





REVIEW

Exploring geographical variances in component-resolved diagnosis within the Asia-Pacific region

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Abstract

Component-resolved diagnostics (CRD) have revolutionized allergy diagnosis, offering enhanced accuracy and insights into allergen sensitization patterns. This review explores geographical variances in CRD for food and aeroallergens across the Asia-Pacific region. We examine the varying prevalence of allergic diseases and the utility of CRD in diagnosing common food allergies, including peanut, shellfish, fish, wheat, and fruits. Notable differences in serum-specific (sp)IgE sensitization patterns and the clinical relevance of particular allergen components are observed between populations in Asian countries and those in Europe and the United States. For food allergies, the literature reports significant differences in allergen components and their diagnostic utility across various countries. Peanut allergy diagnostics, particularly Ara h 2 sIgE, show varying sensitivity and specificity between Asian and Western populations. In shellfish allergy, emerging allergens beyond tropomyosin are gaining importance in the Asia-Pacific region. Fish and wheat allergies also demonstrate unique sensitization patterns, emphasizing the need for region-specific diagnostic approaches. Regarding aeroallergens, pollen sensitization profiles vary widely across the

Abbreviations: AD, atopic dermatitis; AR, allergic rhinitis; AUC, area under the curve; CCDs, cross-reactive carbohydrate determinants; CRD, component-resolved diagnostics; GAN, Global Asthma Network; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; HWP-WDEIA, WDEIA with reactions to hydrolyzed wheat protein; ISAAC, International Study of Asthma and Allergies in Childhood; LFS, Latex-fruit syndrome; LTP, lipid transfer protein; OAS, oral allergy syndrome; PFAS, pollen food allergy syndrome; SCP, sarcoplasmic calcium-binding protein; sIgE, serum specific IgE; SPT, skin prick tests; WDEIA, wheat-dependent exercise-induced anaphylaxis; ω5G, omega-5 gliadin.

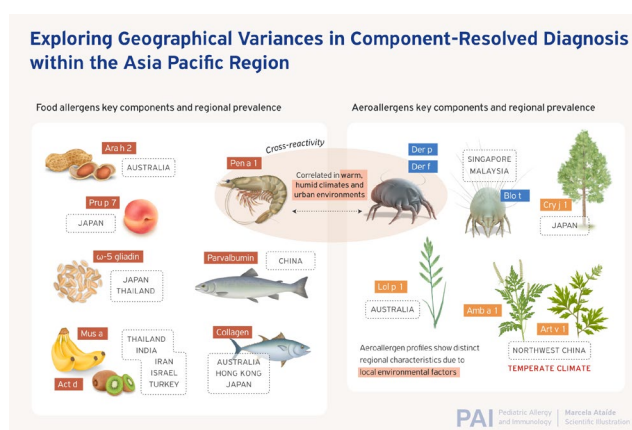
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region, influenced by local flora and climate, to influence symptoms of pollen food allergy syndrome. House dust mite allergens remain a significant concern, with high sensitization rates to major components like Der p 1, Der p 2, and the emerging Der p 23. The cross-reactivity between house dust mite and shellfish allergens is particularly relevant in this region. These findings emphasize the need for region-specific CRD approaches, considering local allergen profiles and sensitization patterns. As the prevalence of allergic diseases continues to rise in the Asia-Pacific, further research into locally relevant allergens and their components is crucial for improving diagnosis, patient management, and targeted immunotherapy strategies in this diverse region.

KEYWORDS

Asia-Pacific, component-resolved diagnosis, geographical variance, molecular allergology



1 | INTRODUCTION

Component-resolved diagnostics (CRD) are increasingly becoming a part of the allergist's toolkit. CRD involves analyzing serum specific (sp) IgE levels directed against individual allergenic proteins or components rather than whole allergen extracts. In many cases, CRD offers greater diagnostic accuracy than traditional skin prick tests or spIgE to whole allergens, such as Ara h 2 and Ana o 3 for peanut and cashew allergies, respectively.¹⁻⁵ Given the known variations in allergens and allergy prevalence observed in the epidemiological literature worldwide, considering the range of allergens specific to the Asia-Pacific region is important for the accurate diagnosis and management of allergic diseases. This review considers geographical variances in CRD for food and aeroallergens in the Asia-Pacific Region.

2 | DIFFERENCE IN PREVALENCE OF ALLERGIC DISEASE ACROSS ASIA-PACIFIC REGION

Although allergies were once thought to be rare in Asia, recent evidence suggests that they are becoming more common. Survey data indicates that the prevalence of food allergy in Taiwan has increased

from 7.7% to 10.4% in children and from 6.4% to 12.5% in adults, between 2004 and 2017.⁶ In the Chongqing region of China, food allergy, confirmed using oral food challenges, increased from 3.5% in 1999 to 7.7% in 2009 and 11.1% in 2019.⁷ This rise mirrors global trends. For example, in Australia, which has one of the highest food allergy rates worldwide, approximately 10% of children have food allergies, including a 3% prevalence of peanut allergy.⁸ Similarly, in the United States, 7.6% of children have food allergies, with shellfish allergy being particularly common.⁹ The triggers of food allergy vary between regions, with seafood allergies being more common in Southeast Asia, while wheat allergies are more common in Japan and Thailand.^{10,11} Egg, milk and seafood are among the most common food allergies in China.¹² In contrast, peanut and tree nut allergies are more prevalent in urbanized regions like Australia and Singapore.^{13,14} Migration from Asia to Australia also influences the prevalence of allergic disease, with babies living in Australia born to Asian parents exhibiting a higher prevalence of food allergy compared to infants living in Australia with Australian-born parents.¹⁵

A recent systematic review found that the prevalence of allergic rhinitis (AR) varies greatly depending on the region and method of defining the outcome, ranging from 1.0% to 47.9% for Asia and 19.2% to 47.5% for Oceania.¹⁶ The International Study of Asthma and Allergies in Childhood (ISAAC) Phase Three survey involving 61 countries showed that 6.9% of

adolescents worldwide experienced severe asthma symptoms, including 3.8% of those in Asia-Pacific and Northern and Eastern Europe and 11.3% in North America.¹⁷ Although the prevalence of asthma in the Asia-Pacific region was comparatively lower compared to that in Western countries, including Australia, where the prevalence of AR is 23.8% (NHS survey 2023), recent evidence shows that the prevalence is on the rise, particularly in countries with rapidly developing economies, such as China.¹⁸

A cross-sectional study conducted across 18 countries found that the prevalence of atopic dermatitis (AD), based on the ISAAC criteria and self-report of AD diagnosis, was between 2.7% and 20.1%; the prevalence of AD was around 11% in Japan and Taiwan.¹⁹ The Global Asthma Network (GAN) assessed eczema prevalence and severity across 27 global centers over time.²⁰ The study found only a small overall 10-year increase in both current eczema and severe eczema prevalence, with no significant increase in Asia. However, there was substantial variation between centers, even within the same income group, highlighting that while income level helps explain some regional patterns, it does not fully account for the observed differences in eczema prevalence globally. Other factors, such as geography, climate, environment, urbanization, and cultural practices, likely play crucial roles. These differences underscore the importance of region-specific studies and tailored strategies to manage allergies effectively in the Asia-Pacific region.

3 | COMPONENT-RESOLVED DIAGNOSTICS FOR FOOD ALLERGY

Diagnosis of food allergy relies heavily on the clinical history of an IgE-mediated allergic reaction. Skin prick tests (SPT) and serum-specific IgE (sIgE) are common first-line diagnostic tests to evaluate food allergen sensitization. These tests can be useful as supporting evidence to confirm an allergy diagnosis or determine when a food may be suitable to reintroduce, but they have limitations as a standalone test.^{21,22} SPT and sIgE have different sensitivity and specificity profiles for different foods. Serum specific IgE to food allergen components is increasingly utilized in the clinical setting and has been shown to have a superior diagnostic performance for certain food sources, notably superior to SPT, particularly in equivocal cases.^{3,23,24} Table 1 summarizes the diverse diagnostic performance of food allergen components with a focus on the Asia-Pacific region.

3.1 | Peanut allergy

Geographic variability in peanut allergy diagnostics underscores the need for region-specific strategies, with component-resolved diagnostics like Ara h 2 showing differing performance between Asia and the Western countries, such as those in North America, Europe, and Australasia. A recent systematic review and meta-analysis of the diagnostic tests used in food allergy provides the most thorough summary of the diagnostic accuracy of several tests, including SPT, sIgE to whole food proteins, and CRD.³ The review included 149 studies, with meta-analysis performed when three or more studies

Key message

Component-resolved diagnostics (CRD) demonstrates significant geographical variations in allergen sensitization patterns between Asia-Pacific populations and Western populations, highlighting the critical need for region-specific diagnostic approaches. The review reveals that established diagnostic markers and their clinical utility differ substantially across regions, as evidenced by the varying sensitivity and specificity of common allergen components such as Ara h 2 for peanut allergy between Asian and Western populations. Additionally, unique sensitization patterns are observed in shellfish, fish, wheat and fruit allergies specific to the Asia-Pacific region, while aeroallergen profiles, particularly for pollens and house dust mites, show distinct regional characteristics influenced by local environmental factors. These findings underscore the importance of developing and validating region-specific CRD approaches rather than universally applying Western diagnostic standards, as this could improve the accuracy of allergy diagnosis and management strategies in the diverse Asia-Pacific region. Further research into locally relevant allergens and their components is essential for advancing targeted immunotherapy and patient care in this region.

were found for a given index test and food. Serum specific IgE to peanut components demonstrated high specificity for several components, including Ara h 2 (92%), Ara h 3 (93%), Ara h 6 (94%) and Ara h 8 (99%). However, the sensitivity was good in only Ara h 2 (82%) and Ara h 6 (87%) at cut-offs of 0.44 and 0.4 kUA/L, respectively. All other components had sensitivity below 50%. Data on Ara h 8 and Ara h 9 sIgE is heterogeneous and appears to be of limited clinical use. Meta-analyses for Ara h 2 sIgE demonstrated maximum sensitivity (0.90) at 0.2 kUA/L and maximum specificity (98%) at 1.15 kUA/L.³ Most of the data analyzed in this systematic review come from Europe and North America, which limits its applicability in other populations.

There is geographic variability in the accuracy of CRD tests worldwide, including within the Asia-Pacific region. In Australia, Ara h 2 sIgE testing, assessed in the HealthNuts study in a random sample of 200 infants (100 with OFC-confirmed peanut allergy and 100 with peanut tolerance), was integrated into a diagnostic algorithm. In this approach, Ara h 2 sIgE testing was only performed for children with skin prick test (SPT) results in an intermediate range (3–8 mm). This algorithm demonstrated higher diagnostic accuracy than whole peanut sIgE, with 60% sensitivity (95% CI, 50–70) at 98% specificity (95% CI, 93–100), and reduced the need for oral food challenges by nearly two-thirds when used in combination.² In contrast, peanut SPT demonstrated the best diagnostic performance in Hong Kong children (3.7 mm threshold, sensitivity 93%, specificity 89%), while

TABLE 1 Key findings in food allergen sensitization and diagnostic patterns across the Asia-Pacific region.

Study Type	Country	Cohort	Number of Participants	Allergens Tested	Frequency of sIgE to Allergen	Key Findings	Ref
Retrospective study	Hong Kong	Peanut allergy	133	Peanut extract, Ara h 2; SPT peanut extract	SPT 3.7 mm threshold, sensitivity 93%, specificity 89%), Ara h 2; good sensitivity with adjusted cut-offs.	95% PPVs for HK: 5.6 mm (SPT), 8.4 kUA/L (peanut sIgE), 2.4 kUA/L (Ara h 2); good sensitivity with adjusted cut-offs.	(116)
Retrospective study	Singapore	Peanut allergy	167	Peanut extract, Ara h 2; SPT peanut extract	Ara h 2 0.35 kUA/L threshold, sensitivity 91%, specificity 87%	95% PPV: 13 mm (SPT), 34 kUA/L (peanut sIgE), 4.26 kUA/L (Ara h 2); Ara h 2 most sensitive (90%).	(116)
Cohort study	Australia	Peanut allergy	100 OFC-confirmed peanut allergy; 100 with peanut tolerance	Ara h 2. Whole peanut	60% sensitivity, 98% specificity for Ara h 2	Ara h 2 superior diagnostic tool compared to whole peanut sIgE, reducing OFCs by two-thirds.	(2)
Prospective study	Hong Kong; Thailand	Shrimp allergy	85 (Hong Kong: 17 OFC-confirmed shrimp allergic; 32 probable shrimp allergy, 20 OFC-confirmed shrimp tolerant. Thailand: 16 physician-diagnosed shrimp allergy)	Pen m 1, Pen m 2, Pen m 3, Pen m 4, Pen m 6, Pen m 7, Pen m 8, Pen m 13, Pen m 14, SPT shellfish extract, sIgE shrimp extract	Hong Kong: Pen m 4, 6, 13 AUC 0.77–0.78; Thailand: Pen m 4 AUC 0.96, Pen m 1 AUC 0.89	Pen m 1, Pen m 6, Pen m 13 and Pen m 14 are major shrimp allergens. Component test is superior compared to shrimp extract sIgE and SPT	(35)
Prospective study	Hong Kong (HK) and Japan	Fish-allergic individuals	45 (30 from HK, 15 from Japan) + 2 non-atopic controls	Salmon and grass carp proteins, including parvalbumin, enolase, collagen, aldolase, and GAPDH	- 60.0% (HK) and 66.7% (Japan) sensitized to 10 kDa parvalbumin - 40.0% (Japan) sensitized to 130 kDa collagen and 58 kDa enolase	Japanese participants showed broader IgE binding, including grass carp, despite lower IgE levels. Parvalbumin is the main allergen in both groups, with higher enolase sensitization in Japan. Monosensitization to parvalbumin is common	(48)
Cross-sectional study	China, Austria, Spain, Norway, Luxembourg	Fish-allergic individuals	Not specified	Parvalbumin (PVs) from various bony fish species	- 74.7% sensitization rate to PV overall - 19% (total) and up to 38% (China) negative to cod PV - 25% negative to sole PV - 78% negative to ray PV	Chinese patients show greater PV-sIgE diversity than Austrian, Spanish, or Norwegian patients, independent of symptoms, age, or gender. Up to 41% of fish-allergic patients may tolerate one bony fish based on PV-sIgE absence. Negative IgE to tuna PV predicts $\geq 90\%$ likelihood of negative IgE to other beta-PVs, excluding salmon (71%), aiding tailored FC panels	(42)
Retrospective study	Japan	Wheat allergy	311	wheat extract, $\omega 5G$	Wheat sIgE 10.1 threshold sensitivity 61%, specificity 74%, $\omega 5G$ sIgE 0.41 threshold sensitivity 72%, specificity 79%	$\omega 5G$ sIgE has better sensitivity than wheat sIgE	(58)
Cross-sectional study	Thailand	Wheat allergy	30	SPT wheat, wheat extract, $\omega 5G$	SPT accuracy 76.6%, wheat sIgE(threshold 0.35) accuracy 90%, $\omega 5G$ sIgE (threshold 0.35) accuracy 76.7%	$\omega 5G$ sIgE has better sensitivity than wheat sIgE (83.3% vs. 50%)	(57)

Ara h2 performed best in Singapore children (0.35 kUA/L threshold, sensitivity 91%, specificity 87%), with both requiring higher thresholds than international standards to achieve a 95% PPV. These findings are based on OFCs conducted in all 300 children, highlighting the robust nature of the diagnostic comparisons.¹¹⁶ Other geographical areas with a Western culture, such as Europe and North America, demonstrate similar sensitivity and specificity to Australia. Sensitivity and specificity are influenced by disease prevalence, which is lower for peanut allergy in Asia (0.27% (95% CI 0.12–0.42) compared to Australia (3.1% (95% CI 2.3–4.1)).^{25,26}

Pollen food allergy syndrome (PFAS) causes oral tingling, itching or soreness of the mouth, ears, or throat when consuming raw plant foods.²⁷ It occurs due to homology between pollen allergens (such as profilins, lipid transfer proteins, and pathogenesis-related class 10 proteins) and food proteins.²⁸ The related oral allergy syndrome (OAS) is a similar phenomenon, but PFAS could be considered a more severe presentation.²⁹ Peanuts have been described in PFAS worldwide, including in Australia, but are uncommon in Asia.^{28,30} Diagnostic recommendations include component sIgE tests such as Ara h 8 sIgE to diagnose PFAS or Ara h 1, 3, and 6 to diagnose IgE-mediated allergy in the absence of a positive Ara h 2 sIgE.²⁷ Given the low prevalence of PFAS in Asian countries and the poor diagnostic accuracy of these components, they are less relevant in diagnostic testing for this area. However, recent studies in this region suggest that in some countries, PFAS may be more prevalent, affecting as many as 10% of adolescents.²⁹ Locally relevant pollen allergens such as *Cyr j 2* or *Art v 3*, in conjunction with *Bet v 1* and *Bet v 2*, may be diagnostically useful.^{29,31} (Also see Table 2 and Pollen allergens below).

3.2 | Shrimp allergy

Research has shown geographic and phenotypic variability in shrimp allergy, particularly in the Asia-Pacific region. The first major allergen identified in crustacean shellfish was tropomyosin, and sensitization to tropomyosin is strongly linked to the incidence of shrimp anaphylaxis.^{32,33} This muscle protein has been considered a good predictor of shrimp allergy, with positive and negative predictive values of 0.72 and 0.91, respectively.³⁴ Tropomyosin remains an important panallergen in the Asia-Pacific region, with sensitization rates around 60% in doctor-diagnosed shrimp allergic patients.³⁵ However, sensitization to Pen a 1 (tropomyosin) varies with geography, with rates ranging from 40% in challenge-proven shrimp allergic patients from China to 82.8%–98% in challenge-proven patients from Brazil and Spain.^{34,36} A study that compared the sensitization patterns and diagnostic values of shrimp allergens in two distinct populations—subjects with challenge-proven or doctor-diagnosed shrimp allergy from Hong Kong ($n=69$) and Thailand ($n=16$), found leading shrimp allergens associated with clinical manifestations of shrimp allergy were sarcoplasmic calcium-binding protein (SCP), troponin C, fatty acid-binding protein, hemocyanin, and the newly identified high molecular weight allergen, glycogen phosphorylase.³⁷ These allergens had greater diagnostic accuracy for shrimp allergy compared to tropomyosin, based

on area under the curve (AUC) values of 0.77–0.96 compared to 0.70 for tropomyosin.³⁵ Hemocyanin, a heat-stable, high molecular weight allergen mainly found in the cephalothorax, was associated with anaphylaxis³⁸ and consistently identified as a clinically important allergen in Italian and Thai shrimp allergic subjects.^{39,40} The recent report indicated that tropomyosin did not appear to be a major allergen for shrimp allergy in Central Europe, particularly in patients with a house dust mite allergy.⁴¹ On the other hand, SCP was considered to be more clinically important, as reported in multi-center studies in Asia and the United States.^{35,41} Emerging research suggests that novel shrimp allergen components may have potential clinical utility in diagnosing shrimp allergies and correlating with patients' clinical presentations. Additional studies will be necessary to fully elucidate the clinical implications of these emerging allergen markers and their relationship to different shrimp allergy phenotypes.

3.3 | Fish allergy

Fish allergy diagnostics in the Asia-Pacific region differ from Western countries, with regional variations in the prevalence and relevance of specific allergens like parvalbumin (PV), collagen, and enolase. Parvalbumin has been known as the major fish allergen in bony fishes for more than 50 years and is included in routine diagnostics. While PV was the sole fish allergen (100% sensitization) in patients from China when tested against carp, mackerel, salmon, swordfish and tuna, up to one-third of patients from Austria, Denmark, Luxembourg, Norway, and Spain were IgE-negative to these PVs.⁴² The varying cross-reactivity among patients may stem from variations in IgE-binding epitopes across PV isoforms.⁴³ For instance, seven PV isoforms were identified in Indian mackerel,⁴⁴ while Japanese scad exhibited four distinct allergenic PVs.⁴⁵ Enolase, aldolase and collagen are other important fish allergens beyond PV, alongside other less characterized allergens such as triosephosphate isomerase (sensitization 19%–34%), tropomyosin (6%–32%), pyruvate kinase, and glyceraldehyde-3-phosphate dehydrogenase (GAPDH).⁴⁶ Unlike the high sensitization rates to aldolase and enolase reported in Europe (50% and 62.9% respectively),⁴⁷ aldolase and enolase were less relevant fish allergens in Asia-Pacific regions. Sensitization to these allergens ranged from 15% in Japanese patients to 35% in Australian patients and could be as low as 4.3% in patients from Hong Kong.^{46,48} Collagen, on the other hand, has been consistently reported as clinically important in the Asia-Pacific region. Apart from being a common sensitizer (21% in Australia, 30.4%–43% in Hong Kong, 17.8%–50% in Japan), monosensitization to collagen was also reported in up to 38% patients.^{48–51} These overall highlight the need to include collagen beyond parvalbumin in the diagnostic workup in the Asia-Pacific region.

3.4 | Wheat allergy

Wheat allergy, including wheat-dependent exercise-induced anaphylaxis (WDEIA), is best diagnosed using specific allergens like omega-5

TABLE 2 Frequency of sIgE reactivity with pollen allergen components in patients with allergic diseases from the Asia-Pacific region.

Study type	Country	Cohort	Number of participants	Testing method
Retrospective	Peking (temperate) China	history of pollinosis with OAS; 68% FA; 10% drug allergy	402 (Mean 30.4 yr, range 8–84; 47% M)	ImmunoCAP,
Retrospective ^a	Beijing, (temperate) China	Allergy department outpatients with pollen allergy	547 Mean age: 28.4 yr SD 14.3 yr M 49.4%	ImmunoCAP
Cross-sectional ^a	Guangzhou, (subtropical) China	Group 1: CCD positive patients Group 2: CCD negative patients	78 (34.6% <18 yr, 44.9% 18–60 yr, 20.5% >60 yr 72% M)	EUROBlotMaster
Cross-sectional ^a	Beijing, (temperate) China	birch pollen allergy with AR, +/- PFAS to apple	58 median 30 (17.8–38.2) yr; 53.4%	ImmunoCAP
Cross-sectional	Guangzhou, (subtropical) China	AD patient serum randomly taken from the serum bank (Jan 2020 to May 2020)	34 Mean age: 18.5 M: 47%	ImmunoCAP, ALEX
Observational ^a	Guangzhou, (subtropical) China	physician-diagnosed mild to moderate AR with/without asthma	258 patients, 88 controls (74.1% >40 yr 60.3% M)	ImmunoCap
Cross-sectional	Guangzhou, (subtropical) China	Any allergic disease with history of contact and sIgE reactivity to at least two pollen allergens	165 (69 <18 yr; mean 7.21 ± 4.26 yr, 96 >18 yr (mean 47.24 ± 15.53 yr) 67.2% M)	ImmunoCAP screening; CRD by EuroImmune
Cross-sectional/cohort	Tokyo, (temperate) Japan	General adolescent cohort checked for pollen allergy; 56.5% AR; 16% OAS; 11.7% PFAS (T-child Study)	506 13 yr M: 49.2%	ISAC
Prospective birth cohort	Japan (Mostly temperate)	General population, School children	984 aged 5 yrs. (2008 and 2010), and 729 aged 9 yrs. (2012 and 2014) 49.8% M	ISAC, ImmunoCAP
Case-control	Machida, (subtropical) Japan	patients with birch pollen allergy-related OAS to apple or peach	30 OAS, 10 controls (median 36.5 yr, range 18–79 yr; 50% M)	ImmunoCAP
Retrospective	Seoul, (temperate) South Korea	birch pollen-sensitized, +/- OAS, or history of anaphylaxis to peanut.	81 Birch pollen-sensitized. 12 with peanut anaphylaxis (Mean 30.3 yr ± 15.3 M/F: 50:50)	ImmunoCAP, SPT
Cross-sectional	Seoul, (temperate) South Korea	A: 18 patients no OAS B: 13 patients with OAS after apple or peach	31 (Mean 8.61 ± 2.81 yr; 67.7% M)	ImmunoCAP, ISAC
Cross-sectional	Seoul, (subtropical) South Korea	allergy patients, plus 12 negative controls	50 (Mean 23 yr, 6–66 yr; 62% M)	ELISA ImmunoCAP, BAT
Cross-sectional	Seoul, (subtropical) Korea	<i>Humulus japonicus</i> (Cannabaceae) weed pollinosis symptoms, plus 22 healthy controls	22 Mean 43.5 yr 12–70 yr; 41% M	SPT or ImmunoCAP screening, ELISA, Immunoblot
Cross-sectional ^a	Queensland Sydney, Perth and Adelaide (temperate - subtropical), Australia	159 GP-allergic AR patients; 50 other allergy donors; 29 non-atopic controls	238 adults (Median 39 yr; IQR: 33.1% M)	ImmunoCAP, SA-ImmunoCAP
Retrospective ^a	Brisbane, (subtropical) Australia	grass pollen-allergic AR patients	48 adults Median 36.5 yr IQR 28.8–44.5 yr 29.7% M	ImmunoCAP ELISA
Cross-sectional ^a	Melbourne, (temperate) Australia	seasonal AR, with or without asthma	51 adults	ELISA sIgE, sIgE cross-reactivity, BAT

Abbreviations: AD, atopic dermatitis; AR, allergic rhinitis; av, average; BAT, basophil activation test; CCD, cross-reactive carbohydrate components; CRD, component-resolved diagnostics; ELISA, enzyme-linked immunosorbent assay; FA, food allergy; GP, grass pollen; ISAC, Immuno-solid-phase Allergen Chip; LTP, lipid transfer proteins; PFAS, pollen food allergy syndrome; PR-10, pathogenesis-related class 10 proteins; SA, streptavidin; sIgE, specific IgE; SPT, skin prick test; yr, years.

^aArticles reporting on pollen CRD in AR in Asia-Pacific were identified with the following search terms: Allergic rhinitis, Rhinitis, Pollen, Allergy, Allergen, CRD, Component-resolved diagnosis, Prevalence, Weed, Grass, Tree, Group 1 allergen, Asia-Pacific, China, Japan, Korea, Australia, Malaysia, Taiwan, and India in PubMed, Google Scholar, and Scopus.

Pollen source/Allergens tested	Frequency of splgE to allergen	Key findings	Ref.
Birch, mugwort, Bet v 1, Bet v 2, Art v 1, Art v 3, Pru p 1, Pru p 3	In PFAS: Art v 3: 79% Bet v 1: 85%	Art v 3 splgE levels might indicate food anaphylaxis in OAS	(31)
Grass, tree and weeds, Bet v 1, Bet v 2, Bet v 4, Amb a 1, Art v 1, Phl p 1, Cyn d 1, Phl p 5b, CCD	Cyn d 1: 34% Phl p 1: 7% Phl p 5b: 2% CCD: 11%	71.1% AR patients sensitized to grass pollen; Bermuda splgE highest (97%), CRD is useful for diagnostics	(95)
Bermuda grass Timothy grass Mugwort, recombinant Cyn d 1, Cyn d 12, Phl p 1, Phl p 12, Art v 4, and nArt v 1, CCD	rCyn d 1: 47.8% in CCD -ve vs. 14.5% in CCD +ve; rPhl p 1: 26.1% in CCD -ve vs. 7.3% in CCD +ve Art v 4: 26.1%	AR patients (esp children) had higher rCyn d 1 splgE; all Cyn d 12 +ve patients had asthma; CRD splgE patterns varied by age	(117)
birch pollen allergen, bet v 1, Bet v 2, Bet v 4 and Bet v 6	Bet v 1: 82.8%, Bet v 2: 29.3%, Bet v 6: 1.7%, Bet v 4: 0%	77.6% splgE to any component; 19% to two; level Bet v 1 splgE associated with PFAS	(118)
Cypress, cedar Ryegrass, ash, 125 allergens	Cup s 1: 14.7% Cry j 1: 14.7% Lol p 1: 11.8% Fra e 1: 8.8%	70.6% positive for >1, 52.9% for >2, 17.6% for >5, and 2.9% for >10 components, Der p1 (20.6%) was highest	(94)
Bermuda, Timothy	Cyn d 1: 53.4%, Phl p 1: 17.1%, Phl p 5: 8.6%, Phl p 4: 41.4% of BeGP; 100% of TGP positive	A higher BeGP-splgE (22.5% v TGP 13.6%); Cyn d 1 splgE may be important, but CCD may confound	(119)
Mugwort, Ragweed, goosefoot, walnut, birch, CCD, allergen molecules	Art v 1: 18.0%, Art v 2 and 3: 24.6% Amb a 1: 7.5% Bet v 2: 40%	Weed pollen splgE higher in adults; extracts strongly correlated with CCD; splgE with allergens not high (Amb a 1: 7.5% - Bet v 2: 40%)	(92)
Japanese cedar, Cypress, white birch, alder, Ragweed, timothy grass, 112 components	Cry j 1: 95.7% Cup a 1: 86.0% Bet v 1: 36.0%; Be tv 2: 10.5%; Bet v 4: 1.5% Aln g 1: 28.3% Amb a 1: 27.9% Phl p 1: 9.0%	72.7% sensitized to pollen; PFAS is common (10%) in adolescents in Japan; kiwi, pineapple and peach common for PFAS, most had poor knowledge about PFAS.	(29)
Japanese cedar Cypress, 112 allergens	Cry j 1: 32.8% at 5 yr 57.8% at 9 yr Cup a 1: 12.3% at 5 yr 46.1% at 9 yr	74.8% sensitized at 9 yrs.; Cry j 1 highest frequency overall; rates of AR and CRD splgE increased with age	(120)
birch pollen, apple, peach, PR-10, profilin, LTP	Bet v 1: 93.3% Bet v 2: 6.7% OAS; Mal d 1: 92.3%; Pru p 1 95.7%	CRD more sensitive and specific than extracts for the diagnosis of birch pollen-related OAS	(85)
Birch and oak pollen	Bet v 1: 82% Bet v 2: 19.7%. 4 with OAS also had Ara h 2 and Ara h 8 splgE	Fagales pollen may influence rates of OAS; Bet v 1, Ara h 2 and Ara h 8 splgE may indicate OAS	(121)
birch pollen, Bet v 1, Bet v 2, and Bet v 4	Bet v 1: A, 61% vs. B, 100% Bet v 2: A, 0% vs. B, 31% Bet v 4: both A and B: 0%	CRD sensitization profiles differed according to the presence of OAS	(91)
Mongolian Oak	Que m 1: 92% Que. a 1: 74% Bet v 1: 38%	Que m 1 and Bet v 1 BAT similar; Que. m 1 may be useful for the diagnosis of tree pollinosis	(99)
Japanese hop (<i>Humulus japonicus</i>)	Hum j 6: 86.4%	Hum j 6 identified as major weed allergen	(98)
Bahia grass, Bio-nPas n 1	Pas n 1: 92.4%	Bahia GP is an important pollen in subtropics; Pas n 1 IgE accounts for most of BaGP-splgE.	(122)
Johnson grass (<i>Sorghum halepense</i>)	Sor h 1: 97.5% Sor h 13: 43.8%	Sor h 1, Sor h 13 are important allergens; Sor h 2 and Sor h 23, but no group 5, allergen present	(123)
Bahia, Bermuda and ryegrass, nPas n 1, nCyn d 1, nLol p 1	Extract frequencies 90%–92%; Lol p 1 splgE > Pas n 1 or Cyn d 1	Sensitization to Ryegrass pollen and Lol p 1 higher in temperate	(124)

gliadin (ω 5G), with in-house extracts offering higher diagnostic accuracy compared to commercial wheat extracts, especially in the Asia-Pacific region. Wheat protein can be classified into either water/salt-soluble albumins and globulins or water/salt-insoluble gliadins and glutenins.⁵² Omega-5 gliadin was identified as the major allergen for IgE-mediated wheat allergy.^{53,54} Recently, a study from Thailand showed that alpha/beta/gamma gliadins were important allergens in children with wheat anaphylaxis.^{55,56} The commercial wheat extract typically contains a mixture of various wheat proteins, which may dilute the allergenic components, including gliadins, leading to a lower diagnostic accuracy. In contrast, the in-house extract is customized to focus more on gliadins and glutenins, which are the key allergens, improving the sensitivity and diagnostic accuracy of the test.⁵⁷ The accuracy for wheat allergy diagnosis by the commercial extract was 76.6% compared to 86.7% using the in-house extract.⁵⁷

Levels of ω 5G-splgE potentially have better diagnostic capacity than wheat splgE, with AUC being slightly greater for ω 5G (78.5%) compared to wheat splgE (73%); sensitivity at the optimal cut-point was also higher for ω 5G (72%) compared to wheat splgE (61%).^{58,59}

WDEIA is a specific clinical phenotype of food allergies. Omega-5 gliadin, gamma gliadin, and high molecular weight glutenin were previously reported as major allergens in WDEIA patients.^{60,61} These allergens have been highlighted in several studies conducted in the Asia-Pacific region. For instance, in Japan, ω 5G is recognized as a critical allergen in WDEIA, with a cut-off value of 0.89 kUA/L for ω 5G, the sensitivity for diagnosing WDEIA is 78%, while the specificity is 96%.⁶² In Thailand, the in-house gliadin/glutenin extract generally has a better diagnostic yield for WDEIA diagnosis than the commercial skin SPT extract.⁶³ A new subtype of WDEIA with reactions to hydrolyzed wheat protein (HWP-WDEIA) has been reported in Japan. Compared to conventional WDEIA, patients with HWP-WDEIA are more sensitized to gluten but not ω 5G-splgE.⁶⁴

3.5 | Fruit allergies

Fruit allergies in the Asia-Pacific region exhibit diverse patterns. Largely based on self-report and/or physician diagnosis but without OFCs, the prevalence of fruit allergy ranged from 0.1 to 4.3% in 2009,⁶⁵ with a recent review showing it now varies from 0.029% to 8%, influenced by settings, populations, and case definitions.⁶⁶ The Rosaceae family (peach, apple, apricot, pear, raspberry, strawberry) and non-Rosaceae fruits, such as kiwifruit, are common causative fruits in Europe.^{67,68} Since the types of tropical fruits vary depending on the region and climate zone, a diverse pattern of Rosaceae and non-Rosaceae allergies is observed in the Asia-Pacific region—Rosaceae fruit allergies are common in Japan,⁶⁹ mango allergies in East Asia (China and Taiwan),^{70–73} and banana and kiwi allergies in Southeast Asia (Thailand),^{74,75} South Asia (India),⁷⁶ and West Asia (Iran, Israel, Turkey).^{77–80} In contrast, a recent US survey (2015–2016) found fruit allergies to be rare, likely due to the exclusion of cases with OAS.^{9,81}

Fruit allergies can range from localized symptoms to anaphylaxis, often due to primary sensitization to pollen allergens, with

cross-reactive IgE binding to plant food, known as Pollen Food Allergy Syndrome (PFAS). Lipid Transfer Protein (LTP) syndrome, a condition involving allergic reactions from at least two unrelated LTP food allergens, is now recognized in countries like China, Japan, South Africa, and Australia.⁸² Latex-fruit syndrome (LFS) is another entity that involves allergic reactions to both latex and various fruits, such as avocado, banana, kiwifruit, and chestnut.⁸³ Class 1 chitinases (Hev b 6) play a significant role in LFS and exhibit high sequence homology with chitinases in related fruits.

Compared to Western countries, there is a paucity of molecular sensitization data on fruit allergies in the Asia-Pacific region. So far, few studies on fruit allergy CRD have been conducted. In China, the major allergenic proteins in mango are Man i 1 (class IV chitinase), Man i 2 (Bet v 1-related protein), and Man i 4 (profilin).⁸⁴ In a large Japanese cohort, Bet v 1 homologs and profilin were sensitized in most cases, indicating the PFAS mechanism. Gibberellin-regulated protein (GRP), represented by peach Pru p 7, is common in non-PFAS patients, while LTP is found in a minority of patients.⁶⁹ The finding is similar to another Japanese group.⁸⁵

Further molecular epidemiology studies are needed for diagnostic utility and interpretation of CRD results in the Asia-Pacific region, as allergen components are specific to certain fruit species, some of which are unique to this region.

4 | COMPONENT-RESOLVED DIAGNOSTICS FOR AEROALLERGENS

Asthma and AR affect millions of children globally, but many asthma and AR patients are not investigated for underlying aeroallergen sensitizations. Timely diagnosis of aeroallergen sensitizations in children with airway allergies can predict asthma development, medication response, and future exacerbation, and is important in guiding personalized treatment plans and the initiation of allergen-specific immunotherapy.^{86,87}

4.1 | Pollens

Pollen is considered a primary cause of allergic rhinitis (AR) in some regions of Asia-Pacific. For instance, the frequency of sensitization to Japanese cedar (*Cryptomeria japonica*) pollen is as high as 68% of the population tested, and this pollen causes AR in as many as 40% of people in Japan.⁸⁸ However, in tropical regions, pollen allergy may be less important; in Malaysia, Taiwan and Thailand, the sensitization of AR patients to grass pollen ranges from 6.5 to 20.5%, 2.5 to 10%, and 16 to 21%, respectively, depending on species tested (Bermuda; *Cynodon dactylon*, Johnson; *Sorghum halepense*, or Bahia; *Paspalum notatum*).⁸⁹ House dust mite showed the highest sensitization rates in southern China, with a subtropical climate, whereas weed pollens of mugwort, ragweed, and dandelion showed the highest frequencies in northwestern China, where the climate is temperate.⁹⁰

Climate, topography, and other biogeographical conditions influence plant distribution, species diversity, pollen abundance, and therefore, pollen allergenicity profiles. Diagnosis has largely relied on SPT with extracts of pollen, particularly those relevant to temperature climate zones of Europe. This may underrepresent true sensitization, and more importantly, fail to reveal the specific pollen sources associated with a person's symptoms in the Asia-Pacific region. Patients with seasonal AR or allergic asthma are often exposed to multiple different pollens, which may contain common allergen families; for example, beta-expansins (Pas n 1, Cyn d 1) and polygalacturonase (Pas n 13) in grass pollen, profilin (e.g., Bet v 2) and pectate lyase (e.g., Cry j 1) in tree pollen, and defensins (Art v 1, Amb a 4) in weed pollen, leading to apparent poly-sensitization by SPT or splgE with multiple pollen extracts. The presence of panallergens in multiple pollen sources, such as profilin (e.g., Phl p 12), polcalcin (e.g., Phl p 7 or Bet v 4), LTP (e.g., Art v 3), and PR-10 (e.g., Bet v 1), may not only confound diagnosis of pollinosis but, in some cases, may be associated with OAS and PFAS in this region, including China,³¹ Japan,²⁹ and Korea⁹¹ (Table 2). A number of pollen allergens, like natural Cyn d 1, Phl p 4, and Cry j 2, contain cross-reactive carbohydrate determinants (CCDs),⁴ potentially confounding diagnosis.⁹² Studies from the Philippines, where grass pollen allergy is low, have shown for unselected allergy patients that grass pollen splgE detected using timothy or Bermuda grass pollen allergens, including natural Cyn d 1 and Phl p 4, can be blocked with other components containing CCDs.⁹³ Use of recombinant allergens, rPas n 1 and rCyn d 1 lacking CCDs, or comparison of allergen component with CCDs may therefore improve specific diagnosis of pollen allergy.^{94–96}

Table 2 collates data from studies within this geographic region that report on splgE to allergen components of tree, weed and grass pollens in general populations, AR patients with or without asthma, OAS, or PFAS.

Considering migration, patients who relocate to a new region may experience changes in pollen exposure and symptoms if pollens to which they were sensitized in their previous residence do not fully overlap with the allergens present in their new environment.⁹⁷ There are continuing processes of characterization of locally relevant pollen allergens including Hum j 6 of Japanese hop,⁹⁸ Que. m 1 of Mongolian oak,⁹⁹ Uro m 1 of Para grass,¹⁰⁰ Zoy m 1 of Manila grass,¹⁰¹ and a number of *Artemisia* allergens related to Mugwort pollen from northern China that have been listed by the International Union of Immunological Society Allergen Committee.¹⁰² It will be important to see further studies on sensitization rates in relevant areas and their clinical importance for particular patient groups across the Asia-Pacific region.

4.2 | House dust mite allergy

Dermatophagoides pteronyssinus (Der p) and *D. farinae* (Der f) are the most common house dust mites (HDM) worldwide. The allergenic reactivity of HDM has been well characterized to be dominated by the Group 1 (Der p 1 and Der f 1) and Group 2 (Der p 2

and Der f 2) allergen components, while Der p 23, a putative chitin-binding protein, is a newly recognized major allergen in HDM that has been found associated with severe asthma and allergic rhinitis symptoms.^{103–105} Sensitization to *Blomia tropicalis* (Blo t) was prevalent in Southeast Asia, particularly Singapore and Malaysia, even more so than to Der p and Der f.^{106,107} Comparison of studies conducted in the West and the East often points to similar sensitization rates and patterns to these allergens, with IgE binding to these allergens well exceeding 50% in HDM-sensitized cohorts. For instance, a recent study in Japan indicated 94.2% and 97.5% sensitization to Der p 1 and Der p 2, respectively,¹⁰⁸ compared to 73% and 80% in a multinational study in Canada, Europe, South Africa, and The States and 74% to >90% in Southern China.^{109,110} The sensitization rate to Der p 23 was 64% in the West, 54.6% in Southern China and 71.7% in Japan. Yet, a comparison of the sensitization profile of cockroach-sensitized individuals showed remarkably lower sensitization to Der p 1 (35%), Der p 2 (48%), Der p 10 (15%) and Der p 23 (31%) in Austria than those from Hong Kong (97%, 88%, 52%, and 79% respectively).¹¹¹ In this context, HDM sensitization highly correlates with shellfish sensitization in warm, humid climates and urban environments where HDM are ubiquitous.¹¹² A prospective study also indicated that HDM sensitization at 18 months was associated with increased odds of shellfish sensitization at 8 years.¹¹³ Der p 10, tropomyosin, has been initially recognized as the allergen responsible for cross-reactivity between HDM and shellfish due to the high sequence and structural homology. Despite its low prevalence among HDM-sensitized patients between 5% and 18%, sensitization to Der p 10 is strongly associated with a history of anaphylaxis to shrimp and shellfish.¹¹⁴ Der p 20, arginine kinase, as well as hemocyanin and paramyosin, are other allergens beyond tropomyosin reported to be involved in the shellfish–HDM interrelationship.¹¹⁵ Yet, whether shellfish allergy is a secondary phenomenon that follows HDM sensitization or a true primary allergy remains an open question to be answered.

5 | CONCLUSION

Component-resolved diagnostics (CRD) are becoming a crucial tool for allergists in research and clinical practice. This review describes the geographical variances in CRD for food and aeroallergens in the Asia-Pacific region and prompts the need to adopt region-specific and tailored strategies in precision diagnosis. While peanut components, such as Ara h 2, significantly improve diagnostic accuracy in Western countries where PFAS is prevalent, their utility in Asia may be limited. Tropomyosin is a major shellfish allergen, especially in the Asia-Pacific region, but it may not be the sole or primary allergen in all populations. Other emerging allergens, such as SCP and troponin C, may be better indicators of clinical shrimp allergy than tropomyosin in some cases.

Pollen is a common cause of allergic rhinitis in parts of Asia-Pacific, but the specific pollen allergens differ by region due to factors like climate and geography. Reliance on pollen extracts

from temperate regions may not capture all relevant sensitivities. The presence of cross-reactive allergen components like profilins and polcalcins in multiple pollen sources can confound the diagnosis of pollinosis and lead to apparent poly-sensitization. Besides, HDM is a major aeroallergen globally, with Der p 1, Der p 2, Der p 23, and Blo t 5 being the dominant allergens in the Asia-Pacific region.

CRD evidently helps in identifying and understanding sensitization to allergens that are unique to particular clinically important allergen sources, which may be more or less prevalent or show biogeographically distinct prognostic utility in the Asia-Pacific region. This information is important for developing region-specific allergy management guidelines and public health policies. We must move forward by adapting CRD panels to fit the regional context and ensuring improved access to and adoption of CRD testing. Additionally, we must prioritize region-specific research and data sharing to expand our understanding of sensitization patterns' epidemiology in diverse Asian populations and ensure the availability for diagnostic use of regionally relevant, well-defined allergen components of food, and aeroallergen sources.

AUTHOR CONTRIBUTIONS

Carmen Riggioni: Conceptualization; writing – original draft; writing – review and editing; supervision. **Agnes Sze-Yin Leung:** Conceptualization; writing – original draft; writing – review and editing; supervision. **Christine Yee-Yan Wai:** Writing – original draft; writing – review and editing. **Janet M. Davies:** Writing – original draft; writing – review and editing. **Mongkhon Sompornrattanaphan:** Writing – original draft; writing – review and editing. **Punchama Pacharn:** Writing – review and editing; writing – original draft. **Sajjad Chamani:** Writing – review and editing; writing – original draft. **Tim Brettig:** Writing – original draft; writing – review and editing. **Rachel L. Peters:** Writing – original draft; writing – review and editing; conceptualization; supervision.

CONFLICT OF INTEREST STATEMENT

The authors declare that they have no conflict of interest.

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REFERENCES

- Dang TD, Peters R, Neeland MR, et al. Ana o 3 sIgE testing increases the accuracy of cashew allergy diagnosis using a two-step model. *Pediatr Allergy Immunol*. 2022;33(1):e13705.
- Dang TD, Tang M, Choo S, et al. Increasing the accuracy of peanut allergy diagnosis by using Ara h 2. *J Allergy Clin Immunol*. 2012;129(4):1056-1063.
- Riggioni C, Ricci C, Moya B, et al. Systematic review and meta-analyses on the accuracy of diagnostic tests for IgE-mediated food allergy. *Allergy*. 2024;79(2):324-352.
- Dramburg S, Hilger C, Santos AF, et al. EAACI molecular allergy user's guide 2.0. *Pediatr Allergy Immunol*. 2023;34(Suppl 28):e13854.
- Matricardi PM, Kleine-Tebbe J, Hoffmann HJ, et al. EAACI molecular Allergy User's guide. *Pediatr Allergy Immunol*. 2016;27(Suppl 23):1-250.
- Su KW, Yan DC, Ou LS, et al. Rising prevalence of food allergies in Taiwan: an epidemiological study. *J Microbiol Immunol Infect*. 2023;56(4):863-870.
- Ma Z, Chen L, Xian R, Fang H, Wang J, Hu Y. Time trends of childhood food allergy in China: three cross-sectional surveys in 1999, 2009, and 2019. *Pediatr Allergy Immunol*. 2021;32(5):1073-1079.
- Osborne NJ, Koplin JJ, Martin PE, et al. Prevalence of challenge-proven IgE-mediated food allergy using population-based sampling and predetermined challenge criteria in infants. *J Allergy Clin Immunol*. 2011;127(3):668-676.e1-2.
- Gupta RS, Warren CM, Smith BM, et al. The public health impact of parent-reported childhood food allergies in the United States. *Pediatrics*. 2018;142(6):e20181235. doi:10.1542/peds.2018.1235
- Leung ASY, Tham EH, Pacharn P, et al. Asia-Pacific Research Network for Anaphylaxis (APRA). Disparities in pediatric anaphylaxis triggers and management across Asia. *Allergy*. 2024;79(5):1317-1328. doi:10.1111/all.16098
- Ebisawa M, Ito K, Fujisawa T. Japanese guidelines for food allergy 2020. *Allergol Int*. 2020;69(3):370-386.
- Feng H, Luo N, Xiong X, Wu Y. Prevalence of food allergy in the Chinese population: a systematic review and meta-analysis of population-based studies. *Allergy Asthma Proc*. 2023;44(5):315-325.
- Tham EH, Lee BW, Chan YH, et al. Low food allergy prevalence despite delayed introduction of allergenic foods-data from the GUSTO cohort. *J Allergy Clin Immunol Pract*. 2018;6(2):466.
- Peters RL, Soriano VX, Allen KJ, et al. The prevalence of IgE-mediated food allergy and other allergic diseases in the first 10 years: the population-based, Longitudinal HealthNuts Study. *J Allergy Clin Immunol Pract*. 2024;12(7):1819-1830.e3. doi:10.1016/j.jaip.2024.03.015
- Koplin JJ, Peters RL, Ponsonby AL, et al. Increased risk of peanut allergy in infants of Asian-born parents compared to those of Australian-born parents. *Allergy*. 2014;69(12):1639-1647.
- Savoure M, Bousquet J, Jaakkola JJK, Jaakkola MS, Jacquemin B, Nadif R. Worldwide prevalence of rhinitis in adults: a review of definitions and temporal evolution. *Clin Transl Allergy*. 2022;12(3):e12130.
- Lai CK, Beasley R, Crane J, Foliaki S, Shah J, Weiland S. Global variation in the prevalence and severity of asthma symptoms: phase three of the International Study of Asthma and Allergies in Childhood (ISAAC). *Thorax*. 2009;64(6):476-483.
- Li X, Song P, Zhu Y, et al. The disease burden of childhood asthma in China: a systematic review and meta-analysis. *J Glob Health*. 2020;10(1):010801.
- Silverberg JI, Barbarot S, Gadkari A, et al. Atopic dermatitis in the pediatric population: a cross-sectional, international epidemiologic study. *Ann Allergy Asthma Immunol*. 2021;126(4):417-428.e2.
- Langan SM, Mulick AR, Rutter CE, et al. Trends in eczema prevalence in children and adolescents: a global asthma network phase I study. *Clin Exp Allergy*. 2023;53(3):337-352.
- Santos AF, Riggioni C, Agache I, et al. EAACI guidelines on the diagnosis of IgE-mediated food allergy. *Allergy*. 2023;78(12):3057-3076.

22. Santos AF, Riggioni C, Du Toit G, Skypala I. An algorithm for the diagnosis and management of IgE-mediated food allergy, 2024 update. *Allergy*. 2024;80(2):629-632. doi:[10.1111/all.16321](https://doi.org/10.1111/all.16321)
23. Greenhawt M, Shaker M, Wang J, et al. Peanut allergy diagnosis: a 2020 practice parameter update, systematic review, and GRADE analysis. *J Allergy Clin Immunol*. 2020;146(6):1302-1334.
24. Brettig T, Dang T, McWilliam V, Peters RL, Koplin JJ, Perrett KP. The accuracy of diagnostic testing in determining tree nut allergy: a systematic review. *J Allergy Clin Immunol Pract*. 2021;9(5):2028-2049.e2.
25. Lee AJ, Tham EH, Goh AEN, et al. Prevalence of IgE-mediated cow milk, egg, and peanut allergy in young Singapore children. *Asia Pac Allergy*. 2022;12(3):e31.
26. Soriano V, Peters R, Ponsonby A-L, et al. Has the prevalence of Peanut allergy changed following earlier introduction of Peanut? The EarlyNuts study. *J Allergy Clin Immunol*. 2021;147(2 Suppl):AB236.
27. Skypala IJ, Hunter H, Krishna MT, et al. BSACI guideline for the diagnosis and management of pollen food syndrome in the UK. *Clin Exp Allergy*. 2022;52(9):1018-1034.
28. Skypala IJ, Cecchi L, Shamji MH, Scala E, Till S. Lipid transfer protein allergy in the United Kingdom: characterization and comparison with a matched Italian cohort. *Allergy*. 2019;74(7):1340-1351.
29. Kiguchi T, Yamamoto-Hanada K, Saito-Abe M, et al. Pollen-food allergy syndrome and component sensitization in adolescents: a Japanese population-based study. *PLoS One*. 2021;16(4):e0249649.
30. Brown CE, Katelaris CH. The prevalence of the oral allergy syndrome and pollen-food syndrome in an atopic paediatric population in south-west Sydney. *J Paediatr Child Health*. 2014;50(10):795-800.
31. Li JD, Du ZR, Liu J, Xu YY, Wang RQ, Yin J. Characteristics of pollen-related food allergy based on individual pollen allergy profiles in the Chinese population. *WAO J*. 2020;13:100120.
32. Tong WS, Yuen AW, Wai CY, Leung NY, Chu KH, Leung PS. Diagnosis of fish and shellfish allergies. *J Asthma Allergy*. 2018;11:247-260.
33. Miraglia Del Giudice M, Dinardo G, Klain A, et al. Anaphylaxis after shrimp intake in a European pediatric population: role of molecular diagnostics and implications for novel foods. *Children (Basel)*. 2023;10(10):1583. doi:[10.3390/children10101583](https://doi.org/10.3390/children10101583)
34. Gamez C, Sanchez-Garcia S, Ibanez MD, et al. Tropomyosin IgE-positive results are a good predictor of shrimp allergy. *Allergy*. 2011;66(10):1375-1383.
35. Wai CY, Leung NYH, Leung ASY, et al. Comprehending the allergen repertoire of shrimp for precision molecular diagnosis of shrimp allergy. *Allergy*. 2022;77:3041-3051.
36. Pascal M, Grishina G, Yang AC, et al. Molecular diagnosis of shrimp allergy: efficiency of several allergens to predict clinical reactivity. *J Allergy Clin Immunol Pract*. 2015;3(4):521-529.e10.
37. Wai CY, Leung PSC. Emerging approaches in the diagnosis and therapy in shellfish allergy. *Curr Opin Allergy Clin Immunol*. 2022;22(3):202-212.
38. Guillen D, Fiandor A, del Pozo V, et al. Anaphylaxis caused by hemocyanin contained in shrimp cephalothorax. *Ann Allergy Asthma Immunol*. 2014;113(6):674.
39. Piboonpocanun S, Jirapongsananuruk O, Tipayanon T, Boonchoo S, Goodman RE. Identification of hemocyanin as a novel non-cross-reactive allergen from the giant freshwater shrimp *Macrobrachium rosenbergii*. *Mol Nutr Food Res*. 2011;55(10):1492-1498.
40. Giuffrida MG, Villalta D, Mistrello G, Amato S, Asero R. Shrimp allergy beyond tropomyosin in Italy: clinical relevance of arginine kinase, sarcoplasmic calcium binding protein and hemocyanin. *Eur Ann Allergy Clin Immunol*. 2014;46(5):172-177.
41. Grilo J, Vollmann U, Aumayr M, Sturm GJ, Bohle B. Tropomyosin is no accurate marker allergen for diagnosis of shrimp allergy in Central Europe. *Allergy*. 2022;77(6):1921-1923.
42. Kalic T, Kuehn A, Aumayr M, et al. Identification of potentially tolerated fish species by multiplex IgE testing of a multinational fish-allergic patient cohort. *J Allergy Clin Immunol Pract*. 2022;10(12):3284-3292.
43. Sharp MF, Lopata AL. Fish allergy: in review. *Clin Rev Allergy Immunol*. 2014;46(3):258-271.
44. Ruethers T, Raith M, Sharp MF, et al. Characterization of Ras k 1 a novel major allergen in Indian mackerel and identification of parvalbumin as the major fish allergen in 33 Asia-Pacific fish species. *Clin Exp Allergy*. 2018;48(4):452-463. doi:[10.1111/cea.13069](https://doi.org/10.1111/cea.13069)
45. Misnan R, Murad S, Jones M, et al. Identification of the major allergens of Indian scad (*Decapterus russelli*). *Asian Pac J Allergy Immunol*. 2008;26(4):191-198.
46. Ruethers T, Taki AC, Karnaneedi S, et al. Expanding the allergen repertoire of salmon and catfish. *Allergy*. 2021;76(5):1443-1453.
47. Kuehn A, Hilger C, Lehnert-Weber C, et al. Identification of enolases and aldolases as important fish allergens in cod, salmon and tuna: component resolved diagnosis using parvalbumin and the new allergens. *Clin Exp Allergy*. 2013;43(7):811-822.
48. Wai CY, Leung NYH, Leung ASY, et al. Differential patterns of fish sensitization in Asian populations: implication for precision diagnosis. *Allergol Int*. 2023;72(3):458-465.
49. Kobayashi Y, Akiyama H, Hige J, et al. Fish collagen is an important panallergen in the Japanese population. *Allergy*. 2016;71(5):720-723.
50. Kalic T, Kamath SD, Ruethers T, et al. Collagen – an important fish allergen for improved diagnosis. *J Allergy Clin Immunol Pract*. 2020;8(9):3084-3092.e10.
51. Kobayashi Y, Hige J, Imamura S, Hamada-Sato N. Study of the cross-reactivity of fish allergens based on a questionnaire and blood testing. *Allergol Int*. 2016;65(3):272-279.
52. Inomata N. Wheat allergy. *Curr Opin Allergy Clin Immunol*. 2009;9(3):238-243.
53. Palosuo K, Varjonen E, Kekki OM, et al. Wheat omega-5 gliadin is a major allergen in children with immediate allergy to ingested wheat. *J Allergy Clin Immunol*. 2001;108(4):634-638.
54. Daengsuwan T, Palosuo K, Phankingthongkum S, et al. IgE antibodies to omega-5 gliadin in children with wheat-induced anaphylaxis. *Allergy*. 2005;60(4):506-509.
55. Pacharn P, Kumjim S, Tattiyapong P, Jirapongsananuruk O, Piboonpocanun S. Identification of wheat sensitization using an in-house wheat extract in Coca-10% alcohol solution in children with wheat anaphylaxis. *Asian Pac J Allergy Immunol*. 2016;34(2):153-158.
56. Rutrakool N, Piboonpocanun S, Wangthan U, et al. Children with wheat anaphylaxis and with low wheat specific IgE have a different IgE immunoblot pattern than those with high wheat specific IgE. *Asian Pac J Allergy Immunol*. 2023. doi:[10.12932/AP-140622-1387](https://doi.org/10.12932/AP-140622-1387)
57. Pacharn P, Siripipattanamongkol N, Pannakapitak N, et al. Accuracy of in-house alcohol-dissolved wheat extract for diagnosing IgE-mediated wheat allergy. *Asian Pac J Allergy Immunol*. 2020;38(2):102-107.
58. Ebisawa M, Shibata R, Sato S, Borres MP, Ito K. Clinical utility of IgE antibodies to omega-5 gliadin in the diagnosis of wheat allergy: a pediatric multicenter challenge study. *Int Arch Allergy Immunol*. 2012;158(1):71-76.
59. Ito K, Futamura M, Borres MP, et al. IgE antibodies to omega-5 gliadin associate with immediate symptoms on oral wheat challenge in Japanese children. *Allergy*. 2008;63(11):1536-1542.
60. Matsuo H, Kohno K, Niihara H, Morita E. Specific IgE determination to epitope peptides of omega-5 gliadin and high molecular weight glutenin subunit is a useful tool for diagnosis of wheat-dependent exercise-induced anaphylaxis. *J Immunol*. 2005;175(12):8116-8122.
61. Piboonpocanun S, Thongngarm T, Wongsas C, Pacharn P, Reamtong O, Sompornrattanaphan M. Omega-5 and gamma gliadin are the

- major allergens in adult-onset IgE-mediated wheat allergy: results from Thai cohort with oral food challenge. *J Asthma Allergy*. 2021;14:907-917.
62. Matsuo H, Dahlstrom J, Tanaka A, et al. Sensitivity and specificity of recombinant omega-5 gliadin-specific IgE measurement for the diagnosis of wheat-dependent exercise-induced anaphylaxis. *Allergy*. 2008;63(2):233-236.
 63. Thongngarm T, Wongs C, Pacharn P, Piboonpocanun S, Sompornrattanaphan M. Clinical characteristics and proposed wheat-cofactor challenge protocol with a high diagnostic yield in adult-onset IgE-mediated wheat allergy. *J Asthma Allergy*. 2020;13:355-368.
 64. Chinuki Y, Morita E. Wheat-dependent exercise-induced anaphylaxis sensitized with hydrolyzed wheat protein in soap. *Allergol Int*. 2012;61(4):529-537.
 65. Zuidmeer L, Goldhahn K, Rona RJ, et al. The prevalence of plant food allergies: a systematic review. *J Allergy Clin Immunol*. 2008;121(5):1210-1218.e4.
 66. Krikeerati T, Rodsaward P, Nawiboonwong J, Pinyopornpanish K, Phusawang S, Sompornrattanaphan M. Revisiting fruit allergy: prevalence across the globe, diagnosis, and current management. *Foods*. 2023;12(22):4083. doi:10.3390/foods12224083
 67. Burney P, Summers C, Chinn S, Hooper R, van Ree R, Lidholm J. Prevalence and distribution of sensitization to foods in the European Community Respiratory Health Survey: a EuroPrevall analysis. *Allergy*. 2010;65(9):1182-1188.
 68. Burney PG, Potts J, Kummeling I, et al. The prevalence and distribution of food sensitization in European adults. *Allergy*. 2014;69(3):365-371. doi:10.1111/all.12341
 69. Inomata N, Miyakawa M, Aihara M. High prevalence of sensitization to gibberellin-regulated protein (peamaclein) in fruit allergies with negative immunoglobulin E reactivity to bet v 1 homologs and profilin: clinical pattern, causative fruits and co-factor effect of gibberellin-regulated protein allergy. *J Dermatol*. 2017;44(7):735-741.
 70. Luo J, Zhang Q, Gu Y, et al. Meta-analysis: prevalence of food allergy and food allergens - China, 2000-2021. *China CDC Wkly*. 2022;4(34):766-770.
 71. Feng H, Zhou J, Lu Y, et al. Prevalence of self-reported food allergy among adults in Jiangxi, China. *World Allergy Organ J*. 2023;16(5):100773.
 72. Feng H, Luo N, Lu Y, et al. Prevalence of parent-reported food allergy among children in China: a population-based cross-sectional survey. *Front Immunol*. 2022;13:982660.
 73. Zeng GQ, Luo JY, Huang HM, et al. Food allergy and related risk factors in 2540 preschool children: an epidemiological survey in Guangdong Province, southern China. *World J Pediatr*. 2015;11(3):219-225.
 74. Sompornrattanaphan M, Ajalasaereewong S, Wongs C, et al. Prevalence and characteristics of adult patients with adult-onset and childhood-onset food allergy. *Asian Pac J Allergy Immunol*. 2023. doi:10.12932/AP-210223-1548
 75. Julanon N, Thiravetyan B, Unhapitpong C, et al. Not just a banana: the extent of fruit cross-reactivity and reaction severity in adults with banana allergy. *Food Secur*. 2023;12(13):2456. doi:10.3390/foods12132456
 76. Mahesh PA, Wong GW, Ogorodova L, et al. Prevalence of food sensitization and probable food allergy among adults in India: the EuroPrevall INCO study. *Allergy*. 2016;71(7):1010-1019. doi:10.1111/all.12868
 77. Ahanchian H, Jafari S, Behmanesh F, et al. Epidemiological survey of pediatric food allergy in Mashhad in Northeast Iran. *Electron Physician*. 2016;8(1):1727-1732. doi:10.19082/1727
 78. Nachshon L, Schwartz N, Elizur A, et al. The prevalence of food allergy in young Israeli adults. *J Allergy Clin Immunol Pract*. 2019;7(8):2782-2789.e4.
 79. Akarsu A, Ocak M, Koken G, Sahiner UM, Soyer O, Sekerel BE. IgE mediated food allergy in Turkey: different spectrum, similar outcome. *Turk J Pediatr*. 2021;63(4):554-563.
 80. Kaya A, Erkokoglu M, Civelek E, Cakir B, Kocabas CN. Prevalence of confirmed IgE-mediated food allergy among adolescents in Turkey. *Pediatr Allergy Immunol*. 2013;24(5):456-462.
 81. Gupta RS, Warren CM, Smith BM, et al. Prevalence and severity of food allergies among US adults. *JAMA Netw Open*. 2019;2(1):e185630.
 82. Breiteneder H, Ebner C. Molecular and biochemical classification of plant-derived food allergens. *J Allergy Clin Immunol*. 2000;106(1 Pt 1):27-36.
 83. Sompornrattanaphan M, Kreetapirom P, Srinoulprasert Y, et al. Severe anaphylaxis after pelvic examination: a case report of dual latex and chlorhexidine allergies. *Allergy Asthma Clin Immunol*. 2019;15:19.
 84. Zhao J, Camus-Ela M, Zhang L, et al. A comprehensive review on mango allergy: clinical relevance, causative allergens, cross-reactivity, influence of processing techniques, and management strategies. *Compr Rev Food Sci Food Saf*. 2024;23(2):e13304.
 85. Shirasaki H, Yamamoto T, Abe S, Kanaizumi E, Kikuchi M, Himi T. Clinical benefit of component-resolved diagnosis in Japanese birch-allergic patients with a convincing history of apple or peach allergy. *Auris Nasus Larynx*. 2017;44(4):442-446.
 86. Casale TB, Pedersen S, Del Rodriguez Rio P, Liu AH, Demoly P, Price D. The role of aeroallergen sensitization testing in asthma management. *J Allergy Clin Immunol Pract*. 2020;8(8):2526-2532.
 87. Dykewicz MS, Wallace DV, Amrol DJ, et al. Rhinitis 2020: a practice parameter update. *J Allergy Clin Immunol*. 2020;146(4):721-767.
 88. Minami T, Fukutomi Y, Inada R, et al. Regional differences in the prevalence of sensitization to environmental allergens: analysis on IgE antibody testing conducted at major clinical testing laboratories throughout Japan from 2002 to 2011. *Allergol Int*. 2019;68(4):440-449.
 89. Davies J. Grass pollen allergens globally; the contribution of subtropical grasses to burden of allergic respiratory diseases. *Clin Exp Allergy*. 2014;44:790-801.
 90. Lou H, Ma S, Zhao Y, et al. Sensitization patterns and minimum screening panels for aeroallergens in self-reported allergic rhinitis in China. *Sci Rep*. 2017;7(1):9286.
 91. Baek HS, Jeong JW, Lee HB, Kwon JW. Molecular sensitization patterns in birch pollen-sensitized Korean children according to the presence of oral allergy syndrome. *Medicine (Baltimore)*. 2020;99(10):e19469.
 92. Xu L, Luo W, Lu Y, et al. A comprehensive analysis of the components of common weed pollen and related allergens in patients with allergic diseases in southern China. *Mol Immunol*. 2022;147:180-186.
 93. Cabauatan CR, Lupinek C, Scheiblhofer S, et al. Allergen microarray detects high prevalence of asymptomatic IgE sensitizations to tropical pollen-derived carbohydrates. *J Allergy Clin Immunol*. 2014;133(3):910-914.e5.
 94. Hu H, Hou X, Luo W, et al. The molecule sensitized pattern of atopic dermatitis patients who co-sensitized to shrimp, cockroaches, crab and house dust mites. *J Asthma Allergy*. 2021;14:993-997. doi:10.2147/JAA.S326411
 95. Li JD, Gu JQ, Xu YY, et al. Serum IgE profiles in Chinese pollinosis patients with grass pollen sensitisation. *World Allergy Organ J*. 2022;15(1):100624.
 96. Davies JM, Pralong C, Tickner J, et al. Nanofluidic point-of-care IgE test for subtropical grass pollen for rapid diagnosis of allergic rhinitis. *Ann Allergy Asthma Immunol*. 2024;132:479-504.
 97. Kailaivasan TH, Timbrell VL, Solley G, et al. Biogeographical variation in specific IgE recognition of temperate and subtropical grass pollen allergens in allergic rhinitis patients. *Clin Transl Immunology*. 2020;9(2):e01103.

98. Jeong KY, Sang M, Lee YS, Gadermaier G, Ferreira F, Park JW. Characterization of hum j 6, a major allergen from *Humulus japonicus* pollen, the primary cause of weed Pollinosis in East Asia. *Allergy Asthma Immunol Res*. 2023;15(6):767-778.
99. Lee JY, Yang M, Jeong KY, et al. Characterization of a major allergen from Mongolian oak, *Quercus mongolica*, a dominant species of oak in Korea. *Int Arch Allergy Immunol*. 2017;174(2):77-85.
100. Dhammachat S, Somkid K, Piboonpocanun S, et al. Isoforms of group 1 allergens from a tropical/subtropical Para grass (*Urochloa mutica*) display different levels of IgE reactivity and cross-reactivity. *Eur Ann Allergy Clin Immunol*. 2019;51(4):174-185. doi:10.23822/EurAnnACI.1764-1489.95
101. Somkid K, Aud-In S, Pinkaew B, Tantilipikorn P, Piboonpocanun S, Songnuan W. Manila grass (*Zoysia matrella*) Zoy m 1 allergen may contribute to allergic sensitization in tropical/subtropical regions due to extensive cross-reactivity with other group-1 grass pollen allergens. *Asian Pac J Allergy Immunol*. 2021. doi:10.12932/AP-250920-0971
102. Pomes A, Davies JM, Gadermaier G, et al. WHO/IUIS allergen nomenclature: providing a common language. *Mol Immunol*. 2018;100:3-13.
103. Thomas WR. Hierarchy and molecular properties of house dust mite allergens. *Allergol Int*. 2015;64(4):304-311.
104. Jiménez-Feijoo R, Pascal M, Moya R, et al. Molecular diagnosis in house dust mite-allergic patients suggests that Der p 23 is clinically relevant in asthmatic children. *J Invest Allergol Clin Immunol*. 2020;30(2):127-132. doi:10.18176/jiaci.0431
105. Gao Y, Li J, Xu X, Wang C, Zhang Y, Zhang L. Sensitisation to house dust mite component Der p 23 is associated with severe symptoms and asthma in allergic rhinitis patients. *Int Arch Allergy Immunol*. 2023;184(9):906-913.
106. Yeoh SM, Kuo IC, Wang DY, et al. Sensitization profiles of Malaysian and Singaporean subjects to allergens from *Dermatophagoides pteronyssinus* and *Blomia tropicalis*. *Int Arch Allergy Immunol*. 2003;132(3):215-220.
107. Wong QYA, Lim JJ, Ng JY, et al. Allergic rhinitis in Chinese young adults from the Singapore/Malaysia Cross-Sectional Genetics Epidemiology Study (SMCGES) cohort: prevalence, patterns, and epidemiology of allergic rhinitis. *World Allergy Organ J*. 2022;15(10):100704.
108. Hasegawa A, Utsumi D, Lund K, Okano M, Ohashi-Doi K, Okubo K. Correlation between sensitization to house dust mite major allergens, age, and symptoms in Japanese house dust mite allergic subjects. *Int Immunopharmacol*. 2022;107:108640.
109. D'Souza N, Weber M, Sarzsinszky E, et al. The molecular allergen recognition profile in China as basis for allergen-specific immunotherapy. *Front Immunol*. 2021;12:719573. doi:10.3389/fimmu.2021.719573
110. Muddaluru V, Valenta R, Vrtala S, et al. Comparison of house dust mite sensitization profiles in allergic adults from Canada, Europe, South Africa and USA. *Allergy*. 2021;76(7):2177-2188. doi:10.1111/all.14749
111. Mittermann I, Lupinek C, Wieser S, et al. IgE reactivity patterns in Asian and central European cockroach-sensitized patients reveal differences in primary sensitizing allergen sources. *J Allergy Clin Immunol Glob*. 2022;1(3):145-153.
112. Wang J, Calatroni A, Visness CM, Sampson HA. Correlation of specific IgE to shrimp with cockroach and dust mite exposure and sensitization in an inner-city population. *J Allergy Clin Immunol*. 2011;128(4):834-837.
113. Loo EXL, Lau HX, Suaini NHA, et al. House dust mite sensitization, eczema, and wheeze increase risk of shellfish sensitization. *Pediatr Allergy Immunol*. 2021;32(5):1096-1099.
114. Indolfi C, Dinardo G, Klain A, et al. Evaluation of Der p 10 in a cohort of European children: role of molecular diagnostics and clinical features. *J Immunol Res*. 2023;2023:5551305.
115. Asero R, Pravettoni V, Scala E, Villalta D. House dust mite-shrimp allergen interrelationships. *Curr Allergy Asthma Rep*. 2020;20(4):9.
116. Tham E, Riggioni C, Leung ASY. Regional disparities in utility of component-resolved diagnostics for peanut allergic patients in Asia. *J Allergy Clin Immunol*. 2025;155(2):AB45.
117. Liao C, Liang CC, Hu H, et al. Major pollen allergen components and CCD detection in Bermuda grass sensitized patients in Guangzhou, China. *J Asthma Allergy*. 2020;13:615-623.
118. Wang XY, Chen LJ, Ding JQ, Wang HT, Wang XY. Profiles of birch allergen component sensitization and its association with pollen food allergy syndrome in Northern China. *J Asthma Allergy*. 2023;16:1241-1250.
119. Luo WT, Huang HM, Zheng PY, et al. Major grass pollen allergens and components detected in a southern Chinese cohort of patients with allergic rhinitis and/or asthma. *Mol Immunol*. 2016;78:105-112.
120. Yamamoto-Hanada K, Borres MP, Aberg MK, et al. IgE responses to multiple allergen components among school-aged children in a general population birth cohort in Tokyo. *World Allergy Organ J*. 2020;13(2):100105.
121. Park KH, Son YW, Lee SC, et al. Clinical significance of component allergens in Fagales pollen-sensitized peanut allergy in Korea. *Allergy Asthma Immun*. 2016;8(6):505-511.
122. Timbrell VL, Riebel L, Simmonds C, et al. An immunodiagnostic assay for quantitation of specific IgE to the major pollen allergen component, Pas n 1, of the subtropical Bahia grass. *Int Arch Allergy Immunol*. 2014;165(4):219-228.
123. Campbell BC, Gilding EK, Timbrell V, et al. Total transcriptome, proteome, and allergome of Johnson grass pollen, which is important for allergic rhinitis in subtropical regions. *J Allergy Clin Immunol*. 2015;135(1):133-142.
124. Davies JM, Dang TD, Voskamp A, et al. Functional immunoglobulin E cross-reactivity between Pas n 1 of Bahia grass pollen and other group 1 grass pollen allergens. *Clin Exp Allergy*. 2011;41(2):281-291.

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