Effects of a sodium glucose co-transporter 2 selective inhibitor, ipragliflozin, on the diurnal profile of plasma glucose in patients with type 2 diabetes: A study using continuous glucose monitoring

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Keywords

3-Hydroxybutyrate, Continuous glucose monitoring, Sodium glucose co-transporter inhibitor

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ABSTRACT

Aims/Introduction: To assess the effects of sodium glucose co-transporter 2 inhibitor therapy on the pathophysiology of type 2 diabetes.

Materials and Methods: We administered ipragliflozin to 21 inpatients with type 2 diabetes for 7 days, and analyzed the diurnal profiles of plasma glucose and 3-hydroxybu-tyrate. A total of 21 age-, sex- and body mass index-matched diabetic patients served as controls.

Results: Continuous glucose monitoring showed that the 24-h glucose curve was shifted downward without hypoglycemia by the administration of ipragliflozin. The average glucose level was reduced from 182 \pm 54 mg/dL to 141 \pm 33 mg/dL (P < 0.0001). The magnitude of the reduction was highly correlated with the baseline average glucose level. Homeostasis model assessment of insulin resistance was decreased, and homeostasis model assessment of β -cell function was increased during the treatment. Urinary glucose excretion was correlated with the average glucose level both on day 0 and on day 7, although the regression line was steeper and shifted leftward on day 7. The ipragliflozintreated patients lost more weight than the control patients (1.4 \pm 0.5 vs 0.5 \pm 0.6 kg, P < 0.0001). Plasma levels of 3-hydroxybutyrate were significantly increased with peaks before breakfast and before dinner. Patient age and bodyweight loss were negatively and positively correlated with the peak levels of 3-hydroxybutyrate on day 7, respectively. Conclusions: The ipragliflozin treatment improved the 24-h glucose curve without causing hypoglycemia. The close correlation between the magnitude of glucose reduction and the baseline plasma glucose concentration suggests that the risk of hypoglycemia is likely low. It might be prudent to monitor ketone body levels in younger patients and in patients with rapid weight loss.

INTRODUCTION

Recently, the role of glucose reabsorption by the renal proximal tubule has emerged as an essential part of glucose homeostasis.

Kentaro Yamada and Hitomi Nakayama contributed equally to this work. Received 25 December 2014; revised 10 March 2015; accepted 22 March 2015 Under normal physiological conditions, approximately 160– 180 g of glucose is filtered by the kidneys in a day, and virtually all glucose filtered is reabsorbed by the renal proximal tubule and returned to the circulation^{1,2}. In a hyperglycemic state, the ability of the proximal tubule to reabsorb glucose increases as the filtered glucose load increases until the

© 2015 The Authors. Journal of Diabetes Investigation published by Asian Association of the Study of Diabetes (AASD) and Wiley Publishing Asia Pty Ltd This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes. maximum glucose transport capacity is reached. When the reabsorption capacity of the proximal tubule is surpassed, glucosuria becomes apparent. The renal threshold of plasma glucose for urinary glucose excretion is 180–200 mg/dL in healthy individuals^{1,3}. Glucose reabsorption from the glomerular filtrate into renal tubular epithelial cells is mediated by sodium glucose co-transporter (SGLT) proteins. Approximately 90% of the filtered glucose is reabsorbed through SGLT2, a low-affinity, high-capacity transporter located predominantly in the S1 segment of the renal proximal tubule, and the remainder is reabsorbed through SGLT1, a high-affinity, low-capacity transporter located in the S2 and S3 segments^{4–6}.

The maximum tubular reabsorption capacity for glucose is significantly increased in patients with diabetes^{7,8}. As a result, renal glucose excretion is often disproportionally low in the hyperglycemic state. The inadequate renal excretion of excess plasma glucose likely promotes the elevation of plasma glucose. The augmented reabsorption of glucose by the renal proximal tubule is partly attributable to a compensatory increase in glucose reabsorption through SGLT1. Furthermore, an increase in SGLT2 expression has been shown to be one of the molecular mechanisms responsible for the increase in the renal threshold in diabetic patients^{9–11}. Thus, the reduction of renal glucose reabsorption through the inhibition of SLGT2 is a rational approach to improving glucose metabolism in diabetic patients.

Several SGLT2 selective inhibitors have come onto the market and are available for the treatment of type 2 diabetes. SGLT2 inhibitors might offer some advantages over other classes of hypoglycemic agents. When used as monotherapy, SGLT2 inhibitors have a low risk for hypoglycemia because of their insulin-independent mode of action¹²⁻¹⁷. The urinary excretion of excess glucose could result in decreased glucose toxicity, and thereby preserve islet mass and improve insulin sensitivity. Another favorable effect of SGLT2 inhibitors is a reduction in bodyweight. In contrast, the known adverse effects of SGLT2 inhibitors include an increased risk of dehydration, urinary tract infections and mycotic genital infections because of the increase in urinary glucose. An add-on therapy of an SGLT2 inhibitor to insulin or insulin secretagogues could increase the risk of hypoglycemia. Furthermore, the increased urinary glucose excretion could result in glucose deficiency, leading to ketoacidosis in some patients with insulin secretory defects or those who ingest low-carbohydrate foods.

Although it has been established that SGLT2 inhibitors have hypoglycemic effects in patients with type 2 diabetes, the effects of SGLT2 inhibitors on continuous glucose monitoring (CGM)-recorded glucose curve remain to be fully described. Furthermore, SGLT2 inhibitor-induced changes in diurnal profiles of plasma ketone bodies have not been reported. The aims of the present study were to elucidate the effects of short-term ipragliflozin therapy on diurnal profiles of glucose and 3-hydroxybutyrate (3HB), bodyweight, blood pressure, hematocrit, serum uric acid, electrolytes, insulin resistance, and β -cell function. In addition, we assessed the factors that contribute to the acceleration of ketogenesis induced by the administration of ipragliflozin.

MATERIALS AND METHODS

The participants of the present study were 21 inpatients with type 2 diabetes, aged 57 ± 10 years, with a body mass index (BMI) of 28.9 ± 4.3 kg/m² (Table 1). The required sample size to assess the hypoglycemic effect was calculated to be 16 using the data of fasting plasma glucose in a double-blind glycemic control trial of ipragliflozin in Japanese patients¹⁸. The participants of this study were recruited from patients admitted to the Kurume University Hospital, Kurume, Japan, for glycemic control. The diagnosis of type 2 diabetes was established based on the American Diabetes Association¹⁹ and Japan Diabetes Society²⁰ criteria for diabetes as well as on the absence of pancreatic autoimmune markers, including glutamic acid decarboxylase antibodies and insulinoma-associated-2 antibodies. Patients with renal failure or severe liver disease were excluded. A total of 21 age-, sex- and BMI-matched patients with type 2 diabetes served as a control group (Table 1). We analyzed the changes of metabolic parameters during the second week of admission. Although this was not a randomized study, there was no significant difference between the ipragliflozin group and the control group in metabolic parameters or pharmacological treatment. The patients of both groups ate a

Table 1 Clinical features of participa
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Characteristic	Ipragliflozin group	Control group
Age (years)	57 ± 10	56 ± 11
Sex (male/female)	10/11	0/11
Body mass index (kg/m ²)	28.9 ± 4.3	28.5 ± 4.2
AST (IU/L)	41 ± 29	39 ± 35
ALT (IU/L)	52 ± 41	44 ± 44
Creatinine (mg/dL)	0.66 ± 0.18	0.66 ± 0.22
eGFR (mL/min/1.73 m ²)	88.2 ± 22.0	90.9 ± 26.1
Total cholesterol (mg/dL)	183 ± 46	183 ± 37
HDL cholesterol (mg/dL)	47.8 ± 8.5	46.2 ± 9.9
LDL cholesterol (mg/dL)	114.8 ± 32.6	116.2 ± 31.4
Triglyceride (mg/dL)	161 ± 103	163 ± 94
Fasting plasma glucose (mg/dL)	181 ± 48	161 ± 36
HbA1c (%)	10.2 ± 2.2	9.3 ± 1.6
Fasting serum CPR (ng/mL)	2.3 ± 1.0	2.5 ± 1.1
Pharmacological treatment (n)		
Insulin	0	2
Metformin	12	15
Glimepiride	7	8
DPP4 inhibitors	6	5
Pioglitazone	1	0
Exenatide	1	0

ALT, alanine transaminase; AST, aspartate transaminase; CPR, C-peptide immunoreactivity; DPP4, dipeptidyl peptidase-4; eGFR, estimated glomerular filtration rate; HbA1c, glycated hemoglobin; HDL, high-density lipoprotein; LDL, low-density lipoprotein. standardized low-fat meal at 08.00, 12.00 and 18.00 h, and carried out only mild exercise, such as walking, during the study period. The maximum dose of glimepiride was 2 mg/day at baseline. The doses of the hypoglycemic agents were unchanged during the study period.

After an approximately 7-day stabilizing period in a diabetes/endocrinology ward, 50 mg of ipragliflozin was given to each patient of the ipragliflozin group after breakfast for 7 days. In our clinical experience, plasma glucose concentration declined on the second day of ipragliflozin treatment and appeared to reach stable levels within a week. Blood samples were obtained on the day before the initiation of ipragliflozin (day 0) and on the 7th day of the ipragliflozin treatment (day 7). We measured the pre- and postprandial levels of plasma glucose, 3HB and free fatty acid (FFA). The 3HB and



Figure 1 | (a) Urinary glucose excretion and (b) urine volume during the ipragliflozin treatment. Mean and standard deviation.

FFA values were measured by enzymatic methods using an auto analyzer (Hitachi High-Tech, Tokyo, Japan). The $iPro^{TM}$ 2 (Medtronic, Northridge, CA, USA) CGM devices were used to monitor subcutaneous glucose levels at 5-min intervals to determine glycemic control on day 0 and day 7. The CGM device was applied to each patient no later than noon of the preceding day, and glucose levels were measured for at least 48 h including day 0 or day 7 during hospitalization. CGM data on day 0 and day 7 were used to calculate mean amplitude of glycemic excursions, mean postprandial glucose



Figure 2 | (a) Changes in bodyweight in the ipragliflozin group (closed circle) and the control group (open circle). Mean and standard deviation. *P < 0.01, **P < 0.001 vs control group. (b) Preprandial plasma glucose levels during the 3 days preceding the initiation of ipragliflozin and during the 7-day ipragliflozin treatment.

excursion, constant variation, percentage of time \geq 140 mg/dL or <70 mg/dL, area under the curve \geq 180 mg/dL and area over the curve <70 mg/dL. Insulin resistance and insulin secretory capacity were assessed by the homeostasis model assessment of estimated insulin resistance (HOMA-IR) and HOMA of β -cell function (HOMA- β), respectively. Bodyweight and blood pressure were measured before breakfast every day. We also measured urinary glucose excretion and urine volume per day. The primary end-point was the improvement of diurnal glucose profile. Secondary end-points included changes in plasma 3HB, bodyweight, blood pressure, hematocrit, uric acid, electrolytes, HOMA-IR, HOMA- β and adverse events including hypoglycemia.

The study was approved by the ethical committee of Kurume University, and was carried out in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments. The procedure was explained, and written informed consent was obtained from all patients.

Statistical Analysis

The data are expressed as the mean \pm standard deviation (SD). Statistical analysis was carried out using SAS v.9.3 (SAS Institute, Cary, NC, USA). Pearson's correlations were used to evaluate the factors associated with the pre-breakfast and

pre-dinner 3HB levels. The paired Student's *t*-test was used to test the difference in the means between baseline and post-treatment data. The serum levels of aspartate transaminase, alanine transaminase, triglyceride, immunoreactive insulin and 3HB were transformed into logarithms to improve the skewed distribution. The results with a P < 0.05 were considered statistically significant.

RESULTS

Urinary glucose excretion and urine volume were increased on the first day of the ipragliflozin treatment, and remained elevated throughout the study period (Figure 1). The average increases in urine glucose and urine volume were 55.4 ± 21.7 g/day and 468 ± 552 mL/day, respectively. The patients lost 1.4 ± 0.5 kg of bodyweight during the 7-day ipragliflozin treatment (Figure 2a). The reduction of bodyweight was significantly greater than that of the control group $(0.5 \pm 0.6$ kg, P < 0.0001). Preprandial glucose levels were approximately stable during the 3-day period preceding the initiation of ipragliflozin (Figure 2b). When the ipragliflozin administration began, its hypoglycemic effect appeared quickly; pre-lunch and pre-dinner glucose levels fell on the first day, and pre-breakfast glucose values were reduced on the second day. The 7-day ipragliflozin treatment resulted in significantly



Figure 3 | The diurnal variations of (a) plasma glucose, (b) immunoreactive insulin (IRI), (c) free fatty acid (FFA) and (d) 3-hydroxybutyrate (3HB) on day 0 (closed circle) and on day 7 (open circle) of the ipragliflozin treatment. Mean and standard deviation. Means and standard deviations of the IRI, FFA, and 3HB levels were calculated after log transformation of the data. *P < 0.05, **P < 0.01, ***P < 0.001. AB, after breakfast; AD, after dinner; AL, after lunch; BB, before breakfast; BD, before dinner; BL, before lunch.

lower plasma glucose levels compared with baseline levels at each time-point of blood sampling (Figure 3a).

The CGM showed a significant reduction in glucose levels throughout the day (Figure 4a,b). The 24-h average glucose level decreased from $182 \pm 54 \text{ mg/dL}$ to $141 \pm 33 \text{ mg/dL}$ (P < 0.0001). No hypoglycemia was detected during the study period. There was no significant difference between patients receiving sulfonvlurea and those without sulfonvlurea in glucose levels at any time-point. Glucose levels below 70 mg/dL were recorded by CGM only in three patients without sulfonylurea treatment. The lowest glucose value was 75 mg/dL in the patients receiving sulfonylurea. The proportion of time spent in the 70-139 mg/dL blood glucose range increased from $29.2 \pm 30.9\%$ to $57.3 \pm 26.5\%$ (*P* < 0.0001; Figure 4c,d). Mean postprandial glucose excursion and area under the curve >180 mg/dL were significantly reduced after the 7-day ipragliflozin treatment (Table 2). The magnitude of reduction in the 24-h average glucose concentration after the ipragliflozin treatment was highly correlated with the average glucose level on day 0 (Figure 5a). Urinary glucose excretion was correlated with the average glucose level on both day 0 and day 7, although the regression line was steeper and shifted leftward on day 7 of the ipragliflozin treatment (Figure 5b).

The pre- and postprandial plasma IRI levels were almost comparable between day 0 and day 7, although IRI levels were slightly but significantly lower before lunch, after lunch and before dinner on day 7 (Figure 3b). Consequently, HOMA-IR was decreased from 3.55 ± 1.84 to 2.11 ± 1.26 (P = 0.001), and HOMA- β was increased from 38.6 ± 28.6 to 51.6 ± 32.4 (P = 0.015). In contrast, plasma FFA levels were significantly higher except after breakfast on day 7 than on day 0 (Figure 3c).

 Table 2 | Parameters obtained from continuous glucose monitoring data of 21 participants treated with ipragliflozin

	Day 0	Day 7	P-value
MAGE (mg/dL)	94.3 ± 64.9	82.0 ± 32.2	NS
MPPGE (mg/dl.)	44.7 ± 29.3	30.9 ± 20.1	0.008
CV	0.222 ± 0.068	0.222 ± 0.064	NS
AUC >180 mg/dL	644 ± 472	230 ± 269	<0.0001
(mg min/dL) AOC <70 mg/dL (mg min/dL)	3 ± 12	19 ± 57	NS

AOC, area over the curve; AUC, area under the curve; CV, constant variation; MAGE, mean amplitude of glycemic excursion; MPPGE, mean of postprandial glucose excursion; NS, not significant.

Similarly, plasma levels of 3HB were significantly elevated at each time-point on day 7, with peaks before breakfast and before dinner (Figure 3d). There were significant correlations between plasma FFA and 3HB levels (P < 0.05) at each point of blood collection on day 7. Pearson's correlation analysis showed that the patient age and bodyweight loss were negatively and positively correlated, respectively, with the pre-breakfast and pre-dinner 3HB levels after the 7-day ipragliflozin treatment (Table 3). No significant changes were observed in blood pressure, hematocrit, uric acid or electrolytes except phosphate (Table 4).

DISCUSSION

We showed, using venous blood sampling and CGM, that the 7-day administration of ipragliflozin shifted the 24-h glucose profile downward without hypoglycemia. Although a number



Figure 4 | Average glucose levels of the 21 participants as measured by continuous glucose monitoring on (a) day 0 and (b) day 7. Mean, mean + standard deviation, and mean – standard deviation. The difference was statistically significant at all time-points. The proportion of time spent at glucose level <70 mg/dL, 70–139 mg/dL and \ge 140 mg/dL on (c) day 0 and (d) day 7.



Figure 5 | (a) Association between the average glucose level by continuous glucose monitoring (CGM) at baseline and the reduction in the average daily glucose concentration during the 7-day ipragliflozin treatment. (b) Association between the average glucose level of the CGM and 24-h urine glucose excretion on day 0 (closed circle) and on day 7 (open circle) of the ipragliflozin treatment.

of clinical trials^{12,14–17,21,22} have reported that SGLT2 inhibitors reduced both fasting and postprandial glucose levels, the effect of SGLT2 inhibitor administration on the nocturnal glucose profile remains to be elucidated. In the present study, the stable nocturnal glucose profile shown by CGM after the ipragliflozin treatment was most impressive. It is also noteworthy that no hypoglycemia was observed despite the reduction in preprandial glucose levels, although the number of the participants was not large. The decrease of mean postprandial glucose excursion shows that postprandial glucose excursion was reduced by the ipragliflozin treatment. The steady control of nocturnal glucose levels and the absence of hypoglycemia were associated with the observation that the magnitude of the hypoglycemic action was closely correlated with the 24-h average plasma glucose level on day 0. This finding is consistent with previous reports in which glycated hemoglobin reductions with SGLT2 inhibitors were greater in the subgroup with higher baseline glycated hemoglobin^{13,18,22}.

A pharmacodynamic study of dapagliflozin reported that treatment with the SGLT2 inhibitor markedly reduced the renal threshold of plasma glucose²³. Although the calculated threshold was extremely low $(21 \pm 46 \text{ mg/dL})$ in the dapagliflozin study, the urinary glucose excretion fell as plasma glucose decreased. Here, we show that ipragliflozin-induced urinary glucose excretion was correlated with the average glucose concentration obtained by the CGM, although the regression line was steeper and left-shifted compared with that before the treatment. It is unknown whether there is some difference between dapagliflozin and ipragliflozin with respect to their pharmacodynamic profiles. However, the correlation between the reduction in plasma glucose and the baseline glucose levels can be attributed to the association between urinary glucose excretion and plasma glucose concentration. Thus, the hypoglycemic risk of ipragliflozin seems to be low, except when it is combined with insulin or large-dose sulfonylureas.

Despite the marked reduction in plasma glucose levels, the 7-day treatment with ipragliflozin had only small effects on pre- and postprandial IRI levels. The pre-breakfast IRI value did not significantly change; therefore, HOMA-IR and HOMA-B was decreased and increased, respectively, after the treatment, likely reflecting reversal of glucose toxicity. Although the validity of the HOMA model is limited in hyperglycemic conditions and the adequacy of HOMA indices has not been established in patients receiving SGLT2 inhibitors, these observations suggest that ipragliflozin treatment ameliorates insulin resistance and insulin secretory capacity in diabetic patients. Thus far, the beneficial effects of SGLT2 inhibitors on insulin secretory capacity and insulin sensitivity have been shown in longer-term administration²⁴⁻²⁷. Our findings indicate that the improvements in β-cell function and insulin sensitivity occurred within a week of initiating SGLT2 inhibitor treatment.

Recently, we have reported that the plasma levels of 3HB and FFA show marked diurnal fluctuation in patients with type 2 diabetes, with two obvious peaks before breakfast and before dinner. Furthermore, the peak levels were inversely correlated with age²⁸. The administration of sulfonylureas or DPP-4 inhibitors was shown to suppress the plasma 3HB levels, likely through their insulinotropic and/or glucagonstatic action. In contrast, the renal glucose loss induced by ipragliflozin might result in increased lipolysis and accelerated ketogenesis. In the present study, the plasma levels of both 3HB and FFA were increased after 7-day ipragliflozin administration, with the most obvious elevations in the pre-breakfast and pre-dinner 3HB levels. The positive correlation between

Table 3 | Pearson's correlation analysis of the factors associated with the pre-breakfast and pre-dinner 3-hydroxybutyrate levels on the 7th day of the ipragliflozin treatment

	Pre-breakfast 3HB		Pre-dinner 3HB	
	Correlation coefficient	<i>P</i> -value	Correlation coefficient	P-value
Age	-0.70833	0.0003	-0.53635	0.0122
Sex (male)	-0.20889	NS	-0.17064	NS
BMI	-0.02143	NS	0.03072	NS
Reduction in bodyweight	0.66052	0.0011	0.64799	0.0015
AST	0.33098	NS	0.38878	NS
ALT	0.27228	NS	0.39473	NS
Total cholesterol	0.28012	NS	0.17225	NS
HDL cholesterol	0.03182	NS	-0.07591	NS
LDL cholesterol	0.22337	NS	0.09282	NS
Triglyceride	0.29377	NS	0.21491	NS
eGFR	0.37494	NS	0.31113	NS
Systolic blood pressure	-0.31088	NS	-0.05392	NS
Diastolic blood pressure	0.25361	NS	0.29684	NS
Pre-breakfast plasma glucose	0.15811	NS	-0.17735	NS
Pre-dinner plasma glucose	0.26006	NS	-0.27157	NS
Pre-breakfast IRI	0.19529	NS	0.21952	NS
Pre-dinner IRI	0.11574	NS	-0.11036	NS
Use of metformin	-0.41113	NS	-0.04505	NS
Use of sulfonylurea	-0.22499	NS	-0.01297	NS
Use of DPP4 inhibitors	-0.27087	NS	0.02257	NS

Aspartate transaminase (AST), alanine transaminase (ALT), triglyceride, immunoreactive insulin (IRI) and 3-hydroxybutyrate (3HB) levels were transformed into logarithms to improve the skewed distribution. BMI, body mass index; DPP4, dipeptidyl peptidase-4; eGFR, estimated glomerular filtration rate; HDL, high-density lipoprotein; LDL, low-density lipoprotein; NS, not significant.

 Table 4 | Changes in blood pressure, hematocrit, uric acid and electrolytes

	lpragliflozin group		Control group	
	Day 0	Day 7	Day 0	Day 7
Systolic blood pressure (mmHg)	117 ± 16	116 ± 13	122 ± 11	123 ± 15
Diastolic blood pressure (mmHg)	78 ± 8	73 ± 11	73 ± 12	71 ± 10
Hematocrit (%)	41.1 ± 3.9	40.8 ± 4.1	39.2 ± 5.1	39.3 ± 4.9
Uric acid (mg/dL)	5.28 ± 1.94	4.87 ± 1.46	6.06 ± 1.51	6.34 ± 1.68
Na (mEq/L)	141 ± 2	141 ± 2	141 ± 2	141 ± 2
K (mEq/L)	4.0 ± 0.3	4.1 ± 0.3	4.0 ± 0.4	4.1 ± 0.4
Cl (mEq/L)	104 ± 2	105 ± 2	104 ± 2	104 ± 2
Ca (mg/dL)	9.11 ± 0.29	9.19 ± 0.31	9.13 ± 0.23	9.04 ± 0.32
Pi (mg/dL)	3.62 ± 0.43	$3.93 \pm 0.35^*$	3.79 ± 0.70	3.80 ± 0.57

*P = 0.002 vs day 0.

plasma FFA and 3HB levels suggest that the enhanced ketogenesis could be partly attributable to the elevation of plasma FFA as a result of accelerated lipolysis. As expected, the 3HB levels before breakfast and before dinner were correlated with the reduction of bodyweight during the treatment. Interestingly, the plasma levels of 3HB were inversely correlated with age, suggesting that younger individuals, rather than elderly persons, are susceptible to hyperketonemia when a SGLT2 inhibitor is administered. Although mild to moderate increases in plasma ketone body levels during SGLT2 inhibitor therapy might be physiological and harmless, as in calorie restriction, marked hyperketonemia with a risk of ketoacidosis should be avoided.

The participants in the ipragliflozin group lost 1.4 ± 0.5 kg of bodyweight during the treatment. When compared with the control participants, the ipragliflozin-induced bodyweight loss was estimated to be approximately 1 kg. Although the elevation of plasma FFA and 3HB, and the correlation

between 3HB levels and bodyweight loss show that lipolysis was largely involved in the reduction of bodyweight, it is likely that water loss partly contributed to the decrease of bodyweight.

SGLT-2 inhibitors have been shown to increase hematocrit^{18,21,24}, and lower blood pressure^{12,13,15,17,24} and serum uric acid^{12,15,24} in clinical trials. However, no significant change was observed in blood pressure, hematocrit or serum uric acid in the present short-term study. Serum phosphate concentration was slightly but significantly increased after the ipragliflozin treatment, although the mechanism and physiological significance of the elevation are not known.

In conclusion, the SGLT2 inhibitor, ipragliflozin, showed a notable hypoglycemic effect in both the preprandial and postprandial states, and improved the 24-h glucose curve without hypoglycemia. The beneficial effects on β -cell function and insulin resistance became apparent within 1 week of the treatment. The magnitude of glucose-lowering action was correlated with plasma glucose concentrations; therefore, the hypoglycemic risk is likely low. When ipragliflozin is added to insulin or large-dose sulfonylureas, the careful prevention of hypoglycemia could be required soon after the initiation, because the hypoglycemic effect appeared quickly. It might be prudent to monitor plasma or urine ketone body levels in younger patients and in patients with rapid weight loss.

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DISCLOSURE

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