



ORIGINAL ARTICLE

Re-biopsy and liquid biopsy for patients with non-small cell lung cancer after EGFR-tyrosine kinase inhibitor failure

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Introduction

In China, lung cancer is the most frequently diagnosed cancer and the leading cause of cancer-related mortality.¹ The five-year survival rate in patients with lung cancer varies from 3.7–32.9%, depending on stage and regional differences.² Non-small cell lung cancer (NSCLC) accounts for 80–85% of lung cancers.³ Personalized therapeutic

Abstract

Background: Re-biopsy is important for exploring resistance mechanisms, especially for non-small cell lung cancer (NSCLC) patients who develop resistance to EGFR-tyrosine kinase inhibitors (TKIs). Liquid biopsy using circulating tumor DNA has come into use for this purpose. This retrospective study investigated the status of re-biopsy and liquid biopsy in NSCLC patients with *EGFR* mutations and evaluated their effect on clinical strategies and prognosis.

Methods: Five hundred fifty-five NSCLC patients with resistance to EGFR-TKIs were included and divided into three groups: re-biopsy, liquid biopsy, and no re-biopsy. Amplification refractory mutation system (ARMS) PCR or super ARMS PCR was used to detect *EGFR* mutations.

Results: Three hundred eight (55.5%) patients underwent re-biopsy; 45.5% (140/308) were positive for T790M. The most common re-biopsy procedure was computed tomography-guided percutaneous core needle biopsy (60.1%), followed by effusion drainage (29.5%) and superficial lymph node biopsy (6.5%). One hundred eighteen (21.3%) patients underwent liquid biopsy; the T790M detection rate was 41.5% (49/118.) Of the 308 patients who underwent re-biopsy, 69 were examined for *EGFR* mutations with plasma. The concordance rate of T790M detection between tissue and plasma was 66.7%. A statistical difference in further treatment after EGFR-TKI failure was observed among all groups ($P = 0.014$). Patients in the biopsy groups were more likely to receive third-generation EGFR-TKIs. Multivariate analysis showed that re-biopsy had a significant impact on overall survival ($P < 0.001$).

Conclusion: Re-biopsy plays a pivotal role in the management of patients with NSCLC and resistance to EGFR-TKIs. Liquid biopsy may be an alternative if difficulties performing re-biopsy exist.

strategies have dramatically improved the clinical outcome of patients with advanced NSCLC in recent years.^{4–6} Activating *EGFR* mutations are the most common therapeutically tractable driver mutation in lung adenocarcinomas with distinct ethnic differences, occurring at higher frequencies in Asian (40–60%) compared to Caucasian populations (7–10%).^{7–10} Many clinical trials have proven that EGFR-tyrosine kinase inhibitors (TKIs) yield a

superior response and more acceptable toxicity than traditional chemotherapy in advanced NSCLC patients with *EGFR* mutations; however patients inevitably develop resistance after achieving 9–13 months of progression-free survival (PFS).^{11–14} Acquired resistance to EGFR-TKIs is mainly attributed to secondary *EGFR* mutations.^{15–17} *EGFR* T790M, a specific point mutation in exon 20, is the most common resistance mutation, and accounts for 33–63%.^{18–24} Third-generation EGFR-TKIs (3-TKIs), such as osimertinib, have proven to have promising activity in advanced NSCLC patients with *EGFR* T790M mutations.^{25–27} Based on these facts, repeat biopsy plays an important role in clinical application for exploring resistance mechanisms and determining further therapeutic strategies.^{28,29} However, obtaining tissue samples from patients for repeat molecular analysis after EGFR-TKI failure remains a challenge. In the present study, we evaluated the current status of re-biopsy and liquid biopsy and their effect on clinical strategies and prognosis in Chinese NSCLC patients with *EGFR* mutations after EGFR-TKI failure.

Methods

Patients

We retrospectively reviewed the medical records of NSCLC patients with *EGFR* sensitive mutations, including 19deletions (19del), L858R, G719X, L861Q, and S768I, treated at the Shanghai Pulmonary Hospital between October 2011 and October 2017. Patients without *EGFR* mutations, or with *EGFR* 20 insert, de novo T790M, or known concomitant gene mutations, were excluded. Patients who were lost follow-up before confirmation of progressive disease (PD) from EGFR-TKIs were also excluded. Patients were divided into two groups according to the method of mutation testing: the rebiopsy group included patients who underwent tissue or cytologic sampling, while the liquid biopsy group included patients who underwent mutation testing with circulating tumor DNA (Fig 1). A control group of patients who did not undergo re-biopsy was also included. If mutation had been tested with tissues and plasma, the tissue results were regarded as standard. Progression-free survival (PFS) was defined as the interval from the initiation of further treatment after EGFR-TKI failure to first confirmed PD according to Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1. Overall survival (OS) was defined as the interval from diagnosis to death from any cause or to 31 December 2018. The primary objective was to evaluate the status of re-biopsy in Chinese patients including frequencies, procedure results, and impact on follow-up treatment strategies. The potential application of plasma-testing methodologies was also

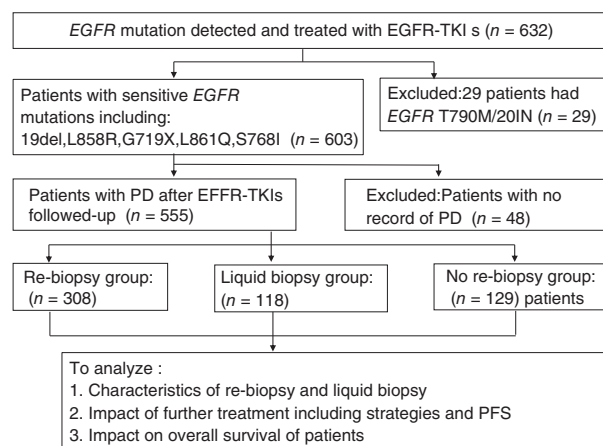


Figure 1 Flow chart of subject enrollment. PFS, progression-free survival; PD, progressive disease; TKIs, tyrosine kinase inhibitors.

assessed. The second objective was to explore whether patients that underwent re-biopsy achieved a better clinical outcome. The need for written informed consent was waived because of the retrospective design of the study. The institutional review board of Shanghai Pulmonary Hospital approved the study.

EGFR mutation analysis

Amplification refractory mutation system (ARMS) PCR was used to detect *EGFR* mutations. Our hospital adopted the ARMS-PCR method on 1 May 2017, thus mutation analysis of plasma was tested by ARMS-PCR before 1 May 2017 and by super ARMS-PCR thereafter. Tissue or cytologic specimens were frozen within 30 minutes of sampling and stored at -80°C . Frozen samples were thawed at 37°C , refrozen rapidly to disrupt the cells, and divided into equal aliquots before DNA was extracted using standard procedures according to *EGFR* mutation test kit instructions.

Statistical analysis

Statistical analyses were performed using SPSS version 19.0. Intergroup comparisons were performed using the Mann–Whitney *U* test for continuous variables, and Pearson's χ^2 or Fisher's exact tests for categorical variables. Multivariate analysis was performed to determine the feasibility of re-biopsy using multinomial logistic regression. The Kaplan–Meier method was used to estimate OS and PFS. Univariate differences in survival were assessed using the stratified log-rank method. Hazard ratios (HRs) for PFS and OS and multivariate differences in survival were estimated using a stratified Cox model. $P < 0.05$ was considered significant.

Results

Patient characteristics

Baseline individual characteristics were retrospectively reviewed in 555 patients with NSCLC after EGFR-TKI failure (Table 1). The median age was 60 years (range: 53–67), with women comprising 59.3% of the total. Adenocarcinoma (89.4%) was the most common histology. Of the total patients, 292 (52.6%) had 19del, 245 (44.1%) had L858R, 11 (2.0%) had uncommon mutations, and 7 (1.3%) had double sensitive mutations. The EGFR-TKIs used were gefitinib (62.0%), erlotinib (18.6%), icotinib (17.5%), and afatinib (2.0%); 73.9% patients were administered EGFR-TKIs at first-line.

Status of re-biopsy and liquid biopsy

A total of 308 (55.5%) patients underwent re-biopsy procedures, including: 185 (60.1%) computed tomography (CT)-guided percutaneous core needle biopsy (PCNB); 91 (29.5%) effusion drainage (including 81 pleural effusion, 5 pericardial effusion, and 5 peritoneal effusion); 20 (6.5%) superficial lymph node biopsy; and 7 (2.3%) transbronchial needle aspiration (TBNA) or endobronchial ultrasonography (EBUS)-guided TBNA (Fig 2a). Of the 308 patients, 134 (43.5%) were positive for *EGFR* T790M, 134 (43.5%) patients only harbored their initial mutation, and 25 (8.1%) patients were wild type (Fig 2b). A total of 118 (21.3%) patients underwent liquid biopsy: 49 (41.5%)

Table 1 Baseline characteristics of patients

Characteristics	All (n = 588)	Univariate			P	Multivariate			
		Re-biopsy	Liquid biopsy	No biopsy		Re-biopsy		Liquid biopsy	
						HR	P	HR	P
Age (years)						1.046	0.847	0.660	0.220
≤ 65	379 (68.3)	208 (67.5)	87 (73.7)	84 (65.1)	0.318				
> 65	176 (31.7)	100 (32.5)	31 (26.3)	45 (34.9)					
Gender						0.889	0.674	1.557	0.135
Male	226 (40.7)	126 (40.9)	50 (42.4)	50 (38.8)	0.842				
Female	329 (59.3)	182 (59.1)	68 (57.6)	79 (61.2)					
Smoking						0.893	0.731	1.066	0.875
Non-smoker	438 (78.9)	242 (78.6)	94 (79.7)	102 (79.1)	0.969				
Current or former smoker	117 (21.1)	66 (21.4)	24 (20.3)	27 (20.9)					
ECOG						0.728	0.365	0.907	0.824
0–1	494 (89.0)	268 (87.0)	111 (94.1)	115 (89.1)	0.114				
2–3	61 (11.0)	40 (13.0)	7 (5.9)	14 (10.9)					
Histology						0.986	0.969	1.854	0.219
Adenocarcinoma	496 (89.4)	276 (89.6)	105 (89.0)	115 (89.1)	0.978				
Non-adenocarcinoma	59 (10.6)	32 (10.4)	13 (11.0)	14 (10.9)					
Gene mutation type									
19del	292 (52.6)	178 (57.8)	54 (45.8)	60 (46.5)	0.096	4.306	0.130	0.955	0.964
L858R	245 (44.1)	121 (39.3)	59 (50.0)	65 (50.4)		3.090	0.241	1.304	0.795
Other	11 (2.0)	7 (2.3)	3 (2.5)	1 (0.8)		11.029	0.095	4.394	0.336
Double mutation	7 (1.3)	2 (0.6)	2 (1.7)	3 (2.3)					
EGFR-TKIs									
Gefitinib	344 (62.0)	198 (64.3)	72 (61.0)	74 (57.4)	0.63	1.729	0.476	1.025	0.977
Erlotinib	103 (18.6)	58 (18.8)	18 (15.3)	27 (20.9)		1.234	0.789	0.740	0.738
Icotinib	97 (17.5)	47 (15.3)	25 (21.2)	25 (19.4)		1.188	0.828	1.093	0.921
2-TKIs	11 (2.0)	5 (1.6)	3 (2.5)	3 (2.3)					
Line of EGFR-TKIs						0.752	0.279	1.208	0.570
1	410 (73.9)	221 (71.8)	93 (78.8)	96 (74.4)	0.328				
≥ 2	145 (26.1)	87 (28.2)	25 (21.2)	33 (25.6)					
Efficacy of EGFR-TKIs									
PR	277 (49.9)	163 (52.9)	70 (59.3)	44 (34.1)	<0.001	1.032	0.953	1.652	0.452
SD	214 (38.6)	112 (36.4)	42 (35.6)	60 (46.5)		0.507	0.205	0.698	0.592
PD	36 (6.5)	15 (4.9)	1 (0.8)	20 (15.5)		0.211	0.013	0.044	0.010
Not clear	28 (5.0)	18 (5.8)	5 (4.2)	5 (3.9)					

Non-adenocarcinoma included: 26 non-small cell lung cancer (NSCLC), 15 adenocarcinoma, 12 squamous carcinoma, 4 poorly differentiated adenocarcinoma, and 2 large cell lung cancer. Other included 5 G719X, 3 L861Q, and 3 S768I. Double mutation included five 19del and L858R, one 19 del and L861Q, and one S768I and G719X. ECOG, Eastern Cooperative Oncology Group; PD, progressive disease; PR, partial response; SD, stable disease; TKI, tyrosine kinase inhibitor.

patients were positive for T790M, 35 (29.7%) patients only harbored their initial mutation, and 32 (27.1%) patients were wild type (Fig 2c). Of the 308 patients who underwent re-biopsy, 69 patients were examined for *EGFR* mutation using plasma. The specificity of liquid biopsy for T790M detection was 84.4% and the sensitivity was 33.3%. The concordance of T790M detection between tissue and plasma was 66.7%. The prevalence of acquired T790M mutation was statistically different between patients with uncommon and common mutations (19del/L858R: 44.8% vs. 11.1%; $P = 0.04$), but further studies are required to verify these results, as our samples of patients with uncommon mutations were few. In addition, univariate and multivariate analysis showed that patients in the re-biopsy and liquid biopsy groups exhibited a better previous response to EGFR-TKIs compared to the control ($P < 0.05$) (Table 1).

Strategies and progression-free survival of further treatment

A total of 414 (74.6%) patients were followed-up after further treatment subsequent to EGFR-TKI failure. Pearson's χ^2 test revealed a statistical difference between the three groups ($P = 0.014$) (Table 2). Further analysis indicated that patients in the biopsy groups were more likely to

receive third-generation TKIs (re-biopsy 26.4% vs. liquid biopsy 31.4% vs. no re-biopsy 6.1%); 87.5% (56/64) of patients administered 3-TKIs were positive for T790M in the re-biopsy group and 85.7% in the liquid biopsy group (Table 2). However, overall only 52.3% (56/107) of patients positive for T790M received 3-TKIs in the re-biopsy and 75.0% (24/32) in the liquid biopsy group (Table 3).

Re-biopsy and liquid biopsy could impact further treatment strategies, as indicated above. To explore the specific impact, we analyzed PFS after further treatment. Univariate analysis proved that the previous efficacy of EGFR-TKIs had a significant impact on PFS ($P = 0.001$) (Table 4, Fig 3a). However, the difference in PFS among the three groups was not statistically significant ($P = 0.123$) (Table 4, Fig 3b). Considering that the deviation resulted from patients who were positive for T790M but did not receive 3-TKIs, we introduced a new variable – treatment-based result (TBR) – and further divided the patients into four groups according to whether T790M was positive and 3-TKIs were administered. Patients in the T790M + 3-TKI + group achieved significantly longer PFS than in other groups ($P < 0.001$) (Table 4, Fig 3c). Multivariate analysis confirmed that the previous efficacy of EGFR-TKIs and TBR were independent significant factors of PFS after further treatment (Table 4).

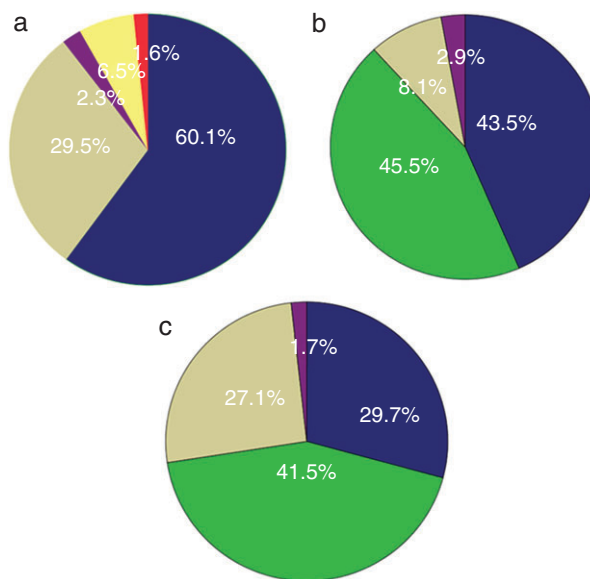


Figure 2 (a) The proportions of different sampling procedures in patients who underwent re-biopsy, (■) CT-guided PCNB, (■) Effusion drainage, (■) TBNA/EBUS-TBNA, (■) SLNB and (■) other metastasis biopsy. The distribution of results in patients who were tested for *EGFR* mutation via (b) tissue (c) or plasma. (■) Consistent with baseline, (■) Mut plus T790M/T790M, (■) Wild type, and (■) others. “Others” in (b) include three patients with small-cell lung cancer, two with baseline mutations and *c-MET*, one with an exon 20 insert, one with translation from 19del to L858R, one with *KRAS* and one with *ALK*. “Others” in (c) include one patient with an exon 20 insert and one with baseline mutations (L858R), S768I, and T790M. CT, computed tomography; EBUS, endobronchial ultrasonography; mut plus T790M, patients harbored a baseline mutation and a T790M mutation when re-tested; PCNB, percutaneous core needle biopsy; SLNB, superficial lymph node biopsy; TBNA, transbronchial needle aspiration; T790M, patients only harbored a T790M mutation and the baseline sensitive mutation disappeared.

Table 2 Further treatment after EGFR-TKI failure

Further treatment	Re-biopsy (n = 242)	Liquid biopsy (n = 90)	No re-biopsy (n = 82)	Total	P
3-TKIs	64 (26.4)	28 (31.1)	5 (6.1)	97 (23.4)	0.014
TKI + chemotherapy/radiotherapy	24 (9.9)	10 (11.1)	10 (12.2)	44 (10.6)	
Other TKIs	7 (2.9)	2 (2.4)	2 (2.4)	12 (2.9)	
Former TKIs	11 (4.5)	7 (7.8)	8 (9.8)	26 (6.3)	
Chemotherapy/radiotherapy	123 (50.8)	40 (44.4)	53 (64.6)	216 (52.2)	
Other	13 (5.4)	2 (2.2)	4 (4.9)	19 (4.6)	

Other tyrosine kinase inhibitors (TKIs) refers to other first or second-generation EGFR-TKIs; Other includes best supportive treatment, traditional Chinese medicine, crizotinib, etc. 3-TKIs, third-generation TKIs.

Table 3 Further treatment after EGFR-TKI failure according to biopsy results

Further treatment	Result of re-biopsy/liquid biopsy				Total	P
	Consistent with baseline	Mut plus T790M/T790M	Wild type	Others		
3-TKIs	6 (5.5)/2 (6.5)	56 (52.3)/24 (75.0)	1 (5.9)/1 (4.0)	1 (12.5)/1 (50.0)	64 (26.4)/28 (31.1)	<0.001/<0.001
TKI + chemotherapy/ radiotherapy	16 (14.5)/6 (19.4)	5 (4.7)/1 (3.1)	2 (11.8)/3 (12.0)	1 (12.5)/0 (0.0)	24 (9.9)/10 (11.1)	
Other TKIs	4 (3.6)/2 (6.5)	1 (0.9)/0 (0.0)	0 (0.0)/1 (4.0)	2 (25.0)/0 (0.0)	7 (2.9)/3 (3.3)	
Former TKIs	6 (5.5)/3 (9.7)	4 (3.7)/1 (3.1)	4 (9.5)/3 (12.0)	0 (0.0)/0 (0.0)	11 (4.5)/7 (7.8)	
Chemotherapy/ radiotherapy	73 (66.4)/17 (54.8)	35 (32.7)/5 (15.6)	12 (70.6)/17 (68.0)	3 (37.5)/0 (0.0)	123 (50.8)/40 (44.4)	
Others	5 (4.5)/1 (3.2)	6 (5.6)/1 (3.1)	1 (5.9)/0 (0.0)	1 (12.5)/0 (0.0)	13 (5.4)/2 (2.2)	

Other tyrosine kinase inhibitors (TKIs) refers to other first or second-generation EGFR-TKIs; Other includes best supportive treatment, traditional Chinese medicine, crizotinib, etc. 3-TKIs, third-generation TKIs.

Table 4 Univariate and multivariate analyses of clinical parameters of PFS of further treatment

Factors	Univariate analysis			Multivariate analysis		
	HR	95% CI	P	HR	95% CI	P
Gender (female/male)	0.881	0.773–1.004	0.057	0.882	0.641–1.214	0.441
Age (< 65/≥ 65)	1.308	0.989–1.730	0.060	1.130	0.847–1.507	0.407
Smoking (never/ever)	0.700	0.512–0.957	0.025	0.849	0.574–1.256	0.413
ECOG (0–1/2–3)	0.785	0.496–1.244	0.319			
Histology (adenocarcinoma/non-adenocarcinoma)	0.771	0.492–1.208	0.256			
EGFR mutation			0.511			
19del/double mutation	0.593	0.188–1.872	0.373			
L858R/double mutation	0.611	0.193–1.933	0.402			
Other/double mutation	1.129	0.252–5.055	0.874			
EGFR-TKIs			0.224			
Gefitinib/afatinib	0.552	0.203–1.502	0.245			
Erlotinib/afatinib	0.455	0.162–1.280	0.136			
Icotinib/afatinib	0.672	0.239–1.889	0.451			
Treatment line of EGFR-TKIs (1/≥ 2)	0.927	0.696–1.235	0.605			
Efficacy of EGFR-TKIs			0.001			0.005
PR/PD	0.336	0.188–0.603	0.000	0.392	0.216–0.712	0.002
SD/PD	0.410	0.227–0.740	0.003	0.500	0.273–0.916	0.025
If re-biopsied			0.123			
Re-biopsy/no re-biopsy	0.709	0.508–0.989	0.043			
Liquid biopsy/no re-biopsy	0.798	0.535–1.190	0.268			
TBR			0.000			0.002
T790M+ 3-TKI+/T790M-3-TKI-	0.467	0.331–0.659	0.000	0.520	0.364–0.745	0.000
T790M + 3-TKI-/T790M-3-TKI-	1.041	0.704–1.542	0.839	1.108	0.741–1.657	0.618
T790M-3-TKI+/T790M-3-TKI-	1.053	0.516–2.146	0.888	1.212	0.590–2.490	0.600

ECOG, Eastern Cooperative Oncology Group; PD, progressive disease; PFS, progression-free survival; PR, partial response; SD, stable disease; TBR treatment-based result; TKI, tyrosine kinase inhibitor.

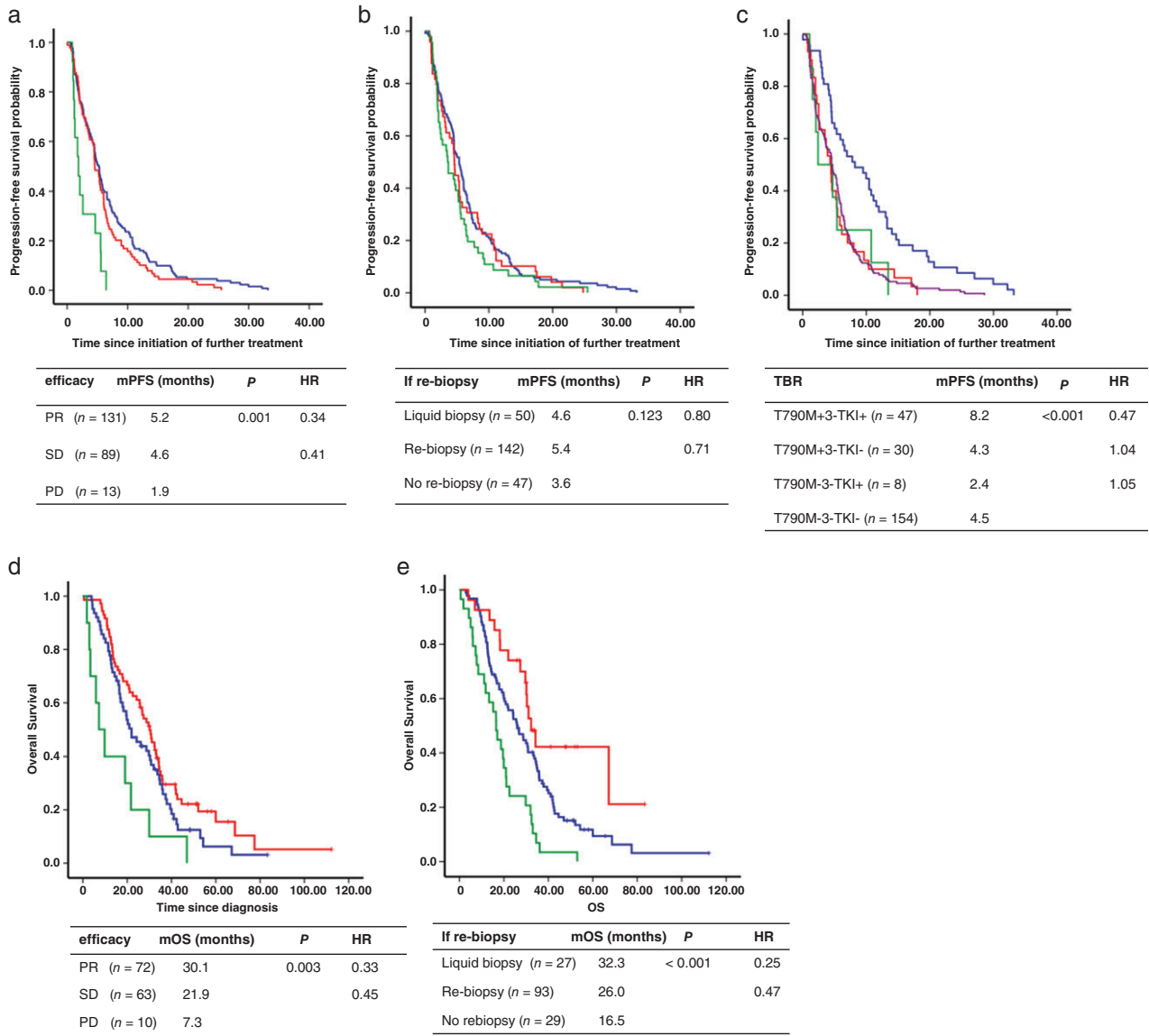


Figure 3 Kaplan–Meier estimates of the duration of progression-free survival (PFS) of further treatment in (a) patients who exhibited an objective response to EGFR-tyrosine kinase inhibitors (TKIs), efficacy (—) PR, (—) SD and (—) PD, (b) patients who underwent re-biopsy or not, if re-biopsy (—) re-biopsy, (—) liquid biopsy and (—) no biopsy and (c) patients divided into different groups according to whether T790M was positive and third-generation TKIs (3-TKIs) were administered, TBR (—) T790M +3-TKI+, (—) T790M +3-TKI-, (—) T790M -3-TKI+ and (—) T790M -3-TKI-. Kaplan–Meier estimates of the duration of overall survival (OS) of (d) patients who exhibited an objective response to EGFR-TKIs, efficacy (—) PR, (—) SD, (—) PD, (—) PR-censored, (—) SD-censored and (—) PD-censored and (e) patients who underwent re-biopsy or not, if re-biopsy (—) re-biopsy, (—) liquid biopsy, (—) no rebiopsy, (—) re-biopsy-censored, (—) liquid biopsy-censored and (—) no rebiopsy-censored. HR, hazard ratio; mOS, median OS; mPFS, median PFS; mut plus T790M, patients harbored a baseline mutation and a T790M mutation when re-tested; PD, progressive disease; PR, partial response; SD, stable disease; T790M, patients only harbored a T790M mutation and the baseline sensitive mutation disappeared.

Overall survival of patients

Overall survival was analyzed in 149 patients and was longer in patients with a previous response to EGFR-TKIs than those with PD (partial response [PR] vs. stable disease [SD] vs. PD: 30.1 vs. 20.9 vs. 7.3 months, respectively; HR 0.33 for PR vs. PD, HR 0.45 for SD vs. PD; *P* = 0.003)

(Fig 3d). The OS of patients in the liquid and re-biopsy groups was superior to patients in the no re-biopsy group (32.3 vs. 26.0 vs. 16.5 months, respectively; HR 0.25 for liquid biopsy vs. no re-biopsy, HR 0.47 for re-biopsy vs. no re-biopsy; *P* < 0.001) (Fig 3e). Multivariate analysis revealed that previous efficacy to EGFR-TKIs and re-biopsy

Table 5 Univariate and multivariate analyses of clinical parameters of OS

Factors	Univariate analysis			Multivariate analysis		
	HR	95% CI	<i>P</i>	HR	95% CI	<i>P</i>
Gender (female/male)	1.002	0.702–1.430	0.991			
Age (< 65/≥ 65)	1.027	0.697–1.514	0.892			
Smoking (never/ever)	0.872	0.580–1.310	0.509			
ECOG (0–1/2–3)	0.604	0.374–0.977	0.040	0.598	0.364–0.980	0.042
Histology (adenocarcinoma/non-adenocarcinoma)	0.962	0.568–1.629	0.884			
EGFR mutation			0.278			
19del/double mutation	1.273	0.460–3.523	0.642			
L858R/double mutation	1.655	0.597–4.591	0.333			
Other/double mutation	2.827	0.624–12.813	0.178			
EGFR-TKIs			0.097			0.029
Gefitinib/2-TKIs	7.595	1.045–55.222	0.045	6.179	0.833–45.820	0.075
Erlotinib/2-TKIs	6.225	0.849–45.635	0.072	4.024	0.538–30.118	0.175
Icotinib/2-TKIs	4.221	0.566–36.114	0.155	2.882	0.345–24.111	0.329
Treatment line of EGFR-TKIs (1/≥ 2)	1.185	0.824–1.704	0.359			
Efficacy of EGFR-TKIs			0.005			0.002
PR/PD	0.334	0.170–0.658	0.002	0.276	0.134–0.569	0.000
SD/PD	0.454	0.231–0.894	0.022	0.319	0.153–0.664	0.002
If re-biopsied			<0.001			0.000
Re-biopsy/no re-biopsy	0.466	0.300–0.725	0.001	0.358	0.220–0.582	0.000
Liquid biopsy/no re-biopsy	0.276	0.146–0.520	<0.001	0.279	0.145–0.535	0.000
TBR			0.450			
T790M+ 3-TKI+/T790M-3-TKI-	0.707	0.407–1.229	0.219			
T790M+ 3-TKI-/T790M-3-TKI-	0.769	0.483–1.225	0.269			
T790M- 3-TKI+/T790M-3-TKI-	0.673	0.246–1.842	0.441			

Variables of *P* < 0.1 in univariate analysis were included into multivariate analysis. Treatment-based result (TBR): patients were divided into four groups according to whether T790M was positive and third-generation tyrosine kinase inhibitors (3-TKIs) were used. 2-TKIs, second-generation TKIs; ECOG, Eastern Cooperative Oncology Group; OS, overall survival; PD, progressive disease; PR, partial response; SD, stable disease.

had a significant impact on OS (Table 5). Multivariate analysis showed that patients with good performance status and those administered 2-TKIs achieved longer survival (Table 5).

Discussion

Re-biopsy has a pivotal role for exploring resistance mechanisms, particularly for patients with resistance to EGFR-TKIs. However, re-biopsy is not always feasible for either patients or physicians. Kawamura *et al.* reported that 120 patients (63%) in their sample had undergone re-biopsy,³⁰ while Ichihara *et al.* reported that 55 (53.9%) patients underwent re-biopsy.²¹ We analyzed re-biopsy status in 555 patients and the rate (55.5%) was similar to the reported data. CT-guided PCNB (60.1%) was the most common re-biopsy procedure used in our study sample, likely because of its safety and operability. The frequency of T790M mutations detected in patients who underwent re-biopsy was 45.5% in the clinical setting at our institution, which is also consistent with previous studies (range: 33–63%).^{18–24} The remaining patients did not undergo re-biopsy because of inaccessible tumors (*n* = 27),

deterioration in performance status (*n* = 21), patient refusal (*n* = 124), or unknown reasons (*n* = 75).

Liquid biopsy has shown promising advantages of non-invasiveness and accessibility. Some studies have suggested that a plasma-based test is a useful method for noninvasive assessment and monitoring of T790M resistance mutations.^{31–33} Our results indicate that detection of T790M in plasma using the super ARMS-PCR method exhibits low sensitivity (33.3%) but high specificity (84.4%), with a satisfactory T790M detection rate (41.5%). The low sensitivity was attributed to the fact that 83 (70.3%) patients were tested by ARMS-PCR. Moreover, these results suggest that re-biopsy and liquid biopsy could provide guidance for further treatment strategies. As expected, patients who harbor T790M are more inclined to receive 3-TKIs.

As the development of new agents has improved the overall survival duration of patients with malignant tumors, management of cancer patients throughout the entire course of disease has become a focus. As an essential part of the process, whether re-biopsy could improve the clinical outcomes of patients with cancer remains to be determined. To the best of our knowledge, our study is the first and the largest to report the relationship between re-biopsy and survival, including PFS of further

treatment and OS in patients with NSCLC after EGFR-TKI failure. As mentioned above, the previous efficacy of EGFR-TKIs is an independent significant factor affecting not only PFS of further treatment but also OS. This may be because tumors with primary resistance to EGFR-TKIs are genetically complex, with high heterogeneity and drug resistance.^{34,35} What is interesting is that our data suggest that re-biopsy has a significant impact on OS but not on PFS of further treatment, while TBR was a significant factor for PFS of further treatment but not for OS. The lack of a significant difference in PFS after further treatment among the three initial groups was attributed to the low proportion of patients positive for T790M that were administered 3-TKIs in the re-biopsy (23.1%) and liquid biopsy (26.7%) groups, but the very value of re-biopsy is to determine the resistance mechanism, on which basis physicians can choose the appropriate therapy, thus the TBR variable was applied. However, any type of treatment a patient is administered can affect OS, which is why re-biopsy is important. Not all of the patients identified with T790M were administered 3-TKIs as further treatment after EGFR-TKI failure, but they were more likely to receive subsequent treatment. Thus, patients who underwent rebiopsy were administered more appropriate subsequent therapies and achieved longer OS.

This study is limited by its retrospective nature and that our cohort was from a single institution. We used ARMs and super ARMS to test *EGFR* mutation in tissues and plasma, which has limited sensitivity. Although we used a large sample overall, the sample sizes of some of the subgroups were small. Our results require further verification in prospective studies.

In conclusion, re-biopsy plays a pivotal role in the management of patients with NSCLC and resistance to EGFR-TKIs and improves the clinical outcome to some extent. Liquid biopsy may be an alternative if difficulties performing re-biopsy exist.

Disclosure

No authors report any conflict of interest.

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