



Review

# Past HIV-1 Medications and the Current Status of Combined Antiretroviral Therapy Options for HIV-1 Patients

Matthew Weichseldorfer <sup>1</sup>, Marvin Reitz <sup>2</sup> and Olga S. Latinovic <sup>1,3,\*</sup>

<sup>1</sup> Institute of Human Virology, School of Medicine, University of Maryland, Baltimore, MD 21201, USA; mweichseldorfer@ihv.umaryland.edu

<sup>2</sup> Department of Medicine, School of Medicine, University of Maryland, Baltimore, MD 21201, USA; MReitz@ihv.umaryland.edu

<sup>3</sup> Department of Microbiology and Immunology, School of Medicine, University of Maryland, Baltimore, MD 21201, USA

\* Correspondence: olatinovic@ihv.umaryland.edu

**Abstract:** Combined antiretroviral therapy (cART) is treatment with a combination of several antiretroviral drugs that block multiple stages in the virus replication cycle. An estimated 60% of the 38 million HIV-1 patients globally receive some form of cART. The benefits of cART for controlling HIV-1 replication, transmission, and infection rates have led to its universal recommendation. Implementation has caused a substantial reduction in morbidity and mortality of persons living with HIV-1/AIDS (PLWHA). More specifically, standard cART has provided controlled, undetectable levels of viremia, high treatment efficacy, reduction in pill burden, and an improved lifestyle in HIV-1 patients overall. However, HIV-1 patients living with AIDS (HPLA) generally show high viral loads upon cART interruption. Latently infected resting CD4+ T cells remain a major barrier to curing infected patients on long-term cART. There is a critical need for more effective compounds and therapies that not only potently reactivate latently infected cells, but also lead to the death of these reactivated cells. Efforts are ongoing to better control ongoing viral propagation, including the identification of appropriate animal models that best mimic HIV-1 pathogenesis, before proceeding with clinical trials. Limited toxicity profiles, improved drug penetration to certain tissues, and extended-release formulations are needed to cover gaps in existing HIV-1 treatment options. This review will cover past, current, and new cART strategies recently approved or in ongoing development.

**Keywords:** HIV-1; AIDS; cART; entry inhibitors; LRA; reverse transcriptase inhibitors; protease inhibitors; integrase inhibitors; HIV-1 latency



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## 1. Introduction

HIV-1 establishes a stably integrated, non-productive latent state of infection of individual cells, mainly long lived CD4+ T cells that are maintained by homeostatic proliferation [1,2]. This latent and stable reservoir is a primary barrier to HIV-1 eradication, despite the evident success of combined antiretroviral therapy (cART) [3]. cART effectively silences HIV-1 replication, but the persistence of latent reservoirs in the myeloid [4,5] and T cells of patients makes HIV-1 infection incurable [3,6–8]. HIV-1 can also replicate in brain microglial cells, which persist despite cART [9,10]. Honeycutt et al. [11] reported that integrated HIV-1 DNA is present in human bone marrow and spleen macrophages even after cART, and that mice with only human myeloid cells allow persistent infection in macrophages during cART in vivo.

The Department of Health and Human Services (DHHS) guidelines on antiretroviral agents for PLWHA encourages treatment for everyone who wants treatment and understands its importance [12]. The WHO has recommended that all PLWHA take cART regardless of their clinical status or CD4+ cell numbers, and the WHO supports promptly starting cART treatment of PLWHA, including administering cART on the same day as the

diagnosis [13]. The DHHS guidelines suggest cART with two nucleoside reverse transcriptase inhibitors (NRTIs) in combination with a third active drug for treatment-naïve HIV positive individuals with completely susceptible virus [14]. The third active drug of choice consists of an integrase strand transfer inhibitor (INSTI).

## 2. First AIDS Drug—AZT

The first AIDS drug, zidovudine (AZT, HIV-1 reverse transcriptase inhibitor (RTI [15])) was released almost five years after the discovery of HIV-1 [15,16]. Upon approval by the US Food and Drug Administration (FDA) in March 1987, AZT-based monotherapy provided the US public confidence that AIDS, considered a death sentence at the time, could be relatively controlled. The lack of major success by AZT (due to drug toxicity causing severe anemia and liver problems in HIV-1 patients [17–19]) was followed by a few additional drugs in the late eighties (Hivid (ddC, zalcitabine), Videx (ddl, didanosine), and Zerit (d4T, stavudine)). All failed to obtain long lasting control of viremia because all had similar mechanisms of action and targeted only one step of the virus replication cycle. An additional issue was the requirement for multiple daily doses, which created complex dosing schedules. A shift toward targeting HIV-1 protease and integrase led to the design of a novel class of antiviral drugs, namely PIs, including Invirase (saquinavir), and the first combined antiretroviral therapy in the 1990s [20,21].

## 3. Highly Active Antiretroviral Therapy—HAART

The impact of highly active antiretroviral therapy (HAART) was evident by the time of the 1996 International AIDS Conference in Vancouver, when HIV-1 mortality rates started to resemble general mortality rates and viral loads became undetectable, largely putting the disease into remission [22]. HAART [23], through the combination of NRTIs, NNRTIs, and PIs, thus, significantly changed the progression and outcome of infection with HIV-1 [24]. The success of HAART changed this originally fatal disease into a treatable chronic infection. For example, a 20-year-old HIV-1 patient on HAART could optimally live into their 50s. By the early 2000s, however, there were obstacles in the use of HAART. High pill burdens, inconvenient dosing, and long-term toxicities contributed to poor compliance and the emergence of drug-resistant virus in many patients [25–27]. For those patients in whom antiviral drug-resistance developed, treatment options become limited and more complicated regimens were necessary to prevent further disease progression. PIs caused insulin resistance, cardiac arrhythmias, unbalanced redistribution of body fat (lipodystrophy), and required three daily intakes. Lactic acidosis [28] and peripheral neuropathy were caused by the use of NRTIs. cART regimens generally consist of a “backbone” of two NRTIs and a “base” of either a PI or NNRTI [28].

## 4. Combined Antiretroviral Therapy—cART

By the mid-2000s, new generations of drugs such as Viread (Tenofovir disoproxil fumarate) showed better antiretroviral performance, safer and longer-lasting regimens, and far fewer side effects with only one daily intake [29]. With this development, HIV-1/AIDS further became a chronic and manageable disease. For example, a 20-year-old HIV-1 patient under cART could optimally live into their 70s [30]. By the late 2000s, the elucidation of HIV-1 entry steps offered new opportunities for therapeutic intervention [31–34]. Two entry inhibitors were licensed in the mid-to-late 2000s, the fusion inhibitor enfuvirtide (T-20) [35] and the small-molecule CCR5 antagonist Maraviroc (MVC) [31,36,37]. For HIV-1 patients resistant to conventional drugs from the NRTI and PI classes, the entry and integration inhibitors approved in 2007 effectively suppressed HIV-1 while offering additional therapeutic options. Entry inhibitors are particularly attractive antiretrovirals because, unlike conventional antiretrovirals, they target HIV-1 extracellularly, thereby sparing cells from both viral- and drug-induced toxicities. In the following sections, some of the current antiretroviral options (Table 1) will be discussed. Table 1 contains a list of the current leading drugs among different antiretroviral groups (Entry Inhibitors, Integrase Strand

Transfer Inhibitors (INSTIs), Nucleoside Reverse Transcriptase Inhibitors (NRTIs), anti-CD4 Monoclonal Antibodies, Nucleoside Reverse Transcriptase Translocation Inhibitors (NRTTIs), Capsid Inhibitors (CAIs), Attachment Inhibitors that bind gp120 on HIV-1, and Latency-Reversing Agents (LRAs) as well as their phase of current development.

**Table 1.** Leading drug options among anti-HIV groups of drugs. Information was retrieved from the NIH US National Library of Medicine (<https://www.clinicaltrials.gov/ct2/home> accessed on 4 June 2021).

Antiretroviral Drug Group	Leading Drug Option	Phase of Current Development	FDA Approval Status	Route of Administration in HIV-1 Patients
	Enfuvirtide (T-20)	IV Completed	Yes	Oral
	Maraviroc	IV Completed	Yes	Oral
	Zinc-Finger Nuclease	II Completed	No	Infusion
<b>Attachment Inhibitors</b>	Fostemsavir (Rukobia)	III Active	Yes	Oral
<b>Anti-CD4 Monoclonal Antibodies</b>	Ibalizumab	III Completed	Yes	Infusion
<b>Nucleoside Reverse Transcriptase Translocation Inhibitors (NRTTI)</b>	Islatravir	I Completed, II Active	No	Oral
<b>Integrase Strand Transfer Inhibitors (INSTI)</b>	Dolutegravir	IV Completed	Yes	Oral
<b>Capsid Inhibitors</b>	GS-6207	Ib Completed, II/III Active	No	Oral/Subcutaneous
<b>Latency-Reversing Agents (LRAs)</b>	Romidepsin	II Completed	No	Infusion

## 5. gp120-Binding Proteins Inhibit HIV-1 Infection

Several HIV entry inhibitors that target HIV Env gp120 and gp41 are potent HIV inhibitors [38]. These include protein-based inhibitors such as soluble CD4 protein (sCD4) [39], eCD4-IgG [40], and antibodies against gp120, N6 [41], and m36.4 [42]. sCD4 targets the CD4-binding site (CD4bs) of gp120 and inhibits HIV entry and infection [39]. Schiavone et al., 2012 [43], designed a peptide mimotope of the HIV-1 gp120 bridging sheet. Their data validated the peptide mimotope approach as a promising tool to obtain an effective HIV-1 vaccine. Most recent studies suggest a potential to expand the previous strategy of combining a gp120-binding protein and a gp41-binding antibody for the treatment of HIV-1 infection [44]. Wang et al., 2021 [44] (originally identified a gp120-binding protein, mD1.22 as an inactivator of laboratory-adapted HIV-1) found that a gp41 N-terminal heptad repeat (NHR)-binding antibody D5 single-chain variable fragment (scFv) alone cannot inactivate HIV-1, even at high concentrations. However, a combination with D5scFv resulted in an enhanced inactivation activity of mD1.22 against divergent HIV-1 strains. These strains include primary HIV-1 isolates, T20- and AZT-resistant strains, HIV-1 laboratory-adapted strains, and LRA-reactivated virions. The authors [44], demonstrated that combining mD1.22 and D5 scFv gave synergistic inhibition of divergent HIV-1 strains.

## 6. Entry Inhibitors

T-20 and MVC are the first entry inhibitors licensed for patients with drug-resistant HIV-1, with MVC restricted to those infected with CCR5-tropic HIV-1 (R5 HIV-1) only. In addition, given that the oral administration of MVC offers high drug levels in cervicovaginal fluid, combinations of MVC and other CCR5 inhibitors have shown high effectiveness in preventing HIV-1 transmission. In addition, CCR5 antagonists prevent rejection of transplanted organs; therefore, MVC suppresses HIV-1 and prolongs organ survival for the growing number of HIV-1 patients with kidney or liver failure necessitating organ transplantation. Thus, MVC offers an important treatment option for HIV-1 patients with drug-resistant R5 HIV-1, who presently account for >40% of drug-resistance cases [45,46].

## 7. Integrase Strand Transfer Inhibitor (INSTI), Dolutegravir (DTG), and Nucleoside Reverse Transcriptase Inhibitor, Lamivudine

The most recent 2018 WHO recommendations list dolutegravir with an NRTI backbone as the preferred first-line ART regimen (WHO from review). This drug belongs to the INSTI class and has high tolerance to resistance. It is commonly recommended by the WHO as the first- and second-line agent for PLWHA [13] in recent triple drug cART, due to DTG's better antiretroviral efficacy over raltegravir (RAL) [47,48]. In treatment-experienced patients, cART regimens based on once-daily DTG showed greater viral suppression when compared to twice-daily RAL (71% DTG versus 64% RAL) [47]. McAllister et al., 2017, showed that DTG with tenofovir disoproxil fumarate and FTC is an effective option for once daily post-exposure prophylaxis in men who have sex with men (MSM) [48]. In addition, DTG exhibits a higher barrier to resistance than RAL does. Additionally, it has a low interaction potential; therefore, there are no food restrictions [49–51]. Based on the most recent Guidelines for the Use of Antiretroviral Agents in Adults and Adolescents with HIV-1 [52], DTG has become a staple in combined and recommended regimens for most people with HIV-1; however, there are competing treatments such as bictegravir with emtricitabine and tenofovir alafenamide [53].

The first two-drug therapy (Juluca) was FDA approved in 2017 for certain HIV patients, followed by Dovato, which was FDA approved in 2019 for both treatment-experienced and treatment-naïve patients [54–56]. Dovato includes the new generation of integrase inhibitor (DTG) with the NRTI, lamivudine. The two-drug combination had fewer side effects than the standard triple drug therapy with the same effectiveness. Darunavir, the first non-INSTI antiretroviral treatment, was approved in 2011 as a single tablet [57]. In 2021, an injectable therapy, Cabenuva (cabotegravir and rilpivirine), was released [58]. Cabenuva is a new and complete prescription regimen and is used to treat HIV-1 infection in adults as a replacement for their current HIV-1 treatment when their healthcare provider determines that they meet certain requirements. Cabenuva is available as an injection that is administered once a month [58].

## 8. Available HIV-1 Treatment Options

### 8.1. Anti-CD4 Monoclonal Antibodies

Viral fusion and entry are primarily achieved by the gp120-gp41-CD4 complex undergoing multiple conformational changes. The first anti-CD4 monoclonal antibodies (mAbs) were introduced in the early 1990s. Ibalizumab is a humanized mAb that binds to the N-terminus of the second CD4 receptor's domain. It has 10-fold higher antiretroviral activity while neutralizing most of the 116 HIV strains with 50% infection suppression [59]. Of the patients examined, 43% had a detectable viral load measuring below 50 copies/mL after 6 weeks of treatment [60]. Resistance to Ibalizumab via loss of glycosylation sites in the envelope V5 loop was evident during phase Ib clinical trials in 80% of the patients experiencing virologic failure. The outcomes of a phase III trial (optimized background regimen in MDR HIV-1) secured US regulatory approval for MDR HIV-1 treatment.

### 8.2. Nucleoside Reverse Transcriptase Translocation Inhibitors (NRTTI)

Unlike NRTIs, the nucleoside reverse transcriptase translocation inhibitors have dual mechanisms of action. In combination with a 3'-hydroxyl group, a 4'-ethynyl group inhibits translocation, resulting in chain termination [61,62]. A first-in-class NRTTI, Islatravir, was used as a monotherapy in Phase I clinical trials with treatment-naïve HIV-1 patients, showing a mean 1.2 log<sub>10</sub> decline in HIV-1 RNA copies daily [63]. Islatravir is an investigational NRTTI currently being evaluated for different frequencies and doses for HIV-1 treatment as both a single drug and in combination with other antiretroviral drugs at the most recent Conference on Retroviruses and Opportunistic Infections, CROI 2021 [64]. The study demonstrates that the implant showed active drug concentrations above the pre-specified PK threshold at 12 weeks. The tested Islatravir doses in the study were 48, 52, and 56 mg. The implant was projected to provide drug concentrations likely above the

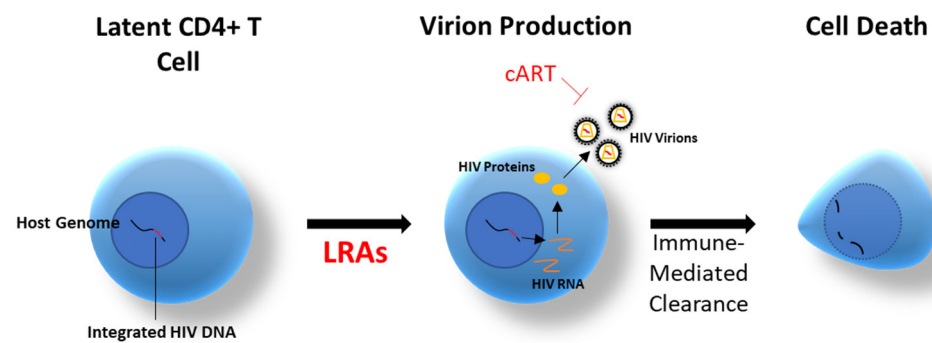
threshold for one year at the highest dose. Based on these findings, Merck plans to initiate a Phase 2 trial. They will explore the potential of a subdermal implant containing Islatravir as a long-acting pre-exposure prophylaxis (PrEP) option for up to a year. There is also an ongoing phase IIa randomized control trial to evaluate oral Islatravir as a PrEP option once monthly [65]. In addition, there is a double-blind randomized control study using Islatravir implants in two different doses, which is slightly different from the Merck study (54 and 62 mg), conducted with 16 healthy individuals for the purposes of evaluating the pharmacokinetics profile and tolerability of the Islatravir implants. This study showed that at 12 weeks, both implants reached above the target PK thresholds in one year. Adverse events were mild to moderate, with no clinical differences between the treatment and placebo experimental groups. The 62-milligram implant group reported higher pain than the 54-milligram implant group did [66].

### 8.3. Capsid Inhibitors

HIV-1 infection depends on the orderly formation and dissolution of the viral capsid. This dual role of the capsid protein in viral maturation and post-entry uncoating makes it a very attractive target of opportunity for antiretroviral therapies. The pre-integration complex (containing the viral DNA in the capsid core) is transported through the cell cytoplasm to the nucleus. The capsid protein is cleaved from Gag polyprotein precursors to form the capsid core during virion maturation. The capsid core undergoes an uncoating process, during which capsid hexamers disassemble after fusion between the virus membrane and the target cell membrane. The released reverse transcription complex is then transported to the cell nucleus. Numerous capsid protein inhibitors (CAIs) have been reported to block uncoating and HIV-1 infection. These include CAP-1, PF74, BI compounds, peptide inhibitors, Coumermycin A1, C1, Ebselen, anti-capsid antibodies, and GS-CA1. CAIs were first introduced at the Conference on Retroviruses and Opportunistic Infections, CROI 2017, as a first-in-class picomolar capsid inhibitor [67]. In 2017, GS-CA1, a highly potent capsid inhibitor, was described as holding promise for clinical development [58]. Although no CAI is currently approved for clinical use, GS-CA1 is a potent CAI that holds much potential for therapeutic development against HIV-1 [68]. CAI's main advantages are a high barrier to resistance and long-acting potential due to low predicted hepatic metabolic clearance, based on cryopreserved hepatocyte models. Good water solubility and the longer half-life of compound GS-6207 (which binds to the linker connecting the N-terminus and C-terminus domains that form the capsid protein) offered the option of monthly subcutaneous dosing. Compound GS-6207 is currently being successfully tested in Phase Ib of randomized controlled trials to estimate its antiretroviral efficacy in treatment-experienced and treatment-naïve HIV-1 patients [69]. This study, thus far, shows a very impressive mean of a 1.8 to 2.2 log<sub>10</sub> decline in HIV-1 RNA copies at day 10 after a single subcutaneous dose of the compound. There were no indications of grade three or four adverse events requiring discontinuation [70]. In summary, capsid-targeting drugs are predicted to have high barriers to HIV-1 resistance.

### 8.4. Latency-Reversing Agents (LRAs)

As mentioned earlier, cART cannot eradicate HIV-1 due to the latent viral DNA present in cell reservoirs. It is also imperative to ensure minimal depletion of non-infected cells. LRAs are considered viable options to cure HIV-1 by destabilizing latency and causing immune clearance of infected cells and could lead to treatment-free remission. HIV-1 latency itself depends in large part on the silencing environment of the infected host cell, which can be chemically altered. The original concept, known as “shock and kill” [3,71], depends upon HIV-1 being functionally reactivated by LRAs in target cells, allowing them to be eliminated by immune effectors such as cytotoxic T lymphocytes or by virus-induced cytopathic damage (Figure 1).



**Figure 1.** The “Shock and Kill” Strategy uses Latency Reversing Agents for Immune-Mediated Clearance.

“Shock and kill” is intended to reverse proviral quiescence by inducing viral transcription and allowing a mixture of antiretroviral therapy, host immune clearance, and HIV-cytolysis to remove the remaining and latently infected cells, leading to a full virologic cure. Timmons et al. [72] showed that for diverse LRAs, latency reversal in model systems involves the heat shock factor 1 (HSF1)-mediated stress pathway. The small-molecule inhibition of HSF1 attenuated HIV-1 latency by histone deacetylase inhibitors, protein kinase C agonists, and proteasome inhibitors without interfering with the known mechanism of action of these LRAs. They demonstrated that *in vitro* models of latency have higher levels of the P-TEFb subunit cyclin T1 than primary cells do, which may explain why many LRAs are functional in model systems but relatively ineffective in primary cells. The failure of LRA in clinical trials was evidenced by a lack of reduction in the size of the reservoirs after LRA implementation. The majority of LRAs identified to date have been relatively ineffective despite activity in model systems.

Alternative therapies such as the HIV-1 transcription-inhibiting “block and lock” strategy to drive the pro-virus into a state of deep latency (utilizing latency promoting agents (LPAs) targeting either HIV or host-specific mechanisms) are, therefore, being considered [73]. Furthermore, the “block and lock” has arguments supporting its use over the “shock and kill” hypothesis due to less adverse side effects. More importantly, the concern that “shock and kill” might never completely eradicate the proviral reservoir reduces its viability for common use. Furthermore, numerous LRAs lack the “kill” aspect; therefore, the “shock and kill” therapy should be supplemented with existing immunomodulatory options. Additional novel strategies to manipulate the latent reservoirs, such as “block and lock”, should be explored further. In addition, there is a need for approaches that induce the death of latently infected cells through the apoptosis and inhibition of cellular pathways critical for cell survival. Those pathways are often hijacked by HIV-1 proteins. Given the advances in the commercial development of compounds that induce apoptosis in cancer chemotherapy, these agents could move rapidly into HIV-1 latency clinical trials, either alone or in combination with existing LRAs in order to target and eliminate latent HIV-1 infection. The development of highly specific LRAs is warranted to contribute to the eradication of HIV-1 [74].

## 9. Remaining Obstacles

Despite enormous successes, the remaining obstacles hindering current cART options are viral persistence, drug toxicities, emergence of drug resistance against existing antiretroviral regimens, and side effects. For example, current cART drug combinations also suffer from increasing pre-existent drug resistance in treated patients and transmission of those resistant variants to others. Drug resistance affects the treatment outcome, and undetectable drug resistant mutants occur even in suppressed patients [75,76], which is why new combinations of antiretrovirals are needed for better long-term safety, tolerability, adherence to cART regimens, and barriers to resistance. Second, multiple studies have demonstrated that activation of viral transcription alone is not sufficient to induce cell death, and some LRAs may have the unwanted effect of counter-acting cell death by

promoting cell survival [77]. In addition, clinical trials with LRAs have demonstrated that activation of viral gene expression is possible *in vivo*, but there is very limited or no clearance at all in the reactivated cells [78–81]. Efforts are being made to develop combinations of two or three synergizing drugs to give the highest possible potency with the lowest side effects, preferably with lower costs. This includes examining the interactions between LRAs and other drugs to determine potential synergy, antagonism, or toxicity.

### 9.1. Viral Persistence

Obstacles such as viral persistence are best considered in the context of viral replication *in vivo*. Viral replication in HIV-1 patients is largely the consequence of a dynamic process involving continuous rounds of *de novo* HIV-1 infection of and replication in activated CD4+ T cells with a rapid turnover of free virus and virus-infected cells. This process is significantly, but not completely, disturbed by effective cART. After a few months of cART therapy, plasma viral loads become undetectable in most patients. Functional assay-based laboratory analysis demonstrating evidence of decreased viral levels initially suggested that eradication of the HIV-1 infection might be achievable. Despite this evidence, there are several potential cellular and anatomical reservoirs for HIV-1 persistence that may contribute to the long-term latency of HIV-1. For example, latently infected resting memory CD4+ T cells carrying integrated HIV-1 DNA, HIV-1 infected cells in the gut-associated lymphoid tissue (GALT), central nervous system (CNS), and the male urogenital tract are active and identified locations for HIV-1 reservoirs. The half-life of resting CD4+ T cells is extremely long (3.7 yrs). That means that the eradication of this reservoir would require over 60 years of cART treatment, which is impractical as a viable eradication strategy. Latently infected resting CD4+ T cells provide a mechanism for life-long persistence of replication-competent forms of HIV-1, rendering hopes of virus eradication with current antiretroviral regimens unrealistic. The extraordinary stability of the reservoir may also reflect gradual reseeding by a very low level of ongoing viral replication and/or mechanisms that contribute to the intrinsic stability of the memory T cell compartment.

### 9.2. Side Effects

Obstacles such as substantial long-term toxicities and the side effects of current cART regimens require developing novel approaches to eradicating latent reservoirs. Solutions are urgently needed and might be addressed by the use of non-invasive CCR5 targeting drugs to intensify standard cART options. The cART refractory latent reservoirs contributing to the rapid virus rebound that occurs when latent cells become reactivated [21] prevent cART from completely eradicating HIV-1.

Efforts to develop alternative strategies have been further stimulated by the apparent cure of the “Berlin patient” [82] by a bone marrow transplant from a donor homozygous for a mutant defective CCR5 gene (CCR5  $\Delta 32$ ), which confers resistance to R5 HIV-1 [83,84]. This confirmed CCR5 as an appealing target for antiviral drugs and suggested that adding CCR5 targeting drugs to cART could increase antiviral efficacy. CCR5 is also an ideal antiviral target due to its relatively disposable role in the human immune system [85,86], suggesting that CCR5 targeting drugs may have low toxicity. By increasing antiviral efficacy, it might be possible to deplete viral reservoirs in gut-associated lymphoid tissue (GALT) and address the issue of inadequate cART distribution into this compartment. CCR5 signaling may facilitate trafficking T cells to areas of inflammation [87] and blocking such trafficking could further reduce viral spread and active replication.

### 9.3. Poor Drug Penetration

cART is not able to suppress HIV-1 replication fully due to poor drug penetration into certain tissues such as the central nervous system [88]. Existing cART drug combinations are effective, but also suffer from increasing pre-existing drug resistance in treated patients and the transmission of those resistant variants to others. Drug resistance can affect treatment outcome and undetectable resistant mutants occur even in successfully

suppressed patients [75,76]. New combinations of antiretrovirals are clearly needed for better long-term safety, tolerability, adherence to cART regimens, and barriers to resistance.

Gastrointestinal tract CCR5+ CD4+ T cells (GALT) are selectively infected and depleted during acute HIV-1 infection. GALT T cell depletion and activation persist despite cART. Persistent infection targets include long-lived CD4+ T cell subpopulations and myeloid lineage cells [89,90]. Provirus in these cells can be transcriptionally inactive for long periods, enabling the virus to evade cART drugs and immune responses [82,91]. It is difficult to reach infected cells harbored in some solid tissues, such as the central nervous system and GALT, with existing antiviral drugs. These are challenging obstacles for virologic cures [92–95]; therefore, identifying and characterizing these sites of viral persistence is a critical step toward a cure. New strategies are needed because neither the combination of latency reversing agents [89] and cART, nor host immune responses [90,91] appear to effectively reduce the pool of latently infected cells in GALT. Drug potency and specificity could be potentially enhanced using drug delivery systems such as nanoparticles coated with specific antibodies targeting CD4 or a latency marker, such as the recently described CD32a [96].

## 10. Summary

Since HIV-1 has become a chronic illness managed with various cART options, the number of patients over the age of 50 years keeps increasing with patients functioning and living successfully. Commitment to cART treatment is essential for long-term success. In addition, many different antiretroviral drugs successfully target different stages of the viral replication cycle with new combined therapeutics with lower side effects. The introduction of a lower cost cART regimen consisting of a single daily pill, injectable medications, and drug-eluting implants may significantly increase the management of HIV-1/AIDS and expand options for the utility of PreP medications for HIV-1. Altogether, this should improve the management of HIV-1/AIDS. Continuous and ongoing efforts are warranted to address eliminating reservoirs causing HIV-1 latency, and we emphasize the importance of investing research efforts into an HIV-1 cure. Identifying novel compounds that can be combined to both induce reactivation and death of HIV-1 latently infected cells are critical in this effort, as this strategy is not dependent upon an effective immune response or understanding immune escape in HIV-1 latently infected human cells.

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## References

1. Chomont, N.; El-Far, M.; Ancuta, P.; Trautmann, L.; Procopio, F.; Yassine-Diab, B.; Boucher, G.; Boulassel, M.-R.; Ghattas, G.; Brechley, J.M.; et al. HIV reservoir size and persistence are driven by T cell survival and homeostatic proliferation. *Nat. Med.* **2009**, *15*, 893–900. [\[CrossRef\]](#)
2. Bosque, A.; Famiglietti, M.; Weyrich, A.S.; Goulston, C.; Planelles, V. Homeostatic Proliferation Fails to Efficiently Reactivate HIV-1 Latently Infected Central Memory CD4+ T Cells. *PLoS Pathog.* **2011**, *7*, e1002288. [\[CrossRef\]](#)



3. Katlama, C.; Deeks, S.G.; Autran, B.; Martinez-Picado, J.; van Lunzen, J.; Rouzioux, C.; Miller, M.; Vella, S.; Schmitz, J.E.; Ahlers, J.; et al. Barriers to a Cure: New Concepts in targeting and eradicating HIV-1 reservoirs. *Lancet* **2013**, *381*, 9883. [CrossRef]
4. Natarajan, V.; Bosche, M.; Metcalf, J.A.; Ward, D.J.; Lane, H.C.; Kovacs, J.A. HIV-1 replication in patients with undetectable plasma virus receiving HAART. *Lancet* **1999**, *353*, 119–120. [CrossRef]
5. Martinez-Picado, J.; Deeks, S. Persistent HIV-1 replication during antiretroviral therapy. *Curr. Opin. HIV AIDS* **2016**, *11*, 417–423. [CrossRef]
6. Ho, Y.-C.; Shan, L.; Hosmane, N.N.; Wang, J.; Laskey, S.B.; Rosenbloom, D.I.; Lai, J.; Blankson, J.N.; Siliciano, J.D.; Siliciano, R.F. Replication-Competent Noninduced Proviruses in the Latent Reservoir Increase Barrier to HIV-1 Cure. *Cell* **2013**, *155*, 540–551. [CrossRef]
7. Richman, D.D.; Margolis, D.M.; Delaney, M.; Greene, W.C.; Hazuda, D.; Pomerantz, R.J. The Challenge of Finding a Cure for HIV Infection. *Science* **2009**, *323*, 1304–1307. [CrossRef] [PubMed]
8. Margolis, D.M. How Might We Cure HIV? *Curr. Infect. Dis. Rep.* **2014**, *16*, 392. [CrossRef]
9. Price, R.W.; Brew, B.; Sidtis, J.; Rosenblum, M.; Scheck, A.C.; Cleary, P. The Brain in AIDS: Central Nervous System HIV-1 Infection and AIDS Dementia Complex. *Science* **1988**, *239*, 586–592. [CrossRef]
10. Gorry, P.R.; Howard, J.L.; Churchill, M.J.; Anderson, J.L.; Cunningham, A.; Adrian, D.; McPhee, D.A.; Purcell, D.F.J. Diminished Production of Human Immunodeficiency Virus Type 1 in Astrocytes Results from Inefficient Translation of gag, env, and nef mRNAs despite Efficient Expression of Tat and Rev. *J. Virol.* **1999**, *73*, 352–361. [CrossRef]
11. Honeycutt, J.B.; Wahl, A.; Baker, C.; Spagnuolo, R.A.; Foster, J.; Zakharova, O.; Wietgreffe, S.; Caro-Vegas, C.; Madden, V.; Sharpe, G.; et al. Macrophages sustain HIV replication in vivo independently of T cells. *J. Clin. Investig.* **2016**, *126*, 1353–1366. [CrossRef]
12. Panel on Antiretroviral Guidelines for Adults and Adolescents. Initiating Antiretroviral Therapy in Treatment-Naive Patients Guidelines for the Use of Antiretroviral Agents in HIV-1-Infected Adults and Adolescents. Department of Health and Human Services. Available online: <http://aidsinfo.nih.gov/contentfiles/lvguidelines/AdultandAdolescentGL.pdf> (accessed on 17 May 2021).
13. World Health Organization. Available online: <https://www.who.int/news-room/fact-sheets/detail/hiv-aids#:~:text=Since%202016%2C%20WHO%20has%20recommended,status%20or%20CD4%20cell%20count> (accessed on 1 September 2021).
14. Burgess, M.; Kasten, M.J.; Zeuli, J.D. Management of HIV/AIDS in older patients—drug/drug interactions and adherence to antiretroviral therapy. *HIV/AIDS-Res. Palliat. Care* **2015**, *7*, 251–264. [CrossRef]
15. Vella, S.; Schwartländer, B.; Sow, S.P.; Eholie, S.P.; Murphy, R. The history of antiretroviral therapy and of its implementation in resource-limited areas of the world. *AIDS* **2012**, *26*, 1231–1241. [CrossRef]
16. GlaxoSmithKline. Package Insert-Retrovir (Zidovudine). Updated September 2018. Available online: [https://gskpro.com/content/dam/global/hcpportal/en\\_NA/PI/Retrovir-GDS31.pdf](https://gskpro.com/content/dam/global/hcpportal/en_NA/PI/Retrovir-GDS31.pdf) (accessed on 17 May 2021).
17. Turner, J.; Badireddy, M. *Anemia*; StatPearls Publishing: Treasure Island, FL, USA, 2019.
18. Abers, M.S.; Shandera, W.X.; Kass, J.S. Neurological and Psychiatric Adverse Effects of Antiretroviral Drugs. *CNS Drugs* **2013**, *28*, 131–145. [CrossRef] [PubMed]
19. Scruggs, E.R.; Naylor, A.J.D. Mechanisms of Zidovudine-Induced Mitochondrial Toxicity and Myopathy. *Pharmacology* **2008**, *82*, 83–88. [CrossRef]
20. Kravcik, S.; Gallicano, K.; Roth, V.; Cassol, S.; Hawley-Foss, N.; Badley, A.; Cameron, D.W. Cerebrospinal fluid HIV RNA and drug levels with combination ritonavir and Saquinavir. *J. Acquir. Immune Defic. Syndr.* **1999**, *21*, 371–375. [CrossRef]
21. Dragsted, U.B.; Gerstoft, J.; Pedersen, C.; Peters, B.; Duran, A.; Obel, N.; Castagna, A.; Cahn, P.; Clumeck, N.; Bruun, J.N.; et al. Randomized Trial to Evaluate Indinavir/Ritonavir versus Saquinavir/Ritonavir in Human Immunodeficiency Virus Type 1—Infected Patients: The MaxCmin1 Trial. *J. Infect. Dis.* **2003**, *188*, 635–642. [CrossRef]
22. Whitesid, A.; Winsbury, R. Vancouver AIDS conference: Special report. The role of the military: To protect society and themselves. *AIDS Anal. Africa.* **1996**, *6*, 4. [PubMed]
23. Da-Yong, L.; Hong-Ying, W.; Yarla, N.S.; Xu, B.; Ding, J.; Lu, T.-R. HAART in HIV/AIDS Treatments: Future Trends. *Infect. Discord Drug Targets* **2018**, *18*, 15–22.
24. El-Sadr, W.M.; Holmes, C.B.; Mugenyi, P.; Thirumurthy, H.; Ellerbrock, T.; Ferris, R.; Sanne, I.; Asiimwe, A.; Hirnschall, G.; Nkambule, N.R.; et al. Scale-up of HIV treatment through PEPFAR: A historic public health achievement. *J. Acquir. Immune Defic. Syndr.* **2012**, *60* (Suppl. 3), S96–S104. [CrossRef]
25. Chow, D. Epidemiological evidence of increasing blood pressure in HIV-1-infected individuals in the era of HAART. *Antivir. Ther.* **2000**, *5*, 13.
26. Maggiolo, F.; Arici, C.; Airoidi, M.; Ripamonti, D.; Quinzan, G.; Gregis, G.; Ravasio, V.; Bombana, E.; Suter, F. Reasons for discontinuation of nevirapine-containing HAART: Results from an unselected population of a large clinical cohort. *J. Antimicrob. Chemother.* **2007**, *59*, 569–572. [CrossRef] [PubMed]
27. Max, B.; Sherer, R. Management of the Adverse Effects of Antiretroviral Therapy and Medication Adherence. *Clin. Infect. Dis.* **2000**, *30*, S96–S116. [CrossRef]
28. Margolis, A.M.; Heverling, H.; Pham, P.A.; Stolbach, A. A Review of the Toxicity of HIV Medications. *J. Med. Toxicol.* **2013**, *10*, 26–39. [CrossRef]
29. The INSIGHT START Study Group. Initiation of Antiretroviral Therapy in Early Asymptomatic HIV Infection. *N. Engl. J. Med.* **2015**, *373*, 795–807. [CrossRef] [PubMed]

30. Marcus, J.L.; Chao, C.R.; Leyden, W.A.; Xu, L.; Quesenberry, C.P.; Klein, D.B.; Towner, W.J.; Horberg, M.A.; Silverberg, M.J. Narrowing the Gap in Life Expectancy Between HIV-Infected and HIV-Uninfected Individuals With Access to Care. *JAIDS J. Acquir. Immune Defic. Syndr.* **2016**, *73*, 39–46. [[CrossRef](#)]
31. Abel, S.; Back, D.J.; Vourvahis, M. Maraviroc: Pharmacokinetics and drug interactions. *Antivir. Ther.* **2009**, *14*, 607–618.
32. Latinovic, O.S.; Le, N.; Reitz, M.; Pal, R.; DeVico, A.; Foulke, J.S.; Redfield, R.R.; Heredia, A. Synergistic Inhibition of CCR5-tropic HIV-1 by Maraviroc and CCR5 Antibody HGS004 in Primary Cells: Potential Implications for Treatment and Prevention. *AIDS* **2011**, *25*, 1232–1235. [[CrossRef](#)]
33. Heredia, A.; Latinovic, O.S.; Gallo, R.C.; Melikian, G.B.; Reitz, M.; Le, N.; Redfield, R.R. Low doses of Rapamycin enhance the antiviral activity of CCR5 antagonist VCV against wild-type and drug resistant R5 HIV-1 (co-first authorship). *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 20476–20481. [[CrossRef](#)]
34. Chan, D.C.; Kim, P.S. HIV Entry and Its Inhibition. *Cell* **1998**, *93*, 681–684. [[CrossRef](#)]
35. Zhang, X.; Ding, X.; Zhu, Y.; Chong, H.; Cui, S.; He, J.; Wang, X.; He, Y. Structural and functional characterization of HIV-1 cell fusion inhibitor T20. *AIDS* **2019**, *33*, 1–11. [[CrossRef](#)]
36. Latinovic, O.; Kuruppu, J.; Davis, C.; Le, N.; Heredia, A. Pharmacotherapy of HIV-1 Infection: Focus on CCR5 Antagonist Maraviroc. *Clin. Med.* **2009**, *1*, 1497–1510. [[CrossRef](#)]
37. Hyland, R.; Dickins, M.; Collins, C.; Jones, H.; Jones, B. Maraviroc: In vitro assessment of drug-drug interaction potential. *Br. J. Clin. Pharmacol.* **2008**, *66*, 498–507. [[CrossRef](#)]
38. Gardner, M.R.; Farzan, M. Engineering antibody-like inhibitors to prevent and treat HIV-1 infection. *Curr. Opin. HIV AIDS* **2017**, *12*, 294–301. [[CrossRef](#)]
39. Deen, K.C.; McDouga, J.S.; Inacker, R.; Folena-Wasserman, G.; Arthos, J.; Rosenberg, J.; Maddon, P.J.; Axel, R.; Sweet, R.W. A soluble form of CD4 (T4) protein inhibits AIDS virus infection. *Nature* **1988**, *331*, 82–84. [[CrossRef](#)]
40. Gardner, M.R.; Kattenhorn, L.M.; Kondur, H.R.; von Schaeuwen, M.; Dorfman, T.; Chiang, J.J.; Haworth, K.G.; Decker, J.M.; Alpert, M.D.; Bailey, C.C.; et al. AAV-expressed eCD4-Ig provides durable protection from multiple SHIV challenges. *Nature* **2015**, *519*, 87–91. [[CrossRef](#)]
41. Huang, J.; Kang, B.H.; Ishida, E.; Zhou, T.; Griesman, T.; Sheng, Z.; Wu, F.; Doria-Rose, N.A.; Zhang, B.; McKee, K.; et al. Identification of a CD4-Binding-Site Antibody to HIV that Evolved Near-Pan Neutralization Breadth. *Immunity* **2016**, *45*, 1108–1121. [[CrossRef](#)]
42. Chen, W.; Zhu, Z.; Feng, Y.; Dimitrov, D.S. Human domain antibodies to conserved sterically restricted regions on gp120 as exceptionally potent cross-reactive HIV-1 neutralizers. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17121–17126. [[CrossRef](#)] [[PubMed](#)]
43. Schiavone, M.; Fiume, G.; Caivano, A.; De Laurentiis, A.; Falcone, C.; Masci, F.F.; Iaccino, E.; Mimmi, S.; Palmieri, C.; Pisano, A.; et al. Design and Characterization of a Peptide Mimotope of the HIV-1 gp120 Bridging Sheet. *Int. J. Mol. Sci.* **2012**, *13*, 5674–5699. [[CrossRef](#)]
44. Wang, X.; Cao, M.; Wu, Y.; Xu, W.; Wang, Q.; Ying, T.; Lu, L.; Jiang, S. Synergistic Effect by Combining a gp120-Binding Protein and a gp41-Binding Antibody to Inactivate HIV-1 Virions and Inhibit HIV-1 Infection. *Molecules* **2021**, *26*, 1964. [[CrossRef](#)]
45. Andreatta, K.; Willkom, M.; Martin, R.; Chang, S.; Wei, L.; Liu, H.; Liu, Y.-P.; Graham, H.; Quirk, E.; Martin, H.; et al. Erratum to: Switching to bictegravir/emtricitabine/tenofovir alafenamide maintained HIV-1 RNA suppression in participants with archived antiretroviral resistance including M184V/I. *J. Antimicrob. Chemother.* **2019**, *74*, 3646–3647. [[CrossRef](#)]
46. Brehm, T.T.; Franz, M.; Hüfner, A.; Hertling, S.; Schmiedel, S.; Degen, O.; Kreuels, B.; Schulze Zur Weisch, J. Safety and efficacy of elvitegravir, dolutegravir, and raltegravir in a real-world cohort of treatment-naïve and -experienced patients. *Medicine* **2019**, *98*, e16721. [[CrossRef](#)]
47. Cahn, P.; Pozniak, A.L.; Mingrone, H.; Shuldyakov, A.; Brites, C.; Andrade-Villanueva, J.F.; Richmond, G.; Buendia, C.B.; Fourie, J.; Ramgopal, M.; et al. Dolutegravir versus raltegravir in antiretroviral-experienced, integrase-inhibitor-naïve adults with HIV: Week 48 results from the randomised, double-blind, non-inferiority SAILING study. *Lancet* **2013**, *382*, 700–708. [[CrossRef](#)]
48. McAllister, J.W.; Towns, J.M.; McNulty, A.; Pierce, A.B.; Foster, R.; Richardson, R.; Carr, A. Dolutegravir with tenofovir disoproxil fumarate-emtricitabine as HIV postexposure prophylaxis in gay and bisexual men. *AIDS* **2017**, *31*, 1291–1295. [[CrossRef](#)]
49. Raffi, F.; Jaeger, H.; Quiros-Roldan, E.; Albrecht, H.; Belonosova, E.; Gatell, J.M.; Baril, J.-G.; Domingo, P.; Brennan, C.; Almond, S.; et al. Once-daily dolutegravir versus twice-daily raltegravir in antiretroviral-naïve adults with HIV-1 infection (SPRING-2 study): 96 week results from a randomised, double-blind, non-inferiority trial. *Lancet Infect. Dis.* **2013**, *13*, 927–935. [[CrossRef](#)]
50. Powderly, W.G. Integrase inhibitors in the treatment of HIV-1 infection. *J. Antimicrob. Chemother.* **2010**, *65*, 2485–2488. [[CrossRef](#)]
51. Venter, E.D.F.; Moorhouse, S.; Sokhela, L.; Fairlie, L.; Mashabane, N.; Masenya, M.; Serenata, C.; Akpomiemie, G.; Qavi, A.; Chandiwana, N.; et al. Dolutegravir plus Two Different Prodrugs of Tenofovir to Treat HIV. *N. Engl. J. Med.* **2019**, *381*, 803–815. [[CrossRef](#)]
52. Guidelines for the Use of Antiretroviral Agents in Adults and Adolescents with HIV. Available online: <https://clinicalinfo.hiv.gov/sites/default/files/guidelines/documents/AdultandAdolescentGL.pdf> (accessed on 3 June 2021).
53. Wohl, D.A.; Yazdanpanah, Y.; Baumgarten, A.; Clarke, A.; Thompson, M.A.; Brinson, C.; Hagins, D.; Ramgopal, M.N.; Antinori, A.; Wei, X.; et al. Bictegravir combined with emtricitabine and tenofovir alafenamide versus dolutegravir, abacavir, and lamivudine for initial treatment of HIV-1 infection: Week 96 results from a randomized, double-blind, multicenter, phase3, non-inferiority trial. *Lancet* **2019**, *6*, e355–e363. [[CrossRef](#)]

54. U.S. Food and Drug Administration. FDA Approves First Two-Drug Regimen for Certain Patients with HIV. Available online: <https://www.fda.gov/news-events/press-announcements/fda-approves-first-two-drug-regimen-certain-patients-hiv> (accessed on 17 May 2021).
55. U.S. Food and Drug Administration. FDA Approves First Two-Drug Complete Regimen for HIV-Infected Patients Who Have Never Received Antiretroviral Treatment. Available online: <https://www.fda.gov/news-events/press-announcements/fda-approves-first-two-drug-complete-regimen-hiv-infected-patients-who-have-never-received> (accessed on 17 May 2021).
56. U.S. Preventive Services Task Force. Final Draft Recommendation: Human Immunodeficiency Virus (HIV) Infection: Screening. Available online: <https://www.uspreventiveservicestaskforce.org/uspstf/recommendation/human-immunodeficiency-virus-hiv-infection-screening> (accessed on 17 May 2021).
57. U.S. Food and Drug Administration. Drug Approval Package. Available online: [https://www.accessdata.fda.gov/drugsatfda\\_docs/nda/2011/202895\\_prezista\\_toc.cfm](https://www.accessdata.fda.gov/drugsatfda_docs/nda/2011/202895_prezista_toc.cfm) (accessed on 7 September 2021).
58. U.S. Food and Drug Administration. FDA Approves Cabenuva and Vocabria for the Treatment of HIV-1 Infection. Available online: <https://www.uspreventiveservicestaskforce.org/Home/GetFileByID/1890> (accessed on 27 January 2021).
59. Pace, C.S.; Fordyce, M.W.; Franco, D.; Kao, C.-Y.; Seaman, M.S.; Ho, D.D. Anti-CD4 Monoclonal Antibody Ibalizumab Exhibits Breadth and Potency Against HIV-1, With Natural Resistance Mediated by the Loss of a V5 Glycan in Envelope. *JAIDS J. Acquir. Immune Defic. Syndr.* **2013**, *62*, 1–9. [[CrossRef](#)]
60. Emu, B.; Fessel, J.; Schrader, S.; Kumar, P.; Richmond, G.; Win, S.; Weinheimer, S.; Marsolais, C.; Lewis, S. Phase 3 Study of Ibalizumab for Multidrug-Resistant HIV-1. *N. Engl. J. Med.* **2018**, *379*, 645–654. [[CrossRef](#)]
61. Grobler, J.A.; Huang, Q.; Hazuda, D.J.; Lai, M.-T. Efficacy of MK-8591 against diverse HIV-1 subtypes and NRTI-resistant clinical isolates. *J. Int. AIDS Soc.* **2018**, *2*, O343.
62. Grobler, J.; Friedman, E.; Barrett, S.E.; Wood, S.L.; Ankrom, W.; Fillgrove, K.L.; Lai, M.-T.; Gindy, M.; Iwamoto, M.; Hazuda, D.J. Long-acting oral and parenteral dosing of MK-8591 for HIV treatment or prophylaxis. In Proceedings of the Conference on Retroviruses and Opportunistic Infections (CROI), Boston, MA, USA, 22–25 February 2016.
63. Matthews, R.P.; Schurmann, D.; Rudd, D.J.; Levine, V.; Fox-Bosetti, S.; Zhang, S.; Robberechts, M.; Huser, A.; Hazuda, D.J.; Iwamoto, M.; et al. Single doses as low as 0.5 mg of the novel NRTTI MK-8591 suppress HIV for at least seven days. In Proceedings of the International AIDS Society Conference, Paris, France, 23–26 July 2017.
64. CROI 2021 Presentation 88. Available online: <https://www.croiconference.org/> (accessed on 3 June 2021).
65. Safety and Pharmacokinetics of Oral Islatravir (MK-8591) Once Monthly in Participants at Low Risk of Human Immunodeficiency Virus 1 (HIV-1) Infection (MK-8591-016). Available online: <https://clinicaltrials.gov/ct2/show/NCT04003103> (accessed on 17 May 2021).
66. Matthews, R. First-in-Human Trial of MK-8591-Eluting Implants Demonstrates Concentrations Suitable for HIV Prophylaxis for at Least One Year. In Proceedings of the International AIDS Society Conference, Mexico City, Mexico, 21–24 July 2019.
67. Tse, W.L.J.O.; Mulato, A.; Niedziela-Majka, A.; Rowe, W.; Somoza, J.R.; Villasenor, A.G.; Yant, S.R.; Zhang, J.R.; Zheng, J. Discovery of Novel Potent HIV Capsid Inhibitors with Long-Acting Potential. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Seattle, Washington, DC, USA, 13–16 February 2017.
68. Carnes, S.K.; Sheehan, J.H.; Aiken, C. Inhibitors of the HIV-1 capsid, a target of opportunity. *Curr. Opin. HIV AIDS* **2018**, *13*, 359–365. [[CrossRef](#)]
69. Safety, Pharmacokinetics, and Antiviral Activity of GS-6207 Administered Subcutaneously in HIV-1 Infected Adults. Available online: <https://ClinicalTrials.gov/show/NCT03739866> (accessed on 17 May 2021).
70. Mascolini, M. *Sharp Drops in HIV Load after 10 Days of Capsid Inhibitor Monotherapy*; International AIDS Society: Mexico City, Mexico, 2019.
71. Hamer, D.H. Can HIV be Cured? Mechanisms of HIV Persistence and Strategies to Combat It. *Curr. HIV Res.* **2004**, *2*, 99–111. [[CrossRef](#)]
72. Timmons, A.; Fray, E.; Kumar, M.; Wu, F.; Dai, W.; Bullen, C.K.; Kim, P.; Hetzel, C.; Yang, C.; Beg, S.; et al. HSF1 inhibition attenuates HIV-1 latency reversal mediated by several candidate LRAs In Vitro and Ex Vivo. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 15763–15771. [[CrossRef](#)]
73. Abner, E.; Jordan, A. HIV “shock and kill” therapy: In need of revision. *Antivir. Res.* **2019**, *166*, 19–34. [[CrossRef](#)] [[PubMed](#)]
74. Ait-Ammar, A.; Kula, A.; Darcis, G.; Verdikt, R.; De Wit, S.; Gautier, V.; Mallon, P.W.G.; Marcello, A.; Rohr, O.; Van Lint, C. Current Status of Latency Reversing Agents Facing the Heterogeneity of HIV-1 Cellular and Tissue Reservoirs. *Front. Microbiol.* **2020**, *10*, 3060. [[CrossRef](#)] [[PubMed](#)]
75. Sengupta, S.; Siliciano, R.F. Targeting the Latent Reservoir for HIV-1. *Immunity* **2018**, *48*, 872–895. [[CrossRef](#)]
76. Vanhamel, J.; Bruggemans, A.; Debyser, Z. Establishment of latent HIV-1 reservoirs: What do we really know? *J. Virus Erad.* **2019**, *5*, 3–9. [[CrossRef](#)]
77. Kim, Y.; Anderson, J.L.; Lewin, S.R. Getting the “kill” into “shock and kill”: Strategies to eliminate latent HIV. *Cell Host Microbe.* **2018**, *23*, 14–26. [[CrossRef](#)] [[PubMed](#)]
78. Archin, N.M.; Kirchherr, J.L.; Sung, J.A.; Clutton, G.; Sholtis, K.; Xu, Y.; Allard, B.; Stuelke, E.; Kashuba, A.D.; Kuruc, J.D.; et al. Interval dosing with the HDAC inhibitor vorinostat effectively reverses HIV latency. *J. Clin. Investig.* **2017**, *127*, 3126–3135. [[CrossRef](#)] [[PubMed](#)]

79. Archin, N.M.; Liberty, A.L.; Kashuba, A.D.; Choudhary, S.K.; Kuruc, J.D.; Crooks, A.M.; Parker, D.C.; Anderson, E.M.; Kearney, M.F.; Strain, M.C.; et al. Administration of vorinostat disrupts HIV-1 latency in patients on antiretroviral therapy. *Nature* **2012**, *487*, 482–485. [[CrossRef](#)]
80. Elliott, J.H.; McMahon, J.; Chang, C.C.; Lee, S.A.; Hartogensis, W.; Bumpus, N.; Savic, R.; Roney, J.; Hoh, R.; Solomon, A.; et al. Short-term administration of disulfiram for reversal of latent HIV infection: A phase 2 dose-escalation study. *Lancet HIV* **2015**, *2*, e520–e529. [[CrossRef](#)]
81. Elliott, J.H.; Wightman, F.; Solomon, A.; Ghneim, K.; Ahlers, J.; Cameron, M.J.; Smith, M.Z.; Spelman, T.; McMahon, J.; Velayudham, P.; et al. Activation of HIV Transcription with Short-Course Vorinostat in HIV-Infected Patients on Suppressive Antiretroviral Therapy. *PLoS Pathog.* **2014**, *10*, e1004473. [[CrossRef](#)] [[PubMed](#)]
82. Hütter, G.; Nowak, D.; Mossner, M.; Ganepola, S.; Müssig, A.; Allers, K.; Schneider, T.; Hofmann, J.; Kücherer, C.; Blau, O.; et al. Long-Term Control of HIV by CCR5Delta32/Delta32 Stem-Cell Transplantation. *N. Engl. J. Med.* **2009**, *360*, 692–698. [[CrossRef](#)] [[PubMed](#)]
83. Michael, N.L.; Louie, L.G.; Sheppard, H.W. CCR5-delta 32 gene deletion in HIV-1 infected patients. *Lancet* **1997**, *350*, 741–742. [[CrossRef](#)]
84. Liu, R.; Paxton, W.A.; Choe, S.; Ceradini, D.; Martin, S.R.; Horuk, R.; MacDonald, M.E.; Stuhlmann, H.; Koup, R.A.; Landau, N.R. Homozygous Defect in HIV-1 Coreceptor Accounts for Resistance of Some Multiply-Exposed Individuals to HIV-1 Infection. *Cell* **1996**, *86*, 367–377. [[CrossRef](#)]
85. Dorr, P.; Westby, M.; Dobbs, S.; Griffin, P.; Irvine, B.; Macartney, M.; Mori, J.; Rickett, G.; Smith-Burchnell, C.; Napier, C.; et al. Maraviroc (UK-427,857), a Potent, Orally Bioavailable, and Selective Small-Molecule Inhibitor of Chemokine Receptor CCR5 with Broad-Spectrum Anti-Human Immunodeficiency Virus Type 1 Activity. *Antimicrob. Agents Chemother.* **2005**, *49*, 4721–4732. [[CrossRef](#)]
86. Ioannidis, J.P.; Rosenberg, P.S.; Goedert, J.J.; Ashton, L.J.; Benfield, T.L.; Buchbinder, S.P.; Coutinho, R.A.; Eugen-Olsen, J.; Gallart, T.; Katzenstein, T.L.; et al. International Meta-Analysis of HIV Host Genetics: Effects of CCR5-Delta32, CCR2-64I, and SDF-13' A alleles on HIV-1 disease progression: An International Meta-Analysis of Individual Patient Data. *Ann. Intern. Med.* **2001**, *135*, 782–795. [[CrossRef](#)]
87. Fera, M.G.; Taborda, N.A.; Hernandez, J.C.; Rugeles, M.T. HIV replication is associated to inflammasomes activation, IL-1 $\beta$ , IL-18 and caspase-1 expression in GALT and peripheral blood. *PLoS ONE* **2018**, *13*, e0192845. [[CrossRef](#)] [[PubMed](#)]
88. Gray, L.R.; Roche, M.; Flynn, J.K.; Wesselingh, S.L.; Gorry, P.R.; Churchill, M.J. Is the central nervous system a reservoir of HIV-1? *Curr. Opin. HIV AIDS* **2014**, *9*, 552–558. [[CrossRef](#)]
89. Walker-Sperling, V.E.; Pohlmeier, C.W.; Tarwater, P.M.; Blankson, J.N. The Effect of Latency Reversal Agents on Primary CD8+ T Cells: Implications for Shock and Kill Strategies for Human Immunodeficiency Virus Eradication. *EBioMedicine* **2016**, *8*, 217–229. [[CrossRef](#)] [[PubMed](#)]
90. Kessing, C.F.; Nixon, C.C.; Li, C.; Tsai, P.; Takata, H.; Mousseau, G.; Ho, P.T.; Honeycutt, J.B.; Fallahi, M.; Trautmann, L.; et al. In Vivo Suppression of HIV Rebound by Didehydro-Cortistatin A, a “Block-and-Lock” Strategy for HIV-1 Treatment. *Cell Rep.* **2017**, *21*, 600–611. [[CrossRef](#)]
91. Kulpa, D.A.; Chomont, N. HIV persistence in the setting of antiretroviral therapy: When, where and how does HIV hide? *J. Virus Erad.* **2015**, *1*, 59–66. [[CrossRef](#)]
92. Shultz, L.D.; Brehm, M.; Garcia-Martinez, J.V.; Greiner, D.L. Humanized mice for immune system investigation: Progress, promise and challenges. *Nat. Rev. Immunol.* **2012**, *12*, 786–798. [[CrossRef](#)] [[PubMed](#)]
93. Siliciano, J.D.; Kajdas, J.; Finzi, D.; Quinn, T.C.; Chadwick, K.; Margolick, J.B.; Kovacs, C.; Gange, S.; Siliciano, R.F. Long-term follow-up studies confirm the stability of the latent reservoir for HIV-1 in resting CD4+ T cells. *Nat. Med.* **2003**, *9*, 727–728. [[CrossRef](#)] [[PubMed](#)]
94. Mok, H.P.; Javed, S.; Lever, A. Stable gene expression occurs from a minority of integrated HIV-1-based vectors: Transcriptional silencing is present in the majority. *Gene Ther.* **2007**, *14*, 741–751. [[CrossRef](#)] [[PubMed](#)]
95. Zhang, L.; Su, L. HIV-1 immunopathogenesis in humanized mouse models. *Cell. Mol. Immunol.* **2012**, *9*, 237–244. [[CrossRef](#)]
96. Descours, B.; Petitjean, G.; Lopez-Zaragoza, J.L.; Bruel, T.; Raffel, R.; Psomas, C.; Reynes, J.; Lacabaratz, C.; Levy, Y.; Schwartz, O.; et al. CD32 is a marker of a CD4 T-cell HIV reservoir harboring replication-competent proviruses. *Nature* **2017**, *543*, 564–567. [[CrossRef](#)]