



# Complete Genome and Plasmid Sequences for *Rhodococcus fascians* D188 and Draft Sequences for *Rhodococcus* Isolates PBTS 1 and PBTS 2

## Rio A. Stamler,<sup>a</sup> Danny Vereecke,<sup>b</sup> Yucheng Zhang,<sup>c</sup> Faye Schilkey,<sup>d</sup> Nico Devitt,<sup>d</sup> Jennifer J. Randall<sup>a</sup>

Department of Entomology, Plant Pathology and Weed Science, New Mexico State University, Las Cruces, New Mexico, USA<sup>a</sup>; Department of Applied Biosciences, Ghent University, Ghent, Belgium<sup>b</sup>; Department of Plant Pathology, University of Florida, Gainesville, Florida, USA<sup>c</sup>; National Center for Genome Resources, Santa Fe, New Mexico, USA<sup>d</sup>

*Rhodococcus fascians*, a phytopathogen that alters plant development, inflicts significant losses in plant production around the world. We report here the complete genome sequence of *R. fascians* D188, a well-characterized model isolate, and *Rhodococcus* species PBTS (pistachio bushy top syndrome) 1 and 2, which were shown to be responsible for a disease outbreak in pistachios.

Received 19 April 2016 Accepted 28 April 2016 Published 9 June 2016

Citation Stamler RA, Vereecke D, Zhang Y, Schilkey F, Devitt N, Randall JJ. 2016. Complete genome and plasmid sequences for *Rhodococcus fascians* D188 and draft sequences for *Rhodococcus* isolates PBTS 1 and PBTS 2. Genome Announc 4(3):e00495-16. doi:10.1128/genomeA.00495-16.

Copyright © 2016 Stamler et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Jennifer J. Randall, jrandall@nmsu.edu.

hodococcus fascians, the sole characterized plant pathogen in the genus *Rhodococcus*, is a Gram-positive pleomorphic bacterium that causes disease through the production and modulation of plant growth regulators, such as cytokinins and auxins (1). R. fascians infects a large variety of plants, including monocots and dicots, often causing cryptic symptomology (2). R. fascians D188, which has intensively been studied for decades, has revealed a wealth of knowledge about this plant pathogen, including chromosomal and plasmid localized virulence genes necessary for symptomatic host colonization and cytokinin production (3, 4). The virulence plasmid pFiD188 was previously sequenced using Sanger technology, and fragmented draft genome sequences for multiple R. fascians isolates, including D188, are available (5, 6). Pistachio bushy top syndrome (PBTS) describes a suite of symptoms affecting clonally propagated UCB-1, an interspecific hybrid rootstock planted in California, Arizona, and New Mexico (7, 8). Two distinct Rhodococcus species isolates (PBTS 1 and PBTS 2) were isolated from diseased trees and shown to cause stunting and abnormal root and shoot development in pathogenicity assays. Given the global scale of nursery plant production and the diverse population structure of R. fascians, as indicated by initial genome sequences (6), high-quality complete reference sequences are needed to fully characterize newly discovered isolates. The reference R. fascians strain D188 (from the private collection of D. Vereecke) and PBTS isolates (7) were stored in glycerol stocks at -80°C. Cultures were grown in 50 ml of LB broth, and DNA was extracted using a Wizard genomic DNA purification kit, according to the manufacturer's instructions (Promega, Madison, WI). DNA was shipped on dry ice to the National Center for Genomic Resources (NCGR, Santa Fe, NM) for single-molecule real-time (SMRT) sequencing. Libraries with a 10-kb insert were prepared, and each isolate was sequenced using one SMRT cell on the PacBio RS II instrument (Pacific Biosciences, Menlo Park, CA). Hierarchical Genome Assembly Process (HGAP) was used for de novo assembly (9) (RS\_HGAP assembly.2; Pacific Biosciences). For

strain D188, an additional round of polishing was performed using only reads with a quality of  $\geq$ 84. The genomes were annotated with Prokka version 1.12-beta (10), protein-coding features were predicted using Prodigal version 2.6 (11), tRNA was predicted by ARAGORN version 1.2 (12), and rRNA was predicted by RNAmmer version 1.2 (13). The genome annotations reveal a wealth of predicted secondary metabolism-related genes, including novel antibiotic synthesis, heavy metal resistance, siderophore production, pilus-like assembly mechanisms, secreted proteins, transposons and other mobile elements, and prophage sequences.

**Nucleotide sequence accession numbers.** The complete genome and plasmid sequences for this whole-genome shotgun project have been deposited at DDBJ/EMBL/GenBank under the following accession numbers: CP015235, CP015236, and CP015237 for D188, CP015219 for PBTS 1, and CP015220 and CP015221 for PBTS 2. The versions described in this paper are CP015235.1, CP015236.1, CP015237.1, CP015219.1, CP015220.1, and CP015221.1, respectively.

## ACKNOWLEDGMENTS

We thank Jordan Martin for computer assistance.

This work was funded by the California Pistachio Research Board and the New Mexico State University Agricultural Experimental Station. The funders had no role in the study design, data collection and interpretation, or the decision to submit the work for publication.

#### FUNDING INFORMATION

This work, including the efforts of Jennifer J. Randall, was funded by California Pistachio Research Board. This work, including the efforts of Jennifer J. Randall, was funded by New Mexico State University Agricultural Experimental Station.

### REFERENCES

1. Stes E, Vandeputte OM, El Jaziri M, Holsters M, Vereecke D. 2011. A successful bacterial coup d'état: how *Rhodococcus fascians* redirects plant

development. Annu Rev Phytopathol **49:**69-86. http://dx.doi.org/ 10.1146/annurev-phyto-072910-095217.

- Putnam ML, Miller ML. 2007. Rhodococcus fascians in herbaceous perennials. Plant Dis 91:1064–1076. http://dx.doi.org/10.1094/PDIS-91-9-1064.
- Vereecke D, Cornelis K, Temmerman W, Jaziri M, Van Montagu M, Holsters M, Goethals K. 2002. Chromosomal locus that affects pathogenicity of *Rhodococcus fascians*. J Bacteriol 184:1112–1120. http:// dx.doi.org/10.1128/jb.184.4.1112-1120.2002.
- 4. Crespi M, Vereecke D, Temmerman W, Van Montagu M, Desomer J. 1994. The *fas* operon of *Rhodococcus fascians* encodes new genes required for efficient fasciation of host plants. J Bacteriol 176:2492–2501.
- Francis I, De Keyser A, De Backer P, Simón-Mateo C, Kalkus J, Pertry I, Ardiles-Diaz W, De Rycke R, Vandeputte OM, El Jaziri M, Holsters M, Vereecke D. 2012. pFiD188, the linear virulence plasmid of *Rhodococcus fascians* D188. Mol Plant Microbe Interact 25:637–647. http:// dx.doi.org/10.1094/MPMI-08-11-0215.
- Creason AL, Vandeputte OM, Savory EA, Davis II, EW, Putnam ML, Hu E, Swader-Hines D, Mol A, Baucher M, Prinsen E, Zdanowska M, Givan SA, El Jaziri M, Loper JE, Mahmud T, Chang JH. 2014. Analysis of genome sequences from plant pathogenic *Rhodococcus* reveals genetic novelties in virulence loci. PLoS One 9:e101996. http://dx.doi.org/ 10.1371/journal.pone.0101996.
- Stamler RA, Kilcrease J, Fichtner E, Kallsen C, Cooke P, Heerema RJ, Randall JJ. 2015. First report of *Rhodococcus* isolates causing pistachio bushy

top syndrome on 'UCB-1' rootstock in California and Arizona. Plant Dis **99:**1468–1476. http://dx.doi.org/10.1094/PDIS-12-14-1340-RE.

- Stamler RA, Heerema R, Randall JJ. 2015. First report of phytopathogenic *Rhodococcus* isolates on pistachio bushy top syndrome 'UCB-1' rootstock in New Mexico. Plant Dis 99:1854. http://dx.doi.org/10.1094/ PDIS-04-15-0471-PDN.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. http://dx.doi.org/ 10.1038/nmeth.2474.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. http://dx.doi.org/10.1093/bioinformatics/btu153.
- Hyatt D, Chen GL, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. http://dx.doi.org/10.1186/ 1471-2105-11-119.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. http://dx.doi.org/10.1093/nar/gkh152.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt H-H, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res 35:3100–3108. http://dx.doi.org/10.1093/ nar/gkm160.