

Review Article

Lippia javanica (Burm.f.) Spreng.: Traditional and Commercial Uses and Phytochemical and Pharmacological Significance in the African and Indian Subcontinent

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Received 5 September 2016; Revised 25 October 2016; Accepted 20 November 2016; Published 1 January 2017

Academic Editor: Rainer W. Bussmann

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Lippia javanica occurs naturally in central, eastern, and southern Africa and has also been recorded in the tropical Indian subcontinent. The potential of *L. javanica* as herbal or recreational tea and herbal medicine and its associated phytochemistry and biological properties are reviewed. The extensive literature survey revealed that *L. javanica* is used as herbal tea and has ethnomedicinal applications such as in colds, cough, fever, malaria, wounds, diarrhoea, chest pains, bronchitis, and asthma. Multiple classes of phytochemicals including volatile and nonvolatile secondary metabolites such as alkaloids, amino acids, flavonoids, iridoids, and triterpenes as well as several minerals have been identified from *L. javanica*. Scientific studies on *L. javanica* indicate that it has a wide range of pharmacological activities which include anticancer, antiamoebic, antidiabetic, antimalarial, antimicrobial, antioxidant, antiplasmodial, and pesticidal effects. Although many of the traditional uses of *L. javanica* have been validated by phytochemical and pharmacological studies, there are still some gaps where current knowledge could be improved. *Lippia javanica* is popular as both herbal and recreational tea, but there is need for more precise studies to evaluate the safety and clinical value of its main active crude and pure compounds and to clarify their mechanisms of action.

1. Introduction

Lippia javanica (Burm.f.) Spreng. (Verbenaceae) (Figure 1(a)) has a long history of traditional uses in tropical Africa as indigenous herbal tea or tisane (Figure 1(b)), refreshing beverage, or food additive based on its perceived health and medicinal properties. *Lippia javanica* is rich in volatile oil, particularly caryophyllene, carvone, ipsenone, ipsdienone, limonene, linalool, myrcene, myrcenone, ocimenone, *p*-cymene, piperitenone, sabinene, and tagetenone [1–5]. Research by Viljoen et al. [3] revealed that the essential oil profiles of *L. javanica* are characterized by inter- and intraspecies variations because they are produced by different metabolic pathways. Using cluster analysis, Viljoen et al. [3] identified five chemotypes of *L. javanica* in South Africa and Swaziland, myrcenone-rich type (36–62%), carvone-rich type (61–73%), piperitenone-rich type (32–48%), ipsdienone-rich type (42–61%), and linalool-rich type (>65%). The

relative proportion of the chemical constituents of *L. javanica* essential oil is important as this determines the biological properties of the species chemotypes.

Lippia javanica belongs to the *Verbena* or vervain family (Verbenaceae) comprising approximately 32 genera and 840 species [97]. The genus *Lippia* L. is named after Augustin Lippi (1678–1701), an Italian botanist and natural historian who was killed in Ethiopia at the age of 23 [98]. Phylogenetic relationships within family Verbenaceae demonstrated that genus *Lippia* and other closely related genera, namely, *Aloysia* Paláu, *Lantana* L., and *Phyla* Lour., are not monophyletic [97]. The boundaries separating these four genera are historically weak, with many taxonomic researchers including species belonging to *Aloysia*, *Lantana*, and *Phyla* in the genus *Lippia* [99]. *Lippia* and *Lantana* genera are the most difficult to separate, as species of these two genera show similarities in their inflorescences that are spicate, often subcapitate during anthesis and elongating in fruit and pedunculated

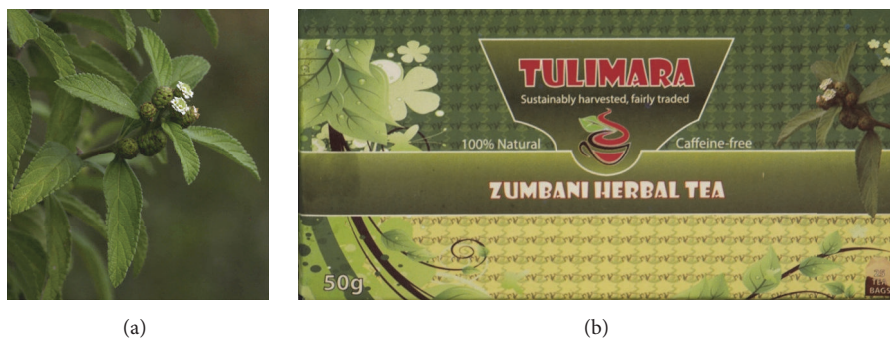


FIGURE 1: (a) *Lippia javanica* (Burm.f.) Spreng. flowers and leaves (photo: BT Wursten) and (b) *L. javanica* herbal tea traded as Zumbani in Zimbabwe (<http://specialityfoods.co.zw/product/zumbani/>).

[98]. According to de Campos et al. [100], the genus *Lippia* comprises about 200 species of herbs, shrubs, and small trees distributed throughout south and central America and tropical Africa. Only 15 *Lippia* species have been recorded in tropical Africa [101]. The specific name “javanica” was given by the Dutch Botanist Nicolaas Laurens Burman (1734–1793) in 1768, who mistakenly thought that the type specimen was collected in Java, Indonesia [102]. He placed the species in the genus *Verbena*, and the German botanist Sprengel (1766–1833) transferred Burman’s name to the genus *Lippia* in 1825 [103]. *Lippia javanica* is morphologically similar to *L. scaberrima* Sond. but is much taller and its bracts are shorter than the flowers (Figure 1(a)), while *L. scaberrima* has many stems arising from ground level and is usually less than 0.5 metres high, and its bracts are not longer than the flowers [104].

Lippia javanica is an erect woody perennial herb or shrub of up to 4.5 m tall, with strong aromatic leaves which give off a lemon-like fragrance when crushed [105]. Stems are brownish, usually erect or spreading with short stiff tubercle-based whitish hairs and small glands, and branched with inflorescences in nearly all axils. Leaves are opposite or in whorls of 3, blades lanceolate to oblong and densely pubescent, rounded and then cuneate at the base, and crenate-serrate or closely serrulate on the margins except near the leaf base [106]. Flowers occur in conical or oblong spikes, purple or dull-reddish in fruit, dark brownish on drying [101, 106]. The flowers are sessile or with short peduncles, lower bracts of spikes ovate and upper bracts smaller, pubescent, glandular, and densely spreading [101]. The calyx is 2-lobed, half as long as the corolla, pubescent, and densely spreading. The corolla is white, yellowish-white to greenish (Figure 1(a)), usually with a yellow throat, glandular and pubescent outside in the upper half, tube narrowly funnel-shaped from a narrow base [101, 106]. Anthers are sulphur-yellow and nutlets are brown and half ovoid [101, 106].

Lippia javanica occurs naturally in central, eastern, and southern Africa (Figure 2) and has also been recorded in the tropical Indian subcontinent [12, 13, 23, 39, 47, 61]. In sub-Saharan Africa, the species is native to Angola, Botswana, Central African Republic, Democratic Republic of Congo, Ethiopia, Kenya, Malawi, Mozambique, South Africa, Swaziland, Tanzania, Uganda, Zambia, Zanzibar, and Zimbabwe.



FIGURE 2: *Lippia javanica* naturally occurs in central, eastern, and southern Africa.

Lippia javanica has been recorded in low to high altitude (0–2350 m above sea level) woodlands and wooded grasslands, scrub bushland, and grassy rocky kopjes, in riverine vegetation, and on margins of dambos and swampy ground, sometimes on termite mounds, in montane grasslands, and on evergreen forest margins, also in disturbed ground beside roads, forest clearings, plantations, and cultivated land and becoming a weed in derived rangelands [101, 106]. This shows that the plant is highly adaptable to a wide range of climatic, soil, and vegetation conditions.

2. Traditional Uses of *Lippia javanica*

Lippia javanica is used for a wide variety of traditional uses (Table 1). Based on literature, the most important traditional applications include its uses as herbal tea and ethnomedicinal applications for (in descending order of importance) colds, cough, fever or malaria, wounds, repelling mosquitos, diarrhoea, chest pains, bronchitis, and asthma (Figure 3). These different uses are discussed in the following seven sections:

TABLE 1: Traditional uses of *Lippia javanica* in central, eastern, and southern Africa.

Use	Plant part(s) used	Mode of use	Country practised	Reference(s)
<i>Food uses</i>				
Food additive	Leaves, twigs	Leaves and twigs boiled together with maize, cassava, groundnuts, and common tea (<i>Camellia sinensis</i> (L.) Kuntze)	Kenya	[6]
Herbal tea	Leaves	Leaves can be sun dried for later use	Botswana, South Africa, and Zimbabwe	[7–11]
Leafy vegetable	Leaves	Eaten as vegetable with meat or fish	India	[12, 13]
<i>Respiratory problems</i>				
Asthma	Leaves, twigs	Decoction taken orally or smoke inhaled	South Africa, Zimbabwe	[14–17]
Blocked nose	Leaves	Leaf decoction of <i>L. javanica</i> alone taken orally or mixed with any of these species: <i>Brachylaena uniflora</i> Harv., <i>Clausena anisata</i> (Willd.) Hook.f. ex Benth., <i>Clematis brachiata</i> Thunb., <i>Combretum molle</i> R. Br. ex G. Don, <i>Eucalyptus grandis</i> W. Hill, <i>Helichrysum kraussii</i> Sch. Bip., <i>Krauseola mosambicina</i> Pax & Hoffm., <i>Plectranthus neochilus</i> Schltr., <i>Senecio serratuloides</i> DC., or <i>Trichilia emetica</i> Vahl taken orally	South Africa	[18]
Bronchitis	Leaves, roots, and stems	Leaf infusion of <i>L. javanica</i> alone taken orally or applied to skin as ointment or leaf and stem infusion of <i>L. javanica</i> mixed with leaves of <i>Artemisia afra</i> Jacq. ex Willd. taken orally	Botswana, South Africa	[15, 19–22]
Chest pains	Leaves, roots	Leaves and roots of <i>L. javanica</i> alone chewed or decoction taken orally or used in washes and poultices or smoke inhaled or “imbiza,” decoction prepared from <i>L. javanica</i> and <i>Cyrtanthus obliquus</i> (L. f.) Aiton, taken orally or leaf decoction of <i>L. javanica</i> mixed with leaves of any of these species: <i>Brachylaena transvaalensis</i> Hutch. ex E. Phillips & Schweick., <i>C. brachiata</i> , <i>E. grandis</i> , <i>H. kraussii</i> , or <i>T. emetica</i> taken orally or leaf juice taken orally with cloves of <i>Allium sativum</i> L.	Bangladesh, Ethiopia, and South Africa	[14, 18, 23–27]
Colds	Leaves, twigs	Leaf infusion of <i>L. javanica</i> alone taken orally or inhaled or applied as an ointment or “imbiza,” decoction prepared from <i>L. javanica</i> and <i>C. obliquus</i> , taken orally	Botswana, Kenya, South Africa, and Zimbabwe	[15, 16, 19, 21, 27–32]
Cough	Leaves, stems, and twigs	Leaves of <i>L. javanica</i> alone chewed or infusion inhaled or taken orally or applied as an ointment or leaf and stem infusion of <i>L. javanica</i> mixed with leaves of any of these species: <i>Acanthospermum glabratum</i> (DC.) Wild., <i>A. afra</i> , <i>B. uniflora</i> , <i>B. transvaalensis</i> , <i>Bridelia cathartica</i> Bertol., <i>C. anisata</i> , <i>C. brachiata</i> , <i>C. molle</i> , <i>E. grandis</i> , <i>H. kraussii</i> , <i>K. mosambicina</i> , <i>P. neochilus</i> , <i>Psidium guajava</i> L., <i>S. serratuloides</i> , <i>Terminalia sericea</i> Burch. ex DC., <i>Tetradenia riparia</i> (Hochst.) Codd, or <i>T. emetica</i> taken orally	Botswana, Ethiopia, South Africa, and Zimbabwe	[15, 16, 18–21, 26, 30, 32]
Influenza	Leaves, roots	Decoction taken orally	Mozambique, South Africa	[7, 33–36]
Lung infections	Leaves	Infusions taken orally	South Africa	[37]

TABLE 1: Continued.

Use	Plant part(s) used	Mode of use	Country practised	Reference(s)
Measles	Leaves, stems	Leaf decoction of <i>L. javanica</i> alone taken orally or body washed with infusion or leaf and stem infusion of <i>L. javanica</i> mixed with leaves of <i>A. afra</i> taken orally	Kenya, South Africa, and Zimbabwe	[14, 20, 38]
Pneumonia	Leaves	Ointment rubbed on chest and abdomen	Zimbabwe	[14]
Respiratory disorders	Leaves	Decoction taken orally	India, South Africa	[39–41]
Runny nose	Leaves, roots	Leaf and root decoction of <i>L. javanica</i> alone taken orally or leaf decoction of <i>L. javanica</i> mixed with leaves of any of these species: <i>B. transvaalensis</i> , <i>B. cathartica</i> , <i>C. anisata</i> , <i>C. brachiata</i> , <i>E. grandis</i> , <i>Hypoxis</i> spp., <i>K. mosambicina</i> , <i>P. guajava</i> , <i>S. serratuloides</i> , <i>P. neochilus</i> , or <i>T. emetica</i> taken orally	South Africa	[18]
Shortness of breath (dyspnoea)	Leaves	Decoction taken orally or body washed with decoction	Zimbabwe	[14]
Sore throat	Leaves	Leaf decoction of <i>L. javanica</i> alone taken orally or leaf decoction of <i>L. javanica</i> mixed with leaves of <i>E. grandis</i> or <i>T. riparia</i> taken orally	South Africa	[18]
Tonsillitis	Leaves	Leaf decoction of <i>L. javanica</i> alone taken orally or leaf decoction of <i>L. javanica</i> mixed with leaves of <i>E. grandis</i> taken orally	South Africa	[18]
Tuberculosis	Leaves	Leaf decoction of <i>L. javanica</i> alone taken orally or “imbiza,” decoction prepared from <i>L. javanica</i> and <i>C. obliquus</i> , taken orally	South Africa, Uganda	[24, 42, 43]
<i>Gastrointestinal diseases</i>				
Amoebiasis	Leaves, twigs	Decoction taken orally	Kenya	[6]
Anthelmintics	Leaf	Infusions taken orally	South Africa	[40]
Diarrhoea	Leaves, roots	Decoction taken orally	Kenya, Mozambique, South Africa	[17, 29, 34, 37, 44]
Gangrenous rectitis	Leaves	Infusions taken orally	South Africa	[45]
Prophylactic against diarrhoea	Leaves	Infusion taken orally	South Africa	[40]
Vomiting	Leaves	Decoction taken orally	Zanzibar, Tanzania	[46]
<i>Fever, malaria, and as insect repellent</i>				
Fever	Leaves, stems, and twigs	Leaf and twig decoction of <i>L. javanica</i> alone taken orally or leaf and stem infusion of <i>L. javanica</i> mixed with leaves of any of these species: <i>A. afra</i> , <i>B. transvaalensis</i> , <i>C. anisata</i> , <i>C. molle</i> , <i>E. grandis</i> , <i>P. guajava</i> , <i>S. serratuloides</i> , <i>T. riparia</i> , or <i>T. emetica</i> taken orally	South Africa, Zimbabwe	[14, 16, 18, 20]
Getting rid of lice, insects, and lice and as mosquito repellent	Leaves, twigs, and whole plant	Used in washes and poultices or as steam or sprayed or burnt to chase away mosquitoes	Ethiopia, South Africa, and Zimbabwe	[4, 26, 34, 47–51]
Malaria	Leaves, roots	Decoction taken orally	Mozambique, South Africa	[17, 33, 52]
Prophylactic against malaria	Leaves	Infusion taken orally	South Africa	[40]

TABLE I: Continued.

Use	Plant part(s) used	Mode of use	Country practised	Reference(s)
<i>Wounds, injuries, pain, and skin infections</i>				
Abdominal pains	Leaves	Leaves chewed and juice swallowed	Zimbabwe	[14]
Acne	Leaves	Not specified	Botswana	[22]
Antidotes	Roots	Used as antidote for food poisoning	Botswana	[22]
Backache	Roots	Infusion taken orally	South Africa, Zimbabwe	[14, 20]
Bleeding from the nose (epistaxis)	Leaves	Fresh leaves inserted into nose or powdered leaves sniffed	South Africa, Zimbabwe	[14, 53]
Boils	Leaves	Infusion taken orally	South Africa	[54]
Chicken pox	Leaves	Used in washes and poultices or as steam	South Africa	[50]
Earache	Leaves	Decoction taken orally mixed with leaves of <i>E. grandis</i>	South Africa	[18]
Febrile rashes	Leaves	Infusions taken orally	South Africa	[40]
Headache, migraine	Leaves, roots	Leaf and root decoction of <i>L. javanica</i> alone taken orally or leaf decoction of <i>L. javanica</i> mixed with leaves of any of these species: <i>B. cathartica</i> , <i>C. brachiata</i> , <i>E. grandis</i> , <i>T. riparia</i> , and <i>T. emetica</i> taken orally	Kenya, South Africa, and Zimbabwe	[14, 18, 33, 34, 38]
Inflammation	Leaves	Not specified	South Africa	[41]
Pubic sores	Leaves	Decoction taken orally mixed with leaves of <i>Acanthospermum australe</i> (Loefl.) Kuntze	Swaziland	[55]
Scabies	Leaves, roots	Infusion taken orally or used in washes and poultices or as steam	South Africa, Zimbabwe	[14, 50, 54]
Shingles	Leaves, twigs, and roots	Applied as an ointment	South Africa	[15]
Skin disorders, such as heat rash scratches, stings, and bites	Leaves, twigs	Applied as an ointment or "imbiza," decoction prepared from <i>L. javanica</i> and <i>C. obliquus</i> , taken orally	South Africa	[21, 27]
Sores	Leaves	Decoction of <i>L. javanica</i> mixed with bark of <i>Acacia burkei</i> Benth., <i>Ozoroa engleri</i> R. Fern. & A. Fern., <i>Sclerocarya birrea</i> (A. Rich.) Hochst., <i>Syzygium cordatum</i> Hochst. ex Krauss, and <i>Tabernaemontana elegans</i> Stapf taken orally	South Africa	[56]
Sore eyes, cataracts	Leaves, roots	Juice squeezed into eyes	Botswana, Zimbabwe	[14, 22]
Sprained joints	Roots	Root powder applied to scarifications around sprained joints	South Africa	[20]
Ulcers	Leaves	Juice taken orally with cloves of <i>A. sativum</i>	Bangladesh	[23]
Wounds	Leaves	Fresh leaves wrapped around wound to enhance healing or infusion taken orally	Kenya, South Africa	[54, 57]
<i>Ethnoveterinary uses</i>				
Disinfecting suspected anthrax-infested meat	Whole plant	Whole plants used to disinfect suspected anthrax-infested meat	South Africa	[20]
Getting rid of ticks and other ectoparasites	Leaves, twigs, and whole plant	Crushed leaves mixed with water and sprayed, twigs used as bedding in fowl runs	South Africa, Zimbabwe	[58–60]
Lice repellent	Whole plant	Whole plant used to repel lice in poultry	India	[61]
Milk preservative	Stem	Stem applied to milk gourd before milk fermentation	Kenya	[62]

TABLE 1: Continued.

Use	Plant part(s) used	Mode of use	Country practised	Reference(s)
<i>Other uses</i>				
Anaemia in pregnancy	Leaves	Decoction taken orally	Zanzibar, Tanzania	[46]
Broom	Whole plant	Whole plants cut and tied together to make rough brooms	South Africa	[53]
Cancer	Leaves	“Imbiza,” decoction prepared from <i>L. javanica</i> and <i>C. obliquus</i> , taken orally	South Africa	[27]
Ceremonial	Leaves, whole plant	Used before and after funerals or placed on patients’ bed after circumcision	Kenya, South Africa	[31, 58, 63]
Convulsions	Leaves	Leaves rubbed on face	Zimbabwe	[14, 64]
Diabetes	Leaves	“Imbiza,” decoction prepared from <i>L. javanica</i> and <i>C. obliquus</i> , taken orally	South Africa	[27]
Fatigue or tiredness	Leaves	Leaf decoction of <i>L. javanica</i> mixed with leaves of <i>A. glabratum</i> or <i>E. grandis</i> or <i>T. riparia</i> taken orally	South Africa	[18]
Fence	Whole plant	Planted around homesteads	Ethiopia	[26]
Fodder	Leaves	Leaves eaten by goats	Kenya	[31]
Fuelwood	Whole plant	Whole plant used as fuelwood	Zanzibar, Tanzania	[46]
Human immunodeficiency virus (HIV) or acquired immunodeficiency syndrome (AIDS) symptoms	Leaves	“Imbiza,” decoction prepared from <i>L. javanica</i> and <i>C. obliquus</i> which is taken orally	South Africa	[65, 66]
Kidney problems	Root	50 g root powder boiled in 2 litres of water; patient takes a cup of this mixture once per day for 3 days	Swaziland	[67]
Madness	Leaves, whole plant	Body washed with leaf infusion	Malawi, Zimbabwe	[14, 68]
Marasmic infants	Roots	Infusions used to bath infants	South Africa, Zimbabwe	[14, 20]
Night blindness	Leaves	Face washed with infusion	Zimbabwe	[14]
Nightmares	Not specified	Not specified	Zimbabwe	[64]
Pre-, intra-, and postpartum complications	Stem, roots	Smear powder of root or stem	Kenya	[69]
Preventing odours or being used indoors to freshen air	Leaves, whole plant	Leaves sprinkled in toilets to prevent odours or whole plant placed in vase or room	Kenya, South Africa	[31, 35, 58]
Protection against dogs and crocodiles	Leaves	Leaves smeared on body	South Africa	[70]
Psychotropic behaviour	Leaves	Infusion taken orally	Swaziland	[71]
Removing bad luck	Leaves	Face and hands washed with infusion of <i>L. javanica</i> leaves mixed with <i>C. molle</i> leaves	Swaziland	[72]
Sleepless nights	Leaves	Leaf decoction of <i>L. javanica</i> mixed with leaves of <i>E. grandis</i> and <i>T. riparia</i> taken orally	South Africa	[18]
To drive away bad spirits	Leaves	Body washed with infusion	Zimbabwe	[14]
To prevent infants from contracting illness caused by father or mother committing adultery	Leaves	Leaves rubbed on forehead, elbows, and knees after committing adultery	Zimbabwe	[14]
Veneral diseases	Roots	Decoction taken orally	Mozambique	[44]
Weak joints	Leaves	Decoction taken orally	Zimbabwe	[14]

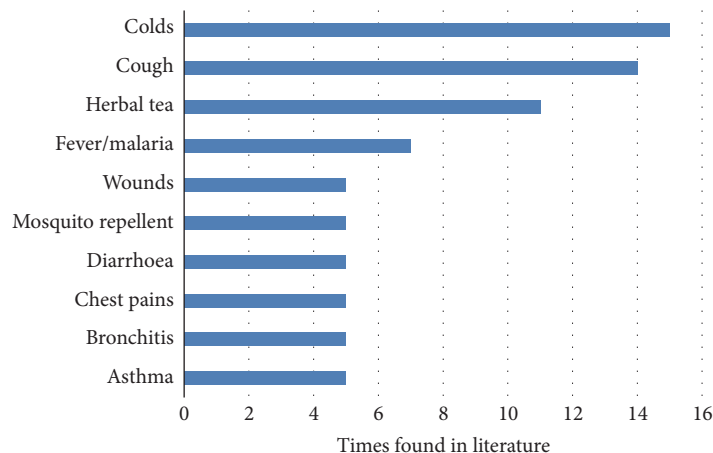


FIGURE 3: Main ethnobotanical applications of *Lippia javanica* in central, eastern, and southern Africa. An ethnobotanical use is counted only once per publication.

Food Uses; Respiratory Problems; Gastrointestinal Diseases; Fever, Malaria, and Insect Repellent; Wounds, Injuries, Pain, and Skin Infections; Ethnoveterinary Uses; and Other Uses.

2.1. Food Uses. Leaves and twigs of *Lippia javanica* are used as food additives in Kenya [6] and leafy vegetable in India [12, 13]. *Lippia javanica* is popular as herbal tea, particularly in Botswana, South Africa, and Zimbabwe [7–11]. *Lippia javanica* herbal tea is prepared by steeping fresh or dried leaves, stems, or twigs in boiling water and letting them stand for two to five minutes to release flavour, with or without milk and sugar added according to taste. A stronger brew, known as a decoction, is prepared by boiling fresh or dried leaves, stems, or twigs for more than five minutes. The herbal tea prepared from *L. javanica* leaves, stems, or twigs has a lemon or vanilla aroma and is often used as a common tea (i.e., *C. sinensis*) substitute or a few leaves, stems, or twigs are added to *C. sinensis* to provide a lemon or vanilla aroma. According to Söukand and Kalle [87], herbal tea or tisane is an English term used to denote a decoction or infusion made of herbs for medicinal purposes. When *L. javanica* is used as medicinal herbal tea, it is consumed for a limited number of days to treat a specific condition like asthma in Zimbabwe [14], bronchitis in South Africa [19], chest pains in Zimbabwe and South Africa [14, 24], colds in Kenya, South Africa, and Zimbabwe [19, 20, 28–30], cough in South Africa and Zimbabwe [19, 20, 30, 53], and other diseases (see Table 1). From literature, it seems *L. javanica* herbal tea evolved over the years from medicinal tea decoctions or infusions to nonmedicinal uses, where the herbal tea is now drunk for recreation and enjoyment. According to Shikanga et al. [7], *L. javanica* tea is appreciated throughout its distributional range as a general health tonic and also because it is naturally caffeine-free and has a calming and relaxing effect. Research done by other workers, for example, Manenzhe et al. [2], Shikanga et al. [7], Parrant [8], Olivier et al. [81], Motlhanka and Makhabu [9], and Bhebhe et al. [10, 11], revealed that *L. javanica* is a popular recreational tea in southern Africa, consumed on a daily basis within a food

context, while medicinal infusions or tisanes of *L. javanica* are taken for a specific medical purpose. During the past 20 years, the use of herbal teas has increased globally [107] because of their functional properties and consumer interest in the health promoting properties of such beverages [108]. The other advantage associated with *L. javanica* usage is that the leaves can be sun-dried and later boiled in water and drunk as herbal tea [9].

Most of *L. javanica* used as herbal tea in rural and periurban communities in central, eastern, and southern Africa is still collected from the wild, although small-scale cultivation has become necessary as it is marketed as herbal tea in Botswana under the brand name “Mosukudu” or “Mosukujane” [109] and in Zimbabwe as “Zumbani” (Figure 1(b)). Although considerable quantities of *L. javanica* are sold in local markets in Botswana and Zimbabwe and also traded on the Internet, there is no data on present production levels, traded volumes, values, and export figures in these two countries. Research by Whiteside [110] showed that sales of *L. javanica* tea bags generated an income of R20300.00 (US\$5718) during 1994/1995 in Botswana. There is increased demand for *L. javanica* herbal tea especially in the light of growing health consciousness worldwide, with the estimated potential demand for the species and its products around 100 tonnes per year on the local market and 1000 tonnes per year on the export market (<http://bio-innovation.org/work/fever-tea-tree/>). Considering the rapid growth of the herbal tea industry worldwide and the increasing demand for *L. javanica* products, there is need for the improvement of *L. javanica* products as well as development of new products. Therefore, *L. javanica* has potential to make the transition from limited local use to commercial and international product. *Lippia javanica* has been identified as one of the few plant species that should be integrated in the domestication process in farming systems in sub-Saharan Africa to support medicinal, nutritional, and income security of local communities [107, 111]. According to Van Wyk [111], *L. javanica* is of commercial value as herbal tea and health drinks. Therefore, commercialization of *L. javanica* is

unlikely to be viable if the product is solely harvested from the wild. In the same line commercialization of *L. javanica* might be more worthy if other products other than tea are derived from the plant.

2.2. Respiratory Problems. The different parts of *L. javanica*, for example, the leaves and twigs, are used for the treatment of asthma, coughs, colds, influenza, pneumonia, tuberculosis, and bronchial problems in Bangladesh, Botswana, Ethiopia, Kenya, South Africa, and Zimbabwe [14–16, 19, 20, 23, 25, 26, 28–32, 53, 112]. In Bangladesh, leaf infusion of *L. javanica* is taken orally mixed with 3–4 pieces of cloves of *A. sativum*, 2–3 times daily as remedy for chest pains [23]. In Ethiopia, leaves of *L. javanica* are chewed with butter as remedy for chest pains and cough [26]. In India, *L. javanica* leaf decoction is taken orally as remedy for respiratory disorders [39]. In Kenya, leaves are sniffed [31] or half glass of hot leaf infusion is taken orally three times a day [28, 29] as remedy for colds and cough. Research done by Davids et al. [19] in South Africa revealed that about 50 g of leaves is added to a cup of boiling water to produce an infusion which is taken orally as remedy for coughs, colds, and bronchial problems or the infusion is applied to the skin or mixed with Vaseline to make an ointment. In South Africa, leaf or twig decoction is taken orally for asthma, colds, and cough [16, 53], leaves are used in washes and poultices for chest pains [25], and leaves are boiled for 5 minutes and one cup of extract is taken orally thrice a day for chest pains and tuberculosis [24, 42]. According to York [18], two handfuls of *L. javanica* leaves are boiled in two litres of water and patient is steamed once or twice a day to treat coughs, chest pain, headaches, fever, chills, a sore throat, or a blocked nose. This decoction can also be taken orally by drinking half a cup daily [18]. Alternatively, a handful of *L. javanica* roots or leaves are mixed with a handful leaves of *A. glabratum* or *B. transvaalensis* or *B. uniflora* or *B. cathartica* or *C. anisata* or *C. brachiata* or *C. molle* or *E. grandis* or *H. kraussii* or *Hypoxis* spp. or *K. mosambicina* or *P. neochilus* or *P. guajava* or *S. serratuloides* or *T. sericea* or *T. riparia* or *T. emetica* to treat blocked nose, chest pain, cough, earache, fatigue, fever, influenza, headache, runny nose, sleepless nights, sore throat, tiredness, and tonsillitis [18]. Leaf and stem infusion of *L. javanica* are taken orally together with *A. afra* by the Zulu people in South Africa as remedy for bronchial ailments, colds, and cough [20]. “*Imbiza*,” a popular herbal decoction prepared from *L. javanica* and *C. obliquus*, is used by the Zulu people in South Africa as herbal tonic and an immune booster and also for the treatment of cancer, chest pains, colds, diabetes, HIV or AIDS symptoms, skin infections, and tuberculosis [27]. In some communities in South Africa, *L. javanica* leaf or root infusion is taken orally as remedy for influenza and malaria [33] and respiratory disorders [40, 41]. In Zimbabwe, leaf or root decoction is taken orally or smoke of burnt leaves and roots is inhaled as remedy for chest pains [14] and leaf and twigs are boiled in water and infusion is taken orally as remedy for colds and cough [30]. According to Gelfand et al. [14], Shona people in Zimbabwe rub leaf ointment of *L. javanica* on the chest and abdomen as remedy for pneumonia and leaf decoction is taken orally and body washed with leaf

decoction as remedy for shortness of breath or dyspnoea. In South Africa, a leaf and stem infusion of *L. javanica* are taken orally together with leaves of *A. afra* as remedy for measles [20].

Based on literature, inhalation of smoke from *L. javanica* appears to be a popular remedy for asthma, chest pains, colds, and chronic coughs in Botswana, South Africa, and Zimbabwe [14, 15, 20, 32]. In Botswana, *L. javanica* leaf infusion vapour is inhaled as remedy for colds and cough [32]. In South Africa, for example, leaves and twigs are burned and smoke is inhaled as remedy for asthma and cough [15] or steam from leaf infusions is inhaled or hot leaf infusions are taken orally against colds and cough [20]. In Zimbabwe, smoke of burnt leaves and roots is inhaled as remedy for chest pains [14].

2.3. Gastrointestinal Diseases. The leaf and root decoction or infusions of *L. javanica* are used as remedies of the digestive system diseases such as cholera, diarrhoea, and dysentery. For example, in Mozambique, root decoction is taken orally as remedy for a type of diarrhoea commonly known as “*chinhamucaca*” which is characterized by milky diarrhoea accompanied by vomiting in children [44]. Leaf decoction is used as herbal medicine for diarrhoea in Kenya [113]. In South Africa, the leaves of *L. javanica* are crushed and mixed with cold or hot water and the mixture is then sieved and a quarter of a cup (75 mL) is taken twice a day until diarrhoea subsides [34]. The Venda people in South Africa use leaf infusions as prophylactic against diarrhoea [40]. Research done by Palgrave et al. [37] in South Africa revealed that tea infusions of the *L. javanica* leaves are used as remedies for HIV/AIDS opportunistic infections such as lung infections and diarrhoea. Previous research in Mozambique [114], South Africa [115], and Zimbabwe [116] showed that gastrointestinal disorders, particularly cholera, diarrhoea, and dysentery are among human diseases often treated with herbal medicines. For those patients diagnosed as having intestinal worms in Venda, South Africa, a leaf infusion of *L. javanica* leaves is used as an anthelmintic [40]. Therefore, these findings illustrate that herbal medicines including *L. javanica* can play an important role in basic health care of local communities through treatment and management of cholera, diarrhoea, and dysentery.

2.4. Fever, Malaria, and Insect Repellent. Traditionally, *L. javanica* is commonly used to treat fever and malaria and repel insects throughout its distributional range [4, 14, 16, 17, 33–35, 40, 48–50, 58–60]. In South Africa, a decoction of fresh or air dried leaves is used to wash or steam body parts infested with lice [50]. In India, the whole plant is used to repel lice in poultry [61]. In South Africa and Zimbabwe, whole plant or leaves are burnt to repel mosquitoes [4, 34, 35, 48, 49, 58]. In South Africa and Zimbabwe, *L. javanica* is widely used to get rid of ticks and other ectoparasites; for example, ticks are sprayed with crushed leaves mixed with water or twigs are used as bedding in fowl runs [58–60]. Leaf and twig decoction of *L. javanica* are taken orally as remedy for fever [16]. The Venda people in South Africa use leaf infusions of *L. javanica* as prophylactic against malaria

[40]. In Zimbabwe, leaf decoction is taken orally as remedy for fever [14].

Lippia javanica is also cultivated on a commercial scale by a rural community in Giyani, the Limpopo province, South Africa, for the production of mosquito-repellent candles [117]. Clinical studies using human volunteers showed that *L. javanica* repels no less than 95% of mosquitos, whereas most mosquito repellents repel only 42% of them [118]. Based on these findings, the Council for Scientific and Industrial Research (CSIR), South Africa, signed a benefit sharing agreement with traditional healers allowing for the commercial cultivation of *L. javanica* aimed at establishing an indigenous oil industry for rural development and large-scale production of antimosquito candles and other insect repellents. The mosquito repellent is registered as a patent under the Fertilisers, Farm Feeds and Stock Remedies Act (Act 36 of 1947) as a pest repellent [119]. These findings and the establishment of a large-scale production of antimosquito candles and other insect repellents strengthen the view that *L. javanica* is a potential source of antipesticidal agents and to some extent validate the traditional use of the plant species for insect pest control. Pesticidal plants such as *L. javanica* are increasingly being used as alternatives where synthetic products are unaffordable or are not available or are ineffective. A number of studies have indicated that the use of *L. javanica* as a pesticide is a long-standing tradition passed down from generation to generation [120].

2.5. Wounds, Injuries, Pain, and Skin Infections. *Lippia javanica* is used as remedy for a variety of skin infections and injuries in Kenya, South Africa, Swaziland, and Zimbabwe. In South Africa, leaf infusions are used to treat skin disorders, such as boils, chicken pox, febrile rashes, heat rashes, measles, scabies, scratches, and stings [20, 21, 40, 50, 54]. In Kenya, about 50 g of fresh leaves is wrapped around a fresh wound to enhance healing [57] and, in South Africa, leaf infusions are taken orally as remedy for wounds [54]. In South Africa, powder from burnt roots of *L. javanica* is applied to scarifications made around sprained joints to facilitate healing [20]. In Kenya and Zimbabwe, leaf infusion is taken orally to treat measles or a patient is washed with leaf infusion [14, 38]. The Zulu people in South Africa take a mixture of chopped handful leaves of *L. javanica* with the same amount of bark of *A. burkei*, *O. engleri*, *S. birrea*, *S. cordatum*, and *T. elegans* boiled in two litres of water as an enema for the treatment of sores [56]. In Swaziland, about 30 g of *L. javanica* leaves and similar amount of *Acanthospermum australe* Kuntze are boiled in 5 litres of water and decoction is taken as remedy for sores [55]. In Zimbabwe, root ashes of *L. javanica* mixed with fat are applied to the skin of a patient with scabies [14]. Wound healing is a process which involves distinct overlapping phases of coagulation, inflammation, proliferation, and tissue remodelling [121]. The same authors argued that a set of complex biochemical events takes place in a closely orchestrated cascade to repair the wound and any errors in the wound healing process can lead to delayed healing or formation of hypertrophic scars. Abubakar [122] argued that colonization of wounds by opportunistic microorganisms usually delays the wound

healing process and/or may lead to infectious condition. Therefore, application of *L. javanica* on wounds and skin infections leads to disinfection, debridement, and provision of a suitable environment for aiding the wound healing process. *Lippia javanica* has potential for therapeutic use in wound and skin diseases management, but there is need for research on the safety, phytochemistry, and biological properties of the species.

2.6. Ethnoveterinary Uses. The leaves, stems, twigs, and whole plants of *L. javanica* are used as ethnoveterinary medicine in India, Kenya, South Africa, and Zimbabwe. The Xhosa people in the Eastern Cape province, South Africa, use *L. javanica* leaves for the disinfection of meat that has been infected with anthrax [20]. In Kenya, the stem of *L. javanica* is used to preserve milk by applying it to the gourd before milk fermentation [62]. In India, whole plants are used as lice repellants while, in South Africa and Zimbabwe, crushed leaves mixed with water are used to get rid of ticks and twigs are used as bedding in fowl runs to get rid of ectoparasites [58–60].

2.7. Other Uses. *Lippia javanica* is used as a good luck charm, to treat persons experiencing bad dreams, to ward off evil spirits, to protect one from lightening, and to protect the home (Table 1). In Zimbabwe, *L. javanica* leaves are prepared as an infusion to treat persons experiencing nightmares [64]. In both South Africa and Zimbabwe, evil spirits are cleansed by washing the body of an affected person by leaf infusion of *L. javanica* [14, 123]. In Swaziland, 50 g leaves of *L. javanica* and *C. molle* are ground into a powder and 5 litres of water is added and face and hands are washed to remove bad luck when exposed to a corpse [71]. In South Africa, the whole plant is placed on a patient's bed after the circumcision ceremony [58], to prevent odours and freshen surrounding air. A mixture of *L. javanica* leaves and roots is used to clean tools and hands before and after funerals, the stems and leaves are used as brooms to sweep grave sites, and the entire plant is also used when coming from the mortuary to remove bad spirits [58, 124]. The corpse is washed with *L. javanica* infusion after death to prevent odours forming or if the corpse has an odour, women place pieces of *L. javanica* in the nostrils of the corpse and sweep the room with twigs where the person was sleeping [58]. In KwaZulu Natal, South Africa, if the meat started to smell it will be boiled with *L. javanica* leaves to take away the smell [58]. The Xhosa people in the Eastern Cape province, South Africa, use *L. javanica* leaves for the disinfection of meat that has been infected with anthrax [20]. In KwaZulu Natal, South Africa, leaves are sprinkled in toilets to prevent odours [58]. Leaves of *L. javanica* are sprinkled in houses for pleasant smell in the Limpopo province, South Africa [35], and Kenya [31]. In KwaZulu Natal, South Africa, it is believed that a person can repel snakes by placing a small stem with leaves on his or her head [58]. In Malawi, Swaziland, and Zimbabwe, people showing sign of mental disorder, madness, or hysterical outbursts are required to wash their bodies with leaf infusions [14, 67, 68, 71].

TABLE 2: Nutritional composition of *Lippia javanica* leaves.

Caloric and nutritional composition	Values	Reference
Ash	1.60 ± 0.001 g	[13]
Calorific value	50.64 ± 5.63 kcal	[13]
Crude fat	0.16 ± 0.001 g	[13]
Crude fibre	2.63 ± 0.002 g	[13]
Crude protein	1.22 ± 0.0004 g	[13]
Dry matter	15.46 ± 1.40 g	[13]
Moisture	90.44 ± 0.26 g	[13]
Total carbohydrate	6.55 ± 0.26 g	[13]
Vitamin C	22.42 ± 0.001 mg	[13]
Ca	2856–9225 µg g ⁻¹	[27]
Mg	1598–5619 µg g ⁻¹	[27]
Fe	75–750 µg g ⁻¹	[27]
Mn	40.1 ± 1.16 µg g ⁻¹	[27]
Zn	15.6–27.3 µg g ⁻¹	[27]
Cu	3.0–8.5 µg g ⁻¹	[27]
Cr	0–2.7 µg g ⁻¹	[27]
Se	2.57 ± 0.19 µg g ⁻¹	[27]
Pb	0.38–1.19 µg g ⁻¹	[27]
Cd	0.53 ± 0.05 µg g ⁻¹	[27]
Co	0.19 ± 0.01 µg g ⁻¹	[27]

3. Phytochemical Constituents and Nutritional Composition of *Lippia javanica*

Multiple classes of phytochemicals including volatile and nonvolatile secondary metabolites, such as alkaloids, amino acids, flavonoids, iridoids, and triterpenes as well as several minerals, have been identified from *L. javanica* [1–5, 7, 13, 27, 73, 78, 81, 83–86, 88–94, 120, 123, 125–127]. Leaves, flowers, and twigs of *L. javanica* have a wide variety of the so-called classic nutrients, such as minerals, carbohydrates, proteins, fats, and vitamins (Table 2). *Lippia javanica* leaves are a good source of minerals such as cadmium, calcium, chromium, cobalt, copper, iron, magnesium, manganese, selenium, and zinc [27]. These authors assessed the levels of the elements in *L. javanica* leaves and found the elements to be in the decreasing order of Ca > Mg > Fe > Zn > Mn > Cu > Se > Cr > Pb > Co > Cd for total concentrations and Ca > Mg > Fe > Zn > Cu > Cr > Pb for water extractable forms. These results corroborate an observation made by Sedaghatthoor et al. [128] that Ca and Mg are among the most abundant elements in tea plants. These mineral elements are important in human nutrition since *L. javanica* is used as herbal tea and food additive (Table 1). Calcium, magnesium, iron, manganese, and zinc play a major role in activating some enzymes and regulating many responses of cells to stimuli [129]. Some of the mineral elements identified from *L. javanica* leaves are required by the human body for repair of worn out cell tissues and strong bones and teeth and building of red blood cells and other related tissues. Therefore, since *L. javanica* has appreciable concentrations of mineral elements such as calcium, magnesium, iron, manganese, and zinc

(Table 2) which are essential for enzyme metabolism, these mineral elements could enhance the nutritional and curative properties of the species.

Bhebhe et al. [10] determined the total phenolic and tannin content and radical scavenging activities of *L. javanica*, comparing it with *Aspalathus linearis* (Burm.f.) R. Dahlgren (Rooibos™), a commercial South African herbal tea, and other popular herbal teas in Zimbabwe (Table 3). *Adansonia digitata* L., *Fadogia ancylantha* Schweinf., *Ficus sycomorus* L., and *Myrothamnus flabellifolius* Welw. are indigenous herbal teas consumed in Zimbabwe. The tannin content of *L. javanica* is very low when compared to *A. linearis* (Rooibos) (Table 3). *Lippia javanica* has higher radical scavenging activity than *A. linearis* (Table 3) which is probably due to higher total phenolic content in comparison to *A. linearis* which is a popular herbal tea consumed by 10% of the global herbal tea market [10]. In another study, Bhebhe et al. [11] determined and compared the effect of several solvents, namely, hot water, 50% methanol, ethanol, 50% ethanol, acetone, 50% acetone, and ethyl acetate, on phenolic composition and free radical scavenging activity in common black tea, *C. sinensis*, and five other well-known herbal teas including *L. javanica*. In all the seven solvents used, *L. javanica* had higher total phenolic content than *C. sinensis* implying that *L. javanica* is competitive to the black tea in terms of phenolic content. Shikanga et al. [7] found leaf extracts of *L. javanica* to have higher phenolic content of 14.8 mg/g gallic acid equivalent of dry weight than flowers (9.9 mg/g) and twigs (8.3 mg/g). Phenolic compounds found in plants are known to play an important role as antioxidants in exhibiting the medicinal properties such as antibiotic, anti-inflammatory, anticancer, and antiallergic properties [10, 66, 130, 131].

The compounds isolated from *L. javanica* are documented and listed in Appendix A and their structures are presented in Appendix B. Simple phenolic compounds and caffeic acid and its derivatives are some of the compounds that have been identified in *L. javanica* and examples include coumarin **1**, 3,4-dihydroxy- β -phenylethoxy-O-[4''- β -caffeoyl- α -rhamnopyranosyl-(1''',3'')-O- β -glucopyranoside], commonly referred to as verbascoside **2**, and 3,4-dihydroxy- β -phenylethoxy-O-[6''- β -caffeoyl- α -rhamnopyranosyl-(1''',3'')-O- β -glucopyranoside] commonly referred to as isoverbascoside **3** isolated by Olivier et al. [81] from the aerial parts of the species. Nonvolatile diterpenes, known as iridoid-glycosides, have also been isolated from *L. javanica* by Rimpler and Sauerbier [82] represented by theveside-Na **4** and theveridoside **5**. Mujovo et al. [83] isolated a long chain alkane “4-ethylnonacosane” **6** and four flavanones apigenin **7**, cirsimaritin **8**, 6-methoxyluteolin 4'-methyl ether **9**, and 6-methoxyluteolin 3',4',7-trimethyl ether **10** from ethanolic extracts of *L. javanica* leaves. Madzimore et al. [120] identified an array of phenolic glycosides and flavonoids which include crassifolioside **11**, luteolin **12**, diosmetin **13**, chrysoeriol **14**, tricetin **15**, isothymusin **16**, eupatorin **17**, 5-dimethyl noboletin **18**, genkwanin **19**, salvigenin **20**, and an alkaloid xanthine **22**. Ludere et al. [84] isolated lippialactone **21** from the ethyl acetate extract of aerial parts of *L. javanica*. Neidlein and Staehle [85] and Dlamini [86] isolated 19 amino acids, compounds **23** to

TABLE 3: Total phenolic content (TPC) as gallic acid equivalents (GAE) and tannin content as leucocyanidin equivalents (LE) and free radical scavenging activity (FRSA) of herbal tea extracts (after Bhebhe et al. [10]).

Plant species	TPC g GAE/100 g	Tannin content of LE/100 g	Percentage FRSA	IC ₅₀ g/mL
<i>Lippia javanica</i>	12.46 ± 4.31	0.12 ± 0.01	83.77 ± 0.8	0.016
<i>Aspalathus linearis</i>	6.69 ± 0.83	0.94 ± 0.04	67.27 ± 0.25	0.053
<i>Adansonia digitata</i>	0.79 ± 0.28	1.69 ± 0.01	43.07 ± 1.0	0.132
<i>Fadogia ancylantha</i>	2.88 ± 0.48	0.20 ± 0.12	70.0 ± 0.40	0.051
<i>Ficus sycamores</i>	14.02 ± 0.01	1.98 ± 0.12	86.13 ± 0.85	0.009
<i>Myrothamnus flabellifolius</i>	4.75 ± 0.33	1.04 ± 0.01	80.93 ± 0.75	0.019

41 in Appendix A, from *L. javanica*. At least 131 different classes of essential oil compounds (compounds 42–172 in Appendices A and B) have been isolated from *L. javanica* by several researchers [3–5, 83, 85, 86, 88–94, 123]. Hutchings and van Staden [96] isolated a toxic triterpenoid saponin, icterogenin 173, from *L. javanica* leaves. These different classes of essential oil compounds have been associated with various therapeutic activities such as anaesthetic, analgesic, anti-inflammatory, antimicrobial, cardiovascular, decongestant, digestive, expectorant, hepatoprotective, and sedative activities as well as stimulant of nervous system and tonifying effects [132]. Meanwhile flavonoids possess several pharmacological properties including antibacterial, anticancer, anti-inflammatory, antioxidant, antiviral, and hepatoprotective effects [133] which play important roles in human health. Flavonoid such as apigenin 7 is reported to possess antibacterial [134] and hepatoprotective [135] properties. Apigenin 7 and luteolin 12 are reported to possess anti-inflammatory and analgesic effects [133], affecting the function of enzyme systems involved in the generation of inflammatory processes, especially tyrosine and serine-threonine protein kinases [136, 137]. It has also been reported that apigenin 7 prevents HIV-1 activation via a novel mechanism that involves inhibition of viral transcription [138] and luteolin 12 demonstrated synergistic effects with another flavonoid kaempferol against herpes simplex virus [133]. Kamiya [139] documented the importance of essential amino acids such as lysine, valine, isoleucine, and histidine in terms of the risks to health if they are deficient. The author also documented their biological effects which include muscle protein maintenance, potentiation of immune function, tissue repair acceleration after burn or trauma, protecting liver from toxic agents, lowering blood pressure, modulating cholesterol metabolism, and stimulating insulin or growth hormone secretion. Therefore, *L. javanica* leaves and other plant parts which have shown to be rich in flavonoid and polyphenolic compounds, amino acids, and essential oil could play an important role in the treatment and management of diseases such as hypertension and inflammation listed in Table 1.

4. Pharmacological Activities

Scientific studies on *L. javanica* indicate that it has a wide range of pharmacological activities (Table 4), which include

anticancer [140], antidiabetic [141], antimalarial [4, 49, 142], antimicrobial [2, 3, 7, 17, 73–75, 77, 83, 84, 126, 143], antioxidant [7, 10, 11, 75, 77, 125], antiplasmodial [2, 79, 80, 84, 144, 145], and pesticidal effects [1, 2, 59, 95, 120, 146–150] and cytotoxicity [2, 73, 79, 120] activities. Table 4 summarizes some of the pharmacological studies undertaken on *L. javanica* extracts aimed at evaluating some of the ethnomedicinal uses of the species documented throughout its distributional range (see Table 1). Some of the listed pharmacological activities may not relate directly to the documented ethnomedicinal uses of the species but may provide some insight into the species' potential therapeutic value and bioactive properties and application.

4.1. Anticancer Activity. Fouche et al. [140] reported anticancer activity of dichloromethane root extract of *L. javanica* against three human cells, exhibiting TGI value of 1.82 $\mu\text{g}/\text{mL}$ for breast MDA-MB-435, 1.86 $\mu\text{g}/\text{mL}$ for breast MDA-N, and 2.09 $\mu\text{g}/\text{mL}$ for melanoma MALME-3M. Based on literature, a couple of terpenoid compounds that have been isolated from *L. javanica* are known to have antitumor properties. For example, linalool 120 is known to have antitumor activity which plays a protective role against hepatotoxicity and the compound has anti-inflammatory activities as well [151]. Research by Yang et al. [152] showed limonene 92 to have inhibitory effect on pancreatic and mammary tumors. Another terpenoid compound, α -pinene 44, is known to inhibit translocation of NF- κ B or p65 protein into nuclei of LPS-stimulated THP-1 cells [153]. These findings serve as a scientific validation for the use of *L. javanica* as a component of a herbal concoction known in KwaZulu Natal province as “imbiza,” prepared by mixing *L. javanica* with *C. obliquus* as herbal medicine for cancer [27].

4.2. Antidiabetic Activity. Arika et al. [141] determined the in vivo antidiabetic activity of aqueous leaf extracts of *L. javanica* in white male alloxan-induced albino mice. The aqueous leaf extracts of *L. javanica* at all dose levels significantly lowered the blood glucose levels in both oral and intraperitoneal routes. The antidiabetic effect of *L. javanica* could have been due to the observed presence of flavonoids. The polyhydroxylated flavonol enhances lipogenesis and glucose uptake in the adipocytes and flavanoids have demonstrated insulinmimetic properties as the compound is known to be effective at controlling blood sugar levels. These findings

TABLE 4: Summary of pharmacological activities of the extracts isolated from different parts of *Lippia javanica*.

Activity tested	Extract	Plant part	Model	Effect	Reference
Antiamoebic	Piperitenone 162	Leaves	Microdilution assay	Active with IC ₅₀ value of 25 µg/mL against <i>E. histolytica</i>	[73]
Antibacterial	Piperitenone 162	Leaves	Microdilution assay	Active with MIC value of 50 µg/mL against <i>A. calcoaceticus</i> , <i>M. kristinae</i> (50 µg/mL), <i>S. typhi</i> (25 µg/mL), and <i>S. aureus</i> (12 µg/mL)	[73]
Antibacterial (Antiproteus)	Methanol	Leaves	Disc diffusion assay	Active with MIC value of 313 µg/mL against <i>P. mirabilis</i> and 926 µg/mL against <i>P. vulgaris</i>	[74]
	Water	Leaves	Disc diffusion assay	Exhibited weak activity against <i>P. mirabilis</i> with MIC value of 1873 µg/mL and 1768 µg/mL against <i>P. vulgaris</i>	[74]
Antibacterial	Acetone	Leaves	Microdilution assay	Active against <i>E. coli</i> with MIC value of 0.64 mg/mL, <i>E. faecalis</i> (0.64), <i>P. aeruginosa</i> (0.32), and <i>S. aureus</i> (0.64). Total activity MIC values were as follows: <i>E. coli</i> (127 mg/mL), <i>E. faecalis</i> (127 mg/mL), <i>P. aeruginosa</i> (253 mg/mL), and <i>S. aureus</i> (127 mg/mL)	[75]
	Acetone	Leaves	Microdilution assay	Active against <i>B. cereus</i> with MIC value of 3 mg/mL, <i>Bacillus pumilus</i> (6 mg/mL), <i>B. subtilis</i> (>12 mg/mL), <i>S. aureus</i> (12 mg/mL), <i>E. faecalis</i> (6 mg/mL), <i>E. cloacae</i> (6 mg/mL), <i>E. coli</i> (6 mg/mL), <i>Pantoea agglomerans</i> (12 mg/mL), <i>P. aeruginosa</i> (12 mg/mL), <i>Shigella flexneri</i> (1.5 mg/mL), <i>Aeromonas hydrophila</i> (12 mg/mL), <i>P. mirabilis</i> (>12 mg/mL), <i>K. pneumoniae</i> (12 mg/mL), <i>Salmonella choleraesuis</i> (6 mg/mL), and <i>Serratia marcescens</i> (>12 mg/mL)	[17]
Antibacterial	Hexane	Leaves	Microdilution assay	Active against <i>B. cereus</i> with MIC value of 12 mg/mL, <i>B. pumilus</i> (12 mg/mL), <i>B. subtilis</i> (12 mg/mL), <i>S. aureus</i> (6 mg/mL), <i>E. faecalis</i> (12 mg/mL), <i>E. cloacae</i> (12 mg/mL), <i>E. coli</i> (6 mg/mL), <i>P. agglomerans</i> (12 mg/mL), <i>P. aeruginosa</i> (12 mg/mL), <i>S. flexneri</i> (6 mg/mL), <i>A. hydrophila</i> (12 mg/mL), <i>P. mirabilis</i> (12 mg/mL), <i>K. pneumoniae</i> (12 mg/mL), <i>S. choleraesuis</i> (6 mg/mL), and <i>antiamoebic effect</i> (12 mg/mL)	[17]
	Methanol	Leaves	Microdilution assay	Active against <i>B. cereus</i> with MIC values of 1.5 mg/mL, <i>B. pumilus</i> (6 mg/mL), <i>B. subtilis</i> (>12 mg/mL), <i>S. aureus</i> (12 mg/mL), <i>E. faecalis</i> (3 mg/mL), <i>E. cloacae</i> (6 mg/mL), <i>E. coli</i> (6 mg/mL), <i>P. agglomerans</i> (12 mg/mL), <i>P. aeruginosa</i> (12 mg/mL), <i>S. flexneri</i> (1.5 mg/mL), <i>A. hydrophila</i> (6 mg/mL), <i>P. mirabilis</i> (>12 mg/mL), <i>K. pneumoniae</i> (12 mg/mL), <i>S. choleraesuis</i> (6 mg/mL), and <i>S. marcescens</i> (12 mg/mL)	[17]
Essential oil	Essential oil	Leaves	Microdilution assay	Active against <i>B. cereus</i> with MIC values of 6 mg/mL, <i>B. pumilus</i> (6 mg/mL), <i>B. subtilis</i> (3 mg/mL), <i>S. aureus</i> (1.5 mg/mL), <i>E. faecalis</i> (3 mg/mL), <i>E. cloacae</i> (12 mg/mL), <i>E. coli</i> (6 mg/mL), <i>P. agglomerans</i> (6 mg/mL), <i>P. aeruginosa</i> (12 mg/mL), <i>S. flexneri</i> (3 mg/mL), <i>A. hydrophila</i> (3 mg/mL), <i>P. mirabilis</i> (>12 mg/mL), <i>K. pneumoniae</i> (3 mg/mL), <i>S. choleraesuis</i> (1.5 mg/mL), and <i>S. marcescens</i> (>12 mg/mL)	[17]
		Leaves	Microdilution assay		

TABLE 4: Continued.

Activity tested	Extract	Plant part	Model	Effect	Reference
	Methanol	Leaves	Microdilution assay	Active against <i>E. faecalis</i> with MIC value of 0.14 mg/mL	[7]
	Methanol	Leaves	Microdilution assay	Active against <i>E. coli</i> with MIC value of 0.31 mg/mL	[7]
	Methanol	Leaves	Microdilution assay	Active against <i>P. aeruginosa</i> with MIC value of 0.42 mg/mL	[7]
	Methanol	Leaves	Microdilution assay	Active against <i>S. aureus</i> with MIC value of 0.13 mg/mL	[7]
	Acetone	Leaves	Microdilution assay	Active against <i>C. albicans</i> with MIC value of >7.5 mg/mL, <i>C. krusei</i> (1.88 mg/mL), and <i>C. neoformans</i> (>7.5 mg/mL)	[7]
	Hexane	Leaves	Microdilution assay	Active against <i>C. albicans</i> with MIC value of >3.75 mg/mL, <i>C. krusei</i> (3.75 mg/mL), and <i>C. neoformans</i> (>7.5 mg/mL)	[7]
	Acetone	Leaves	Microdilution assay	Active against <i>C. krusei</i> with MFC value of 7.5 mg/mL	[7]
Antifungal	Dichloromethane	Aerial parts	Microdilution assay	Active against <i>Fusarium proliferatum</i> with MIC value of 0.14 mg/mL and <i>Fusarium verticillioides</i> (0.19 mg/mL)	[76]
	Hexane	Aerial parts	Microdilution assay	Active against <i>F. proliferatum</i> with MIC value of 0.23 mg/mL and <i>F. verticillioides</i> (0.45 mg/mL)	[76]
	Methanol	Aerial parts	Microdilution assay	Active against <i>F. proliferatum</i> with MIC value of 1.77 mg/mL and <i>F. verticillioides</i> (0.43 mg/mL)	[76]
	Methanol	Aerial parts	Microdilution assay	Active against <i>F. proliferatum</i> with MIC value of >2.50 mg/mL and <i>F. verticillioides</i> (>2.50 mg/mL)	[76]
	Acetone	Leaves	Microdilution assay	Active with MIC value of 0.47 mg/mL and total activity of 10 mL/g against <i>M. smegmatis</i>	[77]
	Dichloromethane	Leaves	Microdilution assay	Active with MIC value of 1.25 mg/mL and total activity of 23 mL/g against <i>M. smegmatis</i>	[77]
Antimycobacterial	Hexane	Leaves	Microdilution assay	Active with MIC value of 0.62 mg/mL and total activity of 13 mL/g against <i>M. smegmatis</i>	[77]
	Methanol	Leaves	Microdilution assay	Active with MIC value of 1.25 mg/mL and total activity of 7 mL/g against <i>M. smegmatis</i>	[77]

TABLE 4: Continued.

Activity tested	Extract	Plant part	Model	Effect	Reference
Antioxidant	Water	Leaves	DPPH assay	Exhibited activity with EC_{50} value of 358 μ g/mL	[7]
	Water	Leaves	—	Exhibited activity with 209 ascorbic acid equivalent (mg/g dry weight)	[7]
	Water	Leaves	ABTS assay	Active with TEAC value of 1.5 mmol/100 g	[78]
	Water	Leaves	DPPH assay	Active with TEAC value of 1462.54 mmol/100 g	[78]
	Water	Leaves	FRAP assay	Active with TEAC value of 2.38 mmol/100 g	[78]
	Antiplasmodial	Hexane, chloroform	Roots	Microdilution assay	Active against <i>P. falciparum</i> chloroquine sensitive with IC_{50} value of 12.25 \pm 0.72 μ g/mL
Roots			Microdilution assay	Active against <i>P. falciparum</i> chloroquine sensitive with IC_{50} value of 12.12 \pm 0.79 μ g/mL	[79]
Roots			Microdilution assay	Active against <i>P. falciparum</i> chloroquine sensitive with IC_{50} value of 1.35 \pm 0.06 μ g/mL	[79]
Hexane, chloroform		Roots	Microdilution assay	Active against <i>P. falciparum</i> chloroquine resistant with IC_{50} value of 18.59 \pm 0.26 μ g/mL	[79]
		Roots	Microdilution assay	Active against <i>P. falciparum</i> chloroquine resistant with IC_{50} value of 15.80 \pm 0.26 μ g/mL	[79]
Methanol		Roots	Microdilution assay	Active against <i>P. falciparum</i> chloroquine resistant with IC_{50} value of 1.75 \pm 0.17 μ g/mL	[79]
		Roots	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of 3.8 μ g/mL	[80]
Dichloromethane/methanol		Roots	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of 27 μ g/mL	[80]
		Roots	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of 24 μ g/mL	[80]
Water		Roots	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of >100 μ g/mL	[80]
		Stems	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of 4.5 μ g/mL	[80]
Dichloromethane/methanol		Stems	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of 21.8 μ g/mL	[80]
		Stems	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of 29.8 μ g/mL	[80]
Methanol		Stems	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of >100 μ g/mL	[80]
		Water	pLDH assay	Active against <i>P. falciparum</i> with IC_{50} value of >100 μ g/mL	[80]
Free radical scavenging activity	Water	—	DPPH assay	Active with IC_{50} value of 0.059 \pm 0.02 g/mL	[11]
		—	DPPH assay	Active with IC_{50} value of 0.04 \pm 0.001 g/mL	[11]
	Ethanol	—	DPPH assay	Active with IC_{50} value of 0.025 \pm 0.001 g/mL	[11]
		—	DPPH assay	Active with IC_{50} value of 0.027 \pm 0.005 g/mL	[11]
	50% ethanol	—	DPPH assay	Active with IC_{50} value of 0.057 \pm 0.004 g/mL	[11]
		—	DPPH assay	Active with IC_{50} value of 0.022 \pm 0.001 g/mL	[11]
	50% acetone	—	DPPH assay	Active with IC_{50} value of 0.066 \pm 0.001 g/mL	[11]
		—	DPPH assay	Active with IC_{50} value of 0.066 \pm 0.001 g/mL	[11]
	Ethyl acetate	—	Brine shrimp lethality assay	40% mortality recorded after 48 h exposure towards <i>Artemia nauplii</i>	[74]
		Methanol	Leaf	Active against <i>P. falciparum</i> chloroquine sensitive with IC_{50} value of 843.0 μ g/mL	[79]
Methanol		Roots	Active against <i>P. falciparum</i> chloroquine resistant with IC_{50} value of 650.3 μ g/mL	[79]	

strengthen the view that *L. javanica* is a potential source of antidiabetic agents and to some extent validate the traditional use of the plant species mixed with *C. obliquus* to form a herbal concoction known in KwaZulu Natal province, South Africa, as “imbiza” used against diabetes [27].

4.3. Antimalarial Activity. Govere et al. [142] found that topical application of *L. javanica* alcohol extract leads to 76.7% protection against *Anopheles arabiensis* for 4 hours. *Lippia javanica* has been used as a mosquito repellent by the rural communities in Zimbabwe for a long time and previous studies have shown that essential oils from the species have very strong and lasting repellent activity against female *A. arabiensis* [64]. Research by Lukwa et al. [4] revealed that topical application of 5 mg/cm² of *L. javanica* leads to 100% protection against *Anopheles aegypti* for 8 hours. Mavundza et al. [154] screened dichloromethane and ethanol leaf extracts of *L. javanica* for adulticidal activity against *A. arabiensis*. The authors observed dichloromethane and ethanol activities of 45% and 55% mosquito mortality, respectively. These findings strengthen the view that *L. javanica* is a potential source of antimalarial agents and to some extent validate the traditional use of the plant species as mosquito repellent in India [47], South Africa [34, 49, 58], and Zimbabwe [4, 48].

4.4. Antioxidant Activity. Leaf infusions of *L. javanica* exhibited antioxidant activity with EC₅₀ value of 358 µg/mL and contained 14.8 mg/mL of dry weight gallic acid equivalent phenolic compounds [7]. The EC₅₀ value of 358 µg/mL obtained for *L. javanica* by Shikanga et al. [7] compares well to those of many commercial teas, including antioxidant capacity of Rooibos Fresh Pack™ herbal teas (*A. linearis*) with the best antioxidant activity of 333 µg/mL. The high antioxidant activities displayed by *L. javanica* infusions can be partially attributed to the high levels of verbascoside 2 (1.5 mg/g dry weight) reported by Olivier et al. [81] in the leaf extract of the species. Earlier research by Muchuweti et al. [125] reported 74.4% inhibition of the DPPH radical by an ethanolic leaf extract of *L. javanica*. Lekganyane et al. [75] reported antioxidant activity in *L. javanica* and the acetone extracts of the species displayed antioxidant activity on BEA chromatogram [77]. Bhebhe et al. [10] reported the antioxidant activities of *L. javanica* based on the DPPH, reducing power and inhibition of phospholipid peroxidation assays. Free radical scavenging activity of *L. javanica* is attributed to phenolic compounds since these compounds have an ideal structural chemistry for free radical scavenging activity [155]. Bhebhe et al. [11] determined the effect of several solvents on the free radical scavenging activity of *L. javanica* using the DPPH assay. Free radical scavenging activity expressed as IC₅₀ ranged from 0.022 ± 0.001 g/mL to 0.066 ± 0.001 g/mL; see Table 4.

4.5. Antiplasmodial Activity. Prozesky et al. [144] evaluated *L. javanica* leaf acetone extract for in vitro antiplasmodial activity using PfUP1, a chloroquine resistant strain of the malaria parasite *Plasmodium falciparum* by means of the flow cytometric test. The IC₅₀ value for *L. javanica* was 4.26 µg/mL. Manenzhe et al. [2] evaluated essential oil,

piperitenone **162**, isolated from *L. javanica* for antiplasmodial activity using chloroquine diphosphate as positive control and found it active against a chloroquine sensitive strain of *P. falciparum* in micromolar concentrations with IC₅₀ of 8 µg/mL. Clarkson et al. [80] evaluated *L. javanica* roots and stems extracts for in vitro activity against *P. falciparum* using the parasite lactase dehydrogenase (pLDH) assay and chloroquine diphosphate (Sigma) as the positive control. The dichloromethane, methane, and water extracts showed IC₅₀ values ranging from 3.8 to >100 µg/mL; see Table 4. Omolo et al. [145] screened the essential oil of *L. javanica* for fumigant toxicity to *Anopheles gambiae* which exhibited LD₅₀ of 4.3 × 10⁻³ mg cm⁻³. Ayuko et al. [79] showed that *L. javanica* root extracts have antiplasmodial activity against *P. falciparum* with IC₅₀ ranging from 1.35 to 18.59 µg/mL; see Table 4. Lippialactone **21**, derived from the ethyl acetate extract of aerial parts of *L. javanica*, exhibited some activity against the chloroquine sensitive D10 strain of *P. falciparum* with an IC₅₀ value of 9.1 µg/mL and is also mildly cytotoxic [84]. Compared to chloroquine, the compound is approximately 2000 times less active against the D10 strain of *P. falciparum* [84].

4.6. Antimicrobial Activities. *Lippia javanica* is widely used in the treatment of a wide range of infectious diseases caused by microorganisms. Viljoen et al. [3] determined the antimicrobial properties of *L. javanica* by evaluating the time kill studies of the species' essential oil using the disc diffusion assay on three respiratory pathogens *Klebsiella pneumoniae*, *Cryptococcus neoformans*, and *Bacillus cereus*. This study showed that the killing rate was greatest for *K. pneumoniae* followed by *C. neoformans* and very little reduction of microbial populations was observed for *B. cereus*. The efficacy of *L. javanica* oil for *K. pneumoniae* showed a killing rate within 30 minutes for the concentrations 0.25, 0.5, 0.75, and 1%, *C. neoformans* showed a killing rate for concentrations 0.5, 0.75, and 1% within 1 hour, and the lowest concentration of 0.25% took 8 hours before a bactericidal effect was noted while *B. cereus* showed some reduction in colonies [3]. The positive antimicrobial activity of *L. javanica* as revealed by the time kill study could be attributed to linalool **120** which averages between 65 and 70% in yield [3, 5, 88, 90] and has known antimicrobial properties [93, 156–159]. These findings somehow corroborate the traditional use of *L. javanica* as herbal medicine for a wide range of bacterial and fungal respiratory ailments indicated in Table 1.

Manenzhe et al. [2] evaluated essential oil, piperitenone **162**, isolated from *L. javanica* for antibacterial activity on cultures of *Bacillus subtilis*, *Staphylococcus aureus*, and *Escherichia coli* using imipenem, cefazolin, and ampicillin as positive controls. The authors found piperitenone **162** to inhibit *S. aureus* and *E. coli* at 1% dilution. Acetone, hexane, and methanol leaf extracts and essential oil isolated from *L. javanica* showed some activity against fifteen Gram-positive and Gram-negative bacteria with MIC values ranging from 1.5 to >12 mg/mL [60]; see Table 4. In a similar study, Samie et al. [73] demonstrated that a pure compound piperitenone **162** isolated from *L. javanica* has antibacterial activities against *Acinetobacter calcoaceticus*, *Micrococcus*

kristinae, *Salmonella typhi*, and *S. aureus* using dimethyl sulphoxide and kanamycin as controls, with MIC values ranging from 12 to 50 $\mu\text{g}/\text{mL}$; see Table 4. Shikanga et al. [7] evaluated the antibacterial activity of *L. javanica* methanolic leaf extract against *S. aureus*, *Enterococcus faecalis*, *E. coli*, and *Pseudomonas aeruginosa* using the serial microdilution method with gentamycin (Virbac®) and acetone as positive and negative controls, respectively. *Lippia javanica* displayed antibacterial activities with MIC values ranging from 0.13 to 0.42 mg/mL against all four pathogens; see Table 4. The obtained minimum inhibitory concentrations are promising, since natural products with MIC values below 1 mg/mL are generally considered to be noteworthy findings [160]. Lippialactone **21**, derived from the ethyl acetate extract of aerial parts of *L. javanica*, exhibited some activity against the *E. coli* and *S. aureus* at a concentration of 10 mg/mL [84]. Lekganyane et al. [75] reported antibacterial activity of *L. javanica* acetone leaf extracts against *E. coli*, *E. faecalis*, *P. aeruginosa*, and *S. aureus* with MIC values ranging from 0.32 to 0.64 mg/mL and total activity of the same species ranging from 127 to 253 mg/mL; see Table 4. Methanol and water leaf extracts of *L. javanica* exhibited some antiproteus activity against *Proteus mirabilis* and *Proteus vulgaris* with MIC values <2000 $\mu\text{g}/\text{mL}$ with standard discs of ampicillin (2 μg) and chloramphenicol (10 μg) as positive controls [74]. Some of the antibacterial properties of *L. javanica* can be attributed to the phytochemical constituents of the species; for example, the phenolic compound apigenin **7** is a well-known antibacterial agent [7, 134]. Apigenin **7** was shown to be highly active against *Vibrio cholera* and *E. faecalis* [161], while Basile et al. [162] reported the inhibition of *S. typhi*, *P. mirabilis*, and *P. aeruginosa* by the compound. Therefore, results from these antibacterial evaluations of *L. javanica* give credence to the use of the species' infusions against bacterial infections and other related diseases.

Shikanga et al. [126] investigated the antifungal activities of leaf extracts and essential oil compounds isolated from *L. javanica* leafy extracts against a Guazatine®-resistant strain of *Penicillium digitatum*. The methanolic leafy extracts, isoverbascoside **3** and verbascoside **2** compounds, isolated from *L. javanica* inhibited fungal growth at concentrations above 0.6 mg·mL⁻¹, causing significant inhibition of mycelial growth. Verbascoside **2** is well-known for its antimicrobial properties and has been found to inhibit viruses, bacteria, and fungi [163]. Similarly, Samie et al. [143] investigated antifungal effects of acetone and hexane leaf extracts of *L. javanica* against *Candida albicans*, *Candida krusei*, and *C. neoformans* isolated from AIDS patients using the microdilution method with nystatin, Roche, and DMSO as positive and negative controls, respectively. Noteworthy moderate antifungal activities were recorded from *C. krusei* with MIC value of 1.88 mg/mL and other recorded MFC and MIC values for other species ranged from 1.88 mg/mL to >7.5 mg/mL; see Table 4. Thembo et al. [76] investigated the antifungal activity of aqueous and organic extracts of *L. javanica* using a serial microdilution assay. Generally, extracts of *L. javanica* exhibited weak activity; see Table 4.

4.7. Antimycobacterial Activity. Mujovo et al. [83] evaluated *L. javanica* compounds against a drug-sensitive strain of *Mycobacterium tuberculosis* using the radiometric respiratory techniques. Of all the isolated compounds, only one triterpenoid carboxylic acid, euscaphic acid **172**, exhibited antimycobacterial activity with MIC value of 50 $\mu\text{g mL}^{-1}$ against this strain. In a similar study, the leaf extract of *L. javanica* exhibited antimycobacterial activity against *M. smegmatis* in an evaluation which used microdilution assay and rifampicin as control [77]. Acetone extract was the best extractant with MIC value of 0.47 mg/mL; it extracted antibacterial agents which was indicated by the lowest MIC value [77]. Masoko and Nxumalo [77] also evaluated the total activity of *L. javanica* which averaged 13 mL/g suggesting that the extract prepared from one gram of *L. javanica* could be diluted to a volume of 13 mL and will still inhibit *M. smegmatis* efficiently. According to Semenya and Maroyi [115], tuberculosis caused by *M. tuberculosis* is a serious disease requiring effective strategies and tools to control and manage it. Therefore, preliminary evaluations done by Mujovo et al. [83] and Masoko and Nxumalo [77] serve as a scientific validation for the use of *L. javanica* in traditional medicine for treatment of tuberculosis and other respiratory ailments in South Africa [24, 27, 115] and Uganda [43] as well as their efficiency in tuberculosis drug discovery.

4.8. Antiviral Activity. Mujovo et al. [83] found that (E)-2(3)-tagetone epoxide **42** and piperitenone **162** inhibited the HIV-1 reverse transcriptase enzyme by 91% and 53%, respectively, at 100 $\mu\text{g mL}^{-1}$ based on a nonradioactive HIV RT colorimetric ELISA kit. Little is known about the HIV RT activity of *L. javanica* extracts or compounds, but flavonoids are known to be active against viral RT and also as potent inhibitors of the cellular alpha and beta DNA polymerase [83], while luteolin **12** is active against HIV RT [164, 165]. *Lippia javanica* is traditionally used to treat HIV/AIDS symptoms in South Africa [65, 66] and several viral and HIV/AIDS opportunistic diseases and infections such as bronchitis [15, 19–22], chicken pox [50], diarrhoea [17, 34, 37, 44, 113], measles [14, 20, 38], pneumonia [14], shingles [15], and venereal diseases [44]. Despite significant advances in the utilization of *L. javanica* in southern Africa over the years for numerous viral diseases (Table 1), very little antiviral evaluations have been done on crude extracts and purified compounds of the species. There is need, therefore, for more pharmacological research as *L. javanica* could be harbouring potent (RT) inhibitors which could be useful for the development of new pharmaceutical products important for use against viral diseases and infections.

4.9. Pesticidal Effects. Magano et al. [95] evaluated the repellent effects of hexane extracts of *L. javanica* essential oil using the in vitro tick climbing repellency bioassay on adults of *Hyalomma marginatum rufipes* Koch ticks. The authors found that 107 mg/mL caused repellency index of 100% at one hour and 30 minutes. In a similar study by Madzimure et al. [120], *L. javanica* aqueous leaf extracts at 10% and 20% w/v were effective at controlling cattle ticks (*Amblyomma* species,

Boophilus species, *Hyalomma* species, *Rhipicephalus appendiculatus*, and *Rhipicephalus evertsi evertsi*) and were as good as the positive control amitraz-based acaricide Trickbuster®. These authors found no parasites on microscopic examination of the Giemsa-stained thin blood smear collected from treated cattle implying that the animals did not suffer from clinical tick-borne diseases. Similarly, Nyahangare et al. [59] tested the pesticidal activity of *L. javanica* water extracts against cattle ticks. The authors found no significant difference between cattle treated with a commercial synthetic acaricide and those under *L. javanica* treatment. Martinez-Velazquez et al. [166] evaluated pesticidal effects of two essential oils, namely, γ -terpinene **82** and p -cymene **99**, isolated from *Lippia graveolens* Kunth against 10-day-old *Rhipicephalus (Boophilus) microplus* (Canestrini) tick larvae using the larval packet test bioassay. The two essential oils produced high mortality ranging from 90 to 100% in all tested concentrations of 20 to 1.25% (v/v). Therefore, *L. javanica* can provide an effective tick control option where synthetic products are unavailable or unaffordable particularly in remote rural areas in sub-Saharan Africa. *Lippia javanica* is popular for tick control and management among resource-poor smallholder farmers in South Africa [58, 95] and Zimbabwe [59, 60, 120]. While these preliminary evaluations may serve as confirmation that *L. javanica* has some bioactivities against ticks, a comprehensive method of tick control is required for the resource-constrained smallholder farmers based on ethnopharmacological properties of *L. javanica*.

McGaw et al. [146] screened *L. javanica* for the anthelmintic test systems using the free living nematode *Caenorhabditis elegans*. The crude ethanol and hexane extracts showed some activity at a concentration of 2 mg/mL, with the 7-day incubation assay appearing to be more sensitive than the shorter assay. Earlier research by Mwangi et al. [1] indicated that *L. javanica* was active against *Aedes aegypti* larvae and *Sitophilus zeamais* Motschulsky (maize weevil). Katsvanga and Chigwiza [147] reported that *L. javanica* is an effective natural pesticide which can be used to control aphid species (*Brevicoryne brassicae*). In their study, Katsvanga and Chigwiza [147] found that 1:1 powdered aqueous leaf extract of *L. javanica* reduced *B. brassicae* by 53.2% against 78.3% and 96.7% of two synthetic pesticides, Aphid kill and Bexadust "L," respectively. Chikukura et al. [149] found *L. javanica* powdered leaf extracts to have insecticidal properties with potential to control grain damage by 21–33%. Mashela et al. [148] reported that the application of ground *L. javanica* leaves to soil in nematode-infested (*Meloidogyne incognita*) pots reduced the nematode numbers by 79–92% and significantly increased fresh fruit yield, dry shoot mass, plant height, and stem diameter of the tomato plants, as well as levels of potassium, nitrogen, and manganese in leaf tissue. Muzemu et al. [150] evaluated water extracts of *L. javanica* leaf powder for pesticidal effects against rape (*Brassica napus* L.) aphids, *B. brassicae*, and tomato (*Solanum lycopersicum* Lam.) red spider mites, and *Tetranychus evansi* as alternatives to conventional pesticides. *Lippia javanica* reduced *B. brassicae* and *Tetranychus evansi* by 12.5% and 63%, respectively [150]. The study demonstrated that *L. javanica* has pesticidal effects on *B. brassicae* and *T.*

evansi. The reduced number of *B. brassicae* and *T. evansi* could be due to *L. javanica*'s extracts having repellent, toxic, and antifeedant effects since the species has essential oils with pesticidal properties [2]. Therefore, these findings indicate that *L. javanica* has both nematocidal and plant growth-promoting properties.

4.10. Toxicity and Cytotoxicity Activity. *Lippia javanica* is known to cause liver damage and photosensitisation in livestock, resulting in stock losses [167]. Triterpenoids isolated from the genus *Lippia* are icterogenic and cause jaundice as a result of liver damage [167]. The characteristic swelling, yellowing, and later peeling of unpigmented skin are due to the presence of phylloerythrin, a photodynamic porphyrin that reacts with sunlight and causes severe cell damage [167]. The compound is normally formed when chlorophyll is broken down by microorganisms in the rumen, but it now accumulates in the liver as a result of the damage caused by triterpenoids [167]. In view of the known toxicity of *Lippia* species, the prolonged use of high doses of *L. javanica* is potentially harmful [167]. *Lippia javanica* showed low toxicity after 48 h exposure with the percentage of mortality below 50% [74].

Ayuko et al. [79] evaluated the toxicity of *L. javanica* using a brine shrimp cytotoxicity assay with LC₅₀ value of 1138 ± 1.33 µg/mL. Ayuko et al. [79] found the cytotoxicity to antiparasitic activity ratios for the methanolic extracts of the two tested strains to be 843.0 and 650.3, and since these are greater than 100, it may be concluded that the extracts are of low toxicity. Samie et al. [73] demonstrated that a pure compound piperitenone **162** isolated from *L. javanica* essential oil has low cytotoxicity activity against intestinal adenocarcinoma cells (i.e., the HCT-8 monolayers with IC₅₀ of 265.6 ± 5.3 µg/mL). Lukwa [64] evaluated the toxicity of *L. javanica* aqueous leaf extracts using sexually mature BALB/c mice with the placebo as control. Within 48 hours, all mice fed with the *L. javanica* leaf aqueous extract at 12.5–37.5% v/v were lethargic, and the overall mortality was 37.5%. Previous research by Manenzhe et al. [2] showed that hydrodistillation of *L. javanica* leaves, flowers, and stems produced oil that was poisonous against *P. falciparum* when diluted to 1% (v/v). These findings imply that, despite its apparent safety, water extracts of *L. javanica* leaves may have deleterious health implications on humans and animals if consumed at very high doses.

Many compounds that have been isolated from *L. javanica* including phenolic glycosides, flavonoids, and essential oils are not known to have acute toxic properties with the exception of icterogenin **173**. Icterogenin **173** has been shown to inhibit biliary excretion in rabbits [168]. Reports in literature indicate that the consumption of xanthine **22** has resulted in mammalian toxicity. According to Madzimure et al. [64], xanthine **22** is a demethylated derivative of caffeine with pharmacological actions such as central nervous system (CNS) stimulation, relaxation of smooth muscle (especially bronchial muscle), myocardial stimulation, peripheral vasoconstriction, and diuresis. Considering the widespread use of *L. javanica* as herbal tea and medicine, it is important

to determine if any toxicological effects can occur from its chronic or subchronic usage.

4.11. Other Activities. Mpofu et al. [169] determined the effect of inclusion of *L. javanica* leaf meal in broiler diets on growth performance, carcass characteristics, and fatty acid profiles. The authors found that the *L. javanica* fed broilers had higher total polyunsaturated fatty acids and n-3 fatty acids. The findings from the study showed that inclusion of *L. javanica* in broiler diets at 5 g/kg feed has positive influences on growth performance, carcass characteristics, and fatty acid profiles of broiler meat and, therefore, *L. javanica* has potential as growth-promoting feed additive in broilers. Samie et al. [73] evaluated antiameobic activity of a pure compound piperitenone **162** isolated from *L. javanica* essential oil against *Entamoeba histolytica* using microdilution method with metronidazole as the positive control, diluents (i.e., culture medium with an appropriate concentration of dimethyl sulphoxide as the negative control), and a blank (i.e., culture medium without dimethyl sulphoxide). Samie et al. [73] demonstrated that piperitenone **162** had marked antiameobic activity with IC₅₀ value of 25 µg/mL. More research is required as *L. javanica* could be harbouring potent antiameobic properties which could be useful for managing amoebiasis, an infection that remains a significant cause of morbidity and mortality worldwide.

5. Conclusion

Lippia javanica has been used in African and Asian countries as herbal tea and medicine for many centuries. Utilization of *L. javanica* because of its flavour and medicinal properties forms the basis of the current demand for the plant species in central, eastern, and southern Africa. Research on *L. javanica* over the past decade on health promoting properties has greatly contributed to the increased consumption of the species as herbal or recreational tea. The focus of this research has been on phytochemical compounds, particularly phenolic content and antioxidant and free radical scavenging activities. Phenolic compounds present in *L. javanica* are largely responsible for the antioxidant properties possessed by the species. More research in this regard is required and future research should focus on more comprehensive chemical characterization of both crude and pure extracts and evaluate potential for commercialization and development of nutraceutical products based on traditional uses of *L. javanica*. Most of the pharmacological researches conducted on *L. javanica* so far have focused on the phytochemistry and biological properties of leaves, and little or no research has been done on roots and other plant parts. Therefore, future research on the species should focus on other plant parts, for example, flowers, roots, and stems, as well as organ-to-organ, age, and seasonal variation evaluations in the phytochemical content and pharmacological activities of the species.

The recent increase in the demand for *L. javanica* products may partly be ascribed to growing body of scientific evidence indicating important health benefits. *Lippia javanica*

is widely sold as herbal tea in Botswana, South Africa, and Zimbabwe. Leaves and stems of *L. javanica* are also sold as herbal medicines in the medicinal plant “muthi” markets in South Africa [35, 170]. For local people who rely on herbal medicines as part of their primary healthcare as well as cultural beliefs, they prefer *L. javanica* harvested from the wild and unprocessed plant parts sold in informal medicinal “muthi” markets [170]. In the past, there were no records of overexploitation of *L. javanica* wild populations in southern Africa, resulting in Raimondo et al. [171] listing the species as Least Concern (LC) under the IUCN Red List Categories and Criteria version 3.1 of threatened species (<http://www.iucnredlist.org>). Recently, signs of overharvesting have been noted, where local people or plant traders have uprooted whole plants to supply medicinal plant “muthi” markets or use the plants as herbal medicines, brooms, or herbal tea. Therefore, large-scale commercial utilization of *L. javanica* is not sustainable if the species is harvested from the wild. Currently, *L. javanica* is cultivated on a commercial scale in Kenya [28] and South Africa [117] for essential oil production for the mosquito-repellent candles and perfume industry. Cultivation of *L. javanica* is therefore, a solution to the sustainability problems associated with harvesting of the species from the wild, and this option is also necessary for establishing commercial scale medicinal production and processing and trade enterprises. The success of commercial cultivation of *L. javanica* will depend on how the species is marketed as herbal tea and medicine and source of essential oil and health care products and as a source of functional foods.

Significant research has been made in the past 50 years into the chemistry and pharmacology of *L. javanica*. These studies have shown *L. javanica* to display various chemical and different biological activities some of which justify its ethnopharmacological utilization in variety of cultures. In the light of the evidence that *L. javanica* is combined with other plant species in traditional medicine, it will be valuable to investigate the possibility of synergistic effects of the different extracts. Deep phytochemical studies of *L. javanica* and its phytochemical properties, especially the mechanisms of action of its bioactive constituents to illustrate the correlation between ethnomedicinal uses and pharmacological activities, should be the focus of further research on the species. There is need for extensive in vivo experiments to validate the existing pharmacological activities. However, because *L. javanica* contains potentially toxic compounds, its toxicological properties need to be properly established via proper quality control of product development to ensure that potentially toxic components are kept below tolerance levels.

Appendix

A. Chemical Compounds Isolated and Characterized from *Lippia javanica*

See Table 5.

TABLE 5

Chemical compound	Reference(s)
<i>Phenolic compounds</i>	
Coumarin 1	[4]
Verbascoside 2	[81]
Isoverbascoside 3	[81]
Theveside-Na 4	[82]
Theveridoside 5	[82]
4-ethylnonacosane 6	[83]
Apigenin 7	[64, 83]
Cirsimaritin 8	[64, 83]
6-Methoxy luteolin 4'-methyl ether 9	[83]
6-Methoxy luteolin 3',4',7-trimethyl ether 10	[83]
Crassifolioside 11	[64]
Luteolin 12	[64]
Diosmetin 13	[64]
Chrysoeriol 14	[64]
Tricin 15	[64]
Isothymusin 16	[64]
Eupatorin 17	[64]
5-Dimethyl noboletin 18	[64]
Genkwanin 19	[64]
Salvigenin 20	[64]
Lippialactone 21	[84]
<i>Alkaloid</i>	
Xanthine 22	[64]
<i>Amino acids</i>	
α -Aminobutyric acid 23	[85, 86]
Valine 24	[86]
Isoleucine 25	[86]
Asparagine 26	[86]
Phenylalanine 27	[86]
α -Amino adipic acid 28	[86]
Lysine 29	[86]
Histidine 30	[86]
Tyrosine 31	[86]
Tryptophan 32	[86]
Alanine 33	[85]
Glycine 34	[85]
Proline 35	[85]
Serine 36	[85]
Glutamine acid 37	[85]
β -Alanine 38	[85]
Glutamine 39	[85]
β -Aminoisobutyric acid 40	[85]
4-hydroxyproline 41	[85]
<i>Essential oil</i>	
(E)-2(3)-tagetone epoxide 42	[83]
4-Methyl-2-pentanone 43	[3]
α -Pinene 44	[3–5, 87–91]

TABLE 5: Continued.

Chemical compound	Reference(s)
1,3-5-Cycloheptatriene 45	[4]
(+)-2-Carene 46	[4]
3-Carene 47	[4]
Eucalyptol 48	[4]
1.8 myrcene 49	[87]
Ipsdienone 50	[87]
Caryophyllene 51	[85]
Geranial 52	[5, 92, 93]
2,6-Dimethylstyrene 53	[92]
Geraniol 54	[92]
Octen-3-one 55	[5]
6-Methyl-5-hepten-2-one 56	[5]
<i>p</i> -Mentha-1(7),8-diene 57	[5]
Artemisia ketone 58	[5]
Linalool oxide 59	[5]
Terpinen-4-ol 60	[5]
(Z)- β -Ocimenone 61	[5]
(E)- β -Ocimenone 62	[5]
Carvyl acetate 63	[5]
α -Cubebene 64	[5]
Sesquithujene 65	[5]
Acora-3,5-diene 66	[5]
β -Bergamotene 67	[5]
Trans-calamenene 68	[5]
β -Alaskene 69	[5]
γ -Cadinene 70	[5]
δ -Cadinene 71	[3, 88, 89, 91]
Cis-calamenene 72	[5]
Nerolidol 73	[5]
(E)-Nerolidol 74	[3, 88, 89]
Spathulenol 75	[5]
Epi- α -muurolol 76	[5]
α -Longipinene 77	[90]
Chrysanthenone 78	[90]
α -Terpineol 79	[3, 5, 90, 91]
α -Amorphene 80	[90]
α -Thujene 81	[91]
γ -Terpinene 82	[91]
α -Cubebene 83	[91]
Linalool acetate 84	[91]
Bicyclosiquiphellandrene 85	[91]
Camphene 86	[3, 5, 88–91]
β -Pinene 87	[3, 5, 91]
Sabinene 88	[3, 5, 87, 90, 91]
Myrcene 89	[3, 5, 87–89, 91–95]
α -Phellandrene 90	[3, 5, 88, 89]
2-Methylbutyl isobutyrate 91	[3]
Limonene 92	[3, 5, 88–94]
1,8-Cineole 93	[3, 5, 90, 91, 95]
β -Phellandrene 94	[3, 4, 88, 89, 91]

TABLE 5: Continued.

Chemical compound	Reference(s)
(Z)-3-Hexenal 95	[3]
(Z)- β -Ocimene 96	[3, 5, 88, 89, 91]
(E)- β -Ocimene 97	[3, 5, 88, 89, 91]
Isomyrcenol 98	[3]
<i>p</i> -Cymene 99	[3, 5, 85, 88–91]
2-Methylbutyl-2-methyl butyrate 100	[3]
Terpinolene 101	[3]
Dihydrotagetone 102	[3, 88, 89, 91, 95]
Cis-alloocimene 103	[3, 88, 89]
(Z)-3-Hexen-1-ol 104	[3]
6,7-Epoxymyrcene 105	[3]
Nonanal 106	[3]
Perillene 107	[3, 91]
Ipsenone 108	[3, 87–89, 95]
<i>Trans</i> -Linalool oxide (<i>furanoide</i>) 109	[3, 88–90]
1-Octen-3-ol 110	[3, 88, 89]
Cis-1,2-limonene epoxide 111	[3]
<i>Trans</i> -1,2-limonene epoxide 112	[3]
<i>cis</i> -Linalool oxide (<i>furanoide</i>) 113	[3, 88–90]
α -Copaene 114	[3, 5, 88–90]
<i>Cis</i> -tagetone 115	[3, 88, 89]
<i>Trans</i> -tagetone 116	[3, 88, 89]
Camphor 117	[3, 5, 88–90, 93, 94]
β -Bourbonene 118	[3, 5, 91]
Benzaldehyde 119	[3, 5]
Linalool 120	[3, 5, 85, 88–90, 92, 93]
<i>Trans</i> - α -bergamotene 121	[3]
α -Cedrene 122	[3, 5]
Myrcenone 123	[3, 5, 83, 91]
β -Caryophyllene 124	[3, 5, 87–91, 93, 94]
2-Methyl-6-methylene-3,7-octadien-2-ol 125	[3]
<i>Trans-p</i> -mentha-2,8-dien-1-ol 126	[3]
<i>Cis-p</i> -mentha-2,8-dien-1-ol 127	[3]
Alloaromadendrene 128	[3, 5, 88, 89]
(Z)- β -Farnesene 129	[3]
(E)- β -Farnesene 130	[3, 5]
(E,E)- α -Farnesene 131	[3, 88, 89]
Ipsdienol 132	[3]
(Z)-3-Hexenyl tiglate 133	[3]
Isovaleric acid 134	[3]
α -Humulene 135	[3, 5, 88, 89, 91]
α -Acoradiene 136	[3]
β -Acoradiene 137	[3]
γ -Muulolene 138	[3, 5, 88, 89]
α -Muulolene 139	[3, 5, 88–90]
<i>Cis</i> -tagetone 140	[3]

TABLE 5: Continued.

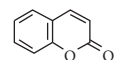
Chemical compound	Reference(s)
<i>Trans</i> -tagetone 141	[3]
Borneol 142	[3, 5, 88–90]
Verbenone 143	[3, 91]
Germacrene-D 144	[3, 87–89, 93]
β -Bisabolene 145	[3, 5]
γ -Bisabolene 146	[3]
<i>Trans</i> -carvyl acetate 147	[3]
Carvone 148	[3, 5, 91]
Bicyclogermacrene 149	[3, 5, 88, 89, 91]
β -Curcumene 150	[3]
Ar-curcumene 151	[3]
<i>Cis</i> -piperitol 152	[3]
2-Methyl-2-butenic acid 153	[3]
<i>Trans-p</i> -mentha-1(7),8-dien-2-ol 154	[3]
<i>Cis-p</i> -mentha-1(7),8-dien-2-ol 155	[3]
2,6-Dimethyl-3(E),5(E),7-octatriene-2-ol 156	[3, 88, 89]
<i>Trans</i> -carveol 157	[3]
Calamenene 158	[3]
Carvone oxide 159	[3]
Isopiperitenone 160	[3]
<i>Cis</i> -carveol 161	[3]
Piperitenone 162	[2, 3, 73, 83]
Isocaryophyllene oxide 163	[3, 88, 89]
Caryophyllene oxide 164	[3–5, 88–90]
Humulene epoxide II 165	[3, 5, 88, 89]
Hexahydrofarnesyl acetone 166	[3]
Spathulenol 167	[3]
Eugenol 168	[3, 5]
Germacrene-D-4-ol 169	[3]
Caryophylla-2(12),6(13)-dien-5 β -ol (=Caryophylladienol I) 170	[3]
Caryophylla-2(12),6(13)-dien-5 α -ol (=Caryophylladienol II) 171	[3]
Euscaphic acid 172	[83]
Icterogenin 173	[96]

B. Chemical Structures of Phenolic Compounds, Alkaloids, Amino Acids, and Essential Oils Isolated from *Lippia javanica*

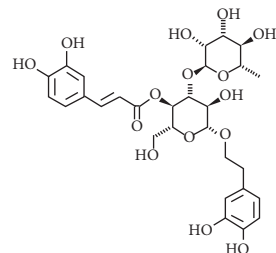
See Figure 4.

Competing Interests

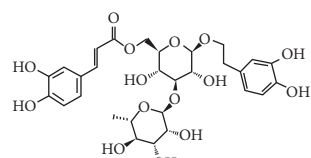
The author declares that there is no conflict of interests regarding the publication of this paper.



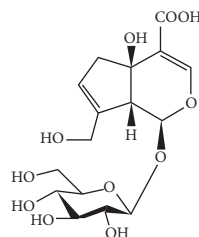
Coumarin 1 [4]



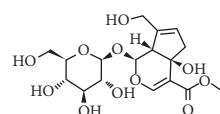
Verbascoside 2 [89]



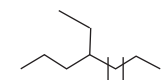
Isoverbascoside 3 [89]



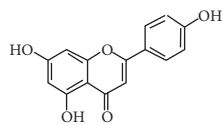
Theveside-Na 4 [90]



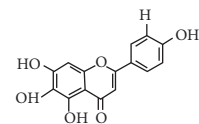
Theveridoside 5 [90]



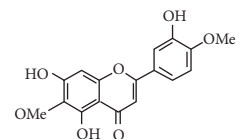
4-Ethylnonacosane 6 [91]



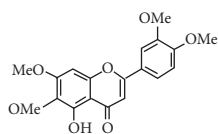
Apigenin 7 [120, 91]



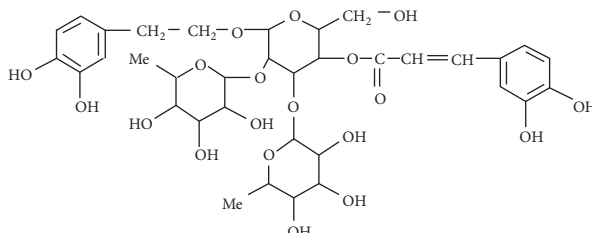
Cirsimaritin 8 [120, 91]



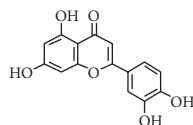
6-Methoxyluteolin 4'-methyl ether 9 [91]



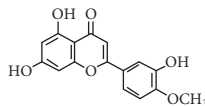
6-Methoxyluteolin 3',4',7-trimethyl ether 10 [91]



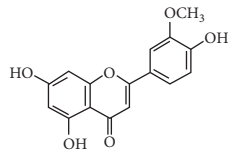
Crassifolioside 11 [120]



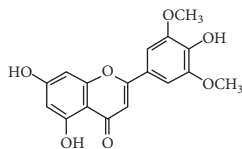
Luteolin 12 [120]



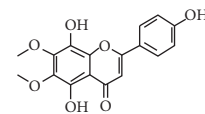
Diosmetin 13 [120]



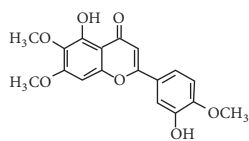
Chrysoeriol 14 [120]



Tricin 15 [120]



Isothymusin 16 [120]



Eupatorin 17 [120]

FIGURE 4: Continued.

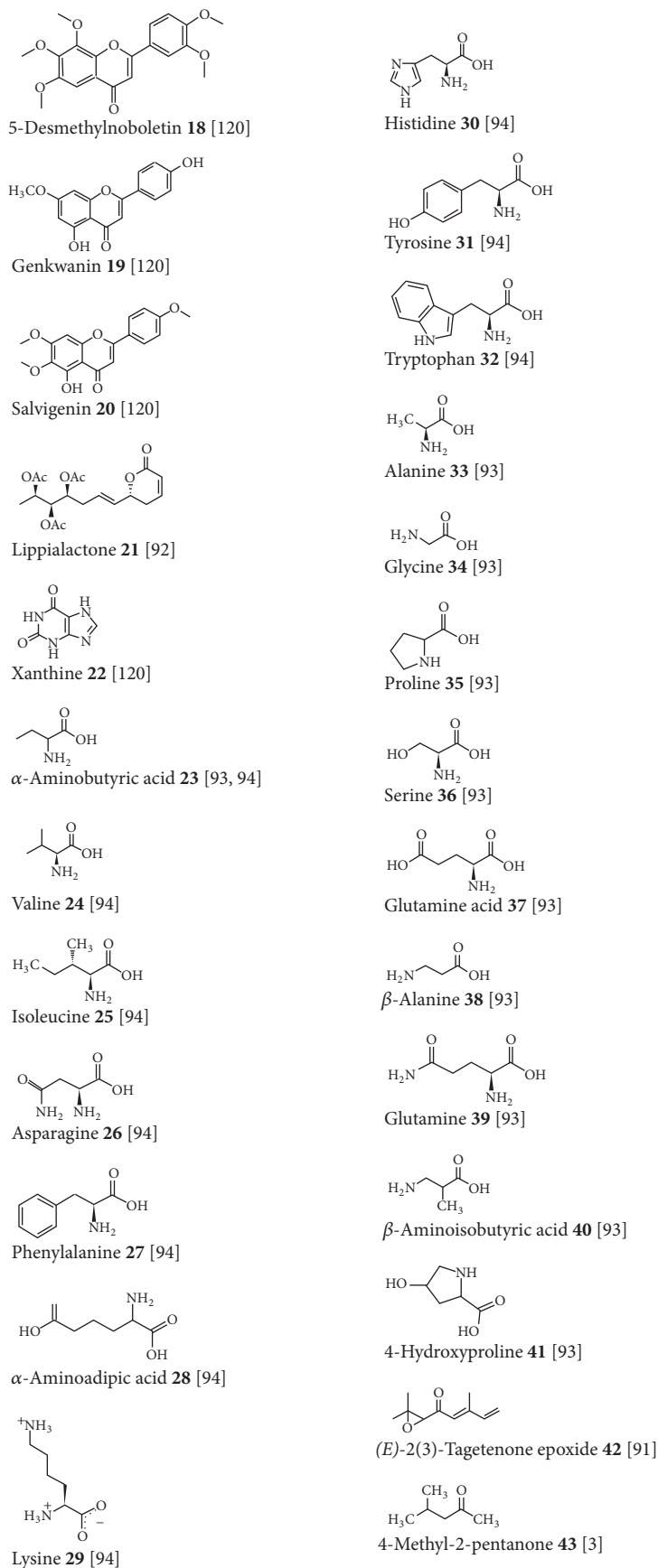


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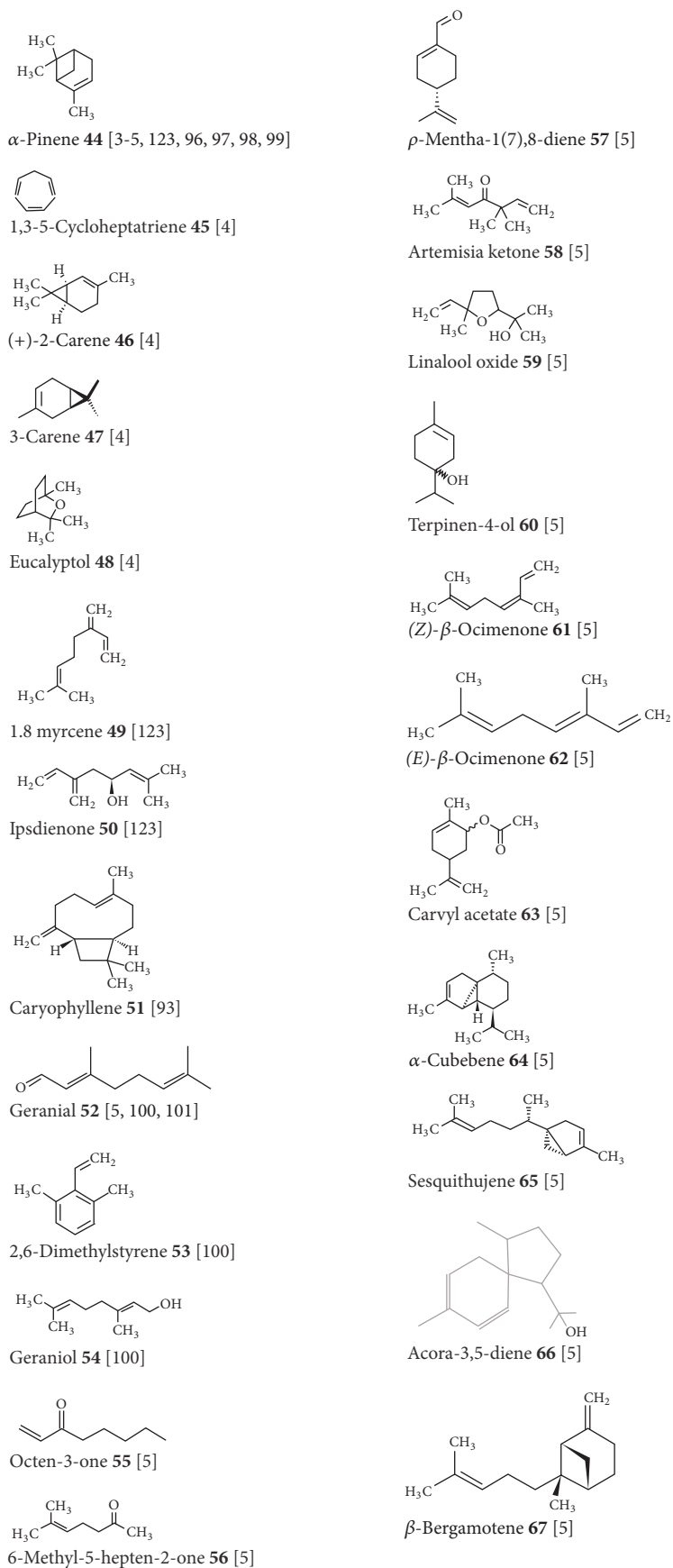


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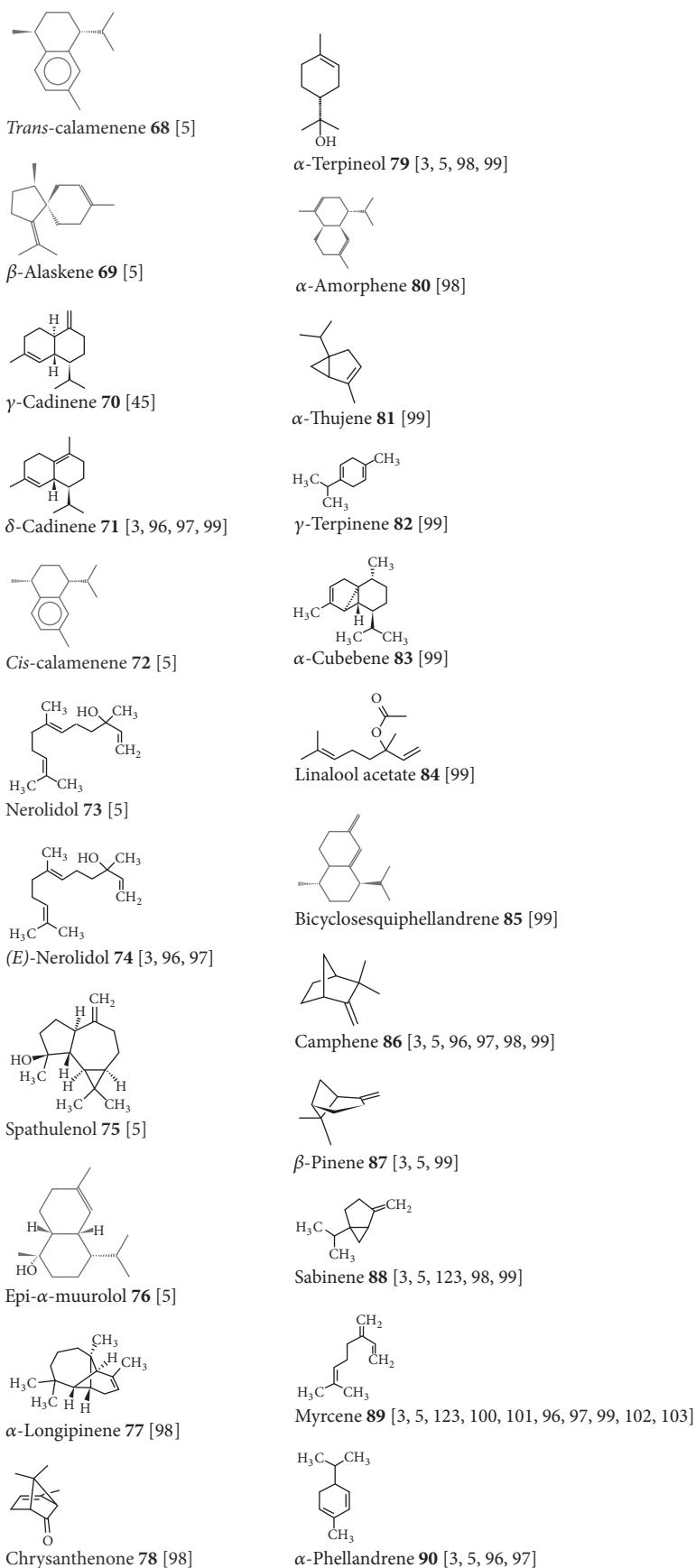


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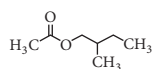
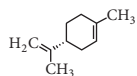
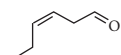
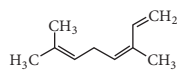
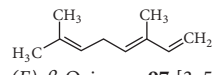
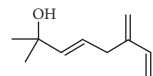
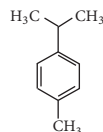
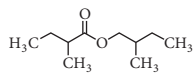
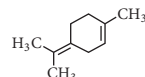
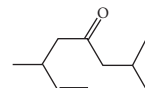
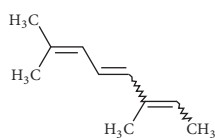
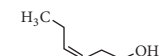
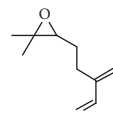
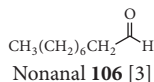
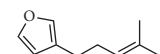
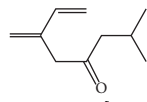
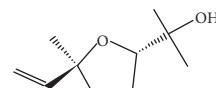
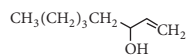
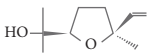
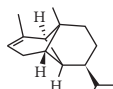
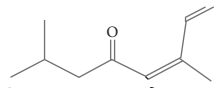
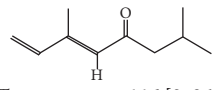
2-Methylbutyl isobutyrate **91** [3]Limonene **92** [3, 5, 100, 101, 96, 97, 98, 99, 102]1,8-Cineole (=eucalyptol) **93** [3, 5, 98, 99, 103] β -Phellandrene **94** [3, 4, 96, 97, 99](Z)-3-Hexenal **95** [3](Z)- β -Ocimene **96** [3, 5, 96, 97, 99](E)- β -Ocimene **97** [3, 5, 96, 97, 99]Isomyrcenol **98** [3]*p*-Cymene **99** [3, 5, 93, 96, 97, 98, 99]2-Methylbutyl-2-methyl butyrate **100** [3]Terpinolene **101** [3]Dihydrotagetone **102** [3, 96, 97, 99, 103]Cis-alloocimene **103** [3, 96, 97](Z)-3-Hexen-1-ol **104** [3]6,7-Epoxymenthene **105** [3]Nonanal **106** [3]Perillene **107** [3, 99]Ipsenone **108** [3, 123, 96, 97, 103]Trans-linalool oxide (furanoid) **109** [3, 96, 97, 98]1-Octen-3-ol **110** [3, 96, 97]Cis-1,2-limonene epoxide **111** [3]Trans-1,2-limonene epoxide **112** [3]Cis-linalool oxide (furanoid) **113** [3, 96, 97, 98] α -Copaene **114** [3, 5, 96, 97, 98]Cis-tagetone **115** [3, 96, 97]Trans-tagetone **116** [3, 96, 97]Camphor **117** [3, 5, 101, 96, 97, 98, 102]

FIGURE 4: Continued.

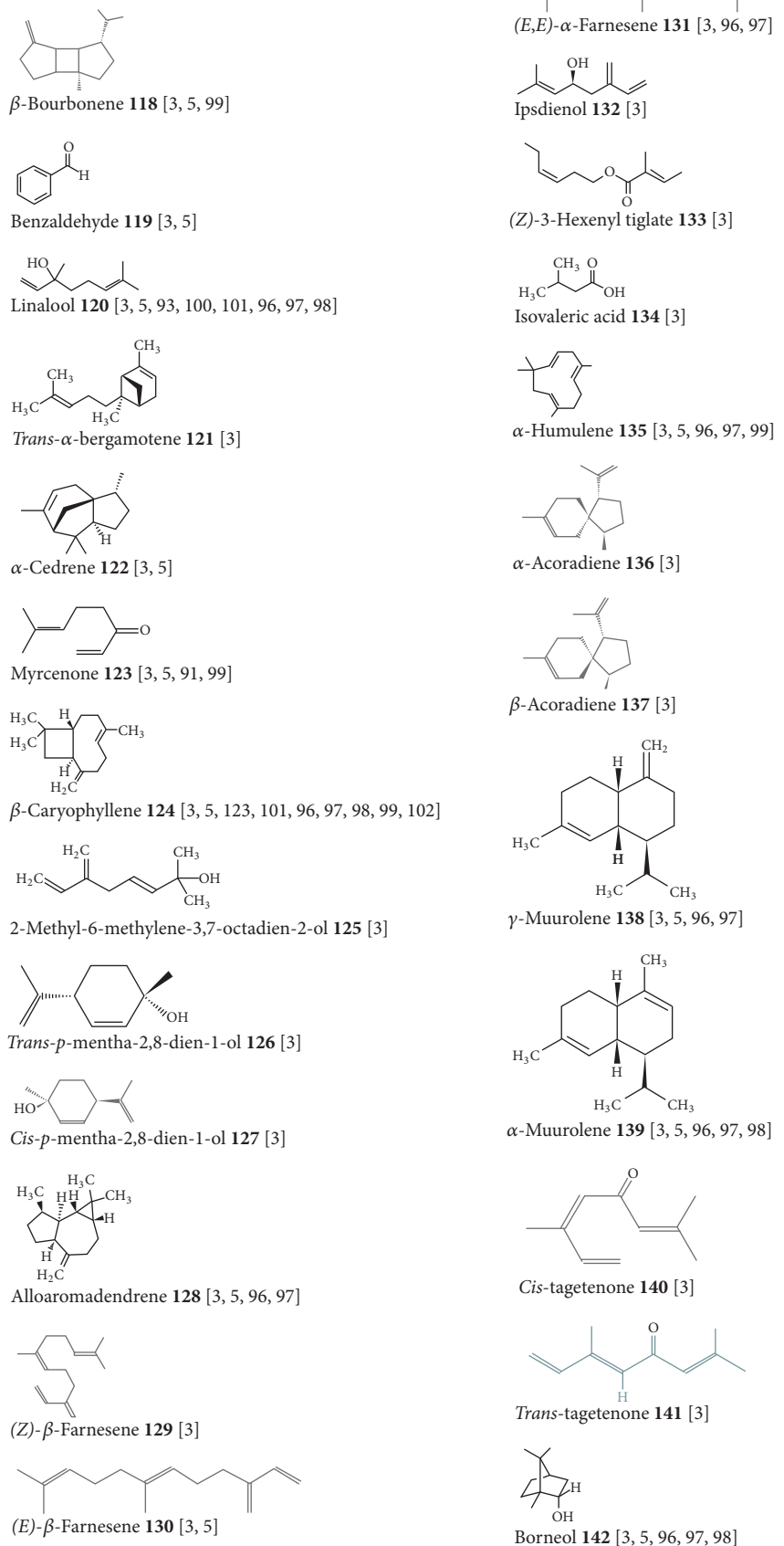


FIGURE 4: Continued.

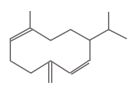
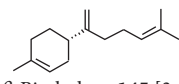
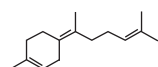
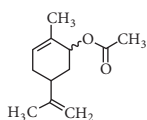
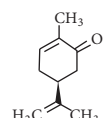
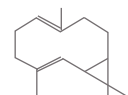
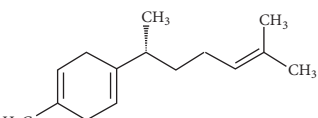
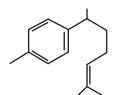
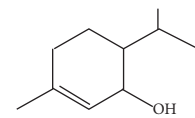
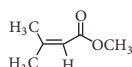
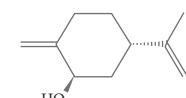
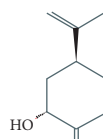
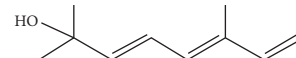
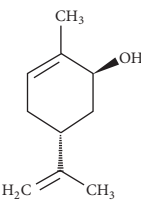
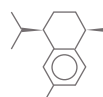
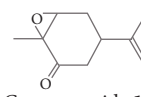
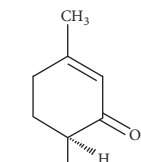
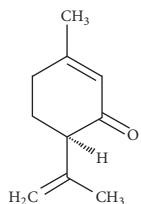
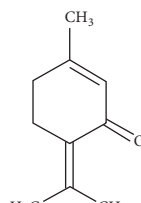
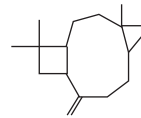
Verbenone **143** [3, 99]Germacrene-D **144** [3, 123, 101, 96, 97] β -Bisabolene **145** [3, 5] γ -Bisabolene **146** [3]*Trans*-carvyl acetate **147** [3]Carvone **148** [3, 5, 99]Bicyclogermacrene **149** [3, 5, 96, 97, 99] β -Curcumene **150** [3]Ar-curcumene **151** [3]*Cis*-piperitol **152** [3]2-Methyl-2-butenoic acid **153** [3]*Trans*-*p*-mentha-1(7),8-dien-2-ol **154** [3]*Cis*-*p*-mentha-1(7),8-dien-2-ol **155** [3]2,6-Dimethyl-3(E),5(E),7-octatriene-2-ol **156** [3, 96, 97]*Trans*-carveol **157** [3]Calamenene **158** [3]Carvone oxide **159** [3]Isopiperitenone **160** [3]*Cis*-carveol **161** [3]Piperitenone **162** [2, 3, 91, 81]Isocaryophyllene oxide **163** [3, 96, 97]

FIGURE 4: Continued.

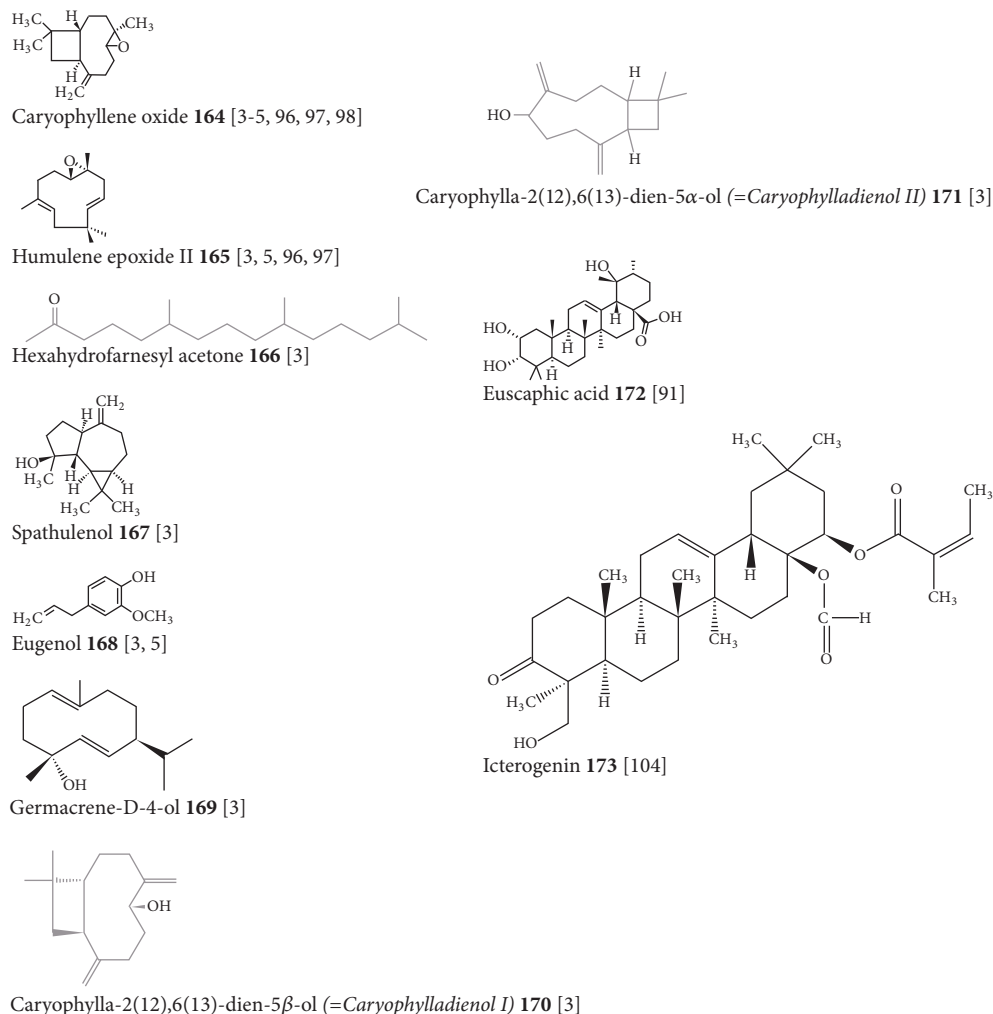


FIGURE 4

Acknowledgments

The author would like to express his gratitude to the National Research Foundation (NRF) and Govan Mbeki Research and Development Centre (GMRDC), University of Fort Hare, for financial support to conduct this research.

References

- [1] J. W. Mwangi, I. Addae-Mensah, R. M. Munavu, and W. Lwande, "Essential oils of Kenyan *Lippia* species. Part III," *Flavour and Fragrance Journal*, vol. 6, no. 3, pp. 221–224, 1991.
- [2] N. J. Manenzhe, N. Potgieter, and T. Van Ree, "Composition and antimicrobial activities of volatile components of *Lippia javanica*," *Phytochemistry*, vol. 65, no. 16, pp. 2333–2336, 2004.
- [3] A. M. Viljoen, S. Subramoney, S. F. V. Vuuren, K. H. C. Başer, and B. Demirci, "The composition, geographical variation and antimicrobial activity of *Lippia javanica* (Verbenaceae) leaf essential oils," *Journal of Ethnopharmacology*, vol. 96, no. 1-2, pp. 271–277, 2005.
- [4] N. Lukwa, P. Mølgaard, P. Furu, and C. Bøgh, "*Lippia javanica* (Burm. f.) Spreng: its general constituents and bioactivity on mosquitoes," *Tropical Biomedicine*, vol. 26, no. 1, pp. 85–91, 2009.
- [5] L. S. Chagonda and J.-C. Chalchat, "Essential oil composition of *Lippia javanica* (Burm.f.) spreng chemotype from Western Zimbabwe," *Journal of Essential Oil Bearing Plants*, vol. 18, no. 2, pp. 482–485, 2015.
- [6] W. Kipkore, B. Wanjohi, H. Rono, and G. Kigen, "A study of the medicinal plants used by the Marakwet Community in Kenya," *Journal of Ethnobiology and Ethnomedicine*, vol. 10, article 24, 2014.
- [7] E. A. Shikanga, S. Combrinck, and T. Regnier, "South African *Lippia* herbal infusions: total phenolic content, antioxidant and antibacterial activities," *South African Journal of Botany*, vol. 76, no. 3, pp. 567–571, 2010.
- [8] N. T. Parrant, "The potential of non-timber forest products of Botswana," *Botswana Notes Records*, vol. 28, no. 1, pp. 203–218, 1996.
- [9] D. M. Motlhanka and S. W. Makhabu, "Medicinal and edible wild fruit plants of Botswana as emerging new crop opportunities," *Journal of Medicinal Plants Research*, vol. 5, no. 10, pp. 1836–1842, 2011.
- [10] M. Bhebhe, T. N. Füller, B. Chipurura, and M. Muchuweti, "Effect of solvent type on total phenolic content and free radical scavenging activity of black tea and herbal infusions," *Food Analytical Methods*, vol. 9, no. 4, pp. 1060–1067, 2016.

- [11] M. Bhebhe, B. Chipurura, and M. Muchuweti, "Determination and comparison of phenolic compound content and antioxidant activity of selected local Zimbabwean herbal teas with exotic *Aspalathus linearis*," *South African Journal of Botany*, vol. 100, no. 1, pp. 213–218, 2015.
- [12] B. Patiri and A. Borah, *Wild Edible Plants of Assam*, Geetakhri Printers, Guwahati, India, 2007.
- [13] H. Narzary and A. Basumatary, "Proximate and vitamin C analysis of wild edible plants consumed by Bodos of Assam, India," *Journal of Molecular Pathophysiology*, vol. 4, no. 4, pp. 128–133, 2015.
- [14] M. Gelfand, S. Mavi, R. B. Drummond, and B. Ndemera, *The Traditional Medical Practitioners in Zimbabwe: His Principles of Practice and Pharmacopoeia*, Mambo Press, Gweru, Zimbabwe, 1985.
- [15] D. Davids, *Materia medica and care: a study of the uses of medicinal herbs and remedies as a form of treatment and negotiating social relationships in cape town and surroundings [M.S. dissertation]*, University of the Western Cape, Cape Town, South Africa, 2012.
- [16] A. Maroyi and G. K. E. Mosina, "Medicinal plants and traditional practices in peri-urban domestic gardens of the Limpopo province, South Africa," *Indian Journal of Traditional Knowledge*, vol. 13, no. 4, pp. 665–672, 2014.
- [17] A. Samie, C. L. Obi, P. O. Bessong, and L. Namrita, "Activity profiles of fourteen selected medicinal plants from Rural Venda communities in South Africa against fifteen clinical bacterial species," *African Journal of Biotechnology*, vol. 4, no. 12, pp. 1443–1451, 2005.
- [18] T. York, *An ethnopharmacological study of plants used for treating respiratory infections in rural Maputaland [M.S. dissertation]*, University of Zululand, KwaDlangezwa, South Africa, 2012.
- [19] D. Davids, T. Blouws, O. Aboyade et al., "Traditional health practitioners' perceptions, herbal treatment and management of HIV and related opportunistic infections," *Journal of Ethnobiology and Ethnomedicine*, vol. 10, article 77, 2014.
- [20] A. Hutchings, A. H. Scott, G. Lewis, and A. Cunningham, *Zulu Medicinal Plants: An Inventory*, University of Natal Press, Pietermaritzburg, South Africa, 1996.
- [21] R. M. Coopoosamy and K. K. Naidoo, "An ethnobotanical study of medicinal plants used by traditional healers in Durban, South Africa," *African Journal of Pharmacy and Pharmacology*, vol. 6, no. 11, pp. 818–823, 2012.
- [22] I. Hedberg and F. Staugard, *Traditional Medicinal Plants in Botswana*, Ipeleng, Gaborone, Botswana, 1989.
- [23] H. Shahriar, A. S. M. H. K. Chowdhury, S. Rahman et al., "Scientific validation of medicinal plants used by a folk medicinal practitioner of Chuadanga District, Bangladesh," *World Journal of Pharmacy and Pharmaceutical Sciences*, vol. 3, no. 11, pp. 13–24, 2014.
- [24] S. S. Semanya, M. J. Potgieter, and M. P. Tshisikhawe, "Use, conservation and present availability status of ethnomedicinal plants of Matebele-Village in the Limpopo Province, South Africa," *African Journal of Biotechnology*, vol. 12, no. 18, pp. 2392–2405, 2013.
- [25] M. Roberts, *Indigenous Healing Plants*, Southern Book Publishers, Halfway House, Johannesburg, South Africa, 1990.
- [26] T. Birhanu, D. Abera, and E. Ejeta, "Ethnobotanical study of medicinal plants in selected Horro Gudurru Woredas, western Ethiopia," *Journal of Biology, Agriculture and Healthcare*, vol. 5, no. 1, pp. 83–93, 2015.
- [27] N. T. Mahlangeni, R. Moodley, and S. B. Jonnalagadda, "Elemental composition of *Cyrtanthus obliquus* and *Lippia javanica* used in South African herbal tonic, Imbiza," *Arabian Journal of Chemistry*, 2013.
- [28] W. Musila, D. Kisangau, and J. Muema, "Conservation status and use of medicinal plants by traditional medical practitioners in Machakos District, Kenya," in *Indigenous Knowledge Conference Proceedings, Pennsylvania State University, May 27–29 2004*, 2004.
- [29] G. N. Njoroge and R. W. Bussmann, "Traditional management of ear, nose and throat (ENT) diseases in Central Kenya," *Journal of Ethnobiology and Ethnomedicine*, vol. 2, article 54, 2006.
- [30] P. Chigora, R. Masocha, and F. Mutenheri, "The role of indigenous medicinal knowledge (IMK) in the treatment of ailments in rural Zimbabwe: the case of Mutirikwi communal lands," *Journal of Sustainable Development in Africa*, vol. 9, no. 2, pp. 26–43, 2007.
- [31] R. W. Bussmann, G. G. Gilbreath, J. Solio et al., "Plant use of the Maasai of Sekenani Valley, Maasai Mara, Kenya," *Journal of Ethnobiology and Ethnomedicine*, vol. 2, article 22, 2006.
- [32] D. Motlhanka and G. P. Nthoiwa, "Ethnobotanical Survey of medicinal plants of Tswapong North, in Eastern Botswana: a case of plants from Mosweu and Seolwane Villages," *European Journal of Medicinal Plants*, vol. 3, no. 1, pp. 10–24, 2013.
- [33] A. Vhurumuku, "Knowledge, use and attitudes towards medicinal plants of pre-service teachers at a South African University," *Global Advanced Research Journal in Environmental Science and Toxicology*, vol. 4, no. 2, pp. 15–24, 2015.
- [34] M. N. Nkwanyana, *An ethnobotanical and anti-diarrhoeal investigation of plants used traditionally in the maputaland area homesteads [M.S. dissertation]*, University of Zululand, KwaDlangezwa, South Africa, 2013.
- [35] T. E. Moeng, *An investigation into the trade of medicinal plants by muthi shops and street vendors in the Limpopo Province, South Africa [M.S. thesis]*, University of Limpopo, Gauteng, South Africa, 2010.
- [36] S. Bandeira, A. M. Manjate, and O. Filipe, "An Ecological Assessment of the Health of the Chibuto-Missavene Wetland in the Dry Season, Mozambique: Emphasis on Resources Assessment, Utilization and Sustainability Analysis," report to Challenge Program for Water and Food, International Water Management Institute, Pretoria, South Africa, 2006, <https://cgspace.cgiar.org/bitstream/handle/10568/21604/21604.pdf?sequence=1>.
- [37] M. C. Palgrave, R. B. Drummond, and E. J. Moll, *Trees of Southern Africa*, Struik, Cape Town, South Africa, 2003.
- [38] M. O. Nanyingi, J. M. Mbaria, A. L. Lanyasunya et al., "Ethnopharmacological survey of Samburu district, Kenya," *Journal of Ethnobiology and Ethnomedicine*, vol. 4, article 14, 2008.
- [39] J. G. J. P. Sankar and K. B. Satapathy, "Weed diversity of Rabi crops and their ethnomedicinal uses in Kendrapara district of Odisha, India," *International Research Journal of Biological Sciences*, vol. 4, no. 3, pp. 33–38, 2015.
- [40] D. E. N. Mabogo, *The ethnobotany of the vhavenda [M.S. dissertation]*, University of Pretoria, Pretoria, South Africa, 1990.
- [41] B.-E. van Wyk, B. van Oudtshoorn, and N. Gericke, *Medicinal Plants of South Africa*, Briza, Pretoria, South Africa, 2013.
- [42] S. S. Semanya and A. Maroyi, "Medicinal plants used for the treatment of tuberculosis by Bapedi traditional healers

- in three districts of the Limpopo Province, South Africa," *African Journal of Traditional, Complementary, and Alternative Medicine*, vol. 10, no. 2, pp. 316–323, 2013.
- [43] L. Bunalema, S. Obakiro, J. R. S. Tabuti, and P. Waako, "Knowledge on plants used traditionally in the treatment of tuberculosis in Uganda," *Journal of Ethnopharmacology*, vol. 151, no. 2, pp. 999–1004, 2014.
- [44] P. Bruschi, M. Morganti, M. Mancini, and M. A. Signorini, "Traditional healers and laypeople: a qualitative and quantitative approach to local knowledge on medicinal plants in Muda (Mozambique)," *Journal of Ethnopharmacology*, vol. 138, no. 2, pp. 543–563, 2011.
- [45] A. T. Bryant, *Zulu Medicine and Medicine-Men*, C. Struik, Cape Town, South Africa, 1966.
- [46] N. S. Mkarafuu, *The role of non wood forest products in improving livelihoods of communities surrounding Jozani and Chwaka Bay National Park, Zanzibar [M.S. thesis]*, Sokoine University of Agriculture, Morogoro, Tanzania, 2010.
- [47] S. Kumar and D. Dash, "Flora of Nandan Kanan sanctuary: medicinal plants with their role in health care," *International Journal of Pharmacy and Life Sciences*, vol. 3, no. 4, pp. 1631–1642, 2012.
- [48] N. Lukwa, N. Z. Nyazema, C. F. Curtis, G. L. Mwaiko, and S. K. Chandiwana, "People's perceptions about malaria transmission and control using mosquito repellent plants in a locality in Zimbabwe," *Central African Journal of Medicine*, vol. 45, no. 3, pp. 64–68, 1999.
- [49] E. J. Mavundza, R. Maharaj, J. F. Finnie, G. Kabera, and J. van Staden, "An ethnobotanical survey of mosquito repellent plants in uMkhanyakude district, KwaZulu-Natal province, South Africa," *Journal of Ethnopharmacology*, vol. 137, no. 3, pp. 1516–1520, 2011.
- [50] J. Mahachi, *Medicinal properties of some plants used for the treatment of skin disorders in the OR Tambo and Amathole municipalities of the Eastern Cape Province, South Africa [M.S. dissertation]*, Walter Sisulu University, Mthatha, South Africa, 2013.
- [51] M. M. Matlebyane, J. W. W. Ng'ambi, and E. M. Aregheore, "Indigenous knowledge (IK) ranking of available browse and grass species and some shrubs used in medicinal and ethno-veterinary practices in ruminant livestock production in Limpopo province, South Africa," *Livestock Research for Rural Development*, vol. 22, no. 3, p. 54, 2010.
- [52] S. O. Bandeira, F. Gaspar, and F. P. Pagula, "African ethnobotany and healthcare: Emphasis on Mozambique," *Pharmaceutical Biology*, vol. 39, pp. 70–73, 2001.
- [53] C. A. Liengme, "Plants used by the Tsonga people of Gazankulu," *Bothalia*, vol. 13, no. 3–4, pp. 501–518, 1981.
- [54] A. J. Afolayan, D. S. Grierson, and W. O. Mbeng, "Ethnobotanical survey of medicinal plants used in the management of skin disorders among the Xhosa communities of the Amathole District, Eastern Cape, South Africa," *Journal of Ethnopharmacology*, vol. 153, no. 1, pp. 220–232, 2014.
- [55] O. O. G. Amusan, N. A. Sukati, and M. S. Shongwe, "Some phytomedicines from Shiselweni region of Swaziland," *Journal of Natural Remedies*, vol. 5, no. 1, pp. 19–25, 2005.
- [56] H. de Wet, S. Nciki, and S. F. van Vuuren, "Medicinal plants used for the treatment of various skin disorders by a rural community in northern Maputaland, South Africa," *Journal of Ethnobiology and Ethnomedicine*, vol. 9, article 51, 2013.
- [57] O. Amuka, P. O. Okemo, A. K. Machocho, and P. K. Mbugua, "Ethnobotanical survey of selected medicinal plants used by Ogiek communities in Kenya against microbial infections," *Ethnobotany Research and Applications*, vol. 12, pp. 627–641, 2014.
- [58] A. Hutchings, "Ritual cleansing, incense and the tree of life: observations on some indigenous plant usage in traditional Zulu and Xhosa purification and burial rites," *Alternation*, vol. 14, no. 2, pp. 189–218, 2007.
- [59] E. T. Nyahangare, B. M. Mvumi, and T. Mutibvu, "Ethnoveterinary plants and practices used for ecto-parasite control in semi-arid smallholder farming areas of Zimbabwe," *Journal of Ethnobiology and Ethnomedicine*, vol. 11, article 30, 2015.
- [60] E. T. Nyahangare, B. M. Mvumi, and P. C. Stevenson, "Tick control measures from nature," *Afgriland*, vol. 56, no. 3, pp. 76–77, 2012.
- [61] V. L. Narayana and G. M. Narasimharao, "Plants used in ethnoveterinary medicine by tribals of Visakhapatnam and Vizianagaram districts, Andhra Pradesh, India," *International Journal of Pure and Applied Bioscience*, vol. 3, no. 2, pp. 432–439, 2015.
- [62] C. A. Onyango, L. W. Gakuya, F. M. Mathooko et al., "Preservative effect of various indigenous plants on fermented milk from Maasai community of Kajiado County," *Journal of Applied Biosciences*, vol. 73, pp. 5935–5941, 2014.
- [63] I. T. Rampedi, *Indigenous plants in the Limpopo Province: potential for their commercial beverage production [Ph.D. thesis]*, University of South Africa, Pretoria, South Africa, 2010.
- [64] N. Lukwa, "Do traditional mosquito repellent plants work as mosquito larvicides?" *Central African Journal of Medicine*, vol. 40, no. 11, pp. 306–309, 1994.
- [65] United Nations Programme on HIV/AIDS, "AIDS epidemic update: Special Report on HIV/AIDS," 2006, http://data.unaids.org/pub/EpiReport/2006/2006.EpiUpdate_en.pdf.
- [66] A. R. Ndhkala, G. I. Stafford, J. F. Finnie, and J. van Staden, "In vitro pharmacological effects of manufactured herbal concoctions used in KwaZulu-Natal, South Africa," *Journal of Ethnopharmacology*, vol. 122, no. 1, pp. 117–122, 2009.
- [67] O. O. G. Amusan, P. S. Dlamini, J. D. Msonthi, and L. P. Makhubu, "Some herbal remedies from Manzini region of Swaziland," *Journal of Ethnopharmacology*, vol. 79, no. 1, pp. 109–112, 2002.
- [68] J. Williamson, *Useful Plants of Malawi*, University of Malawi, Limbe, Cameroon, 1974.
- [69] C. K. Kaingu, J. A. Oduma, and T. I. Kanui, "Practices of traditional birth attendants in Machakos District, Kenya," *Journal of Ethnopharmacology*, vol. 137, no. 1, pp. 495–502, 2011.
- [70] C. M. Doke and B. W. Vilakazi, *Zulu-English Dictionary*, Witwatersrand University Press, Johannesburg, South Africa, 1972.
- [71] O. O. Amusan, "Herbal medicine in Swaziland: an overview," in *African Natural Plant Products: New Discoveries and Challenges in Chemistry and Quality*, vol. 1021 of ACS Symposium Series, pp. 31–49, American Chemical Society, Washington, DC, USA, 2010.
- [72] O. O. G. Amusan, "Ethical and environmental issues in bioprospecting for drugs through traditional medicine: the case of Swaziland," *The African Journal of Plant Sciences and Biotechnology*, vol. 2, no. 1, pp. 1–9, 2008.
- [73] A. Samie, A. Housein, N. Lall, and J. J. M. Meyer, "Crude extracts of, and purified compounds from, *Pterocarpus angolensis*, and the essential oil of *Lippia javanica*: their in-vitro cytotoxicities

- and activities against selected bacteria and *Entamoeba histolytica*,” *Annals of Tropical Medicine and Parasitology*, vol. 103, no. 5, pp. 427–439, 2009.
- [74] I. E. Cock and S. F. van Vuuren, “Anti-proteus activity of some South African medicinal plants: their potential for the prevention of rheumatoid arthritis,” *Inflammopharmacology*, vol. 22, no. 1, pp. 23–36, 2014.
- [75] M. A. Lekganyane, T. M. Matsebatlela, R. L. Howard, L. J. Shai, and P. Masoko, “The phytochemical, antibacterial and antioxidant activity of five medicinal plants against the wound infecting bacteria,” *African Journal of Biotechnology*, vol. 11, no. 68, pp. 13210–13219, 2012.
- [76] K. M. Thembo, H. F. Vismer, N. Z. Nyazema, W. C. A. Gelderblom, and D. R. Katerere, “Antifungal activity of four weedy plant extracts against selected mycotoxigenic fungi,” *Journal of Applied Microbiology*, vol. 109, no. 4, pp. 1479–1486, 2010.
- [77] P. Masoko and K. M. Nxumalo, “Validation of antimycobacterial plants used by traditional healers in three districts of the Limpopo Province (South Africa),” *Evidence-based Complementary and Alternative Medicine*, vol. 2013, Article ID 586247, 7 pages, 2013.
- [78] D. R. Katerere, G. Graziani, K. M. Thembo, N. Z. Nyazema, and A. Ritieni, “Antioxidant activity of some African medicinal and dietary leafy African vegetables,” *African Journal of Biotechnology*, vol. 11, no. 17, pp. 4103–4108, 2012.
- [79] T. A. Ayuko, R. N. Njau, W. Cornelius, N. Leah, and I. O. Ndiege, “*In vitro* antiplasmodial activity and toxicity assessment of plant extracts used in traditional malaria therapy in the Lake Victoria Region,” *Memórias do Instituto Oswaldo Cruz*, vol. 104, no. 5, pp. 689–694, 2009.
- [80] C. Clarkson, V. J. Maharaj, N. R. Crouch et al., “*In vitro* antiplasmodial activity of medicinal plants native to or naturalised in South Africa,” *Journal of Ethnopharmacology*, vol. 92, no. 2–3, pp. 177–191, 2004.
- [81] D. K. Olivier, E. A. Shikanga, S. Combrinck, R. W. M. Krause, T. Regnier, and T. P. Dlamini, “Phenylethanoid glycosides from *Lippia javanica*,” *South African Journal of Botany*, vol. 76, no. 1, pp. 58–63, 2010.
- [82] H. Rimpler and H. Sauerbier, “Iridoid glucosides as taxonomic markers in the genera *Lantana*, *Lippia*, *Aloysia* and *Phyla*,” *Biochemical Systematics and Ecology*, vol. 14, no. 3, pp. 307–310, 1986.
- [83] S. F. Mujovo, A. A. Hussein, J. J. M. Meyer, B. Fourie, T. Muthivhi, and N. Lall, “Bioactive compounds from *Lippia javanica* and *Hoslundia opposita*,” *Natural Product Research*, vol. 22, no. 12, pp. 1047–1054, 2008.
- [84] M. T. Ludere, T. van Ree, and R. Vleggaar, “Isolation and relative stereochemistry of lippialactone, a new antimalarial compound from *Lippia javanica*,” *Fitoterapia*, vol. 86, no. 3, pp. 188–192, 2013.
- [85] R. Neidlein and R. Staehle, “Constituents of *Lippia javanica*. III,” *Deutsche Apotheker Zeitung*, vol. 40, pp. 1588–1592, 1974.
- [86] T. P. Dlamini, *Isolation and characterization of bioactive compounds from Lippia javanica [M.S. thesis]*, University of Johannesburg, Johannesburg, South Africa, 2006.
- [87] R. Sõukand and R. Kalle, “Where does the border lie: locally grown plants used for making tea for recreation and/or healing, 1970s–1990s Estonia,” *Journal of Ethnopharmacology*, vol. 150, no. 1, pp. 162–174, 2013.
- [88] Y. Frum and A. M. Viljoen, “*In vitro* 5-lipoxygenase and antioxidant activities of South African medicinal plants commonly used topically for skin diseases,” *Skin Pharmacology and Physiology*, vol. 19, no. 6, pp. 329–335, 2006.
- [89] S. F. van Vuuren, *The antimicrobial activity and essential oil composition of medicinal aromatic plants used in african traditional healing [Ph.D. thesis]*, University of the Witwatersrand, Johannesburg, South Africa, 2007.
- [90] S. Suliman, *Antimicrobial interactions of Artemisia afra used in african traditional medicine [M.S. dissertation]*, University of the Witwatersrand, Johannesburg, South Africa, 2011.
- [91] S. Virijevec, *Antimicrobial and chemical properties of essential oils from indigenous South African lippia species [M.S. dissertation]*, Tshwane University of Technology, Tshwane, South Africa, 2012.
- [92] L. S. Chagonda, C. D. Makanda, and J.-C. Chalchat, “Essential oils of wild and cultivated *Lippia javanica* (Spreng) and *L. oatesii* (Rolfe) from Zimbabwe,” *Journal of Essential Oil Research*, vol. 12, no. 1, pp. 1–6, 2000.
- [93] O. Ngassapa, D. K. B. Runyoro, E. Harvala, and I. B. Chinou, “Composition and antimicrobial activity of essential oils of two populations of Tanzanian *Lippia javanica* (Burm. f.) Spreng. (Verbenaceae),” *Flavour and Fragrance Journal*, vol. 18, no. 3, pp. 221–224, 2003.
- [94] O. Amuka, A. K. Machochi, P. O. Okemo, and P. K. Mbugua, “Antibacterial and antifungal activities of essential oils from *Satureia biflora* D. Don, Benth, Speng (Chepsagitiet), *Lippia javanica* Burm. f. (Labotuet) and *Toddalia asiatica* (L) Lam. Rutaceae (Chepindoruet),” *Global Journal of Science Frontier Research: B Chemistry*, vol. 14, no. 4, pp. 45–51, 2014.
- [95] S. R. Magano, F. Nchu, and J. N. Eloff, “*In vitro* investigation of the repellent effects of the essential oil of *Lippia javanica* on adults of *Hyalomma marginatum rufipes*,” *African Journal of Biotechnology*, vol. 10, no. 44, pp. 8970–8975, 2011.
- [96] A. Hutchings and J. van Staden, “Plants used for stress-related ailments in traditional Zulu, Xhosa and Sotho medicine. Part 1: plants used for headaches,” *Journal of Ethnopharmacology*, vol. 43, no. 2, pp. 89–124, 1994.
- [97] H. E. Marx, N. O’Leary, Y.-W. Yuan et al., “A molecular phylogeny and classification of Verbenaceae,” *American Journal of Botany*, vol. 97, no. 10, pp. 1647–1663, 2010.
- [98] A. A. Munir, “A taxonomic revision of the genus *Lippia* (Hout. ex) Linn. (Verbenaceae) in Australia,” *Journal of the Adelaide Botanic Garden*, vol. 15, no. 2, pp. 129–145, 1993.
- [99] R. W. Sanders, “The genera of Verbenaceae in the southeastern United States,” *Harvard Papers in Botany*, vol. 5, no. 2, pp. 303–358, 2001.
- [100] J. M. S. de Campos, S. M. Sousa, P. S. Silva, L. C. Pinheiro, F. Sampaio, and L. F. Viccini, “Chromosome numbers and DNA C values in the genus *Lippia* (Verbenaceae),” *Plant Systematics and Evolution*, vol. 291, no. 1, pp. 133–140, 2011.
- [101] B. Verdcourt, “Verbenaceae,” in *Flora of Tropical Africa*, R. M. Polhill, Ed., p. 155, A. A. Balkema, Rotterdam, The Netherlands, 1992.
- [102] M. Hyde, B. Wursten, P. Ballings, and M. C. Palgrave, “Flora of Zimbabwe: species information: *Lippia javanica* (Burm. f.) Spreng,” http://www.zimbabweflora.co.zw/speciesdata/species.php?species_id=148720.
- [103] IPNI (International Plant Name Index), 2016, <http://www.ipni.org>.
- [104] H. H. W. Pearson, “Verbenaceae,” *Flora Capensis*, vol. 5, no. 3, pp. 192–197, 1912.

- [105] B.-E. van Wyk and N. Gericke, *People's Plants: A Guide to Useful Plants of Southern Africa*, Briza, Pretoria, South Africa, 2000.
- [106] R. Fernandes, "Verbenaceae," in *Flora Zambesiaca*, G. V. Pope and E. S. Martins, Eds., vol. 8, part 7, pp. 6–61, Royal Botanic Gardens, London, UK, 2005.
- [107] B.-E. Van Wyk, "The potential of South African plants in the development of new food and beverage products," *South African Journal of Botany*, vol. 77, no. 4, pp. 857–868, 2011.
- [108] H. R. Juliani, Y. Koelliker, M. Bucuk et al., "Quality and consumer studies in the USA of African herbal teas for the natural product industry development in sub-Sahara Africa," in *Proceedings of the African Natural Plant Products: New Discoveries and Challenges in Chemistry and Quality Symposium*, H. R. Juliani, J. E. Simon, and C.-T. Ito, Eds., vol. 1021 of *ACS Symposium Series*, pp. 403–420, American Chemical Society Publications, Washington, DC, USA, 2009.
- [109] C. Pretorius, *Antioxidant properties of Lippia javanica (Burm. f.) Spreng [M.S. thesis]*, North West University, Potchefstroom, South Africa, 2010.
- [110] M. Whiteside, *Encouraging Sustainable Family Sector Agriculture in Botswana*, Cooperation for Research, Development and Education (CORDE), Gaborone, Botswana, 1997.
- [111] B.-E. Van Wyk, "A broad review of commercially important southern African medicinal plants," *Journal of Ethnopharmacology*, vol. 119, no. 3, pp. 342–355, 2008.
- [112] H. Wild and M. Gelfand, "Some native herbal remedies at present in use in Mashonaland," *The Central African journal of medicine*, vol. 5, no. 6, pp. 292–305, 1959.
- [113] G. N. Njoroge and J. W. Kibunga, "Herbal medicine acceptance, sources and utilization for diarrhoea management in a cosmopolitan urban area (Thika, Kenya)," *African Journal of Ecology*, vol. 45, no. 1, pp. 65–70, 2007.
- [114] A. Ribeiro, M. M. Romeiras, J. Tavares, and M. T. Faria, "Ethnobotanical survey in Canhane village, district of Massingir, Mozambique: medicinal plants and traditional knowledge," *Journal of Ethnobiology and Ethnomedicine*, vol. 6, article 33, 2010.
- [115] S. S. Semenya and A. Maroyi, "Medicinal plants used by the Bapedi traditional healers to treat diarrhoea in the Limpopo Province, South Africa," *Journal of Ethnopharmacology*, vol. 144, no. 2, pp. 395–401, 2012.
- [116] A. Maroyi, "Traditional use of medicinal plants in south-central Zimbabwe: review and perspectives," *Journal of Ethnobiology and Ethnomedicine*, vol. 9, article 31, 2013.
- [117] V. J. Maharaj, G. Fouche, J. Senabe, R. Nthambeleni, and F. Kotze, "Agro-processing opportunities identified through a novel mosquito repellent from a medicinal plant," in *Proceedings of the 2nd CSIR Biennial Conference on Science Real and Relevant*, Pretoria, South Africa, November 2008, <http://hdl.handle.net>.
- [118] J. McCosh, Z. Nxele, B. Letty, E. de Beer, N. Majara, and K. Swanepoel, "Study to develop a strategy for the development of a viable essential oils industry in South Africa: review and situation analysis," INR Report 442/11, Institute of Natural Resources, Pietermaritzburg, South Africa, 2011.
- [119] CSIR (Council for Scientific and Industrial Research), CSIR patent, mosquito repellent, South African patent no. 95/9583, 1995.
- [120] J. Madzimure, E. T. Nyahangare, H. Hamudikuwanda et al., "Acaricidal efficacy against cattle ticks and acute oral toxicity of *Lippia javanica* (Burm F.) Spreng," *Tropical Animal Health and Production*, vol. 43, no. 2, pp. 481–489, 2011.
- [121] Q. Hou, W.-J. He, H.-J. Hao et al., "The four-herb chinese medicine ANBP enhances wound healing and inhibits scar formation via bidirectional regulation of transformation growth factor pathway," *PLoS ONE*, vol. 9, no. 12, Article ID e112274, 2014.
- [122] E.-M. M. Abubakar, "The use of *Psidium guajava* Linn. in treating wound, skin and soft tissue infections," *Scientific Research and Essays*, vol. 4, no. 6, pp. 605–611, 2009.
- [123] N. N. Mokoka, *Indigenous knowledge of fever tea (Lippia javanica) and effect of shade netting on plant growth, oil yield and compound composition [M.S. thesis]*, University of Pretoria, Pretoria, South Africa, 2005.
- [124] M. A. Ngwenya, A. Koopman, and R. Williams, *Zulu Botanical Knowledge: An Introduction*, National Botanical Institute, Pretoria, South Africa, 2003.
- [125] M. Muchuweti, L. Nyamukonda, L. S. Chagonda, A. R. Ndhkala, C. Mupure, and M. Benhura, "Total phenolic content and antioxidant activity in selected medicinal plants of Zimbabwe," *International Journal of Food Science and Technology*, vol. 41, no. 1, pp. 33–38, 2006.
- [126] E. Shikanga, T. Regnier, S. Combrinck, and B. Botha, "Polar *Lippia* extracts as alternatives for the postharvest control of Guazatine®-resistant strains of *Penicillium digitatum* in citrus," *Fruits*, vol. 64, no. 2, pp. 75–82, 2009.
- [127] M. Sandasi, G. P. P. Kamatou, S. Combrinck, and A. M. Viljoen, "A chemotaxonomic assessment of four indigenous South African *Lippia* species using GC-MS and vibrational spectroscopy of the essential oils," *Biochemical Systematics and Ecology*, vol. 51, no. 3, pp. 142–152, 2013.
- [128] S. Sedaghatoor, A. M. Torkashvand, D. Hashemabadi, and B. Kaviani, "Yield and quality response of tea plant to fertilizers," *African Journal of Agricultural Research*, vol. 4, no. 6, pp. 568–570, 2009.
- [129] K. O. Soetan, C. O. Olaiya, and O. E. Oyewole, "The importance of mineral elements for humans, domestic animals and plants: a review," *African Journal of Food Science*, vol. 4, no. 5, pp. 200–222, 2008.
- [130] R. Borneo, A. E. León, A. Aguirre, P. Ribotta, and J. J. Cantero, "Antioxidant capacity of medicinal plants from the Province of Córdoba (Argentina) and their in vitro testing in a model food system," *Food Chemistry*, vol. 112, no. 3, pp. 664–670, 2009.
- [131] W.-Y. Huang, Y.-Z. Cai, and Y. Zhang, "Natural phenolic compounds from medicinal herbs and dietary plants: potential use for cancer prevention," *Nutrition and Cancer*, vol. 62, no. 1, pp. 1–20, 2010.
- [132] A. Djlani and A. Dicko, "The therapeutic benefits of essential oils, nutrition, well-being and health," 2012, <http://www.intechopen.com/books/nutrition-well-being-and-health/the-therapeutic-benefits-of-essential-oils>.
- [133] S. Kumar and A. K. Pandey, "Chemistry and biological activities of flavonoids: an overview," *The Scientific World Journal*, vol. 2013, Article ID 162750, 16 pages, 2013.
- [134] T. P. T. Cushnie and A. J. Lamb, "Antimicrobial activity of flavonoids," *International Journal of Antimicrobial Agents*, vol. 26, no. 5, pp. 343–356, 2005.
- [135] A. Tapas, D. Sakarkar, and R. Kakde, "Flavonoids as nutraceuticals: a review," *Tropical Journal of Pharmaceutical Research*, vol. 7, no. 3, pp. 1089–1099, 2008.
- [136] Y. Nishizuka, "The molecular heterogeneity of protein kinase C and its implications for cellular regulation," *Nature*, vol. 334, no. 6184, pp. 661–665, 1988.

- [137] T. Hunter, "Protein kinases and phosphatases: the Yin and Yang of protein phosphorylation and signaling," *Cell*, vol. 80, no. 2, pp. 225–236, 1995.
- [138] J. W. Critchfield, S. T. Butera, and T. M. Folks, "Inhibition of HIV activation in latently infected cells by flavonoid compounds," *AIDS Research and Human Retroviruses*, vol. 12, no. 1, pp. 39–46, 1996.
- [139] T. Kamiya, "Biological functions and health benefits of amino acids," *Food Fermentation Journal Japan*, vol. 206, pp. 33–44, 2002.
- [140] G. Fouche, G. M. Cragg, P. Pillay, N. Kolesnikova, V. J. Maharaj, and J. Senabe, "In vitro anticancer screening of South African plants," *Journal of Ethnopharmacology*, vol. 119, no. 3, pp. 455–461, 2008.
- [141] W. M. Arika, Y. A. Abdirahman, M. M. Mawia et al., "Hypoglycemic effect of *Lippia javanica* in alloxan induced diabetic mice," *Journal of Diabetes Metabolism*, vol. 6, article 624, 2015.
- [142] J. Govere, D. N. Durrheim, N. du Toit, R. H. Hunt, and M. Coetzee, "Local plants as repellents against *Anopheles arabiensis*, in Mpumalanga Province, South Africa," *Central African Journal of Medicine*, vol. 46, no. 8, pp. 213–216, 2000.
- [143] A. Samie, T. Tambani, E. Harshfield, E. Green, J. N. Ramalivhana, and P. O. Bessong, "Antifungal activities of selected vanda medicinal plants against *candida albicans*, *Candida krusei* and *cryptococcus neoformans* isolated from South African AIDS patients," *African Journal of Biotechnology*, vol. 9, no. 20, pp. 2965–2976, 2010.
- [144] E. A. Prozesky, J. J. M. Meyer, and A. I. Louw, "In vitro antiplasmodial activity and cytotoxicity of ethnobotanically selected South African plants," *Journal of Ethnopharmacology*, vol. 76, no. 3, pp. 239–245, 2001.
- [145] M. O. Omolo, D. Okinyo, I. O. Ndiege, W. Lwande, and A. Hassanali, "Fumigant toxicity of the essential oils of some African plants against *Anopheles gambiae* sensu stricto," *Phytomedicine*, vol. 12, no. 3, pp. 241–246, 2005.
- [146] L. J. McGaw, A. K. Jäger, and J. van Staden, "Antibacterial, anthelmintic and anti-amoebic activity in South African medicinal plants," *Journal of Ethnopharmacology*, vol. 72, no. 1-2, pp. 247–263, 2000.
- [147] C. A. T. Katsvanga and S. Chigwiza, "Effectiveness of natural herbs, fever tea (*Lippia javanica*) and mexican marigold (*Tagetes minuta*) as substitutes to synthetic pesticides in controlling aphid species (*Brevicoryne brassica*) on cabbage (*Brassica capitata*)," *Tropical and Subtropical Agroecosystems*, vol. 4, no. 3, pp. 101–106, 2004.
- [148] P. W. Mashela, H. A. Shimelis, D. de Waele, M. N. Mokgalong, F. N. Mudau, and L. G. Ngobeni, "Fever tea (*Lippia javanica*) as a root-knot nematode suppressant in tomato production," *African Plant Protection*, vol. 16, no. 1, pp. 1–6, 2010.
- [149] L. Chikukura, B. M. Mvumi, R. Chikonzo, and C. Chenzara, "Evaluation of selected indigenous pesticidal plant powders against stored maize and cowpeas insect pests," in *Proceedings of the African Crop Science Conference*, vol. 10, pp. 191–194, 2011.
- [150] S. Muzemu, B. M. Mvumi, S. P. M. Nyirenda et al., "Pesticidal effects of indigenous plants extracts against rape aphids and tomato red spider mites," in *Proceedings of the African Crop Science Conference Proceedings*, vol. 10, pp. 169–171, 2012.
- [151] D. W. Nyamai, W. Arika, P. E. Ogola, E. N. M. Njagi, and M. P. Ngugi, "Medicinally important phytochemicals: an untapped research avenue," *Research and Reviews: Journal of Pharmacology and Phytochemistry*, vol. 4, no. 1, pp. 35–49, 2016.
- [152] C. S. Yang, J. M. Landau, M.-T. Huang, and H. L. Newmark, "Inhibition of carcinogenesis by dietary polyphenolic compounds," *Annual Review of Nutrition*, vol. 21, pp. 381–406, 2001.
- [153] A. Salminen, M. Lehtonen, T. Suuronen, K. Kaarniranta, and J. Huuskonen, "Terpenoids: natural inhibitors of NF- κ B signaling with anti-inflammatory and anticancer potential," *Cellular and Molecular Life Sciences*, vol. 65, no. 19, pp. 2979–2999, 2008.
- [154] E. J. Mavundza, R. Maharaj, J. C. Chukwujekwu, J. F. Finnie, and J. Van Staden, "Screening for adulticidal activity against *Anopheles arabiensis* in ten plants used as mosquito repellent in South Africa," *Malaria Journal*, vol. 13, article 173, 2014.
- [155] S. Chanda and R. Dave, "In vitro models for antioxidant activity evaluation and some medicinal plants possessing antioxidant properties: an overview," *African Journal of Microbiology Research*, vol. 3, no. 13, pp. 981–996, 2009.
- [156] K. Knobloch, A. Pauli, B. Iberl, H. Weigand, and N. Weis, "Antibacterial and antifungal properties of essential oil components," *Journal of Essential Oil Research*, vol. 1, no. 3, pp. 119–128, 1989.
- [157] J. B. Hinou, C. E. Harvala, and E. B. Hinou, "Antimicrobial activity screening of 32 common constituents of essential oils," *Pharmazie*, vol. 44, no. 4, pp. 302–303, 1989.
- [158] J. Kim, M. R. Marshall, and C.-I. Wei, "Antibacterial activity of some essential oil components against five foodborne pathogens," *Journal of Agricultural and Food Chemistry*, vol. 43, no. 11, pp. 2839–2845, 1995.
- [159] S. Pattnaik, V. R. Subramanyam, M. Bapaji, and C. R. Kole, "Antibacterial and antifungal activity of aromatic constituents of essential oils," *Microbios*, vol. 89, no. 358, pp. 39–46, 1997.
- [160] S. Gibbons, "Anti-staphylococcal plant natural products," *Natural Product Reports*, vol. 21, no. 2, pp. 263–277, 2004.
- [161] N. D. Martini, D. R. P. Katerere, and J. N. Eloff, "Biological activity of five antibacterial flavonoids from *Combretum erythrophyllum* (Combretaceae)," *Journal of Ethnopharmacology*, vol. 93, no. 2-3, pp. 207–212, 2004.
- [162] A. Basile, S. Giordano, J. A. López-Sáez, and R. C. Cobiánchi, "Antibacterial activity of pure flavonoids isolated from mosses," *Phytochemistry*, vol. 52, no. 8, pp. 1479–1482, 1999.
- [163] C. S. A. Lima, E. L. C. de Amorim, K. R. de Sena et al., "Antimicrobial activity of a mixture of two isomeric phenylpropanoid glycosides from *Arrabidaea harleyi* A. H. Gentry (Bignoniaceae)," *Brazilian Journal of Pharmaceutical Sciences*, vol. 39, no. 1, pp. 77–81, 2003.
- [164] K. Ono, H. Nakane, M. Fukushima, J. C. Chermann, and F. Barré-Sinoussi, "Differential inhibitory effects of various flavonoids on the activities of reverse transcriptase and cellular DNA and RNA polymerases," *European Journal of Biochemistry*, vol. 190, no. 3, pp. 469–476, 1990.
- [165] A. J. Vlietinck, T. de Bruyne, S. Apers, and L. A. Pieters, "Plant-derived leading compounds for chemotherapy of human immunodeficiency virus (HIV) infection," *Planta Medica*, vol. 64, no. 2, pp. 97–109, 1998.
- [166] M. Martinez-Velazquez, R. Rosario-Cruz, G. Castillo-Herrera, J. M. Flores-Fernandez, A. H. Alvarez, and E. Lugo-Cervantes, "Acaricidal effect of essential oils from *Lippia graveolens* (Lamiaceae: Verbenaceae), *Rosmarinus officinalis* (Lamiaceae: Lamiaceae), and *Allium sativum* (Liliales: Liliaceae) against *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae)," *Journal of Medical Entomology*, vol. 48, no. 4, pp. 822–827, 2011.
- [167] B.-E. van Wyk, F. van Heerden, and B. van Oudtshoorn, *Poisonous Plants of South Africa*, Briza Publications, Pretoria, South Africa, 2009.

- [168] T. Heikel, B. C. Knight, C. Rimington, H. D. Ritchie, and E. J. Williams, "Studies on biliary excretion in the rabbit. 1: the effect of icterogenin and rehmmanic acid on bile flow and the excretion of bilirubin, phylloerythrin, coproporphyrin, alkaline phosphatase and bromosulfalein," *Proceedings of the Royal Society of London: Series B, Biological Sciences*, vol. 153, no. 950, pp. 47-79, 1960.
- [169] D. A. Mpofu, U. Marume, V. Mlambo, and A. Hugo, "The effects of *Lippia javanica* dietary inclusion on growth performance, carcass characteristics and fatty acid profiles of broiler chickens," *Animal Nutrition*, vol. 2, no. 3, pp. 160-167, 2016.
- [170] V. L. Williams, K. Balkwill, and E. T. F. Witkowski, "A lexicon of plants traded in the Witwatersrand umuthi shops, South Africa," *Bothalia*, vol. 31, no. 1, pp. 71-98, 2001.
- [171] D. Raimondo, L. von Staden, W. Foden et al., *Red List of South African Plants*, Strelitzia 25, South African National Biodiversity Institute, Pretoria, South Africa, 2009.