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## The myeloid binding peptide adenoviral vector enables multi-organ vascular endothelial gene targeting

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### Abstract

Vascular endothelial cells (ECs) are ideal gene therapy targets as they provide widespread tissue access and are the first contact surfaces following intravenous vector administration. Human recombinant adenovirus serotype 5 (Ad5) is the most frequently used gene transfer system because of its appreciable transgene payload capacity and lack of somatic mutation risk. However, standard Ad5 vectors predominantly transduce liver but not the vasculature following intravenous administration. We recently developed an Ad5 vector with a myeloid cell-binding peptide (MBP) incorporated into the knob-deleted, T4 fibritin chimeric fiber (Ad.MBP)<sup>1</sup>. This vector was shown to transduce pulmonary ECs presumably via a vector handoff mechanism. Here we tested the body wide tropism of the Ad.MBP vector, its myeloid cell necessity, and vector-EC expression dose response. Using comprehensive multi-organ co-immunofluorescence analysis, we discovered that Ad.MBP produced widespread EC transduction in lung, heart, kidney, skeletal muscle, pancreas, small bowel, and brain. Surprisingly, Ad.MBP retained hepatocyte tropism albeit at a reduced frequency compared with standard Ad5. While binding specifically to myeloid cells *ex vivo*, multi-organ Ad.MBP expression was not dependent on circulating monocytes or macrophages. Ad.MBP dose de-escalation maintained full lung targeting capacity but drastically reduced transgene expression in other organs. Swapping the EC-specific *ROBO4* for the *CMV* promoter/enhancer

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### DISCLOSURE

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abrogated hepatocyte expression but also reduced gene expression in other organs. Collectively, our multilevel targeting strategy could enable therapeutic biologic production in previously inaccessible organs that initiate the most debilitating or lethal human diseases.

## Keywords

adenovirus vector; gene therapy; endothelial cell; gene transfer

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Despite development of targeted small molecule inhibitors or biologics, both benign and malignant diseases remain challenging or even recalcitrant to control or cure. One problem is that therapies are not tightly targeted to disease and produce systemic complications. Another obstacle is disease resistance emergence. Both of these challenges could be addressed by development of agents that would be systemically administered, home to the disease site, and express high levels of a therapeutic either persistently or under switchable control<sup>2-6</sup>.

One option for targeted therapeutic delivery is adenoviral (Ad) vectors. These biologics can be genetically manipulated for disease site homing, infection of relevant target cells, and high-level transgene expression<sup>3, 7-9</sup>. Disease site-specific transductional targeting can be accomplished by genetic manipulation of the viral capsid<sup>6, 9</sup>. Transcriptional targeting for expression within specific cell types can be accomplished using DNA enhancer/promoter elements incorporated into the vector transgene<sup>10</sup>. Regulated targeted gene expression may be achieved through drug-inducible enhancers/promoters<sup>11</sup>, or DNA secondary structural elements<sup>12</sup> or microRNA seed sequences placed up or downstream of the transgene open reading frame<sup>13</sup>.

One portal to disease sites marginally accessed by vectors is vascular endothelium. As many diseases evoke expansion of existent or creation of new blood vessels, endothelial cell (EC) targeted viral vectors could produce therapeutics titrated to disease severity. Moreover, as the vascular ECs are the initial cell layer in contact with intravenously injected therapeutics, first-pass organ targeting can be achieved<sup>5</sup>. Several viral vectors have been engineered for vascular EC targeting as potential novel therapeutics for both benign and malignant diseases<sup>14, 15</sup>. In oncology, the predominant goal has been vascular ablation. However, the strategy of “starving” tumors has elicited resistance either due to coordinate upregulation of untargeted factors by malignant cells, or dynamic micro environmental remodeling<sup>2</sup>. Other than hypervascular intraocular disease, vessel ablation would be contraindicated for most benign diseases. An alternative to vascular ablation is manipulation of the molecular secretion repertoire of ECs. Angiocrine has been the term assigned to EC production or display of membrane tethered growth factors, cyto-/chemokines, and stem cell nurturing factors<sup>16</sup>. Engineering angiocrine functions is a compelling opportunity for vector-based, novel disease therapeutics both for benign and malignant disease.

Here we tested the body-wide cell type expression of a previously reported vascular EC targeted Ad5 vector containing a heptapeptide, “myeloid binding peptide” (MBP). Ad.MBP was engineered for MBP display at the tip of a “de-knobbed” chimeric fiber<sup>5, 17</sup>. This vector was shown to bind specifically to myeloid cells *ex vivo* but predominantly transduced lung

vascular endothelium following systemic administration<sup>1</sup>. Here, we discovered that the Ad.MBP vector affected multi-organ EC specific expression. Most striking was the finding that hitherto inaccessible organs evidenced EC transgene expression including brain, small and large bowel mucosa, kidney glomeruli, medulla, and papilla, skeletal muscle, and cardiac subendocardium and myocardium. The Ad.MBP vector presents an opportunity for targeting the most prominent and vexing human diseases.

## MATERIALS AND METHODS

### Animals

All mice were of C57BL/6J background and seven to fourteen weeks of age. Mice were obtained from Jackson Laboratory (Bar Harbor, ME) or through breeding in authors' animal facility. Experimental procedures involving mice were carried out under protocols #20120029 and #20110035 approved by the Washington University Animal Studies Committee.

### Cells and Adenovirus vectors

Human embryonic kidney HEK293 cells were purchased from Microbix Biosystems (Ontario, Canada). Cells were cultured in DMEM/F12 (Mediatech, Herndon, VA) media containing 10% fetal bovine serum (FBS) (Summit Biotechnology, Fort Collins, CO), in a humidified atmosphere with 5% CO<sub>2</sub> at 37°C. Replication incompetent *E1* - and *E3* -deleted Ad5 vectors were created using a two-plasmid rescue method. Plasmids encoded expression cassettes containing either the cytomegalovirus major immediate-early enhancer/promoter (CMV), or the human *roundabout4* (*ROBO4*) enhancer/promoter, each cloned upstream of enhanced green fluorescent protein (EGFP) followed by the bovine growth hormone polyadenylation signal. These expression cassettes were cloned into a shuttle plasmid (pShuttle, Qbiogene, Carlsbad, CA) to generate the pShuttleCMV-EGFP and pShuttleROBO4-EGFP plasmids, respectively, and inserts were confirmed by using restriction enzyme mapping and partial sequence analysis.

The shuttle plasmids were linearized with *Pme* I and integrated into the Ad5 genome by homologous recombination with apAd5 plasmid, encoding the native Ad5 fiber, or a pAdMBP plasmid, encoding an MBP-fiber-fibrin chimera, in the *E. coli* strain BJ5183. To rescue Ad.MBP.ROBO4, the recombinant viral genome was linearized with *Pac* I and then transfected into 293F28 cells using SuperFect Transfection Reagent (Qiagen, Chatsworth, CA). 293F28 cells stably express the native Ad5 fiber; thus, viruses rescued at this point were mosaic in the sense that the Ad5 virions randomly incorporated a mixture of native Ad5 fibers and MBP-fiber-fibrin chimeras<sup>18</sup>. After an additional round of amplification on 293F28 cells, the viruses were amplified in HEK293 cells, which do not express native Ad5 fiber, to obtain virus particles containing only MBP-fiber-fibrin proteins. The Ad.MBP.CMV vector containing the sequence WTLDRGY on a T4 fibrin chimeric fiber knob was created as described previously<sup>1, 17</sup>. Recombinant viruses were purified by two rounds of CsCl density ultracentrifugation and dialyzed in storage buffer containing 10 mmol/L HEPES, 1 mmol/L MgCl<sub>2</sub>, pH 7.8 with 10% glycerol as previously described<sup>19</sup>.

The viral particle (vp) concentration was determined by absorbance of dissociated virus at  $A_{260\text{nm}}$  using a conversion factor of  $1.1 \times 10^{12}$  vp/absorbance unit.

### Warfarin and clodronate-liposome treatment

Mice were subcutaneously injected with warfarin, 5 mg/kg in peanut oil, 72 hours and 24 hours prior to virus injection<sup>20</sup>. Clodronate-liposomes, 10  $\mu\text{L/g}$  body weight, ([ClodronateLiposomes.com](http://ClodronateLiposomes.com), Netherlands) or saline buffer were injected into the tail vein 48 and 24 hours prior to vector injection<sup>21</sup>. Twenty-four hours later, peripheral blood was collected by cheek pouch bleeding, and then Ad.MBP was injected.

### Virus injection and host organ harvest

Mice were tail-vein injected with  $1 \times 10^{11}$  or  $2 \times 10^{10}$  particles of virus in 200  $\mu\text{L}$  of saline. Seventy-two hours post virus administration, mice were anesthetized with 2.5% 2, 2, 2-tribromoethanol (Avertin, Sigma-Aldrich, St. Louis, MO), perfused via the left ventricle with phosphate-buffered saline (PBS) followed by 10% neutral buffered formalin. Harvested organs were post-fixed in formalin at room temperature for 2 to 4 hours, cryo-preserved in 30% sucrose in PB Sat 4°C overnight. Lung was further inflated and fixed by injecting formalin solution into trachea followed by closing the trachea by ligature and then processed as above. Treated tissues were embedded in NEG50 (Thermo Fisher Scientific, Waltham, MA) or Tissue-Tek OCT mounting medium (Sakura Torrance, CA, USA), and frozen in a liquid nitrogen pre-chilled, 2-methylbutane-containing glass beaker.

### Immunofluorescence staining

All mouse tissues were cryosectioned at 16  $\mu\text{m}$ . Lung was also cut at 5  $\mu\text{m}$  for determination of transgene microvessel co-localization. Frozen section slides were air-dried for ten minutes, washed three times in PBS, blocked with protein block solution (5% donkey serum and 0.1% Triton X-100 in PBS) for one hour, and incubated at 4°C overnight in protein block containing primary antibodies including: rat anti-endomucin 1:1,000, rat anti-PDGFR $\beta$  1:200 (#14-5851-81, and #14-1402-81, eBioscience, San Diego, CA), Armenian hamster anti-CD31 1:1,000, rabbit anti-NG2 chondroitin sulfate proteoglycan 1:100 (#MAB1398Z and #AB5320, EMD-Millipore, Billerica, MA), rat anti-CD45 1:100 (#550539, BD Biosciences, San Jose, California), rat anti-F4/80 1:500 (#MCA497R, AbD Serotec-BioRad, Raleigh, NC), rabbit anti-GFP1:400, and chicken anti-GFP 1:400 (#A11122 and #A10262, Life Technologies, Carlsbad, CA). The two GFP antibodies performed equally well; the chicken anti-GFP antibody was used in the clodronate-liposome experiment, and the rabbit antibody was used throughout the rest of the study. On day 2, the slides were washed three times in PBS, incubated with corresponding 1:400 diluted Alexa Fluor 488- and Alexa Fluor 594-conjugated secondary antibodies (Jackson ImmunoResearch Laboratories, West Grove, PA), and counterstained with SlowFade Gold Antifade mounting reagent with 4',6-diamidino-2-phenylindole (DAPI) (Life Technologies).

### Immunofluorescence microscopy-based analysis of viral reporter gene expression

Immunofluorescence images were collected using an Olympus BX61 microscope equipped with an FV7II digital camera (Olympus America, Center Valley, PA). The Extended Focal

Imaging (EFI) function was used in collecting all high-magnification micrographs to allow the creation of a single in-focus image from a series of views of the same field at different z-dimensional focal planes at 2 $\mu$ m intervals. EFI was carried out in a live-processing mode during image acquisition. Camera acquisition time for EGFP immunofluorescence was optimized and set *a priori* for each organ through independent experiments where the collected data were pooled for statistical analyses. The optimized acquisition time for EGFP immunofluorescence display was 200 msec for liver, 400 msec for spleen, 300 msec for lung, 300 msec for heart, 300 msec for kidney, 1 sec for muscle, 500 msec for pancreas, 1 sec for small bowel, 1 sec for large bowel, and 500msec for brain. Where vera figure contains micrographs collected using a different level of exposure for EGFP, the setting is indicated in the figure legend. Immunofluorescence micrographs were subjected to measurement of both color intensity and color-positive area using MicroSuite Biological Suite image analysis software Version 5 (Olympus). To determine the EGFP fluorescence intensity, a threshold defining the background green fluorescence color for each pixel was set at 70 while the possible range of intensity values were from 0 representing a complete absence of green color intensity to 255 taken to be of full intensity. A region of interest (ROI) was drawn over the tissue compartment in each image, and positive ID particles in the ROI, defined as containing at least 5 connected pixels all with above the background color intensity, were identified. The color intensity values from every pixel of positive ID particles were summed and normalized by the tissue ROI area (per  $\mu$ m<sup>2</sup>). To evaluate the fraction of tissue vascular area expressing EGFP, the endothelial marker-positive area and EGFP-positive area within the tissue ROI were quantified by summing up the areas of all positive ID particles based on the color detection threshold of 70 for both the green and red colors. The percentage ratio of EGFP-positive area to EC-positive area in each organ was calculated for evaluating the vascular EC vector gene expression. Mean and standard deviation of data points in each organ derived from experiment mice were plotted.

### Quantitative flow cytometry

Peripheral blood was collected from mice treated with vehicle or clodronate liposomes and 50  $\mu$ L of each sample was spiked with re-fluorescent beads (Invitrogen, CA) as internal standards for absolute counts. Red blood cells were lysed with red blood cell lysis buffer (BioLegend, San Diego, CA), and mononuclear cells (MNCs) were isolated. MNCs were then washed with cold PBS and stained with CD11b-fluorescein isothiocyanate (FITC) and CD45-phycoerythrin (PE) (BD Pharmingen, BD Biosciences, San Jose, CA) for 1 hr on ice. Then, cells were washed, re-suspended in PBS and analyzed by flow cytometry. Forward scatter (FSC) and side scatter (SSC) were used to gate monocytes as high-size (FSC)/ low-granulation (SSC) population; moreover, the monocyte population was further characterized as CD11b-positive/CD45-positive. The count of the FSC-high/SSC-low/CD11b-positive/CD45-positive monocyte population was normalized to the count of the fluorescent beads. Results were presented as the % of average of vehicle treated mice.

### Statistical analysis

All data are reported as mean  $\pm$  standard deviation. Significance of the means between the mouse groups was determined using unpaired Student's test for each organ with Bonferroni correction for multiple independent comparisons carried out on different organs from the

same cohort of mice. Statistical significance was defined as adjusted  $P < 0.05$  (GraphPad Prism, San Diego, CA).

## RESULTS

### **MBP pseudotyping attenuated hepatocyte vector expression while producing widespread multi-organ vascular EC expression**

Our initial work with the Ad.MBP.CMV vector focused on differential liver and lung transduction<sup>1</sup>. Here we sought to test the multiorgan biodistribution of Ad.MBP.CMV using semi quantitative tissue section immunofluorescence analysis. As expected, Ad5.CMV-mediated expression was predominantly localized in liver hepatocytes and readily detectable in reticuloendothelial system and endothelial cells (ECs) of spleen (Figure S1A). Vector expression was scarcely found in lung, heart, kidney, gastrocnemius muscle, pancreas, small bowel, large bowel, and not detectable in any part of the brain (Figure 1B). In contrast, Ad.MBP.CMV produced EC expression throughout the microvasculature of heart, kidney, muscle, pancreas, intestine, and brain (Figure 1A). Vector EC co-localization was confirmed using high-magnification EFI imaging in these organs (Figure S2). Surprisingly robust transgene expression was detectable in ECs within all brain regions including cerebrum, cerebellum, hippocampus, and medulla (Figure S1B). To quantify vector transgene expression, EGFP fluorescence intensity was summed in a tissue region of interest (ROI) and normalized by the ROI area (per  $\mu\text{m}^2$ ) in each organ. Liver sections from Ad.MBP.CMV-injected mice exhibited a 5-fold reduction in the EGFP fluorescence intensity compared with the Ad5.CMV counterparts (Figure 1B). Liver detargeting was associated with 2-fold increase in vector expression in splenic reticuloendothelial cells and ECs (Figure 1B). A slung, heart, kidney, pancreas, small and large bowel, and brain were barely or not transducible by the Ad5.CMV, the retargeting enhancement of the Ad.MBP.CMV to these organs was remarkable, ranging from greater than 10-fold increase in pancreas, small bowel, and large bowel, greater than 100-fold increase in lung and kidney, greater than 1,000-fold increase in heart and muscle, and greater than 10,000-fold increase in brain (Figure 1B, red bars versus blue bars for Lu, H, K, M, P, SB, LB, and B).

We next determined the EC-expression efficiency of the Ad.MBP.CMV versus Ad5.CMV in multiple organs by quantifying the percentage of total tissue EC area expressing EGFP in each organ. Ad.MBP.CMV targeted greater than 62% of blood vessels in all regions of the brain (B), 21% in lung (Lu), 26% heart (H), 33% in kidney (K), 38% in muscle (M), 30% in pancreas (P), 16% in small bowel (SB), and 6% in large bowel (LB) (Figure 1C). Other than liver and spleen, pancreas and small bowel were the only organs where Ad5.CMV produced an appreciable but still rare vascular EC expression (Figure 1C).

To further test EC-specific expression, immunofluorescence for the pericyte markers, PDGFR $\beta$  or proteoglycan nerve-gial antigen 2 (NG2), and the pan-hematopoietic lineage cell marker CD45 or macrophage marker F4/80 were performed. High-magnification revealed that the EGFP-expressing cells were clearly distinct from the PDGFR $\beta$ -positive or NG2-positive cells in all organs (Figure S2). Ad.MBP.CMV was expressed in rare CD45-positive hematopoietic cells and F4/80-positive macrophages in liver and spleen, but not in any other sampled organs (Figure S3).

### Warfarin liver detargeting failed to increase Ad.MBP.CMV multiorgan EC expression

While the Ad.MBP.CMV yielded an impressive level of hepatocyte detargeting, liver remained a substantial transductional and transcriptional target (Figure 2). As the major pathway directing Ad5hepatocyte sequestration is mediated by coagulation Factor X-viral hexon binding, we tested whether warfarin could affect diminution in the level of Ad.MBP.CMV expression in hepatocytes<sup>22</sup>. Warfarin pretreatment diminished the number of EGFP-expressing hepatocytes (Figure 2A). The residual number of vector expressing hepatocytes was similar to our previous work with Ad5 based vectors with wild type capsids. This residual hepatocyte vector expression following warfarin treatment has been seen by others and likely represents a “floor” for the efficacy of pharmacological blockade<sup>22</sup>. However, in contrast to our previous work with vectors with wild type capsids<sup>23</sup>, warfarin failed to enhance EC expression in all other organs (Figure 2A and 2B). These data suggested that either the Ad.MBP.CMV peripheral vascular EC vector expression was saturated at our  $1 \times 10^{11}$  viral particle dose, or that complement opsonization facilitated destruction of the vector dose increment that escaped hepatocyte transduction<sup>24</sup>.

### Ad.MBP.CMV dose reduction produced organ-specific non-linear EC expression reduction

To test the sensitivity of each organ vascular bed for Ad.MBP.CMV expression we challenged mice with injection of  $2 \times 10^{10}$  viral particles. The lower viral dose drastically reduced tissue Ad.MBP.CMV expression in liver, spleen, pancreas, heart, kidney, muscle, pancreas, small bowel and brain. Strikingly, the frequency and EC expression level in the lung remained unaffected by vector dose reduction (Figure 3A and 3B). Comparison of EGFP fluorescence intensity of low- versus high-dose tissue samples revealed that splenic and brain transgene expression was 16% and 31% of the high-dose counterparts (Figure 3C, S and B). These levels of reduction in EGFP expression were within the range of linear response to the viral dose difference (20%). However, the drastically diminished expression in liver (5% of high-dose level), heart (0.4% of high-dose level), kidney (0.5% of high dose level), muscle (0.1% of high-dose level), pancreas (0.4% of high-dose level), and small bowel (3% of high-dose level) was non-linear. Similar to EC expression frequency analysis, vector dose reduction failed to significantly diminish transgene expression in lung (91% of high-dose level,  $p=0.588$ ). These results suggested that Ad.MBP. CM Vlung-specific EC expression targeting may be achievable through vector dose fine tuning.

### Mononuclear cell depletion failed to diminish Ad.MBP.CMV EC transgene expression

The Ad.MBP.CMV acquired a specific and high affinity binding to myeloid cells *ex vivo*, compared with the Ad5<sup>1, 17</sup>. To test the necessity of mononuclear cells for Ad.MBP.CMV EC transgene expression in intact mice, we administered two daily doses of intravenous clodronate liposomes and measured the percentage of circulating CD11b-positive cells and the level of multi-organ tissue section EGFP fluorescence (Figure 4A–C). Clodronate treatment depleted circulating CD11b-positive leukocytes by 77% and completely depleted F4/80-positivemacrophages in liver (Kupffer cells) and spleen (Figures 4A and S4). In contrast, clodronate barely reduced resident macrophages in lung, small bowel, heart, and kidney (data not shown). Clodronate increased Ad.MBP.CMV EC lung expression by 2-fold but did not significantly alter EC transgene expression level or the EC-specific expression

pattern in liver, spleen, heart, kidney, muscle, pancreas, small bowel, or brain (Figure 4B). The lack of increase in hepatocyte was surprising given prior reports on the scavenging function of liver Kupffer cells<sup>25</sup>, however, others have also reported a modest, though statistically insignificant level of clodronate-mediated Ad vector liver expression enhancement<sup>26</sup>. Most importantly our data demonstrate that circulating monocytes and macrophages were dispensable for Ad.MBP.CMV organ EC expression. However, the persistence of extrahepatic organ Cd11b and F4/80 cells following clodronate depletion does not rule out the marginated myeloid cell pool as a mechanism for vector-EC handoff<sup>1</sup>.

### **EC-specific *ROBO4* gene promoter/enhancer ablated the MBP vector hepatocyte expression but reduced EC expression in other organs**

We recently engineered an Ad5 vector for transcriptional targeting of ECs using the EC-specific human *ROBO4* gene enhancer/promoter fragment<sup>10, 23</sup>. To test whether the combination of transcriptional with transductional targeting could produce enhanced multi-organ EC expression, we replaced the *CMV* promoter with the *ROBO4* enhancer/promoter. The dual targeted Ad.MBP.*ROBO4* vector was administered intravenously and organs were analyzed for vector transgene expression (Figure 5A). Ad.MBP.*ROBO4* abrogated hepatocyte expression and instead EGFP was detectable in a scattered population of liver ECs (Figure 5A). The Ad.MBP.*ROBO4* also produced EC transgene expression in an appreciable vascular area fraction in spleen (23%), kidney (23%), lung (10%), muscle (9%), heart (10%), and brain (15%) but produced very low expression in small bowel and large bowel (1% and 2% respectively) (Figure 5B, S, Lu, H, K, M, SB, LB, and B). Collectively, the *ROBO4* enhancer/promoter produced an appreciably lower host organ EC expression compared to Ad.MBP.CMV in each organ. However, the undetectable vector transgene expression in hepatocytes highlighted the enhanced endothelial cell type stringency of the *ROBO4* compared to the *CMV* promoter in the Ad.MBP vector.

## **DISCUSSION**

Vascular endothelium has been a sought after gene therapy target because of its immediacy to blood-borne therapeutics and its pathophysiological role in a wide range of benign and malignant diseases<sup>2, 5, 15, 27</sup>. Despite their accessibility, vascular ECs are poor transduction targets for unmodified Ad5 vectors<sup>14</sup>. In addition, systemically administered Ad5 is rapidly opsonized by circulating IgM antibodies and complement components, leading to virus clearance by liver Kupffer cells<sup>8</sup>. Ad5 also avidly binds to blood coagulation factor X, which bridges the virus to hepatocytes by interacting with cell surface heparan sulfate proteoglycans<sup>22</sup>. Liver Kupffer cell clearance and hepatocyte transduction greatly limit circulating Ad5 vector efficacy. Thus, molecular engineering efforts to achieve Ad vector vascular targeting have focused on diminishing or abrogating liver tropism and opsonization while increasing EC transduction<sup>8, 10</sup>. Liver detargeting engaged genetic capsid modification. Virus opsonization diminution has been addressed using chemical shielding of Ad5 capsid proteins<sup>8</sup>. One approach to EC transductional targeting has been vector pseudotyping. Ad5 vectors pseudotyped with fibers or fiber knobs from different human, or from non-human, serotypes exhibited improved transduction efficiency of cultured human or rodent (rat) ECs<sup>28-30</sup>. Another approach has been addition of “angio-adenobodies” onto the



Ad vector capsid. These bispecific biologics shield the native tropism receptors by one antibody site and target EC luminal surface receptors using another antibody site<sup>6</sup>. EC receptors targeted by this mechanism include VEGFR-2, Tie-2, E-selectin, endoglin, and membrane-bound angiotensin converting enzyme<sup>7, 31–33</sup>. EC transduction has also been achieved through capsid fiber knob display of peptide ligands such as the arginine-glycine-aspartate (RGD) motif cognate for the angiogenesis associated integrins  $\alpha_v/\beta_5$  and  $\alpha_v/\beta_3$ <sup>29, 34</sup>, or the DDTRHWG peptide<sup>3</sup>. A parallel strategy for EC specificity has been transcriptional targeting using enhancer/promoter elements of endothelial-specific genes such as *VEGFR-2*, *VEGFR-1*, *preproendothelin-1*, and *roundabout-4*<sup>10, 23, 35–38</sup>. Transcriptional targeting restricts vector transgene expression to specific EC populations that in most instances are angiogenic and in some cases also hypoxic. However, the transcriptional strategy, when applied alone, does not alter the Kupffer cell sequestration or hepatocyte transduction. Recent efforts have focused on the combination of transductional and transcriptional strategies to achieve enhanced organ or disease specific EC vector transgene expression<sup>10, 30</sup>. Despite progress, systemically administered Ad vectors are still ineffective in gene transfer to some clinically important organs. Dose escalation to achieve appreciable vector expression in marginally accessible organs likely will fail due to dose-limiting adverse effects such as liver toxicity, cytokine storm, or organ imperviousness to the vector<sup>39</sup>. Collectively, the limitations of current EC targeting efforts reinforce the need for further vector improvement.

Ad.MBP was previously shown to preserve the myeloid cell-binding specificity of the WTLDRGY MBP peptide *ex vivo*<sup>17</sup>, but efficiently and preferentially target gene expression to the lung microvessel ECs *in vivo*<sup>1</sup>. The latter work used single-cell lung suspensions and confirmed that Ad.MBP solely bound to myeloid cells and not to ECs. Co-culture of virus-loaded myeloid cells on an EC monolayer provided indirect evidence supporting a myeloid cell-mediated viral “hand-off” mechanism for potentiating the EC transduction<sup>1</sup>. Similar carrier cell hand-off or “hitchhiking” target cell transduction was proposed for other viruses *in vivo*<sup>40, 41</sup>. A central tenet of vector hand-off postulates close contact of virus-carrier cells to the target cells enabling viral penton access with target cell integrins for internalization, bypassing the requirement of an initial attachment step in cell transduction<sup>41</sup>. Indeed, the previous lung work revealed that the Ad.MBP virions rapidly bound to lung following intravenous injection. There the hypothesis was that vector attachment and “hand-off” to lung ECs was mediated by marginated neutrophils<sup>1</sup>. Our current data extended these findings and revealed that the MBP peptide possessed a much broader EC-specific tropism *in vivo*, and many of the permissive recipient organs exhibit a wide range of tissue-specific forms of resident myeloid cells. Our clodronate study provided evidence suggesting that circulating mononuclear cells and tissue resident macrophages in liver and spleen were dispensable or redundant for mediating the Ad.MBP EC expression. Nevertheless, it remains to be demonstrated whether the resident myeloid populations in other target organs are essential for the virus EC targeting. Importantly, the existing data on Ad.MBP transduction may also be explained by an alternative mechanism to hand-off, in which a yet to identified MBP cognate cell surface receptor or blood-borne factors also contribute Ad.MBP EC transduction. Of additional interest was the fact that the Ad.MBP vector produced multi-organ vascular expression following warfarin-mediated Factor X depletion. Indeed previous

work demonstrated that Factor X-virus hexon binding “shielded” the vector from peripheral natural antibody-mediated destruction in immuno competent mice<sup>24</sup>. One explanation could be that the fiber modification of the Ad.MBP vector somehow protected the vector in the peripheral circulation. That said, overall our work additionally differs from this paper in that we repeatedly demonstrated a robust warfarin effect in Rag2 knockout complement-deficient mice, whereas warfarin was ineffectual in the reported Rag-deficient background<sup>24</sup>. As such, the role of Factor X-complement-mediated systemic vector opsonization will certainly need to be independently tested.

Multiorgan expression analysis also enabled us to expand on previous observations and discover the exquisite lung tropism of our Ad.MBP vector. Viral particle dose reduction essentially eliminated gene transfer to most organs while maintaining robust lung expression. This apparent pulmonary vascular avidity suggests that the Ad.MBP vector could be an ideal vehicle for treatment of pulmonary diseases, particularly those initiated by single gene mutations. Over the past two decades, several gene therapy trials have been accomplished for alpha-1 antitrypsin deficiency (AATD). AATD is caused by a mutation in the *alpha-1 antitrypsin (AAT)* gene that produces a misfolded, nonfunctional protease inhibitor<sup>42</sup>. Affected individuals develop emphysema due to pulmonary tissue destruction and cirrhosis from hepatocyte AAT protein aggregation. Gene therapy has sought to restore lung AAT levels via elevation of circulating wild-type enzyme and inhibition of hepatocyte mutant protein accumulation<sup>43</sup>. Recently a combinatorial approach using microRNA (miR) targeting the AAT mutant protein and a transgene encoding a miR-resistant wild-type protein was highly efficacious in a mouse genetic model<sup>44, 45</sup>. The dual hepatocyte and lung EC expression profile of our Ad.MBP vector suggests that it could be an ideal vehicle for AATD therapy with a similar miR/wild-type bicistronic transgene. Likewise, cystic fibrosis (CF) is a lung disease caused by disrupted chloride channel function particularly affecting the airway epithelium due to mutations in the gene *cystic fibrosis transmembrane conductance regulator (CFTR)* gene. So far, 25 viral and non-viral gene therapy clinical trials for CF have been conducted or are in progress aiming at direct gene transfer to the airway epithelium<sup>46</sup>. Some the studies evidenced partial correction of the chloride transport defect, but clinical benefits have not been equivocal<sup>46</sup>. While the primary CF defect is abnormal epithelial mucous production, marked neutrophil recruitment, neutrophil elastase production, and overall inflammatory hypersensitivity accompany the disease and lead to bronchial wall destruction (bronchiectasis). Here, the perialveolar and peribronchial Ad.MBP vascular EC expression could be a platform for secretion of vector transgenes inhibiting neutrophil elastase or inflammatory mediator expression<sup>47</sup>. In addition, the unique multi-organ vascular expression of the Ad.MBP vector, in particular within the intestinal and pancreatic vasculature, presents a tantalizing opportunity to treat nearly all of the major organ systems affected by CF. Again, however, the epithelial-centric nature of CF will require a creative choice of transgenes to achieve multi-site efficacy.

While Ad.MBP has many conceivable applications in other organs, its widespread expression in cardiac and brain vasculature is particularly exciting. In the heart, gene therapy has focused on ischemic disease<sup>48</sup>. While the immediate cause of cardiac ischemia is coronary artery atherosclerosis, myocardial remodeling is the principal mechanism for

development of chronic congestive heart failure<sup>49</sup>. Restoration of blood flow has been approached using gene therapy as a surgical adjuvant or as primary treatment<sup>4, 50</sup>. So far, more than a dozen of trials using first-generation Ad5-based vectors, have been or are being carried out, to deliver angiogenic factors such as the VEGF family members, FGF-4, HGF, or angiogenic regulator HIF1 $\alpha$  to ischemic myocardium via the intracoronary or intramuscular route. While a few Phase I and II studies resulted in encouraging endpoints such as statistically significant increase in regional myocardial perfusion<sup>51, 52</sup>, or improved exercise time and angina<sup>53-55</sup>, several phase III trials yielded overall disappointing results<sup>56, 57</sup>. The reasons for the variegated therapeutic outcomes have not been fully understood, but vector types, therapeutic genes, and routes of administration, and the presence of anti-Ad5 neutralizing antibodies may have contributed<sup>4</sup>. Our Ad.MBP vector could solve the dual challenge of coronary perfusion and myocardial remodeling. Coronary perfusion could be increased using Ad.MBP vector armed with constitutively active hypoxia-inducible factors (HIFs)<sup>38</sup>. The advantage of HIFs is that these transcription factors upregulate a collection, rather than a single angiogenic factor, and induce a leakage resistant hypervascularity<sup>58, 59</sup>. The widespread myocardial vascular distribution of Ad.MBP presents the opportunity to capitalize on EC angiocrine functions such that ECs are transformed into local sources of HIF-mediated angiogenic factors to both preserve marginal zone myocardial viability, and potentially augment arteriogenesis. Similarly, Ad.MBP vectors containing polycistronic transgenes encoding the same molecules apparently secreted by MSCs or ESCs could possibly effect restorative rather than pathological myocardial remodeling by inducing expansion and myocardial differentiation of perivascular resident cardiac stem cells<sup>60, 61</sup>.

In contrast to the goal of improvement of vascularity through transient angiogenic gene expression for heart gene therapy, brain gene therapy strives to achieve long-term expression in neurological disorders such as Alzheimer's, amyotrophic lateral sclerosis(ALS), or brain cancer<sup>62-64</sup>. Benign neurological diseases have been approached using intraventricular vector injection. While intraventricular vector injection has produced some efficacy preclinically, this strategy predominantly targets the brain surface rather than the brain parenchyma. Recent studies have highlighted the potential for local production of secreted antibodies targeting mutant SOD in amyotrophic lateral sclerosis and tau protein in Alzheimer's<sup>65, 66</sup>. Here again, ability of our Ad.MBP vector to target greater than 62% of blood vessel beds in all regions of the brain offers the intriguing potential for treating the multifocal intraparenchymal mechanisms for both diseases. In brain cancer, glioblastoma (GBM) in particular, gene therapy has predominantly been restricted to vector injections into the surgical bed<sup>67</sup>. This approach ultimately fails due to the invasiveness of brain cancer wherein host vessel cooption leads to widespread permeation. Moreover, GBM is notoriously resistant to either conventional chemo-irradiation or molecularly targeted therapies. This resistance is based in part on GBM stem cells that have been shown to reside in perivascular niches throughout the brain<sup>68</sup>. Collectively, the remarkable tropism of the Ad.MBP vector for brain vascular ECs offers the possibility for targeting perivascular GBM stem cells by angiocrine mediated secretion of secreted cytotoxics or molecules blocking signaling pathways maintaining this therapy resistant cell population<sup>69, 70</sup>. Indeed, we have preclinical studies underway to test these hypotheses.

In summary, our Ad.MBP vector enables unprecedented multi-organ vascular access. This vector can be used to harness ECs for production of a variety of therapeutic molecules for a diverse collection of benign and malignant diseases. While promiscuous vascular vector expression might be detrimental, its multi-organ tropism may in fact be uniquely beneficial. In cases where in greater disease specificity is required, the inherent EC vector tropism enables swapping in enhancer/promoters tailored to the altered microenvironment created by each disease in each organ. Clearly, considerable preclinical modeling, functional testing, and possibly additional genetic engineering will need to be done to validate the Ad.MBP vector's potential and possibly increase its expression duration. However, its potential to treat multiple diseases in previously inaccessible organs is compelling.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## ACKNOWLEDGEMENTS

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## Abbreviations

<b>MBP</b>	myeloid cell binding peptide
<b>EC</b>	endothelial cell
<b>Ad5</b>	adenovirus serotype 5
<b>HIFs</b>	hypoxia-inducible factors

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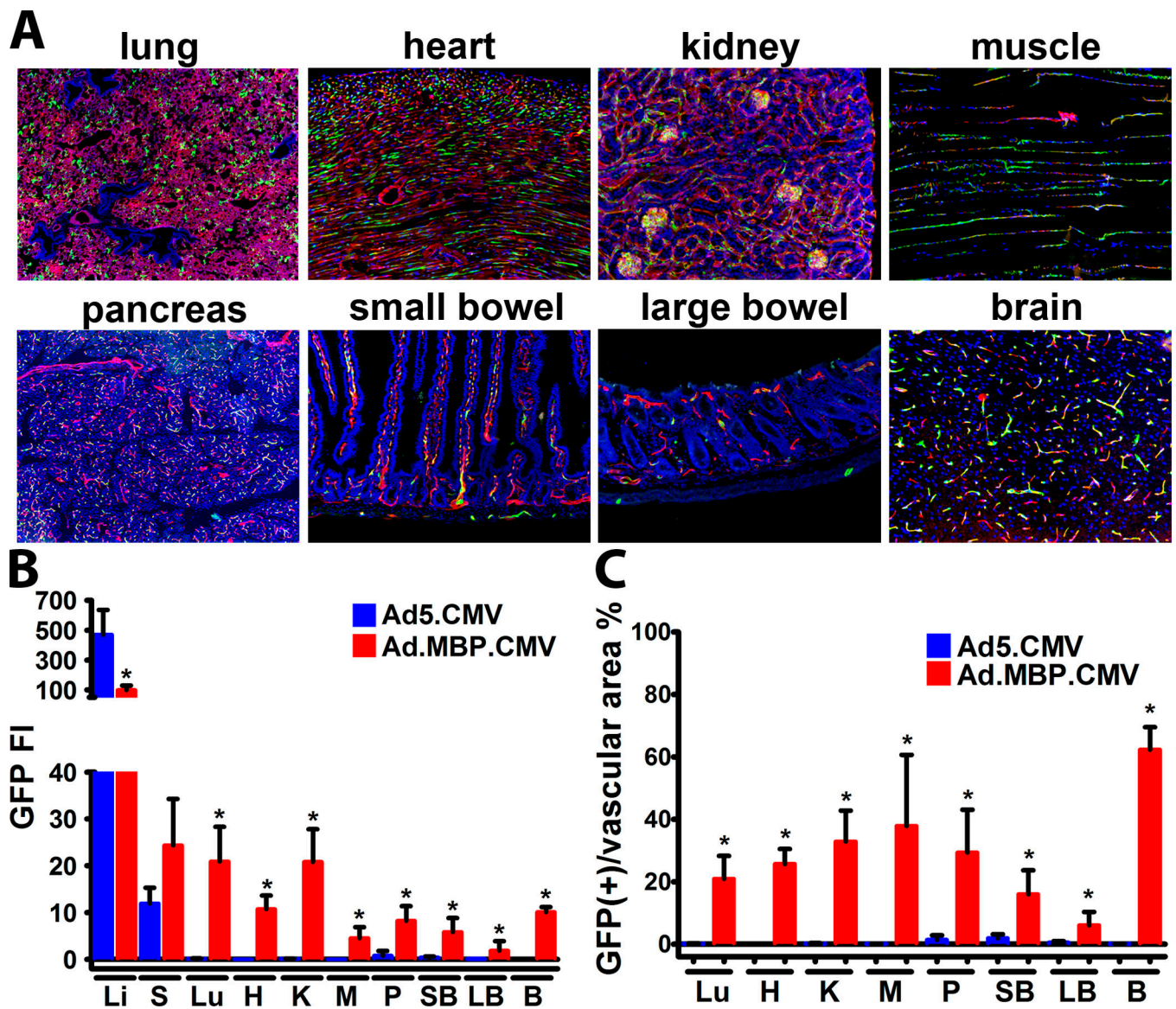
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**Figure 1. Incorporation of MBP into Ad5 drastically increased viral gene expression to vascular beds of multiple host organs**

(A) Immunofluorescence microscopy analysis of vector EGFP expression in host organs following intravenous injection of  $1 \times 10^{11}$  viral particles (vp) of Ad.MBP.CMV into adult C57BL/6J mice revealed prominent transgene expression in lung, heart, kidney, gastrocnemius muscle, pancreas, small and large bowel, and brain. Co-staining of tissue sections with an EC-specific endomucin/CD31 cocktail revealed that EGFP expression was restricted to the vasculature. (B) EGFP fluorescence per  $\mu\text{m}^2$  of tissue section area (FI, fluorescence intensity) in each organ derived from Ad5.CMV-injected mice ( $n=4$  for all organs) versus that from Ad.MBP.CMV-injected mice ( $n=10$  for liver, spleen, heart, kidney, muscle, small bowel, and brain;  $n=7$  for lung, pancreas, and large bowel). (C) The percentage of vascular EC area expressing EGFP in each organ derived from Ad5.CMV-injected mice ( $n=4$  for all organs) versus that from Ad.MBP.CMV-injected mice ( $n=10$  for heart, kidney, muscle, small bowel, and brain;  $n=7$  for lung, pancreas, and large bowel). Bar

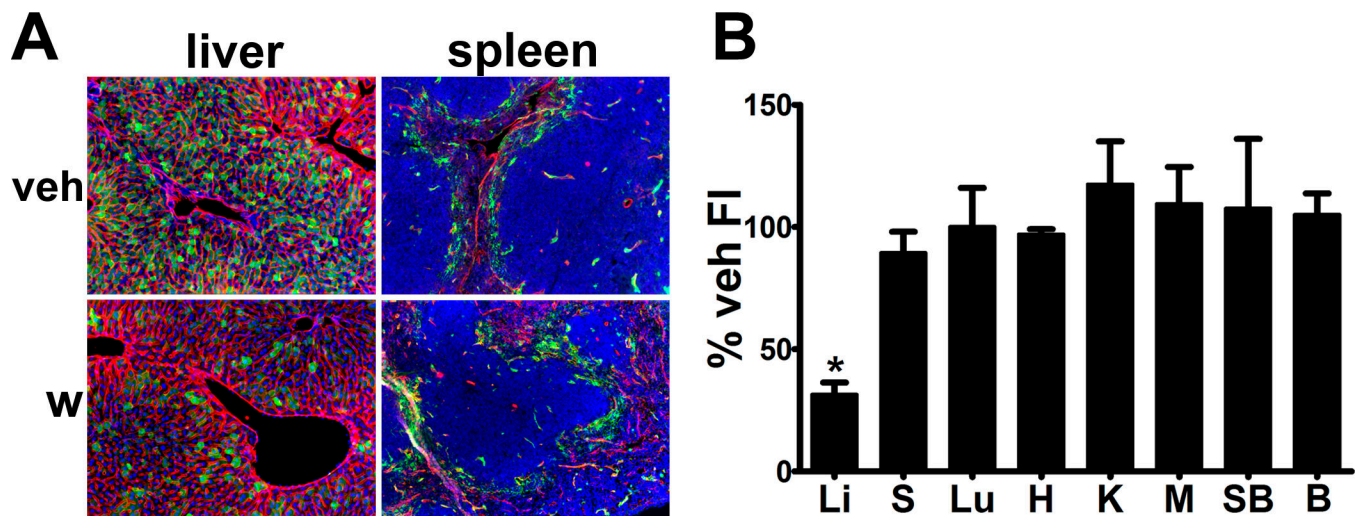
graph is mean  $\pm$  standard deviation asterisk: adjusted  $p < 0.05$ . Magnification: 100 $\times$ , Red: endomucin/CD31, Green: EGFP immunofluorescence, Blue: DAPI. Li: liver, S: spleen, Lu: lung, H: heart, K: kidney, M: muscle, P: pancreas, SB: small bowel, LB: large bowel, B: brain.

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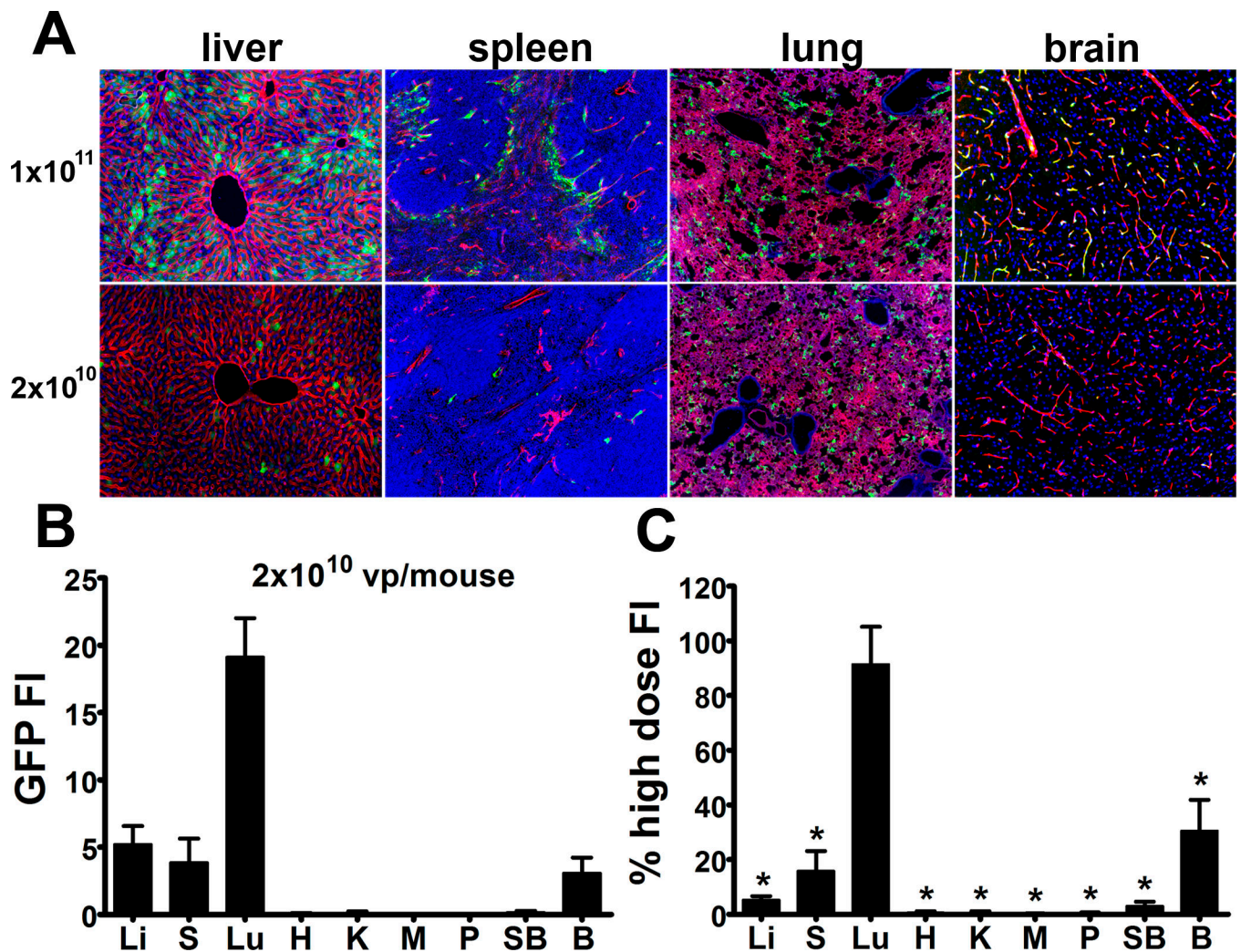
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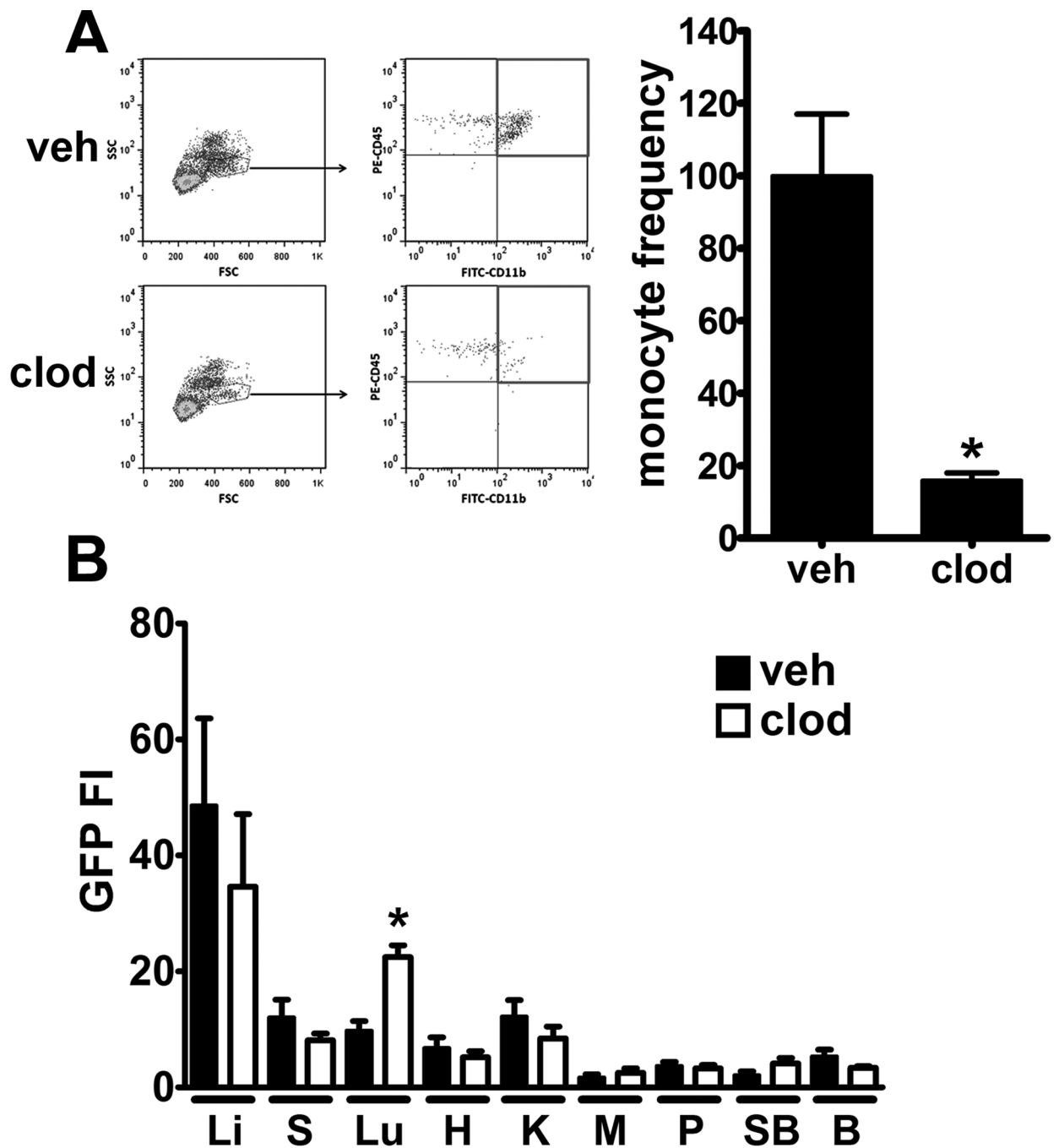
**Figure 2. Warfarin pretreatment reduced Ad.MBP.CMV liver tropism but did not alter gene expression in other host organs**

(A) Warfarin, 5 mg/kg, on day -3 and -1 before vector injection diminished hepatocyte expression but did not change transgene expression in spleen. (B) EGFP fluorescence per  $\mu\text{m}^2$  of tissue area in each organ derived from warfarin-treated mice ( $n=3$  for all organs) normalized as percentage of the mean value of vehicle-treated or untreated counterparts ( $n=10$  for liver, spleen, heart, kidney, muscle, small bowel, and brain;  $n=7$  for lung) with standard deviation. Warfarin pretreatment reduced vector liver expression by 68% (Li) but did not lead to a significant change in gene expression in spleen (S), lung (Lu), heart (H), kidney (K), muscle (M), small bowel (SB), or brain (B). Asterisk indicates adjusted  $p < 0.05$ . Magnification: 100 $\times$ , Red: CD31/endomucin, Green: EGFP immunofluorescence, Blue: DAPI.



**Figure 3. Systemic administration of a low dose of Ad.MBP.CMV into adult mice produced differential and non-linear reduction in gene expression in host organs**

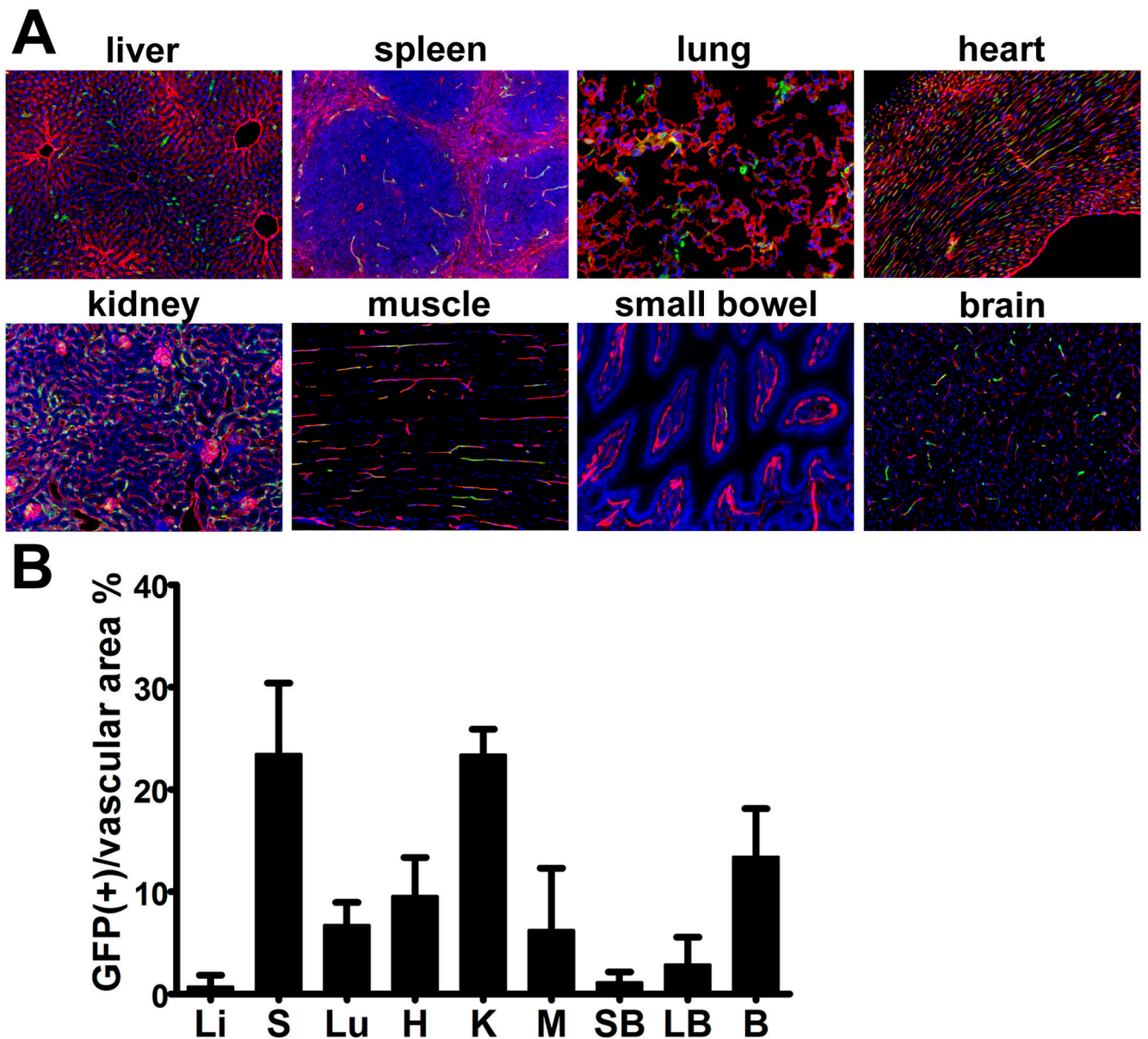
(A) EGFP expression in host liver, spleen, lung, and brain following intravenous injection of  $1 \times 10^{11}$  or  $2 \times 10^{10}$  vp of Ad.MBP.CMV into adult mice. Lowering vector dosage significantly reduced EGFP expression in vascular ECs of liver, spleen, and brain but did not change the expression in lung. (B) EGFP fluorescence per  $\mu\text{m}^2$  of tissue area in each organ derived from the low-dose group ( $n=6$  for each organ). (C) Normalization of the tissue EGFP fluorescence intensity values in (B) to the mean value of the high-dose counterparts. The spleen and brain EGFP expression in low-dose group was 16% and 31% of the high-dose counterparts. However, low virus dose drastically diminished the transgene expression in, heart, kidney, muscle, pancreas, and small bowel (3% of high-dose level). The low dose did not significantly alter the transgene expression in lung (91% of high-dose level). Asterisk indicates  $p < 0.05$ . Magnification:  $100\times$ , Red: endomucin/CD31, Green: EGFP immunofluorescence, Blue: DAPI. Li: liver, S: spleen, Lu: lung, H: heart, K: kidney, M: muscle, P: pancreas, SB: small bowel, B: brain.



**Figure 4. Depletion of circulating monocytes and hepatic and splenic macrophages lead to an increased Ad.MBP.CMV gene expression in the lung without a significant change in gene expression in other organs**

(A) Representative flow cytometry plots (left panel) quantifying the FSC-high/SSC-low/CD11b-positive/CD45-positive monocyte population in circulation. Relative frequency (right panel) of circulating monocytes from clodronate liposome-treated mice (clod, n=3) versus saline-treated mice (veh, n=3). Intravenous injection of clodronate liposomes depleted circulating CD11b-positive cells by 84%. (B) EGFP fluorescence per  $\mu\text{m}^2$  of tissue area in each organ derived from the saline-injected mice (n= 7 for liver, spleen, heart,

kidney, muscle, pancreas, small bowel, and brain; n=4 for lung) versus clodronate-liposome-injected mice (n=8 for liver, spleen, heart, kidney, pancreas, small bowel, and brain; n=7 for muscle; n= 5 for lung). Intravenous clodronate increased Ad.MBP.CMV lung expression by 2-fold (Lu) but did not result in a significant change in gene expression in liver (Li), spleen (S), heart (H), kidney (K), muscle (M), pancreas (P), small bowel (SB), or brain (B). Asterisk indicates adjusted  $p < 0.05$ .



**Figure 5. Ad.MBP.ROBO4 detargeted hepatocyte expression but reduced levels of vascular EC expression in other host organs**

(A) EGFP expression following intravenous injection of  $1 \times 10^{11}$  vp of Ad.MBP. ROBO4 into adult mice. Ad.MBP.ROBO4 yielded punctate vascular EC expression in liver but showed a reduced targeting efficiency to vascular ECs in spleen, lung, heart, kidney, muscle, small bowel, and brain. (B) The EGFP-positive vascular area analysis was performed as shown in Figure 1C. Magnification: 100 $\times$ , Red: endomucin/CD31, Green: EGFP immunofluorescence, Blue: DAPI. Li: liver, S: spleen, Lu: lung, H: heart, K: kidney, M: muscle, SB: small bowel, LB: large bowel, B: brain.