

electrical activity of the diaphragm (1). Whether (central) inhibition (or inhibition of the supplementary motor cortex) is present in ventilated patients with abnormal sleep remains to be determined.

The elegant investigation of Rault and colleagues (1) is provocative. The investigators have set the stage for the objective study of the physiologic maze that accompanies sleep deprivation. One challenge will be to unravel the sex-specific effect of sleep deprivation on dyspnea, spinal and supraspinal reflex inhibition, and function of the primary motor cortex. Another challenge will be to determine the effect of sleep deprivation in critically ill patients, including those who fail invasive and noninvasive ventilation. The challenge is formidable, but now is the time to tackle it. ■

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## References

- Rault C, Sangaré A, Diaz V, Ragot S, Frat J-P, Raux M, *et al*. Impact of sleep deprivation on respiratory motor output and endurance: a physiological study. *Am J Respir Crit Care Med* 2020;201:976–983.
- Banzett RB, O'Donnell CR, Guilfoyle TE, Parshall MB, Schwartzstein RM, Meek PM, *et al*. Multidimensional dyspnea profile: an instrument for clinical and laboratory research. *Eur Respir J* 2015;45:1681–1691.
- Parshall MB, Schwartzstein RM, Adams L, Banzett RB, Manning HL, Bourbeau J, *et al*. American Thoracic Society Committee on Dyspnea. An official American Thoracic Society statement: update on the mechanisms, assessment, and management of dyspnea. *Am J Respir Crit Care Med* 2012;185:435–452.
- Elvsåshagen T, Mutsaerts HJ, Zak N, Norbom LB, Quraishi SH, Pedersen PØ, *et al*. Cerebral blood flow changes after a day of wake, sleep, and sleep deprivation. *Neuroimage* 2019;186:497–509.
- Rosales-Lagarde A, Armony JL, Del Río-Portilla Y, Trejo-Martínez D, Conde R, Corsi-Cabrera M. Enhanced emotional reactivity after selective REM sleep deprivation in humans: an fMRI study. *Front Behav Neurosci* 2012;6:25.
- Laghi F, Shaikh HS, Morales D, Sinderby C, Jubran A, Tobin MJ. Diaphragmatic neuromechanical coupling and mechanisms of hypercapnia during inspiratory loading. *Respir Physiol Neurobiol* 2014;198:32–41.
- Raux M, Demoule A, Redolfi S, Morelot-Panzini C, Similowski T. Reduced phrenic motoneuron recruitment during sustained inspiratory threshold loading compared to single-breath loading: a twitch interpolation study. *Front Physiol* 2016;7:537.
- De Gennaro L, Marzano C, Veniero D, Moroni F, Fratello F, Curcio G, *et al*. Neurophysiological correlates of sleepiness: a combined TMS and EEG study. *Neuroimage* 2007;36:1277–1287.
- Civardi C, Boccagni C, Vicentini R, Bolamperti L, Tarletti R, Varrasi C, *et al*. Cortical excitability and sleep deprivation: a transcranial magnetic stimulation study. *J Neurol Neurosurg Psychiatry* 2001;71:809–812.
- Kreuzer P, Langguth B, Popp R, Raster R, Busch V, Frank E, *et al*. Reduced intra-cortical inhibition after sleep deprivation: a transcranial magnetic stimulation study. *Neurosci Lett* 2011;493:63–66.
- Roche Campo F, Drouot X, Thille AW, Galia F, Cabello B, d'Ortho M-P, *et al*. Poor sleep quality is associated with late noninvasive ventilation failure in patients with acute hypercapnic respiratory failure. *Crit Care Med* 2010;38:477–485.
- Dres M, Younes M, Rittayamai N, Kendzerska T, Telias I, Grieco DL, *et al*. Sleep and pathological wakefulness at the time of liberation from mechanical ventilation (SLEEWE): a prospective multicenter physiological study. *Am J Respir Crit Care Med* 2019;199:1106–1115.
- Olbrich E, Landolt HP, Achermann P. Effect of prolonged wakefulness on electroencephalographic oscillatory activity during sleep. *J Sleep Res* 2014;23:253–260.
- Jubran A, Tobin MJ. Pathophysiologic basis of acute respiratory distress in patients who fail a trial of weaning from mechanical ventilation. *Am J Respir Crit Care Med* 1997;155:906–915.
- Doorduyn J, Roesthuis LH, Jansen D, van der Hoeven JG, van Hees HWH, Heunks LMA. Respiratory muscle effort during expiration in successful and failed weaning from mechanical ventilation. *Anesthesiology* 2018;129:490–501.

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## Latent Tuberculosis Infection–associated Immunodiagnostic Test Responses as Biomarkers of Incipient Tuberculosis: Fruitful or Futile?

Latent tuberculosis infection (LTBI) constitutes part of the TB disease spectrum (1, 2). The diagnosis and treatment of LTBI is important, as global eradication targets will not be attainable

without treating LTBI (3). These considerations also apply to drug-resistant TB, which threatens to derail control efforts (4). The World Health Organization has recently recommended that close contacts of index cases of TB, even in TB endemic countries, should be considered for LTBI treatment (even if they are HIV-uninfected or not children) (5). However, the diagnosis of LTBI is challenging. Unlike with active TB, in humans there is no microbiological or histopathological reference standard for LTBI, and one can only infer the potential presence of LTBI using immunodiagnostic tests, which enumerate the magnitude of

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relatively antigen-specific T-helper cell type 1 effect or T-cell responses (6). However, it is epidemiologically well-recognized that only a small proportion of individuals with presumed LTBI (~5–10%) will progress to active TB over a lifetime (7). Thus, the more important public health question is whether, and how, we can accurately target treatment by identifying individuals who are most likely to progress to active TB. This state is broadly defined as “incipient TB” and is characterized by a lack of TB-related symptoms and appropriate chest radiographic abnormalities at the time of testing, as well as lack of any microbiological evidence of active TB, but a high likelihood of progression to active TB in the short-term, with the potential for perpetuating the transmission cycle (2). The duration from initial exposure to incipient TB or active disease is variable and will depend on several host, mycobacterial, and environmental factors. Epidemiological data suggest that of those infected, ~5% will progress to active TB over the course of a 5-year period, with the highest risk being within the first 2 years of exposure (8, 9). Biomarkers to identify incipient TB has remained one of the “Holy Grails” of TB research.

Given these considerations, it has often been asked whether a higher magnitude of the IFN- $\gamma$  release assay (IGRA) response, or larger tuberculin skin test (TST) induration diameter, reflecting a higher burden of circulating effector T cells and inferring a higher burden of *Mycobacterium tuberculosis*, predicts a higher likelihood of incipient TB. Indeed, serial IGRA responses increasing in magnitude over time were associated with the development of active disease in several reports, suggesting that antigen-driven T-cell responses could be a marker of incipient TB (10). However, there are limited and conflicting data regarding this point. Zellweger and Haldar found no association between the magnitude of the IFN- $\gamma$  response and progression to active TB (11, 12). In contrast, Winje and colleagues interrogated a large population-based cohort using QuantiFERON Gold In-Tube (QFT-GIT) and found that a quantitative IGRA readout >4.0 IU/L was highly associated (>30-fold risk compared with QFT-GIT negativity) with the development of active TB (13). Using a different metric, Andrews and coworkers from South Africa found that QFT-GIT conversion at IFN- $\gamma$  values higher than 4.00 IU/L (but not below this threshold) was associated with substantially increased risk (42-fold higher risk than nonconverters) of developing active TB over the course of a ~2-year period (14). However, although these data collectively suggested that the magnitude of the T-cell response was associated with higher rates of downstream active TB, there remained several unanswered questions. Did this relationship hold true for other immunodiagnostic readouts (such as T-SPOT.TB and the TST), what are the implications for clinical practice, and is this relationship meaningful and clinically useful?

The study by Gupta and coworkers in this issue of the *Journal* (pp. 984–991) provides answers to some of these questions (15). Their findings were based on the results of the prospective UK PREDICT (UK Prognostic Evaluation of Diagnostic IGRAs Consortium) study that evaluated three immunodiagnostic tests (T-SPOT.TB, QFT-GIT, and TST) in almost 10,000 participants who were at high risk for LTBI (close contacts of active TB cases or recent migrants) sequentially recruited from 54 centers in the United Kingdom (16). They found that although the magnitude of the IGRA (both QFT-GIT and T-SPOT.TB) and the TST response was a biomarker of incipient TB, the threshold-specific positive

predictive value for all three immunodiagnostic tests for active TB over a median follow-up of ~3 years was poor at <5%. This is because there were many nonprogressors who had a magnitude of response at or above the threshold identifying incipient TB. Moreover, using this higher threshold in clinical practice would result in a substantial drop in test sensitivity to detect active TB cases, making the usefulness of such an approach redundant. This is because IGRAs and TSTs are simply poor tests of incipient TB. This is not surprising, as only a small proportion of those with LTBI (~5%) will progress to active disease.

The authors must be commended on undertaking such a challenging study both in terms of recruitment and analysis. The findings are helpful to clinicians and public health physicians who are using immunodiagnostic tests in screening programs. It suggests that alternative biomarkers of incipient TB are urgently needed. A weakness of the study, however, despite the drawbacks of the IGRAs, was the lack of serial testing (discussed here). Such an approach would have only been feasible if the TST was not performed at baseline (as tuberculin contains RD-1 antigen and can boost downstream IGRA responses) (17). To try and circumvent the poor predictive value and specificity, alternative immunodiagnostic readouts have been investigated including different cytokine readouts (e.g., combination of IL-2/IFN- $\gamma$ ), T-cell responses to alternative antigens (e.g., HBHA and Ag85a [18–20]), cell activation markers (e.g., CD4<sup>+</sup> HLA-DR<sup>+</sup> T cells [21]), and readouts from alternative compartments including RD-1–based skin tests that are being commercialized (22).

Other investigators have uncovered biosignatures of incipient TB. Several studies have identified blood-based transcriptional signatures associated with progression to active TB (23–26) with a positive predictive value ~10-fold higher than the IGRAs. These genomic biosignatures, consisting of 3–16 gene transcripts, were able to predict TB progression in participants with LTBI, although a recent systematic review found that performance was variable and better reflected the short-term risk of TB (over 3 to 6 mo). Suliman and colleagues (27) derived a 4-gene signature, which correlated with TB disease progression and performed well when validated against other transcriptomic signatures. However, using RT-PCR–based readouts may not be user-friendly or cost-effective for TB-endemic settings. Very recently, a three- to five-protein biosignature of incipient TB was derived and validated (28), and a novel ultrasensitive phage-based amplification assay for incipient TB was described (29). These data suggest that a point-of-care assay may be a realistic goal once better biomarkers are developed and validated.

Another broader issue raised by this study is the ambiguous and confusing interpretation of IGRA readouts. On the one hand, positive IGRA responses are often interpreted as a marker of LTBI, and hence “protection,” given that ~95% never progress to active disease, and serial IGRA responses often decrease during the course of successful TB treatment (30). In contrast, the work of Gupta and others suggests that IGRAs are a biomarker of incipient TB, and hence TB risk. Which is it, protection or risk? The answer is both, depending on the clinical context. Thus, the conundrum can be resolved by recognizing that TB is a spectrum of infection, which is a dynamic interplay between host and pathogen at the level of the granuloma, and this may change over time, reflecting time point–specific host immunity

and mycobacterial disease burden (and hence changing levels of TB-specific effector T cells in blood). A temporal compartment-specific effect may also influence interpretation as a result of translocation of antigen-specific T cells from the blood to the disease site (e.g., the lung) (31). Thus, serial measurements may often be required to determine whether IGRA responses are stable, increase in magnitude over time (conversion), or reduce in magnitude over time (reversion), possibly suggesting clearance of infection (32). This concept has been well outlined in a recent review (6). Thus, IGRA readouts can be a marker of protection or susceptibility depending on the context. This will explain why patients with stable, persistently positive IGRA responses remain asymptomatic and do not progress to active TB over many years, whereas those with increasing IFN $\gamma$ -specific spot-forming units and/or responses progress to active TB, and others may revert to presumably clear their infection. Thus, selecting vaccine candidates simply on their ability to induce or drive antigen-specific IFN- $\gamma$  responses is counterintuitive; rather, selection based on preventing sustained conversion seems more logical and is an approach that has recently been used (33).

For now, the findings of Gupta and colleagues are clinically useful and point us in the right direction. The bottom line is that better biomarkers of incipient TB are required, and nascent biomarker signatures require urgent prospective clinical validation. It is hoped that these resource-intensive and challenging prospective validation studies (e.g., the CORTIS [The Correlate of Risk Targeted Intervention Study] study [34]) will be fruitful rather than futile, as TB remains the foremost infectious disease killer globally. ■

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## References

1. Dheda K, Barry CE III, Maartens G. Tuberculosis. *Lancet* 2016;387:1211–1226.

2. Drain PK, Bajema KL, Dowdy D, Dheda K, Naidoo K, Schumacher SG, *et al.* Incipient and subclinical tuberculosis: a clinical review of early stages and progression of infection. *Clin Microbiol Rev* 2018;31:e00021–18.
3. Houben RMGJ, Menzies NA, Sumner T, Huynh GH, Arinaminpathy N, Goldhaber-Fiebert JD, *et al.* Feasibility of achieving the 2025 WHO global tuberculosis targets in South Africa, China, and India: a combined analysis of 11 mathematical models. *Lancet Glob Health* 2016;4:e806–e815.
4. Dheda K, Gumbo T, Maartens G, Dooley KE, Murray M, Furin J, *et al.*; Lancet Respiratory Medicine drug-resistant tuberculosis Commission group. The Lancet Respiratory Medicine Commission: 2019 update: epidemiology, pathogenesis, transmission, diagnosis, and management of multidrug-resistant and incurable tuberculosis. *Lancet Respir Med* 2019;7:820–826.
5. World Health Organization. Latent TB infection: updated and consolidated guidelines for programmatic management. Geneva, Switzerland: WHO; 2018.
6. Pai M, Denkinger CM, Kik SV, Rangaka MX, Zwering A, Oxlade O, *et al.* Gamma interferon release assays for detection of *Mycobacterium tuberculosis* infection. *Clin Microbiol Rev* 2014;27:3–20.
7. Schwander S, Dheda K. Human lung immunity against *Mycobacterium tuberculosis*: insights into pathogenesis and protection. *Am J Respir Crit Care Med* 2011;183:696–707.
8. Ferebee SH. Controlled chemoprophylaxis trials in tuberculosis: a general review. *Bibl Tuberc* 1970;26:28–106.
9. Sloot R, Schim van der Loeff MF, Kouw PM, Borgdorff MW. Risk of tuberculosis after recent exposure: a 10-year follow-up study of contacts in Amsterdam. *Am J Respir Crit Care Med* 2014;190:1044–1052.
10. Pai M, O'Brien R. Serial testing for tuberculosis: can we make sense of T cell assay conversions and reversions? *PLoS Med* 2007;4:e208.
11. Haldar P, Thuraingam H, Patel H, Pereira N, Free RC, Entwisle J, *et al.* Single-step QuantiFERON screening of adult contacts: a prospective cohort study of tuberculosis risk. *Thorax* 2013;68:240–246.
12. Zellweger J-P, Sotgiu G, Block M, Dore S, Altet N, Blunski R, *et al.*; TBNET. Risk assessment of tuberculosis in contacts by IFN- $\gamma$  release assays: a Tuberculosis Network European Trials Group study. *Am J Respir Crit Care Med* 2015;191:1176–1184.
13. Winje BA, White R, Syre H, Skutlberg DH, Oftung F, Mengshoel AT, *et al.* Stratification by interferon- $\gamma$  release assay level predicts risk of incident TB. *Thorax* 2018;73:652–661.
14. Andrews JR, Nemes E, Tameris M, Landry BS, Mahomed H, McClain JB, *et al.* Serial QuantiFERON testing and tuberculosis disease risk among young children: an observational cohort study. *Lancet Respir Med* 2017;5:282–290.
15. Gupta RK, Lipman M, Jackson C, Sitch AJ, Southern J, Drobniewski F, *et al.* Quantitative IFN- $\gamma$  release assay and tuberculin skin test results to predict incident tuberculosis: a prospective cohort study. *Am J Respir Crit Care Med* 2020;201:984–991.
16. Abubakar I, Drobniewski F, Southern J, Sitch AJ, Jackson C, Lipman M, *et al.*; PREDICT Study Team. Prognostic value of interferon- $\gamma$  release assays and tuberculin skin test in predicting the development of active tuberculosis (UK PREDICT TB): a prospective cohort study. *Lancet Infect Dis* 2018;18:1077–1087.
17. van Zyl-Smit RN, Pai M, Pehrah K, Meldau R, Kieck J, Juritz J, *et al.* Within-subject variability and boosting of T-cell interferon- $\gamma$  responses after tuberculin skin testing. *Am J Respir Crit Care Med* 2009;180:49–58.
18. Temmerman ST, Place S, Debie A-S, Loch C, Mascart F. Effector functions of heparin-binding hemagglutinin-specific CD8 $^{+}$  T lymphocytes in latent human tuberculosis. *J Infect Dis* 2005;192:226–232.
19. Macedo GC, Bozzi A, Weinreich HR, Bafica A, Teixeira HC, Oliveira SC. Human T cell and antibody-mediated responses to the *Mycobacterium tuberculosis* recombinant 85A, 85B, and ESAT-6 antigens. *Clin. Dev. Immunol.* 2011;2011:351573.
20. Suter-Riniker F, Berger A, Mayor D, Bittel P, Iseli P, Bodmer T. Clinical significance of interleukin-2/gamma interferon ratios in *Mycobacterium tuberculosis*-specific T-cell signatures. *Clin Vaccine Immunol* 2011;18:1395–1396.

21. Fletcher HA, Snowden MA, Landry B, Rida W, Satti I, Harris SA, *et al.* T-cell activation is an immune correlate of risk in BCG vaccinated infants. *Nat Commun* 2016;7:11290.
22. Hoff ST, Peter JG, Theron G, Pascoe M, Tingskov PN, Aggerbeck H, *et al.* Sensitivity of C-Tb: a novel RD-1-specific skin test for the diagnosis of tuberculosis infection. *Eur Respir J* 2016;47:919–928.
23. Maertzdorf J, McEwen G, Weiner J III, Tian S, Lader E, Schriek U, *et al.* Concise gene signature for point-of-care classification of tuberculosis. *EMBO Mol Med* 2016;8:86–95.
24. Sweeney TE, Braviak L, Tato CM, Khatri P. Genome-wide expression for diagnosis of pulmonary tuberculosis: a multicohort analysis. *Lancet Respir Med* 2016;4:213–224.
25. Zak DE, Penn-Nicholson A, Scriba TJ, Thompson E, Suliman S, Amon LM, *et al.*; ACS and GC6-74 cohort study groups. A blood RNA signature for tuberculosis disease risk: a prospective cohort study. *Lancet* 2016;387:2312–2322.
26. Roe J, Venturini C, Gupta RK, Gurry C, Chain BM, Sun Y, *et al.* Blood transcriptomic stratification of short-term risk in contacts of tuberculosis. *Clin Infect Dis* 2020;70:731–737.
27. Suliman S, Thompson E, Sutherland J, Weiner Rd J, Ota MOC, Shankar S, *et al.*; GC6-74 and ACS cohort study groups. Four-gene pan-African blood signature predicts progression to tuberculosis. *Am J Respir Crit Care Med* 2018;197:1198–1208.
28. Penn-Nicholson A, Hraha T, Thompson EG, Sterling D, Mbandi SK, Wall KM, *et al.*; ACS and GC6-74 cohort study groups. Discovery and validation of a prognostic proteomic signature for tuberculosis progression: a prospective cohort study. *PLoS Med* 2019;16:e1002781.
29. Verma R, Swift BMC, Handley-Hartill W, Lee JK, Woltmann G, Rees CED, *et al.* A novel, high-sensitivity bacteriophage-based assay identifies low-level *Mycobacterium tuberculosis* bacteremia in immunocompetent patients with active and incipient tuberculosis. *Clin Infect Dis* 2020;70:933–936.
30. Theron G, Peter J, Lenders L, van Zyl-Smit R, Meldau R, Govender U, *et al.* Correlation of mycobacterium tuberculosis specific and non-specific quantitative Th1 T-cell responses with bacillary load in a high burden setting. *PLoS One* 2012;7:e37436.
31. Dheda K, van Zyl-Smit RN, Meldau R, Meldau S, Symons G, Khalfey H, *et al.* Quantitative lung T cell responses aid the rapid diagnosis of pulmonary tuberculosis. *Thorax* 2009;64:847–853.
32. Dheda K, Schwander SK, Zhu B, van Zyl-Smit RN, Zhang Y. The immunology of tuberculosis: from bench to bedside. *Respirology* 2010;15:433–450.
33. Nemes E, Geldenhuys H, Rozot V, Rutkowski KT, Ratangee F, Bilek N, *et al.*; C-040-404 Study Team. Prevention of *M. tuberculosis* infection with H4:IC31 vaccine or BCG revaccination. *N Engl J Med* 2018;379:138–149.
34. Fiore-Gartland A, Carpp LN, Naidoo K, Thompson E, Zak DE, Self S, *et al.* Considerations for biomarker-targeted intervention strategies for tuberculosis disease prevention. *Tuberculosis (Edinb)* 2018;109:61–68.

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