ^{13}C NMR (126 MHz, acetone-d₆) of 7y.



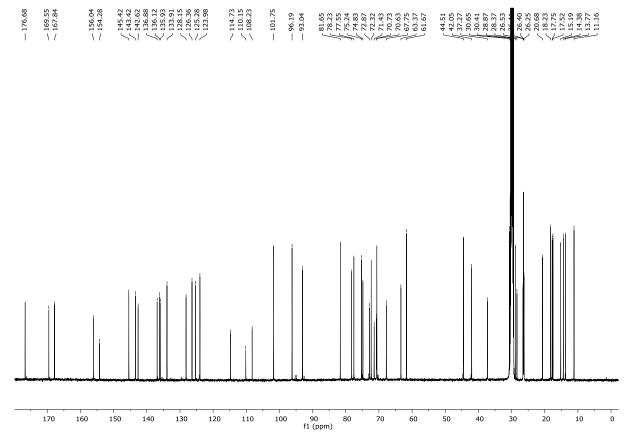
Supplementary Figure 57. 1 H NMR (500 MHz, acetone- d_{6}) of 7z.



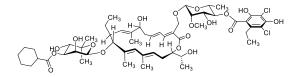
Supplementary Figure 58. 13 C NMR (126 MHz, acetone- d_6) of 7z.

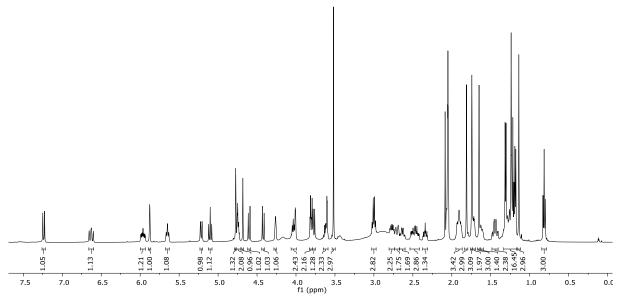


Supplementary Figure 59. ¹H NMR (500 MHz, acetone-d₆) of 7aa.

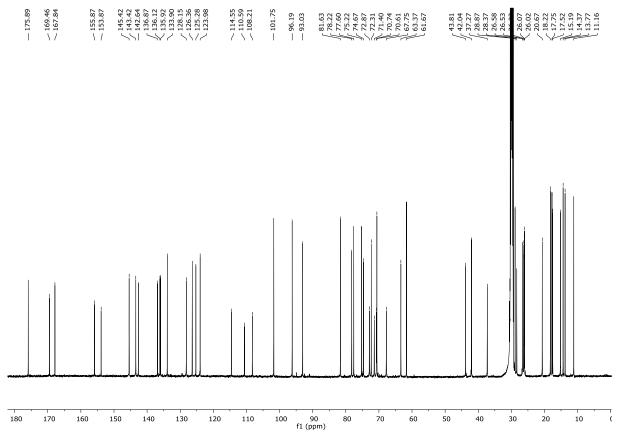


Supplementary Figure 60. ¹³C NMR (126 MHz, acetone-d₆) of 7aa.

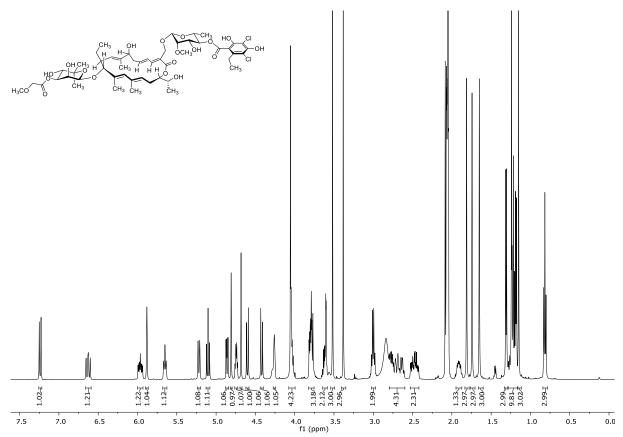




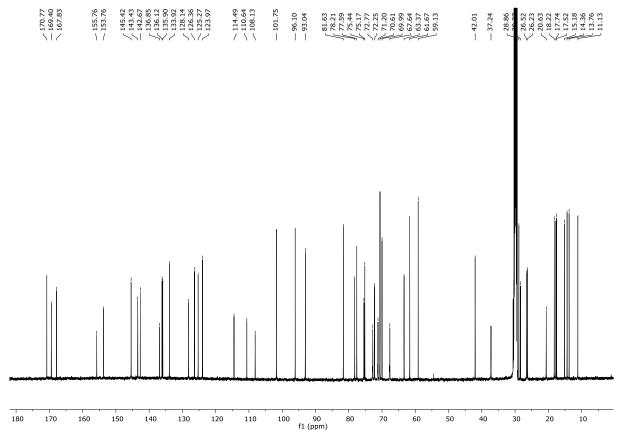
Supplementary Figure 61. ¹H NMR (500 MHz, acetone-*d*₆) of **7ab**.



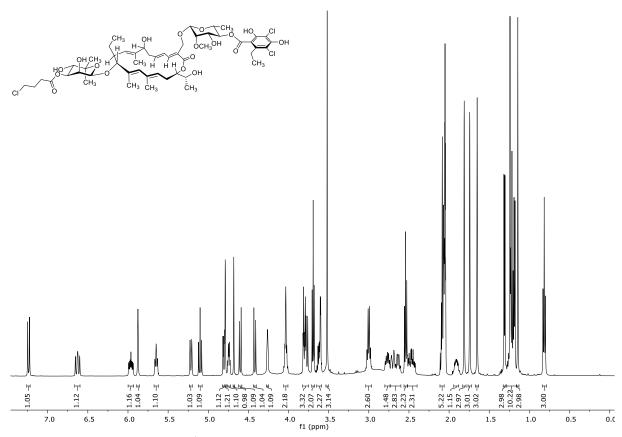
Supplementary Figure 62. ¹³C NMR (126 MHz, acetone-d₆) of **7ab**.



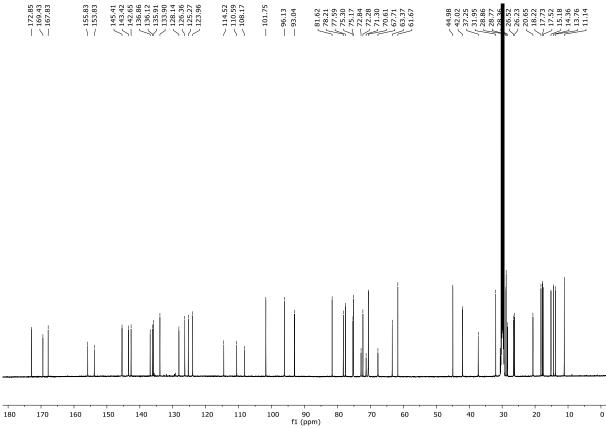
Supplementary Figure 63. ¹H NMR (500 MHz, acetone-d₆) of 7ad.



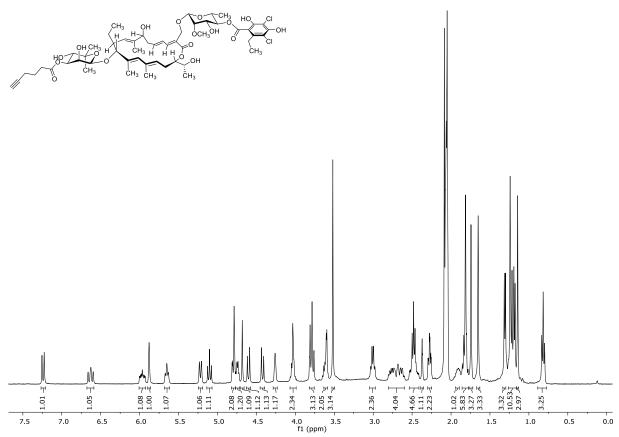
Supplementary Figure 64. ¹³C NMR (126 MHz, acetone-d₆) of 7ad.



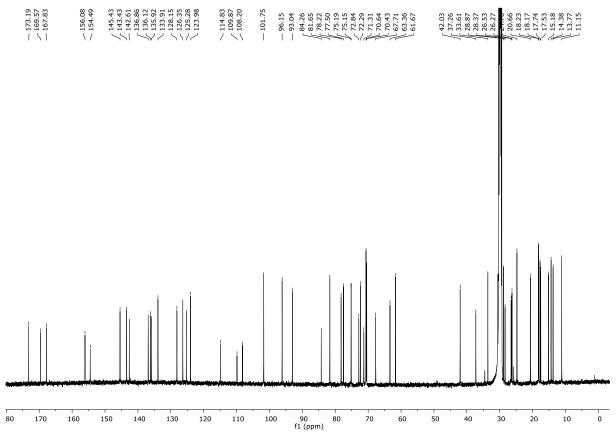
Supplementary Figure 65. ¹H NMR (500 MHz, acetone-d₆) of 7ag.



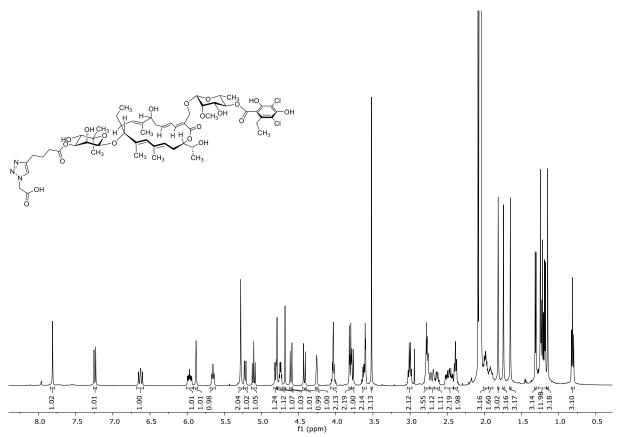
Supplementary Figure 66. ¹³C NMR (126 MHz, acetone-d₆) of **7ag**.



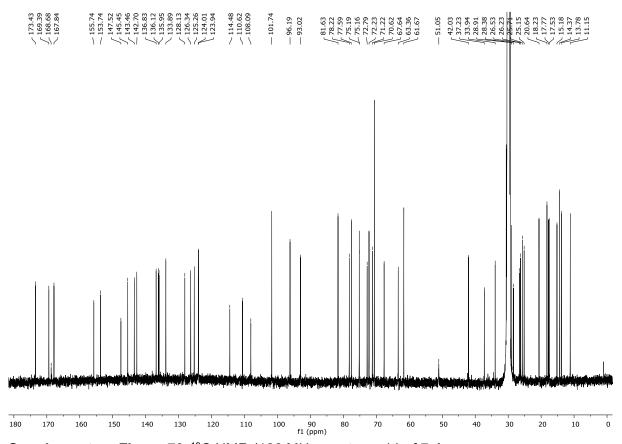
Supplementary Figure 67. ¹H NMR (400 MHz, acetone-*d*₆) of 7ah.



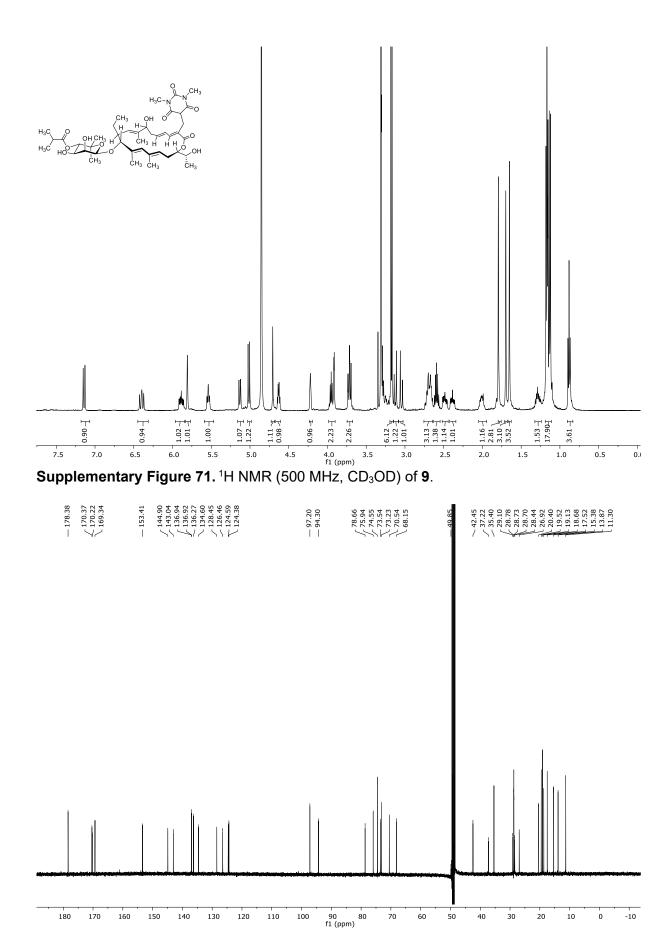
Supplementary Figure 68. ¹³C NMR (126 MHz, acetone-d₆) of **7ah**.



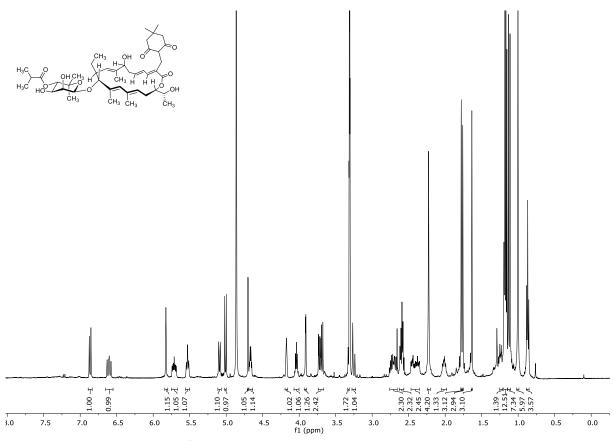
Supplementary Figure 69. ¹H NMR (500 MHz, acetone-*d*₆) of **7ai**.



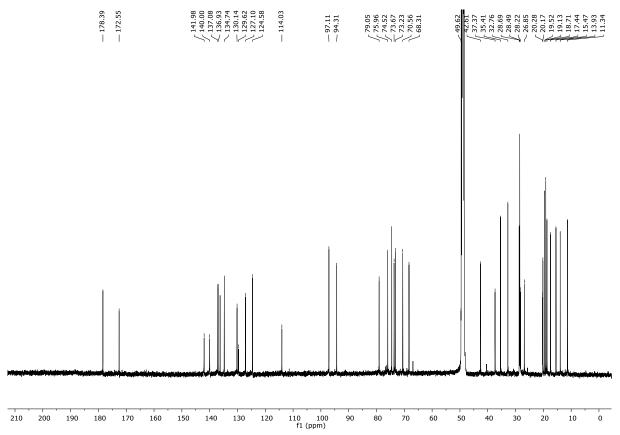
Supplementary Figure 70. ^{13}C NMR (126 MHz, acetone-d₆) of 7ai.



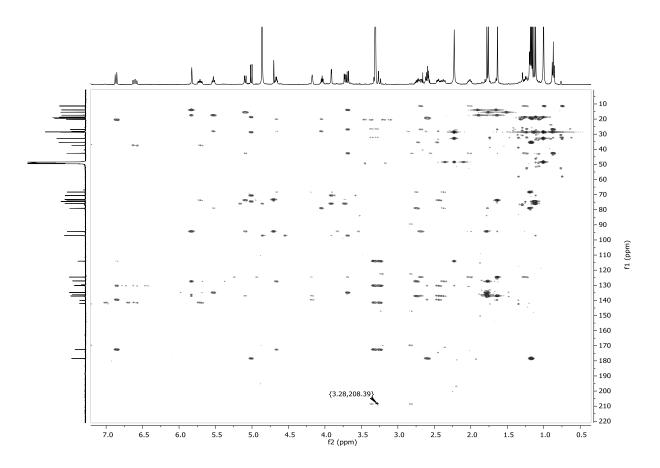
Supplementary Figure 72. ^{13}C NMR (126 MHz, CD₃OD) of 9.



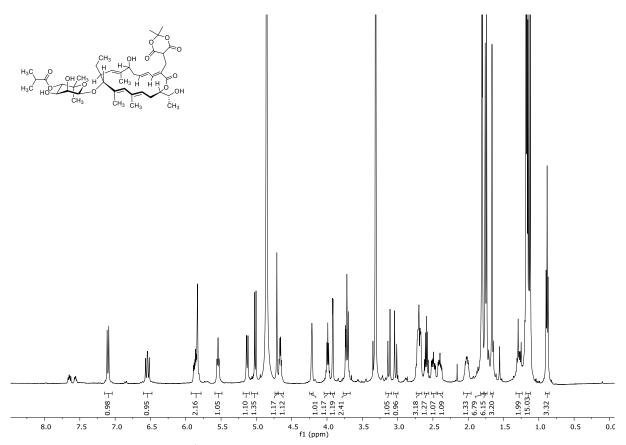
Supplementary Figure 73. ¹H NMR (500 MHz, CD₃OD) of 10.



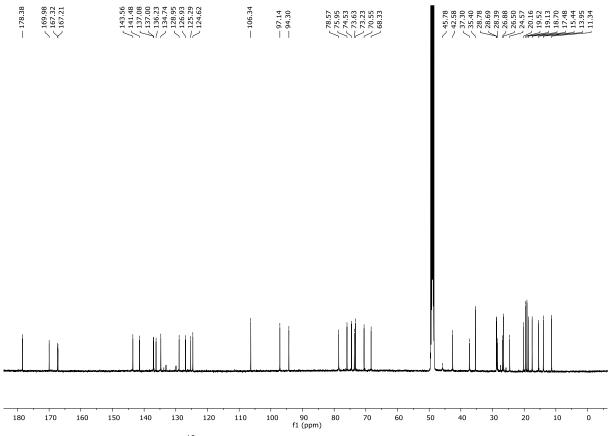
Supplementary Figure 74. ^{13}C NMR (126 MHz, CD₃OD) of 10.



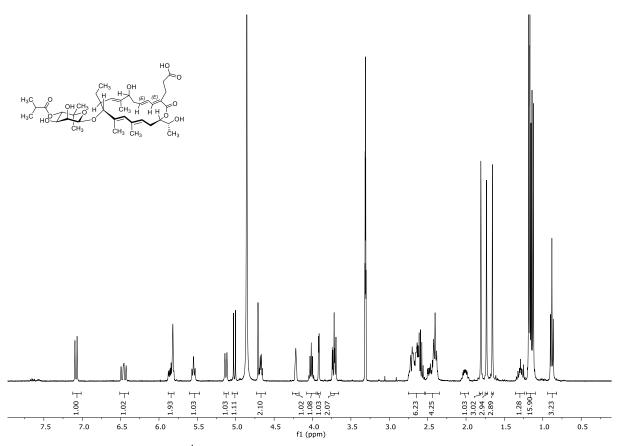
Supplementary Figure 75. HMBC NMR (500 MHz, 126 MHz, CD_3OD) of 10.



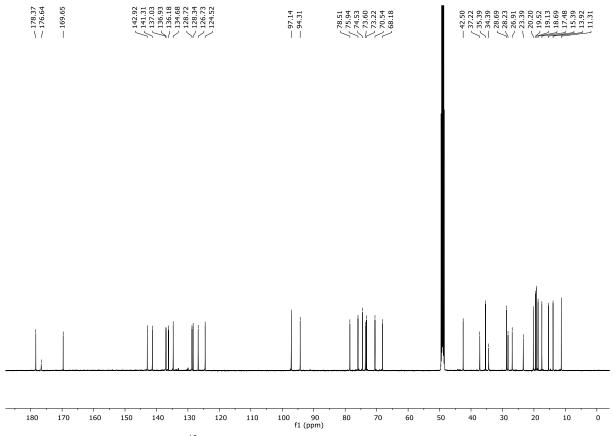
Supplementary Figure 76. ¹H NMR (500 MHz, CD₃OD) of 11.



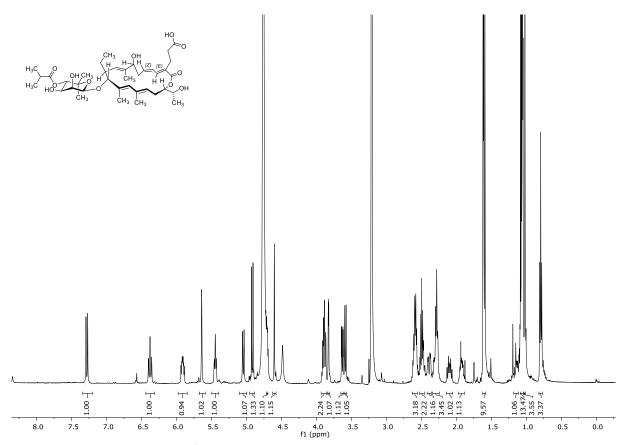
Supplementary Figure 77. ^{13}C NMR (126 MHz, CD $_{3}OD)$ of 11.



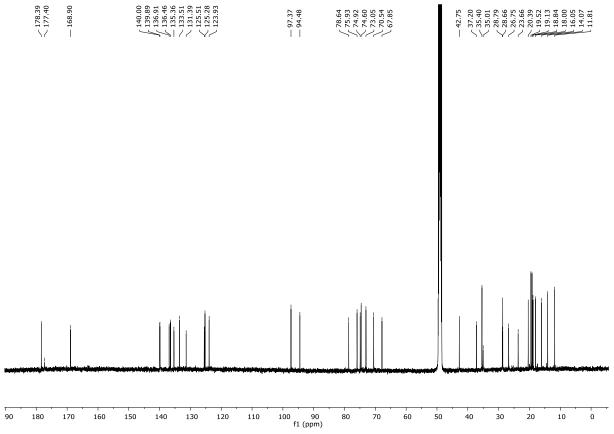
Supplementary Figure 78. ¹H NMR (400 MHz, CD₃OD) of (*E*)-13.



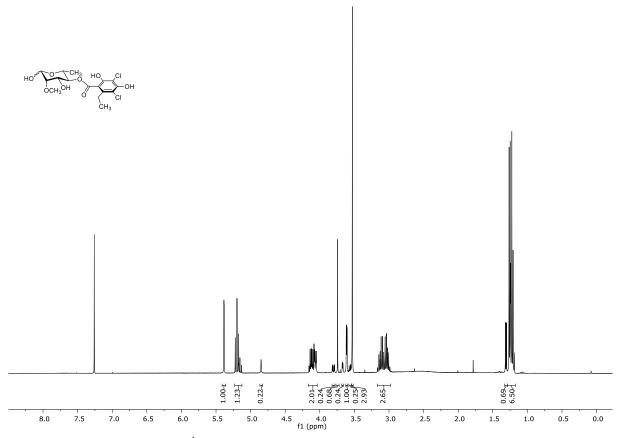
Supplementary Figure 79. ¹³C NMR (126 MHz, CD₃OD) of (*E*)-13.



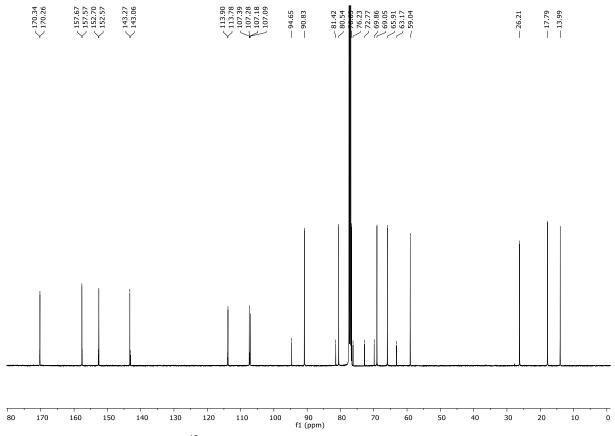
Supplementary Figure 80. ¹H NMR (500 MHz, CD₃OD) of (*Z*)-13.



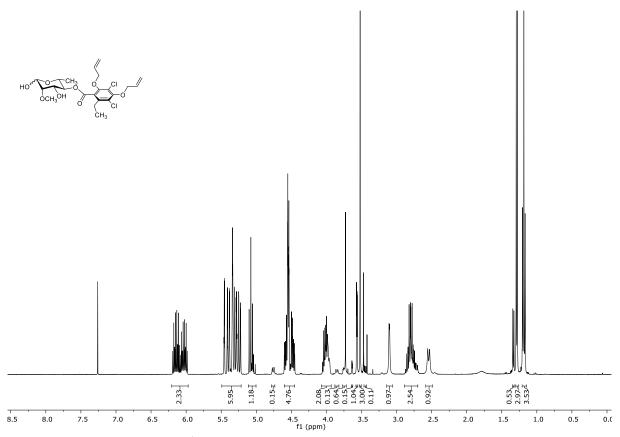
Supplementary Figure 81. ¹³C NMR (126 MHz, CD₃OD) of (*Z*)-13.



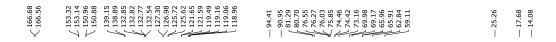
Supplementary Figure 82. ¹H NMR (400 MHz, CDCl₃) of 12.

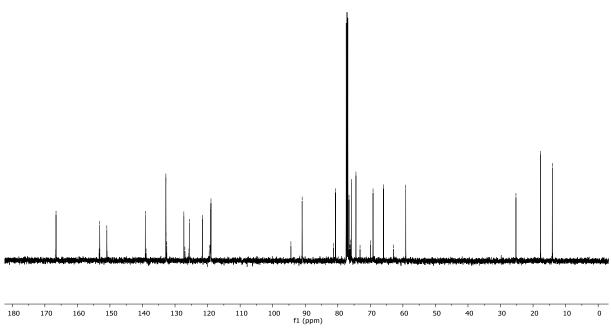


Supplementary Figure 83. ^{13}C NMR (126 MHz, CDCl₃) of 12.

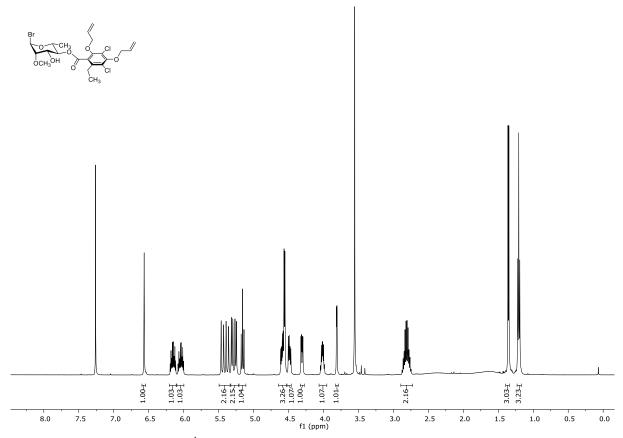


Supplementary Figure 84. ^1H NMR (400 MHz, CDCl₃) of S1.

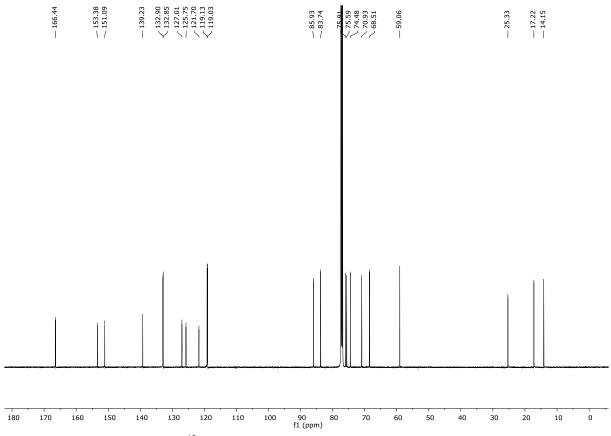




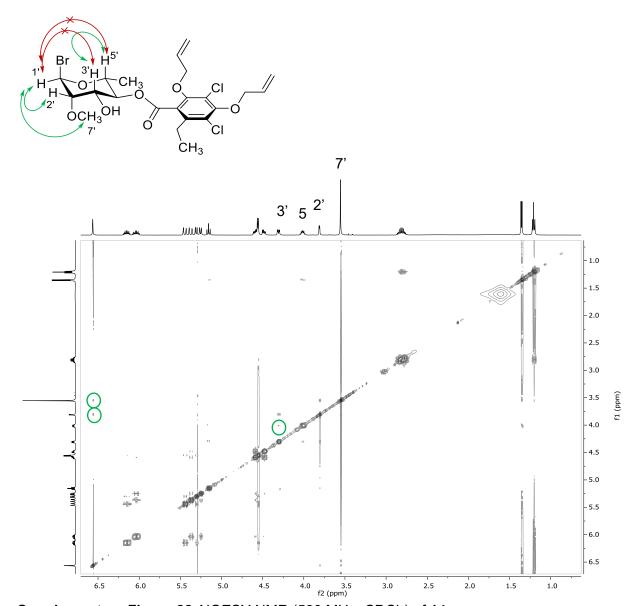
Supplementary Figure 85. 13 C NMR (126 MHz, CDCl₃) of S1.



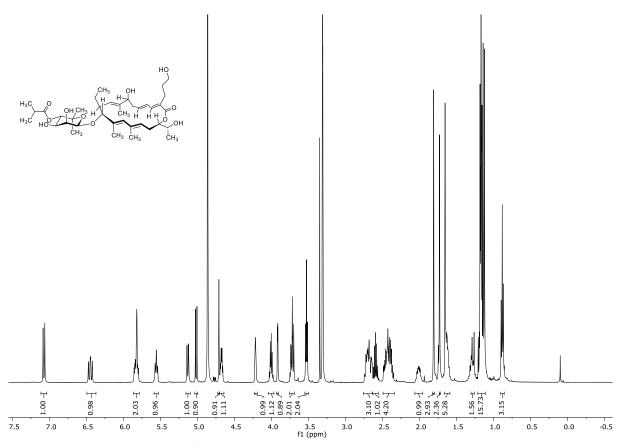
Supplementary Figure 86. ¹H NMR (500 MHz, CDCl₃) of 14.



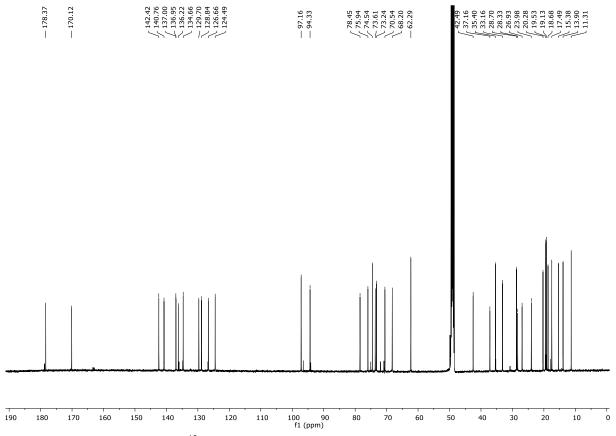
Supplementary Figure 87. 13 C NMR (126 MHz, CDCl₃) of 14.



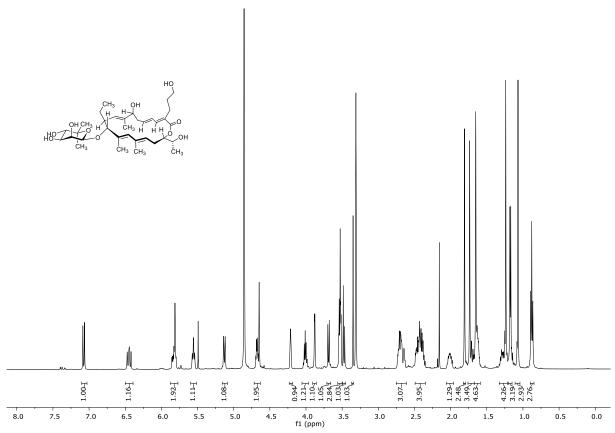
Supplementary Figure 88. NOESY NMR (500 MHz, CDCl $_3$) of 14.



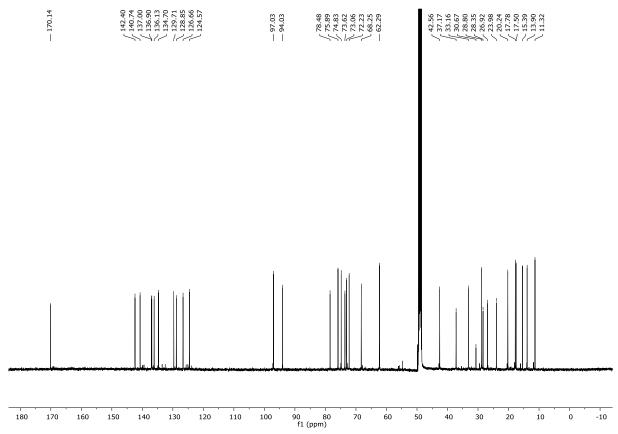
Supplementary Figure 89. ¹H NMR (500 MHz, CD₃OD) of 15.



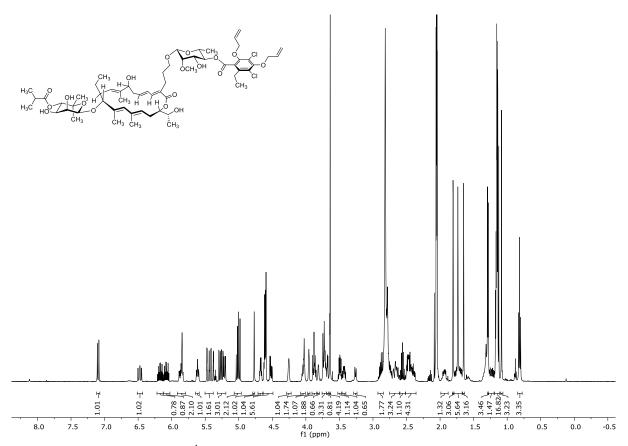
Supplementary Figure 90. ¹³C NMR (126 MHz, CD₃OD) of 15.



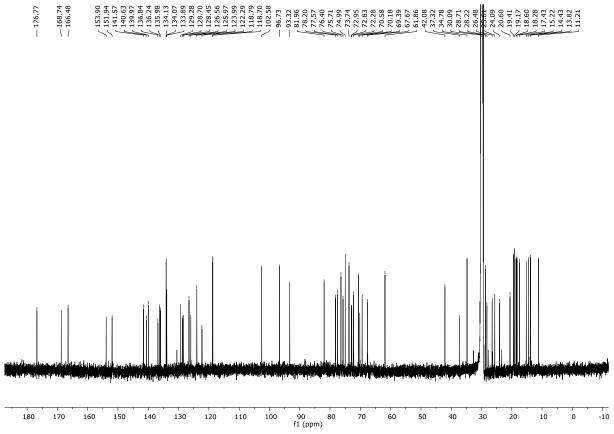
Supplementary Figure 91. 1H NMR (500 MHz, CD $_3OD$) of S2.



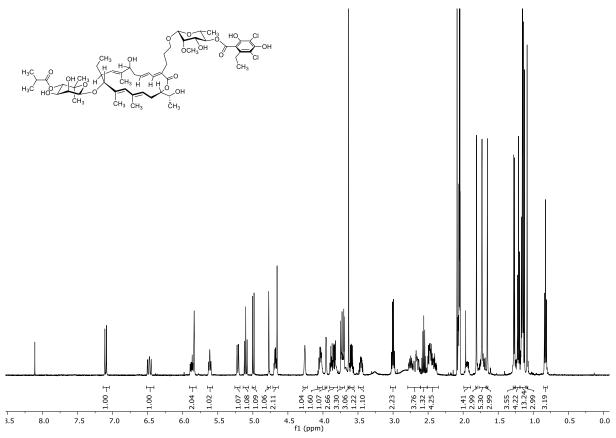
Supplementary Figure 92. ¹³C NMR (126 MHz, CD₃OD) of S2.



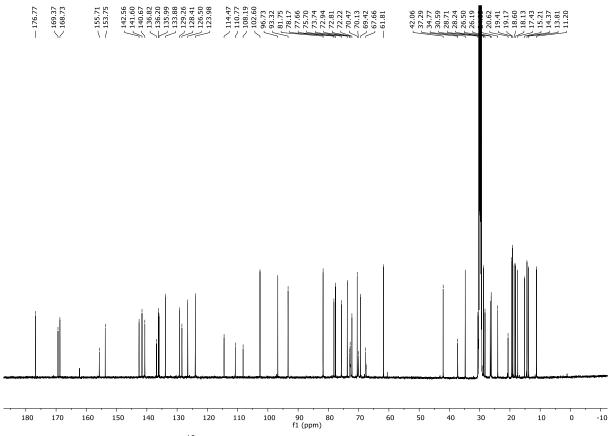
Supplementary Figure 93. 1 H NMR (500 MHz, acetone- d_{6}) of S3.



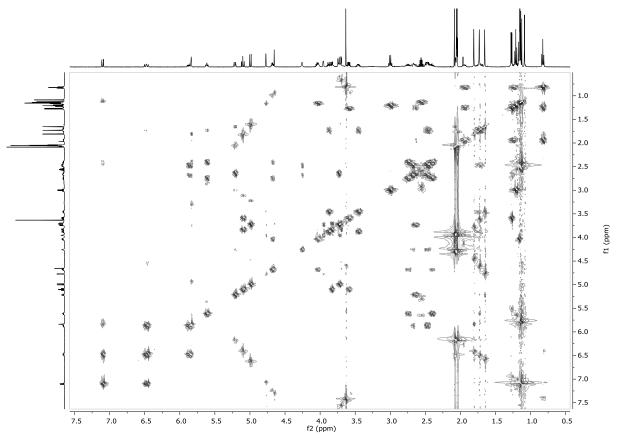
Supplementary Figure 94. 13 C NMR (126 MHz, acetone- d_6) of S3.



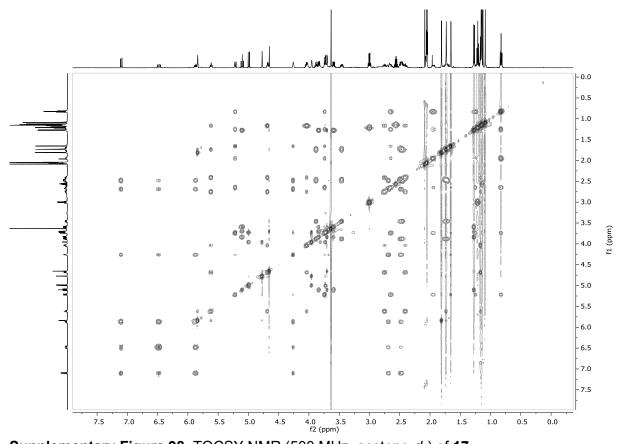
Supplementary Figure 95. ¹H NMR (500 MHz, acetone-*d*₆) of 17.



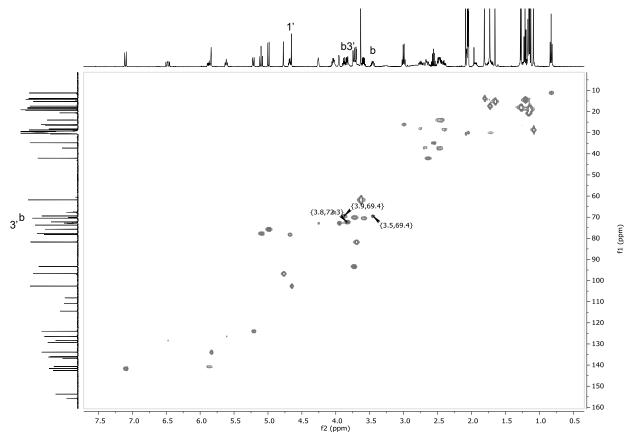
Supplementary Figure 96. 13 C NMR (126 MHz, acetone- d_6) of **17**.



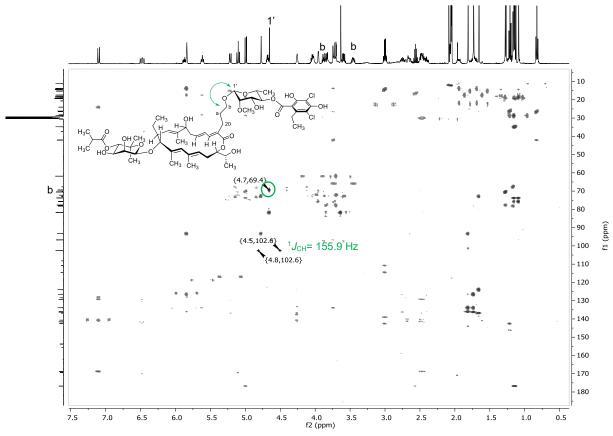
Supplementary Figure 97. COSY NMR (500 MHz, acetone- d_6) of 17.



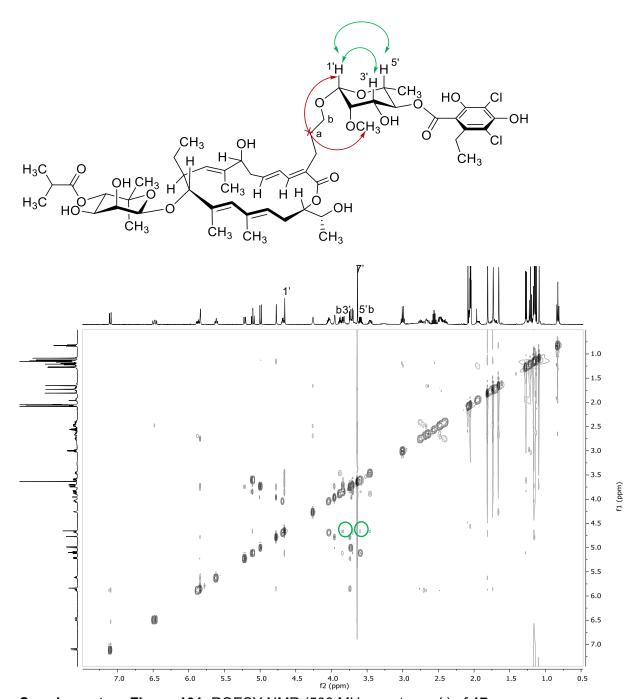
Supplementary Figure 98. TOCSY NMR (500 MHz, acetone- d_6) of 17.



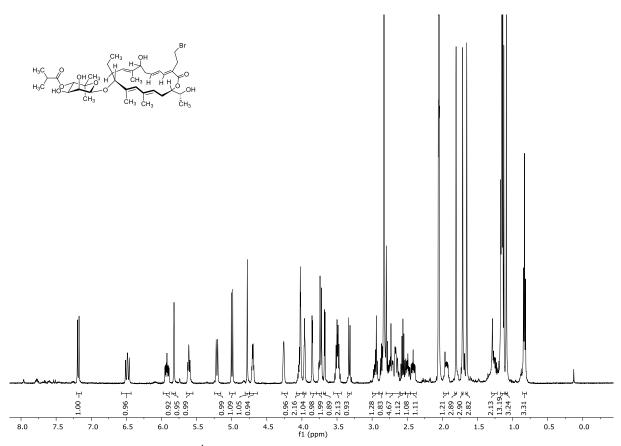
Supplementary Figure 99. HSQC NMR (500 MHz, 126 MHz, acetone-d₆) of 17.



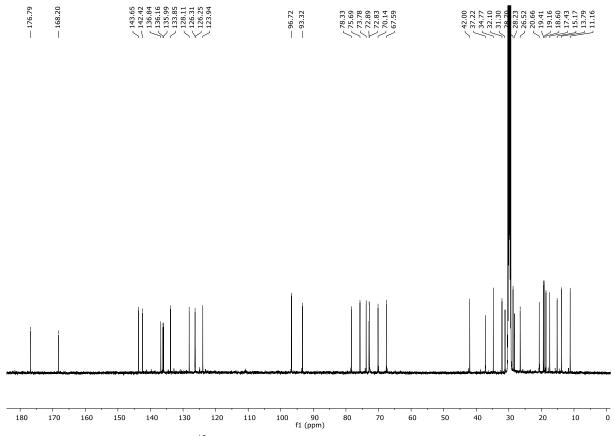
Supplementary Figure 100. HMBC NMR (500 MHz, 126 MHz, acetone- d_6) of 17.



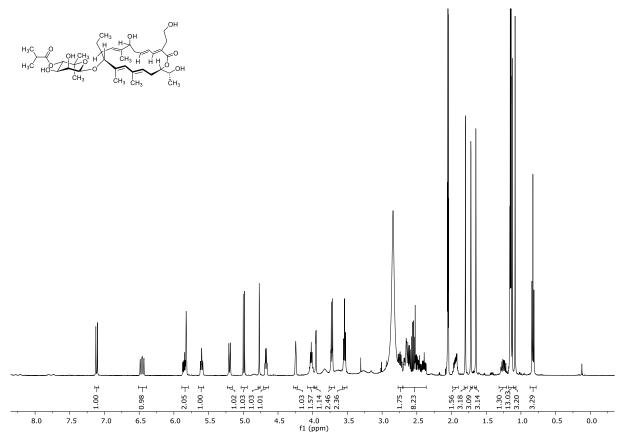
Supplementary Figure 101. ROESY NMR (500 MHz, acetone- d_6) of **17.**



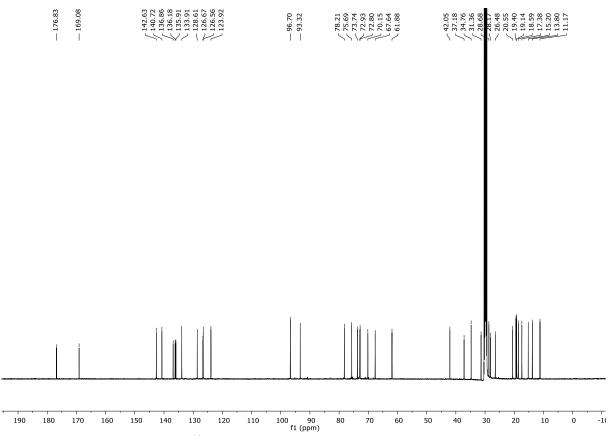
Supplementary Figure 102. ¹H NMR (500 MHz, acetone- d_6) of 16.



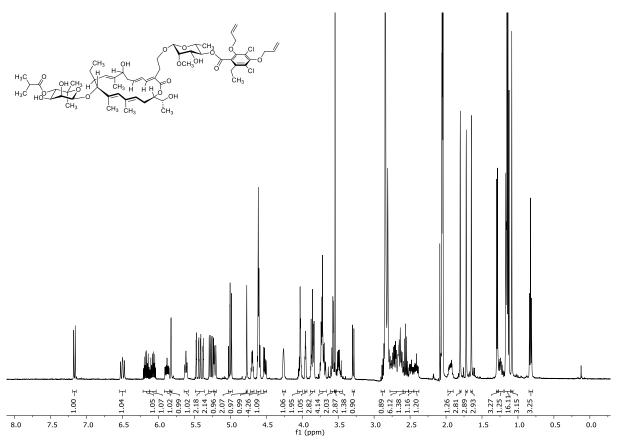
Supplementary Figure 103. 13 C NMR (126 MHz, acetone- d_6) of 16.



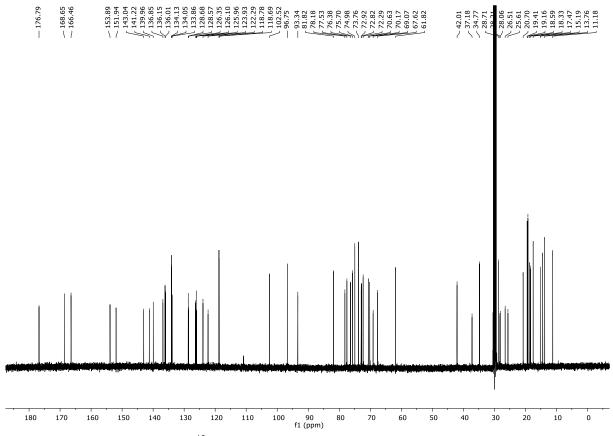
Supplementary Figure 104. ¹H NMR (500 MHz, acetone-*d*₆) of 16.1.



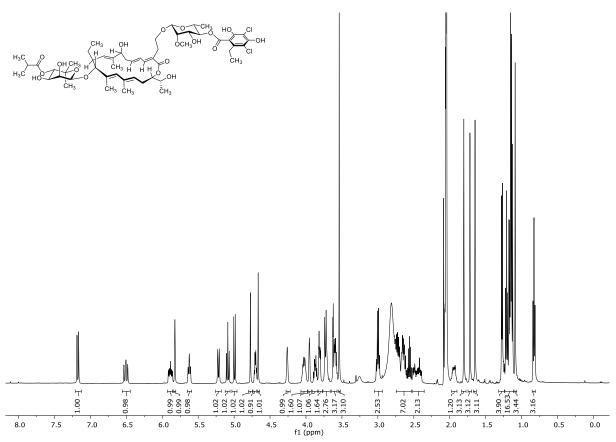
Supplementary Figure 105. 13 C NMR (126 MHz, acetone- d_6) of 16.1.



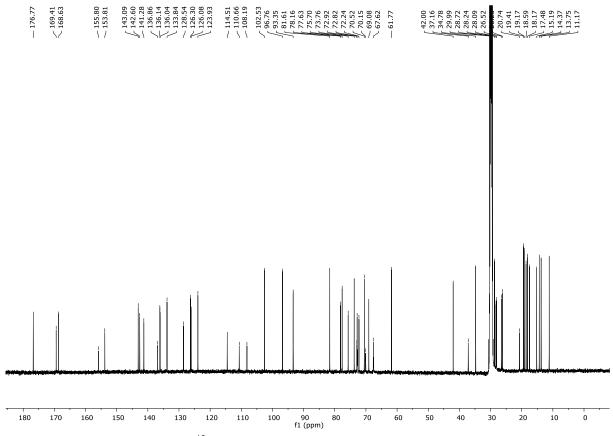
Supplementary Figure 106. ¹H NMR (500 MHz, acetone- d_6) of S4.



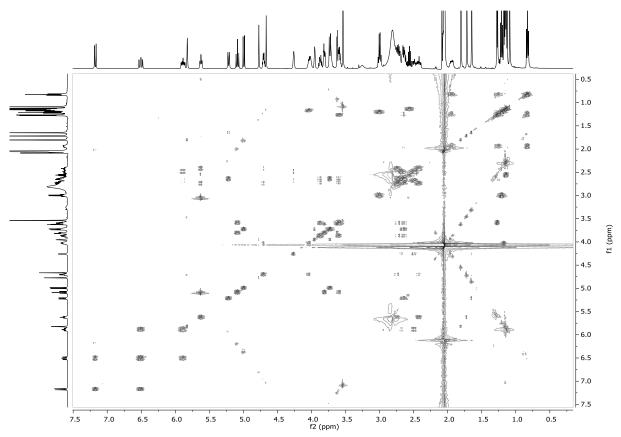
Supplementary Figure 107. 13 C NMR (126 MHz, acetone- d_6) of S4.



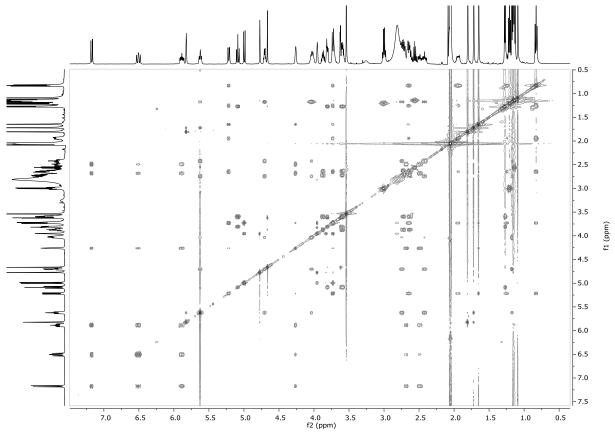
Supplementary Figure 108. ¹H NMR (500 MHz, acetone-*d*₆) of 18.



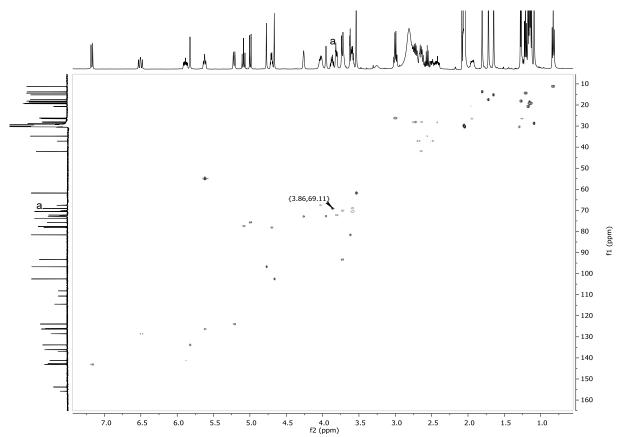
Supplementary Figure 109. 13 C NMR (126 MHz, acetone- d_6) of 18.



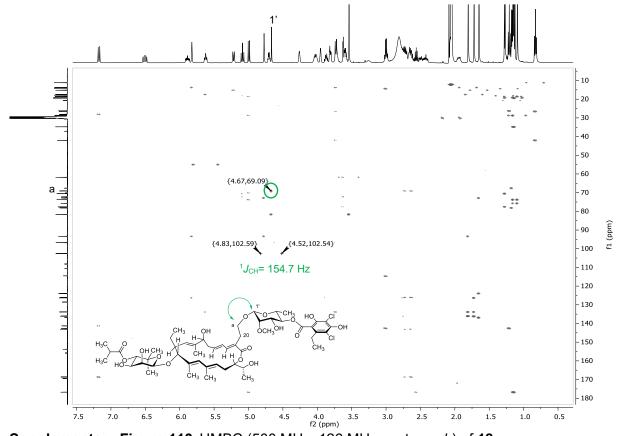
Supplementary Figure 110. COSY NMR (500 MHz, acetone- d_6) of 18.



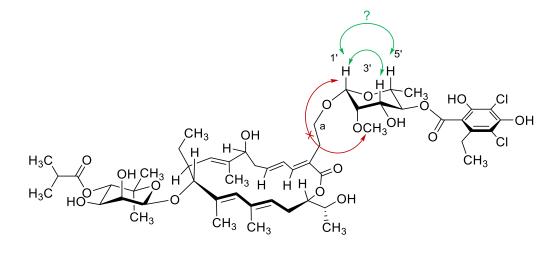
Supplementary Figure 111. TOCSY NMR (500 MHz, acetone- d_6) of 18.

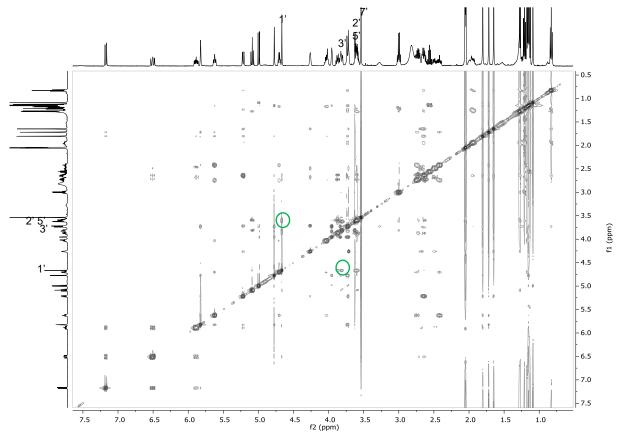


Supplementary Figure 112. HSQC (500 MHz, 126 MHz acetone- d_6) of 18.

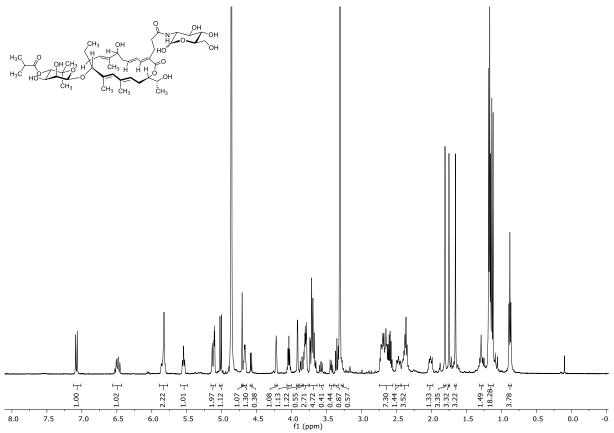


Supplementary Figure 113. HMBC (500 MHz, 126 MHz acetone- d_6) of 18.

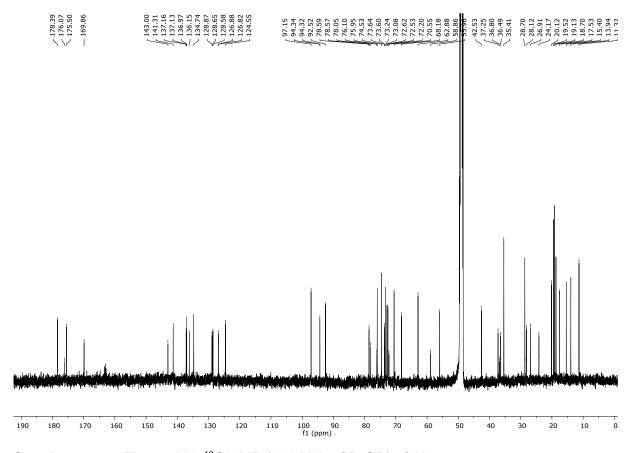




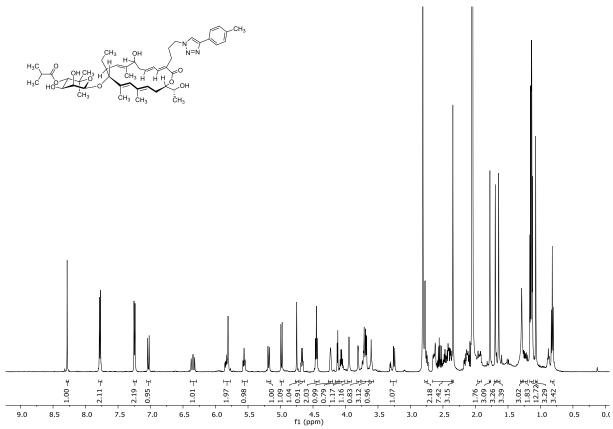
Supplementary Figure 114. ROESY (500 MHz, acetone- d_6) of **18.**



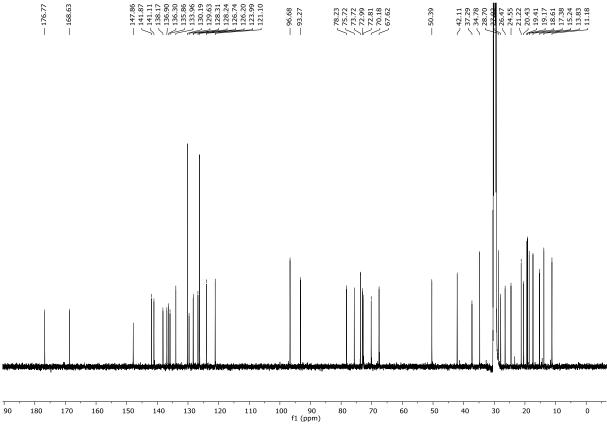
Supplementary Figure 115. ¹H NMR (500 MHz, CD₃OD) of 19.



Supplementary Figure 116. ¹³C NMR (126 MHz, CD₃OD) of 19.



Supplementary Figure 117. ¹H NMR (500 MHz, acetone- d_6) of 20.



Supplementary Figure 118. 13 C NMR (126 MHz, acetone- d_6) of 20.