

# Synthesis of biocompatible iron oxide nanoparticles as a drug delivery vehicle

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**Abstract:** Over the last decade, there has been growing interest in developing novel nanoparticles (NPs) for biomedical applications. A safe-by-design approach was used in this study to synthesize biocompatible iron oxide NPs. The size of the particles obtained was ~100 nm. Although these NPs were significantly ( $P < 0.05$ ) internalized in MCF-7 (human breast adenocarcinoma cell line) cells, no adverse effect was observed in the cells as assessed by cytotoxicity assays (neutral red uptake and 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) and cell cycle analysis. Our data demonstrate the potential of iron oxide NPs as a biocompatible carrier for targeted drug delivery. **Keywords:** biocompatible iron oxide nanoparticles, human breast adenocarcinoma cells, coprecipitation

## Introduction

Iron-based magnetic nanoparticles (NPs) such as magnetite ( $\text{Fe}_3\text{O}_4$ ) have been studied in detail due to their unique properties, such as stability over time, biocompatibility, and sensitivity to magnetic field.<sup>1-3</sup> They can potentially be used as magnetic targeted drug delivery carriers and magnetic resonance imaging contrast agents due to their high saturation magnetization, low toxicity, and biocompatibility.<sup>4</sup> The magnetic properties of  $\text{Fe}_3\text{O}_4$  NPs are attributed to their size and size distribution, which, in turn, is dependent on the route of synthesis.

Therefore, in this study, an attempt was made to synthesize  $\text{Fe}_3\text{O}_4$  NPs using a safe-by-design approach by the coprecipitation method. Polyethylene glycol (PEG) was used as surfactant to control the particle size and narrow size distribution. The biocompatibility of  $\text{Fe}_3\text{O}_4$  NPs was evaluated by cytotoxicity assays and cell cycle analysis in the human breast adenocarcinoma cell line (MCF-7).

## Materials and methods

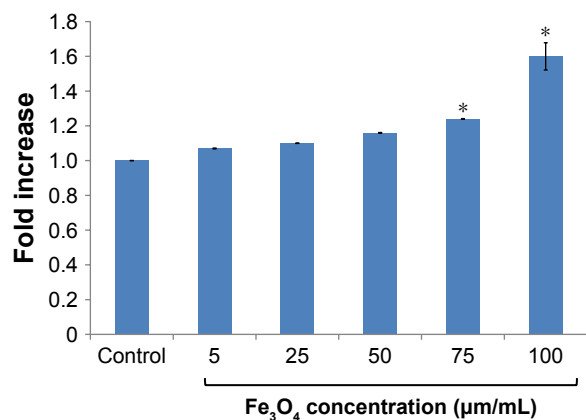
### Materials

Ferric chloride hexahydrate and ferrous sulfate were purchased from SD-Fine-Chem. Ltd, Mumbai, India. Polyethyleneglycol (PEG-6000), dimethylsulphoxide, sodium hydroxide (NaOH), minimum essential medium eagle, phosphate-buffered saline, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), and antibiotic-antimycotic solution were purchased from HiMedia Laboratories Pvt. Ltd., (Mumbai, India). The MCF-7 cell line was purchased from the National Centre for Cell Sciences, Pune, India.

### $\text{Fe}_3\text{O}_4$ NP synthesis

The preparation of  $\text{Fe}_3\text{O}_4$  NPs was performed by a chemical coprecipitation method of  $\text{Fe}^{2+}$  and  $\text{Fe}^{3+}$  ions (1:2 molar ratios) by the addition of NaOH.<sup>5</sup> A total volume of 15 mL

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**Figure 1** Internalization of Fe<sub>3</sub>O<sub>4</sub> NPs in MCF-7 cells using flow cytometry.  
**Notes:** Data are expressed as mean ± standard error of the mean from three independent experiments. \**P*<0.05, when compared with control.  
**Abbreviations:** NPs, nanoparticles; MCF-7, human breast adenocarcinoma cell line.

of 0.25 M Fe<sup>2+</sup> and 0.5 M Fe<sup>3+</sup> solutions were prepared in deionized water. PEG was then added and the temperature slowly risen up to 80°C. During the initial 2 minutes of the reaction, NaOH was added to achieve a pH of 10. The reaction was allowed to continue on a magnetic stirrer for 2 hours. Thereafter, the suspension was centrifuged and washed several times with deionized water to lower the pH to 7. Finally, the particles were suspended in 10 mL of dimethylsulphoxide.

## Characterization of Fe<sub>3</sub>O<sub>4</sub> NPs

One milliliter of the stock suspension of Fe<sub>3</sub>O<sub>4</sub> NPs was diluted in 10 mL complete minimum essential medium eagle. The hydrodynamic size and zeta potential were determined using Zetasizer Nano ZS.

## Cytotoxicity assessment

The cytotoxic potential of Fe<sub>3</sub>O<sub>4</sub> NPs was assessed in MCF-7 cells after 6 and 24 hours of treatment using MTT and neutral

red uptake (NRU) assays as described by Mosmann<sup>6</sup> and Borenfreund and Puerner, respectively.<sup>7</sup>

## Cellular internalization of NPs

The internalization of Fe<sub>3</sub>O<sub>4</sub> NPs in MCF-7 cells was assessed according to the method described in our earlier study.<sup>8</sup>

## Cell cycle analysis

The effect of Fe<sub>3</sub>O<sub>4</sub> NPs on cell cycle was assessed according to the method described in our earlier study.<sup>8</sup>

## Results and discussion

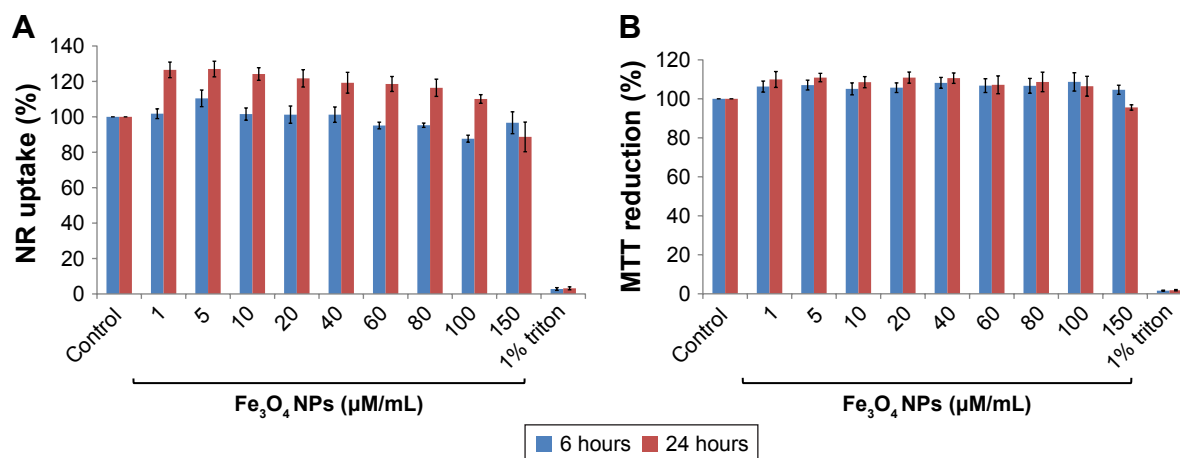
The mean hydrodynamic size and zeta potential of synthesized Fe<sub>3</sub>O<sub>4</sub> NPs were 98.19±1.0 nm and 36±2 mV, respectively. Flow cytometric analysis revealed a significant (*P*<0.05) increase in the internalization of Fe<sub>3</sub>O<sub>4</sub> NPs in MCF-7 cells after 24 hours exposure at the two higher concentrations, as evident by an increase in the side scatter intensity (Figure 1).

In the cytotoxicity assays, namely NRU and MTT, Fe<sub>3</sub>O<sub>4</sub> NPs were found to be biocompatible as there was no significant increase in the NRU (88% at concentration 150 µM/mL) and a reduction (96%) in the mitochondrial succinate dehydrogenase activity was observed at the highest concentration after 24-hour exposure (Figures 2A and B).

Additionally, no change in cell cycle progression was observed in Fe<sub>3</sub>O<sub>4</sub> NPs-treated MCF-7 cells, after 24-hour exposure (Figure 3).

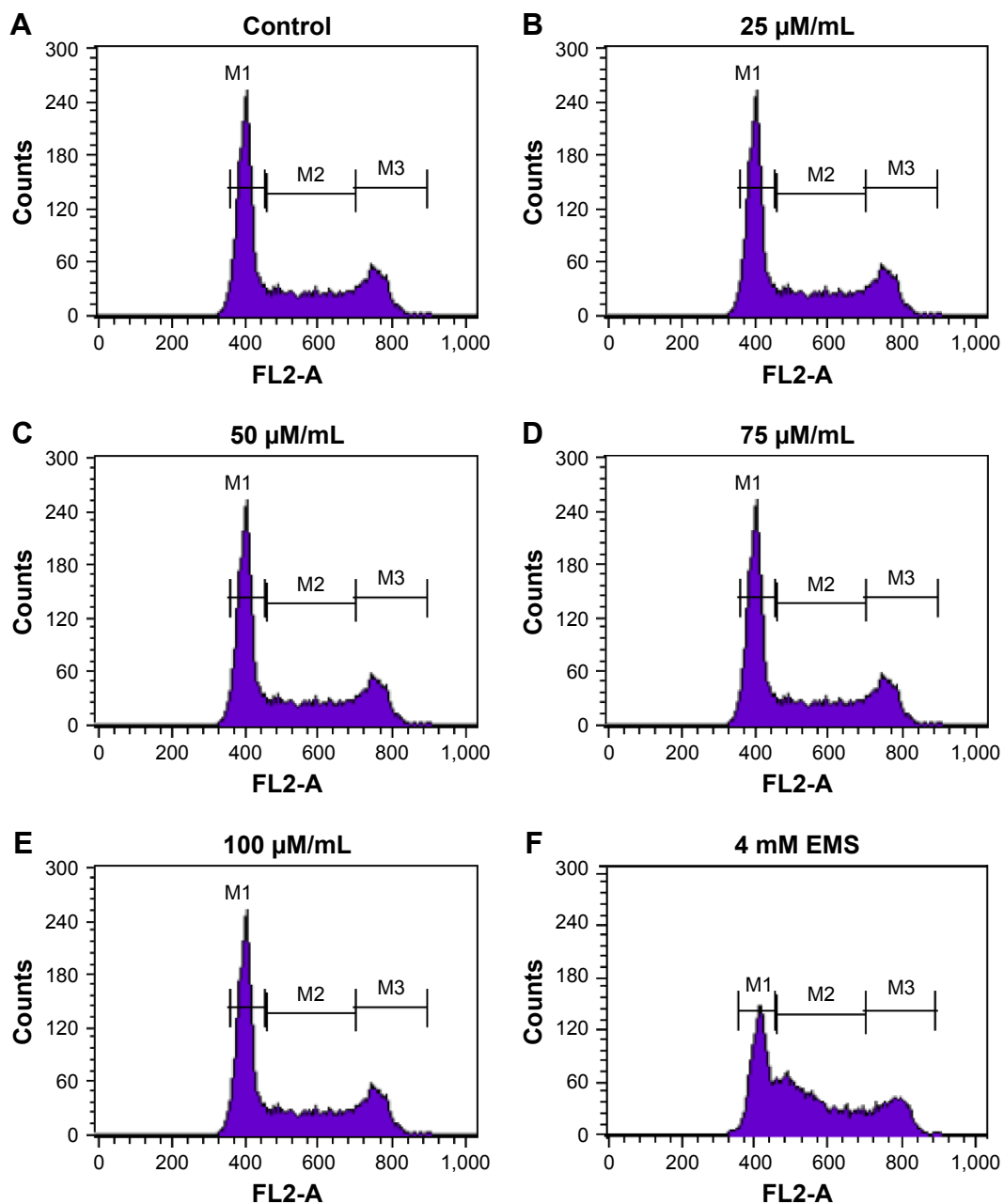
## Conclusion

Our results demonstrated that Fe<sub>3</sub>O<sub>4</sub> NPs synthesized using the safe-by-design approach showed no adverse effect on cells, as assessed by cytotoxicity assays and cell cycle analysis in MCF-7 cells, even though they are significantly



**Figure 2** Cytotoxicity of Fe<sub>3</sub>O<sub>4</sub> NPs in MCF-7 cells. (A) NR uptake (%); (B) MTT reduction (%).

**Notes:** The viability of the control cells was considered as 100%. Data are expressed as mean ± standard error of the mean from three independent experiments.  
**Abbreviations:** NPs, nanoparticles; MCF-7, human breast adenocarcinoma cell line; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NR, neutral red.



**Figure 3** Effect of  $\text{Fe}_3\text{O}_4$  NPs on cell cycle progression in MCF-7 cells (A) control (B–E) cells treated with different concentration of  $\text{Fe}_3\text{O}_4$  NPs and (F) cells treated with 4 mM EMS.

**Abbreviations:** EMS, ethylmethane sulfonate; NPs, nanoparticles; MCF-7, human breast adenocarcinoma cell line.

internalized. Therefore, these NPs have a potential to be used as a carrier for targeted drug delivery.

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## Disclosure

The authors report no conflicts of interest in this work.

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