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Review Article

Functional compounds in tropical fruit processing by-products and intrinsic factors affecting their composition: A review

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ABSTRACT

Tropical fruits, highly demanded in the food industry, generate a considerable amount of waste during processing. These traditionally discarded by-products, such as peels, seeds and pomace, are rich in bioactive compounds, natural molecules that have beneficial properties for human health, as they participate in various metabolic processes in the organism. Among the most prominent compounds are flavonoids, carotenoids, phenolic compounds, tannins and vitamin C. Beyond their health benefits, these compounds have significant industrial value and are widely used in the textile, pharmaceutical, cosmetic, biotechnological and food fields, in the latter especially as preservatives, additives, colorants and others. This review explores the main bioactive compounds found in fruit by-products, highlighting their functional relevance and analyzing the intrinsic or fruitderived factors that influence the composition of these compounds, such as the type of by-product (peels, seeds, bagasse, pomace), the variety of fruit, and the state of maturity at the time of processing. In addition, the extraction methods used to obtain these compounds are addressed, differentiating between conventional techniques, such as solvent extraction, and emerging methods, such as ultrasound-assisted extraction and supercritical fluid extraction, which offer advantages in terms of efficiency and sustainability. The diversity of bioactive compounds and their potential application in various industries highlight the importance of ongoing research in this field. It is necessary to further study the factors that influence the composition of these compounds, as well as the development of more efficient and sustainable extraction methods. These advances will not only add value to food industry waste, but will also contribute to the development of natural products with health benefits

1. Introduction

Tropical fruits are one of the most important crops worldwide (Villacís-Chiriboga et al., 2020), and have been part of the group of basic products since 1970, due to advances in transport, trade agreements and changes in consumer preferences. Tropical fruits represent 3 % of world agricultural food products, positioning them as the third most valuable group within the fruit sector, due to their high export price per unit (Cádiz et al., 2020). They can be classified according to the extent of their cultivation, volume of production and market, in main and secondary (among which are the wild ones). The main ones include mango (Mangifera indica L.), pineapple (Ananas comosus L. Merr), avocado (Persea americana Mill.), papaya (Carica papaya L.) and banana (Musa paradisiaca L.), among others. Secondary fruits include guava (Psidium guajava L.), tamarind (Tamarindus indica L.), açai (Euterpe oleracea Mart.), passion fruit (Passiflora edulis Sims), pomegranate (Punica

granatum L.), coconut (Cocos nucifera Linn.), acerola (Malpighia emarginata D.C.), noni (Morinda citrifolia L.), camu-camu (Myrciaria dubia McVaugh), dragon fruit (Hylocereus spp.), mangosteen (Garcinia mangostana L.), among others.

These fruits are consumed fresh and in products such as juices, sauces, jams, chutneys, ready-to-use peeled fruit, ice creams and others. Their industrial processing generates significant amounts of food by-products, such as trimmings, peels, pomace, seeds, and leaves, which normally have no additional food use (Vilas et al., 2024). The by-product fraction can reach approximately up to 60 % of the whole fruit (J. F. Ayala et al., 2011), which implies enormous losses and waste, being an environmental problem that requires adequate management (Trigo et al., 2020). However, the efficient management of these wastes faces significant challenges, such as the lack of effective technologies to manage large volumes of biomass, the lack of suitable processing methods, and the high costs associated with their treatment (Sharma

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et al., 2016).

The use of by-products or waste from the tropical fruit industry is an alternative that allows the implementation of an industrial circular economy, which leads to an increase in profitability and environmental sustainability (Routray and Orsat, 2019), ultimately giving rise to a greater diversity of products. The use of such waste has been taking place for decades, mainly for animal feed (Cádiz et al., 2020; Clemente et al., 2015), and in some cases its potential as an alternative source of energy production (biogas, bioethanol or biohydrogen) has been explored (Domínguez-Bocanegra et al., 2015). In particular, papaya and pineapple peels have been used for the production of methane through anaerobic digestion (Romero et al., 2020; Arifan et al., 2022); melon, pineapple and mango waste, for the production of bioethanol using a non-conventional yeast Wickerhamomyces sp. (Zanivan et al., 2022) and cashew nut shells for biomethane production (Nikiema et al., 2020). However, in recent years, the use of these by-products has focused on the formulation of products intended for human consumption (Durán-Aranguren et al., 2023), due to their great nutritional quality derived from the high content of bioactive compounds (Sood and Saini, 2022), which provide significant antioxidant and antimicrobial properties (Peschel et al., 2006; O'Shea et al., 2012; Trigo et al., 2020; Sayago-Averdi et al., 2021; Belmonte et al., 2022), preventing them from leaving the food supply chain (Thorsen et al., 2022).

Bioactive compounds are present in all anatomical parts of the fruit (Lachowicz et al., 2019) and the composition of these compounds varies between different fractions, often being significantly higher in the so-called inedible parts of the fruit (Banerjee et al., 2017). Bioactive compounds obtained from tropical fruit residues can act through several pathways in the human body, including interacting with proteins, DNA and other biological molecules. These processes stop oxidation, which inactivates free radicals and increases the potential to modify the composition and metabolic activity of the intestinal microbiota (Ajikumar et al., 2008; El-Hadary and Ramadan, 2019; Azizan et al., 2020). This results in health benefits by reducing the risk of diseases (Gil-Chávez et al., 2013), among which diabetes, cancer and cardiovascular diseases stand out (Şahin et al., 2018; Dobrinčić et al., 2020; Pavlović et al., 2020; Pirozzi et al., 2020), thus contributing to the revalorization of underutilized waste or by-products.

Current literature reviews on bioactive compounds in tropical fruit by-products focus on composition and their application as ingredients in food products (Khan et al., 2024; Chel et al., 2022; Oliveira et al., 2022; Marcillo et al., 2021a; Maqsood et al., 2020). This review stands out by its comprehensive approach, analyzing not only the presence of bioactive compounds in tropical fruit by-products, but also the intrinsic factors that affect their composition, such as the type of by-product, the fruit variety and its maturity stage, aspects rarely explored in previous reviews. It emphasizes on the valorization of agro-industrial waste within a circular economy approach, promoting its use in the development of functional and sustainable products.

2. Bioactive compounds present in tropical fruit by-products

Tropical fruit by-products are a significant source of bioactive compounds, which stand out mainly for their content of polyphenols or phenolic compounds, including flavonoids, phenolic acids and tannins. Also, carotenoids and ascorbic acid or vitamin C, among others (Marcillo et al., 2021a), have been classified as potential sources of antioxidants and contribute to improving health (Kumar et al., 2017). Table 1 presents information on the bioactive compounds extracted from different tropical fruit residues, highlighting the extraction method and the amount of each compound. The analyzed residues come from different parts of the fruits, such as peels, seeds, pomace, flavedo and albedo, and correspond to strawberry, raspberry, acerola, guava, melon, avocado, mango, passion fruit, lemon, papaya, plum, pineapple, watermelon, cashew, seriguela and noni.

 Table 1

 Bioactive compounds in the by-products of tropical fruits

Bioactive compounds	Residues	Quantity	References
Phenolic compounds	Strawberry waste	$7157\pm160~\mu g$ GAE/ $100~g$ $5761\pm12~\mu g$ GAE/	Vázquez et al. (2020)
	Raspberry waste	100 g $19405 \pm 377 \text{ μg}$ $GAE/100 \text{ g}$	
	Acerola pomace	$49644 \pm 760 \mu g$ GAE/100 g $444.01 \pm 45.99 mg$ GAE/100 g	Carvalho et al. (2021)
	Guava pomace	419.35 ± 7.4 mg GAE/100 g 160.33 ± 5.77 mg	(2021)
		GAE/100 g 172.56 \pm 15.13 mg GAE/100 g	
	Acerola waste Melon peels	106.8 ± 5.5 mg GAE/ 100 g 296.03 ± 7.75 mg	Poletto et al. (2021) Gómez et al.
	Hass avocado peel	GAE/100 g 190 ± 3 mg GAE/g dry extract	(2021) Rojas et al. (2022)
	Hass avocado seed	60 ± 2 mg GAE/g dry extract	(2022)
	Mango peels	2931 ± 175–5149 ± 154 mg GAE/100 g 22.57 ± 0.38 mg GAE/100 g	Marcillo et al. (2021b) Zahid et al. (2022)
	Cashew bagasse	$293 \pm 2.84 – 265 \pm 3.88$ mg GAE/100 g	Cruz et al. (2022)
	Lemon Flavedo Lemon Albedo	2.89 ± 0.27 mg GAE/ 100 g 2.42 ± 0.26 mg GAE/	Rosa et al. (2023)
	Passion Fruit	100 g 2.07 ± 0.05 mg GAE/	Vitor et al.
	Peels Pomegranate Peel	$\begin{array}{c} \text{g} \\ 25.92 \pm 0.13 \text{ mg} \\ \text{GAE/g} \\ 40.55 \pm 0.23 \text{ mg} \\ \text{GAE/g} \\ 42.92 \pm 0.65 \text{ mg} \end{array}$	(2022) Kupnik et al. (2022)
	Sweet apple peel	$\begin{array}{l} \text{GAE/g} \\ \text{1043} \pm 224 \text{ mg GAE/} \\ \text{L} \end{array}$	Tlais et al. (2023)
	Papaya peel	$\begin{array}{c} 2.79 \pm 0.12 \text{ mg GAE/} \\ \text{g} \end{array}$	Zhou et al. (2023)
	Papaya seed Plum waste	2.80 ± 0.11 mg GAE/ g 10.24 ± 0.37 mg	Xiang et al.
	Purple passion fruit seeds Sweet apple peel	GAE/100 g 112.81 ± 13 mg GAE/100 g 770.18 ± 6.13 mg	(2023) da Costa et al. (2023)
	Watermelon rind	GAE/100 g $7.07 \pm 0.86~\mu g~GAE/mg$	Yusoff et al. (2023)
	Noni seeds	$15.10 \pm 0.58 \ \mu g$ GAE/mg $12.48 \pm 0.23 \ mg$ GAE/100 g	Dzah et al. (2024)
	Lemon pomace	$1.47 \pm 0.08~\mu g$ GAE/ mg $1.18 \pm 0.04~\mu g$ GAE/	Iervese et al. (2024)
	Seriguela peel	$\begin{array}{l} \text{mg} \\ \text{2563.30} \pm 331.60 \text{ mg} \\ \text{GAE/100 g} \end{array}$	Cangussu et al. (2024)
	Seriguela seeds	991.18 ± 52.33 mg GAE/100 g	
Flavonoids	Acerola pomace	213.38 ± 35.8 mg QE/100 g 207.82 ± 21.09 mg QE/100 g	Carvalho et al. (2021)

Table 1 (continued)

Bioactive compounds	Residues	Quantity	References	
	Guava pomace	44.45 ± 4.00 mg QE/		
	1	100 g		
		22.26 ± 0.20 mg QE/		
		100 g		
	Mango peels	502 \pm 23.2–779 \pm	Marcillo et al.	
		11.9 mg QE/100 g	(2021b)	
	Watermelon peel	4.97 \pm 0.29 μ g RE/g	Yusoff et al.	
		$6.16\pm0.09~\mu g$ RE/g	(2023)	
	Papaya peel	1.25 ± 0.05 mg QE/g	Zhou et al.	
	Papaya seed	0.16 ± 0.01 mg QE/g	(2023)	
	Seriguela skin	80.50 ± 3.56 mg	Cangussu et al	
	Coriguala sonda	QCE/100 g 28.96 ± 0.48 mg	(2024)	
	Seriguela seeds	QCE/100 g		
	Lemon pomace	0.47 ± 0.02 mg CE/	Iervese et al.	
	Demon pointee	mL	(2024)	
		0.33 ± 0.02 mg CE/	(2021)	
		mL		
Anthocyanins	Strawberry waste	$0.49 \pm 0.03 \text{ mg/}100$	Vázquez et al.	
		g	(2020)	
		$2.83\pm0.03~\text{mg/}100$		
		g		
	Raspberry waste	$0.18\pm0.02~\text{mg/}100$		
		8		
		$1.28 \pm 0.06 \ \text{mg/}100$		
	Pineapple peel	g 208 mg/g	Luan et al.	
	гшеарріе рееі	206 Hig/g	(2023)	
	Plum waste	$68.95\pm1.03~\mu\text{g/g}$	Xiang et al.	
	Tium waste	00.90 ± 1.00 μ ₈ / δ	(2023)	
Tanins	Cashew bagasse	299 ± 3.09–288 ±	Cruz et al.	
		4.50 mg TA/100 g	(2022)	
	Papaya peel	$1.88\pm0.06~\text{mg CE/g}$	Zhou et al.	
	Papaya seed	$0.19\pm0.01~\text{mg CE/g}$	(2023)	
	Seriguela skin	$1849.29 \pm 333.29 \ \text{mg}$	Cangussu et al	
		TAE/100 g	(2024)	
	Seriguela seeds	718.50 ± 98.65 mg		
		TAE/100 g		
Carotenoids	Cashew fibers	0.29 mg/100g	Servent et al.	
	Managa shalla	0.38 mg/100g	(2020)	
	Mango shells	4.53 ± 0.03 – 5.69 ± 0.20 mg/ 100 g	Marcillo et al. (2021a)	
	Cashew	$134.41 \pm 1.40 \mu \text{g/g}$	Coelho et al.	
	pseudofruit	$165.47 \pm 1.79 \mu\text{g/g}$	(2022).	
	Cashew bagasse	1.71 ± 0.28 $\pm 4.72 \pm$	Cruz et al.	
		0.60 μg/100g	(2022)	
	Pineapple shells	71 pg/mL	Luan et al.	
	Seriguela seeds	$991.18 \pm 52.33 \text{ mg}$	(2023)	
		GAE/100 g		
Ascorbic acid	Cashew bagasse	88 \pm 3.81–204 \pm	Cruz et al.	
		4.26 mg/100g	(2022)	
	Noni seeds	$1.79\pm0.11~\text{mg/g}$	Dzah et al.	
			(2024)	

2.1. Polyphenols or phenolic compounds

In tropical fruit by-products, phenolic compounds or polyphenols (characterized by having at least one phenol group and an aromatic ring that is attached to a hydroxyl group) are found in greater quantities, including phenolic acids, flavonoids and tannins (J. Ayala et al., 2011; Belmonte et al., 2022; López et al., 2021; Pereira, 2018). Among the residues with high polyphenol content, citrus residues (Rosa et al., 2023; Vitor et al., 2022), mango peels (Marcillo et al., 2021a; Zahid et al., 2022), seriguela (Cangussu et al., 2024), passion fruit (da Costa et al., 2023), acerola (Poletto et al., 2021), avocado (Rojas et al., 2022), among others, stand out. Among the residues analyzed, avocado peel had the highest content of these compounds, followed by mango and seriguela peel, indicating that the polyphenol content is higher in the peel than in the seed (Table 1).

2.1.1. Phenolic acids

The term "phenolic acids" mainly describes phenolic compounds that have a carboxylic acid group. They are secondary metabolites produced by plants and found in the plant cell wall (Mandal et al., 2010). They are found in a variety of plant-based foods (seeds, fruit peels, and vegetable leaves). They are usually present bound forms, such as amides, esters, or glucosides, and are rarely in free form (Pereira et al., 2009). They are characterized by being the main polyphenols, which are effortlessly absorbed in the intestine (Kumar and Goel, 2019) and have great antioxidant activity, providing health benefits, significantly reducing the risk of many diseases related to oxidative stress (cancers, diabetes, and cardiovascular diseases) (Sehrawat et al., 2022).

In the food industry, they have been used to improve the organoleptic (flavor, astringency, and hardness), nutritional, and antioxidant properties of foods (Shahidi and Naczk, 2003). They have also been recognized as antimicrobial compounds due to their ability to be metabolized by microorganisms, thus providing an alternative as a preservative (Faustino et al., 2019), which reduces the use of chemicals harmful to the environment. The application of these compounds in high concentrations in food matrices is limited, because unpleasant changes are generated in their sensory properties, promoting the encapsulation or immobilization of said compounds with a view to improving their functionality (Gutiérrez et al., 2018).

The most common phenolic acid in these residues is gallic acid, which has a high antioxidant, anti-inflammatory and neuroprotective capacity due to its ability to neutralize free radicals (Frešer et al., 2021; Xu et al., 2021). It is primarily present in guava waste (4.69 \pm 0.01 $\mu g/g$) (Carvalho et al., 2021), mango peels (28.6 \pm 2.22–28.6 \pm 2.22 mg/100g) (Marcillo et al., 2021a), and papaya peels (0.33 \pm 0.20 μ g/g) (Khan et al., 2021). Chlorogenic acid is another phenolic acid that protects cells from free radical damage and, at the same time, exhibits antimicrobial activity against various bacteria, especially Gram-positive bacteria (Chaves and Esquivel, 2019; X. Guo et al., 2019). It is present in acerola and guava waste (42.54 \pm 0.05 $\mu g/g$ and 6.89 \pm 0.28 $\mu g/g$, respectively) (Carvalho et al., 2021). Likewise, ferulic acid shows high antioxidant properties related to its phenolic core and an unsaturated side chain that forms resonance-stabilized structures (Celebioglu and Uyar, 2020; Gu et al., 2021). It is commonly found in papaya peels and seeds (0.3 \pm 0.16 mg/100g and 3.86 \pm 0.53 mg/100g respectively) (Khan et al., 2021) and in lemon pomace (25.45 \pm 1.36 mg/100g) (Iervese et al., 2024).

2.1.2. Flavonoids

Flavonoids are low molecular weight polyphenolic phytochemicals secreted as a secondary metabolite of plants (Donadio et al., 2021). They are responsible for color, fragrance and flavor characteristics and are considered the most abundant pigments along with chlorophyll and carotenoids (Rodríguez et al., 2020). The main sources of these compounds are fruits, vegetables, seeds and flowers. Although they are also found in beer, wine, green tea and agro-industrial waste derived from the same primary sources (Stefanello et al., 2018).

Flavonoids are considered natural antioxidants for their ability to eliminate free radicals and inhibit their formation. As well as the chelation of metal ions to inhibit lipid peroxidation (Chamira and Preethi, 2014). Due to these characteristics, they have a wide variety of physiological functions in the human body, such as anti-inflammatory, anticancer, cardioprotective, antimicrobial and antiviral (Dias et al., 2021). In the food field, their use as preservatives focuses on the prevention of lipid oxidation and the inhibition of microbial growth. They are also used for the protection of vitamins and enzymes, as additives in human dietary supplements and animal feed, flavorings and colorings (e.g., anthocyanins) (Ruiz et al., 2017). In tropical fruit residues, flavonoids are mainly found in mango peels (Marcillo et al., 2021a), acerola pomace and guava (Carvalho et al., 2021; Rezende et al., 2018), with the first two having the highest amount of flavanoids (Table 1).

Anthocyanins are part of the flavonoid family, which constitute one

of the largest groups of water-soluble pigments in the plant kingdom. They are responsible for giving color to flowers and fruits (Santos and González, 2019) and are widely distributed in fruits, vegetables and cereals with reddish tones. They are generally used as natural coloring agents replacing synthetic ones (Albuquerque et al., 2020). Epidemiological studies, in vitro trials and dietary intervention have supported the possibility that they play multiple roles as antivirals, antioxidants and anti-inflammatories, and promoters of intestinal health (Gull et al., 2022). Anthocyanins have also proven to have great potential as substitutes for antibiotics, but their use is limited due to their unstable structure and low bioavailability (Guo and Shahidi, 2024). Their stability is affected by external factors such as pH, temperature, light, solvent, oxygen, enzymes, other flavonoids, proteins and metal ions (Fang, 2014). These compounds are found mainly in tropical fruits waste with red, purple and blue peels, such as strawberries and raspberries (Vázquez et al., 2020) and plums (Xiang et al., 2023).

2.1.3. Tannins

Tannins are categorized within water-soluble polyphenols that differ in terms of solubility and site of accumulation in the plant (Petchidurai et al., 2019). They are classified into three different chemical structures: hydrolysable tannins, flavonoid-based tannins and phlorotannins. Hydrolysable tannins are derived from simple phenolic acids and carbohydrates, where hydroxyl groups are partially or completely esterified with phenolic groups. Flavonoid-based tannins are synthesized through flavins and catechins, which are well known for their antioxidant properties. For their part, phlorotannins are characteristic of brown algae (*Phaeophyceae*), they are composed of phloroglucinol units linked by carbon-carbon bonds. Phlorotannins present a wide range of biological activities, including antioxidant, anti-inflammatory, antiviral and anticancer properties (Lopes et al., 2012).

The astringency of most foods is attributed to their tannin content (Dheeraj and Mishra, 2023), which occurs when salivary proteins precipitate mainly as a result of the formation of insoluble complexes between proteins and tannins (astringent polyphenolic compounds) (Tezotto et al., 2018). This process depends on the average degree of polymerization and the compound; the higher the degree of tannins, the greater their affinity to bind to salivary proteins and cause astringency (Soares et al., 2020). Some tropical fruit waste with high tannin content includes Cashew nut pomace (Cruz et al., 2022), papaya peel and seed (Zhou et al., 2023), as well as seriguela skin and seed (Cangussu et al., 2024). Among them, the seriguela skin stands out for having the highest concentration of this compound (Table 1).

2.2. Carotenoids

Carotenoids are considered precursors of vitamin A (Meléndez, 2019; Meléndez et al., 2023), and are responsible for the yellow, orange, and red colors in fruits (Meléndez et al., 2023). β -carotene, α -carotene, β -cryptoxanthin, lycopene, lutein, and zeaxanthin are the carotenoids most frequently found in foods and represent the greatest benefits to human health (Beltrán et al., 2015). They participate in important biological actions that contribute to reducing the risk of diseases such as cancer, cardiovascular, skin, bone, eye, and metabolic diseases, and are beneficial for cognitive development (Meléndez, 2019).

They are widely used in the food, supplement, and pharmaceutical industries for their coloring and nutraceutical properties. They exhibit stronger antioxidant properties than other pigments, are considered prooxidants, and enhancers of communication between junctional cells, but they degrade rapidly when exposed to light, oxygen, moisture and heat during processing and storage, causing undesirable changes and reduced bioactivity (Meléndez et al., 2023). Among the changes that occur are color loss and the production of unpleasant flavors and aromatic compounds, which reduce consumer acceptability and shorten the shelf life of the final product (Vieira et al., 2020).

The intensity of the yellow, orange or red color of the fruit peel

depends largely on the composition and concentration of the colored carotenoids (Ma et al., 2013; Luan et al., 2024). The color of the peel is an important characteristic of appearance, which is an indicator of fruit ripeness and quality, directly affecting consumer preference. Fruit waste such as cashew, seriguela and mango present shades ranging from yellow to orange, being great representatives of these compounds (Cangussu et al., 2024; Coelho et al., 2022; Cruz et al., 2022; Marcillo et al., 2021a). Among them, cashew waste stands out for their high content of carotenoids, with a higher concentration in the pomace compared to the pseudofruit, whose shades vary from yellow to red (Table 1).

2.3. Ascorbic acid

Ascorbic acid is commonly known as vitamin C and is one of the basic and necessary compounds for the proper functioning of the human body. It contributes to many processes, such as the strengthening and sealing blood vessels, the regulation of microbial absorption by leukocytes, the reduction of cholesterol levels, and is involved in skin rejuvenation and acceleration of wound healing (Gegotek and Skrzydlewska, 2023).

It is considered an important water-soluble antioxidant due to its great capacity to donate electrons. It has a protective effect on cellular oxidation, is capable of reducing the number of free radicals produced during metabolic processes, and stimulates iron absorption. It is thermolabile and easily oxidized in the air. It is bioavailable in fruits, vegetables, juices, and fortified foods (Ramírez and Quiles, 2005). The waste of citrus, cashew, guava, acerola and passion fruit seeds contain ascorbic acid (Barros et al., 2017; Carvalho et al., 2021, Cruz et al., 2022). Table 1 shows the high content of these compounds in uncommon waste such as noni seeds and cashew bagasse.

When analyzing the tropical fruit waste, it is observed that mango, acerola, seriguela and cashew nuts have the highest concentrations of bioactive compounds, standing out for their richness in phenolic compounds, flavonoids, tannins and carotenoids. The high concentration of these compounds suggests a great potential for their valorization in the food, pharmaceutical and cosmetic industries, contributing to the development of functional and sustainable products. In addition, the optimization of extraction methods could further improve the recovery and utilization of these residues, strengthening the circular economy and the reduction of agro-industrial waste.

3. Extraction methods of bioactive compounds

Extraction is essential for the recovery of bioactive compounds, for which conventional techniques have been developed (e.g., Soxhlet extraction, maceration and hydro-distillation) and extended to an industrial scale, but have generated environmental problems due to $\rm CO_2$ emissions and the use of dangerous chemical solutions. Therefore, in recent years alternative techniques have been used, such as ultrasound-assisted extraction, ohmic extraction, pulsed electric fields, high pressures, supercritical fluid extraction and others, seeking high yields and recovery effectiveness. This results in shorter extraction times and less environmental damage, tending towards green and environmentally friendly extraction techniques (Vânia and Ramin, 2018; Zhang et al., 2018).

Table 2 shows comparison of techniques used for the extraction of different bioactive compounds from tropical fruit waste. Phenolic compounds, flavonoids, anthocyanins and carotenoids are analyzed using various extraction methods.

Most of the bioactive compounds from tropical fruits are extracted by conventional methods, including maceration, digestion, percolation, decoction, hydrodistillation, and Soxhlet extraction, which usually require a large volume of chemical solvents (acetone, methanol, ethanol, and others). Despite being low-cost techniques, they exhibit long extraction times, low extraction efficiency and degradation of the compounds due to increased temperature. As a result, other extraction

Table 2Comparison of techniques used for the extraction of bioactive compounds in tropical fruit by-products.

Bioactive compounds	Residues	Extraction methods ^a	Quantity	References
Phenolic	Strawberry	EES	7.157 \pm	Vázquez
compounds	waste		0.160 mg	et al. (2020)
			GAE/100 g	
		OES	5.761 ± 0.0	
	n 1		12 mg GAE/	
			100 g	
	Raspberry	EES	19.405 ±	
	waste	OES	0.377 mg	
			GAE/100 g 49.644 \pm	
		OES	0.760 mg	
			GAE/100 g	
	Acerola	ECS - A	444.01 ±	Carvalho
	pomace	EGD 11	45.99 mg	et al. (2021)
	pomace		GAE/100 g	ct ai. (2021)
		ECS - US	419.35 ± 7.4	
		200 00	mg GAE/100	
			g gran, 100	
	Guava	ECS - A	$^{8}_{160.33~\pm}$	
	pomace		5.77 mg GAE/	
	P		100 g	
		ECS - US	$172.56 \pm$	
		200 00	15.13 mg	
			GAE/100 g	
	Mango peels	OES - A	2931 \pm	Marcillo
	0 1		175–5149 \pm	et al. (2021a)
			154 mg GAE/	
			100 g	
		ECS	22.57 ± 0.38	Zahid et al.
			mg GAE/100	(2022)
			g	
	Pomegranate	ECS - US	25.92 ± 0.13	Kupnik
	peel		mg GAE/g	et al. (2022)
		ECS - SE	40.55 ± 0.23	
			mg GAE/g	
		ECS - CM	42.92 ± 0.65	
			mg GAE/g	
	Watermelon rind	ECS - CM	0.00707 \pm	Yusoff et al
			0.00086 mg	(2023)
			GAE/mg	
		ECS - US	$0.01510~\pm$	
			0.00058 mg	
			GAE/mg	
	Lemon	ECS - CM	0.00147 \pm	Iervese
	pomace		0.08 mg GAE/	et al. (2024)
			mg	
		ECS - US	0.00118 ±	
			0.04 mg GAE/	
			mg	
Flavonoids	Acerola	ECS- A	213.38 \pm	Carvalho
	pomace		35.8 mg QE/	et al. (2021)
			100 g	
		ECS - US	207.82 \pm	
			21.09 mg QE/	
			100 g	
	Guava	ECS- A	$100~{\rm g} \\ 44.45 \pm 4.00$	
	Guava pomace		_	
		ECS - US	$\begin{array}{c} 44.45 \pm 4.00 \\ \text{mg QE/100 g} \\ 22.26 \pm 0.20 \end{array}$	
	pomace	ECS - US	$\begin{array}{c} 44.45 \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \end{array}$	
	pomace Watermelon		$\begin{array}{c} 44.45 \pm 4.00 \\ \text{mg QE/100 g} \\ 22.26 \pm 0.20 \\ \text{mg QE/100 g} \\ 0.00497 \pm \end{array}$	
	pomace	ECS - US	$\begin{array}{c} 44.4\bar{5} \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \ \pm \\ 0.00029 \ mg \end{array}$	Yusoff et al (2023)
	pomace Watermelon	ECS - US	$\begin{array}{c} 44.45 \pm 4.00 \\ \text{mg QE/100 g} \\ 22.26 \pm 0.20 \\ \text{mg QE/100 g} \\ 0.00497 \pm \end{array}$	
	pomace Watermelon	ECS - US	$\begin{array}{c} 44.4\bar{5} \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \ \pm \\ 0.00029 \ mg \end{array}$	
	pomace Watermelon	ECS - US ECS- M	$\begin{array}{c} 44.4\bar{5} \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \pm \\ 0.00029 \ mg \\ RE/g \end{array}$	
	pomace Watermelon	ECS - US ECS - US	$\begin{array}{c} 44.45 \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \pm \\ 0.00029 \ mg \\ RE/g \\ 0.00616 \pm \end{array}$	
	pomace Watermelon	ECS - US ECS- M	$\begin{array}{c} 44.45 \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \pm \\ 0.00029 \ mg \\ RE/g \\ 0.00616 \pm \\ 0.0009 \ mg \end{array}$	
	pomace Watermelon rind	ECS - US ECS - US	$\begin{array}{c} 44.45 \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \pm \\ 0.00029 \ mg \\ RE/g \\ 0.00616 \pm \\ 0.0009 \ mg \\ RE/g \end{array}$	(2023) Iervese
	pomace Watermelon rind Lemon	ECS - US ECS - US	$\begin{array}{c} 44.45 \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \pm \\ 0.00029 \ mg \\ RE/g \\ 0.00616 \pm \\ 0.0009 \ mg \\ RE/g \\ 0.47 \pm 0.02 \end{array}$	(2023) Iervese
	pomace Watermelon rind Lemon	ECS - US ECS - US ECS - M	$\begin{array}{c} 44.45 \pm 4.00 \\ \text{mg QE/100 g} \\ 22.26 \pm 0.20 \\ \text{mg QE/100 g} \\ 0.00497 \pm \\ 0.00029 \text{ mg} \\ \text{RE/g} \\ 0.00616 \pm \\ 0.0009 \text{ mg} \\ \text{RE/g} \\ 0.47 \pm 0.02 \\ \text{mg CE/mL} \end{array}$	(2023) Iervese
Anthocyanins	pomace Watermelon rind Lemon	ECS - US ECS - US ECS - M	$\begin{array}{c} 44.45 \pm 4.00 \\ mg \ QE/100 \ g \\ 22.26 \pm 0.20 \\ mg \ QE/100 \ g \\ 0.00497 \pm \\ 0.00029 \ mg \\ RE/g \\ 0.00616 \pm \\ 0.0009 \ mg \\ RE/g \\ 0.47 \pm 0.02 \\ mg \ CE/mL \\ 0.33 \pm 0.02 \\ \end{array}$	

Table 2 (continued)

Bioactive compounds	Residues	Extraction methods ^a	Quantity	References
		OSE	2.83 ± 0.03 mg/100 g	
	Raspberry waste	EES	0.18 ± 0.02 mg/100 g	
		OSE	1.28 ± 0.06 mg/100 g	
Carotenoids	Cashew fibers	M	0.29 mg/100g	Servent
		M - ET	0.38 mg/100g	et al. (2020)
	Cashew	ECS- M	$0.13441~\pm$	Coelho
	pseudofruit		0.00140 mg/g	et al.
	-	ECS- US	$0.16547~\pm$	(2022).
			0.00179 mg/g	

^a ECS: Extraction with chemical solvents; A: Agitation; OSE: Organic solvent extraction; EES: Extraction with eutectic solvent; US: Ultrasound; M: Maceration; ET: Enzyme treatment; SE: Soxlet.

methodologies have been investigated that are more sustainable, faster, more reliable, energy efficient and environmentally friendly (Kaderides et al., 2019). Within these emerging methodologies, ultrasound is the most frequently used, as it is more cost-effective (Kupnik et al., 2022).

Ultrasound-assisted extraction allows to increase extraction performance due to the formation of microscopic bubbles that release large amounts of energy (Briones et al., 2015). It uses acoustic-mechanical waves to generate high shear, turbulence (Shirsath et al., 2017), and cavitation, causing several effects in the liquid, such as shock waves and high-velocity liquid jets (Sharma et al., 2023). It is widely used in the extraction of volatile compounds due to the mechanical effect generated by acoustic cavitation, which causes the release of intracellular products by breaking the cell wall, thus increasing the extraction of volatile substances (Zamanipoor et al., 2020). This effect can be evidenced in the carotenoid content of the cashew pseudofruit (Coelho et al., 2022), in the phenolic compounds of guava residues (Carvalho et al., 2021) and watermelon (Yusoff et al., 2023). However, the flavonoid content in acerola, guava (Carvalho et al., 2021) and lemon (Iervese et al., 2024) residues decreased, which may be due to the extraction time, since high exposure to sonication increases the temperature and consequently the material to be extracted degrades (Qiu et al., 2020).

The use of deep eutectic solvents allowed for increased extraction of phenolic compounds (Vázquez et al., 2020), as they have much lower melting points compared to the individual components of the food matrix, which gives them high thermal stability and biodegradability (Cao et al., 2022, 2024; Płotka-Wasylka et al., 2021). However, it should not be applicable for anthocyanins, as the recovery of this compound is not efficient with respect to the traditional method using acetone.

Gas-expanded liquids are a novel extraction technique, using a liquid solvent that acts as the main solvent and a compressible gas to assist in the extraction (Strieder et al., 2024), making it an environmentally friendly method, resulting in a solvent with higher extraction power, even at low temperatures (Abderrezag et al., 2022; Herrero et al., 2017; Zabot et al., 2021). By using this technique, the yield of obtaining phenolic compounds was increased compared to conventional methods in acerola waste (106.8 \pm 5.5 mg GAE/100 g) (Poletto et al., 2021).

Another technique is pressure-driven membranes, which are an alternative to high temperature treatments because they avoid damaging the functional characteristics of the products (di Corcia et al., 2022). Among these membrane processes is cross-flow microfiltration, which allows the separation of particles with very small diameters (Khan et al., 2023), where the important thing is the retentivity, since the bioactive compounds accumulate there (di Corcia et al., 2020; di Corcia et al., 2022). This technique has been applied in conjunction with enzymatic liquefaction in cashew fiber extract, allowing to multiply the carotenoid content, reducing energy costs and increasing the production volume (Servent et al., 2020).

These characteristics have allowed alternative methodologies to awaken the interest of researchers as future application prospects, as they are more efficient due to high yields for the recovery of bioactive compounds (Vidal et al., 2024). However, the choice of the appropriate method depends on factors such as the type of bioactive compound, the plant matrix and the available resources. The industrial implementation of these emerging techniques requires a careful evaluation of the costs, since the investment in equipment may be higher, the reduction in operating costs and processing times may compensate this investment. Likewise, industrial-scale implementation is feasible, but may require technical adaptations in the case of ultrasound, deep eutectic solvents and gas-expanded liquids (Enríquez et al., 2023; Khan et al., 2024; Wong et al., 2020).

4. Factors affecting the content of bioactive compounds in tropical fruit by-products

The composition of bioactive compounds from tropical fruit by-products is influenced by extrinsic factors such as climate, soil type and geographical location (Ahmed et al., 2020; Vignesh et al., 2024), and intrinsic or fruit-derived factors such as type of by-product (peel, pomace or seed), plant variety and fruit maturity at harvest (Fig. 1) (Ceccarelli et al., 2021; Lolletti et al., 2021; Marcillo et al., 2021a).

4.1. By-product type

The peel, seed and pomace of the fruits have different compositions of bioactive compounds. In the outermost parts of the fruits (peels), there is usually a higher content of bioactive compounds due to the differences in the composition of their cell wall (Resende et al., 2019; Cangussu et al., 2021). The high levels of bioactive compounds present in the fruit peel act as a defensive barrier, helping to protect and preserve the internal parts and seeds against possible threats from insects and microorganisms (Jayarathne et al., 2024).

The peel is the barrier between the fruit and the environment. It is exposed to different environmental factors (sunlight, insect bites, temperature, wind and others) that cause stress in plants, which induces the

secondary metabolism of bioactive compounds, mainly for defense purposes, thus increasing the levels of these metabolites, among which phenolic compounds stand out (Ashraf et al., 2018; Dzah et al., 2024). Likewise, flavonoids and anthocyanins are significantly higher in the peel than in the seeds, which is attributed to the accumulation of pigments in this part of the fruit (Jayarathne et al., 2024).

Seeds are vital structures that contain the reserves needed for the germination of a new plant. They contain bioactive compounds, mainly phenolic compounds, which help protect the embryo (Samtiya et al., 2021). In seeds, the accumulation of these compounds is related to protection against abiotic and biotic stress during their development and germination (Jeon et al., 2022), and the expenditure of the compounds in these processes decreases their composition.

The pomace consists of a mixture of shell and seed residues, characterized by a distinctive chemical composition that distinguishes it from the shell and seed individually. The bioactive compounds from the pomace, such as flavonoids, carotenoids and polyphenols, act as an immune system for the plant, protecting it from various threats and ensuring its survival (How and Nyam, 2024).

The levels of phenolic compounds, anthocyanins, flavonoids, tannins and ascorbic acid in different parts of the fruit can fluctuate in response to environmental factors such as drought, temperature variations, pollution, ultraviolet light and pathogen attacks, which significantly influence the synthesis and accumulation of bioactive compounds (Altaf et al., 2024).

For instance, the peel of papaya and seriguela have higher flavonoid content than the seeds of both fruits (Table 1). Also, phenolic compounds are higher in the peel of purple passion fruit, seriguela, and Hass avocado (Rojas et al., 2022; da Costa et al., 2023; Zhou et al., 2023), which is associated with their antioxidant effect mainly attributed to the chelating potential conferred by the chemical structure of phenolic compounds (Prietsch et al., 2014).

4.2. Variety

Within the same species of plants, different varieties can be found, which differ in size, color, shape, genotype and, above all, adaptation to

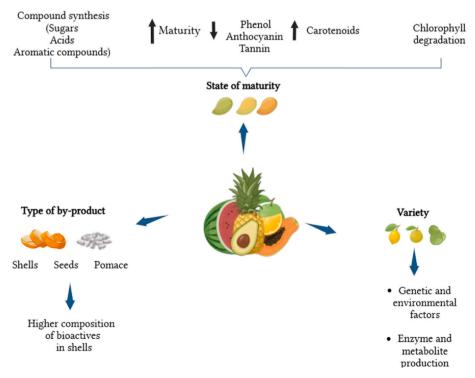


Fig. 1. Intrinsic factors affecting the content of bioactive compounds in fruit by-products.

different climates (Muhie, 2023). Variety is a factor that influences the composition of bioactive compounds (Ceccarelli et al., 2021; Sahie et al., 2023), due to a complex interaction of genetic and environmental factors (Peron et al., 2024). These characteristics confer biochemical properties to the fruit, influencing the production and accumulation of bioactive compounds (Ghahremani et al., 2023).

Each variety has a characteristic set of genes that code for the production of specific enzymes and secondary metabolites. These genetic differences translate into variations in the concentration and type of bioactive compounds present in the fruit (Sheidai et al., 2024). The genes code for enzymes that catalyze reactions in metabolic pathways leading to the synthesis of bioactive compounds. Variations in these genes can alter the speed and efficiency of these pathways, affecting the composition of the compounds produced (Kashyap et al., 2022).

The dynamic interactions between multiple hormones, transcription factors and genetic modifications establish the complex regulatory network that controls the expression levels of genes related to the composition of bioactive compounds. Also, interactions between genotype and environment have a pronounced effect on fruit carotenoids, anthocyanins, and phenolic compounds (Palmieri et al., 2017; Kim et al., 2024).

Particularly, the Aliança variety papaya has higher phenolic contents in peels (8.9 mg GAE/g) and seeds (4.7 mg GAE/g) than the Formosa variety (8.0 and 4.3 mg GAE/g for peels and seeds, respectively). Likewise, the flavonoid content is higher in the Aliança variety than in Formosa (2.3 and 2.5 mg CE/g respectively) (Vinha et al., 2024) (Table 1). Similar to papaya varieties, the bioactive composition of bagasse from different cashew varieties was evaluated, reporting differences in the compositions of carotenoids, tannins and phenolic compounds, which are attributed to their genetic differences (Cruz et al., 2022).

4.3. Maturity stage

The maturity stage of the fruit is an important factor when evaluating bioactive compounds (Ceccarelli et al., 2021; Khan et al., 2021). Fruit ripening is a complex process, which involves several physiological, biochemical and molecular mechanisms (Bhatla and Lal, 2023). As the fruit matures, the phenolic, anthocyanin and tannin content decreases, which may be associated with the biosynthesis processes and color changes that occur in the fruit (López et al., 2019; de Souza Silva et al., 2020; Gadelha et al., 2024).

During maturity, the synthesis of sugars, acids, aromatic compounds (Altaf et al., 2024) and the degradation of chlorophyll occur, and as a result of these biological reactions, free radicals are generated, which are counteracted by bioactive compounds, thus minimizing the content of the latter (Dzah, 2014; Martínez et al., 2017). Senescence in fruits affects their metabolic integrity, due to physiological disorders that eventually weaken antioxidant defense systems, making fruits vulnerable to oxidative stress induced by free radicals and depletion of antioxidant compounds (Zheng et al., 2016).

With maturation there is a greater probability of genetic errors induced by reactive oxygen species that can hinder the adequate synthesis of bioactive compounds. Also, with the degradation of components of the cellular structure such as pectin, other compartmentalized biomolecules and enzymes are exposed to the internal cellular environment, which inhibits the specialized biosynthesis of antioxidant compounds and enzymes, causing a reduction in the levels of phenolic compounds. Furthermore, as maturity-related loss of cellular integrity occurs, plant parts experience a loss of water and nutrients that are crucial for biosynthetic mechanisms. This phenomenon also negatively affects the biosynthesis of bioactive compounds of interest (Idris et al., 2023; Dzah et al., 2024).

On the other hand, carotenoids increase with fruit ripening, as color changes occur during this process. These changes typically begin with the degradation of chlorophyll, which results in the appearance of several darker pigments, accompanied by an increase in these bioactive compounds (Ma et al., 2023).

The extraction procedure varies according to the type of by-product; however, for the same by-product at different stages of maturation, the same extraction process is used. Therefore, according to Table 1, the phenolic content in purple passion fruit seeds decreases with the ripening of the fruit, where the decrease is 2.89 mg GAE/100g of sample (da Costa et al., 2023), in noni seeds the content decreased by 1.68 mg GAE/g of sample (Dzah et al., 2024) and in Hass avocado peel is 157.76 mg GAE/100g of sample (Alkaltham et al., 2021).

In addition to the aforementioned intrinsic factors, it is important to note that bioactive compounds are susceptible to various degradation mechanisms during storage and processing, which can affect their stability and functionality. Exposure to oxygen and high temperatures during processing can reduce the efficacy of these compounds, generating undesirable degradation products. Due to their high reactivity, compounds such as ascorbic acid, phenols and flavonoids undergo changes in their structure when reacting with oxygen. Likewise, elevated temperatures can induce thermal degradation of sensitive compounds, such as anthocyanins and some polyphenols, affecting their stability and biological activity (Elgamal et al., 2023; Mieszczakowska et al., 2021; Song et al., 2022).

Besides, variations in pH can destabilize bioactive compounds, leading to degradation or loss of functionality and bioactivity. Another key factor is exposure to light, especially ultraviolet radiation, which can trigger photochemical reactions that alter the structure and properties of the compounds, affecting their stability, as occurs with phenolic compounds in the presence of oxygen and light. On the other hand, high humidity levels can favor hydrolytic reactions and microbial growth, compromising the integrity and quality of these compounds. Increased water activity can accelerate their deterioration through oxidation and affect the dynamics of enzymatic reactions (Sepúlveda and Zapata, 2019; de Benedictis et al., 2023; García et al., 2025; Wang et al., 2025).

In order to improve the stability of bioactive compounds, which is affected by each of the above factors, microencapsulation (particles 1–1000 μm) and nanoencapsulation (particles <1000 nm) techniques have been applied (Hasan et al., 2024; Noore et al., 2021; Zabot et al., 2022). These techniques provide advantages such as protection against environmental degradation, controlled release and ease of handling in solid or liquid products. Nanoencapsulation offers advantages over microencapsulation in terms of increased solubility, improved absorption and targeted release in biological systems, especially for polyphenols and carotenoids (Chowdhury et al., 2024; Hasan et al., 2024; Matra et al., 2024). However, its disadvantages include higher production costs and greater complexity in industrial scale-up (Martins et al., 2022; Phupaboon et al., 2024). Despite their differences, both techniques represent promising strategies to improve the efficacy and stability of bioactive compounds, broadening their application in different sectors.

5. Conclusions and future perspectives

Tropical fruit by-products have a high content of bioactive compounds that give them a high nutritional value. However, they are often underused in the food field, so it is essential to investigate their application in this sector, to generate and develop new food products with excellent nutritional quality that contribute to improving and maintaining health.

The use of tropical fruit by-products contributes to improving the circular economy by using low-cost waste, thereby revaluing these by-products. Therefore, it is recommended to develop effective strategies to maximize their use and conduct a life cycle analysis to assess the environmental impact and economic benefits of the derived products.

The type of fruit by-product, the variety and the state of maturity are influential factors in the composition of bioactive compounds. These can be extracted by different methods, with green technologies being the

ones used in the long term and on a large scale, due to their great environmental benefits, balancing their high costs with the benefits they provide.

The valorization of tropical fruit by-products in the formulation of functional foods represents a key strategy to take advantage of their high content of bioactive compounds and respond to the growing demand for products with health benefits. Future research should focus on the development of technologies that allow the efficient incorporation of these compounds into foods without compromising their stability and bioavailability. To this end, it is essential to optimize extraction and purification methods, as well as to explore strategies such as microencapsulation and nanotechnology, which can improve the protection, release and absorption of bioactive compounds in the organism. In addition, it is essential to conduct studies on interactions between bioactive compounds and the food matrix, their stability during processing and storage, as well as their impact on the functionality of the final product. Scientific validation of their beneficial effects through clinical studies and bioavailability tests will strengthen their acceptance in the market and facilitate their integration into value-added products.

From a regulatory perspective, the use of by-products in functional foods must align with regulatory frameworks and food safety policies, ensuring compliance with the standards established by bodies such as the European Food Safety Authority (EFSA), the U.S. Food and Drug Administration (FDA) and the Food Code (Codex Alimentarius). It is necessary to promote the development of specific regulations that facilitate the approval and marketing of these products, guaranteeing their safety and functionality without generating excessive regulatory barriers.

In conclusion, the future of the valorization of by-products in functional foods depends on the combination of scientific advances in extraction, formulation and stability, together with the evolution of regulatory frameworks that facilitate their commercialization and guarantee their safety and efficacy.

CRediT authorship contribution statement

María Romero-Martínez: Conceptualization, Methodology, Writing – original draft, Visualization, Writing – review & editing. Ricardo Andrade-Pizarro: Methodology, Writing – review & editing, Visualization. Claudia De Paula: Writing – review & editing, Visualization, All authors have read and agree with the published version of the manuscript.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Data availability

Data will be made available on request.

References

Abderrezag, N., Montenegro, Z.J.S., Louaer, O., Meniai, A.-H., Cifuentes, A., Ibáñez, E., Mendiola, J.A., 2022. One-step sustainable extraction of Silymarin compounds of wild Algerian milk thistle (Silybum marianum) seeds using gas expanded Liquids. J. Chromatogr. A 1675, 463147. https://doi.org/10.1016/j.chroma.2022.463147.

- Ahmed, A., Hameed, A., Saeed, S., 2020. Biochemical profile and bioactive potential of thirteen wild folk medicinal plants from Balochistan, Pakistan. PLoS One 15 (8), e0231612. https://doi.org/10.1371/journal.pone.0231612.
- Ajikumar, P., Tyo, K., Carlsen, S., Mucha, O., Phon, T.H., Stephanopoulos, G., 2008. Terpenoids: opportunities for biosynthesis of natural product drugs using engineered microorganisms. Mol. Pharm. 5 (2), 167–190. https://doi.org/10.1021/mp700151b.
- Albuquerque, B., Pinela, J., Barros, L., Oliveira, M., Ferreira, I., 2020. Anthocyanin-rich extract of jabuticaba epicarp as a natural colorant: Optimization of heat- and ultrasound-assisted extractions and application in a bakery product. Food Chem 316, 126364. https://doi.org/10.1016/j.foodchem.2020.126364.
- Alkaltham, M., Uslu, N., Özcan, M., Salamatullah, A., Mohamed Ahmed, I., Hayat, K., 2021. Effect of drying process on oil, phenolic composition and antioxidant activity of avocado (cv. Hass) fruits harvested at two different maturity stages. Lebensm. Wiss. Technol. 148, 111716. https://doi.org/10.1016/j.lwt.2021.111716.
- Altaf, L., Wani, S.A., Hussain, P.R., Suradkar, P., Baqual, M.F., Bhat, A.A., 2024. Bioactive compounds and antioxidant activity in various parts of *Morus alba* L. Cv. ichinose: a comparative analysis. Discover Life 54 (1), 7. https://doi.org/10.1007/ s11084.024.09550.9
- Arifan, F., Broto, R., Sumardiono, S., Sutaryo, S., Dewi, A., Yudanto, Y., Sapatra, E., 2022. Effect of thermal pretreatment of pineapple peel waste in biogas production using response surface methodology. International Journal of Technology 13 (3), 619. https://doi.org/10.14716/ijtech.v13i3.4747.
- Ayala, J.F., Vega, V., Rosas, C., Palafox, H., Villa, J.A., Siddiqui, Md, Dávila, J.E., González, G.A., 2011. Agro-industrial potential of exotic fruit byproducts as a source of food additives. Food Res. Int. 44 (7), 1866–1874. https://doi.org/10.1016/j. foodres 2011.02.021
- Ashraf, M.A., Iqbal, M., Rasheed, R., Hussain, I., Riaz, M., Arif, M.S., 2018. Environmental stress and secondary metabolites in plants. In: Plant Metabolites and Regulation Under Environmental Stress. Elsevier, pp. 153–167. https://doi.org/10.1016/B978-0-12-812689-9.00008-X.
- Ayala, J., Vega, V., Rosas, C., Palafox, H., Villa, J., Siddiqui, Md, Dávil, J., González, G., 2011. Agro-industrial potential of exotic fruit byproducts as a source of food additives. Food Res. Int. 44 (7), 1866–1874. https://doi.org/10.1016/j. foodres.2011.02.021.
- Azizan, A., Lee, A., Abdul, N.A., Maulidiani, M., Mediani, A., Abdul, S., Zolkeflee, N., Abas, F., 2020. Potentially bioactive metabolites from pineapple waste extracts and their antioxidant and α -Glucosidase inhibitory activities by 1H NMR. Foods 9 (2), 173. https://doi.org/10.3390/foods9020173.
- Banerjee, J., Singh, R., Vijayaraghavan, R., MacFarlane, D., Patti, A.F., Arora, A., 2017. Bioactives from fruit processing wastes: green approaches to valuable chemicals. Food Chem. 225, 10–22. https://doi.org/10.1016/j.foodchem.2016.12.093.
- Barros, E., Carvalho, V., Rodrigues, H., Ponte, M., Gonçalves, L., 2017. Comparison of strategies for the simultaneous saccharification and fermentation of cashew apple bagasse using a thermotolerant *Kluyveromyces marxianus* to enhance cellulosic ethanol production. Chem. Eng. J. 307, 939–947. https://doi.org/10.1016/j. cei.2016.09.006.
- Belmonte, B., Domínguez, J., Wall, A., Ayala, J.F., Preciado, A., Salazar, N., López, L., Yahia, E., Robles, R., González, G., 2022. Lesser-consumed tropical fruits and their by-products: phytochemical content and their antioxidant and anti-inflammatory potential. Nutrients 14 (17), 3663. https://doi.org/10.3390/nu14173663.
- Beltrán, B., Estévez, R., Olmedilla, B., 2015. Assessment of dietary vitamin A intake (retinol, α-carotene, β-carotene, β-cryptoxanthin) and its sources in the National Survey of Dietary Intake in Spain (2009-2010). Int. J. Food Sci. Nutr. 66 (6), 706–712. https://doi.org/10.3109/09637486.2015.1077787.
- Bhatla, S.C., Lal, M.A., 2023. Fruit development and ripening. In: Plant Physiology, Development and Metabolism. Springer Nature, Singapore, pp. 607–624. https://doi.org/10.1007/978-981-99-5736-1-27.
- Briones, V., Plaza, M., Giovagnoli, C., Jamett, F., 2015. High hydrostatic pressure and ultrasound extractions of antioxidant compounds, sulforaphane and fatty acids from Chilean papaya (*Vasconcellea pubescens*) seeds: effects of extraction conditions and methods. Lebensm. Wiss. Technol. 60 (1), 525–534. https://doi.org/10.1016/j. lwt.2014.07.057.
- Cádiz, M., Villegas, M. del C., Leyva, F.J., Pimentel, S., Fernández, Á., Alañón, M.E., Segura, A., 2020. Revalorization of bioactive compounds from tropical fruit byproducts and industrial applications by means of sustainable approaches. Food Res. Int. 138, 109786. https://doi.org/10.1016/j.foodres.2020.109786.
- Cangussu, L.B., Fronza, P., Franca, A.S., Oliveira, L.S., 2021. Chemical characterization and bioaccessibility assessment of bioactive compounds from umbu (Spondias tuberosa A.) Fruit peel and pulp flours. Foods 10 (11), 2597. https://doi.org/ 10.3390/foods10112597.
- Cangussu, L., Costa, A., Franca, A., de Oliveira, L., 2024. Chemical characterization and the bioaccessibility of bioactive compounds from seriguela (*Spondias purpurea L.*) pulp and by-products flours. Bioactive Carbohydrates and Dietary Fibre 31, 100404. https://doi.org/10.1016/j.bcdf.2024.100404.
- Cao, J., Wang, C., Shi, L., Cheng, Y., Hu, H., Zeng, B., Zhao, F., 2022. Water based-deep eutectic solvent for ultrasound-assisted liquid-liquid microextraction of parabens in edible oil. Food Chem. 383, 132586. https://doi.org/10.1016/j. foodchem.2022.132586
- Cao, Y., Wang, H., Jian, Y., Hao, G., Di, D., Liu, J., 2024. Temperature-switchable deep eutectic solvent extraction of polysaccharides from wolfberry fruits: process optimization, structure characterization, and antioxidant activity. J. Mol. Liq. 398, 124352. https://doi.org/10.1016/j.molliq.2024.124352.
- Carvalho, N., Santos, C., Pedreira, J., Silva, M., Caroline, H., Rajan, M., Terezinha Santos Leite Neta, M., Narain, N., 2021. Bioactive compounds and antioxidant activities in the agro-industrial residues of acerola (Malpighia emarginata L.), guava (Psidium guajava L.), genipap (Genipa americana L.) and umbu (Spondias tuberosa L.) fruits

- assisted by ultrasonic or shaker extraction. Food Res. Int. 147, 110538. https://doi.org/10.1016/j.foodres.2021.110538.
- Ceccarelli, D., Antonucci, F., Talento, C., Ciccoritti, R., 2021. Chemical characterization in the selection of Italian autochthonous genotypes of plum. Sci. Hortic. 281, 109922. https://doi.org/10.1016/j.scienta.2021.109922.
- Celebioglu, A., Uyar, T., 2020. Development of ferulic acid/cyclodextrin inclusion complex nanofibers for fast-dissolving drug delivery system. Int. J. Pharm. 584, 119395. https://doi.org/10.1016/j.ijpharm.2020.119395.
- Chamira, F., Preethi, S., 2014. Total phenolic, flavonoid contents, in-vitro antioxidant activities and hepatoprotective effect of aqueous leaf extract of Atalantia ceylanica. BMC Compl. Alternative Med. 14 (1), 395. https://doi.org/10.1186/1472-6882-14-395
- Chaves, E.C., Esquivel, P., 2019. Ácidos clorogénicos presentes en el café: capacidad antimicrobiana y antioxidante. Agron. Mesoam. 299–311. https://doi.org/ 10.15517/am.y30i1.32974
- Chel, L.D., Cuevas, L.F., Sauri, E., Sierra, E., de León, F.D., Mendoza, J.A., 2022. Tropical fruit peels as sources of bioactive compounds: a review. Pakistan J. Bot. 54 (3). https://doi.org/10.30848/PJB2022-3, 7.
- Chowdhury, S., Kar, K., Mazumder, R., 2024. Exploration of different strategies of nanoencapsulation of bioactive compounds and their ensuing approaches. Fut. J. Pharma. Sci 10 (1), 72. https://doi.org/10.1186/s43094-024-00644-y.
- Clemente, R., Pardo, T., Madejón, P., Madejón, E., Bernal, M., 2015. Food byproducts as amendments in trace elements contaminated soils. Food Res. Int. 73, 176–189. https://doi.org/10.1016/j.foodres.2015.03.040.
- Coelho, T., Silva, D., dos Santos, J., Dantas, C., Nogueira, A., Lopes, C., Vieira, E., 2022. Multivariate optimization and comparison between conventional extraction (CE) and ultrasonic-assisted extraction (UAE) of carotenoid extraction from cashew apple. Ultrason. Sonochem. 84, 105980. https://doi.org/10.1016/j.ultsonch.2022.105980.
- Corcia, S., Dornier, M., Palmade, L., Dhuique-Mayer, C., 2022. Enhancement of the in vitro bioavailable carotenoid content of a citrus juice combining crossflow microfiltration and high-pressure treatments. Food Res. Int. 156, 111134. https:// doi.org/10.1016/j.foodres.2022.111134.
- Cruz, L., Durán, D., Forero, L., Tarapuez, L., Durán, D., Carazzone, C., Sierra, R., 2022. Chemical composition and bioactive compounds of cashew (*Anacardium occidentale*) apple juice and bagasse from Colombian varieties. Heliyon 8 (5), e09528. https://doi.org/10.1016/j.heliyon.2022.e09528.
- da Costa, C., Machado, G., Rodrigues, L., de Barros, H., Natarelli, C., Boas, E.V., 2023. Phenolic compounds profile and antioxidant activity of purple passion fruit's pulp, peel and seed at different maturation stages. Sci. Hortic. 321, 112244. https://doi.org/10.1016/j.scienta.2023.112244.
- de Benedictis, J.N., de Kok, T.M., van Breda, S.G., 2023. Impact of processing method and storage time on phytochemical concentrations in an antioxidant-rich food mixture. Antioxidants 12 (6), 1252. https://doi.org/10.3390/antiox12061252.
- de Souza Silva, G., da Silva Campelo Borges, G., Pinho da Costa Castro, C.D., de Tarso Aidar, S., Telles Biasoto Marques, A., Tonetto de Freitas, S., Poloni Rybka, A.C., Cardarelli, H.R., 2020. Physicochemical quality, bioactive compounds and in vitro antioxidant activity of a new variety of passion fruit cv. BRS Sertão Forte (*Passiflora cincinnata* Mast.) from Brazilian Semiarid region. Sci. Hortic. 272, 109595. https://doi.org/10.1016/j.scienta.2020.109595.
- Dheeraj, S., Mishra, A., 2023. Mitigation of cashew apple fruits astringency. Environ. Sustainability 6 (3), 319–329. https://doi.org/10.1007/s42398-023-00276-7.
- di Corcia, S., Dhuique-Mayer, C., Dornier, M., 2020. Concentrates from citrus juice obtained by crossflow microfiltration: Guidance of the process considering carotenoid bioaccessibility. Innovative Food Science & Emerging Technologies. https:// doi.org/10.1016/j.ifset.2020.102526
- Dias, M., Pinto, D., Silva, A., 2021. Plant flavonoids: chemical characteristics and biological activity. Molecules 26 (17), 5377. https://doi.org/10.3390/
- Dobrinčić, A., Repajić, M., Garofulić, I.E., Tuđen, L., Dragović, V., Levaj, B., 2020. Comparison of different extraction methods for the recovery of olive leaves polyphenols. Processes 8 (9), 1008. https://doi.org/10.3390/pr8091008.
- Domínguez-Bocanegra, A.R., Torres-Muñoz, J.A., López, R.A., 2015. Production of bioethanol from agro-industrial wastes. Fuel 149, 85–89. https://doi.org/10.1016/j. fuel.2014.09.062.
- Donadio, G., Mensitieri, F., Santoro, V., Parisi, V., Bellone, M.L., De Tommasi, N., Izzo, V., Dal Piaz, F., 2021. Interactions with microbial proteins driving the antibacterial activity of flavonoids. Pharmaceutics 13 (5), 660. https://doi.org/ 10.3390/pharmaceutics13050660.
- Durán-Aranguren, D.D., Muñoz-Daza, L.F., Castillo-Hurtado, L.J., Posada, J.A., Mussatto, S.I., Sierra, R., Hernández-Carrión, M., 2023. Design of a baked good using food ingredients recovered from agro-industrial by-products of fruits. Lebensm. Wiss. Technol. 185, 115174. https://doi.org/10.1016/j.lwt.2023.115174.
- Dzah, C., 2014. Influence of fruit maturity on antioxidant potential and chilling injury resistance of peach fruit (*Prunus persica*) during cold storage. Afr. J. Food Nutr. Sci. 14 (67), 9578–9591. https://doi.org/10.18697/ajfand.67.13600.
- Dzah, C., Kpodo, F., Asante-Donyinah, D., Boateng, N., 2024. The influence of Morinda citrifolia fruit maturity level, parts and storage length on total phenols, ascorbic acid, antioxidant activity and ethylene gas emission. Food Chemistry Advances 4, 100599. https://doi.org/10.1016/j.focha.2023.100599.
- Elgamal, R., Song, C., Rayan, A.M., Liu, C., Al-Rejaie, S., Elmasry, G., 2023. Thermal degradation of bioactive compounds during drying process of horticultural and agronomic products: a comprehensive overview. Agronomy 13 (6), 1580. https:// doi.org/10.3390/agronomy13061580.
- El-Hadary, A.E., Ramadan, M.F., 2019. Phenolic profiles, antihyperglycemic, antihyperlipidemic, and antioxidant properties of pomegranate (*Punica granatum*) peel extract. J. Food Biochem. 43 (4), e12803. https://doi.org/10.1111/jfbc.12803.

- Enríquez, M., Villafuerte, F., Figueroa, A., 2023. Efectos de los componentes bioactivos de frutas, vegetales, lácteos y plantas medicinales en la nutrición humana. Revista de Ciencias Agropecuarias ALLPA 6 (11), 2–24. https://doi.org/10.56124/allpa. v6i11.0055
- Fang, J., 2014. Bioavailability of anthocyanins. Drug Metab. Rev. 46 (4), 508–520. https://doi.org/10.3109/03602532.2014.978080.
- Faustino, M., Veiga, M., Sousa, P., Costa, E.M., Silva, S., Pintado, M., 2019. Agro-food byproducts as a new source of natural food additives. Molecules 24 (6), 1056. https://doi.org/10.3390/molecules24061056.
- Frešer, F., Hostnik, G., Tošović, J., Bren, U., 2021. Dependence of the Fe (II) gallic acid coordination compound formation constant on the pH. Foods 10 (11), 2689. https://doi.org/10.3390/foods10112689.
- Gadelha, T.M., do Nascimento, A.M., da Silva, T.I., da Silva, J.L., Sales, G.N.B., Ribeiro, W.S., de Queiroga, R.C.F., de Almeida, F.A., de Souza, P.A., Ribeiro, J.E. da S., Nóbrega, J.S., de Lima, G.S., Soares, L., da Costa, F.B., 2024. Comprehensive analysis of quality and bioactive compounds in umbu-cajá fruit during different ripening stages. Sci. Hortic. 327, 112886. https://doi.org/10.1016/j. scienta.2024.112886.
- García, D.F., Valdiviezo, J., Rodrigues, C., Viganó, J., Oliveira, A. L. de, 2025. Advances and trends in compressed fluid extraction to recovering bioactive compounds from avocado: effect of extraction parameters, biological properties, and applications. J. Supercrit. Fluids 218, 106505. https://doi.org/10.1016/j.supflu.2024.106505.
- Gegotek, A., Skrzydlewska, E., 2023. Ascorbic acid as antioxidant, 247–270. https://doi. org/10.1016/bs.vh.2022.10.008.
- Ghahremani, A., anji Moghaddam, E., Marjani, A., 2023. Growth, yield, and biochemical behaviors of important stone fruits affected by plant genotype and environmental conditions. Sci. Hortic. 321, 112211. https://doi.org/10.1016/j. scienta.2023.112211.
- Gil-Chávez, G., Villa, J., Ayala, J., Heredia, J., Sepulveda, D., Yahia, E.M., González, G., 2013. Technologies for extraction and production of bioactive compounds to be used as nutraceuticals and food ingredients: an overview. Compr. Rev. Food Sci. Food Saf 12 (1), 5–23, 10.1111/1541-4337.12005.
- Gómez, R., Campos, D.A., Oliveira, A., Aguilar, C., Madureira, A.R., Pintado, M., 2021. A chemical valorisation of melon peels towards functional food ingredients: bioactives profile and antioxidant properties. Food Chem. 335, 127579. https://doi. org/10.1016/j.foodchem.2020.127579.
- Gu, Y., Zhang, Y., Li, M., Huang, Z., Jiang, J., Chen, Y., Chen, J., Jia, Y., Zhang, L., Zhou, F., 2021. Ferulic acid ameliorates atherosclerotic injury by modulating gut microbiota and lipid metabolism. Front. Pharmacol. 12. https://doi.org/10.3389/fblar.2021.621339.
- Gull, A., Sheikh, M.A., Kour, J., Zehra, B., Zargar, I.A., Wani, A.A., Bhatia, S., Lone, M.A., 2022. Anthocyanins. In: In Nutraceuticals and Health Care. Elsevier, pp. 317–329. https://doi.org/10.1016/B978-0-323-89779-2.00018-1.
- Guo, F., Shahidi, F., 2024. Can anthocyanins replace antibiotics in food and animal feed? A review. Trends Food Sci. Technol. 143, 104219. https://doi.org/10.1016/j. tifs.2023.104219.
- Guo, X., Qiu, H., Deng, X., Mao, X., Guo, X., Xu, C., Zhang, J., 2019. Effect of chlorogenic acid on the physicochemical and functional properties of *Coregonus* peled myofibrillar protein through hydroxyl radical oxidation. Molecules 24 (17), 3205. https://doi.org/10.3390/molecules/24173/205
- Gutiérrez, I., Fernández, J., Lombó, F., 2018. Plant nutraceuticals as antimicrobial agents in food preservation: terpenoids, polyphenols and thiols. Int. J. Antimicrob. Agents 52 (3), 309–315. https://doi.org/10.1016/j.ijantimicag.2018.04.024.
- Hasan, Md, Islam, Md, Haque, A., Kabir, Md, Khushe, K., Hasan, S., 2024. Trends and challenges of fruit by-products utilization: insights into safety, sensory, and benefits of the use for the development of innovative healthy food: a review. Bioresources and Bioprocess 11 (1), 10. https://doi.org/10.1186/s40643-023-00722-8.
 Herrero, M., Mendiola, J.A., Ibáñez, E., 2017. Gas expanded liquids and switchable
- Herrero, M., Mendiola, J.A., Ibánez, E., 2017. Gas expanded líquids and switchable solvents. Curr. Opin. Green Sustainable Chem. 5, 24–30. https://doi.org/10.1016/j. cogsc.2017.03.008.
- How, Y.H., Nyam, K.L., 2024. Reutilization of fruit waste as potential prebiotic for probiotic or food-grade microorganisms in food applications: a review. Probiotics and Antimicrobial Proteins. https://doi.org/10.1007/s12602-024-10375-4.
- Idris, N.S., Khandaker, M.M., Rashid, Z.M., Majrashi, A., Alenazi, M.M., Nor, Z.M., Mohd Adnan, A.F., Mat, N., 2023. Polyphenolic compounds and biological activities of leaves and fruits of Syzygium samarangense cv. 'Giant Green' at three different maturities. Horticulturae 9 (3), 326. https://doi.org/10.3390/ horticulturae9030326.
- Iervese, F., Flamminii, F., D'Alessio, G., Neri, L., De Bruno, A., Imeneo, V., Valbonetti, L., Di Mattia, C.D., 2024. Flavonoid- and limonoid-rich extracts from lemon pomace byproducts: technological properties for the formulation of o/w emulsions. Food Biosci. 59, 104030. https://doi.org/10.1016/j.fbio.2024.104030.
- Jayarathne, P.S.I.U., Dharmadasa, R.M., Abeysinghe, D.C., Weerawarna, S.W.A., 2024. Changes of physicochemical parameters, bioactive compounds and antioxidant capacity of four different cultivars of *Punica granatum* (L.) at four distinct maturity stages. Food Chemistry Advances 4, 100640. https://doi.org/10.1016/j. focha.2024.100640.
- Jeon, Y.A., Chung, S.W., Kim, S.C., Lee, Y.J., 2022. Comprehensive assessment of antioxidant and anti-inflammatory properties of papaya extracts. Foods 11 (20), 3211. https://doi.org/10.3390/foods11203211.
- Kaderides, K., Papaoikonomou, L., Serafim, M., Goula, A.M., 2019. Microwave-assisted extraction of phenolics from pomegranate peels: optimization, kinetics, and comparison with ultrasounds extraction. Chemical Engineering and Processing -Process Intensification 137, 1–11. https://doi.org/10.1016/j.cep.2019.01.006.
- Kashyap, A.K., Dubey, S.K., Shah, S., Kumar, A., 2022. A short review on genes regulating biosynthesis of major secondary metabolites. In: Phytochemical

- Genomics. Springer Nature, Singapore, pp. 501–519. https://doi.org/10.1007/978-981-19-5779-6-20
- Khan, A., Zahiruddin, S., Ibrahim, M., Basist, P., Gaurav, Parveen, R., Umar, S., Ahmad, S., 2021. Thin layer chromatography-mass spectrometry bioautographic identification of free radical scavenging compounds and metabolomic profile of Carica papaya linn. fruit and seeds using high-performance thin-layer chromatography, gas chromatography-mass spectrometry and ultra-performance liquid chromatography-mass spectrometry. Pharmacogn. Mag. 17 (5), 21. https://doi.org/10.4103/pm.pm 326 20.
- Khan, S., Das, P., Abdulquadir, M., Thaher, M., Al-Ghasal, G., Hafez Mohammed Kashem, A., Faisal, M., Sayadi, S., Al-Jabri, H., 2023. Pilot-scale crossflow ultrafiltration of four different cell-sized marine microalgae to assess the ultrafiltration performance and energy requirements. Separ. Purif. Technol. 315, 123681. https://doi.org/10.1016/j.seppur.2023.123681.
- Khan, R., Anwar, F., Ghazali, F.M., Mahyudin, N.A., 2024. Valorization of waste: innovative techniques for extracting bioactive compounds from fruit and vegetable peels - a comprehensive review. Innov. Food Sci. Emerg. Technol. 97, 103828. https://doi.org/10.1016/j.ifset.2024.103828.
- Kim, I.J., Park, S., Kyoung, H., Song, M., Kim, S.R., 2024. Microbial valorization of fruit processing waste: opportunities, challenges, and strategies. Curr. Opin. Food Sci. 56, 101147. https://doi.org/10.1016/j.cofs.2024.101147.
- Kumar, N., Goel, N., 2019. Phenolic acids: natural versatile molecules with promising therapeutic applications. Biotechnology Reports 24, e00370. https://doi.org/ 10.1016/j.btre.2019.e00370.
- Kumar, K., Yadav, A.N., Kumar, V., Vyas, P., Dhaliwal, H.S., 2017. Food waste: a potential bioresource for extraction of nutraceuticals and bioactive compounds. Bioresources and Bioprocessing 4 (1), 18. https://doi.org/10.1186/s40643-017-0148-6.
- Kupnik, K., Leitgeb, M., Primožić, M., Postružnik, V., Kotnik, P., Kučuk, N., Knez, Ž., Marevci, M.K., 2022. Supercritical fluid and conventional extractions of high valueadded compounds from pomegranate peels waste: production, quantification and antimicrobial activity of bioactive constituents. Plants 11 (7), 928. https://doi.org/ 10.3390/plants11070928.
- Lachowicz, S., Oszmiański, J., Wiśniewski, R., Seliga, Ł., Pluta, S., 2019. Chemical parameters profile analysis by liquid chromatography and antioxidative activity of the Saskatoon berry fruits and their components. Eur. Food Res. Technol. 245 (9), 2007–2015. https://doi.org/10.1007/s00217-019-03311-2.
- Lolletti, D., Principio, L., Ciorba, R., Mitrano, F., Ceccarelli, D., Antonucci, F., Manganiello, R., Ciccoritti, R., 2021. Asimina triloba: crop years, cultivars and ripening time influence on qualitative parameters. Sci. Hortic. 289, 110481. https:// doi.org/10.1016/j.scienta.2021.110481.
- Lopes, G., Sousa, C., Silva, L.R., Pinto, E., Andrade, P.B., Bernardo, J., Mouga, T., Valentão, P., 2012. Can phlorotannins purified extracts constitute a novel pharmacological alternative for microbial infections with associated inflammatory conditions? PLoS One 7 (2), e31145. https://doi.org/10.1371/journal. pone.0031145.
- López, C., Aguirre, C., Ramírez, J., Raya, J., Santiago, Y., Gutiérrez, J., Hernández, A., 2019. Compuestos bioactivos y actividad antioxidante en tres estados de madurez de Myrtillocactus geometrizans provenientes del Valle del Mezquital, Hidalgo. Investigación y Desarrollo En Ciencia y Tecnología de Alimentos 4, 317–322.
- López, L., Enríquez, S., González, G., 2021. Tropical fruits and by-products as a potential source of bioactive polysaccharides. Biotec 23 (3). https://doi.org/10.18633/ biotecnia.v23i3.1450.
- Luan, Y., Wang, S., Wang, P., Ke, F., Zhu, C., Xu, C., 2024. Accumulation of β-citraurin and other carotenoids in relation to the expression of PSY1 and CCD4b1 in peel of twelve red-peeled citrus cultivars. Sci. Hortic. 330, 113094. https://doi.org/10.1016/j.scienta.2024.113094.
- Luan, A., Zhang, W., Yang, M., Zhong, Z., Wu, J., He, Y., He, J. 2023. Unveiling the molecular mechanism involving anthocyanins in pineapple peel discoloration during fruit maturation. Food Chem. 412, 135482. https://doi.org/10.1016/j.foodchem.20 23.135482.
- Ma, G., Zhang, L., Matsuta, A., Matsutani, K., Yamawaki, K., Yahata, M., Wahyudi, A., Motohashi, R., Kato, M., 2013. Enzymatic formation of β-Citraurin from β-Cryptoxanthin and zeaxanthin by carotenoid cleavage dioxygenase4 in the flavedo of citrus fruit. Plant Physiol. 163 (2), 682–695. https://doi.org/10.1104/pp.113.293907
- Ma, B., Zhao, X., Dai, B., Zhao, M., Yang, X., Cui, L., Shao, H., 2023. Changes in phytochemical compositions and antioxidant activity of *Schisandra sphenathera* fruit during ripening. Ind. Crop. Prod. 199, 116773. https://doi.org/10.1016/j. indcrop.2023.116773.
- Mandal, S.M., Chakraborty, D., Dey, S., 2010. Phenolic acids act as signaling molecules in plant-microbe symbioses. Plant Signal. Behav. 5 (4), 359–368. https://doi.org/ 10.4161/psb.5.4.10871.
- Maqsood, S., Adiamo, O., Ahmad, M., Mudgil, P., 2020. Bioactive compounds from date fruit and seed as potential nutraceutical and functional food ingredients. Food Chem. 308, 125522. https://doi.org/10.1016/j.foodchem.2019.125522.
- Marcillo, V., Anaguano, M., Molina, M., Tupuna, D., Ruales, J., 2021a. Characterization and quantification of bioactive compounds and antioxidant activity in three different varieties of mango (*Mangifera indica* L.) peel from the Ecuadorian region using HPLC-UV/VIS and UPLC-PDA. NFS Journal 23, 1–7. https://doi.org/10.1016/j. nfs.2021.02.001.
- Marcillo, V., Tupuna, D., González, Z., Ruales, J., 2021b. Encapsulation of bioactive compounds from fruit and vegetable by-products for food application – a review. Trends Food Sci. Technol. 116, 11–23. https://doi.org/10.1016/j.tifs.2021.07.009.

- Martínez, M.E., Balois, R., Alia, I., Cortes, M.A., Palomino, Y.A., López, G.G., 2017.
 Postcosecha de frutos: maduración y cambios bioquímicos. Revista Mexicana de Ciencias Agrícolas 19, 4075–4087. https://doi.org/10.29312/remexca.v0i19.674.
- Martins, V., Pintado, M., Morais, R., Morais, A., 2022. Valorisation of micro/ nanoencapsulated bioactive compounds from plant sources for food applications towards sustainability. Foods 12 (1), 32. https://doi.org/10.3390/foods12010032.
- Matra, M., Phupaboon, S., Totakul, P., Prommachart, R., Shah, A.A., Shah, A.M., Wanapat, M., 2024. Microencapsulation of Mitragyna leaf extracts to be used as a bioactive compound source to enhance in vitro fermentation characteristics and microbial dynamics. Animal Bioscience 37 (1), 74–83. https://doi.org/10.5713/ ab.23.0200.
- Meléndez, A.J., 2019. An overview of carotenoids, apocarotenoids, and vitamin A in agro-food, nutrition, health, and disease. Mol. Nutr. Food Res. 63 (15). https://doi. org/10.1002/mnfr.201801045.
- Meléndez, A.J., Esquivel, P., Rodriguez, D., 2023. Comprehensive review on carotenoid composition: transformations during processing and storage of foods. Food Res. Int. 169, 112773. https://doi.org/10.1016/j.foodres.2023.112773.
- Mieszczakowska, M., Celejewska, K., Płocharski, W., 2021. Impact of i nnovative technologies on the content of vitamin c and its bioavailability from processed fruit and vegetable products. Antioxidants 10 (1), 54. https://doi.org/10.3390/ artioxidants/10010054
- Muhie, S.H., 2023. Concepts, principles, and application of biodynamic farming: a review. Circular Economy and Sustainability 3 (1), 291–304. https://doi.org/ 10.1007/s43615-022-00184-8.
- Nikiema, M., Somda, M.K., Sawadogo, J.B., Dianou, D., Traoré, A.S., Ouattara, A.S., 2020. Valorization of agricultural waste: theoretical estimation and experimental biomethane yield from cashew nut hulls. J. Sustain. Bioenergy Syst. 10 (4), 113–130. https://doi.org/10.4236/jsbs.2020.104009.
- Noore, S., Rastogi, N.K., O'Donnell, C., Tiwari, B., 2021. Novel bioactive extraction and nano-encapsulation. Encyclopedia 1 (3), 632–664. https://doi.org/10.3390/ encyclopedia1030052.
- Oliveira, F., Arruda, T., Morzelle, M., Pereira, A., Casarotti, S., 2022. Fruit by-products as potential prebiotics and promising functional ingredients to produce fermented milk. Food Res. Int. 161, 111841. https://doi.org/10.1016/j.foodres.2022.111841.
- O'Shea, N., Arendt, E.K., Gallagher, E., 2012. Dietary fibre and phytochemical characteristics of fruit and vegetable by-products and their recent applications as novel ingredients in food products. Innov. Food Sci. Emerg. Technol. 16, 1–10. https://doi.org/10.1016/j.ifset.2012.06.002.
- Palmieri, L., Masuero, D., Martinatti, P., Baratto, G., Martens, S., Vrhovsek, U., 2017. Genotype-by-environment effect on bioactive compounds in strawberry (*Fragaria x ananassa* Duch.). J. Sci. Food Agric. 97 (12), 4180–4189. https://doi.org/10.1002/isfa.8290.
- Pavlović, N., Jokić, S., Jakovljević, M., Blažić, M., Molnar, M., 2020. Green extraction methods for active compounds from food waste—cocoa bean shell. Foods 9 (2), 140. https://doi.org/10.3390/foods9020140.
- Pereira, A., 2018. Tropical fruits as natural, exceptionally rich, sources of bioactive compounds. Int. J. Fruit Sci. 18 (3), 231–242. https://doi.org/10.1080/ 15538362.2018.1444532.
- Pereira, D., Valentão, P., Pereira, J., Andrade, P., 2009. Phenolics: from chemistry to biology. Molecules 14 (6), 2202–2211. https://doi.org/10.3390/ ppaleules/14062302
- Peron, G., Bernabé, G., Marcheluzzo, S., Zengin, G., Ibrahime Sinan, K., Hošek, J., Treml, J., Kaja, I., Paccagnella, M., Brun, P., Castagliuolo, I., Zancato, M., Dall'Acqua, S., 2024. Orange fruit peels from PDO varieties of Ribera (Sicily, Italy): an insight into the chemistry and bioactivity of volatile and non-volatile secondary metabolites extracted using a microwave-assisted method. J. Funct.Foods 116, 106147. https://doi.org/10.1016/i.iff.2024.106147.
- 106147. https://doi.org/10.1016/j.jff.2024.106147.
 Peschel, W., Sánchez, F., Diekmann, W., Plescher, A., Gartzía, I., Jiménez, D., Lamuela, R., Buxaderas, S., Codina, C., 2006. An industrial approach in the search of natural antioxidants from vegetable and fruit wastes. Food Chem. 97 (1), 137–150. https://doi.org/10.1016/j.foodchem.2005.03.033.
- Petchidurai, G., Nagoth, J.A., John, M.S., Sahayaraj, K., Murugesan, N., Pucciarelli, S., 2019. Standardization and quantification of total tannins, condensed tannin and soluble phlorotannins extracted from thirty-two drifted coastal macroalgae using high performance liquid chromatography. Bioresour. Technol. Rep. 7, 100273. https://doi.org/10.1016/j.biteb.2019.100273.
- Phupaboon, S., Matra, M., Sommai, S., Dagaew, G., Suriyapha, C., Prachumchai, R., Wanapat, M., 2024. Microencapsulation efficiency of fruit peel phytonutrient-based antimicrobial to mitigate rumen emission using in vitro fermentation technique. Ital. J. Anim. Sci. 23 (1), 664–677. https://doi.org/10.1080/1828051X.2024.2337688. Pirozzi, A., Imbimbo, P., D'Agostino, A., Tirino, V., Finamore, R., Monti, D.M.,
- Pirozzi, A., Imbilindo, P., D Agostino, A., Tirino, V., Finantore, R., Moliti, D.M., Piccoli, R., Schiraldi, C., 2020. Antioxidant and hypolipidemic activity of Açai fruit makes it a valuable functional food. Antioxidants 10 (1), 40. https://doi.org/ 10.3390/antiox10010040.
- Płotka-Wasylka, J., Rutkowska, M., de la Guardia, M., 2021. Are deep eutectic solvents useful in chromatography? A short review. J. Chromatogr. A 1639, 461918. https://doi.org/10.1016/j.chroma.2021.461918.
- Poletto, P., Álvarez-Rivera, G., López, G.-D., Borges, O.M.A., Mendiola, J.A., Ibáñez, E., Cifuentes, A., 2021. Recovery of ascorbic acid, phenolic compounds and carotenoids from acerola by-products: an opportunity for their valorization. Lebensm. Wiss. Technol. 146, 111654. https://doi.org/10.1016/j.lwt.2021.111654.
- Prietsch, R.F., Monte, L.G., da Silva, F.A., Beira, F.T., Del Pino, F.A.B., Campos, V.F., Collares, T., Pinto, L.S., Spanevello, R.M., Gamaro, G.D., Braganhol, E., 2014. Genistein induces apoptosis and autophagy in human breast MCF-7 cells by modulating the expression of proapoptotic factors and oxidative stress enzymes. Mol. Cell. Biochem. 390 (1–2), 235–242. https://doi.org/10.1007/s11010-014-1974-x.

- Qiu, L., Zhang, M., Chitrakar, B., Bhandari, B., 2020. Application of power ultrasound in freezing and thawing Processes: effect on process efficiency and product quality. Ultrason. Sonochem. 68, 105230. https://doi.org/10.1016/j.ultsonch.2020.105230.
- Ramírez, M.C., Quiles, J., 2005. Tratado de nutrición. Tomo 1: Vitamina C, Vitamina E y otros antioxidantes de origen alimentario" 1 (Editorial Acción Médica, Ed.; 1st ed.
- Resende, L., Franca, A., Oliveira, L., 2019. Buriti (Mauritia flexuosa L. f.) fruit by-products flours: evaluation as source of dietary fibers and natural antioxidants. Food Chem. 270, 53–60. https://doi.org/10.1016/j.foodchem.2018.07.079.
- Rezende, Y., Nogueira, J., Narain, N., 2018. Microencapsulation of extracts of bioactive compounds obtained from acerola (*Malpighia emarginata* DC) pulp and residue by spray and freeze drying: chemical, morphological and chemometric characterization. Food Chem. 254, 281–291. https://doi.org/10.1016/j.foodchem.2018.02.026.
- Rodríguez, S., Ramírez, R., Serna, S., 2020. Environmentally friendly methods for flavonoid extraction from plant material: impact of their operating conditions on yield and antioxidant properties. Sci. World J. 1–38. https://doi.org/10.1155/2020/ 6792069, 2020.
- Rojas, A., Fuentes, E., Cádiz-Gurrea, M. de la L., Rodriguez, L., Villegas-Aguilar, M. del C., Palomo, I., Arráez-Román, D., Segura-Carretero, A., 2022. Biological evaluation of avocado residues as a potential source of bioactive compounds. Antioxidants 11 (6), 1049. https://doi.org/10.3390/antiox11061049.
- Romero, H., Vega, C., Feijoó, V., Villacreses, D., Sarmiento, C., 2020. Methane production through anaerobic co-digestion of tropical fruit biomass and urban solid waste. Energy Rep. 6, 351–357. https://doi.org/10.1016/j.egyr.2020.11.170.
- Rosa, A., Petretto, G.L., Maldini, M., Tirillini, B., Chessa, M., Pintore, G., Sarais, G., 2023. Chemical characterization, antioxidant and cytotoxic activity of hydroalcoholic extract from the albedo and flavedo of *Citrus limon* var. pompia Camarda. J. Food Meas. Char. 17 (1), 627–635. https://doi.org/10.1007/s11694-022-01659-w.
- Routray, W., Orsat, V., 2019. Agricultural and food industry By-products: source of bioactive components for functional beverages. In: Nutrients in Beverages. Elsevier, pp. 543–589. https://doi.org/10.1016/B978-0-12-816842-4.00015-0.
- Ruiz, S., Chaparro Saraí, Ruiz, K., Cira, L.A., Estrada, M.I., Ortega, L.E., Ornelas, J. de J., Mata, M., 2017. Flavonoids: Important biocompounds in food. In: Flavonoids - From biosynthesis to human health. InTech. https://doi.org/10.5772/67864.
- Sahie, L.B.C., Soro, D., Kone, K.Y., Assidjo, N.E., Yao, K.B., 2023. Some processing steps and uses of cashew apples: a review. Food Nutr. Sci. 14 (1), 38–57. https://doi.org/ 10.4236/fns.2023.141004.
- Şahin, S., İlbay, Z., Kırbaşlar, Ş.İ., 2018. Pulsed ultrasound-assisted extraction of natural antioxidants from Mandarin (*Citrus deliciosa* Tenore) leaves: experimental and modeling study. Chem. Eng. Commun. 205 (6), 717–726. https://doi.org/10.1080/ 00986445.2017.1328414.
- Samtiya, M., Acharya, S., Pandey, K.K., Aluko, R.E., Udenigwe, C.C., Dhewa, T., 2021. Production, purification, and potential health applications of edible seeds' bioactive peptides: a concise review. Foods 10 (11), 2696. https://doi.org/10.3390/ foods10112696.
- Santos, C., González, A.M., 2019. Anthocyanins. In: Encyclopedia of Food Chemistry. Elsevier, pp. 10–21. https://doi.org/10.1016/B978-0-08-100596-5.21609-0.
- Sayago-Ayerdi, S., García-Martínez, D.L., Ramírez-Castillo, A.C., Ramírez-Concepción, H. R., Viuda-Martos, M., 2021. Tropical fruits and their co-products as bioactive compounds and their health effects: a review. Foods 10 (8). https://doi.org/ 10.3390/foods10081952
- Sehrawat, R., Rathee, P., Akkol, E.K., Khatkar, S., Lather, A., Redhu, N., Khatkar, A., 2022. Phenolic acids versatile natural moiety with numerous biological applications. Curr. Top. Med. Chem. 22 (18), 1472–1484. https://doi.org/10.2174/1568026622666220623114450.
- Sepúlveda, C.T., Zapata, J.E., 2019. Efecto de la temperatura, el pH y el contenido en sólidos sobre los compuestos fenólicos y la actividad antioxidante del extracto de *Bixa orellana* L. Inf. Tecnol. 30 (5), 57–66. https://doi.org/10.4067/S0718-07642019000500057
- Servent, A., Abreu, F.A.P., Dhuique, C., Belleville, M., Dornier, M., 2020. Concentration and purification by crossflow microfiltration with diafiltration of carotenoids from a by-product of cashew apple juice processing. Innov. Food Sci. Emerg. Technol. 66, 102519. https://doi.org/10.1016/j.ifset.2020.102519.
- Shahidi, F., Naczk, M., 2003. Phenolics in Food and Nutraceuticals. CRC Press. https://doi.org/10.1201/9780203508732.
- Sharma, S., Rana, S., Patial, V., Gupta, M., Bhushan, S., Padwad, Y., 2016. Antioxidant and hepatoprotective effect of polyphenols from apple pomace extract via apoptosis inhibition and Nrf2 activation in mice. Human Exp. Toxicol. 35 (12), 1264–1275. https://doi.org/10.1177/0960327115627689.
- Sharma, N., Sahil, Madhumita, M., Kumar, Y., Prabhakar, P.K., 2023. Ultrasonic modulated rice bran protein concentrate: induced effects on morphological, functional, rheological, and thermal characteristics. Innov. Food Sci. Emerg. Technol. 85, 103332. https://doi.org/10.1016/j.ifset.2023.103332.
- Sheidai, M., Noormohammadi, Z., Vazifeshenas, M.R., 2024. A new look at genetic variability in pomegranate: landscape genetics and structural equation modeling (SEM). Sci. Hortic. 334, 113323. https://doi.org/10.1016/j.scienta.2024.113323.
- Shirsath, S., Sable, S., Gaikwad, S., Sonawane, S., Saini, D., Gogate, P., 2017. Intensification of extraction of curcumin from Curcuma amada using ultrasound assisted approach: effect of different operating parameters. Ultrason. Sonochem. 38, 437–445. https://doi.org/10.1016/j.ultsonch.2017.03.040.
- Soares, S., Brandão, E., Guerreiro, C., Soares, S., Mateus, N., de Freitas, V., 2020. Tannins in food: insights into the molecular perception of astringency and bitter taste. Molecules 25 (11), 2590. https://doi.org/10.3390/molecules25112590.
- Song, C., Liu, C., Chen, J., 2022. Relationship between heat/mass transfer and ingredient degradation during drying process. J. Food Process. Eng. 45 (1). https://doi.org/ 10.1111/jfpe.13917.

- Sood, A., Saini, C.S., 2022. Red pomelo peel pectin based edible composite films: effect of pectin incorporation on mechanical, structural, morphological and thermal properties of composite films. Food Hydrocoll. 123. https://doi.org/10.1016/j. foodbyd/2021.107135
- Stefanello, F.S., dos Santos, C.O., Bochi, V.C., Fruet, A.P.B., Soquetta, M.B., Dörr, A.C., Nörnberg, J.L., 2018. Analysis of polyphenols in brewer's spent grain and its comparison with corn silage and cereal brans commonly used for animal nutrition. Food Chem. 239, 385–401. https://doi.org/10.1016/j.foodchem.2017.06.130.
- Strieder, M.M., Bragagnolo, F.S., Pizani, R.S., Rostagno, M.A., 2024. The impact of using an expansion gas with high-intensity ultrasound on the pressurized liquid extraction of bioactive compounds from an industrial coffee solid residue. Innov. Food Sci. Emerg. Technol. 92, 103575. https://doi.org/10.1016/j.ifset.2024.103575.
- Tezotto, J.V., de Paula, J.T., Tessmer, M.A., Kluge, R.A., 2018. Ethanol vapor is efficient for reduction of astringency compounds in cashew apple. Postharvest Biol. Technol. 145, 117–124. https://doi.org/10.1016/j.postharvbio.2018.07.002.
- Thorsen, M., Skeaff, S., Goodman-Smith, F., Thong, B., Bremer, P., Mirosa, M., 2022. Upcycled foods: a nudge toward nutrition. Front. Nutr. 9. https://doi.org/10.3389/fnut.2022.1071829.
- Tlais, A., Rantsiou, K., Filannino, P., Cocolin, L., Cavoski, I., Gobbetti, M., Di Cagno, R., 2023. Ecological linkages between biotechnologically relevant autochthonous microorganisms and phenolic compounds in sugar apple fruit (*Annona squamosa* L.). Int. J. Food Microbiol. 387, 110057. https://doi.org/10.1016/j.iifoodmicro.2022.110057
- Trigo, J.P., Alexandre, E.M.C., Saraiva, J.A., Pintado, M.E., 2020. High value-added compounds from fruit and vegetable by-products-Characterization, bioactivities, and application in the development of novel food products. In: Critical Reviews in Food Science and Nutrition. Taylor and Francis Inc, pp. 1388–1416. https://doi.org/10.1080/10408398.2019.1572588, 60 (8).
- Vânia, Z., Ramin, L.Z., 2018. Green and sustainable separation of natural products from agro-industrial waste: challenges, potentialities, and perspectives on emerging approaches. In: Chemistry and Chemical Technologies in Waste Valorization, pp. 229–282. https://doi.org/10.1007/978-3-319-90653-9_8.
- Vázquez, M., Fernández, Á., Bermúdez, A., Rodríguez, E., Pérez, A., Fernández, J., Rodríguez, G., 2020. Utilization of strawberry and raspberry waste for the extraction of bioactive compounds by deep eutectic solvents. Lebensm. Wiss. Technol. 130, 109645. https://doi.org/10.1016/j.lwt.2020.109645.
- Vidal, C., Bastías, J.M., Tamarit, Y., Reyes, J.E., Pérez, M., 2024. Effect of Pulsed Electric Field (PEF) and cryoconcentration (c) in aqueous extracts of maqui (Aristotelia chilensis (Mol.) Stuntz) and calafate (Berberis microphylla G.) on physicochemical properties, microbiological quality, and bioactive compounds. Food Bioprod. Process. 148, 587–601. https://doi.org/10.1016/j.fpp.2024.10.021.
- Vieira, M.V., Pastrana, L.M., Fuciños, P., 2020. Microalgae encapsulation systems for food, pharmaceutical and cosmetics applications. Mar. Drugs 18 (12), 644. https:// doi.org/10.3390/md18120644.
- Vignesh, A., Amal, T.C., Sivalingam, R., Selvakumar, S., Vasanth, K., 2024. Influence of ecological factors on the phytochemical composition and bioactivity of *Berberis* tinctoria Lesch. wild edible fruits. Biochem. Systemat. Ecol. 112, 104771. https://doi. org/10.1016/j.bsc.2023.104771
- Vilas, A., Montemurro, M., Casertano, M., Fogliano, V., 2024. The food by-products bioprocess wheel: a guidance tool for the food industry. Trends Food Sci. Technol. 152, 104652. https://doi.org/10.1016/j.tifs.2024.104652.
- Villacís-Chiriboga, J., Elst, K., van Camp, J., Vera, E., Ruales, J., 2020. Valorization of byproducts from tropical fruits: Extraction methodologies, applications, environmental, and economic assessment: A review (Part 1: General overview of the byproducts, traditional biorefinery practices, and possible applications). In: Comprehensive Reviews in Food Science and Food Safety, 19. Blackwell Publishing Inc, pp. 405–447. https://doi.org/10.1111/1541-4337.12542.
- Vinha, A.F., Costa, A.S.G., Espírito Santo, L., Ferreira, D.M., Sousa, C., Pinto, E., Almeida, A., Oliveira, M.B.P.P., 2024. High-value compounds in papaya by-products (*Carica papaya* L. var. Formosa and Aliança): potential sustainable use and exploitation. Plants 13 (7), 1009. https://doi.org/10.3390/plants13071009.
- Vitor, D., Barrales, F., Pereira, E., Viganó, J., Iglesias, A., Reyes, F., Martínez, J., 2022. Phenolic compounds from passion fruit rinds using ultrasound-assisted pressurized liquid extraction and nanofiltration. J. Food Eng. 325, 110977. https://doi.org/ 10.1016/j.jfoodeng.2022.110977.
- Wang, Z., Yu, W., Liu, M., Wu, Y., Ouyang, J., 2025. Inhibitory effect of bioactive compounds from quinoa of different colors on the in vitro digestibility of starch. Int. J. Biol. Macromol. 297, 139918. https://doi.org/10.1016/j.ijbiomac.2025.139918.
- Wong, J.E., Aguilar-Zárate, P., Veana, F., Muñiz-Márquez, D.B., 2020. Impacto de las tecnologías de extracción verdes para la obtención de compuestos bioactivos de los residuos de frutos cítricos. Tip. Rev. Espec. Ciencias Químico-Biol. 23. https://doi.org/10.22201/fesz.23958723e.2020.0.255.
- Xiang, N., Chang, X., Qin, L., Li, K., Wang, S., Guo, X., 2023. Insights into tissue-specific anthocyanin accumulation in Japanese plum (*Prunus salicina* L.) fruits: a comparative study of three cultivars. Food Chem.: Molecular Sciences 7, 100178. https://doi.org/ 10.1016/j.fochms.2023.100178.
- Xu, Y., Tang, G., Zhang, C., Wang, N., Feng, Y., 2021. Gallic acid and diabetes mellitus: its association with oxidative stress. Molecules 26 (23), 7115. https://doi.org/ 10.3390/molecules26237115.
- Yusoff, I., Chua, L., Taher, Z., 2023. Valorization of fruit waste from Cucurbitaceae family: profiling of phytoconstituent of *Benincasa hispida* and *Citrullus lanatus* rinds using ultrasound-assisted extraction. Food Biosci. 51, 102190. https://doi.org/ 10.1016/j.fbio.2022.102190.
- Zabot, G.L., Viganó, J., Silva, E.K., 2021. Low-frequency ultrasound coupled with highpressure technologies: impact of hybridized techniques on the recovery of

- phytochemical compounds. Molecules 26 (17), 5117. https://doi.org/10.3390/
- Zabot, G., Schaefer, F., Polano, L., Vinícius, M., Herrera, E., Palacin, H., Córdova, J.S., Best, I., Olivera, L., 2022. Encapsulation of bioactive compounds for food and agricultural applications. Polymers 14 (19), 4194. https://doi.org/10.3390/ polym14194194.
- Zahid, H., Ali, A., Ranadheera, C., Fang, Z., Dunshea, F., Ajlouni, S., 2022. In vitro bioaccessibility of phenolic compounds and alpha-glucosidase inhibition activity in yoghurts enriched with mango peel powder. Food Biosci. 50, 102011. https://doi. org/10.1016/j.fbio.2022.102011.
- Zamanipoor, M.H., Yakufu, B., Tse, E., Rezaeimotlagh, A., Hook, J.M., Bucknall, M.P., Thomas, D.S., Trujillo, F.J., 2020. Brewing coffee? Ultrasonication has clear beneficial effects on the extraction of key volatile aroma components and triglycerides. Ultrason. Sonochem. 60, 104796. https://doi.org/10.1016/j.ultsonch.2019.104796.
- Zanivan, J., Bonatto, C., Scapini, T., Dalastra, C., Bazoti, S.F., Júnior, S.L.A., Fongaro, G., Treichel, H., 2022. Evaluation of bioethanol production from a mixed fruit waste by Wickerhamomyces sp. BioEnergy Research 15 (1), 175–182. https://doi.org/ 10.1007/s12155-021-10273-5.
- Zhang, Q., Lin, L., Ye, W., 2018. Techniques for extraction and isolation of natural products: a comprehensive review. Chin. Med. 13 (1), 20. https://doi.org/10.1186/s13020-018-0177-x
- Zheng, J.-L., Hu, L.-Y., Hu, K.-D., Wu, J., Yang, F., Zhang, H., 2016. Hydrogen sulfide alleviates senescence of fresh-cut apple by regulating antioxidant defense system and senescence-related gene expression. Hortscience 51 (2), 152–158. https://doi.org/ 10.21273/HORTSCI.51.2.152.
- Zhou, Y., Cao, Y., Li, J., Agar, O.T., Barrow, C., Dunshea, F., Suleria, H.A.R., 2023. Screening and characterization of phenolic compounds by LC-ESI-QTOF-MS/MS and their antioxidant potentials in papaya fruit and their by-products activities. Food Biosci. 52, 102480. https://doi.org/10.1016/j.fbio.2023.102480.